

Review Article

The dysfunctional lipids in prostate cancer

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Received July 31, 2019; Accepted August 2, 2019; Epub August 15, 2019; Published August 30, 2019

Abstract: Prostate cancer (PCa) is well-recognized as a lipid-enriched tumor. Lipids represent a diverse array of molecules essential to the cellular structure, defense, energy, and communication. Lipid metabolism can often become dysregulated during tumor development. The increasing body of knowledge on the biological actions of steroid hormone-androgens in PCa has led to the development of several targeted therapies that still represent the standard of care for cancer patients to this day. Sequencing technologies for functional analyses of androgen receptors (ARs) have revealed that AR is also a master regulator of cellular energy metabolism such as fatty acid β -oxidation, and de novo lipid synthesis. In addition, bioactive lipids are also used as physiological signaling molecules, which have been shown to be involved in PCa progression. This review discusses the potent player(s) in altered lipid metabolism of PCa and describes how lipids and their interactions with proteins can be used for therapeutic advantage. We also discuss the possibility that the altered bioactive lipid mediators affect intracellular signaling pathway and the related transcriptional regulation be of therapeutic interest.

Keywords: Lipid metabolism, bioactive lipids, prostate cancer, cancer progression

Introduction

A key player in prostate cancer (PCa) development and progression is the androgen receptor (AR). Pathologically, PCa is known as a lipid-rich tumor [1]. Indeed, several genes encoding lipogenic enzymes can be regulated by androgen [2-7]; increased synthesis of fatty acids and cholesterol is governed by androgens through stimulation of the expression of whole sets of lipogenic enzymes, covering the entire pathways of fatty acid (FA) pathway. The resulting increase in lipogenesis helps synthesise the synthesis of key membrane components (phospholipids, cholesterol) and is a major hallmark of cancer cells. In addition, an increase in total cholesterol and in triglycerides duration of androgen deprivation therapy (ADT) that ranges from 24 weeks to 12 months [8-12]. While increased lipogenesis is initially androgen-responsive it persists or re-emerges with the development of castration resistant PCa (CR-PC), indicating that lipid metabolism is a fundamental aspect of PCa cell biology. In this review, we discuss the lipid landscape and the possible underlying mechanisms mediating PCa development and progression.

Lipogenesis in prostate carcinogenesis

Cancer cells usually exhibit the ability of rapid proliferation. In order to deal with this altered growth rate, changes in the cellular metabolic pathways are always displayed [13, 14]. Since 1950s, researchers have noticed the metabolic dysregulation in cancer cells and it has been widely studied. Some of the most well-known alterations include Warburg effect [15] and increased glutamine metabolism [16]. Recently, lipid metabolism emerges as a more and more important role in cancer. Since lipids supply energy, provide signaling molecules and synthesize the cellular membrane [16-18], highly proliferative cancer cells often have a higher demand for lipids and exhibit an abnormally active lipogenesis [19]. Unlike most normal somatic cells, which mainly utilize the exogenous lipids, studies have shown that many cancer cells mainly use de novo FA synthesis to increase total FA [8], regardless of abundant extracellular lipid content [20-22]. Fatty acid synthase (FASN) is the key enzyme in fatty acid synthesis. Since the identification of onco-antigen OA-519 as a FASN in breast cancer 20 years ago, it has now become a well-established

lished oncogene in numerous types of cancer, including prostate, ovary, colon, endometrium, lung, bladder, stomach, esophagus and pancreas [23, 24]. Higher FASN expression is associated with poor survival and disease recurrence in PCa patients [25-27]. Even in some pre-invasive lesions, elevated FASN can be detected [27]. Consistent with the clinical observation, immortalized prostate epithelial cells with overexpressing FASN exhibit increased invasion ability [28]. In addition, ATP citrate lyase (ACLY) is the first enzyme of the reaction chain and serves as an important bridge between glycolysis and lipogenesis by catalyzing coenzyme A (CoA) and citrate that is a product of glycolysis [29, 30]. Clinically, increased ACLY represents an unfavorable biomarker for PCa and several other cancer types including bladder, renal, non-small cell lung, colorectal, breast, liver and gastric cancers [31, 32].

Although lipogenesis is considered to be the major source of FA in cancer cell, it is reported that some cancers may also adopt lipolysis as an additional method to acquire FA [33]. During certain metabolic stresses, cancer cells may switch from de novo FA synthesis to scavenging extracellular lipids [34]. Lipoprotein lipase (LPL) is the key enzyme for extracellular lipolysis, which hydrolyzes the triglycerides (TGs) in chylomicrons or very low-density lipoproteins (VLDL), and the FA produced from hydrolysis are then taken up by the cancer cells through the transmembrane channel CD36 [35, 36]. Indeed, both LPL and CD36 are ubiquitously expressed in PCa tissues [33, 36], suggesting lipolysis may also contribute to PCa development.

Bioactive lipid mediators in PCa progression

Phosphoinositides (PIs) are major second messengers, which transmit signals from activated growth factor receptors on the cellular surface to the interior of the cells. Saturated and unsaturated fatty acids combine with glycerol-3-phosphate in glycerolipid biosynthesis which is highly dependent on glycerol-3-phosphate acyltransferase (GPAT) to produce PIs and phosphoglycerides [37]. One of the most prominent lipids of this class is phosphatidylinositol (3,4,5)-trisphosphate [PtdIns (3,4,5) P3; PIP3], which is produced by PI3K in response to growth factor signaling and mediates the recruitment and activation of Akt [38]. In con-

trast, PTEN (phosphatase and tensin homolog deleted on chromosome ten) is a PIP3 phosphatase and commonly downregulated in PCa [39]. Other lipid second messengers, such as lysophosphatidic acid (LPA), phosphatidic acid (PA) and diacylglycerol (DAG), which are produced by the different phospholipases [40]. LPA can be produced by the extracellular lysophospholipase or autotaxin, which can activate cell proliferation, migration and survival via binding to G-protein-coupled receptors [41]. PA can bind to the mTOR polybasic domain, which is essential for its activation. The phosphoinositide-specific phospholipases C (PLC) can transform phosphatidylinositol 4,5-bisphosphate [PtdIns (4,5) P2] into the DAG and inositol 1,4,5-trisphosphate. Several studies demonstrated that PLC γ 1 plays an important role in PCa metastasis [42, 43].

Ceramide as the central molecule in the sphingolipid metabolism. The balance between the levels of sphingosine-1-phosphate (S1P) and its metabolic precursors ceramide and sphingosine has been regarded as a rheostat that could determine cell fate [44, 45]. For example, ceramide mediates numerous cell-stress responses such as induction of apoptosis and cell senescence, whereas S1P plays a pivotal role in cell survival, migration, and inflammation. Ceramide production was correlated with enhanced apoptosis in LNCaP cells treated with TNF- α and irradiation. Ceramide treatment can specifically kill PCa cells but not normal prostate epithelial cells by decreasing c-myc expression [46].

Accumulating evidence links S1P produced by Sphingosine kinase 1 (Sphk1) with PCa; Sphk1 is elevated in primary PCa lesion compared to adjacent benign tissue [47, 48]. Using PCa cell culture models, elevated Sphk1 can promote PCa invasion, which is mediated by Sphingosine-1-phosphate receptor 4 (S1PR4)-Matriptase activation [49]. Also, upregulation of the SphK1-S1P pathway is associated with chemoresistance in PCa cells [50]. A selective SphK1 inhibitor (such as FTY720) can trigger apoptosis of a variety of PCa cells including androgen-responsive LNCaP, androgen variant-expressing 22RV1 and castration resistant PC3 cell [51-53]. Furthermore, hypoxia can activate Sphk1 enzyme activity leading to the stabilization of HIF-1 α levels, which could lead to radio-

resistant PCa [54]. These results indicate the critical role of Sphk1 in PCa survival.

Regulators of lipid metabolism in PCa

Sterol regulatory element-binding proteins (SREBPs) are transcription factors that can mediate the homeostasis of cholesterol and fatty acids [55]. SREBP-1 mainly regulates genes in FA synthesis and is highly elevated in PCa [56]. SREBP-2 is responsible for cholesterol synthesis. Upregulated of SREBP-2 has been found in PCa patient tumor tissues [57]. Several lines of evidence indicate that androgens activate the SREBP pathway (1) Androgens stimulate the nuclear accumulation of mature SREBP [2]. (2) Androgen stimulation of key lipogenic genes (fatty acid synthase, HMG-CoA synthase) is abolished when the SREBP binding sites in the proximal promoter are deleted or mutated [2-5]. (3) Introduction of a dominant-negative SREBP strongly suppresses the lipogenic effects of androgens [2-5]. In several instances, the lipogenic effects of androgens are more pronounced than estimated from the changes in mRNA levels of lipogenic genes, suggesting that also translational and/or post-translational mechanisms are involved [3]. Moreover, PI3K/Akt/mTOR can upregulate the SREBPs expression and stabilize the nuclear form of SREBP-1 nuclear form, and then promote its target gene expression via decrease in the expression of Fbw-7 in mediating fatty acid synthesis and cholesterol uptake. SREBPs can be activated by SREBP-cleavage-activating protein (SCAP) to drive expression of enzymes needed for lipid syntheses, such as fatty acid synthase (FAS), 3-hydroxy-3-methylglutaryl coenzyme A reductase (HMG CoA-R) and low-density lipoprotein receptor (LDLR) [58]. SREBP-1 can upregulate the expression of ATP citrate lyase (ACL), acetyl-CoA carboxylase (ACC) and FAS to promote fatty acid synthesis and enhance cholesterol uptake via upregulation of LDLR, thereby promoting the cancer tumor growth [59, 60]. Plk1 can induce activation of the PI3K/AKT/mTOR/GSK3 β and AR pathways and increase of lipid biosynthesis [58]. Fatostatin suppresses Plk1/SREBP, which leads to the inhibition of cell proliferation, invasion, and migration, and to arrest cancer cells at the G2/M checkpoint in both of androgen-responsive LNCaP and androgen-insensitive C4-2B PCa cells [61]. The PI3K/Akt/mTOR/SREBP signaling pathway has a positive

feedback regulatory loop, which can boost Akt expression in cell migration, tumor growth, and metastasis [62].

Peroxisome proliferator-activated receptor gamma (PPAR γ) is, a ligand-dependent transcription factor belonging to the nuclear hormone receptor superfamily, considered a master regulator for the genes involved in FA synthesis and lipogenesis. PPAR γ protein level is found significantly elevated in advanced PCa when compared to localized PCa or benign prostatic hyperplasia [63, 64]. Higher protein expression of PPAR γ is also associated with shorter patient survival duration [65]. In prostate-specific *Pten*^{-/-} mouse model, over-expression of the PPAR γ protein is associated with significantly decreased survival and increased metastases to the lungs and lymph nodes compared to littermate controls [66]. The PPAR γ inhibitor (antagonist GW9662) also decreases the growth of human PCa cells in culture [66]. Furthermore, there is a reciprocal regulation between PPAR γ and androgen receptor (AR) activity; DHT treatment decreased PPAR γ mRNA and protein levels in LNCaP C4-2 and VCaP cell lines [67]. Noticeably, PPAR γ plays a key role in IL-6-elicited neuroendocrine differentiation of PCa (NEPC) [68]. Altogether, these data support the development of PPAR γ inhibition as a new strategy of PCa treatment.

The family of PPAR γ coactivator 1-alpha coactivators (PGC1) have two isoforms, PGC1 α and β , that are transcriptional coactivators and can regulate the mitochondrial biogenesis and functions including FA and lipid metabolism. PGC1 α plays a major role in the rapid metabolically active tissues such as liver, cardiac, skeletal muscle, kidneys, and adipose tissue [69, 70] in energy-demanding situations. The PGC1 α protein is associated with PPAR γ (the role in adipogenesis, thermogenesis, and mitobiogenesis), nuclear respiratory factor 1-2 (Nrf1-2), Forkhead box O3 (FoxO3a), cyclic-AMP (cAMP) response element-binding protein (CREB) and estrogen-related receptor- α (ERR α) [69-71]. Androgens signaling can increase the expression of PGC1 α in PCa cells [72]. Clinically, in PCa patient specimens, a significant correlation between PGC1 α with tumor proliferation was reported [72].

Mammalian silent information regulator 1 (SIRT1) is a nicotinamide adenine dinucleotide

(NAD)-dependent histone deacetylase, which plays a major role in multiple physiological processes such as stress responses, metabolism, apoptosis, and calorie restriction, etc [73-77]. SIRT1 has been demonstrated to be an oncogene in mouse PCa model with PTEN deficiency [78]. Several studies indicated that SIRT1 was associated with the class III deacetylases and its targets, such as p53, PPAR γ , PGC1 α , Beclin 1 and β -catenin [79-82]. SIRT1 overexpression induces epithelial-to-mesenchymal transition (EMT) in epithelial prostate cells and increases PCa cell migration in vitro and metastasis in vivo. In contrast, inhibiting the expression of SIRT1 in PCa cells restores cell-cell adhesion and reverses EMT. Thus, SIRT1 can regulate the expression of the E-cadherin epithelial markers and γ -catenin, and the mesenchymal markers fibronectin and N-cadherin [83].

AMP-activated protein kinase (AMPK) is a serine/threonine protein kinase consisting of a catalytic subunit (α) and two regulatory subunits (β and γ) [84, 85]. Once the activation of AMPK redirects lipid metabolism towards increased catabolic fatty acid oxidation and decreased anabolic lipid synthesis via the phosphorylation of acetyl-CoA carboxylases (ACCs) phosphorylation. ACCs represents the first step in de novo lipid synthesis, which responsible for the carboxylation of acetyl-CoA to form malonyl-CoA [86]. Several studies showed that knockout of one of the catalytic subunits of AMPK, which support a tumor suppressive role for AMPK in PCa [87-89]. Elevation of both activated AMPK (Threonine 172 phosphorylation) and acetyl-CoA carboxylase (Serine 80 phosphorylation) were detected in PCa clinical samples compared to surrounding benign tissues. AMPK was also associated with the progression of PCa with higher Gleason grades and advanced stages [72, 88, 89]. Collectively, these studies support the clinical relevance of AMPK in PCa development.

Conclusion

Increased lipogenesis is an important hallmark in PCa development and androgen plays a critical role to stimulate lipogenesis. The resulting increase in the coordinate expression of multiple regulators or enzymes involved in the gene transcription, metabolism and transport of FAs and cholesterol mainly results in the synthesis

of phospholipids partitioning in various malignant activities. While increased lipogenesis is initially androgen-responsive it persists or re-emerges with the development of CRPC or NEPC, indicating that lipogenesis is a fundamental aspect of PCa cell biology and is a potential target for anti-neoplastic therapy in advanced PCa.

Disclosure of conflict of interest

None.

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References

- [1] Wu X, Daniels G, Lee P, Monaco ME. Lipid metabolism in prostate cancer. *Am J Clin Exp Urol* 2014; 2: 111-20.
- [2] Swinnen JV, Ulrix W, Heyns W, Verhoeven G. Coordinate regulation of lipogenic gene expression by androgens: evidence for a cascade mechanism involving sterol regulatory element binding proteins. *Proc Natl Acad Sci U S A* 1997; 94: 12975-80.
- [3] Swinnen JV, Van Veldhoven PP, Esquenet M, Heyns W, Verhoeven G. Androgens markedly stimulate the accumulation of neutral lipids in the human prostatic adenocarcinoma cell line LNCaP. *Endocrinology* 1996; 137: 4468-74.
- [4] Swinnen JV, Esquenet M, Goossens K, Heyns W, Verhoeven G. Androgens stimulate fatty acid synthase in the human prostate cancer cell line LNCaP. *Cancer Res* 1997; 57: 1086-90.
- [5] Heemers H, Maes B, Fougelle F, Heyns W, Verhoeven G, Swinnen JV. Androgens stimulate lipogenic gene expression in prostate cancer cells by activation of the sterol regulatory element-binding protein cleavage activating protein/sterol regulatory element-binding protein pathway. *Mol Endocrinol* 2001; 15: 1817-28.
- [6] Myers RB, Oelschlager DK, Weiss HL, Frost AR, Grizzle WE. Fatty acid synthase: an early molecular marker of progression of prostatic adenocarcinoma to androgen independence. *J Urol* 2001; 165: 1027-32.
- [7] Heemers H, Vanderhoydonc F, Roskams T, Shechter I, Heyns W, Verhoeven G, Swinnen JV. Androgens stimulate coordinated lipogenic gene expression in normal target tissues in vivo. *Mol Cell Endocrinol* 2003; 205: 21-31.

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- [8] Smith JC, Bennett S, Evans LM, Kynaston HG, Parmar M, Mason MD, Cockcroft JR, Scanlon MF, Davies JS. The effects of induced hypogonadism on arterial stiffness, body composition, and metabolic parameters in males with prostate cancer. *J Clin Endocrinol Metab* 2001; 86: 4261-7.
- [9] Smith MR. Changes in fat and lean body mass during androgen-deprivation therapy for prostate cancer. *Urology* 2004; 63: 742-5.
- [10] Nishiyama T, Ishizaki F, Anraku T, Shimura H, Takahashi K. The influence of androgen deprivation therapy on metabolism in patients with prostate cancer. *J Clin Endocrinol Metab* 2005; 90: 657-60.
- [11] Torimoto K, Samma S, Kagebayashi Y, Chihara Y, Tanaka N, Hirayama A, Fujimoto K, Hirao Y. The effects of androgen deprivation therapy on lipid metabolism and body composition in Japanese patients with prostate cancer. *Jpn J Clin Oncol* 2011; 41: 577-81.
- [12] Mitsuzuka K and Arai Y. Metabolic changes in patients with prostate cancer during androgen deprivation therapy. *Int J Urol* 2018; 25: 45-53.
- [13] DeBerardinis RJ, Lum JJ, Hatzivassiliou G, Thompson CB. The biology of cancer: metabolic reprogramming fuels cell growth and proliferation. *Cell Metab* 2008; 7: 11-20.
- [14] Benjamin DI, Cravatt BF and Nomura DK. Global profiling strategies for mapping dysregulated metabolic pathways in cancer. *Cell Metab* 2012; 16: 565-77.
- [15] Warburg O. On the origin of cancer cells. *Science* 1956; 123: 309-14.
- [16] DeBerardinis RJ, Mancuso A, Daikhin E, Nissim I, Yudkoff M, Wehrli S, Thompson CB. Beyond aerobic glycolysis: transformed cells can engage in glutamine metabolism that exceeds the requirement for protein and nucleotide synthesis. *Proc Natl Acad Sci U S A* 2007; 104: 19345-50.
- [17] Yue S, Li J, Lee SY, Lee HJ, Shao T, Song B, Cheng L, Masterson TA, Liu X, Ratliff TL, Cheng JX. Cholesteryl ester accumulation induced by PTEN loss and PI3K/AKT activation underlies human prostate cancer aggressiveness. *Cell Metab* 2014; 19: 393-406.
- [18] Accioly MT, Pacheco P, Maya-Monteiro CM, Carrossini N, Robbs BK, Oliveira SS, Kaufmann C, Morgado-Diaz JA, Bozza PT, Viola JP. Lipid bodies are reservoirs of cyclooxygenase-2 and sites of prostaglandin-E2 synthesis in colon cancer cells. *Cancer Res* 2008; 68: 1732-40.
- [19] Beloribi-Djefaffia S, Vasseur S and Guillaumond F. Lipid metabolic reprogramming in cancer cells. *Oncogenesis* 2016; 5: e189.
- [20] Medes G, Thomas A and Weinhouse S. Metabolism of neoplastic tissue. IV. A study of lipid synthesis in neoplastic tissue slices in vitro. *Cancer Res* 1953; 13: 27-9.
- [21] Ookhtens M, Kannan R, Lyon I, Baker N. Liver and adipose tissue contributions to newly formed fatty acids in an ascites tumor. *Am J Physiol* 1984; 247: R146-53.
- [22] Mashima T, Seimiya H and Tsuruo T. De novo fatty-acid synthesis and related pathways as molecular targets for cancer therapy. *Br J Cancer* 2009; 100: 1369-72.
- [23] Kuhajda FP, Jenner K, Wood FD, Hennigar RA, Jacobs LB, Dick JD, Pasternack GR. Fatty acid synthesis: a potential selective target for anti-neoplastic therapy. *Proc Natl Acad Sci U S A* 1994; 91: 6379-83.
- [24] Kuhajda FP. Fatty-acid synthase and human cancer: new perspectives on its role in tumor biology. *Nutrition* 2000; 16: 202-8.
- [25] Visca P, Sebastiani V, Botti C, Diodoro MG, Lasagni RP, Romagnoli F, Brenna A, De Joannon BC, Donnorso RP, Lombardi G, Alo PL. Fatty acid synthase (FAS) is a marker of increased risk of recurrence in lung carcinoma. *Anticancer Res* 2004; 24: 4169-73.
- [26] Corominas-Faja B, Vellon L, Cuyàs E, Buxó M, Martin-Castillo B, Serra D, García J, Lupu R, Menendez JA. Clinical and therapeutic relevance of the metabolic oncogene fatty acid synthase in HER2+ breast cancer. *Histol Histopathol* 2017; 32: 687-698.
- [27] Flavin R, Peluso S, Nguyen PL, Loda M. Fatty acid synthase as a potential therapeutic target in cancer. *Future Oncol* 2010; 6: 551-62.
- [28] Fiorentino M, Zadra G, Palescandolo E, Fedele G, Bailey D, Fiore C, Nguyen PL, Migita T, Zamponi R, Di Vizio D, Priolo C, Sharma C, Xie W, Hemler ME, Mucci L, Giovannucci E, Finn S, Loda M. Overexpression of fatty acid synthase is associated with palmitoylation of Wnt1 and cytoplasmic stabilization of beta-catenin in prostate cancer. *Lab Invest* 2008; 88: 1340-8.
- [29] Chypre M, Zaidi N and Smans K. ATP-citrate lyase: a mini-review. *Biochem Biophys Res Commun* 2012; 422: 1-4.
- [30] Zaidi N, Swinnen JV and Smans K. ATP-citrate lyase: a key player in cancer metabolism. *Cancer Res* 2012; 72: 3709-14.
- [31] Granchi C. ATP citrate lyase (ACLY) inhibitors: an anti-cancer strategy at the crossroads of glucose and lipid metabolism. *Eur J Med Chem* 2018; 157: 1276-1291.
- [32] Qian X, Hu J, Zhao J, Chen H. ATP citrate lyase expression is associated with advanced stage and prognosis in gastric adenocarcinoma. *Int J Clin Exp Med* 2015; 8: 7855-60.
- [33] Zaidi N, Lupien L, Kuemmerle NB, Kinlaw WB, Swinnen JV, Smans K. Lipogenesis and lipolysis: the pathways exploited by the cancer cells

- to acquire fatty acids. *Prog Lipid Res* 2013; 52: 585-9.
- [34] Boroughs LK and DeBerardinis RJ. Metabolic pathways promoting cancer cell survival and growth. *Nat Cell Biol* 2015; 17: 351-9.
- [35] Goldberg IJ, Eckel RH and Abumrad NA. Regulation of fatty acid uptake into tissues: lipoprotein lipase- and CD36-mediated pathways. *J Lipid Res* 2009; 50 Suppl: S86-90.
- [36] Kuemmerle NB, Rysman E, Lombardo PS, Flanagan AJ, Lipe BC, Wells WA, Pettus JR, Froehlich HM, Memoli VA, Morganelli PM, Swinnen JV, Timmerman LA, Chaychi L, Fricano CJ, Eisenberg BL, Coleman WB, Kinlaw WB. Lipoprotein lipase links dietary fat to solid tumor cell proliferation. *Mol Cancer Ther* 2011; 10: 427-36.
- [37] Chen YQ, Kuo MS, Li S, Bui HH, Peake DA, Sanders PE, Thibodeaux SJ, Chu S, Qian YW, Zhao Y, Bredt DS, Moller DE, Konrad RJ, Beigneux AP, Young SG, Cao G. AGPAT6 is a novel microsomal glycerol-3-phosphate acyltransferase. *J Biol Chem* 2008; 283: 10048-57.
- [38] Manna P and Jain SK. Phosphatidylinositol-3,4,5-triphosphate and cellular signaling: implications for obesity and diabetes. *Cell Physiol Biochem* 2015; 35: 1253-75.
- [39] Engelman JA, Luo J and Cantley LC. The evolution of phosphatidylinositol 3-kinases as regulators of growth and metabolism. *Nat Rev Genet* 2006; 7: 606-19.
- [40] Giusto NM, Pasquaré SJ, Salvador GA, Ilincheta de Boschero MG. Lipid second messengers and related enzymes in vertebrate rod outer segments. *J Lipid Res* 2010; 51: 685-700.
- [41] Yang F and Chen GX. Production of extracellular lysophosphatidic acid in the regulation of adipocyte functions and liver fibrosis. *World J Gastroenterol* 2018; 24: 4132-4151.
- [42] Shepard CR, Kassis J, Whaley DL, Kim HG, Wells A. PLC gamma contributes to metastasis of in situ-occurring mammary and prostate tumors. *Oncogene* 2007; 26: 3020-6.
- [43] Raimondi C, Chikh A, Wheeler AP, Maffucci T, Falasca M. A novel regulatory mechanism links PLCgamma1 to PDK1. *J Cell Sci* 2012; 125: 3153-63.
- [44] Cuvillier O, Pirianov G, Kleuser B, Vanek PG, Coso OA, Gutkind S, Spiegel S. Suppression of ceramide-mediated programmed cell death by sphingosine-1-phosphate. *Nature* 1996; 381: 800-3.
- [45] Brizuela L, Martin C, Jeannot P, Ader I, Gstalder C, Andrieu G, Bocquet M, Laffosse JM, Gomez-Brouchet A, Malavaud B, Sabbadini RA, Cuvillier O. Osteoblast-derived sphingosine 1-phosphate to induce proliferation and confer resistance to therapeutics to bone metastasis-derived prostate cancer cells. *Mol Oncol* 2014; 8: 1181-95.
- [46] Liu B, Fu X, Wang D, Zhang W, Yang X. Synthesis of organic-inorganic hybrid microspheres and the corresponding mesoporous silica nanoparticles. *J Colloid Interface Sci* 2013; 411: 98-104.
- [47] Brizuela L, Ader I, Mazerolles C, Bocquet M, Malavaud B, Cuvillier O. First evidence of sphingosine 1-phosphate lyase protein expression and activity downregulation in human neoplasm: implication for resistance to therapeutics in prostate cancer. *Mol Cancer Ther* 2012; 11: 1841-51.
- [48] Malavaud B, Pchejetski D, Mazerolles C, de Paiva GR, Calvet C, Doumerc N, Pitson S, Rischmann P, Cuvillier O. Sphingosine kinase-1 activity and expression in human prostate cancer resection specimens. *Eur J Cancer* 2010; 46: 3417-24.
- [49] Lee CF, Dang A, Hernandez E, Pong RC, Chen B, Sonavane R, Raj G, Kapur P, Lin HY, Wu SR, Ko CJ, Lo UG, Lee HY, Hsieh JT, Lee MS. Activation of sphingosine kinase by lipopolysaccharide promotes prostate cancer cell invasion and metastasis via SphK1/S1PR4/matrix metalloproteinase. *Oncogene* 2019; 38: 5580-5598.
- [50] Akao Y, Banno Y, Nakagawa Y, Hasegawa N, Kim TJ, Murate T, Igarashi Y, Nozawa Y. High expression of sphingosine kinase 1 and S1P receptors in chemotherapy-resistant prostate cancer PC3 cells and their camptothecin-induced up-regulation. *Biochem Biophys Res Commun* 2006; 342: 1284-90.
- [51] Furuya H, Shimizu Y and Kawamori T. Sphingolipids in cancer. *Cancer Metastasis Rev* 2011; 30: 567-76.
- [52] Wang JD, Takahara S, Nonomura N, Ichimaru N, Toki K, Azuma H, Matsumiya K, Okuyama A, Suzuki S. Early induction of apoptosis in androgen-independent prostate cancer cell line by FTY720 requires caspase-3 activation. *Prostate* 1999; 40: 50-5.
- [53] Chua CW, Lee DT, Ling MT, Zhou C, Man K, Ho J, Chan FL, Wang X, Wong YC. FTY720, a fungus metabolite, inhibits in vivo growth of androgen-independent prostate cancer. *Int J Cancer* 2005; 117: 1039-48.
- [54] Ader I, Brizuela L, Bouquerel P, Malavaud B, Cuvillier O. Sphingosine kinase 1: a new modulator of hypoxia inducible factor 1alpha during hypoxia in human cancer cells. *Cancer Res* 2008; 68: 8635-42.
- [55] Brown MS and Goldstein JL. The SREBP pathway: regulation of cholesterol metabolism by proteolysis of a membrane-bound transcription factor. *Cell* 1997; 89: 331-40.
- [56] Ettinger SL, Sobel R, Whitmore TG, Akbari M, Bradley DR, Gleave ME, Nelson CC. Dysregulation

- tion of sterol response element-binding proteins and downstream effectors in prostate cancer during progression to androgen independence. *Cancer Res* 2004; 64: 2212-21.
- [57] Li X, Wu JB, Li Q, Shigemura K, Chung LW, Huang WC. SREBP-2 promotes stem cell-like properties and metastasis by transcriptional activation of c-Myc in prostate cancer. *Oncotarget* 2016; 7: 12869-84.
- [58] Zhang Z, Hou X, Shao C, Li J, Cheng JX, Kuang S, Ahmad N, Ratliff T, Liu X. PIK1 inhibition enhances the efficacy of androgen signaling blockade in castration-resistant prostate cancer. *Cancer Res* 2014; 74: 6635-47.
- [59] Pan Z, Zheng W, Zhang J, Gao R, Li D, Guo X, Han H, Li F, Qu S, Shao R. Down-regulation of the expression of CCAAT/enhancer binding protein alpha gene in cervical squamous cell carcinoma. *BMC Cancer* 2014; 14: 417.
- [60] Guo D, Reinitz F, Youssef M, Hong C, Nathanson D, Akhavan D, Kuga D, Amzajerdi AN, Soto H, Zhu S, Babic I, Tanaka K, Dang J, Iwanami A, Gini B, Dejesus J, Lisiero DD, Huang TT, Prins RM, Wen PY, Robins HI, Prados MD, Deangelis LM, Mellinghoff IK, Mehta MP, James CD, Chakravarti A, Cloughesy TF, Tontonoz P, Mischel PS. An LXR agonist promotes glioblastoma cell death through inhibition of an EGFR/AKT/SREBP-1/LDLR-dependent pathway. *Cancer Discov* 2011; 1: 442-56.
- [61] Li X, Chen YT, Hu P, Huang WC. Fatostatin displays high antitumor activity in prostate cancer by blocking SREBP-regulated metabolic pathways and androgen receptor signaling. *Mol Cancer Ther* 2014; 13: 855-66.
- [62] Yamauchi Y, Furukawa K, Hamamura K, Furukawa K. Positive feedback loop between PI3K-Akt-mTORC1 signaling and the lipogenic pathway boosts Akt signaling: induction of the lipogenic pathway by a melanoma antigen. *Cancer Res* 2011; 71: 4989-97.
- [63] Nakamura Y, Suzuki T, Sugawara A, Arai Y, Sasano H. Peroxisome proliferator-activated receptor gamma in human prostate carcinoma. *Pathol Int* 2009; 59: 288-93.
- [64] Rogenhofer S, Ellinger J, Kahl P, Stoehr C, Hartmann A, Engehausen D, Wieland WF, Müller SC, Hofstädter F, Walter B. Enhanced expression of peroxisome proliferator-activated receptor gamma (PPAR-gamma) in advanced prostate cancer. *Anticancer Res* 2012; 32: 3479-83.
- [65] Forootan FS, Forootan SS, Malki MI, Chen D, Li G, Lin K, Rudland PS, Foster CS, Ke Y. The expression of C-FABP and PPARgamma and their prognostic significance in prostate cancer. *Int J Oncol* 2014; 44: 265-75.
- [66] Ahmad I, Mui E, Galbraith L, Patel R, Tan EH, Salji M, Rust AG, Repiscak P, Hedley A, Markert E, Loveridge C, van der Weyden L, Edwards J, Sansom OJ, Adams DJ, Leung HY. Sleeping Beauty screen reveals Pparg activation in metastatic prostate cancer. *Proc Natl Acad Sci U S A* 2016; 113: 8290-5.
- [67] Olokpa E, Bolden A and Stewart LV. The androgen receptor regulates PPARgamma expression and activity in human prostate cancer cells. *J Cell Physiol* 2016; 231: 2664-72.
- [68] Lin LC, Gao AC, Lai CH, Hsieh JT, Lin H. Induction of neuroendocrine differentiation in castration resistant prostate cancer cells by adipocyte differentiation-related protein (ADRP) delivered by exosomes. *Cancer Lett* 2017; 391: 74-82.
- [69] Villena JA. New insights into PGC-1 coactivators: redefining their role in the regulation of mitochondrial function and beyond. *FEBS J* 2015; 282: 647-72.
- [70] Tan Z, Luo X, Xiao L, Tang M, Bode AM, Dong Z, Cao Y. The role of PGC1alpha in cancer metabolism and its therapeutic implications. *Mol Cancer Ther* 2016; 15: 774-82.
- [71] Ranhotra HS. Estrogen-related receptor alpha and mitochondria: tale of the titans. *J Recept Signal Transduct Res* 2015; 35: 386-90.
- [72] Tennakoon JB, Shi Y, Han JJ, Tsouko E, White MA, Burns AR, Zhang A, Xia X, Ilkayeva OR, Xin L, Ittmann MM, Rick FG, Schally AV, Frigo DE. Androgens regulate prostate cancer cell growth via an AMPK-PGC-1alpha-mediated metabolic switch. *Oncogene* 2014; 33: 5251-61.
- [73] Smith J. Human Sir2 and the 'silencing' of p53 activity. *Trends Cell Biol* 2002; 12: 404-6.
- [74] Bordone L and Guarente L. Calorie restriction, SIRT1 and metabolism: understanding longevity. *Nat Rev Mol Cell Biol* 2005; 6: 298-305.
- [75] Giannakou ME and Partridge L. The interaction between FOXO and SIRT1: tipping the balance towards survival. *Trends Cell Biol* 2004; 14: 408-12.
- [76] Guarente L. Sirtuins in aging and disease. *Cold Spring Harb Symp Quant Biol* 2007; 72: 483-8.
- [77] Guarente L and Picard F. Calorie restriction-the SIR2 connection. *Cell* 2005; 120: 473-82.
- [78] Herranz D, Maraver A, Cañamero M, Gómez-López G, Inglada-Pérez L, Robledo M, Castellblanco E, Matias-Guiu X, Serrano M. SIRT1 promotes thyroid carcinogenesis driven by PTEN deficiency. *Oncogene* 2013; 32: 4052-6.
- [79] Frazzi R, Valli R, Tamagnini I, Casali B, Latruffe N, Merli F. Resveratrol-mediated apoptosis of hodgkin lymphoma cells involves SIRT1 inhibition and FOXO3a hyperacetylation. *Int J Cancer* 2013; 132: 1013-21.
- [80] Ma CH, Chiua YC, Wu CH, Jou IM, Tu YK, Hung CH, Hsieh PL, Tsai KL. Homocysteine causes dysfunction of chondrocytes and oxidative stress through repression of SIRT1/AMPK

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- pathway: a possible link between hyperhomocysteinemia and osteoarthritis. *Redox Biol* 2018; 15: 504-512.
- [81] Sun T, Jiao L, Wang Y, Yu Y, Ming L. SIRT1 induces epithelial-mesenchymal transition by promoting autophagic degradation of E-cadherin in melanoma cells. *Cell Death Dis* 2018; 9: 136.
- [82] Lv J, Jiang S, Yang Z, Hu W, Wang Z, Li T, Yang Y. PGC-1 α sparks the fire of neuroprotection against neurodegenerative disorders. *Ageing Res Rev* 2018; 44: 8-21.
- [83] Byles V, Zhu L, Lovaas JD, Chmielewski LK, Wang J, Faller DV, Dai Y. SIRT1 induces EMT by cooperating with EMT transcription factors and enhances prostate cancer cell migration and metastasis. *Oncogene* 2012; 31: 4619-29.
- [84] Hardie DG. AMP-activated/SNF1 protein kinases: conserved guardians of cellular energy. *Nat Rev Mol Cell Biol* 2007; 8: 774-85.
- [85] Habegger KM, Penque BA, Sealls W, Tackett L, Bell LN, Blue EK, Gallagher PJ, Sturek M, Al-loosh MA, Steinberg HO, Considine RV, Elmen-dorf JS. Fat-induced membrane cholesterol accrual provokes cortical filamentous actin destabilisation and glucose transport dysfunction in skeletal muscle. *Diabetologia* 2012; 55: 457-67.
- [86] Herzig S and Shaw RJ. AMPK: guardian of metabolism and mitochondrial homeostasis. *Nat Rev Mol Cell Biol* 2018; 19: 121-135.
- [87] Zadra G, Photopoulos C, Tyekucheva S, Heidari P, Weng QP, Fedele G, Liu H, Scaglia N, Priolo C, Sicinska E, Mahmood U, Signoretti S, Birnberg N, Loda M. A novel direct activator of AMPK inhibits prostate cancer growth by blocking lipogenesis. *EMBO Mol Med* 2014; 6: 519-38.
- [88] Park HU, Suy S, Danner M, Dailey V, Zhang Y, Li H, Hyduke DR, Collins BT, Gagnon G, Kallakury B, Kumar D, Brown ML, Fornace A, Dritschilo A, Collins SP. AMP-activated protein kinase promotes human prostate cancer cell growth and survival. *Mol Cancer Ther* 2009; 8: 733-41.
- [89] Choudhury Y, Yang Z, Ahmad I, Nixon C, Salt IP, Leung HY. AMP-activated protein kinase (AMPK) as a potential therapeutic target independent of PI3K/Akt signaling in prostate cancer. *Oncoscience* 2014; 1: 446-56.