Review Article Role of prostate stem cells and treatment strategies in benign prostate hyperplasia

Kalyan J Gangavarapu¹, Peter F Jowdy^{2,3}, Barbara A Foster¹, Wendy J Huss^{2,4}

¹Department of Pharmacology and Therapeutics, Roswell Park Comprehensive Cancer Center, Buffalo, NY 14263, USA; ²Department of Dermatology, Roswell Park Comprehensive Cancer Center, Buffalo, NY 14263, USA; ³Jacobs School of Medicine and Biomedical Sciences, University at Buffalo, Buffalo, NY 14203, USA; ⁴Department of Cell Stress Biology, Roswell Park Comprehensive Cancer Center, Buffalo, NY 14263, USA

Received October 27, 2021; Accepted April 25, 2022; Epub June 15, 2022; Published June 30, 2022

Abstract: Benign prostate hyperplasia (BPH) is a progressive disease with a direct correlation between incidence and age. Since the treatment and management of BPH involve harmful side effects and decreased quality of life for the patient, the primary focus of research should be to find better and longer-lasting therapeutic options. The mechanisms regulating prostate stem cells in development can be exploited to decrease prostate growth. BPH is defined as the overgrowth of the prostate, and BPH is often diagnosed when lower urinary tract symptoms (LUTS) of urine storage or voiding symptoms cause patients to seek treatment. While multiple factors are involved in the hyperplastic growth of the stromal and epithelial compartments of the prostate, the clonal proliferation of stem cells is considered one of the main reasons for BPH initiation and regrowth of the prostate after therapies for BPH fail. Several theories explain possible reasons for the involvement of stem cells in the development, progression, and pathogenesis of BPH. The aim of the current review is to discuss current literature on the fundamentals of prostate development and the role of stem cells in BPH. This review examines the rationale for the hypothesis that unregulated stem cell properties can lead to BPH and therapeutic targeting of stem cells may reduce treatment-related side effects and prevent the regrowth of the prostate.

Keywords: Prostate, BPH, stem cells, androgen

Introduction

Prevention strategies for BPH need further investigation. The current treatment strategies to reduce BPH symptoms are falling short, given the increasing incidence of BPH as men age and the current rising aging population. Strategies to pharmacologically reduce BPH symptoms are treatment with α-adrenergic antagonists, that relax smooth muscles and improve urine flow, either alone or in combination with 5- α reductase types 1 and 2 inhibitors, that block the conversion of T to DHT and reduce prostate size [1-5]. If BPH symptoms do not improve with pharmacologic treatment, then surgical options are explored, including transurethral resection of the prostate (TURP), transurethral incision of the prostate (TUIP), prostatic urethral lift, aquablation, prostatic stenting, and prostatic artery embolization [6, 7]. Discontinuation of pharmaceutical treatment in BPH can result in regrowth of the prostate and return of symptoms in both humans and rodents [8, 9], suggesting there is a component of the disease that is never targeted or is not sufficiently treated. We propose that the role of prostate stem cells in BPH is similar their role in prostate development and therapeutic strategies that target prostate stem cells would provide a needed prevention strategy.

Targeting prostate stem cells alone is unlikely to be curative, in our opinion, but when used in combination with current strategies could enhance therapeutic response and reduce the incidence of regrowth when treatment is stopped. Specificity of targeting prostate stem cells, without targeting all stem cells remains a barrior, in our opinion. There are several ways to target stem cells, but two key mechanisms are: inhibition of key stem cell signaling pathways

and direct targeting of stem cells via surface markers. Signaling pathways involved in stem cell biology include hedgehog, phosphoinositide 3-kinase (PI3K)-AKT, and Wnt [10]. Surface markers can be used for targeted delivery of therapeutic agents to stem cells. In recent studies antibodies against stem cell surface markers are currently being used to deliver nanoparticles with cytotoxic agents specifically to stem cells expressing CD44 and CD133 [11]. In addition to targeting stem cell signaling pathways and directing therapy with stem cell surface markers, several naturopathic medicinal therapies are thought to target prostate stems cells. Qianliening capsule, saw palmetto, Pygeum Africanum, and Hypoxis rooperi which have been used to target prostate stem cells in BPH with few side effects, may provide an opportunity for early prevention strategies [12, 13]. Even though BPH is a very prevalent disease in older men, not much is known about the biological factors involved in its pathology. In our opinion, given the high incidence of BPH in the aging population, treatment strategies should be developed based on the role of stem cells in prostate development to prevent BPH with aging. In this article the role of prostate stem cells in the etiology and treatment options of BPH in relationship to prostate development are reviewed.

Etiology of BPH

BPH is a common disease in elderly men and is the most frequent disease treated by urologists. While BPH has not been associated with progression to prostatic carcinoma [14-17], several key regulatory functions are shared between BPH and prostate cancer.

The frequency and incidence of benign nodular hyperplasia increase with age and have a higher prevalence in men over 50 years of age [18-21]. BPH is often accompanied by LUTS, which, if left untreated, can eventually lead to acute urinary retention that necessitates surgical intervention [21-24]. BPH is caused primarily by hyperplasia of the prostatic stromal and epithelial cells paired with a decrease in apoptosis of prostatic epithelial cells. BPH arises in the periurethral region of the transitional zone of the human prostate, whereas prostate cancer mainly arises in the peripheral zone of the prostate [25-30].

Although the symptoms of BPH cannot be simplified as just urethral resistance, a primary symptom caused by BPH is urinary outlet obstruction. Urinary outlet obstruction, also known as bladder outlet obstruction (BOO), can cause changes in the anatomical features of the prostate gland. Specifically, this occurs as changes in the transitional zone and not in the overall volume of the prostate [29, 30]. The volume of the periurethral region may correlate with the degree of outlet obstruction. Assessment of BPH is based upon clinical history, digital rectal examination (DRE), and urodynamic studies that evaluate urinary flow patterns [31]. Several problematic symptoms of BOO include: poor flow of urine, terminal dribbling, straining, hesitancy, nocturia, and increased frequency and urgency of urination [31, 32]. In addition to DRE, the size of the prostate can also be estimated by a perirectal ultrasound, which is critical in both determining treatment modalities for BPH and evaluating potential underlying prostate cancer. If left untreated, BPH can eventually lead to acute urinary retention and chronic kidney problems [33-35].

Even though BPH is a very prevalent disease in men, not much is known about the biological factors involved in its pathology. Several theories in the scientific community attempt to explain the possible etiology and pathology of BPH. A primary concept suggests that more dihydrotestosterone (DHT) is produced in the aging prostate, which then leads to enlargement [33, 36-38]. However, Walsh et al. demonstrated that the amount of DHT in the hyperplastic prostate is not different from that seen in the normal prostate [39]; suggesting that the presence of a critical amount of DHT is not the only requirement for developing BPH. Several studies have suggested that the amount of DHT present in the prostate is critical in developing BPH by promoting stromal-epithelial interactions and by increasing the number of prostate cells due to stem cell proliferation [16, 33]. Prostate stem cells are thought to undergo clonal expansion and produce transit-amplifying cells which give rise to terminally differentiated luminal cells that are then programmed to undergo apoptosis. In BPH, this process is unbalanced resulting in an enlargement of the prostate. Thus, targeting of prostate stem cells to manage BPH at the source of the increased number of cells could be an effective therapeutic strategy.

In BPH, some cells in the basal compartment have stem cell features, including genomic protection and suppression of cell death. Even though genomic mutations are rare in BPH, epithelial and stromal cell proliferation is increased several-fold in BPH [40]. The stromal to epithelial cell ratio can increase 5-fold in BPH (10:1) compared to normal prostate (2:1), resulting in common BPH symptoms [41]. The interaction between the epithelial compartment and the stroma increases signaling factors that promote epithelial expansion. Signaling between the stromal and epithelial cells is critical in prostatic development, as demonstrated by Cuhna et al. [42]. The theory of stromal and epithelial cell interaction is postulated as "embryonic reawakening" whereby stromal cells secrete growth factors that stimulate adjacent epithelial cell proliferation and vice versa [16] and will be further discussed in section 3b) Prostate Stem Cells.

Aging and androgens

Aging and hormone variations are two main etiological factors of BPH. Evidence suggests that BPH is a hormonally regulated disease. In studies by Bianch-Frias et al., aging was identified as another risk factor for BPH that may affect the microenvironment and promote pathology in the human prostate. The authors observed that with aging, the prostate stroma shows increased smooth muscle disorientation and decreased expression of collagen-related genes, further suggesting extracellular matrix disorientation [43]. Furthermore, Bianch-Frias et al. observed an increase in the infiltration of inflammatory cells into the inter-glandular space of the aging stroma; with an increase in B-cells, T-cells, macrophages, the chemokine (C-C motif) ligand 8 (CCL8), and the stress response protein, apolipoprotein D (ApoD) [43]. Several studies have suggested a delicate balance of androgens and growth factors within the microenvironment that regulates autocrine/ paracrine signaling is critical for maintenance of normal cellular proliferation and tissue homeostasis [43, 44]. Alterations in the cytoskeletal framework can lead to prostate pathologies, including BPH and prostate cancer [43, 45]. Cytokines, growth factors, and matrix components can be considered critical in maintaining the prostate microenvironment and are potential targets for treating BPH.

Prostatic growth is dependent on androgens. Men castrated before puberty do not develop BPH in their later age [33, 46, 47]. Geller further described how high levels of DHT, an active form of testosterone (T), plays a central role in the development and pathogenesis of BPH, including: "embryonic re-awakening" as explained by McNeal [16], increasing NADPH, and increasing the estrogen-to-testosterone ratio in plasma [47]. Despite a low concentration of testicular androgens in aging men, there is increased activity of $5-\alpha$ reductase, an enzyme that metabolizes T to DHT, and elevated DHT levels promote the stromal and epithelial growth that leads to BPH [46]. $5-\alpha$ reductase deficiency, or inhibition of the enzyme, reduces conversion of testosterone to DHT and disrupts many androgen-controlled developmental processes [46, 48]. In adult men inhibition of 5- α reductase results in a decrease in the size of the prostate [46, 48]. The DHT levels and conversion from testosterone were increased in prostatic hypertrophy further supporting the role of DHT in BPH [49]. Later several studies measured elevated levels of DHT in BPH prostates when compared to normal prostates [46, 49-53]. Studies conducted by Walsh et al. refute the idea that low levels of DHT are simply an artifact in a normal post-mortem prostate [39]. Furthermore, separate studies conducted by Morfin [50] and Isaacs' groups [36] support the idea that elevated DHT in BPH prostatic tissue is related to an increase in 5-α reductase enzyme activity, thus demonstrating the effects of androgen-induced metabolic activities on normal and hyperplastic prostates. In addition to androgens, Devlin et al. review the roles of estrogens, insulin, and growth factors in prostate development and growth regulation associated with BPH [54] and are discussed below in section 3) Role of prostate compartments in development and BPH. Although the circulating levels of testicular androgens decrease with age, levels of DHT and AR signaling are high in the aging prostate. Studies show a role for androgens and AR in promoting BPH development [55, 56].

AR regulation of the prostate microenvironment includes the immune system and vascularization within the stroma. Lai and colleagues demonstrated that AR expression in both epithelial and stromal cells attracts infiltrating macrophages [57-60]. The eventual interac-

tions between macrophages and epithelial/ stromal cells lead to increased expression of transforming growth factor β -2 (TGF β -2) and chemokine CCL3 respectively [55, 56]. Androgens also regulate the vascular system, where epithelial cellular death during prostatic involution following castration was first described by Kerr et al. [61] and was confirmed as apoptosis [62], and this has been modeled in the human prostate [63]. The epithelial cell apoptosis induced by castration is actually preceded by a dramatic change in the stroma vasculature that may facilitate directly, or indirectly, the signaling of apoptosis of epithelial cells [64]. While dysregulation of androgen signaling is clearly a major contributing factor to BPH, the specific effects on and between multiple cellular compartments within the prostate still need elucidation, and the role of androgen signaling in the prostate stem cell compartment is discussed below.

Inflammation

Inflammation is a risk factor in BPH by many mechanisms in addition to androgen signaling. McLauren et al. proposed that inflammation and the presence of stem cells in the prostate are the mechanisms triggering the re-awakening of developmental growth in the prostate [65]. Inflammation of the prostate can lead to activation of interleukins-triggered growth pathways in the stroma and epithelium of the prostate [65]. Jerde et al. showed that interleukin-1 (IL-1) induces growth signaling in BPH via activation of insulin-like growth factor (IGF)dependent signaling during prostate development [66]. Along with IL-1, IL-6, IL-8, IL-12, other interleukins are involved in growth signaling during prostate development, and reactivation of growth signaling during inflammation results in regenerate of prostate tissue. Separate studies have emphasized the role of IGF-1 signaling in prostate development and growth [67-69]. Shah et al. reviewed molecular pathways to target in BPH, although targeting inflammation in BPH is not currently thought to be an appropriate therapeutic target [70]. Growth factors and interleukins secreted into the prostate stroma promote several proliferation pathways such as mitogen-activated protein kinase (MAPK), PI3K signaling pathways, and the activation of transcription factors that induce proliferation promoting genes leading to stroma proliferation [71]. Effector molecules, along with autocrine and paracrine growth factors, work together to cause increased stromal and epithelial cell proliferation, regulated by stem cell maintenance in our opinion, thus leading to the development and progression of BPH.

Abdominal obesity

Increased waist circumference correlates with BPH and is associated with increased inflammation. Abdominal obesity is defined as a waist circumference that is >102 cm (40 inches) and has been linked to the progression of BPH [72]. As the prevalence of a large waist circumference has increased, so has awareness of the associated adverse health factors [73]. A metaanalysis of cohort and case-control studies has identified a positive association with BMI and increased risk of BPH and LUTS [74]. While the mechanisms are not fully understood, Gotera et al. have been studying possible etiologies. They have concluded that waist circumference >102 cm indirectly increases the risk of developing BPH through both IL-6 and hyperinsulinemia secondary to insulin resistance [75]. IL-6, an acute phase reactant, is increased with inflammation, an established risk factor for BPH, and elevated IL-6 is common in abdominal obesity [65]. Thus, an increased waist circumference, which is associated with inflammation, promotes growth signaling pathways that further reinforce the epithelial/stromal interactions and potentially stem cell proliferation that are hypothesized to be a driving factor in developing BPH.

Role of prostate compartments in development and BPH

Stromal and epithelial paracrine signaling

Prostate development is initiated through paracrine interactions between the stromal and epithelial compartments. Epithelial and stromal cells show heterogeneous expression of AR [76, 77]. AR expression in the stroma is instrumental in regulating paracrine signaling to induce epithelial cell proliferation. In classic studies utilizing tissue recombination with androgen-insensitive AR^{tfm} and androgen-sensitive AR^{wt} prostate tissue, Cunha et al. demonstrated that a functional AR is not required in the epithelium but is required in the mesenchyme for androgen-induced growth and branching morphogenesis [42]. However, a functional AR in the epithelium is required for the differentiation of the epithelium into mature prostatic secretory cells [42]. Thus, AR is required for functional differentiation in the adult prostate, and during prostatic development AR regulates many stages including proliferation, branching morphogenesis, and maturation of secretory epithelial cells. Estrogens have long been associated with hyperplasia in rodent models [78] and BPH risks in men [79]. Stromal estrogen receptor (ER)- α and ER- β signaling are both required to regulate androgen signaling [80, 81]. Dr. Cunha thoroughly reviewed the androgen and estrogen-regulated epithelial/mesenchymal interactions [82], and Delvin et al. provide a robust review of the current understanding of the roles of androgen, estrogen, and growth factors in BPH initiation and progression [54].

AR signaling in stromal cells also promotes the secretion of growth factors that act on the epithelial compartment, the immune system, extracellular matrix remodeling, and neovascularization. Paracrine signaling from the prostate stromal compartment is a key regulator of epithelial stem cells during epithelial branching in prostate development [83, 84]. Recently, single-cell RNA sequencing of the developing prostate has further elucidated specific cell populations that express developmental regulators [85]. Regulation of prostate epithelial branching during development initially occurs through paracrine signaling of growth factors from the stroma. Conversely, paracrine signaling from the epithelium also triggers stromal stem cells to differentiate into stromal smooth muscle cells and fibroblasts [22, 86]. The stromal stem cell-derived smooth muscle cell progenv express AR protein and 5-α reductase enzyme [22]. In smooth muscle cells, DHT binding to the AR receptor triggers the expression of several growth and differentiation factors, including IGF-1 [57], fibroblast growth factor (FGF-7) [87], nerve growth factor-β (NGF-β) [42, 88], and vascular endothelial growth factor (VEGF) [89]. TGF β inhibits and rogen signaling in prostate stromal cells [89], and FGF-7 can induce epithelial branching independent of androgen signaling [90]. Additionally, prostate epithelial stem cells initiate prostate proliferation and epithelial branching by Nkx3.1 [91], Notch [81, 92, 93], Foxa1 [83, 94], Shh [95-97], and Wnt [83, 94] signaling. Thus, stromal and epithelial cell proliferation control is a multifactorial process regulated by several growth factors that interact at multiple levels including through the stem cell compartment further discussed below.

Prostate stem cells

In the prostate both the stromal and epithelial compartments contain stem cells, and adult prostate stem cells play a critical role in differentiation and maintenance. Expansion of prostate stem cells in both the epithelial and stromal compartments can lead to hyperplasia in the prostate gland; thus, the size of the prostate is dictated by both the stromal and epithelial compartments. As postulated by Coffey and Walsh, the increase in the size of an organ depends on the balance between cell proliferation and cell death [17, 33]. Studies conducted by Coffey et al. suggested that androgens regulate cellular proliferation and cell death in the prostate and that BPH results from an unregulated stem cell compartment [17, 22, 33]. For example, in the epithelial compartment, stem cells divide and give rise to a compartment of transit-amplifying cells that eventually terminally differentiate into mature luminal cells with an average lifespan of 500 days [98]. The terminally differentiated luminal cells have a finite lifespan and eventually undergo apoptosis. However, due to either a delay or a block in the maturation process as postulated by Isaacs and colleagues [22, 33], this may lead to an increased number of transit-amplifying cells, thus leading to an enlargement in the size of the gland. As a result, the size of the prostate may be defined by the presence of epithelial stem cells and transit-amplifying cells [33]. Historically, prostate epithelial stem cells were thought to reside in the basal laver and give rise to distinct types of progenies which have varying proliferation potentials. Transit-amplifying cells divide and give rise to intermediate cells [22, 33]. Intermediate cells respond to androgens and terminally differentiate into luminal cells [22, 23]. There is evidence of multiple types of epithelial stem cells, including a common basal and luminal stem cell and only luminal stem cells that have been proposed to be a source of recurrent prostate cancer [99-101]. Thus, the plasticity of the prostate cell compartments is a likely contributor to different cell types identified in BPH by single-cell RNA sequencing [102].

The stromal stem cells in the prostate have demonstrated mesenchymal stem cell properties. The stromal stem cells give rise to smooth muscle cells, fibroblasts, adipose, and myoadipose cells with mesenchymal stem cell markers [22, 86]. Additionally, prostate stroma derived from BPH specimens has multipotent properties upon treatment with differentiation agents, suggesting the presence of adult stromal stem cells in the bulk of the prostate stroma [85]. Alternatively, Brennen et al. demonstrated that BPH tissue was infiltrated with mesenchymal stem cells [103]. While the presence of stem cells is hypothesized to be one of the factors leading to BPH, the factors that stimulate differentiation into mature stromal and epithelial cells are not fully understood, particularly as associated with BPH.

Androgen regulation of the prostate stem cell compartment

The response of epithelial prostate stem cells to androgen levels was shown in seminal work conducted by English et al. and others in the late 1980s. English et al. showed that the prostate gland can undergo regression upon castration and re-growth with subsequent re-administration of testosterone [9]. This regression and re-growth suggest that stem cells not only survive androgen deprivation but also have the capability to respond to androgen stimulation and regenerate an entire prostate gland [9, 104, 105]. Thus, prostate stem cells that survive androgen deprivation therapy (ADT) can regulate prostate size through the uncontrolled generation of stromal and epithelial cells.

ADT interrupts stromal-epithelial interactions and interrupts cell expansion in BPH. Interruption of stromal-epithelial interactions decreases the level of several growth factors, hierarchical differentiation and the total number of stromal stem cells [22, 106]. There are prostate stem cells that survive ADT, and prolonged ADT exposure can conversely cause the stroma to induce proliferation of epithelial cells. ADT alone is not sufficient as a treatment for BPH since resistance mechanisms do not eliminate the stem cells, the cells responsible for prostate growth and stroma signaling. Therefore, understanding the basic mechanisms that trigger the proliferation and/or apoptosis of cells are critically needed to increase our understanding of factors that control the existence and number of stem cells in the prostate. While basal epithelial stem cells of the prostate respond to proliferative signaling and give rise to transit-amplifying/intermediate cells which terminally differentiate into luminal cells, distinct prostate luminal stem cells that bypass the traditional differentiation path were first described in 2014 [107, 108], and were later analyzed with single-cell RNA sequencing [109]. Defects in the maturation process of epithelial stem cells to luminal cells can cause an increase in the number of epithelial cells.

BPH initiation and progression depend on the fine balance between stromal and epithelial cell proliferation and apoptosis. An imbalance in the paracrine signaling that promote proliferation and/or induce apoptosis can lead to abnormal proliferation of stromal cells leading to BPH [110-113]. In BPH, "reawakening" of the embryonic growth, development, and differentiation occurs wherein the stromal compartment reacquires the properties of its embryonic anlagen. In development, the urogenital mesenchyme actively induces the development of the urogenital epithelium into the prostate in the presence of androgens [16, 103]. The proliferation and cell death mechanisms are tightly regulated in the developing prostate, but in BPH, these mechanisms are thought to be deregulated. Uncontrolled proliferation led to the continued proliferation and clonal expansion of stromal and epithelial cells. Additionally, androgen ablation may affect stromal cell function but not the actual number of stromal cells. In BPH nodules, stromal cells have high levels of proliferation and low levels of apoptosis. For example, in BPH the expression of the apoptotic inducer TGF-B1 is decreased, and the anti-apoptotic protein Bcl-2 is increased [110]. Zhang et al. observed a positive correlation between the density of stromal cells and proliferation with an inverse relationship between the density of epithelial cells and apoptosis [114]. Many additional mechanisms regulating epithelial and stromal proliferation and apoptosis have been associated with BPH, including estrogen and hedgehog signaling. Differential regulation of ER- α and ER- β on AR signaling has been shown to regulate prostate growth [81, 92, 93]. Stromal hedgehog signaling is also required for prostate growth and regeneration [95-97] and is currently under investigation in

BPH and in development as potential therapeutic target to treat BPH [115, 116].

We hypothesize that the total number of stem cells present in the prostate modulates prostatic growth and BPH development. A pool of stem cells is accumulated in the postnatal prostate, which is critical for the development of the prostate to its normal size. Critical physiological levels of androgens are also important for generating the pool of stem cells. The importance of these stem cells in the prostate is evidenced by experiments by Coffey and his colleagues. Boys castrated before puberty do not get BPH even if exposed to high levels of androgens later in life [33, 46, 47]. This suggests that the presence of a critical number of stem cells is important for abnormal development and that this number of prostate stem cells is reached only after puberty. There are also molecular factors that help maintain the pool of stem cells in the prostate. Targeting these molecular factors could help deplete the stem cell pool and reduce prostate size when combined with ADT. BPH patients may be given interventions that target prostate stem cells in combination with ADT to reduce prostate size. The pool of stem cells in the adult prostate is maintained through stromal-epithelial interactions, mediated by growth and survival factors regulated by androgens. Thus, in our opinion targeting prostate stem cells in combination with ADT or other therapeutic interventions like $5-\alpha$ reductase inhibitor to deplete the stem cell pool to a critical level may result in improved treatment of BPH.

Rationale for targeting stem cells to treat BPH

Given the decrease in androgen levels with age, a subsequent decrease in proliferation of prostatic epithelium and stroma would be expected. However, Bushman et al. observed that castration of adult mice led to the induction of cells expressing various stem cell markers, including CD44, CD133, Sca-1, and CD117 indicating that a decrease in androgens with age may be compensated by an increase in proliferation of stem/progenitor cells [65]. Crowell et al. demonstrated that Trop2+ luminal progenitor cells expand in the prostate of aged mice and men [100]. PSCA-expressing prostate luminal progenitor cells were increased in BPH tissue compared to normal prostate [100]. These studies suggest that prostate stem cells increase with age and in BPH and that castration, or ADT, alone may not be sufficient for effective BPH treatment. We propose that inhibiting mechanisms that promote proliferation of prostate stem cells, such as hedgehog signaling and Wnt signaling, may increase the efficacy of BPH treatment.

Current BPH treatment considerations and options

Once BPH is diagnosed, there are two different options for the patients with bothersome symptoms: (i) medicinal treatments are suggested for patients with symptoms that are bothersome and impede quality of life; and (ii) surgical treatment is usually suggested for patients with recurrent disease who can tolerate a surgical procedure [117, 118].

Current BPH pharmaceutical treatments include finasteride (a form of ADT), and surgical interventions, including TURP and TUIP [119, 120]. ADT and TURP/TUIP have quality of life impacting side effects such as impotence, decreased libido, and abnormal ejaculation. Finasteride or other pharmacologic forms of ADT are used continuously to keep the prostate size under control [119, 120]. Discontinuation of pharmaceutical treatment often results in regrowth of the prostate and return of symptoms, suggesting there is a component of the disease that is never targeted or is not sufficiently treated. Stromal-epithelial interactions and prostate stem cells are two components of the prostate we propose are resisting/surviving the effects of hormone ablation. Though ADT reduces the prostate size, the number of stem cells in the prostate is not decreased since most stem cells are not affected by ADT. Thus, the remaining stem cells regenerate the prostatic tissue when androgen levels return to normal [22]. In addition, prostate stem cells are resistant to apoptosis by α -adrenergic antagonists. Resistance to apoptosis prevents a reduction in prostate size and/or increases the chances of recurrence [121-123]. Therefore, we propose targeting the prostate epithelial and stromal stem cells is important to circumvent prostate regeneration. As suggested by Isaacs [9], ADT treatment followed by radiation therapy is one approach that could inhibit the stromal-epithelial interactions and thereby

inhibit signaling that triggers proliferation of the stem cell compartment. The enlarged prostate should be targeted in a way as to reduce stem cell numbers to a critically low level, without complete depletion of the stem cell pool. This critically low level of the stem cell pool will leave the remaining stem cells presumably unable to induce BPH development in the future, even if triggered by different autocrine and paracrine signaling factors. Thus, we propose that BPH treatment should focus on reducing prostate size, reducing the number of prostatic stem cells and relieving the associated clinical symptoms of lower urinary tract. Though different options are available for BPH treatment, some significantly impact the patient's quality of life. Several avenues that can be targeted pharmacologically to reduce BPH symptoms by reducing the size of the enlarged prostate include: i) targeting DHT synthesis; ii) targeting smooth muscle function; iii) surgical interventions; iv) targeting stromal-epithelial interactions; v) targeting stem cells.

i) Targeting DHT synthesis: Several clinical trials compromised of patients with moderate-tosevere clinical BPH symptoms demonstrated a reduction in BPH symptoms and prostate size by treatment with α -adrenergic antagonists in combination with either $5-\alpha$ reductase types 1 and 2 inhibitors [1-5]. α -Adrenergic antagonists are drugs that block α -adrenoceptors that function to constrict smooth muscles surrounding the urethra and bladder. Relaxation of smooth muscles with α -adrenergic antagonists improves urine flow. Examples of α -adrenergic blockers include tamsulosin (FlomaxÒ), alfuzosin, terazosin, and doxazosin. $5-\alpha$ reductase types 1 and 2 inhibitors inhibit the 5- α reductase enzymes, thereby blocking the conversion of T to DHT. 5- α reductase inhibitors, finasteride and dutasteride, further reduce prostate size, thus, improving the BPH symptoms. Clinical trials demonstrated that treating patients with BPH with α -adrenergic antagonists and/or $5-\alpha$ reductase inhibitors may have long-term control at reducing the prostate size and improving BPH symptoms. However, such treatments are not effective at completely eliminating the need for surgery or the risk of acute urinary retention. BPH is a clinically progressive disease. While pharmaceutical treatment alleviates symptoms of BPH, the chances of return of symptoms are high, suggesting targeting additional mechanisms of BPH disease would be more successful.

ii) Targeting smooth muscle function: a. αadrenergic antagonists are drugs that block α -adrenoceptors that function to constrict smooth muscles surrounding the urethra and bladder. Relaxation of smooth muscles with α-adrenergic antagonists improves urine flow and relieves BPH symptoms. Examples of α-adrenergic blockers include tamsulosin (FlomaxO), alfuzosin, terazosin, and doxazosin. b. Other medications that target smooth muscle activity in the urinary track include anticholinergic drugs that inhibit muscarinic receptors in the bladder urothelium which limit detrusor overactivity associated with BPH. While these medications can show some benefit, they need to be used with caution, especially in elderly patients, due to the numerous side effects, most notably confusion [6].

iii) Surgical interventions: In addition to TURP procedures, multiple surgical interventions have been developed to treat BPH, including endoscopic procedures such as a prostatic urethral lift. This procedure can alleviate adverse effects such as erectile and ejaculatory dysfunction but is less efficacious than a TURP [120]. Other surgical procedures with limited evidence of benefit include aquablation, prostatic stenting, and prostatic artery embolization. Each of these procedures is less invasive than TURP; however, long-term data and success rates have not been sufficiently measured [6].

iv) Targeting stromal-epithelial interactions: Stromal-epithelial interactions regulate prostate development and contribute to prostatic diseases. Animal and in vitro models provide evidence that targeting growth factor signaling pathways between stromal-epithelial compartments, such as hedgehog, notch, and estrogen receptor signaling, would decrease prostate growth and BPH symptoms. Growth factor signaling regulates stromal-epithelial interactions leading to stromal and epithelial cell proliferation [124]. For instance, stromal and epithelial cell proliferation and differentiation were observed to be dependent on Notch signaling [125, 126]. ER- α promotes proliferation, and ER-ß induces apoptosis in normal prostate epithelium and prostate cancer [92]. ER- α is overexpressed in BPH stroma, whereas in BPH

ER- β is minimally expressed in stromal cells and highly expressed in basal and luminal cells [92]. Furthermore, ER- α , not ER- β , increases urine retention in mice which can be relieved with treatment with the ER- α antagonist, raloxifene [127]. Additionally, in the prostate of aging men, the levels of prostatic testosterone decrease while the levels of estrogens increase [92, 128, 129]. A loss of ER-B expression and increased expression of ER- α is observed in BPH and prostate cancer [129]. Therefore, targeting the estrogen receptor pathway is another strategy to treat BPH. Paracrine signaling, such as PI3K/AKT/mTOR, hedgehog signaling, and IGF signaling also plays a role in regulating stromal-epithelial interactions. Deregulation of these pathways that mediate paracrine signaling can lead to BPH [10]. Studying the signaling pathways involved in stromal-epithelial proliferation and differentiation may identify new targets for more effective therapies for BPH.

v) Targeting stem cells: While most therapies focus on reducing prostate size and temporarily alleviating the patient's symptoms, an emphasis on targeting stem cells may help prevent regrowth of the prostate and return of BPH symptoms, in our opinion. BPH stem cells can be targeted by several mechanisms. In studies from our laboratory, prostate stem cells that have a functionally active ABCG2 transporter and aldehyde dehydrogenase 1A1 (ALDH1A1) can serially regenerate the prostate when combined with embryonic rat urogenital mesenchyme [130, 131]. Additionally, we showed ABCG2 regulates the maintenance of mouse and human prostate stem cells and can be pharmaceutically inhibited to decrease regeneration of the mouse prostate [132, 133]. Additionally, targeting stem cells can be accomplished by inhibiting signaling pathways such as hedgehog, PI3K/AKT and Wnt, which promote self-renewal and proliferation of prostate stem cells [10], and can be investigated as potential targets for therapeutic interventions. Recent studies take advantage of linking antibodies against cancer stem surface markers to nanoparticles to target cytotoxic agents specifically to cancer stem cells as described in a recent review [134]. Many of the surface markers used to target nanoparticles are found on prostate stem cells, including CD44 and CD133 [11]. Specifically, nanoparticles linked

with CD44 antibodies have successfully targeted therapy to prostate cancer stem cells [135]. We propose that nanoparticles linked to antibodies to prostate stem cell markers would target prostate stem cells in BPH and have therapeutic benefits.

Future directions

While there are several treatment options for BPH, most are not curative for the majority of patients and significantly decrease in quality of life of patients with BPH. Given the significant number of men affected by BPH, prevention and curative treatments should be the focus of future research. Targeting prostate stem cells provides an opportunity to develop curative treatments. While some studies have focused on preventing the growth of the prostate and reducing the symptoms of BPH, or finding therapies with fewer side effects; these studies have not focused on targeting prostate stem cells. More studies are needed to evaluate novel therapies in preclinical models with the goal of rapid movement toward clinical trials. Silymarin is a phytoestrogen compound that can prevent testosterone-induced BPH in rats [136]. In this study, silymarin is both pro-apoptotic and anti-oxidative to prevent BPH [136]. Better nutrition and exercise are also indicated in the prevention of BPH. Dietary supplements including zinc, *β*-sitosterol, saw palmetto, Pygeum Africannum, and Cernilton can help relieve symptoms of BPH and have fewer side effects. Additionally, studies have shown that intake of dietary supplements, such as isoflavones (genistein) or medicinal herbs (saw palmetto), significantly improve symptoms of BPH, including a decrease in LUTS [137]. Diets containing a high intake of vegetables, polyunsaturated fats, and moderate alcohol intake also decrease the risk of BPH. In contrast, diets associated with a high intake of starches and red meats increase the risk of BPH [138, 139]. More BPH prevention studies need to be performed to assess the role of diet in disrupting prostate stem cell regulation of prostate growth. Specifically, more studies are needed examining the impact of dietary supplements specifically on prostatic stem cells are need.

Naturopathic medicinal therapies, including qianliening capsule (a natural product and a Chinese formulation) and saw palmetto are

options that target prostate stem cells in BPH and have fewer side effects. Qianliening, in capsule form, acts through suppression of the EGFR/STAT3 signaling pathway leading to decreased expression of Bcl-2 resulting in decreased prostate epithelial proliferation [12]. Other natural products, including saw palmetto, Pygeum Africanum, and Hypoxis rooperi, have few side effects and are being used to successfully treat BPH [13]. Disulfiram is a naturopathic therapy that inhibits ALDH detoxification of alcohol and has been used as an anti-alcoholic drug for decades. Disulfiram directly targets stem cells with high ALDH activity and has been shown to specifically target cancer stem cells, recently reviewed [140]. The ALDH3A1 isoform is elevated in BPH, making disulfiram an exciting new treatment to test in BPH [141]. The role of ALDH in stem in BPH warrants further studies. Targeting prostate stem cells in BPH can help manage the disease and delay the growth of the prostate. Additionally, in our opinion, therapy strategies that target prostate stem cells are likely effective therapies in the prevention of BPH.

Conclusions

In the absence of symptoms that disrupt lifestyle, management of BPH by watchful waiting is an option for patients [117]. Current BPH therapeutic and surgical therapies reduce the prostate burden and alleviate the associated symptoms of BPH but do not reduce the risk of disease progression and return of BPH symptoms with the regrowth of the prostate. Furthermore, current pharmaceutical and surgical therapy options have harmful side effects that can greatly impact the patient's quality of life. In our opinion, there is a need for better therapeutic options to effectively prevent and treat BPH. To achieve better therapeutic outcomes, BPH treatment strategies should focus more on the root problem in the development, progression, and pathogenesis of BPH [9, 16, 21, 103] and focusing on stem cell biology in BPH provides an attractive therapeutic target. The vast knowledge of the prostatic stem cells in development and growth needs to be exploited in order to better manage the inevitable emergence of BPH in aging men.

Disclosure of conflict of interest

None.

Address correspondence to: Wendy J Huss, Departments of Dermatology and Cell Stress Biology, Roswell Park Comprehensive Cancer Center, Buffalo, NY 14263, USA. E-mail: wendy.huss@roswellpark.org

References

- [1] Andersen JT, Ekman P, Wolf H, Beisland HO, Johansson JE, Kontturi M, Lehtonen T and Tveter K. Can finasteride reverse the progress of benign prostatic hyperplasia? A two-year placebo-controlled study. The Scandinavian BPH Study Group. Urology 1995; 46: 631-637.
- [2] Marberger MJ. Long-term effects of finasteride in patients with benign prostatic hyperplasia: a double-blind, placebo-controlled, multicenter study. PROWESS Study Group. Urology 1998; 51: 677-686.
- [3] Kirby RS, Roehrborn C, Boyle P, Bartsch G, Jardin A, Cary MM, Sweeney M and Grossman EB. Efficacy and tolerability of doxazosin and finasteride, alone or in combination, in treatment of symptomatic benign prostatic hyperplasia: the Prospective European Doxazosin and Combination Therapy (PREDICT) trial. Urology 2003; 61: 119-126.
- [4] Lepor H, Williford WO, Barry MJ, Brawer MK, Dixon CM, Gormley G, Haakenson C, Machi M, Narayan P and Padley RJ. The efficacy of terazosin, finasteride, or both in benign prostatic hyperplasia. Veterans Affairs Cooperative Studies Benign Prostatic Hyperplasia Study Group. N Engl J Med 1996; 335: 533-539.
- [5] McConnell JD, Roehrborn CG, Bautista OM, Andriole GL Jr, Dixon CM, Kusek JW, Lepor H, McVary KT, Nyberg LM Jr, Clarke HS, Crawford ED, Diokno A, Foley JP, Foster HE, Jacobs SC, Kaplan SA, Kreder KJ, Lieber MM, Lucia MS, Miller GJ, Menon M, Milam DF, Ramsdell JW, Schenkman NS, Slawin KM and Smith JA. The long-term effect of doxazosin, finasteride, and combination therapy on the clinical progression of benign prostatic hyperplasia. N Engl J Med 2003; 349: 2387-2398.
- [6] Bortnick E, Brown C, Simma-Chiang V and Kaplan SA. Modern best practice in the management of benign prostatic hyperplasia in the elderly. Ther Adv Urol 2020; 12: 1756287220929486.
- [7] Lerner LB, McVary KT, Barry MJ, Bixler BR, Dahm P, Das AK, Gandhi MC, Kaplan SA, Kohler TS, Martin L, Parsons JK, Roehrborn CG, Stoffel JT, Welliver C and Wilt TJ. Management of lower urinary tract symptoms attributed to benign prostatic hyperplasia: AUA GUIDELINE PART I-initial work-up and medical management. J Urol 2021; 206: 806-817.
- [8] Shindo T, Hashimoto K, Shimizu T, Itoh N and Masumori N. Significance of intraprostatic architecture and regrowth velocity for consider-

ing discontinuation of dutasteride after combination therapy with an alpha blocker: a prospective, pilot study. Korean J Urol 2015; 56: 305-309.

- [9] English HF, Santen RJ and Isaacs JT. Response of glandular versus basal rat ventral prostatic epithelial cells to androgen withdrawal and replacement. Prostate 1987; 11: 229-242.
- [10] Blum R, Gupta R, Burger PE, Ontiveros CS, Salm SN, Xiong X, Kamb A, Wesche H, Marshall L, Cutler G, Wang X, Zavadil J, Moscatelli D and Wilson EL. Molecular signatures of the primitive prostate stem cell niche reveal novel mesenchymal-epithelial signaling pathways. PLoS One 2010; 5: e13024.
- [11] Chen H, Lin J, Shan Y and Zhengmao L. The promotion of nanoparticle delivery to two populations of gastric cancer stem cells by CD133 and CD44 antibodies. Biomed Pharmacother 2019; 115: 108857.
- [12] Lin J, Zhou J, Xu W, Zhong X, Hong Z and Peng J. Qianliening capsule treats benign prostatic hyperplasia via suppression of the EGF/STAT3 signaling pathway. Exp Ther Med 2013; 5: 1293-1300.
- [13] Rick FG, Abi-Chaker A, Szalontay L, Perez R, Jaszberenyi M, Jayakumar AR, Shamaladevi N, Szepeshazi K, Vidaurre I, Halmos G, Krishan A, Block NL and Schally AV. Shrinkage of experimental benign prostatic hyperplasia and reduction of prostatic cell volume by a gastrinreleasing peptide antagonist. Proc Natl Acad Sci U S A 2013; 110: 2617-2622.
- [14] Chang RT, Kirby R and Challacombe BJ. Is there a link between BPH and prostate cancer? Practitioner 2012; 256: 13-16, 2.
- [15] Eaton CL. Aetiology and pathogenesis of benign prostatic hyperplasia. Curr Opin Urol 2003; 13: 7-10.
- [16] McNeal JE. Origin and evolution of benign prostatic enlargement. Invest Urol 1978; 15: 340-345.
- [17] Walsh PC. Human benign prostatic hyperplasia: etiological considerations. Prog Clin Biol Res 1984; 145: 1-25.
- [18] Wei JT, Calhoun E and Jacobsen SJ. Urologic diseases in America project: benign prostatic hyperplasia. J Urol 2005; 173: 1256-1261.
- [19] Ekman P. BPH epidemiology and risk factors. Prostate Suppl 1989; 2: 23-31.
- [20] Tang J and Yang J. Etiopathogenesis of benign prostatic hypeprlasia. Indian J Urol 2009; 25: 312-317.
- [21] Berry SJ, Coffey DS, Walsh PC and Ewing LL. The development of human benign prostatic hyperplasia with age. J Urol 1984; 132: 474-479.
- [22] Isaacs JT. Prostate stem cells and benign prostatic hyperplasia. Prostate 2008; 68: 1025-1034.

- [23] Notara M and Ahmed A. Benign prostate hyperplasia and stem cells: a new therapeutic opportunity. Cell Biol Toxicol 2012; 28: 435-442.
- [24] Prajapati A, Gupta S and Mistry B. Prostate stem cells in the development of benign prostate hyperplasia and prostate cancer: emerging role and concepts. Biomed Res Int 2013; 2013: 107954.
- [25] Schuster GA and Schuster TG. The relative amount of epithelium, muscle, connective tissue and lumen in prostatic hyperplasia as a function of the mass of tissue resected. J Urol 1999; 161: 1168-1173.
- [26] McNeal JE. Regional morphology and pathology of the prostate. Am J Clin Pathol 1968; 49: 347-357.
- [27] McNeal JE, Redwine EA, Freiha FS and Stamey TA. Zonal distribution of prostatic adenocarcinoma. Correlation with histologic pattern and direction of spread. Am J Surg Pathol 1988; 12: 897-906.
- [28] McNeal JE. Normal anatomy of the prostate and changes in benign prostatic hypertrophy and carcinoma. Semin Ultrasound CT MR 1988; 9: 329-334.
- [29] McNeal JE. Normal histology of the prostate. Am J Surg Pathol 1988; 12: 619-633.
- [30] McNeal J. Pathology of benign prostatic hyperplasia. Insight into etiology. Urol Clin North Am 1990; 17: 477-486.
- [31] Peeling WB. Diagnostic assessment of benign prostatic hyperplasia. Prostate Suppl 1989; 2: 51-68.
- [32] Tanguay S, Awde M, Brock G, Casey R, Kozak J, Lee J, Nickel JC and Saad F. Diagnosis and management of benign prostatic hyperplasia in primary care. Can Urol Assoc J 2009; 3 Suppl 2: S92-S100.
- [33] Isaacs JT and Coffey DS. Etiology and disease process of benign prostatic hyperplasia. Prostate Suppl 1989; 2: 33-50.
- [34] McConnell JD. The pathophysiology of benign prostatic hyperplasia. J Androl 1991; 12: 356-363.
- [35] Schroder FH and Blom JH. Natural history of benign prostatic hyperplasia (BPH). Prostate Suppl 1989; 2: 17-22.
- [36] Isaacs JT, Brendler CB and Walsh PC. Changes in the metabolism of dihydrotestosterone in the hyperplastic human prostate. J Clin Endocrinol Metab 1983; 56: 139-146.
- [37] Brendler CB, Follansbee AL and Isaacs JT. Discrimination between normal, hyperplastic and malignant human prostatic tissues by enzymatic profiles. J Urol 1985; 133: 495-501.
- [38] Wilson JD. The pathogenesis of benign prostatic hyperplasia. Am J Med 1980; 68: 745-756.
- [39] Walsh PC, Hutchins GM and Ewing LL. Tissue content of dihydrotestosterone in human pros-

tatic hyperplasis is not supranormal. J Clin Invest 1983; 72: 1772-1777.

- [40] Claus S, Wrenger M, Senge T and Schulze H. Immunohistochemical determination of age related proliferation rates in normal and benign hyperplastic human prostates. Urol Res 1993; 21: 305-308.
- [41] Shapiro E, Becich MJ, Hartanto V and Lepor H. The relative proportion of stromal and epithelial hyperplasia is related to the development of symptomatic benign prostate hyperplasia. J Urol 1992; 147: 1293-1297.
- [42] Cunha GR and Lung B. The possible influence of temporal factors in androgenic responsiveness of urogenital tissue recombinants from wild-type and androgen-insensitive (Tfm) mice. J Exp Zool 1978; 205: 181-193.
- [43] Bianchi-Frias D, Vakar-Lopez F, Coleman IM, Plymate SR, Reed MJ and Nelson PS. The effects of aging on the molecular and cellular composition of the prostate microenvironment. PLoS One 2010; 5: e12501.
- [44] Bhowmick NA, Chytil A, Plieth D, Gorska AE, Dumont N, Shappell S, Washington MK, Neilson EG and Moses HL. TGF-beta signaling in fibroblasts modulates the oncogenic potential of adjacent epithelia. Science 2004; 303: 848-851.
- [45] Morrison C, Thornhill J and Gaffney E. The connective tissue framework in the normal prostate, BPH and prostate cancer: analysis by scanning electron microscopy after cellular digestion. Urol Res 2000; 28: 304-307.
- [46] Geller J. Pathogenesis and medical treatment of benign prostatic hyperplasia. Prostate Suppl 1989; 2: 95-104.
- [47] Geller J. Overview of benign prostatic hypertrophy. Urology 1989; 34: 57-63.
- [48] Imperato-McGinley J, Guerrero L, Gautier T and Peterson RE. Steroid 5alpha-reductase deficiency in man: an inherited form of male pseudohermaphroditism. Science 1974; 186: 1213-1215.
- [49] Siiteri PK and Wilson JD. Dihydrotestosterone in prostatic hypertrophy. I. The formation and content of dihydrotestosterone in the hypertrophic prostate of man. J Clin Invest 1970; 49: 1737-1745.
- [50] Morfin RF, Di Stefano S, Bercovici JP and Floch HH. Comparison of testosterone, 5alpha-dihydrotestosterone and 5alpha-adrostane-3beta, 17beta-diol metabolisms in human normal and hyperplastic prostates. J Steroid Biochem 1978; 9: 245-252.
- [51] Wilkin RP, Bruchovsky N, Shnitka TK, Rennie PS and Comeau TL. Stromal 5 alpha-reductase activity is elevated in benign prostatic hyperplasia. Acta Endocrinol (Copenh) 1980; 94: 284-288.

- [52] Habib FK, Lee IR, Stitch SR and Smith PH. Androgen levels in the plasma and prostatic tissues of patients with benign hypertrophy and carcinoma of the prostate. J Endocrinol 1976; 71: 99-107.
- [53] Hammond GL. Endogenous steroid levels in the human prostate from birth to old age: a comparison of normal and diseased tissues. J Endocrinol 1978; 78: 7-19.
- [54] Devlin CM, Simms MS and Maitland NJ. Benign prostatic hyperplasia - what do we know? BJU Int 2021; 127: 389-399.
- [55] Izumi K, Mizokami A, Lin WJ, Lai KP and Chang C. Androgen receptor roles in the development of benign prostate hyperplasia. Am J Pathol 2013; 182: 1942-1949.
- [56] Lai KP, Huang CK, Chang YJ, Chung CY, Yamashita S, Li L, Lee SO, Yeh S and Chang C. New therapeutic approach to suppress castration-resistant prostate cancer using ASC-J9 via targeting androgen receptor in selective prostate cells. Am J Pathol 2013; 182: 460-473.
- [57] Yu S, Zhang C, Lin CC, Niu Y, Lai KP, Chang HC, Yeh SD, Chang C and Yeh S. Altered prostate epithelial development and IGF-1 signal in mice lacking the androgen receptor in stromal smooth muscle cells. Prostate 2011; 71: 517-524.
- [58] Lai KP, Yamashita S, Vitkus S, Shyr CR, Yeh S and Chang C. Suppressed prostate epithelial development with impaired branching morphogenesis in mice lacking stromal fibromuscular androgen receptor. Mol Endocrinol 2012; 26: 52-66.
- [59] Yu S, Yeh CR, Niu Y, Chang HC, Tsai YC, Moses HL, Shyr CR, Chang C and Yeh S. Altered prostate epithelial development in mice lacking the androgen receptor in stromal fibroblasts. Prostate 2012; 72: 437-449.
- [60] Wu CT, Altuwaijri S, Ricke WA, Huang SP, Yeh S, Zhang C, Niu Y, Tsai MY and Chang C. Increased prostate cell proliferation and loss of cell differentiation in mice lacking prostate epithelial androgen receptor. Proc Natl Acad Sci U S A 2007; 104: 12679-12684.
- [61] Kerr JF and Searle J. Deletion of cells by apoptosis during castration-induced involution of the rat prostate. Virchows Arch B Cell Pathol 1973; 13: 87-102.
- [62] Kyprianou N and Isaacs JT. Activation of programmed cell death in the rat ventral prostate after castration. Endocrinology 1988; 122: 552-562.
- [63] Staack A, Kassis AP, Olshen A, Wang Y, Wu D, Carroll PR, Grossfeld GD, Cunha GR and Hayward SW. Quantitation of apoptotic activity following castration in human prostatic tissue in vivo. Prostate 2003; 54: 212-219.

- [64] Shabsigh A, Chang DT, Heitjan DF, Kiss A, Olsson CA, Puchner PJ and Buttyan R. Rapid reduction in blood flow to the rat ventral prostate gland after castration: preliminary evidence that androgens influence prostate size by regulating blood flow to the prostate gland and prostatic endothelial cell survival. Prostate 1998; 36: 201-206.
- [65] McLaren ID, Jerde TJ and Bushman W. Role of interleukins, IGF and stem cells in BPH. Differentiation 2011; 82: 237-243.
- [66] Jerde TJ and Bushman W. IL-1 induces IGFdependent epithelial proliferation in prostate development and reactive hyperplasia. Sci Signal 2009; 2: ra49.
- [67] Ruan W, Powell-Braxton L, Kopchick JJ and Kleinberg DL. Evidence that insulin-like growth factor I and growth hormone are required for prostate gland development. Endocrinology 1999; 140: 1984-1989.
- [68] DiGiovanni J, Kiguchi K, Frijhoff A, Wilker E, Bol DK, Beltran L, Moats S, Ramirez A, Jorcano J and Conti C. Deregulated expression of insulin-like growth factor 1 in prostate epithelium leads to neoplasia in transgenic mice. Proc Natl Acad Sci U S A 2000; 97: 3455-3460.
- [69] Hahn AM, Myers JD, McFarland EK, Lee S and Jerde TJ. Interleukin-driven insulin-like growth factor promotes prostatic inflammatory hyperplasia. J Pharmacol Exp Ther 2014; 351: 605-615.
- [70] Shah A, Shah AA, K N and Lobo R. Mechanistic targets for BPH and prostate cancer - a review. Rev Environ Health 2021; 36: 261-270.
- [71] Kassen A, Sutkowski DM, Ahn H, Sensibar JA, Kozlowski JM and Lee C. Stromal cells of the human prostate: initial isolation and characterization. Prostate 1996; 28: 89-97.
- [72] Lean ME, Han TS and Morrison CE. Waist circumference as a measure for indicating need for weight management. BMJ 1995; 311: 158-161.
- [73] Ross R, Neeland IJ, Yamashita S, Shai I, Seidell J, Magni P, Santos RD, Arsenault B, Cuevas A, Hu FB, Griffin BA, Zambon A, Barter P, Fruchart JC, Eckel RH, Matsuzawa Y and Despres JP. Waist circumference as a vital sign in clinical practice: a Consensus Statement from the IAS and ICCR Working Group on Visceral Obesity. Nat Rev Endocrinol 2020; 16: 177-189.
- [74] Wang S, Mao Q, Lin Y, Wu J, Wang X, Zheng X and Xie L. Body mass index and risk of BPH: a meta-analysis. Prostate Cancer Prostatic Dis 2012; 15: 265-272.
- [75] Gotera W, Mahadita GW, Bakta IM, Oka AA, Budiartha AAG, Putra Manuaba IB and Maliawan S. Waist circumference increased risk of benign prostatic hyperplasia through an increase in the level of interleukin-6 and insu-

lin resistance in abdominal obesity patients. Bali Medical Journal 2017; 6: 204.

- [76] Ricke EA, Williams K, Lee YF, Couto S, Wang Y, Hayward SW, Cunha GR and Ricke WA. Androgen hormone action in prostatic carcinogenesis: stromal androgen receptors mediate prostate cancer progression, malignant transformation and metastasis. Carcinogenesis 2012; 33: 1391-1398.
- [77] Tanner MJ, Welliver RC Jr, Chen M, Shtutman M, Godoy A, Smith G, Mian BM and Buttyan R. Effects of androgen receptor and androgen on gene expression in prostate stromal fibroblasts and paracrine signaling to prostate cancer cells. PLoS One 2011; 6: e16027.
- [78] Ho SM, Tang WY, Belmonte de Frausto J and Prins GS. Developmental exposure to estradiol and bisphenol A increases susceptibility to prostate carcinogenesis and epigenetically regulates phosphodiesterase type 4 variant 4. Cancer Res 2006; 66: 5624-5632.
- [79] Giusti RM, Iwamoto K and Hatch EE. Diethy-Istilbestrol revisited: a review of the long-term health effects. Ann Intern Med 1995; 122: 778-788.
- [80] McPherson SJ, Ellem SJ, Simpson ER, Patchev V, Fritzemeier KH and Risbridger GP. Essential role for estrogen receptor beta in stromal-epithelial regulation of prostatic hyperplasia. Endocrinology 2007; 148: 566-574.
- [81] Prins GS and Korach KS. The role of estrogens and estrogen receptors in normal prostate growth and disease. Steroids 2008; 73: 233-244.
- [82] Cunha GR. Mesenchymal-epithelial interactions: past, present, and future. Differentiation 2008; 76: 578-586.
- [83] Joesting MS, Cheever TR, Volzing KG, Yamaguchi TP, Wolf V, Naf D, Rubin JS and Marker PC. Secreted frizzled related protein 1 is a paracrine modulator of epithelial branching morphogenesis, proliferation, and secretory gene expression in the prostate. Dev Biol 2008; 317: 161-173.
- [84] Sugimura Y, Foster BA, Hom YK, Lipschutz JH, Rubin JS, Finch PW, Aaronson SA, Hayashi N, Kawamura J and Cunha GR. Keratinocyte growth factor (KGF) can replace testosterone in the ductal branching morphogenesis of the rat ventral prostate. Int J Dev Biol 1996; 40: 941-951.
- [85] Lin VK, Wang SY, Vazquez DV, C Xu C, Zhang S and Tang L. Prostatic stromal cells derived from benign prostatic hyperplasia specimens possess stem cell like property. Prostate 2007; 67: 1265-1276.
- [86] Karhadkar SS, Bova GS, Abdallah N, Dhara S, Gardner D, Maitra A, Isaacs JT, Berman DM and Beachy PA. Hedgehog signalling in pros-

tate regeneration, neoplasia and metastasis. Nature 2004; 431: 707-712.

- [87] Peehl DM and Rubin JS. Keratinocyte growth factor: an androgen-regulated mediator of stromal-epithelial interactions in the prostate. World J Urol 1995; 13: 312-317.
- [88] Cunha GR, Hayward SW, Wang YZ and Ricke WA. Role of the stromal microenvironment in carcinogenesis of the prostate. Int J Cancer 2003; 107: 1-10.
- [89] Montecinos VP, Godoy A, Hinklin J, Vethanayagam RR and Smith GJ. Primary xenografts of human prostate tissue as a model to study angiogenesis induced by reactive stroma. PLoS One 2012; 7: e29623.
- [90] Gerdes MJ, Dang TD, Larsen M and Rowley DR. Transforming growth factor-beta1 induces nuclear to cytoplasmic distribution of androgen receptor and inhibits androgen response in prostate smooth muscle cells. Endocrinology 1998; 139: 3569-3577.
- [91] Bhatia-Gaur R, Donjacour AA, Sciavolino PJ, Kim M, Desai N, Young P, Norton CR, Gridley T, Cardiff RD, Cunha GR, Abate-Shen C and Shen MM. Roles for Nkx3.1 in prostate development and cancer. Genes Dev 1999; 13: 966-977.
- [92] Royuela M, de Miguel MP, Bethencourt FR, Sanchez-Chapado M, Fraile B, Arenas MI and Paniagua R. Estrogen receptors alpha and beta in the normal, hyperplastic and carcinomatous human prostate. J Endocrinol 2001; 168: 447-454.
- [93] Ellem SJ and Risbridger GP. The dual, opposing roles of estrogen in the prostate. Ann N Y Acad Sci 2009; 1155: 174-186.
- [94] Huang L, Pu Y, Hu WY, Birch L, Luccio-Camelo D, Yamaguchi T and Prins GS. The role of Wnt5a in prostate gland development. Dev Biol 2009; 328: 188-199.
- [95] Le V, He Y, Aldahl J, Hooker E, Yu EJ, Olson A, Kim WK, Lee DH, Wong M, Sheng R, Mi J, Geradts J, Cunha GR and Sun Z. Loss of androgen signaling in mesenchymal sonic hedgehog responsive cells diminishes prostate development, growth, and regeneration. PLoS Genet 2020; 16: e1008588.
- [96] Olson AW, Le V, Wang J, Hiroto A, Kim WK, Lee DH, Aldahl J, Wu X, Kim M, Cunha GR, You S and Sun Z. Stromal androgen and hedgehog signaling regulates stem cell niches in pubertal prostate development. Development 2021; 148: dev199738.
- [97] Peng YC, Levine CM, Zahid S, Wilson EL and Joyner AL. Sonic hedgehog signals to multiple prostate stromal stem cells that replenish distinct stromal subtypes during regeneration. Proc Natl Acad Sci U S A 2013; 110: 20611-20616.

- [98] Berges RR, Vukanovic J, Epstein JI, CarMichel M, Cisek L, Johnson DE, Veltri RW, Walsh PC and Isaacs JT. Implication of cell kinetic changes during the progression of human prostatic cancer. Clin Cancer Res 1995; 1: 473-480.
- [99] Beltran H, Hruszkewycz A, Scher HI, Hildesheim J, Isaacs J, Yu EY, Kelly K, Lin D, Dicker A, Arnold J, Hecht T, Wicha M, Sears R, Rowley D, White R, Gulley JL, Lee J, Diaz Meco M, Small EJ, Shen M, Knudsen K, Goodrich DW, Lotan T, Zoubeidi A, Sawyers CL, Rudin CM, Loda M, Thompson T, Rubin MA, Tawab-Amiri A, Dahut W and Nelson PS. The role of lineage plasticity in prostate cancer therapy resistance. Clin Cancer Res 2019; 25: 6916-6924.
- [100] Crowell PD, Fox JJ, Hashimoto T, Diaz JA, Navarro HI, Henry GH, Feldmar BA, Lowe MG, Garcia AJ, Wu YE, Sajed DP, Strand DW and Goldstein AS. Expansion of luminal progenitor cells in the aging mouse and human prostate. Cell Rep 2019; 28: 1499-1510.
- [101] Karthaus WR, Iaquinta PJ, Drost J, Gracanin A, van Boxtel R, Wongvipat J, Dowling CM, Gao D, Begthel H, Sachs N, Vries RG, Cuppen E, Chen Y, Sawyers CL and Clevers HC. Identification of multipotent luminal progenitor cells in human prostate organoid cultures. Cell 2014; 159: 163-175.
- [102] Middleton LW, Shen Z, Varma S, Pollack AS, Gong X, Zhu S, Zhu C, Foley JW, Vennam S, Sweeney RT, Tu K, Biscocho J, Eminaga O, Nolley R, Tibshirani R, Brooks JD, West RB and Pollack JR. Genomic analysis of benign prostatic hyperplasia implicates cellular re-landscaping in disease pathogenesis. JCI Insight 2019; 5: e129749.
- [103] Brennen WN and Isaacs JT. Mesenchymal stem cells and the embryonic reawakening theory of BPH. Nature reviews. Urology 2018; 15: 703-715.
- [104] Evans GS and Chandler JA. Cell proliferation studies in the rat prostate: II. The effects of castration and androgen-induced regeneration upon basal and secretory cell proliferation. Prostate 1987; 11: 339-351.
- [105] Verhagen AP, Aalders TW, Ramaekers FC, Debruyne FM and Schalken JA. Differential expression of keratins in the basal and luminal compartments of rat prostatic epithelium during degeneration and regeneration. Prostate 1988; 13: 25-38.
- [106] Ravenna L, Lubrano C, Di Silverio F, Vacca A, Felli MP, Maroder M, D'Eramo G, Sciarra F, Frati L, Gulino A, et al. Androgenic and antiandrogenic control on epidermal growth factor, epidermal growth factor receptor, and androgen receptor expression in human prostate cancer cell line LNCaP. Prostate 1995; 26: 290-298.

- [107] Chua CW, Shibata M, Lei M, Toivanen R, Barlow LJ, Bergren SK, Badani KK, McKiernan JM, Benson MC, Hibshoosh H and Shen MM. Single luminal epithelial progenitors can generate prostate organoids in culture. Nat Cell Biol 2014; 16: 951-961, 951-954.
- [108] Karthaus WR, Iaquinta PJ, Drost J, Gracanin A, van Boxtel R, Wongvipat J, Dowling CM, Gao D, Begthel H, Sachs N, Vries RGJ, Cuppen E, Chen Y, Sawyers CL and Clevers HC. Identification of multipotent luminal progenitor cells in human prostate organoid cultures. Cell 2014; 159: 163-175.
- [109] Crowley L, Cambuli F, Aparicio L, Shibata M, Robinson BD, Xuan S, Li W, Hibshoosh H, Loda M, Rabadan R and Shen MM. A single-cell atlas of the mouse and human prostate reveals heterogeneity and conservation of epithelial progenitors. Elife 2020; 9: e59465.
- [110] Claus S, Berges R, Senge T and Schulze H. Cell kinetic in epithelium and stroma of benign prostatic hyperplasia. J Urol 1997; 158: 217-221.
- [111] Kyprianou N, Tu H and Jacobs SC. Apoptotic versus proliferative activities in human benign prostatic hyperplasia. Hum Pathol 1996; 27: 668-675.
- [112] Colombel M, Vacherot F, Diez SG, Fontaine E, Buttyan R and Chopin D. Zonal variation of apoptosis and proliferation in the normal prostate and in benign prostatic hyperplasia. Br J Urol 1998; 82: 380-385.
- [113] Thompson TC and Yang G. Regulation of apoptosis in prostatic disease. Prostate Suppl 2000; 9: 25-28.
- [114] Zhang X, Zhang Q, Zhang Z, Na Y and Guo Y. Apoptosis profiles in benign prostatic hyperplasia: close associations of cell kinetics with percent area density of histologic composition. Urology 2006; 68: 905-910.
- [115] Chen M, Tanner M, Levine AC, Levina E, Ohouo P and Buttyan R. Androgenic regulation of hedgehog signaling pathway components in prostate cancer cells. Cell Cycle 2009; 8: 149-157.
- [116] Fibbi B, Penna G, Morelli A, Adorini L and Maggi M. Chronic inflammation in the pathogenesis of benign prostatic hyperplasia. Int J Androl 2010; 33: 475-488.
- [117] Rosenberg MT, Miner MM, Riley PA and Staskin DR. STEP: simplified treatment of the enlarged prostate. Int J Clin Pract 2010; 64: 488-496.
- [118] AUA Practice Guidelines Committee. AUA guideline on management of benign prostatic hyperplasia (2003). Chapter 1: diagnosis and treatment recommendations. J Urol 2003; 170: 530-547.
- [119] Mirone V, Sessa A, Giuliano F, Berges R, Kirby M and Moncada I. Current benign prostatic hy-

perplasia treatment: impact on sexual function and management of related sexual adverse events. Int J Clin Pract 2011; 65: 1005-1013.

- [120] Lerner LB, McVary KT, Barry MJ, Bixler BR, Dahm P, Das AK, Gandhi MC, Kaplan SA, Kohler TS, Martin L, Parsons JK, Roehrborn CG, Stoffel JT, Welliver C and Wilt TJ. Management of lower urinary tract symptoms attributed to benign prostatic hyperplasia: AUA GUIDELINE PART II-surgical evaluation and treatment. J Urol 2021; 206: 818-826.
- [121] Bajek A, Pokrywka L, Wolski Z, Debski R and Drewa T. Prostate epithelial stem cells are resistant to apoptosis after alpha1-antagonist treatment. The impact for BPH patients. Cent European J Urol 2011; 64: 256-257.
- [122] Chlosta P, Drewa T and Kaplan S. Alphablockade, apoptosis, and prostate shrinkage: how are they related? Cent European J Urol 2013; 66: 189-194.
- [123] Kyprianou N. Doxazosin and terazosin suppress prostate growth by inducing apoptosis: clinical significance. J Urol 2003; 169: 1520-1525.
- [124] Lucia MS and Lambert JR. Growth factors in benign prostatic hyperplasia: basic science implications. Curr Urol Rep 2008; 9: 272-278.
- [125] Orr B, Grace OC, Vanpoucke G, Ashley GR and Thomson AA. A role for notch signaling in stromal survival and differentiation during prostate development. Endocrinology 2009; 150: 463-472.
- [126] Wu X, Xu K, Zhang L, Deng Y, Lee P, Shapiro E, Monaco M, Makarenkova HP, Li J, Lepor H and Grishina I. Differentiation of the ductal epithelium and smooth muscle in the prostate gland are regulated by the Notch/PTEN-dependent mechanism. Dev Biol 2011; 356: 337-349.
- [127] Nicholson TM, Moses MA, Uchtmann KS, Keil KP, Bjorling DE, Vezina CM, Wood RW and Ricke WA. Estrogen receptor- α is a key mediator and therapeutic target for bladder complications of benign prostatic hyperplasia. J Urol 2015; 193: 722-729.
- [128] Bushman W. Etiology, epidemiology, and natural history of benign prostatic hyperplasia. Urol Clin North Am 2009; 36: 403-415.
- [129] Bonkhoff H and Berges R. The evolving role of oestrogens and their receptors in the development and progression of prostate cancer. Eur Urol 2009; 55: 533-542.
- [130] Foster BA, Gangavarapu KJ, Mathew G, Azabdaftari G, Morrison CD, Miller A and Huss WJ. Human prostate side population cells demonstrate stem cell properties in recombination with urogenital sinus mesenchyme. PLoS One 2013; 8: e55062.
- [131] Gangavarapu KJ, Azabdaftari G, Morrison CD, Miller A, Foster BA and Huss WJ. Aldehyde de-

hydrogenase and ATP binding cassette transporter G2 (ABCG2) functional assays isolate different populations of prostate stem cells where ABCG2 function selects for cells with increased stem cell activity. Stem Cell Res Ther 2013; 4: 132.

- [132] Sabnis NG, Miller A, Titus MA and Huss WJ. The efflux transporter ABCG2 maintains prostate stem cells. Mol Cancer Res 2017; 15: 128-140.
- [133] Samant MD, Jackson CM, Felix CL, Jones AJ, Goodrich DW, Foster BA and Huss WJ. Multidrug resistance ABC transporter inhibition enhances murine ventral prostate stem/progenitor cell differentiation. Stem Cells Dev 2015; 24: 1236-1251.
- [134] Ertas YN, Abedi Dorcheh K, Akbari A and Jabbari E. Nanoparticles for targeted drug delivery to cancer stem cells: a review of recent advances. Nanomaterials (Basel) 2021; 11: 1755.
- [135] Wei J, Sun J and Liu Y. Enhanced targeting of prostate cancer-initiating cells by salinomycinencapsulated lipid-PLGA nanoparticles linked with CD44 antibodies. Oncol Lett 2019; 17: 4024-4033.
- [136] Atawia RT, Tadros MG, Khalifa AE, Mosli HA and Abdel-Naim AB. Role of the phytoestrogenic, pro-apoptotic and anti-oxidative properties of silymarin in inhibiting experimental benign prostatic hyperplasia in rats. Toxicol Lett 2013; 219: 160-169.

- [137] Djavan B, Eckersberger E, Finkelstein J, Espinosa G, Sadri H, Brandner R, Shah O and Lepor H. Benign prostatic hyperplasia: current clinical practice. Prim Care 2010; 37: 583-597.
- [138] Espinosa G. Nutrition and benign prostatic hyperplasia. Curr Opin Urol 2013; 23: 38-41.
- [139] Kim TH, Lim HJ, Kim MS and Lee MS. Dietary supplements for benign prostatic hyperplasia: an overview of systematic reviews. Maturitas 2012; 73: 180-185.
- [140] Lu C, Li X, Ren Y and Zhang X. Disulfiram: a novel repurposed drug for cancer therapy. Cancer Chemother Pharmacol 2021; 87: 159-172.
- [141] Le Magnen C, Bubendorf L, Rentsch CA, Mengus C, Gsponer J, Zellweger T, Rieken M, Thalmann GN, Cecchini MG, Germann M, Bachmann A, Wyler S, Heberer M and Spagnoli GC. Characterization and clinical relevance of ALDHbright populations in prostate cancer. Clin Cancer Res 2013; 19: 5361-5371.