# Review Article Extracellular vesicles in Inter-Kingdom communication in gastrointestinal cancer

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Abstract: The production and secretion of extracellular vesicles (EVs) are common features of cells (including various normal cells, neoplastic cell lines as well as bacteria) that span all domains of life. Tumor-derived exosomes are enriched with kinds of tumorigenesis mediators which are derived from the cytoplasm of cancer cells and fully reflect the tumor conditions. Indeed, the major topics and challenges on current oncological research are the identification of tumorigenic and metastatic molecules in tumor-cell-derived exosomes as well as elucidating the pathways that guarantee these components to be included in exosomes. The bacterial EVs have also been implicated in the pathogenesis of gastrointestinal (GI) tumors and chronic inflammatory diseases; however, the possible function of outer membrane vesicles (OMVs) in tumorigenesis remains largely underestimated. We suggest that EVs from both eukaryotic cells and different microbes in GI tract act as regulators of intracellular and cross-species communication, thus particularly facilitate tumor cell survival and multi-drug resistance. Therefore, our review introduces comprehensive knowledge on the promising role of EVs (mainly exosomes and OMVs) production of GI cancer development and gut microbiome, as well as its roles in developing novel therapeutic strategies.

Keywords: Extracellular vesicles, exosomes, gastrointestinal cancer, gut microbiome

#### Introduction

EVs can be divided into three types according to the different molecular sizes and release modes, namely exosomes, microvesicles, and apoptotic bodies [1, 2]. Exosomes are small monolayer secretory organelles, 30-200 nm in diameter, with the same topological structure as cells and rich in selected nucleic acids, proteins, lipids, and glycoconjugates. The molecular diameter of microvesicles is 100-1000 nm. and they are produced directly from the surface of the mother cell membrane by budding. They are heterogeneous in size and highly express phosphatidylserine. They do not have specific surface molecular markers, but like exosomes, they express surface markers derived from the mother cell. Bacteria, on the other hand, release membrane sacs (MVs), with diameters ranging from 20 to 400 nm, that affect a variety of biological processes (including virulence factor transport, DNA transfer, phage interception, antibiotic and eukaryotic host defense factors, cell detoxification, cell metabolite output, and cell-to-cell communication). It was first found that MVs are produced through controlled vesicles of the outer membrane of G-bacteria, so it is often called outer-membrane vesicles (OMV). In addition to OMVs, other types of MVs have been recently discovered, including outer-inner membrane vesicles (OIMV), cytoplasmic membrane vesicles (CMV), and tube-shaped membranous structures (TSMS). MVs can also be formed by cell lysis triggered by lysin in phages. Unfortunately, many studies have confused exosomes with EVs. The EVs discussed in this paper are mainly exosomes and OMVs, which are also the focus of EV research at present.

Ten years ago, due to the availability of nextgeneration-sequencing (NGS) technologies, we come to realize that we harbor 'another' genome (namely, the microbiome) [3, 4]. The significance of the gut microbiome is self-evident since an imbalance of microbiota leads healthy individuals to physiological disorders and even tumorigenesis [5, 6].

There are growing evidence of EV-mediated "guest/host dialogue" from gut microbiota, opening the way for other interesting findings. Bacteria, both G+ and G- bacteria, produce EVs in much the similar way that our human cells produce EVs. Now that we have known more about our "other genome", we think it's time to decide how to interact with it.

## Characteristics of different extracellular vesicles

Molecular structure and composition of EVs

Evidently, EVs can be found in almost all living cells [1, 2], demonstrating that EVs are highly evolutionarily conserved as a ubiquitous communication pattern among species.

According to the online exosome database (www.exocarta.org), the most recent update lists 3,408 mRNAs, 2,838 miRNAs, 9,769 proteins, and 1,116 lipids [7] (Figure 1). Exosomes are heterogeneous in composition and size, and enriched in membrane-associated protein complexes. What exosomes carry depends on the functions and states of the original cell types [8-12]. The distinct heterogeneity of exosomes is due to their limited load capacity, the mechanistic forces that lead to the variant protein distribution and the differential gene expression. The secretion of exosomal DNA may facilitate DNA quality control, regulation of inflammation, and perhaps may play a powerful role in tumor biomarkers or chemotherapeutic resistance. Exosomal RNA contains non-coding RNAs (ncRNAs). However, most studies focus only on the exosomal overall RNA composition, not single-particle levels, and thus may have underestimated the actual complexity of RNAs in exosomes [13].

OMVs are mainly composed of proteins, lipids and various pathogen-associated molecular patterns (PAMPs) [14, 15]. As far as the proteins in OMVs are concerned, most of the proteins are virus-related factors, such as enzymes, molecular chaperones, toxins, etc. [16-18]. The composition of OMVs protein showed

great heterogeneity among different strains [19]. Phospholipids and LPS are the main lipids contained in OMVs. Moreover, other PAMPs, including peptidoglycan, lipoprotein, DNA and RNA, are also presented in OMVs. The co-existence of bacterial antigens and abundant PAMPs endows OMVs the potential for superior vaccines

Biogenesis, secretion, and release of EVs

Exosomes are produced by vesicle budding into the endosomal membrane, with subsequent accumulation and fusion at large multivesicular bodies (MVBs) [20-22]. However, the presence of an endosomal pattern of exosome biogenesis does not imply that all exosomes are produced by endosome budding alone. In fact, multiple evidences suggest that exosomes also germinate from the plasma membrane [23-27] (Figure 1). Unfortunately, this pattern is largely overlooked by the majority cartoon models of exosome biogenesis, while the view of the endodermal body of exosome biogenesis is widely accepted [28, 29]. Stephen J. Gould considers this may be due to observational bias [30].

Besides, the exosomal secretion mechanism has also been extensively studied. Ras associated proteins of the Rab family are regarded as important modulators in exosomal secretion pathways [31, 32]. Exosomes can communicate with their recipient cells by sending signals directly through the interaction of receptor molecules or ligand on their respective surfaces.

For bacteria and archaea, EVs are released outward from their membrane, whereas eukaryotes can also produce EVs from the endocytic pathway and release them through the multivesicles [33]. Up to now, EV release in eukaryotes and archaea has been understood to be mediated by ESCRT-related proteins and homologues. However, the biogenic mechanisms of bacterial EVs are largely vague, and many scholars speculate that there may be more diverse mechanisms [34]. It is noteworthy that the number of EVs generated by G+ bacteria is naturally lower than that of G- bacteria. Our understanding of the biological origin and composition of the bacterial EVs is primarily from the research of the OMVs of G-bacterial EVs (Figure 2).

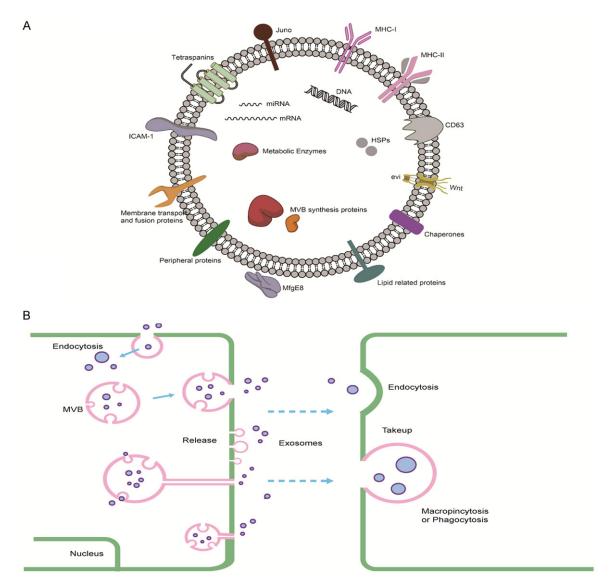


Figure 1. Composition, structure, biogenesis, and uptake of exosomes. A. Exosomes are heterogeneous in size and composition and are rich in protein complexes, DNA, RNA molecules, etc. B. Exosomes are produced in three ways (left): (a) After endocytosis, scattered endosomes develop into mature MVB, which then fuse with the cell membrane and release exosomes; (b) The plasma membrane secretes exosomes directly; (c) The intracellular-plasma-membrane connected compartments (IPMCs) bud out and subsequently release exosomes through the IPMC necks. After exosomes are released, they can interact with recipient cells through ligand/receptor signaling. Exosomes can enter recipient cells in different ways (right), such as endocytosis, direct membrane fusion, micropinocytosis, and even phagocytosis. HSP, heat shock protein; ICAM-1, intracellular, adhesion molecule-1; MfgE8, milk fat globule protein E8; Wnt proteins, wingless proteins.

#### Purification and identification of EVs

To understand the physiological and pathological functions of EVs, the purification, identification, and quantitative analysis of EVs are the basis of basic research and clinical application. Based on some specific characteristics of EVs, such as their morphology, density, size, or surface protein, there are generally five types of EV

isolation methods (**Table 1**), among which the UC technique is the most traditional and widely recognized method [35-37]. However, there is no single protocol that is universally applicable for the analysis of various body fluids, such as saliva, plasma and feces. The isolation method must base on the complexity of the samples (e.g., composition and sample size). Bacterial EVs in human body fluids has rarely been stud-

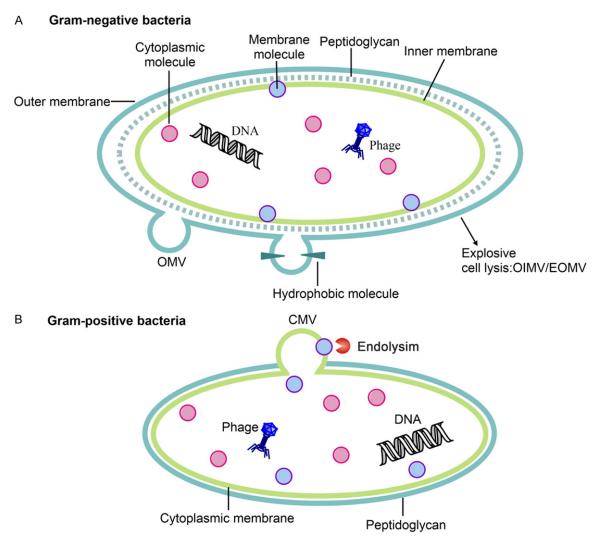


Figure 2. Different routine of bacterial membrane vesicles. A. There are two main ways for G- bacteria to form vesicles: the blebbing and formation of OMVs, or explosive cell lysis, producing OIMVs or EOMVs. B. The bubbling of G+ bacteria leads to the formation of cytoplasmic CMVs. The OMVs of G- bacteria harbor an internal phospholipid lobule and an external LPS lobule, which activates immune cells through TLR-4. At present, our understanding of the composition, molecular structure, and function of OMVs mainly comes from bacteria cultured in the lab. OIMVs: outer-inner membrane vesicles; EOMVs: explosive outer-membrane vesicles; LPS: lipopolysaccharide. TLR-4: toll-like receptor 4.

ied, possibly due to methodological challenges in isolating them from stromal- and host-related eukaryotic EVs, such as human exosomes and micro-vesicles [38, 39].

## EVs derived from gastrointestinal tumors and gut microbiota

EVs derived from gastrointestinal tumors

There have been many studies focusing on the role of exosomes in GI cancer tumorigenesis, progress, and metastasis. Zhou et al. [40] stud-

ied patients with different types of esophageal diseases, and reported two kinds of exosome miRNAs (miR-223-3p and miR-584), which can be satisfactorily used for the detection of esophageal squamous cell carcinoma. Exosomes loading mutant p53 DNA were separated from the serum of patients with pancreatic cancer, which contribute to the diagnosis, treatment of cancer [41]. Costa Silva b. et al. confirmed that macrophage migration inhibitor (MIF) was elevated in PDAC-derived exosomes from stage I PDAC patients and can block the formation and metastasis of pre-liver niches.

### EV in gastrointestinal cancers and its application

Table 1. Isolation methods of EVs

Isolation method	Principle	Advantages	Disadvantages
Ultracentrifugation	Particle density and size	Simple procedure and good extract-uniformity	Limited efficiency and purity
Ultrafiltration	Molecular size and morphology	Simple extraction process, high uniformity, and tiny effect on the bio-activity of EVs	Limited product loss and purity
Density-gradient centrifugation	Density	High purity	Time-consuming and multi-step process
Protein precipitation	Ammonium sulfate precipitates proteins	Economical and is promising for large sample separation	Time-consuming, low separation efficiency and purity
Immunoaffinity capture	The interaction of the molecule with a specific ligand	High purity	Expensive and not suitable for mass extraction of OMVs

Table 2. Comparison of bacterial and human EVs application

	EVs in human (mainly exosomes)	Bacterial EVs (mainly OMVs)
Application fields	Mediate ECM regulation, signaling and molecular transfer	Vaccine (bacterial OMVs are promising vaccines induce humoral and cellular immune responses humans and animals)
	Mediate signaling and molecular transfer	Adjuvants that can enhance and regulate immune responses to specific antigens
	A noninvasive and efficient approach for tumor diagnosis and prognostic monitoring	Cancer immunotherapy drugs to target tumor tissues
	Activate the immune response to enhance immunity	As a carrier of chemotherapy drugs, OMVs increase the accumulation of chemotherapy drugs in tumors
	Exosomes serves as delivery systems for drug therapy	OMVs is a bacterial mimic that competitively inhibits parental pathogen attacks on host cells

These results suggest that MIF derived from PDAC exosomes may be an effective prognostic marker for the occurrence of PDAC liver metastases [42]. It is worth noting that EVs in quantitative peritoneal lavage fluid (PLF) may have an advantage over blood exosomal levels. Tokuhisa et al. recently identified the expression of miR-21 and miR-1225-5p in PLF exosomes and suggested that they could indicate the metastatic stage of gastric cancer (GC) [43]. In addition, the levels of these two miRNAs in T4 patients were higher than those in T1-T3 patients. GC diagnostic tests will probably include the detection of EVs in gastric fluid in the future [44]. These data deserve further investigation, and the sensitivity and specificity of the research should be validated by large and multicenter studies. The findings of Herrera m. et al. suggest that exosome-loaded ncRNAs are potential biomarkers of CRC, while CAF (cancer-associated fibroblasts)-derived exosomes are specific communication mediators between CAFs and colon cancer cells [45]. Exosomes-derived miR-10b from CRC microenvironment cells, such as CAFs, can also accelerate proliferation and promote tumor progression by influencing stromal cells. These CAFs have been proved to enhance CRC growth in vitro and in vivo [46]. Some studies demonstrated that after the p53 R273H mutation was improved, exosomal miR-21-3p and miR-769-3p can motivate the activation of fibroblasts in lung tissues and its tumor microenvironment. Furthermore, they demonstrate that this process consolidates the formation of CRC pre-metastatic niches and enhances its lung metastases [47]. Cook T. et al. observed that exosomes rich in miR-1246 can be selectively released by CRC cells carrying mutant p53. Derived from unique mutant 53 exosomes, miR-1246 may be suitable for the treatment and diagnosis of CRC. They discovered that the ingestion of mutated p53-derived exosomes activated the formation of a tumorrelated macrophage subgroup that is involved in tumor progression and metastasis [48]. Another notable molecule is heat shock protein 60 (HSP60). After surgical resection of CRC tumor, the level of HSP60 in exosomes is reduced, so this assay can be applied to monitor treatment response [49]. It should be emphasized that most of the researchers believe that the expression of exosomal antigens in the blood of GI cancer patients is increased and are associated with patient survival and reflect disease severity. Zhou J. and

colleagues proved that miRNA-21 from hepatocellular carcinoma (HCC) cell exosomes was able to significantly transform normal hepatic stellate cells (HSCs) into CAFs [50]. Also, exosomes can regulate Wnt pathway receptors by influencing stem cell-associated signaling pathways [51] to regenerate the phenotypes of stem cells and transform them into tumor stem cells. Recently, the function of both FZD10 and FZD10-mRNA which were reported to be associated with GI cancer cells were investigated. The results demonstrated that FZD10 and exosomal FZD10-mRNA may be potential deliverers of cell transformation in the distant metastasis process [52]. Other studies suggest that exosomes containing FZD10 were involved in controlling cancer progression and cancer cell modification and can be an indicator of the pathological condition [53]. Notably, the panel of exosome molecules and CEA may increase the efficacy of CRC diagnosis. In addition, some research has shown that exosomes have the potential to be biomarkers for the early diagnosis of CRC [54, 55]. The oncogenic miRNA-203 in CRC exosomes may enhance liver metastasis by eliciting the activation of tumor-associated macrophages [55]. Additionally, a report demonstrated that miR-25-3p supports liver and lung metastasis via enhancing vascular permeability and promoting angiogenesis, and is associated with the establishment of premetastasis niches in vivo [56]. Importantly, EVs is involved in GI tumor genesis and development as a drug resistance modulator. Exosomal miR-155-5p is associated with paclitaxel resistance in GC [57]. Bhome et al. [58] demonstrated that miR-21 is transferred to CRC cells via fibroblast exosomes, leading to oxaliplatin resistance in these cells.

However, due to the differences in the approaches of separation and purification, sample size, sample population, sample types, and other experimental conditions, the results of different experiments are still lack repeatability and consistency. Stratified analysis indicators such as age, sex, and race may be important for the identification of diagnostic biomarkers, however, few investigations have focused on these possible factors.

#### EVs derived from gut microbiota

Harboring about  $3 \times 10^{13}$  bacteria, the GI tract is guarded by epithelial cells, which are main-

tained by constant interaction between the GI microbiota, the mucosal barrier, and immune cells [59]. Dysregulation of gut microbiota has been shown to occur in a variety of tumors, particularly in the GI tract [60-64]. In different correlation and mechanism studies, Bacteroides fragilis, Enterococcus faecalis, Escherichia coli and Streptococcus gallolyticus, etc. were reported to be individually associated with CRC [65-68]. For example, the fecal abundance of Enterococcus, Fusobacteria, Escherichia coli, and Streptococcus in CRC patients is abnormal [69, 70]. Viljoen etc. al. [71] found that, compared with healthy individuals, the number of Fusobacterium spp. and Bacteroides fragilis were significantly higher in patients with advanced CRC. Fusobacterium spp. does not have direct carcinogenicity, but it may indirectly lead to tumorigenicity by enhancing inflammatory response and promoting the proliferation of tumor cells [72]. Fusobacterium spps. can activate the adhesion of FadA, trigger the Wnt signaling pathway of colonic epithelial cells, and promote the proliferation of epithelial cells. Another important participant, enterotoxigenic Bacteroides fragilis (ETBF), produces B. fragilis toxin (BFT), which plays a significant role in colorectal cancer, diarrhea, and other diseases [73].

The OMVs/MVs production of G- and G+ bacteria are one of the main mechanisms of the gastrointestinal microbiota [74]. OMVs are about 40-300 nm in size and are part of the bacterial release and transport system that transfer their cargos such as nucleic acids, proteins, etc. [75] to other bacteria, fungi, or host cells. OMVs are also rich in bioactive components such as immune-regulating lipopolysaccharides, lipoproteins, peptidoglycans, etc. These molecules have been reported to exist in a variety of G-bacteria in the human GI tract, including Escherichia coli, Fusobacterium nucleatum, Helicobacter pilori, B. fragilis and others [75-77]. Bacteroides fragilis can secrete OMVs to deliver immune molecules to human immune cells. These secreted OMVs and IBD (Inflammatory bowel disease, IBD)-related genes (ATG16L1 and NOD2) play an important protective role in inflammatory bowel diseases that activate non-classical autophagy pathways [78]. Recently, researchers at New York University have found that pancreatic ductal adenocarcinoma (PDA) tumors contain many fungi that originate in the gut and have shown that these fungi can induce normal cells to turn into PDA. It also suggests that an anti-fungal approach may be a promising treatment for pancreatic cancer [79]. OMVs also play a critical role in the pathogenesis of chronic inflammation of GI tract, including Crohn's disease (CD) [80] and Hp. associated inflammation [81].

In addition to the secretion of carcinogenic toxic components and the direct effect of specific microorganisms on the host, microorganisms may also play a critical role through inflammatory and metabolism-associated pathways. Notably, the occurrence of GI tumors is caused by microorganisms, inflammation, and intestinal immune regulation. Among them, the pathogenicity of microbial EVs cannot be ignored. This intercellular transport medium can induce continuous inflammation and even cancer, leading to the establishment of the original tumor environment.

#### EVs communicate across boundaries

Interspecies and even inter-boundary communication and interaction occur continuously in the GI tract. EVs of different origins (host eukaryotic cells/pathogenic cells, fungi, viruses, worm and edible plants [82, 83], etc.) meet in the three-dimensional space of the gastrointestinal cavity and interact with intestinal cells. EVs can be secreted by many human parasites, which play a significant role in stimulating and sustaining parasite infections [84]. Bacterial DNA integration and associated mutations through horizontal gene transfer are presented in cancer cells [85]. As a container of nucleic acid molecules, EVs have the potential to be carriers of such transfer. In addition, this transportation enables protein epitopes to be shared between self-antigens and microbial molecules, resulting in a cross-reaction that leads to tissue destruction, apoptosis, and the accompanying expression of self-antigens and microbe antigens [86]. We hypothesized that the EVs released by the GI cancer cells and the gut microbes might have a certain level of matching protein sequences. Barteneva et al. [87]. compared the protein sequences of the colorectal EV protein group [88] and different symbiotic bacteria and viruses studied by Choi and colleagues, and found a large number of matching protein sequences. Recently, the

authors [89] analyzed the gut microbes and related human proteins in the pediatric IBD population and identified the characteristics of new host-microbial interaction (including microbial metabolism). In addition, the intestinal mucosa of patients with IBD can secrete EVs containing host defense proteins which, when ingested by microorganisms, will cause the response of microbial defense stress and functional adaptation, leading to the imbalance of intestinal microbiota and the subsequent development of mucosal inflammation. Moreover, CRC derived EVs can influence symbiotic bacteria in the GI microenvironment. These novel analytical approaches contribute to the understanding of overall composition and similarities between bacteria and human eukaryotic proteins, as well as their functions.

Intriguing, in addition to bacteria, fungus and protozoa, plant-derived exosome-like nanoparticles (ELNs) also play a role in disease development. Recently, Yun Teng and Huang-ge Zhang et al. [90] found that small RNAs in ginger ELNs can affect GI microbiota, thus improving intestinal barrier function and alleviating colitis in mice.

Malignancies include not only tumor cells with genetic and phenotypic heterogeneity, but also heterogeneous healthy cell populations involved in the anti-tumor immune response that forms a specific extracellular matrix and its EVs guarantee tumor evolution and development [91]. Another key aspect of tumor evolution is the epigenetic regulation at the gene transcription level that influences proliferation, differentiation, and the fate of tumor cells. The initial strategy of using abnormal methylation to diagnose tumors is challenging since chronic inflammation can also lead to changes in gene methylation expression levels [92, 93].

#### Clinical application of EVs

Multiple cell types [94, 95] have been applied as donor cells for EVs production, including autologous ascites, autologous monocytes, bacteria, and worms, among others (**Table 2**). Mesenchymal stem/stromal cells derived EVs (MSC-EVs) is promising and effective in animal models of 30 human diseases. However, several key issues including the efficacy, safety, characteristics, and heterogeneity of MSC-EVs need to be seriously addressed before they can

be successfully converted to clinical practice [96]. Some institutions have proposed several indicators to define the potential of MSC-S EVs to treat COVID-19 [97, 98], which clinical trials are urgently needed to assess and confirm the potential. EV from non-MSC cell sources have also been reported and are in ongoing preclinical studies [99]. EVs can be loaded with antisense oligonucleotides (ASOs) or Cas9 mRNA and gRNA and can deliver them to cancer cells [100]. RBC-EVs can be absorbed by leukemia cells and has the advantages of high efficiency and low cytotoxicity. In xenograft mouse models, ASO-loaded RBC-EVs can effectively knock out miR-125b and inhibit the development of leukemia or breast cancer, demonstrating the potential of EV for the treatment of cancers. EVs can be utilized as carriers for anti-tumor drugs, small RNAs, and anti-inflammatory therapeutic agents. A series of studies have shown that EVs from different sources can cross tissue barriers, and carry cargos and deliver them to target cells [101-103]. Professor Robert Blelloch's team found that exosomes secreted by prostate cancer may enter the draining lymph nodes and spleen of tumors and suppress immune cells. They also found that inhibiting the formation and release of tumor exosomes eliminates the resistance of many tumors to PD-L1 inhibitors, and allows the immune system to form a long-term immune memory of the tumor that acts as a tumor vaccine [31]. EVs carrying pathogen-specific antigens can be used as new vaccines for human and animal infectious diseases. Parasitederived EVs modifies intestinal inflammatory molecular (including cytokines and signaling molecules) in preclinical mouse models [104]. In a preclinical model of inflammatory bowel disease (IBD) in mice, EV secreted by worm parasites is beneficial in the inflammatory response, resulting in a substantial reduction of pro-inflammatory cytokines [105, 106]. However, due to the large number of active macromolecules carried by parasitic EV, the therapeutic use of parasitic EVs needs to be thoroughly evaluated.

However, there is still a lot of room for further development of targeted strategies, we can improve the targeting efficiency by engineering the surface molecules. Adhering to the highest standards of EVs separation and production is critical for clinical applications. The European

Medicines Agency (EMA) and the US Food and Drug Administration (FDA) have published guidelines covering the manufacture and clinical evaluation of novel EVs therapeutic agents. In addition to general requirements such as EVs isolation and storage, safety and efficacy evaluation of EVs-based therapies are also required. Besides, the safety of donors and recipients is also an important issue to consider. Therefore, EVs must be manufactured per Good Practice principles aimed at ensuring product safety, meeting its intended use, and following quality control processes during manufacturing, monitoring, storage, and distribution. In the future, we should provide advice to research institutions and clinical trial sponsors at the national or international level, which will facilitate interdisciplinary collaboration between academia and industry and accelerate the success of preclinical development and clinical transformation (Figure 3).

#### Conclusion and future outlooks

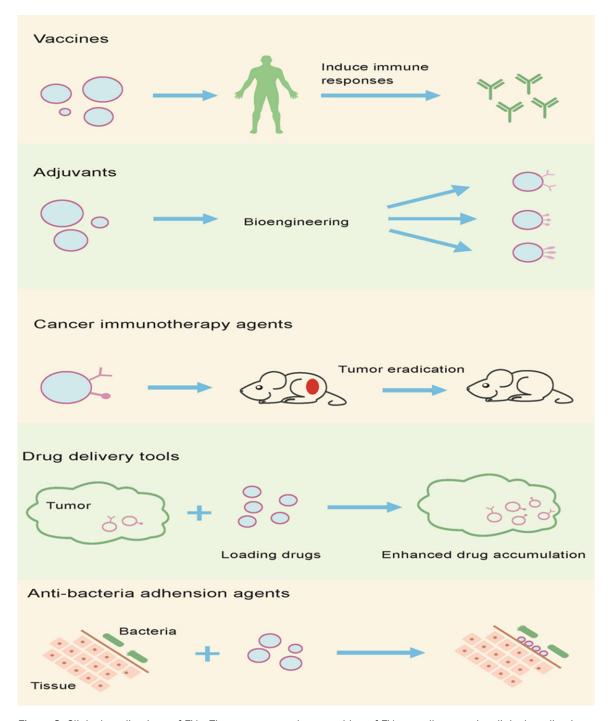
Over the years, the field of EVs has made considerable epoch-making and milestone progress, revealing the immeasurable role of EVs in microbe-host and microbe-microbe cross-border communication, but it is still a daunting task to recognize and translate various research results into clinical practice.

Other critical issues in EVs biogenesis and delivery

Although the basic framework of EVs has been understood over the past decade or so, it is not deep and comprehensive enough. (a) The clear function of ESCRT proteins in EV biogenesis is unclear; (b) Due to the lack of endogenous control to standardize the expression of exosomal nucleic acids and the contradictory results of various studies, its clinical application remains limited. (c) Intensively investigating the physical and chemical properties of EVs, as well as the positive and negative regulatory factors involved in the occurrence of EVs; (d) Explaining the protein-protein interaction networks involved in EV biological processes; (e) Determining the delivery mechanisms that mediate the interactions between bioactive proteins, nucleic acids and lipids in the host and microbial EVs and target cells. (f) Studying the changes of host-microbial EVs composition and function in both health and disease conditions, and mapping their flow through the body-fluids. Meanwhile, we also need new experimental methods and techniques to assist us in studying the composition and biogenesis of EVs at single-vesicle and single-cell levels. These advances will accelerate our comprehension of EVs and facilitate clinical translation of EVs into diagnosis and therapies (Figure 3).

Challenges in the application of EVs in tumor therapy

Firstly, since EV can be used as clinical diagnostic and prognostic biomarkers, vaccines, or drug delivery devices, more precise and standardized isolation and purification methods are urgently needed. Secondly, the loading efficiency of exosome antigen should be improved in order to improve the odds of immunotherapy or exosome-based immunization. Thirdly, it is of importance how to massively produce EVs in clinical application. Currently, EVs of good manufacturing practice grade have been reported from certain cells (such as MSCs) [107], and in the future, we will need to extract EVs from more different cell types and identify which cell types are best suited to produce clinically grade EVs. Besides, the combination of EVs and other anti-tumor therapies has a broad and promising application prospect and may contribute to making a breakthrough in the bottleneck of tumor treatment. Recently, EVs have been reported as siRNA delivery carriers to silence oncogenes in cancer cells [108]. The activation of GTPase KRAS is very common in pancreatic cancer [109, 110]. Due to the low stability and uncontrollability of KRAS-targeted nucleic acids, it has become a difficult problem to promote efficacy by targeting KRAS. Delivery of KRAS siRNA using EVs from normal human prepuce fibroblasts has been reported to be highly efficient, significantly inhibiting pancreatic tumor progression and improving overall survival in mice [111]. Shortly, immune-cellderived EVs will be applied in cancer immunotherapy. For example, NK cell EVs have cytotoxic effects on different human tumor cells. Activated EVs of CD8+ cells can also deplete the mesenchymal cells in the mesenchymal tumors and inhibit cancer progression [112]. At present, liquid biopsy, a non-invasive and simple method, acts as an important biomarker for tumor diagnosis and prognosis evaluation. EVs have sufficient concentration and high stability in human circulation, thus they have obvious advantages over other liquid biopsy methods



**Figure 3.** Clinical applications of EVs. The structure and composition of EVs contributes to its clinical applications. Multiple proteins, PAMPs, lipids and other EV molecules make it promising vaccines, cancer immunotherapy agents, adjuvants, drug delivery tools and anti-bacteria agents and endow it broad prospects.

[113]. Also, EV contains a variety of cargos, such as proteins and nucleic acids (miRNAs), which can be used as biomarkers for disease diagnosis [114, 115]. The GPC1+ EVs can be applied as potential non-invasive biomarkers to

detect early pancreatic cancer. Overwhelming studies have indicated that miRNAs in EVs contribute to immune regulation, drug resistance, and cancer metastasis of various tumor types [116-118]. In addition to miRNAs, circular RNAs

from cancer cells and patient serum may serve as novel biomarkers [119, 120].

In summarize, EVs from different sources have great potential for early cancer diagnosis, tracking chemotherapy drug resistance and therapeutic responses, and tailoring precision treatment strategies for patients with cancers. Nevertheless, there are many problems in EV field that need to be solved urgently, and it is worth us to further study its mechanism and carry out cross-field and cross-disciplinary research to enrich its application prospects.

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#### Disclosure of conflict of interest

None.

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#### References

- [1] Pegtel DM and Gould SJ. Exosomes. Annu Rev Biochem 2019; 88: 487-514.
- [2] Thery C, Ostrowski M and Segura E. Membrane vesicles as conveyors of immune responses. Nat Rev Immunol 2009; 9: 581-593.
- [3] Qin JJ, Li RQ, Raes J, Arumugam M, Burgdorf KS, Manichanh C, Nielsen T, Pons N, Levenez F, Yamada T, Mende DR, Li JH, Xu JM, Li SC, Li DF, Cao JJ, Wang B, Liang HQ, Zheng HS, Xie YL, Tap J, Lepage P, Bertalan M, Batto JM, Hansen T, Le Paslier D, Linneberg A, Nielsen HB, Pelletier E, Renault P, Sicheritz-Ponten T, Turner K, Zhu HM, Yu C, Li ST, Jian M, Zhou Y, Li YR, Zhang XQ, Li SG, Qin N, Yang HM, Wang J, Brunak S, Dore J, Guarner F, Kristiansen K, Pedersen O, Parkhill J, Weissenbach J, Bork P, Ehrlich SD, Wang J and Consortium M. A hu-

- man gut microbial gene catalogue established by metagenomic sequencing. Nature 2010; 464: 59-65.
- Nelson KE, Weinstock GM, Highlander SK, Worley KC. Creasy HH. Wortman JR. Rusch DB. Mitreva M, Sodergren E, Chinwalla AT, Feldgarden M, Gevers D, Haas BJ, Madupu R, Ward DV, Birren B, Gibbs RA, Methe B, Petrosino JF, Strausberg RL, Sutton GG, White OR, Wilson RK, Durkin S, Gujja S, Howarth C, Kodira CD, Kyrpides N, Madupu R, Mehta T, Mitreva M, Muzny DM, Pearson M, Pepin K, Pati A, Qin X, Yandava C, Zeng QD, Zhang L, Berlin AM, Chen L, Hepburn TA, Johnson J, McCorrison J, Miller J, Minx P, Nusbaum C, Russ C, Sutton GG, Sykes SM, Tomlinson CM, Young S, Warren WC, Badger J, Crabtree J, Madupu R, Markowitz VM, Orvis J, Rusch DB, Sutton GG, Cree A, Ferriera S, Gillis M, Hemphill LD, Joshi V, Kovar C, Wetterstrand KA, Abouellleil A, Wollam AM, Buhay CJ, Ding Y, Dugan S, Fulton LL, Fulton RS, Holder M. Hostetler J. Sutton GG, Allen-Vercoe E, Badger J, Clifton SW, Earl AM, Farmer CN, Giglio MG, Liolios K, Surette MG, Sutton GG, Torralba M, Xu Q, Pohl C, Durkin S, Sutton GG, Wilczek-Boney K, Zhu DH and Jumpstart HM. A catalog of reference genomes from the human microbiome. Science 2010; 328: 994-999.
- [5] Althani AA, Marei HE, Hamdi WS, Nasrallah GK, El Zowalaty ME, Al Khodor S, Al-Asmakh M, Abdel-Aziz H and Cenciarelli C. Human microbiome and its association with health and diseases. J Cell Physiol 2016; 231: 1688-1694.
- [6] Biedermann L and Rogler G. The intestinal microbiota: its role in health and disease. Eur J Pediatr 2015; 174: 151-167.
- [7] Mathivanan S, Fahner CJ, Reid GE and Simpson RJ. ExoCarta 2012: database of exosomal proteins, RNA and lipids. Nucleic Acids Res 2012; 40: D1241-1244.
- [8] Pisitkun T, Shen RF and Knepper MA. Identification and proteomic profiling of exosomes in human urine. Proc Natl Acad Sci U S A 2004; 101: 13368-13373.
- [9] Admyre C, Johansson SM, Qazi KR, Filen JJ, Lahesmaa R, Norman M, Neve EP, Scheynius A and Gabrielsson S. Exosomes with immune modulatory features are present in human breast milk. J Immunol 2007; 179: 1969-1978.
- [10] Michael A, Bajracharya SD, Yuen PS, Zhou H, Star RA, Illei GG and Alevizos I. Exosomes from human saliva as a source of microRNA biomarkers. Oral Dis 2010; 16: 34-38.
- [11] Zijlstra C and Stoorvogel W. Prostasomes as a source of diagnostic biomarkers for prostate cancer. J Clin Invest 2016; 126: 1144-1151.
- [12] Hiemstra TF, Charles PD, Gracia T, Hester SS, Gatto L, Al-Lamki R, Floto RA, Su Y, Skepper JN,

- Lilley KS and Karet Frankl FE. Human Urinary exosomes as innate immune effectors. J Am Soc Nephrol 2014; 25: 2017-2027.
- [13] Verweij FJ, Bebelman MP, Jimenez CR, Garcia-Vallejo JJ, Janssen H, Neefjes J, Knol JC, de Goeij-de Haas R, Piersma SR, Baglio SR, Verhage M, Middeldorp JM, Zomer A, van Rheenen J, Coppolino MG, Hurbain I, Raposo G, Smit MJ, Toonen RFG, van Niel G and Pegtel DM. Quantifying exosome secretion from single cells reveals a modulatory role for GPCR signaling (vol 217, pg 1129, 2018). J Cell Biol 2018; 217: 1129-1142.
- [14] Kaparakis-Liaskos M and Ferrero RL. Immune modulation by bacterial outer membrane vesicles. Nat Rev Immunol 2015; 15: 375-387.
- [15] Jan AT. Outer Membrane Vesicles (OMVs) of gram-negative bacteria: a perspective update. Front Microbiol 2017; 8: 1053.
- [16] Kulkarni HM and Jagannadham MV. Biogenesis and multifaceted roles of outer membrane vesicles from Gram-negative bacteria. Microbiology 2014; 160: 2109-2121.
- [17] Haurat MF, Aduse-Opoku J, Rangarajan M, Dorobantu L, Gray MR, Curtis MA and Feldman MF. Selective sorting of cargo proteins into bacterial membrane vesicles. J Biol Chem 2011; 286: 1269-1276.
- [18] Elhenawy W, Debelyy MO and Feldman MF. Preferential packing of acidic glycosidases and proteases into Bacteroides outer membrane vesicles. mBio 2014; 5: e00909-e00914.
- [19] Kikuchi Y, Obana N, Toyofuku M, Kodera N, Soma T, Ando T, Fukumori Y, Nomura N and Taoka A. Diversity of physical properties of bacterial extracellular membrane vesicles revealed through atomic force microscopy phase imaging. Nanoscale 2020; 12: 7950-7959.
- [20] Thery C, Zitvogel L and Amigorena S. Exosomes: composition, biogenesis and function. Nat Rev Immunol 2002; 2: 569-579.
- [21] van Niel G, D'Angelo G and Raposo G. Shedding light on the cell biology of extracellular vesicles. Nat Rev Mol Cell Biol 2018; 19: 213-228.
- [22] Mathieu M, Martin-Jaular L, Lavieu G and Thery C. Specificities of secretion and uptake of exosomes and other extracellular vesicles for cell-to-cell communication. Nat Cell Biol 2019; 21: 9-17.
- [23] Anderson HC, Garimella R and Tague SE. The role of matrix vesicles in growth plate development and biomineralization. Allergy Clin Immunol 2005; 10: 822-837.
- [24] Bianchi E, Doe B, Goulding D and Wright GJ. Juno is the egg Izumo receptor and is essential for mammalian fertilization. Nature 2014; 508: 483-7.

- [25] Booth AM, Fang Y, Fallon JK, Yang JM, Hildreth JEK and Gould SJ. Exosomes and HIV Gag bud from endosome-like domains of the T cell plasma membrane. J Cell Biol 2006; 172: 923-935
- [26] Fang Y, Wu N, Gan X, Yan WH, Morrell JC and Gould SJ. Higher-order oligomerization targets plasma membrane proteins and HIV gag to exosomes. PLoS Biol 2007; 5: 1267-1283.
- [27] Nager AR, Goldstein JS, Herranz-Perez V, Portran D, Ye F, Garcia-Verdugo JM and Nachury MV. An actin network dispatches ciliary GPCRs into extracellular vesicles to modulate signaling. Cell 2017; 168: 252-263, e14.
- [28] Egea-Jimenez AL and Zimmermann P. Phospholipase D and phosphatidic acid in the biogenesis and cargo loading of extracellular vesicles. J Lipid Res 2018; 59: 1554-1560.
- [29] Colombo M, Raposo G and Thery C. Biogenesis, secretion, and intercellular interactions of exosomes and other extracellular vesicles. Annu Rev Cell Dev Biol 2014; 30: 255-89.
- [30] Pegtel DM and Gould SJ. Exosomes. Annu Rev Biochem 2019; 88: 487-514.
- [31] Poggio M, Hu T, Pai CC, Chu B, Belair CD, Chang A, Montabana E, Lang UE, Fu Q, Fong L and Blelloch R. Suppression of exosomal PD-L1 induces systemic anti-tumor immunity and memory. Cell 2019; 177: 414-427, e13.
- [32] Ostrowski M, Carmo NB, Krumeich S, Fanget I, Raposo G, Savina A, Moita CF, Schauer K, Hume AN, Freitas RP, Goud B, Benaroch P, Hacohen N, Fukuda M, Desnos C, Seabra MC, Darchen F, Amigorena S, Moita LF and Thery C. Rab27a and Rab27b control different steps of the exosome secretion pathway. Nat Cell Biol 2010; 12: 19-30.
- [33] Yáñez-Mó M, Siljander PR, Andreu Z, Zavec AB, Borràs FE, Buzas EI, Buzas K, Casal E, Cappello F, Carvalho J, Colás E, Cordeiro-da Silva A, Fais S, Falcon-Perez JM, Ghobrial IM, Giebel B, Gimona M. Graner M. Gursel I. Gursel M. Heegaard NH, Hendrix A, Kierulf P, Kokubun K, Kosanovic M, Kralj-Iglic V, Krämer-Albers EM, Laitinen S, Lässer C, Lener T, Ligeti E, Linē A, Lipps G, Llorente A, Lötvall J, Manček-Keber M, Marcilla A, Mittelbrunn M, Nazarenko I, Nolte-'t Hoen EN, Nyman TA, O'Driscoll L, Olivan M, Oliveira C. Pállinger É. Del Portillo HA. Reventós J, Rigau M, Rohde E, Sammar M, Sánchez-Madrid F, Santarém N, Schallmoser K, Ostenfeld MS, Stoorvogel W, Stukelj R, Van der Grein SG, Vasconcelos MH, Wauben MH and De Wever O. Biological properties of extracellular vesicles and their physiological functions. J Extracell Vesicles 2015; 4: 27066.
- [34] Deatherage BL and Cookson BT. Membrane vesicle release in bacteria, eukaryotes, and archaea: a conserved yet underappreciated as-

- pect of microbial life. Infect Immun 2012; 80: 1948-1957.
- [35] Li P, Kaslan M, Lee SH, Yao J and Gao Z. Progress in exosome isolation techniques. Theranostics 2017; 7: 789-804.
- [36] Momen-Heravi F, Balaj L, Alian S, Mantel PY, Halleck AE, Trachtenberg AJ, Soria CE, Oquin S, Bonebreak CM, Saracoglu E, Skog J and Kuo WP. Current methods for the isolation of extracellular vesicles. Biol Chem 2013; 394: 1253-1262.
- [37] Xu R, Greening DW, Zhu HJ, Takahashi N and Simpson RJ. Extracellular vesicle isolation and characterization: toward clinical application. J Clin Invest 2016; 126: 1152-1162.
- [38] Hong J, Dauros-Singorenko P, Whitcombe A, Payne L, Blenkiron C, Phillips A and Swift S. Analysis of the Escherichia coli extracellular vesicle proteome identifies markers of purity and culture conditions. J Extracell Vesicles 2019; 8: 1632099.
- [39] Crescitelli R, Lasser C, Jang SC, Cvjetkovic A, Malmhall C, Karimi N, Hoog JL, Johansson I, Fuchs J, Thorsell A, Gho YS, Olofsson Bagge R and Lotvall J. Subpopulations of extracellular vesicles from human metastatic melanoma tissue identified by quantitative proteomics after optimized isolation. J Extracell Vesicles 2020; 9: 1722433.
- [40] Zhou X, Wen W, Zhu J, Huang Z, Zhang L, Zhang H, Qi LW, Shan X, Wang T, Cheng W, Zhu D, Yin Y, Chen Y, Zhu W, Shu Y and Liu P. A six-microR-NA signature in plasma was identified as a potential biomarker in diagnosis of esophageal squamous cell carcinoma. Oncotarget 2017; 8: 34468-34480.
- [41] Kahlert C, Melo SA, Protopopov A, Tang J, Seth S, Koch M, Zhang J, Weitz J, Chin L, Futreal A and Kalluri R. Identification of double-stranded genomic DNA spanning all chromosomes with mutated KRAS and p53 DNA in the serum exosomes of patients with pancreatic cancer. J Biol Chem 2014; 289: 3869-3875.
- [42] Costa-Silva B, Aiello NM, Ocean AJ, Singh S, Zhang H, Thakur BK, Becker A, Hoshino A, Mark MT, Molina H, Xiang J, Zhang T, Theilen TM, García-Santos G, Williams C, Ararso Y, Huang Y, Rodrigues G, Shen TL, Labori KJ, Lothe IM, Kure EH, Hernandez J, Doussot A, Ebbesen SH, Grandgenett PM, Hollingsworth MA, Jain M, Mallya K, Batra SK, Jarnagin WR, Schwartz RE, Matei I, Peinado H, Stanger BZ, Bromberg J and Lyden D. Pancreatic cancer exosomes initiate pre-metastatic niche formation in the liver. Nat Cell Biol 2015; 17: 816-26.
- [43] Tokuhisa M, Ichikawa Y, Kosaka N, Ochiya T, Yashiro M, Hirakawa K, Kosaka T, Makino H, Akiyama H, Kunisaki C and Endo I. Exosomal

- miRNAs from peritoneum lavage fluid as potential prognostic biomarkers of peritoneal metastasis in gastric cancer. PLoS One 2015; 10: e0130472.
- [44] Kagota S, Taniguchi K, Lee SW, Ito Y, Kuranaga Y, Hashiguchi Y, Inomata Y, Imai Y, Tanaka R, Tashiro K, Kawai M, Akao Y and Uchiyama K. Analysis of extracellular vesicles in gastric juice from gastric cancer patients. Int J Mol Sci 2019; 20: 953.
- [45] Herrera M, Llorens C, Rodriguez M, Herrera A, Ramos R, Gil B, Candia A, Larriba MJ, Garre P, Earl J, Rodriguez-Garrote M, Caldes T, Bonilla F, Carrato A, Garcia-Barberan V and Pena C. Differential distribution and enrichment of noncoding RNAs in exosomes from normal and Cancer-associated fibroblasts in colorectal cancer. Mol Cancer 2018; 17: 114.
- [46] Dai GY, Yao XG, Zhang YB, Gu JB, Geng YF, Xue F and Zhang JC. Colorectal cancer cell-derived exosomes containing miR-10b regulate fibroblast cells via the PI3K/Akt pathway. Bulletin Du Cancer 2018; 105: 336-349.
- [47] Ju Q, Zhao LN, Gao JJ, Zhou LP, Xu Y, Sun YL and Zhao XH. Mutant p53 increases exosomemediated transfer of miR-21-3p and miR-769-3p to promote pulmonary metastasis. Chin J Cancer Res 2019; 31: 533-546.
- [48] Cooks T, Pateras IS, Jenkins LM, Patel KM, Robles AI, Morris J, Forshew T, Appella E, Gorgoulis VG and Harris CC. Mutant p53 cancers reprogram macrophages to tumor supporting macrophages via exosomal miR-1246. Nat Commun 2018; 9: 771.
- [49] Campanella C, Rappa F, Sciume C, Marino Gammazza A, Barone R, Bucchieri F, David S, Curcuru G, Caruso Bavisotto C, Pitruzzella A, Geraci G, Modica G, Farina F, Zummo G, Fais S, Conway de Macario E, Macario AJ and Cappello F. Heat shock protein 60 levels in tissue and circulating exosomes in human large bowel cancer before and after ablative surgery. Cancer 2015; 121: 3230-3239.
- [50] Zhou Y, Ren HZ, Dai B, Li J, Shang LC, Huang JF and Shi XL. Hepatocellular carcinoma-derived exosomal miRNA-21 contributes to tumor progression by converting hepatocyte stellate cells to cancer-associated fibroblasts. J Exp Clin Cancer Res 2018; 37: 324.
- [51] Xu JS, Liao KL and Zhou WM. Exosomes regulate the transformation of cancer cells in cancer stem cell homeostasis. Stem Cells Int 2018; 2018: 4837370.
- [52] Scavo MP, Depalo N, Rizzi F, Ingrosso C, Fanizza E, Chieti A, Messa C, Denora N, Laquintana V, Striccoli M, Curri ML and Giannelli G. FZD10 carried by exosomes sustains cancer cell proliferation. Cells 2019; 8: 777.

- [53] Scavo MP, Cigliano A, Depalo N, Fanizza E, Bianco MG, Denora N, Laquintana V, Curri ML, Lorusso D, Lotesoriere C, Panarese A and Giannelli G. Frizzled-10 extracellular vesicles plasma concentration is associated with tumoral progression in patients with colorectal and gastric cancer (vol 2019, 2715968, 2019). J Oncol 2019; 2019: 2715968.
- [54] Min L, Zhu S, Chen L, Liu X, Wei R, Zhao L, Yang Y, Zhang Z, Kong G, Li P and Zhang S. Evaluation of circulating small extracellular vesicles derived miRNAs as biomarkers of early colon cancer: a comparison with plasma total miR-NAs. J Extracell Vesicles 2019; 8: 1643670.
- [55] Tian Y, Ma L, Gong M, Su G, Zhu S, Zhang W, Wang S, Li Z, Chen C, Li L, Wu L and Yan X. Protein profiling and sizing of extracellular vesicles from colorectal cancer patients via flow cytometry. ACS Nano 2018; 12: 671-680.
- [56] Zeng Z, Li Y, Pan Y, Lan X, Song F, Sun J, Zhou K, Liu X, Ren X, Wang F, Hu J, Zhu X, Yang W, Liao W, Li G, Ding Y and Liang L. Cancer-derived exosomal miR-25-3p promotes pre-metastatic niche formation by inducing vascular permeability and angiogenesis. Nat Commun 2018; 9: 5395.
- [57] Wang M, Qiu R, Yu S, Xu X, Li G, Gu R, Tan C, Zhu W and Shen B. Paclitaxelresistant gastric cancer MGC803 cells promote epithelialtomesenchymal transition and chemoresistance in paclitaxelsensitive cells via exosomal delivery of miR1555p. Int J Oncol 2019; 54: 326-338.
- [58] Bhome R, Goh RW, Bullock MD, Pillar N, Thirdborough SM, Mellone M, Mirnezami R, Galea D, Veselkov K, Gu Q, Underwood TJ, Primrose JN, De Wever O, Shomron N, Sayan AE and Mirnezami AH. Exosomal microRNAs derived from colorectal cancer-associated fibroblasts: role in driving cancer progression. Aging (Albany NY) 2017; 9: 2666-2694.
- [59] Wells JM, Rossi O, Meijerink M and van Baarlen P. Epithelial crosstalk at the microbiotamucosal interface. Proc Natl Acad Sci U S A 2011; 108 Suppl 1: 4607-4614.
- [60] Meng C, Bai C, Brown TD, Hood LE and Tian Q. Human gut microbiota and gastrointestinal cancer. Methods Mol Biol 2018; 16: 33-49.
- [61] Coker OO, Dai Z, Nie Y, Zhao G, Cao L, Nakatsu G, Wu WK, Wong SH, Chen Z, Sung JJY and Yu J. Mucosal microbiome dysbiosis in gastric carcinogenesis. Gut 2018; 67: 1024-1032.
- [62] Tsoi H, Chu ESH, Zhang X, Sheng J, Nakatsu G, Ng SC, Chan AWH, Chan FKL, Sung JJY and Yu J. Peptostreptococcus anaerobius induces intracellular cholesterol biosynthesis in colon cells to induce proliferation and causes dysplasia in mice. Gastroenterology 2017; 152: 1419-1433, e5.

- [63] Péré-Védrenne C, Prochazkova-Carlotti M, Rousseau B, He W, Chambonnier L, Sifré E, Buissonnière A, Dubus P, Mégraud F, Varon C and Ménard A. The cytolethal distending toxin subunit CdtB of promotes senescence and endoreplication in xenograft mouse models of hepatic and intestinal cell lines. Front Cell Infect Microbiol 2017; 7: 268.
- [64] Michaud DS. Role of bacterial infections in pancreatic cancer. Carcinogenesis 2013; 34: 2193-2197.
- [65] Wu S, Rhee KJ, Albesiano E, Rabizadeh S, Wu X, Yen HR, Huso DL, Brancati FL, Wick E, McAllister F, Housseau F, Pardoll DM and Sears CL. A human colonic commensal promotes colon tumorigenesis via activation of T helper type 17 T cell responses. Nat Med 2009; 15: 1016-1022.
- [66] Arthur JC, Perez-Chanona E, Mühlbauer M, Tomkovich S, Uronis JM, Fan TJ, Campbell BJ, Abujamel T, Dogan B, Rogers AB, Rhodes JM, Stintzi A, Simpson KW, Hansen JJ, Keku TO, Fodor AA and Jobin C. Intestinal inflammation targets cancer-inducing activity of the microbiota. Science 2012; 338: 120-123.
- [67] Huycke MM, Abrams V and Moore DR. Enterococcus faecalis produces extracellular superoxide and hydrogen peroxide that damages colonic epithelial cell DNA. Carcinogenesis 2002; 23: 529-536.
- [68] Boleij A and Tjalsma H. The itinerary of Streptococcus gallolyticus infection in patients with colonic malignant disease. Lancet Infect Dis 2013; 13: 719-724.
- [69] Castellarin M, Warren RL, Freeman JD, Dreolini L, Krzywinski M, Strauss J, Barnes R, Watson P, Allen-Vercoe E, Moore RA and Holt RA. Fusobacterium nucleatum infection is prevalent in human colorectal carcinoma. Genome Res 2012; 22: 299-306.
- [70] Wang TT, Cai GX, Qiu YP, Fei N, Zhang MH, Pang XY, Jia W, Cai SJ and Zhao LP. Structural segregation of gut microbiota between colorectal cancer patients and healthy volunteers. ISME J 2012; 6: 320-329.
- [71] Viljoen KS, Dakshinamurthy A, Goldberg P and Blackburn JM. Quantitative profiling of colorectal cancer-associated bacteria reveals associations between fusobacterium spp., enterotoxigenic Bacteroides fragilis (ETBF) and clinicopathological features of colorectal cancer. PLoS One 2015; 10: e0119462.
- [72] Rubinstein MR, Wang X, Liu W, Hao Y, Cai G and Han YW. Fusobacterium nucleatum promotes colorectal carcinogenesis by modulating E-cadherin/beta-catenin signaling via its FadA adhesin. Cell Host Microbe 2013; 14: 195-206.

- [73] Boleij A, Hechenbleikner EM, Goodwin AC, Badani R, Stein EM, Lazarev MG, Ellis B, Carroll KC, Albesiano E, Wick EC, Platz EA, Pardoll DM and Sears CL. The Bacteroides fragilis toxin gene is prevalent in the colon mucosa of colorectal cancer patients. Clin Infect Dis 2015; 60: 208-215.
- [74] Kim JH, Lee J, Park J and Gho YS. Gram-negative and Gram-positive bacterial extracellular vesicles. Semin Cell Dev Biol 2015; 40: 97-104.
- [75] Celluzzi A and Masotti A. How our other genome controls our epi-genome. Trends Microbiol 2016; 24: 777-787.
- [76] Kinder SA and Holt SC. Localization of the Fusible Scheme Scheme
- [77] Patrick S, McKenna JP, O'Hagan S and Dermott E. A comparison of the haemagglutinating and enzymic activities of Bacteroides fragilis whole cells and outer membrane vesicles. Microb Pathog 1996; 20: 191-202.
- [78] Chu H, Khosravi A, Kusumawardhani IP, Kwon AH, Vasconcelos AC, Cunha LD, Mayer AE, Shen Y, Wu WL, Kambal A, Targan SR, Xavier RJ, Ernst PB, Green DR, McGovern DP, Virgin HW and Mazmanian SK. Gene-microbiota interactions contribute to the pathogenesis of inflammatory bowel disease. Science 2016; 352: 1116-1120.
- [79] Aykut B, Pushalkar S, Chen R, Li Q, Abengozar R, Kim JI, Shadaloey SA, Wu D, Preiss P, Verma N, Guo Y, Saxena A, Vardhan M, Diskin B, Wang W, Leinwand J, Kurz E, Kochen Rossi JA, Hundeyin M, Zambrinis C, Li X, Saxena D and Miller G. The fungal mycobiome promotes pancreatic oncogenesis via activation of MBL. Nature 2019; 574: 264-267.
- [80] Rolhion N, Barnich N, Claret L and Darfeuille-Michaud A. Strong decrease in invasive ability and outer membrane vesicle release in Crohn's disease-associated adherent-invasive Escherichia coli strain LF82 with the yfgL gene deleted. J Bacteriol 2005; 187: 2286-2296.
- [81] Keenan J, Day T, Neal S, Cook B, Perez-Perez G, Allardyce R and Bagshaw P. A role for the bacterial outer membrane in the pathogenesis of Helicobacter pylori infection. FEMS Microbiol Lett 2000; 182: 259-264.
- [82] Simbari F, McCaskill J, Coakley G, Millar M, Maizels RM, Fabriás G, Casas J and Buck AH. Plasmalogen enrichment in exosomes secreted by a nematode parasite versus those derived from its mouse host: implications for exosome stability and biology. J Extracell Vesicles 2016; 5: 30741.

- [83] Song MK, Kim HY, Choi BK and Kim HH. Filifactor alocis-derived extracellular vesicles inhibit osteogenesis through TLR2 signaling. Mol Oral Microbiol 2020; 35: 202-210.
- [84] Barteneva NS, Maltsev N and Vorobjev IA. Microvesicles and intercellular communication in the context of parasitism. Front Cell Infect Microbiol 2013; 3: 49.
- [85] Robinson KM and Dunning Hotopp JC. Mobile elements and viral integrations prompt considerations for bacterial DNA integration as a novel carcinogen. Cancer Lett 2014; 352: 137-144.
- [86] Ruff WE and Kriegel MA. Autoimmune hostmicrobiota interactions at barrier sites and beyond. Trends Mol Med 2015; 21: 233-244.
- [87] Barteneva NS, Baiken Y, Fasler-Kan E, Alibek K, Wang S, Maltsev N, Ponomarev ED, Sautbayeva Z, Kauanova S, Moore A, Beglinger C and Vorobjev IA. Extracellular vesicles in gastrointestinal cancer in conjunction with microbiota: on the border of Kingdoms. Biochim Biophys Acta Rev Cancer 2017; 1868: 372-393.
- [88] Choi DS, Choi DY, Hong BS, Jang SC, Kim DK, Lee J, Kim YK, Kim KP and Gho YS. Quantitative proteomics of extracellular vesicles derived from human primary and metastatic colorectal cancer cells. J Extracell Vesicles 2012; 1.
- [89] Zhang X, Deeke SA, Ning Z, Starr AE, Butcher J, Li J, Mayne J, Cheng K, Liao B, Li L, Singleton R, Mack D, Stintzi A and Figeys D. Metaproteomics reveals associations between microbiome and intestinal extracellular vesicle proteins in pediatric inflammatory bowel disease. Nat Commun 2018; 9: 2873.
- [90] Teng Y, Ren Y, Sayed M, Hu X, Lei C, Kumar A, Hutchins E, Mu J, Deng Z, Luo C, Sundaram K, Sriwastva MK, Zhang L, Hsieh M, Reiman R, Haribabu B, Yan J, Jala VR, Miller DM, Van Keuren-Jensen K, Merchant ML, McClain CJ, Park JW, Egilmez NK and Zhang HG. Plant-derived exosomal microRNAs shape the gut microbiota. Cell Host Microbe 2018; 24: 637-652, e638.
- [91] Polyak K. Tumor heterogeneity confounds and illuminates: a case for Darwinian tumor evolution. Nat Med 2014; 20: 344-346.
- [92] Mege D, Panicot-Dubois L, Ouaissi M, Robert S, Sielezneff I, Sastre B, Dignat-George F and Dubois C. The origin and concentration of circulating microparticles differ according to cancer type and evolution: a prospective singlecenter study. Int J Cancer 2016; 138: 939-48.
- [93] Nakajima T, Enomoto S, Yamashita S, Ando T, Nakanishi Y, Nakazawa K, Oda I, Gotoda T and Ushijima T. Persistence of a component of DNA methylation in gastric mucosae after Helico-

- bacter pylori eradication. J Gastroenterol 2010; 45: 37-44.
- [94] Dai S, Wei D, Wu Z, Zhou X, Wei X, Huang H and Li G. Phase I clinical trial of autologous ascitesderived exosomes combined with GM-CSF for colorectal cancer. Mol Ther 2008; 16: 782-790.
- [95] Lugini L, Cecchetti S, Huber V, Luciani F, Macchia G, Spadaro F, Paris L, Abalsamo L, Colone M, Molinari A, Podo F, Rivoltini L, Ramoni C and Fais S. Immune surveillance properties of human NK cell-derived exosomes. J Immunol 2012; 189: 2833-2842.
- [96] Lai RC, Yeo RW and Lim SK. Mesenchymal stem cell exosomes. Semin Cell Dev Biol 2015; 40: 82-88.
- [97] Witwer KW, Van Balkom BWM, Bruno S, Choo A, Dominici M, Gimona M, Hill AF, De Kleijn D, Koh M, Lai RC, Mitsialis SA, Ortiz LA, Rohde E, Asada T, Toh WS, Weiss DJ, Zheng L, Giebel B and Lim SK. Defining mesenchymal stromal cell (MSC)-derived small extracellular vesicles for therapeutic applications. J Extracell Vesicles 2019; 8: 1609206.
- [98] Börger V, Weiss DJ, Anderson JD, Borràs FE, Bussolati B, Carter DRF, Dominici M, Falcón-Pérez JM, Gimona M, Hill AF, Hoffman AM, de Kleijn D, Levine BL, Lim R, Lötvall J, Mitsialis SA, Monguió-Tortajada M, Muraca M, Nieuwland R, Nowocin A, O'Driscoll L, Ortiz LA, Phinney DG, Reischl I, Rohde E, Sanzenbacher R, Théry C, Toh WS, Witwer KW, Lim SK and Giebel B. International society for extracellular vesicles and international society for cell and gene therapy statement on extracellular vesicles from mesenchymal stromal cells and other cells: considerations for potential therapeutic agents to suppress coronavirus disease-19. Cytotherapy 2020; 22: 482-485.
- [99] Dellett M, Brown ED, Guduric-Fuchs J, O'Connor A, Stitt AW, Medina RJ and Simpson DA. MicroRNA-containing extracellular vesicles released from endothelial colony-forming cells modulate angiogenesis during ischaemic retinopathy. J Cell Mol Med 2017; 21: 3405-3419.
- [100] Usman WM, Pham TC, Kwok YY, Vu LT, Ma V, Peng B, Chan YS, Wei L, Chin SM, Azad A, He AB, Leung AYH, Yang M, Shyh-Chang N, Cho WC, Shi J and Le MTN. Efficient RNA drug delivery using red blood cell extracellular vesicles. Nat Commun 2018; 9: 2359.
- [101] Tian Y, Li S, Song J, Ji T, Zhu M, Anderson GJ, Wei J and Nie G. A doxorubicin delivery platform using engineered natural membrane vesicle exosomes for targeted tumor therapy. Biomaterials 2014; 35: 2383-2390.
- [102] Sun D, Zhuang X, Xiang X, Liu Y, Zhang S, Liu C, Barnes S, Grizzle W, Miller D and Zhang HG. A novel nanoparticle drug delivery system: the anti-inflammatory activity of curcumin is en-

- hanced when encapsulated in exosomes. Mol Ther 2010; 18: 1606-1614.
- [103] Vader P, Mol EA, Pasterkamp G and Schiffelers RM. Extracellular vesicles for drug delivery. Adv Drug Deliv Rev 2016; 106: 148-156.
- [104] Barbosa FMC, Dupin TV, Toledo MDS, Reis NFDC, Ribeiro K, Cronemberger-Andrade A, Rugani JN, De Lorenzo BHP, Novaes E Brito RR, Soares RP, Torrecilhas AC and Xander P. Extracellular vesicles released by leishmania (Leishmania) amazonensis promote disease progression and induce the production of different cytokines in macrophages and B-1 cells. Front Microbiol 2018; 9: 3056.
- [105] Eichenberger RM, Ryan S, Jones L, Buitrago G, Polster R, Montes de Oca M, Zuvelek J, Giacomin PR, Dent LA, Engwerda CR, Field MA, Sotillo J and Loukas A. Hookworm secreted extracellular vesicles interact with host cells and prevent inducible colitis in mice. Front Immunol 2018; 9: 850.
- [106] Roig J, Saiz ML, Galiano A, Trelis M, Cantalapiedra F, Monteagudo C, Giner E, Giner RM, Recio MC, Bernal D, Sanchez-Madrid F and Marcilla A. Extracellular vesicles from the helminth fasciola hepatica prevent DSS-induced acute ulcerative colitis in a T-Lymphocyte independent mode. Front Microbiol 2018; 9: 1036.
- [107] Mendt M, Kamerkar S, Sugimoto H, McAndrews KM, Wu CC, Gagea M, Yang S, Blanko EVR, Peng Q, Ma X, Marszalek JR, Maitra A, Yee C, Rezvani K, Shpall E, LeBleu VS and Kalluri R. Generation and testing of clinical-grade exosomes for pancreatic cancer. JCI Insight 2018; 3: e99263.
- [108] Pi F, Binzel DW, Lee TJ, Li Z, Sun M, Rychahou P, Li H, Haque F, Wang S, Croce CM, Guo B, Evers BM and Guo P. Nanoparticle orientation to control RNA loading and ligand display on extracellular vesicles for cancer regression. Nat Nanotechnol 2018: 13: 82-89.
- [109] Chang DK, Grimmond SM and Biankin AV. Pancreatic cancer genomics. Curr Opin Genet Dev 2014; 24: 74-81.
- [110] Collins MA, Brisset JC, Zhang Y, Bednar F, Pierre J, Heist KA, Galbán CJ, Galbán S and di Magliano MP. Metastatic pancreatic cancer is dependent on oncogenic Kras in mice. PLoS One 2012; 7: e49707.
- [111] Kamerkar S, LeBleu VS, Sugimoto H, Yang S, Ruivo CF, Melo SA, Lee JJ and Kalluri R. Exosomes facilitate therapeutic targeting of oncogenic KRAS in pancreatic cancer. Nature 2017; 546: 498-503.
- [112] Seo N, Shirakura Y, Tahara Y, Momose F, Harada N, Ikeda H, Akiyoshi K and Shiku H. Activated CD8 T cell extracellular vesicles prevent tumour progression by targeting of lesional mesenchymal cells. Nat Commun 2018; 9: 435.

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- [113] Im H, Lee K, Weissleder R, Lee H and Castro CM. Novel nanosensing technologies for exosome detection and profiling. Lab Chip 2017; 17: 2892-2898.
- [114] Zijlstra C and Stoorvogel W. Prostasomes as a source of diagnostic biomarkers for prostate cancer. J Clin Invest 2016; 126: 1144-1151.
- [115] Taylor DD and Gercel-Taylor C. MicroRNA signatures of tumor-derived exosomes as diagnostic biomarkers of ovarian cancer. Gynecol Oncol 2008; 110: 13-21.
- [116] Alexander M, Hu R, Runtsch MC, Kagele DA, Mosbruger TL, Tolmachova T, Seabra MC, Round JL, Ward DM and O'Connell RM. Exosome-delivered microRNAs modulate the inflammatory response to endotoxin. Nat Commun 2015; 6: 7321.
- [117] Taylor DD and Gercel-Taylor C. Exosomes/microvesicles: mediators of cancer-associated immunosuppressive microenvironments. Semin Immunopathol 2011; 33: 441-454.
- [118] Peinado H, Alečković M, Lavotshkin S, Matei I, Costa-Silva B, Moreno-Bueno G, Hergueta-Redondo M, Williams C, García-Santos G, Ghajar C, Nitadori-Hoshino A, Hoffman C, Badal K, Garcia BA, Callahan MK, Yuan J, Martins VR, Skog J, Kaplan RN, Brady MS, Wolchok JD, Chapman PB, Kang Y, Bromberg J and Lyden D. Melanoma exosomes educate bone marrow progenitor cells toward a pro-metastatic phenotype through MET. Nat Med 2012; 18: 883-891.

- [119] Jie M, Wu Y, Gao M, Li X, Liu C, Ouyang Q, Tang Q, Shan C, Lv Y, Zhang K, Dai Q, Chen Y, Zeng S, Li C, Wang L, He F, Hu C and Yang S. Circ-MRPS35 suppresses gastric cancer progression via recruiting KAT7 to govern histone modification. Mol Cancer 2020; 19: 56.
- [120] He JH, Han ZP, Luo JG, Jiang JW, Zhou JB, Chen WM, Lv YB, He ML, Zheng L, Li YG and Zuo JD. Hsa\_Circ\_0007843 acts as a mir-518c-5p sponge to regulate the migration and invasion of colon cancer SW480 cells. Front Genet 2020; 11: 9.