Review Article Application of animal and human PET in cardiac research

Quan Wang^{1*}, Zhi-Gang He^{2*}, Shun-Yuan Li³, Mao-Hui Feng⁴, Hong-Bing Xiang¹

Departments of ¹Anesthesiology and Pain Medicine, ²Emergency Medicine, Tongji Hospital of Tongji Medical College, Huazhong University of Science and Technology, Wuhan 430030, Hubei, PR China; ³Department of Anesthesiology, The First Affiliated Quanzhou Hospital of Fujian Medical University, Quanzhou 362000, PR China; ⁴Department of Gastrointestinal Surgery, Zhongnan Hospital, Wuhan University, No. 169 Donghu Road, Wuhan 430071, PR China. *Equal contributors.

Received February 25, 2018; Accepted April 19, 2018; Epub June 15, 2018; Published June 25, 2018

Abstract: Purpose of Review: After a warm-up period of imaging research, several modalities of positron emission tomography (PET) are under development for evaluating ischemic heart disease. Recent Findings: Several types of well-documented stem/progenitor PET imaging have been utilized for changes in myocardial blood flow and carbo-hydrate metabolism. Some recent experimental and human studies reported that these data may have beneficial effects on cardiac research. Summary: Although the role of PET in the pathology of ischemic heart disease has not been sufficiently elucidated, many studies attempting imaging research of myocardial metabolism and neural regulation have been reported. Further studies are needed to better evaluate the potential of PET in evaluating ischemic heart disease.

Keywords: Positron emission tomography, cardiac research, ischemic heart disease

Introduction

Cardiac ischemia is the serious event in heart surgery. A great need exists for improved formulations and mechanisms to prevent and protect the myocardial tissues from reperfusion damage caused by myocardial ischemia. Current efforts to prevent reperfusion damage to the myocardial tissues, which in many cases leads to myocardial infarction and circulatory arrest [1-3]. The neural regulation is involved in an imbalance in metabolic supply and demand within the ischemic myocardial tissues [4-8], which is a natural prevention from ischemia and reperfusion-associated tissue inflammation and organ dysfunction. Modern imaging, such as positron emission tomography (PET), has revolutionized our view of ischemic heart disease [9-13], allowing the opportunity to investigate the metabolic regulation mechanisms of heart by measuring the changes in myocardial blood flow or carbohydrate metabolism, and to offer potential information to further improve prognostic outcome of ischemic heart disease [14-17].

Positron-emitting tracers

Today, the field of PET medicine is undergoing great development [18]. Traditionally, there have been several options for positron-emitting tracers, i.e., ¹⁵O-water, ¹³N-ammonia and ⁸²Rubidium. However, some new sources of positron-emitting tracers were successively accumulated, such as ¹⁸F-labeled myocardial flow radiotracer flurpiridaz [14], and the potential for PET in conjunction with several radiotracers seems to be expanding very rapidly. Rapid development of labeling biologic chemistry gives great potential for the development of new PET tracer candidates [15]. It is known that nanoparticle imaging rely on MRI detection of the iron oxide cores [19, 20], and a study of Ueno [21] showed that dextran nanoparticles the PET isotope copper-64 detected heart transplant rejection and predicted organ survival by reporting on myeloid cells.

The applications of an $^{18}\text{F-labeled}$ perfusion agent [^{18}F fluorodeoxyglucose ($^{18}\text{F-FDG}$) and $^{18}\text{F-sodium}$ fluoride ($^{18}\text{F-NaF}$)] for PET have

Researcher	Species	Radionuclide	Application
Daly [24]	C57BL/6 mice	$[^{13}\text{N}]\text{NH}_3$ and $[^{18}\text{F}]\text{FDG}$	Monitoring the development of cardiac allograft rejection
Hoff [16]	Rat	¹³ NH3 and [¹⁸ F]FDG	A potential diagnostic role of PET
Ueno [21]	Female 57BL/6 mice	Isotope copper-64	Predicting organ survival by reporting on myeloid cells
Srivatsava [17]	Patients	[¹⁸ F]FDG	The assessment of myocardial viability in patients with left ventricular dysfunction
Meeder [13]	Patients	[¹³ N]NH ₃	Exploring the pathophysiology of smoking-related coronary events
Gerber [25]	Patients	[¹⁸ F]FDG	Predicting recovery of global cardiac function
Siebelink [26]	Patients	$[^{13}\text{N}]\text{NH}_3$ and $[^{18}\text{F}]\text{FDG}$	The assessment of revascularization with suspicion of jeopardized myocardium
De Jong [27]	Patients	[(11)C]CGP 12177	Measurement of myocardial beta-adrenoceptor density
De Boer [28]	Patients	Tc-MIBI and [18F]FDG	The assessment of myocardial viability

Table 1. Radiotracer characteristics and application of PET in cardiac Research

revealed details on the pathophysiology of cardiovascular diseases [22]. Some radiotracers have unique effects, such as F-labeled fluorodeoxyglucose ([F]FDG) reflects glucose flux and N-labeled ammonia ([N]NH3) stand for a biomarker of blood flow [23].

The relevant characteristics of radiotracer, animal models compared with human disease are listed in **Table 1** and discussed below.

PET imaging to monitor the allograft rejection

Owing to have the potential to be a specific, sensitive and quantitative diagnostic test, PET imaging in conjunction with radiotracers such as F-labeled fluorodeoxyglucose ([F]FDG) reflecting glucose flux and N-labeled ammonia ([N] NH3) reflecting blood flow, is increasingly used in clinical routine for transplant rejection detection [29, 30], yielding high diagnostic information, while providing valuable outcome in human transplant recipients [31]. Hoff [16] evaluated for the ability of positron-emitting tracers [13NH3 and 18F 2-fluoro 2-deoxyglucose (18F-FDG)] to detect acute allograft rejection after heterotopic cardiac transplantation in the rat with sham-operated controls, nonrejecting isografts, and rejecting allografts, and found that uptake of ¹⁸F-FDG and ¹³NH3 in native hearts of animals from all experimental groups is not significantly different from that in shamoperated controls, suggesting that glucose may be a preferred metabolic substrate during rejection, which supports a humoral mechanism for substrate preference during transplant rejection and a potential diagnostic role for PET.

Daly [24] evaluated N-labeled ammonia ([N] NH3) reflecting myocardial perfusion and ¹⁸F-labeled fluorodeoxyglucose ([¹⁸F]FDG) small

animal PET imaging in a well-established murine cardiac rejection model, and found that there was a significant increase in [F]FDG uptake in allografts from 14 d to 21 d, and [F]FDG uptake correlated with an increase in rejection grade within allografts between 14 d and 28 d after transplantation; whereas the uptake of [N]NH3 was significantly lower relative to the native heart in allografts with chronic vasculopathy compared to isograft controls on 28 d, suggesting that PET imaging with [F]FDG can be used after transplantation to monitor the evolution of rejection, and decreased uptake of [N]NH3 in rejecting allografts may be reflective of decreased myocardial blood flow. These data suggest that combined [F]FDG and [N]NH3 PET imaging could contribute to unravel pathophysiological mechanisms underlying allograft rejection as a noninvasive, quantitative technique, and has potential application for serial monitoring of allograft rejection in human transplant recipients.

PET imaging to monitor the cerebral glucose metabolic change after cardiac ischemia/reperfusion

There is a growing concern about heart-brain neural crosstalk. Understanding neural mechanisms could lead to a better comprehension of cerebral circuit structure and function after cardiac ischemia/reperfusion injury. We used PET imaging to monitor the cerebral glucose metabolic changes after cardiac ischemia/reperfusion (**Figure 1**). Surgical procedures of myocardial ischemia-reperfusion injury models were performed following previously described methods [32-35]. After reperfusion, approximate $500 \pm 50 \ \mu$ Ci 18-fluoro-6-deoxy-glucose (¹⁸F-FDG) was injected via the tail vein. After 1 h of ¹⁸F-FDG uptake, rats were anesthetized with 2% isoflurane. Images were obtained with the

Cardiac research by PET



Figure 1. Alternations of glucose metabolism by small animal PET scanning. Representative images of ¹⁸F-FDG accumulation in the rats' brains of two groups (Control group and Model group). The images were displayed in three planes: coronal, horizontal, and sagittal planes.

whole body scanning pattern (5 min per scanning bed) by the Trans-PET BioCaliburn 700 system (Raycan Technology Co., Ltd, Suzhou, China). The PET images were reconstructed using the three-dimensional (3D) OSEM method with a voxel size of $0.5 \times 0.5 \times 0.5$ mm³. A volume-of-interest (VOI) analysis was conducted using the AMIDE software package (The Free Software Foundation Inc., Boston, Massachusetts, USA).

PET imaging in the assessment of sympathetic re-innervation after heart transplantation

Some reports show that structural sympathetic re-innervation of the transplanted heart can develop after cardiac transplantation [36-39], but the evidence can be difficult to diagnose. Schwaiblmair [40] investigated the influence of sympathetic re-innervation on cardiopulmonary exercise testing after orthotopic heart transplantation in 35 patients underwent PET, and found that sympathetic re-innervation enabled an increased peak oxygen uptake, suggesting that partial sympathetic reinnervation after cardiac transplantation is of functional significance. Schwaiger [41] studied possible

re-innervation of the human transplant after cardiac transplant by PET imaging approach in combination with catecholamine analogue [11C] hydroxyephedrine ([11C]HED), and found that there is the presence of sympathetic neuronal tissue in the terminals of transplanted human heart, which may reflect local sympathetic re-innervation. Bengel [42] described individual growth of sympathetic terminals late after cardiac transplantation by a longitudinal quantitative assessment, and found that sympathetic re-innervation was happened in the basal anterior region, apex, septal, and lateral wall, whereas inferior wall remained denervated; the largest reinnervated area surveyed in an individuum was 66% of the left ventricle, suggesting that re-innervation remained regionally heterogeneous.

PET imaging to predict recovery of global cardiac function

The past decade has seen strong progress in understanding PET imaging accuracy for predicting recovery of cardiac function after revascularization [43-45]. Gerber [25] assessed the accuracy of PET to predict recovery of global

cardiac function after revascularization in 157 male patients with coronary artery disease, and found that the highest sensitivity (79%) and specificity (55%) predicted postoperative ejection fraction improvement by using ¹⁸F-FDG PET, suggesting that FDG positron emission tomography can predict improvement of cardiac function coronary patients with impaired ejection fraction. Srivatsava [17] prospectively studied 120 patients with left ventricular (LV) dysfunction who underwent 99mTechnetium-Sestamibi myocardial perfusion SPECT-CT and 18FFDG cardiac PET-CT, and indicated that the change in LV impaired ejection after surgical management was statistically significant compared to medical management, and the assessment of myocardial viability was performed in patients who present after 12 h of acute myocardial infarction or with LV dysfunction due to ischemic heart disease to decide upon appropriate surgical management, suggest that there is an important role of PET-CT in assessment of myocardial viability in patients with LV dysfunction.

PET imaging to evaluate the cardiovascular effects of drugs and stimulation

Within the last decade, PET imaging has translated from a mere research tool to the cardiovascular efficacy of drugs by myocardial perfusion imaging and flow quantification. Molecular imaging tools including PET are increasingly applied in the drug development process [46].

Ueno [21] imaged the effects of angiotensinconverting enzyme inhibitor (5 mg/kg enalapril) in mice with heart allografts, and found that enalapril significantly decreased macrophagesavid nanoparticle signal by using sensitive positron emission tomography-computed tomography (PET-CT) imaging, and reduced a number of myeloid cells in the graft, blood, and lymph nodes by histology and flow cytometry, suggesting that angiotensin-converting enzyme inhibitor significantly prolong allograft survival.

Spinal cord stimulation causes significant symptomatic improvement in many patients with refractory angina pectoris [37, 47-49], and the mechanism underling this beneficial response is not fully known [50-53]. Hautvast [54] assessed the effect of spinal cord stimulation on myocardial blood flow by positron emission tomography in patients with refractory angina

pectoris, and found that after 6 weeks of stimulation, both frequency of daily anginal attacks and nitrogen consumption decreased, and the coefficient of variation of flow, representing flow heterogeneity, decreased after treatment, both at rest and after dipyridamole stress, suggesting that spinal cord stimulation is clinically effective due to homogenization of myocardial blood flow. Posma et al. [55] also reported a redistribution of myocardial flow during dual chamber pacing in a patient with non-obstructive hypertrophic cardiomyopathy by positron emission tomography, suggesting that early septal activation reduced septal fibre strain and blood flow and increased septal perfusion reserve.

Acknowledgements

This work was supported in part by grants from the National Natural Science Foundation of China (No. 81670240, 81770283, 81072152) and the Clinical Medical Research Center of Peritoneal Cancer of Wuhan (No. 201506091-1020462) and the Natural Science Foundation of Hubei Province (No. 2015CFA027), the Research Foundation of Health and Family Planning Commission of Hubei Province (No. WJ2015MA010, WJ2017M249) and Medical innovation project in Fujian Province (No. 2017-CX-48).

Disclosure of conflict of interest

None.

Address correspondence to: Dr. Mao-Hui Feng, Department of Gastrointestinal Surgery, Zhongnan Hospital, Wuhan University, No. 169 Donghu Road, Wuhan 430071, Hubei, PR China. E-mail: fengmh5690@163.com; Dr. Hong-Bing Xiang, Department of Anesthesiology and Pain Medicine, Tongji Hospital, Tongji Medical College, Huazhong University of Science and Technology, Wuhan 430030, Hubei, PR China. E-mail: xhbtj2004@163. com

References

- [1] Li R, Wong GT, Wong TM, Zhang Y, Xia Z and Irwin MG. Intrathecal morphine preconditioning induces cardioprotection via activation of delta, kappa, and mu opioid receptors in rats. Anesth Analg 2009; 108: 23-29.
- [2] Wong GT, Yao L, Xia Z and Irwin MG. Intrathecal morphine remotely preconditions the heart via

a neural pathway. J Cardiovasc Pharmacol 2012; 60: 172-178.

- [3] Lu Y, Hu J, Zhang Y, Dong CS and Wong GT. Remote intrathecal morphine preconditioning confers cardioprotection via spinal cord nitric oxide/cyclic guanosine monophosphate/protein kinase G pathway. J Surg Res 2015; 193: 43-51.
- [4] Xu LJ, Liu TT, He ZG, Hong QX and Xiang HB. Hypothesis: CeM-RVLM circuits may be implicated in sudden unexpected death in epilepsy by melanocortinergic-sympathetic signaling. Epilepsy Behav 2015; 45: 124-127.
- [5] Foreman RD, Garrett KM and Blair RW. Mechanisms of cardiac pain. Compr Physiol 2015; 5: 929-960.
- [6] Santos SF, Rebelo S, Derkach VA and Safronov BV. Excitatory interneurons dominate sensory processing in the spinal substantia gelatinosa of rat. J Physiol 2007; 581: 241-254.
- [7] Franco-Cereceda A, Kallner G and Lundberg JM. Capsazepine-sensitive release of calcitonin gene-related peptide from C-fibre afferents in the guinea-pig heart by low pH and lactic acid. Eur J Pharmacol 1993; 238: 311-316.
- [8] Steagall RJ, Sipe AL, Williams CA, Joyner WL and Singh K. Substance P release in response to cardiac ischemia from rat thoracic spinal dorsal horn is mediated by TRPV1. Neuroscience 2012; 214: 106-119.
- [9] Pirich C and Schwaiger M. The clinical role of positron emission tomography in management of the cardiac patient. Rev Port Cardiol 2000; 19 Suppl 1: I89-100.
- [10] Elsinga PH, Doze P, van Waarde A, Pieterman RM, Blanksma PK, Willemsen AT and Vaalburg W. Imaging of beta-adrenoceptors in the human thorax using (S)-[(11)C]CGP12388 and positron emission tomography. Eur J Pharmacol 2001; 433: 173-176.
- [11] Meeder JG, Peels HO, Blanksma PK, Tan ES, Pruim J, van der Wall EE, Vaalburg W and Lie KI. Comparison between positron emission tomography myocardial perfusion imaging and intracoronary Doppler flow velocity measurements at rest and during cold pressor testing in angiographically normal coronary arteries in patients with one-vessel coronary artery disease. Am J Cardiol 1996; 78: 526-531.
- [12] Meeder JG, Blanksma PK, van der Wall EE, Willemsen AT, Pruim J, Anthonio RL, de Jong RM, Vaalburg W and Lie KI. Coronary vasomotion in patients with syndrome X: evaluation with positron emission tomography and parametric myocardial perfusion imaging. Eur J Nucl Med 1997; 24: 530-537.
- [13] Meeder JG, Blanksma PK, van der Wall EE, Anthonio RL, Willemsen AT, Pruim J, Vaalburg W and Lie KI. Long-term cigarette smoking is as-

sociated with increased myocardial perfusion heterogeneity assessed by positron emission tomography. Eur J Nucl Med 1996; 23: 1442-1447.

- [14] Schindler TH. Positron-emitting myocardial blood flow tracers and clinical potential. Prog Cardiovasc Dis 2015; 57: 588-606.
- [15] Bergstrom M, Awad R, Estrada S, Malman J, Lu L, Lendvai G, Bergstrom-Pettermann E and Langstrom B. Autoradiography with positron emitting isotopes in positron emission tomography tracer discovery. Mol Imaging Biol 2003; 5: 390-396.
- [16] Hoff SJ, Stewart JR, Frist WH, Kessler RM, Sandler MP, Atkinson JB, Votaw J, Carey JA, Ansari MS and Merrill WH. Noninvasive detection of heart transplant rejection with positron emission scintigraphy. Ann Thorac Surg 1992; 53: 572-577.
- [17] Srivatsava MK, Indirani M, Sathyamurthy I, Sengottuvelu G, Jain AS and Shelley S. Role of PET-CT in the assessment of myocardial viability in patients with left ventricular dysfunction. Indian Heart J 2016; 68: 693-699.
- [18] Reddan MC and Wager TD. Modeling pain using fMRI: from regions to biomarkers. Neurosci Bull 2018; 34: 208-215.
- [19] Christen T, Nahrendorf M, Wildgruber M, Swirski FK, Aikawa E, Waterman P, Shimizu K, Weissleder R and Libby P. Molecular imaging of innate immune cell function in transplant rejection. Circulation 2009; 119: 1925-1932.
- [20] Ye Q, Wu YL, Foley LM, Hitchens TK, Eytan DF, Shirwan H and Ho C. Longitudinal tracking of recipient macrophages in a rat chronic cardiac allograft rejection model with noninvasive magnetic resonance imaging using micrometer-sized paramagnetic iron oxide particles. Circulation 2008; 118: 149-156.
- [21] Ueno T, Dutta P, Keliher E, Leuschner F, Majmudar M, Marinelli B, Iwamoto Y, Figueiredo JL, Christen T, Swirski FK, Libby P, Weissleder R and Nahrendorf M. Nanoparticle PET-CT detects rejection and immunomodulation in cardiac allografts. Circ Cardiovasc Imaging 2013; 6: 568-573.
- [22] Lee WW. Recent advances in nuclear cardiology. Nucl Med Mol Imaging 2016; 50: 196-206.
- [23] Blanksma PK and Pruim J. Positron emission tomography: measurement of myocardial perfusion using (13)N-labeled ammonia and (15) O-labeled water. Methods 2002; 27: 226-227.
- [24] Daly KP, Dearling JL, Seto T, Dunning P, Fahey F, Packard AB and Briscoe DM. Use of [18F] FDG positron emission tomography to monitor the development of cardiac allograft rejection. Transplantation 2015; 99: e132-139.
- [25] Gerber BL, Ordoubadi FF, Wijns W, Vanoverschelde JL, Knuuti MJ, Janier M, Melon P,

Blanksma PK, Bol A, Bax JJ, Melin JA and Camici PG. Positron emission tomography using(18) F-fluoro-deoxyglucose and euglycaemic hyperinsulinaemic glucose clamp: optimal criteria for the prediction of recovery of post-ischaemic left ventricular dysfunction. Results from the European community concerted action multicenter study on use of(18)F-fluoro-deoxyglucose positron emission tomography for the detection of myocardial viability. Eur Heart J 2001; 22: 1691-1701.

- [26] Siebelink HM, Blanksma PK, Crijns HJ, Bax JJ, van Boven AJ, Kingma T, Piers DA, Pruim J, Jager PL, Vaalburg W and van der Wall EE. No difference in cardiac event-free survival between positron emission tomography-guided and single-photon emission computed tomography-guided patient management: a prospective, randomized comparison of patients with suspicion of jeopardized myocardium. J Am Coll Cardiol 2001; 37: 81-88.
- [27] de Jong RM, Blanksma PK, van Waarde A and van Veldhuisen DJ. Measurement of myocardial beta-adrenoceptor density in clinical studies: a role for positron emission tomography? Eur J Nucl Med Mol Imaging 2002; 29: 88-97.
- [28] De Boer J, Slart RH, Blanksma PK, Willemsen AT, Jager PL, Paans AM, Vaalburg W and Piers DA. Comparison of 99mTc-sestamibi-18F-fluorodeoxyglucose dual isotope simultaneous acquisition and rest-stress 99mTc-sestamibi single photon emission computed tomography for the assessment of myocardial viability. Nucl Med Commun 2003; 24: 251-257.
- [29] Kashiyama N, Miyagawa S, Fukushima S, Kawamura T, Kawamura A, Yoshida S, Harada A, Watabe T, Kanai Y, Toda K, Hatazawa J and Sawa Y. Development of PET imaging to visualize activated macrophages accumulated in the transplanted iPSc-derived cardiac myocytes of allogeneic origin for detecting the immune rejection of allogeneic cell transplants in mice. PLoS One 2016; 11: e0165748.
- [30] Wareham NE, Lundgren JD, Da Cunha-Bang C, Gustafsson F, Iversen M, Johannesen HH, Kjaer A, Rasmussen A, Sengelov H, Sorensen SS and Fischer BM. The clinical utility of FDG PET/CT among solid organ transplant recipients suspected of malignancy or infection. Eur J Nucl Med Mol Imaging 2017; 44: 421-431.
- [31] Chen Y, Zhang L, Liu J, Zhang P, Chen X and Xie M. Molecular imaging of acute cardiac transplant rejection: animal experiments and prospects. Transplantation 2017; 101: 1977-1986.
- [32] Murry CE, Jennings RB and Reimer KA. Preconditioning with ischemia: a delay of lethal cell injury in ischemic myocardium. Circulation 1986; 74: 1124-1136.
- [33] Huang CH, Lai CC, Yang AH and Chiang SC. Myocardial preconditioning reduces kidney in-

jury and apoptosis induced by myocardial ischaemia and reperfusion. Eur J Cardiothorac Surg 2015; 48: 382-391.

- [34] Li ZX, Lin Q, He ZG, Wang Q, Chen YL, Feng MH, Li SY and Xiang HB. Altered myocardial gene expression profiling in the ischemic tissues at different time points after cardiac ischemia/ reperfusion in rats. Oncotarget 2018; 9.
- [35] Wang Q, Li ZX, Liu BW, He ZG, Liu C, Chen M, Liu SG, Wu WZ and Xiang HB. Altered expression of differential gene and IncRNA in the lower thoracic spinal cord on different time courses of experimental obstructive jaundice model accompanied with altered peripheral nociception in rats. Oncotarget 2017; 8: 106098-106112.
- [36] Buendia-Fuentes F, Almenar L, Ruiz C, Vercher JL, Sanchez-Lazaro I, Martinez-Dolz L, Navarro J, Bello P and Salvador A. Sympathetic reinnervation 1 year after heart transplantation, assessed using iodine-123 metaiodobenzylguanidine imaging. Transplant Proc 2011; 43: 2247-2248.
- [37] Singh H, Merry AF, Ruygrok P and Ruttley A. Treatment of recurrent chest pain in a heart transplant recipient using spinal cord stimulation. Anaesth Intensive Care 2008; 36: 242-244.
- [38] De Marco T, Dae M, Yuen-Green MS, Kumar S, Sudhir K, Keith F, Amidon TM, Rifkin C, Klinski C, Lau D, et al. Iodine-123 metaiodobenzylguanidine scintigraphic assessment of the transplanted human heart: evidence for late reinnervation. J Am Coll Cardiol 1995; 25: 927-931.
- [39] Kaye DM, Esler M, Kingwell B, McPherson G, Esmore D and Jennings G. Functional and neurochemical evidence for partial cardiac sympathetic reinnervation after cardiac transplantation in humans. Circulation 1993; 88: 1110-1118.
- [40] Schwaiblmair M, von Scheidt W, Uberfuhr P, Ziegler S, Schwaiger M, Reichart B and Vogelmeier C. Functional significance of cardiac reinnervation in heart transplant recipients. J Heart Lung Transplant 1999; 18: 838-845.
- [41] Schwaiger M, Hutchins GD, Kalff V, Rosenspire K, Haka MS, Mallette S, Deeb GM, Abrams GD and Wieland D. Evidence for regional catecholamine uptake and storage sites in the transplanted human heart by positron emission tomography. J Clin Invest 1991; 87: 1681-1690.
- [42] Bengel FM, Ueberfuhr P, Ziegler SI, Nekolla S, Reichart B and Schwaiger M. Serial assessment of sympathetic reinnervation after orthotopic heart transplantation. A longitudinal study using PET and C-11 hydroxyephedrine. Circulation 1999; 99: 1866-1871.
- [43] Raja S, Mittal BR, Santhosh S, Bhattacharya A and Rohit MK. Comparison of LVEF assessed

by 2D echocardiography, gated blood pool SPECT, 99mTc tetrofosmin gated SPECT, and 18F-FDG gated PET with ERNV in patients with CAD and severe LV dysfunction. Nucl Med Commun 2014; 35: 1156-1161.

- [44] Uebleis C, Hellweger S, Laubender RP, Becker A, Sohn HY, Lehner S, Haug A, Bartenstein P, Cumming P, Van Kriekinge SD, Slomka PJ and Hacker M. The amount of dysfunctional but viable myocardium predicts long-term survival in patients with ischemic cardiomyopathy and left ventricular dysfunction. Int J Cardiovasc Imaging 2013; 29: 1645-1653.
- [45] Raja S, Singh B, Rohit MK, Manohar K, Kashyap R, Bhattacharya A and Mittal BR. Comparison of nitrate augmented Tc-99m tetrofosmin gated SPECT imaging with FDG PET imaging for the assessment of myocardial viability in patients with severe left ventricular dysfunction. J Nucl Cardiol 2012; 19: 1176-1181.
- [46] Fernandes E, Barbosa Z, Clemente G, Alves F and Abrunhosa AJ. Positron emitting tracers in pre-clinical drug development. Curr Radiopharm 2012; 5: 90-98.
- [47] Lopshire JC, Zhou X, Dusa C, Ueyama T, Rosenberger J, Courtney N, Ujhelyi M, Mullen T, Das M and Zipes DP. Spinal cord stimulation improves ventricular function and reduces ventricular arrhythmias in a canine postinfarction heart failure model. Circulation 2009; 120: 286-294.
- [48] Issa ZF, Zhou X, Ujhelyi MR, Rosenberger J, Bhakta D, Groh WJ, Miller JM and Zipes DP. Thoracic spinal cord stimulation reduces the risk of ischemic ventricular arrhythmias in a postinfarction heart failure canine model. Circulation 2005; 111: 3217-3220.
- [49] Mannheimer C, Eliasson T, Augustinsson LE, Blomstrand C, Emanuelsson H, Larsson S, Norrsell H and Hjalmarsson A. Electrical stimulation versus coronary artery bypass surgery in severe angina pectoris: the ESBY study. Circulation 1998; 97: 1157-1163.

- [50] Ding X, Ardell JL, Hua F, McAuley RJ, Sutherly K, Daniel JJ and Williams CA. Modulation of cardiac ischemia-sensitive afferent neuron signaling by preemptive C2 spinal cord stimulation: effect on substance P release from rat spinal cord. Am J Physiol Regul Integr Comp Physiol 2008; 294: R93-101.
- [51] Hua F, Ardell JL and Williams CA. Left vagal stimulation induces dynorphin release and suppresses substance P release from the rat thoracic spinal cord during cardiac ischemia. Am J Physiol Regul Integr Comp Physiol 2004; 287: R1468-1477.
- [52] Gibbons DD, Southerland EM, Hoover DB, Beaumont E, Armour JA and Ardell JL. Neuromodulation targets intrinsic cardiac neurons to attenuate neuronally mediated atrial arrhythmias. Am J Physiol Regul Integr Comp Physiol 2012; 302: R357-364.
- [53] Ardell JL, Cardinal R, Beaumont E, Vermeulen M, Smith FM and Andrew Armour J. Chronic spinal cord stimulation modifies intrinsic cardiac synaptic efficacy in the suppression of atrial fibrillation. Auton Neurosci 2014; 186: 38-44.
- [54] Hautvast RW, Blanksma PK, DeJongste MJ, Pruim J, van der Wall EE, Vaalburg W and Lie KI. Effect of spinal cord stimulation on myocardial blood flow assessed by positron emission tomography in patients with refractory angina pectoris. Am J Cardiol 1996; 77: 462-467.
- [55] Posma JL, Blanksma PK and van der Wall EE. Redistribution of myocardial perfusion during permanent dual chamber pacing in symptomatic non-obstructive hypertrophic cardiomyopathy: a quantitative positron emission tomography study. Heart 1996; 75: 522-524.