Review Article Systemic therapies for salivary gland adenoid cystic carcinoma

Sosuke Sahara^{1,2}, Alexandra E Herzog¹, Jacques E Nör^{1,3,4,5}

¹Department of Cariology, Restorative Sciences, and Endodontics, University of Michigan School of Dentistry, Ann Arbor, Michigan 48109-1078, USA; ²Department of Otorhinolaryngology/Head and Neck Surgery, Hamamatsu University School of Medicine, Hamamatsu 431-3192, Japan; ³Department of Otolaryngology-Head & Neck Surgery, University of Michigan School of Medicine, Ann Arbor, Michigan 48109-1078, USA; ⁴Department of Biomedical Engineering, University of Michigan College of Engineering, Ann Arbor, Michigan 48109, USA; ⁵University of Michigan Rogel Cancer Center, Ann Arbor, Michigan 48109, USA

Received June 18, 2021; Accepted August 9, 2021; Epub September 15, 2021; Published September 30, 2021

Abstract: Adenoid cystic carcinoma (ACC) is a slow growing, but relentless cancer. Due to its rarity and lack of understanding of its molecular etiology, no standard chemotherapy for ACC currently exists and many patients suffer from recurrent and/or metastatic disease. As such, development of safe and effective therapies is imperative. To describe and summarize existing clinical trial studies and preclinical discoveries, we surveyed the PubMed on developmental therapeutics for ACC. Objective response rates to monotherapy with cytotoxic agents were approximately 10% with cisplatin, 5-FU, gemcitabine, mitoxantrone, epirubicin, vinorelbine and paclitaxel. The most studied combination therapies were cyclophosphamide-doxorubicin-cisplatin (CAP) and cisplatin-vinorelbine, with an objective response rate of 18-31%. Among molecularly targeted drugs, the most studied drugs are inhibitors targeting the vascular endothelial growth factor receptor (VEGFR) to inhibit tumor angiogenesis. Among those, lenvatinib and axitinib showed a relatively high objective response rate of 11-16% and 9-17%, respectively. Given high recurrence rates and chemoresistance of ACC, treatments targeting cancer stem cells (CSC), which function as tumor-initiating cells and drive chemoresistance, may be particularly valuable. CSC have been shown to be targetable via MYB, Notch1, p53 and epigenetic mechanisms. Myb overexpression is characteristic in ACC but was previously thought to present a difficult target due to its nature as a transcription factor. However, due to the development Myb-targeted inhibitors and an ongoing clinical trial of MYB-targeted cancer vaccine therapy, MYB is becoming an increasingly attractive therapeutic target. Drugs targeting NOTCH signaling demonstrated 5-17% response rate in phase I clinical trials. Within the field of epigenetics, treatment with PRMT5 inhibitors has shown 21% partial response rate in phase I clinical trial. Immunotherapies, such as PD-1 inhibitors, are also associated with CSC, but have not been effective against ACC. However, clinical trials of cancer vaccine therapies are actively being conducted. In addition to conventional chemotherapies and inhibitors of angiogenesis, the emergence of new therapies such as immunotherapy and those targeting cancer stemness is expected to bring clinical benefits to patients in the future.

Keywords: Adenoid cystic carcinoma, cancer stem cells, systemic therapy, chemotherapy, immunotherapy, salivary gland cancer

Introduction

Adenoid cystic carcinoma (ACC) is a relatively rare cancer, accounting for 1% of head and neck tumors and 10% of salivary gland tumors [1]. Salivary glands present the most common primary site for ACC, but tumors may also occur in the lacrimal glands, paranasal sinuses, mammary glands, skin, and genital organs [2-5]. The disease progresses relatively slowly, but continues to grow relentlessly, giving rise to a 5-year and 10-year patient survival rate of about 60% and 50%, respectively. Notably, the 20-year survival rate is only 20%, due to high incidences of recurrence and metastasis [6]. Lungs constitute the most common site for metastases, while perineural invasion is characteristic of local progression for ACC [7-9].

At present, the standard treatment for ACC remains surgery with or without radiation therapy, as no approved systemic therapy currently exists. Although radiation therapy has a positive effect for local disease control, its effect on prolonging overall survival is unclear [10, 11]. Distant metastasis is the most common type of relapse, with about 30% of patients with distant metastasis not experiencing recurrence of the primary lesion [10, 12].

The need for a robust preclinical ACC model is apparent to advance basic and translational research within the field. However, it was reported that 6 types of ACC cell lines (ACC2, ACC3, ACCM, ACCNS, ACCS and CAC2) commonly used by many laboratories were crosscontaminated with other cells. This poses a large question mark on the research resulting from use of these cells, while leaving in vitro studies reliant on use of low passage primary cells [13]. Recently, Moskaluk and collaborators reported a xenograft model that reproduces the characteristics of human ACC [14], from which several cell lines have been established, thereby raising expectations that the understanding of pathological conditions underlying ACC will be accelerated [15]. More recently, a MYB-NFIB fusion-positive ACC cell line was developed and characterized as well suited for developmental therapeutics studies [16]. In this review, we will focus on the current state of pharmacological treatment of ACC. We surveyed the PubMed, summarized results of clinical trials, and discussed the potential clinical benefit of these therapies to ACC patients in the future.

Clinical trial data for cytotoxic chemotherapy

Single agent

In 10 studies focusing on single agent chemotherapeutic treatment, only 22 out of 163 (16%) patients demonstrated objective responses. No response was seen in the 21 patients who received gemcitabine or the 14 patients who received paclitaxel [17, 18]. Cisplatin and 5-FU showed relatively high objective response rates. Response rates to cisplatin vary widely from report to report, ranging from 0 to 70%. In addition, a 33% (4 of 12) objective response rate was observed upon 5-FU treatment [19-22]. Regarding the anthracycline cytotoxic agents mitoxantrone and epirubicin, the number of patients participating in the study was relatively large, whereas the objective response rate was only 5-12% [2325] (**Table 1**). Almost half of patients showed stable disease (65 of 113). However, it was difficult to determine whether the disease was stable due to the drug's effect or the natural history of ACC.

Combination chemotherapy

The combination of cisplatin, doxorubicin and cyclophosphamide (CAP) was the most common regimen for ACC, sometimes combined with 5-FU. From 5 studies, 27% patients (12 of 43) presented objective responses [26-30]. Patients treated with a CAP plus 5-FU regimen demonstrated the longest duration of objective response. However, the authors mentioned that this treatment was too toxic to be considered standard treatment [30]. Cisplatin and doxorubicin were used in combination with bleomycin, in which 3 of 9 patients showed objective response [21]. The combination of cyclophosphamide, vincristine, and 5-FU showed a rather long duration of objective responses [31]. The above studies have been conducted relatively early. However, the regimens of paclitaxel-carboplatin and cisplatin-vinorelbine are newer and have been reported since 2000. Two studies investigated the combination of carboplatin with paclitaxel, where 2 of 10 and 1 of 9 patients showed objective responses [32, 33]. A randomized phase II trial was completed to compare single-agent vinorelbine with cisplatin plus vinorelbine, but included only 36 patients (of which 22 patients had ACC) [34]. Another study showed that 7 of 34 (20%) and 6 of 19 (31%) patients demonstrate objective responses [35, 36]. In several studies, the combination therapy groups demonstrated greater toxicities, particularly nausea, vomiting, myelosuppression, and neuropathy. Notably, the vinorelbine plus cisplatin regimen shows higher objective response rate compared to the response to the CAP regimen. Overall, when focusing on studies with 10 or more patients, the best objective response rates were obtained with cisplatin-vinorelbine combination [28, 29, 33, 35-37] (Table 1).

Targeted therapies

c-Kit and EGFR

As in many other cancers, treatment with molecularly targeted drugs has been attempted in ACC. It is known that high c-Kit expression

single agent	authors	year	# of patients with ACC	objective responses	duration	stable disease	median survival (months)
cisplatin	Schramm et al. [19]	1981	10	7 (70%)	7-18 months	NS	NS
	Licitra et al. [20]	1991	13	2 (15%)	5-8 months	6 (46%)	20
	Dick Haan et al. [21]	1992	10	0		5 (50%)	78
5-FU	Tannock et al. [22]	1980	12	4 (33%)	5-24 months	2 (17%)	NS
gemcitabine	van Herpen et al. [18]	2008	21	0		11 (52%)	NS
mitoxantrone	Verweij et al. [23]	1996	32	4 (12%)	3-13 months	22 (69%)	18
	Mattox et al. [24]	1990	18	1 (5%)		12 (66%)	19
epirubicin	Vermorken et al. [25]	1993	20	2 (10%)	7.5, 20 months	NS	16
vinorelvine	Airoldi et al. [34]	2001	13	2 (15%)	NS	NS	NS
paclitaxel	Gilbert et al. [17]	2006	14	0		7 (50%)	25
combination therapy							
CAP	Licitra et al. [29]	1996	12	3 (25%)	5, 9, 13 months	5 (41%)	34
	Creagan et al. [28]	1988	11	2 (18%)	12, 12 months	NS	22.5
	Belani et al. [26]	1988	4	1 (25%)	16 months	NS	13
	Dreyfuss et al. [27]	1987	9	3 (33%)	NS	NS	NS
CAP + 5-FU	Dimery et al. [30]	1990	7	3 (42%)	6, 13, 18 months	2 (28%)	29
P + A + bleomycin	Dick Haan et al. [21]	1992	9	3 (33%)	6, 21, 77 months	5 (55%)	67
CVF	Triozzi et al. [31]	1987	8	2 (25%)	107, 28 months	4 (50%)	NS
P + 5-FU	Hill et al. [37]	1997	11	0		9 (81%)	12
carboplatin, paclitaxel	Airoldi et al. [33]	2000	10	2 (20%)	5, 12 months	NS	NS
	Nakano et al. [32]	2016	9	1 (10%)	NS	NS	21.9
P + vinorelvine	Airoldi et al. [34]	2001	9	4 (44%)	NS	NS	NS
	Airoldi et al. [35]	2017	34	7 (20%)	NS	NS	10.2
	Hong et al. [36]	2018	19	6 (31%)	NS	10 (52%)	NS

Table 1. Studies reporting cytotoxic chemotherapy for ACC

*P: cisplatin, C: cyclophosphamide, A: doxorubicin, V: vincristine, F: 5-fluorouracil.

was confirmed in 90% of ACC tumors [38]. But, single drug imatinib (targets c-Kit) resulted in no objective responses in ACC [39-41]. Dasatinib, which also targets c-Kit, failed to show any activity in ACC [42]. In one trial, combination treatment with cisplatin and imatinib was conducted and showed 3 (10%) cases of partial response [43]. Increased expression of epidermal growth factor receptor (EGFR) was also observed in ACC [44]. However, clinical trials with EGFR inhibitors (e.g. cetuximab, gefitinib, lapatinib) did not provide a positive therapeutic responses [45-47] (Table 2). Given the results of these trials, molecular targets other than c-Kit and EGFR have been considered in ACC.

Targeting angiogenesis

Vascular endothelial growth factor (VEGF) is highly expressed in approximately 76% of ACC patients, raising the possibility that VEGF function may be associated with recurrence and metastases [48]. It has also been shown that

overexpression of MYB promotes expression of several target genes including VEGF [49]. Therefore, VEGF receptor (VEGFR) could be considered as a potential therapeutic target for ACC. Chen and colleagues [50] have shown the efficacy of regorafenib in a preclinical study, where it led to delayed tumor growth and metastasis in 2 patient-derived xenograft (PDX) mouse models. To observe impacts of treatment of metastasis, they implanted labeled ACC cells in zebrafish embryos. Regorafenib treatment inhibited ACC cell migration and intravascular invasion as compared to the control group. Several VEGFR-targeted drugs are being tested in clinical trials. No objective response was observed to sunitinib, regorafenib, and nintedanib [51-53], but 9-16% objective response rate was observed with sorafenib, axitinib, and Lenvatinib. These data suggest limited efficacy of VEGFR-targeted treatment [54-59] (Table 2). Particularly, Tchekmedyian and collaborators showed that lenvatinib leads to the longest median progression-free survival (mPFS) of 17.5 months [55].

Table 2. Studies reporting targeted therapies for ACC

phase II studies	target	authors	year	# of patients	# of patients with ACC	objective responses	stable disease	median survival (months)	median PFS (months)
imatinib	c-kit, bcrabl, PDGFR	Hotte et al. [41]	2005	16	16	0	9 (56%)	7	2
		Pfeffer et al. [39]	2007	10	10	0	2 (20%)	NS	NS
		Bahl et al. [40]	2019	8	8	0	2 (25%)	NS	NS
imatinib + cisplatin		Ghosal et al. [43]	2011	28	28	3 (11%)	19 (67%)	35	15
dasatinib	c-kit, bcrabl, SRC family, PDGF β , EPHA2	Wong et al. [42]	2016	54	40	1 (2.5%)	20 (50%)	14.5	4.8
cetuximab	EGFR	Locati et al. [46]	2009	30	23	0	20 (87%)	NS	NS
gefitinib	EGFR	Jakob et al. [45]	2015	36	19	0	13 (68%)	25.9	4.3
lapatinib	HER-2, EGFR	Agulnik et al. [47]	2007	40	19	0	15 (79%)	NS	3.5
dovitinib	FGFR, VEGFR, PDGFR, c-Kit	Keam et al. [68]	2015	32	32	1 (3%)	30 (93%)	NS	6
		Dillon et al. [69]	2017	34	34	2 (6%)	22 (65%)	20.6	8.2
sunitinib	VEGFR, c-KIT, PDGFR	Chau et al. [52]	2012	13	13	0	11 (85%)	18.7	7.2
regorafenib	VEGFR, FGFR, PDGFR	Ho et al. [51]	2017	38	38	0	17 (45%)	NS	NS
nintedanib	VEGFR, FGFR, PDGFR	Kim et al. [53]	2017	20	13	0	10 (77%)	10	7.9
lenvatinib	VEGFR, FGFR, PDGFR and etc.	Tchekmedyian et al. [55]	2019	32	32	5 (16%)	24 (75%)	NS	17.5
		Locati et al. [54]	2020	28	28	3 (11%)	20 (71%)	27	9.1
axitinib	VEGFR, PDGFR, c-KIT	Locati et al. [57]	2019	26	6	1 (17%)	3 (50%)	NS	NS
		Ho et al. [56]	2016	33	33	3 (9%)	25 (76%)	NS	5.7
sorafenib	VEGFR, PDGFR, c-Kit and etc.	Thomson et al. [58]	2015	23	23	2 (11%)	13 (57%)	19.6	11.3
		Locati et al. [59]	2016	37	19	2 (11%)	11 (58%)	26.4	8.9
vorinostat	histone deacetylase inhibitor	Goncalves et al. [114]	2017	30	30	2 (7%)	27 (90%)	11.5	10
everolimus	mTOR	Kim et al. [123]	2014	34	34	0	27 (79%)	NS	11.2
nelfinavir	Akt pathway inhibitor	Hoover et al. [124]	2015	15	15	0	7 (47%)	NS	5.5
phase I studies									
figitumab + dacomitinib	IGF1R + EGFR inhibitor	Calvo et al. [92]	2017	74	3	1 (33%)	2 (67%)		
R1507 + sorafenib	IGF1R + multikinase inhibitor	Mahadevan et al. [93]	2014		NS		1		
BMS-986115	pan-NOTCH inhibitor	Aung et al. [99]	2018	36	NS	0	2 (5.6%)		
crenigacestat (LY3039478) expansion of phase I	pan-NOTCH inhibitor	Even et al. [100]	2020	22	22	1 (5%)	15 (68%)	NS	5.3
brontictuzumab	NOTCH1 inhibitor	Ferrarotto et al. [101]	2018	48	12	2 (17%)	3 (25%)	NS	NS
GSK3326595	PRMT5 inhibitor	Siu et al. [119]	2019	44	14	3 (21%)	NS	NS	NS

ongoing phase II	Target or drug types	dose	clinical.gov
ATRA	retinoic acid receptor (RAR)	NS	NCT03999684
AL101	pan-NOTCH inhibitor	4 mg IV weekly	NCT03691207
CB-103	pan-NOTCH inhibitor	NS	NCT03422679
APG-115 ± carboplatin	MDM2-p53 interaction	150 mg every other day	NCT03781986
chidamide + cisplatin	histone deacetylase inhibitor	Chidamide: 30 mg orally two times per week, one week before cycle 1 treatment Cisplatin 25 mg/m 2 iv	NCT03639168
lenvatinib + pembrolizumab	VEGFR inhibitor + PD-1 antibody	Lenvatinib 20 mg daily, Pembrolizumab (200 mg)	NCT04209660
rivoceranib (apatinib)	VEGFR-2 inhibitor	oral rivoceranib, 700 mg daily	NCT04119453
ongoing phase I			
PRT543	PRMT5 inhibitor		NCT03886831
CV8102	intratumor therapy TLR7/8 agonist adjuvant		NCT03291002
VMD-928	TrkA inhibitor		NCT03556228
TetMYB Vaccine	cancer therapy vaccine with PD-1 antibody		NCT03287427

 Table 3. Ongoing clinical trials for ACC

Currently, the VEGFR-2 inhibitor rivoceranib (apatinib) is in phase II trial (NCT04119453) (**Table 3**).

Tumor cells release VEGF and other pro-angiogenic factors to promote angiogenesis which enables influx of oxygen and nutrients to support the high metabolic demands of tumor growth. Interestingly, VEGF induces B-cell lymphoma (Bcl)-2 expression and enhances both endothelial and tumor cell viability in several tumor types [60]. The anti-apoptotic Bcl-2 and Bcl-x, proteins are overexpressed in adenoid cystic carcinomas [61]. Acasigua and colleagues [62] showed the effect of a BH3mimetic small molecule inhibitor (BM-1197) on ACC tumor suppression, where they demonstrated that small molecule inhibitors of Bcl-2 induce apoptosis of tumor cells by suppressing the heterodimerization of Bcl-2 and Bcl-x. In a PDX model of ACC, the BM-1197-treated group showed an increase in apoptotic rate and growth inhibition as compared to the control group. However, clinical studies targeting Bcl-2 have not been conducted yet for ACC.

Overexpression of fibroblast growth factor (FGF) has been observed in salivary gland cancers including ACC [63]. Increased FGF, as well as VEGF, has been shown to be associated with overexpression of MYB [64]. FGF is also involved in angiogenesis by promoting proliferation and migration of vascular endothelial cells [65]. Doddapaneni and colleagues reported responses of ACC of the lacrimal gland in protein expression due to intra-arterial cytoreductive chemotherapy (IACC) with cisplatin and doxorubicin. According to the study, immunofluorescence and immunohistochemical analyses demonstrate a significant increase of FGFR1 in post-IACC tissues, when comparing to pre-IACC. They also performed an *in vitro* study on ACC cells using cisplatin and AZD4547, a FGFR1 inhibitor. The cells treated with the FGFR1 inhibitor and cisplatin showed a lower cell proliferation rate and cell migration compare to the control group or the cisplatin alone group [66].

The clinical trial for dovitinib is one of the trials targeting FGFR in ACC. Dovitinib is also a drug that targets VEGFR, but it has a higher affinity for FGFR when compared to sorafenib and lenvatinib [67]. In the phase II clinical study on dovitinib, the objective response rate was 3-6%, which was lower than that of Lenvatinib [68, 69].

Targeting stemness

Cancer stem cells in adenoid cystic carcinoma

Cancer Stem Cells (CSCs) constitute a small fraction of the entire tumor cell population (typically ≥5%). These cells are highly tumorigenic, capable of self-renewal, and capable of (re-) generating the various cell phenotypes that make up a tumor [70]. It has been suggested that CSCs are resistant to conventional chemotherapy and are strongly associated with recurrence and metastasis [71, 72]. Methods for identifying CSCs have been suggested in various types of carcinoma, and the markers identified for labeling differ depending on the type of carcinoma. In a study using a PDX model of ACC, when ALDH^{high} and ALDH^{low} cell



Figure 1. Representative photomicrographs of immunofluorescence staining of putative cancer stem-like cells in Adenoid Cystic Carcinoma. Histological section prepared from a surgical specimen of human ACC (A) and from a patient-derived xenograft UM-PDX-HACC-5 (B). Tissues were stained for the CSC marker ALDH (red) and for DAPI (blue). Images were taken at 400× magnification. Scale bars represent 50 µm.

populations were subcutaneously injected into mice, ALDH^{high} cell populations were capable of forming tumors with a smaller number of cells than ALDH^{low} cells. This suggested that ALDH^{high} cells have high tumorigenicity, and that ALDH is an effective CSC marker in ACC [73] (Figure 1). In another study, when the ALDH^{high}CD44^{high} and ALDH^{low}CD44^{low} cells were compared for tumorigenicity, the ALDHhigh CD44^{high} cells were more tumorigenic. From this, it was suggested that CD44 in addition to ALDH can be a marker for CSCs in ACC [74]. It has further been reported that CD44 and CD133 have overlapping expression in ACC, suggesting that CD133 may also serve as a marker to identify ACC CSCs [75].

As mentioned above, CSCs selected by the aforementioned markers constitute a very small population [74]. As such, an effective treatment targeting CSCs might eliminate this small cell population but would not result in immediate tumor regression. Thus, treatments targeting CSCs are typically accompanied by strategies (e.g. cytotoxic therapies) aiming at the elimination of bulk tumor cells [72]. Importantly, CSC-targeted treatment has the potential to reduce ACC recurrence rates, as it eliminates tumor-initiating cells. In a preclinical recurrence study using PDX models of ACC, inhibition of the MDM2-p53 interaction with a

small molecule inhibitor reduced the fraction of CSCs and enhanced sensitivity to cisplatin. In addition, no tumor recurrence was observed after tumor resection in neoadjuvant administration of the targeted therapy group (MI-773), whereas 63% of the vehicle control group showed recurrence [76]. Altogether, treatment targeting CSCs can potentially be beneficial in ACC, as tumor recurrence is frequently observed.

MYB as a stemness target

A recurrent t(6;9)(q22-23; p23-24) translocation is a common chromosomal abnormality in ACC and results in the fusion of the protooncogene (MYB) with the transcriptional factor gene (NFIB) [49]. The MYB-NFIB gene fusion is observed in about 50% of all ACC tumors. Interestingly, 35% of MYB-NFIB gene fusion negative ACC have MYBL1 gene alterations. Therefore, it is suggested that approximately 80% of all ACC have MYB or MYBL1 gene alterations, and that MYB-like signaling might be involved in the oncogenesis and maintenance of ACC [77]. In tumors with MYB-NFIB translocation, the fusion is detected in all tumor cells. However, the expression of Myb protein is detected only in limited subsets of ACC cells, primarily with a basal phenotype. In the tumor cells with a basal phenotype, p63 has been

suggested as a CSC marker [78] and is typically co-expressed with Myb [79]. In addition, sphere formation in ultra-low attachment plates (functional *in vitro* assessment of stemness) was suppressed by MYB gene silencing. This suggests that MYB promotes tumorigenesis by enhancing cancer stemness [80].

Since MYB is a transcription factor, this genetic abnormality is particularly difficult to target clinically. However, initial approaches to inhibit MYB activity or expression by small molecule or peptide-mimetic inhibitors have already proven successful and have shown that MYB inhibition is feasible clinically. The main strategy to inhibit MYB remains targeting MYB's interaction with partner proteins or inducing direct degradation of MYB. CBP/p300 was the first identified coactivator to interact with Myb and influence its transcriptional activity [81, 82]. Recent studies have proposed that Celastrol, a natural low-molecular-weight compound, can inhibit MYB function through disruption of its interaction with the KIX domain of p300/CBP [83]. However, Celastrol reduced viability not only of MYB-positive ACC cells but also of cells not associated with MYB activation at the same concentration [84]. This suggests that Celastrol inhibits non-selective cell viability. To improve the therapeutic specificity for MYB, a peptidomimetic inhibitor (MYBMIM) that was designed to target and interfere with the assembly of the MYB:CBP/P300 co-transcriptional protein complex using structure-guided molecular design was utilized [85]. A dosedependent tumor cell viability reduction was observed in MYB-activated tumor cells by MYBMIM. On the other hand, incubating MYBMIM with cells that are not associated with MYB activation does not decrease cell viability. This suggests that MYBMIM may be a viable selective MYB inhibitor for the treatment of ACC patients [84]. Yusenko and colleagues showed that the polyether ionophore monensin A (referred to monensin) has an inhibitory effect on MYB and induces its degradation in vitro. Using MYB-NFIB mutation-positive ACC cells, the expression of MYB was suppressed under the administration of monensin. Under similar conditions, the expression of VEGF, which is typically induced by MYB, was also suppressed [86].

One drug already in clinical use has been shown to have an inhibitory effect on MYB.

Mandelbaum and collaborators demonstrated that All-trans Retinoic Acid (ATRA), clinically available for treatment of acute promyelocytic leukemia (APL), decreases c-MYB expression in myeloid leukemia cell via retinoic acid receptor (RAR). In a PDX model of MYB translocationpositive ACC, it was shown that ATRA and retinoic acid agonist suppresses tumor growth. They also examined apoptosis (cleaved caspase-3) and proliferation (Ki-67) in ACC xenograft tumors, and observed that ATRA treatment induced tumor cell death, but had no significant effect on tumor cell proliferation [87]. To verify these results in humans, a phase II trial of ATRA in Advanced Adenoid Cystic Carcinoma (NCT03999684) has been initiated (Table 3).

Immunomodulatory therapy to target MYB is another area of ongoing research. The TetMYB vaccine is a DNA vaccine targeting MYB. It was generated using a full-length MYB complementary DNA (cDNA) bound by two potent CD-4 epitopes derived from the tetanus toxin, which was then cloned into the FDA-compliant DNA vaccine vector pVAX1 [88]. Following several studies demonstrating that the TetMYB vaccine has a tumor suppressing effect for colorectal cancer [89, 90], a phase I clinical trial for colorectal carcinoma or adenoid cystic carcinoma is currently active [91] (NCT03287427) (**Table 3**).

Lastly, MYB has also been reported to be downregulated by inhibiting IGF1R signaling. In MYB-NFIB fusion-positive ACC cells, IGFR1/ EGFR/MET are consistently activated. These receptors stimulate proliferation of ACC cells through AKT signaling. Inhibition of these signaling events upon treatment with linsitinib, crizotinib, or gefitinib (IGF1R/INSR, MET, and EGFR inhibitors, respectively) significantly decreased tumor growth in ACC xenograft models [80]. Currently, there are two phase I clinical trials IGF1R targeting. One trial tests figitumab combined with the EGFR inhibitor dacomitinib, and the another uses R1507 combined with sorafenib. In those studies, 1 patient with partial response for 1.5 years and 3 patients with stable disease were observed. Although clinical development of figitumumab has been discontinued, these clinical studies support that IGF1R signaling has the potential to inhibit ACC growth and progression [92, 93] (Table 2).

NOTCH1 as a stemness target

NOTCH signaling is an evolutionarily conserved cell fate determinant pathway that regulates stem cells in many adult tissues, as well as in pathological conditions such as cancer [94]. More recent data have established a correlation between NOTCH1 signaling and cancer stemness. Higher expression of NOTCH1 was observed in CD133⁺ cells when compared to CD133⁻ and unsorted cells. Silencing of NOTCH1 suppresses spheroid formation in low attachment culture conditions [95]. NOTCH1 signaling was also shown to indicate poor patient prognosis. NOTCH1 mutation leads to its activation as demonstrated by a luciferase reporter assay bearing the promoter of HES1, which is a Notch1 transcriptional target. The NOTCH1 mutation was found in approximately 13-14% of ACC patients [9, 96]. Patients harboring NOTCH1 mutations showed more aggressive histologies with a solid subtype, shorter relapse-free survival, and shorter overall survival when compared with NOTCH1 wildtype tumors. Although distant metastases with organ involvement outside the lungs predicted poorer outcomes [97], patients with NOTCH1 mutation exhibited a higher likelihood of developing metastasis in the liver and/or bone [9].

The NOTCH-pathway can be targeted with pannotch inhibitors such as gamma-secretase inhibitors or Notch1 inhibitors. Preclinical studies demonstrated significant activity of Notch inhibitors (e.g. brontictuzumab, Notch1 inhibitor; AL101, pan-Notch inhibitor) in a NOTCH1 mutant ACC PDX. In both studies, brontictuzumab and AL101 had no significant effect on tumors lacking Notch1 activating mutations. For AL101, together with cisplatin or everolimus (mTOR inhibitor) was also studied, but this combination therapy had no additional benefit in Notch1 mutation-positive tumors [9, 98]. These preclinical studies suggested that when treating patients with Notch inhibitors, it may be more effective to select and treat patients who exhibit NOTCH1 mutations. There is some emerging data showing that Notch inhibitors benefit ACC patients. In a phase I study of BMS-986115 (pan-NOTCH inhibitor), some clinical benefits including 2 stable disease (SD) patients with ACC were shown. However, the study was terminated early [99]. In an expansion of a phase I study on crenigacestat (pan-

Notch-inhibitor), a cohort of 22 ACC patients was enrolled. From this cohort, 14 (64%) patients were positive for Notch by immunohistochemistry, but mutation status was not given. In the trial, 1 patient had an unconfirmed partial response (PR) (15%) while 15 patients showed SD [100] (Table 2). Treatment using brontictuzumab also demonstrated clinical benefit with 2 PR patients and 3 SD patients out of 12 patients enrolled in a phase I trial. Bronticutuzumab (specific Notch1 inhibitor), was expected to reduce systemic toxicity (e.g. diarrhea) related to Notch inhibition in the intestinal crypts. However, the frequency and grade of diarrhea was comparable with that reported with other pan-Notch inhibitors [101]. Currently, one phase II clinical trial with AL101 (Gamma Secretase Inhibitor) is recruiting ACC patients with NOTCH1 mutation (NCT03691207), and another phase I/IIA study with CB-103 (targeting assembly of the NOTCH transcription complex in the cell nucleus) is recruiting patients with advanced or metastatic solid tumors including ACC (NCT03422679) [102] (**Table 3**).

p53 as a stemness target

Mutations in the TP53 gene are the most frequently found in human cancer. Indeed, p53 is known as the "guardian of the genome" playing a key role in the determination of cell fate. Under normal conditions, p53 is under tight molecular regulation. However, once the gene is mutated, it is released from the degradation mechanism of p53 and accumulates rapidly in the cell to promote various transcription factors involved in senescence and apoptosis. Mouse double minute 2 (MDM2) is a major negative regulator of p53 that promotes degradation of p53 upon direct binding [103]. It is considered that malignant tumors survive because p53 itself does not have proper function due to an abnormality in itself or an abnormality in the binding to MDM2 [104].

There is some relationship between p53 and CSCs regarding stem cell self-renewal and differentiation [105]. In studies with mammary stem cells, p53 deficiency increased self-renewal capacity, increased the stem cell pool, and promoted symmetric division of cells. p53 knockout mice were found to have a higher proportion of cells capable of generating mam-

mospheres, supporting the concept that p53 plays an important role in the generation and maintenance of the cancer stem cell pool [106, 107]. In addition, p53 promotes differentiation by suppressing the expression of Nanog in mouse embryonic stem cells [108]. Given these findings, it is concluded that p53 controls the balance between self-renewal and differentiation of stem cells.

Although TP53 is mutated in many cancers, it is not as frequently mutated in salivary gland tumors when compared to other neoplasms [109]. In addition, high expression of p53 was noted in 19 out of 21 ACC cases (90%) via immunohistochemistry [61]. This indicates that increased accumulations of p53 might have a therapeutic effect within ACC. In fact, inhibition of the MDM2-p53 interaction with small molecule (MI-773) activates downstream effectors of apoptosis and causes tumor regression in PDX models of ACC [110]. MI-773 also sensitized ACC PDX tumors to cisplatin and successfully reduced the fraction of CSCs (ALDHhigh CD44^{high} cancer cells). Furthermore, inhibition of MDM2-p53 prevented tumor recurrence in preclinical trials. No recurrence was observed upon tumor resection after neoadjuvant administration of MI-773, whereas 63% of the mice in the control group showed recurrence [76]. Given these results, a Phase I/II trial of APG-115 (small molecule inhibitor of MDM2-p53) in patients with salivary gland tumors including ACC (NCT03781986) is currently ongoing (Table 3).

The epigenome as a stemness target

Treatments targeting the epigenome have been studied in many carcinomas, with ACC being no exception. Mutations related to the epigenome such as histone acetylation and methylation, as well as chromatin remodeling have been reported [111]. CBP/P300 described in MYB also has a histone acetyltransferase (HAT) function to acetylate H3K18 and H3K27 [112]. Histone deacetylase (HDAC), which has the opposite effect to HAT, has been shown to have an effect as a pro-oncogene [111]. A preclinical study showed that Vorinostat, an HDAC inhibitor (HDACi), has an effect of depressing CSC in ACC. In an in vitro study, Vorinostat reduced CSCs as identified by CD44 expression and ALDH activity in primary ACC cells.

Furthermore, combination treatment with cisplatin reduced the CSC fraction when compared with Vorinostat monotherapy. From these data, it was shown that Vorinostat demonstrates not only the tumor suppressive effect of a single agent but also the effect of sensitizing cells to cisplatin [113]. Vorinostat has also been tested in clinical trial. In a phase II study in which 90% (27/30) of the patients exhibited disease progression prior to enrollment, there were 2 patients out of 30 (7%) with PR and 27 patients with SD [114] (Table 2). Although the PR rate was inferior when compared with anti-VEGFR drugs such as Lenvatiniv, considering the results of preclinical studies, it is possible that the combination treatment with conventional chemotherapy may be effective. To answer this question, another phase II trial of combination therapy with HDACi (chidamide, orally active histone deacetylase inhibitor) and cisplatin is being conducted (NCT03639168) (Table 3). Chidamide has been approved for the treatment of relapsed and refractory peripheral T-cell lymphoma and is expected to have a tumor suppressing effect on various types of carcinoma [115].

The inhibitor of the protein arginine methyltransferase 5 (PRMT5) may also have a therapeutic effect in ACC. PRMTs are involved in various signal transduction cascades by catalyzing methylation of specific arginine residues. Overexpression of PRMT5 has been confirmed in many carcinomas and is thought to have a role in cancer progression by inhibiting tumor suppressor gene expression via methylation of transcription factors and chromatin associated proteins [116]. In oropharyngeal cancers, high expression of PRMT5 in the nucleus indicates poor prognosis. Notably, and it has been shown that IL-6 promotes PRMT5 translocation into the nucleus [117]. In breast cancer, PRMT5 was overexpressed in chemoresistant cell lines compared to non-resistant cell lines. Conversely, it was observed that knocking down PRMT5 enhances drug sensitivity. In addition, PRMT5 knockdown inhibits sphere formation and decreases the CSC fraction identified as CD44⁺CD22⁻ [118]. Therefore, it is likely that PRMT5 has an association with carcinogenesis and cancer stemness. Preclinical studies showing a relationship between salivary gland carcinoma and PRMT5 are inadequate, but clinical trials have shown some promise for ACC. Although it is an intermediate result of a Phase I trial, PR was confirmed in 3 out of 14 patients in the ACC patient group treated with GSK3326595, which is a PRMT5 inhibitors [119] (NCT02783300) (**Table 2**). In addition, another PRMT5 inhibitor (*i.e.* PRT543) is also undergoing phase I studies in patients with ACC (NCT03886831) (**Table 3**).

PI3K/Akt/mTOR pathway as a stemness target

Phosphatidylinositol 3-kinase (PI3K) activates Akt by phosphorylation. Furthermore, it activates transcriptional factors such as mammalian target of rapamycin (mTOR), an Akt substrate. Inhibition of mTOR has been shown to decrease expression of CSC markers and inhibit sphere formation in various cancer types including breast cancer and colorectal cancer. This indicates that inhibition of mTOR has a suppressive effect on CSCs [120]. In ACC, p-Akt is overexpressed when compared to normal salivary gland tissue, and therefore it may play a role in ACC carcinogenesis [121, 122]. However, high p-Akt expression was associated with lower grade histology, and it was even found that the p-Akt high-expressing group had a better prognosis [122]. In a phase II study of the mTOR inhibitor everolimus, 27 of 34 (79%) had SD, but no PR was seen [123]. Of note, nelfinavir (anti-HIV drug) has a tumor-suppressing effect by inhibiting Akt. As such, a phase II trial of nelfinavir was conducted in ACC, but only 7 out of 15 (46%) patients showed SD, none showed PR, and clinical prognosis was not affected [124] (Table 2).

Immunotherapy and cancer stemness

It is known that cancer cells can be eliminated by immune cells, creating a surveillance by the immune system that can inhibit tumor formation. However, tumorigenic cells might evade the immune surveillance and grow into tumors. CSCs are known to be able to evade immunological recognition and to exhibit tumorigenic potential. Immunosuppressive mechanisms have been characterized as key for tumor generation and progression, including programmed cell death 1 (PD-1), programmed cell death 1 ligand 1 (PD-L1), transforming growth factor β (TGF- β) and cytotoxic Tlymphocyte-associated antigen 4 (CTLA-4) [125]. Therapeutic efficacy of PD-1/PDL-1 inhibitors has been demonstrated in head and neck squamous cell carcinoma (HNSCC), with higher levels of PD-L1 expressed in the CD44^{high} CSCs group [126]. However, the therapeutic potential of inhibiting the PD-1/PD-L1 pathway in ACC remains unclear. A clinical trial of pembrolizumab (PD-1 inhibitor) was conducted, but no objective response was observed in ACC patients [127]. A combination of pembrolizumab and radiotherapy did not result in tumor regression [128]. Even with the combination therapy of vorinostat and pembrolizumab, the therapeutic effect was 8% (1 out of 12) in ACC, and no additional therapeutic effect for pembrolizumab was observed [129]. A potential reason for the lack of response to PD-1 inhibition might be because PD-L1 is rarely expressed in ACC and tumor-infiltrating lymphocytes (TILs) [130, 131]. Further, expression of PD-L2 alone was shown to aid in evasion of immune surveillance mechanisms [132]. PD-L2 expression tends to be relatively high in ACC [130, 131].

Emerging evidence suggests the possibility of developing anti-CSC vaccines. Cancer therapeutic vaccines activate a number of tumor antigen-specific cytotoxic T lymphocytes (CTLs) to eliminate the tumor by recognizing tumorassociated antigens [125]. Studies using the xenograft model of ovarian cancer have shown that the CD117⁺CD44⁺ CSC vaccine inhibits tumorigenicity [133]. A clinical trial of a MYBtargeted vaccine is being conducted in MYBexpressing tumors, such as ACC and colorectal cancer [91]. The results of this trial should shed light on the therapeutic potential of this new treatment modality.

Ongoing clinical trials

Intra-tumor therapy

When discussing immunotherapy, we mentioned anti-cancer vaccines. But vaccines using tumor-associated antigens may not elicit enough immunostimulatory action to evoke a sufficient immune response, which is required to have a strong and persistent tumoricidal effect. Most adjuvants have been developed for prophylactic vaccines and have a low Th1activating effect. However, CV8102, a non-coding, long-chain RNA molecule, is an adjuvant that stimulates TLR7/8 and induces a robust



Figure 2. Graphical illustration depicting cytotoxic chemotherapy, anti-angiogenic therapy, stemness-targeted therapy and immunotherapy as potential strategies for treatment of therapy Adenoid Cystic Carcinoma.

immune response [134, 135]. In a preclinical study using an HPV-related cervical cancer model, comparing the vaccine containing HPV-16 E7 protein-derived long peptides alone with the vaccine and CV8102 together, the combination group showed a longer and more potent anti-tumor effect. It also demonstrated a stronger immunostimulatory effect than the existing adjuvant, i.e. poly (I:C) [136]. Furthermore, it has been reported that intratumoral administration of adjuvants alone has an anti-tumor effect by presenting the tumor cell itself as an antigen to immune cells [137]. Intra-tumoral administration of adjuvants also showed increased activation of immune evasion mechanisms such as PD-L1. Therefore, it has been suggested that combination with PD-L1 inhibitors may enhance antitumor effects [138]. Based on this evidence, a clinical trial exploring a combination therapy that includes intra-tumoral administration of CV8102 and PD-1 inhibitor for solid tumors including ACC is ongoing (NCT03291002) (Table 3). In the mid-term report, complete regression was confirmed in one patient with melanoma, and tumor shrinkage was observed in one patient with head and neck squamous cell carcinoma and another in a patient with melanoma [139]. Considering the potential immunostimulatory effect, combination therapies involving anti-cancer vaccines may become a novel strategy for ACC.

TRKA

Perineural invasion, a characteristic invasion pattern of ACC, poses a major challenge in the treatment of patients with ACC [140]. Neural invasion is also a prognosis factor of local control rate [141]. It is believed that neural invasion is associated with nerve growth factor (NGF) and its receptor Tropomysin receptor kinase A (TrkA). In ACC, high expression of NGF and TrkA displays significant correlation with neural invasion and disseminated disease [142, 143]. A Phase I study of oral small-molecule TrkA inhibitor

has been conducted in many carcinomas including ACC is actively recruiting patients at this time (NCT03556228) [144] (**Table 3**).

Combination therapy

The observation that vorinostat enhanced the effect of cisplatin in preclinical studies [113], suggests that this drug combination may be effective in salivary gland cancer. The combination of either conventional chemotherapy with targeted therapies, or the combination of a molecular targeting drug with immunotherapy, are being actively pursued in clinical trials. The former is presented by combination therapies using HDACi and cisplatin (NCT03639-168), as well as APG-115 and carboplatin (NCT03781986), while the latter is presented by a combination therapy using lenvatinib and pembrolizumab (NCT04209660) (**Table 3**).

Conclusion

ACC is a rare disease found primarily in the salivary glands. Initial treatment for ACC is often surgery with or without radiation therapy, but most patients eventually experience tumor recurrence and/or metastasis. Chemotherapy is administered to control advanced, unresectable, recurrent and/or metastatic ACC, but a standard-of-care chemotherapy protocol is yet to be determined. Although inhibitors of tumor angiogenesis demonstrated relatively high effective response rates, they were at best comparable to conventional chemotherapy in patients with ACC. However, NOTCH signaling inhibitors and PRMT5 inhibitors have higher efficacy response rates and show promise as putative therapeutic targets in ACC. Furthermore, clinical studies are being conducted on cancer vaccines targeting MYB and small molecule inhibitors of the p53-MDM2p53 interaction. In addition, the emergence of new therapies targeting stemness and the maturation of immunotherapy protocols is expected to bring benefits to clinical patients in the future (Figure 2). The recent development of validated ACC cell lines and preclinical models that are suitable for developmental therapeutics studies, raise the speed of discovery and should enable discoveries towards safer and more effective therapies for ACC patients.

Acknowledgements

This work was funded by grants from Society for Promotion of International Oto-Rhino-Laryngology (SS) and the NIH/NIDCR grants F30-DE029097 (AEH), R01-DE021139 and R01-DE023220 (JEN).

Disclosure of conflict of interest

None.

Address correspondence to: Dr. Jacques E Nör, Dentistry and Otolaryngology, University of Michigan, 1011 N, University Rm, G049, Ann Arbor, Michigan 48109-1078, USA. Tel: 734-936-9300; E-mail: jenor@umich.edu

References

- [1] Spiro RH, Huvos AG and Strong EW. Adenoid cystic carcinoma of salivary origin: a clinicopathologic study of 242 cases. Am J Surg 1974; 128: 512-520.
- [2] Martelotto LG, De Filippo MR, Ng CK, Natrajan R, Fuhrmann L, Cyrta J, Piscuoglio S, Wen HC, Lim RS, Shen R, Schultheis AM, Wen YH, Edelweiss M, Mariani O, Stenman G, Chan TA, Colombo PE, Norton L, Vincent-Salomon A, Reis-Filho JS and Weigelt B. Genomic landscape of adenoid cystic carcinoma of the breast. J Pathol 2015; 237: 179-189.
- [3] Woida FM and Ribeiro-Silva A. Adenoid cystic carcinoma of the bartholin gland: an overview. Arch Pathol Lab Med 2007; 131: 796-798.

- [4] Wiseman SM, Popat SR, Rigual NR, Hicks WL, Orner JB, Wein RO, McGary CT and Loree TR. Adenoid cystic carcinoma of the paranasal sinuses or nasal cavity: a 40-year review of 35 cases. Ear Nose Throat J 2002; 81: 510-4, 516-7.
- [5] Goto H, Yamamoto T, Ishiyama Z, Usui M and Okada S. Adenoid cystic carcinoma arising from the lower eyelid. Jpn J Ophthalmol 2006; 50: 374-376.
- [6] Dodd RL and Slevin NJ. Salivary gland adenoid cystic carcinoma: a review of chemotherapy and molecular therapies. Oral Oncol 2006; 42: 759-769.
- [7] Khan AJ, Digiovanna MP, Ross DA, Sasaki CT, Carter D, Son YH and Haffty BG. Adenoid cystic carcinoma: a retrospective clinical review. Int J Cancer 2001; 96: 149-158.
- [8] Lim WS, Oh JS, Roh JL, Kim JS, Kim SJ, Choi SH, Nam SY and Kim SY. Prediction of distant metastasis and survival in adenoid cystic carcinoma using quantitative 18F-FDG PET/CT measurements. Oral Oncol 2018; 77: 98-104.
- [9] Ferrarotto R, Mitani Y, Diao L, Guijarro I, Wang J, Zweidler-McKay P, Bell D, William WN Jr, Glisson BS, Wick MJ, Kapoun AM, Patnaik A, Eckhardt G, Munster P, Faoro L, Dupont J, Lee JJ, Futreal A, El-Naggar AK and Heymach JV. Activating NOTCH1 mutations define a distinct subgroup of patients with adenoid cystic carcinoma who have poor prognosis, propensity to bone and liver metastasis, and potential responsiveness to notch1 inhibitors. J Clin Oncol 2017; 35: 352-360.
- [10] Chummun S, McLean NR, Kelly CG, Dawes PJDK, Fellows S, Meikle D and Soames JV. Adenoid cystic carcinoma of the head and neck. Br J Plast Surg 2001; 54: 476-480.
- [11] Chen AM, Bucci MK, Weinberg V, Garcia J, Quivey JM, Schechter NR, Phillips TL, Fu KK and Eisele DW. Adenoid cystic carcinoma of the head and neck treated by surgery with or without postoperative radiation therapy: prognostic features of recurrence. Int J Radiat Oncol Biol Phys 2006; 66: 152-159.
- [12] Garden AS, Weber RS, Morrison WH, Ang KK and Peters LJ. The influence of positive margins and nerve invasion in adenoid cystic carcinoma of the head and neck treated with surgery and radiation. Int J Radiat Oncol 1995; 32: 619-626.
- [13] Phuchareon J, Ohta Y, Woo JM, Eisele DW and Tetsu O. Genetic profiling reveals cross-contamination and misidentification of 6 adenoid cystic carcinoma cell lines: ACC2, ACC3, ACCM, ACCNS, ACCS and CAC2. PLoS One 2009; 4: e6040.
- [14] Moskaluk CA, Baras AS, Mancuso SA, Fan H, Davidson RJ, Dirks DC, Golden WL and Frier-

son HF. Development and characterization of xenograft model systems for adenoid cystic carcinoma. Lab Investig 2011; 91: 1480-1490.

- [15] Li J, Perlaky L, Rao P, Weber RS and El-Naggar AK. Development and characterization of salivary adenoid cystic carcinoma cell line. Oral Oncol 2014; 50: 991-999.
- [16] Warner KA, Oklejas AE, Pearson AT, Zhang Z, Wu W, Divi V, Rodriguez-Ramirez C, Castilho RM, Polverini PJ and Nör JE. UM-HACC-2A: MYB-NFIB fusion-positive human adenoid cystic carcinoma cell line. Oral Oncol 2018; 87: 21-28.
- [17] Gilbert J, Li Y, Pinto HA, Jennings T, Kies MS, Silverman P and Forastiere AA. Phase II trial of taxol in salivary gland malignancies (E1394): a trial of the Eastern cooperative oncology group. Head Neck 2006; 28: 197-204.
- [18] van Herpen CM, Locati LD, Buter J, Thomas J, Bogaerts J, Lacombe D, de Mulder P, Awada A, Licitra L, Bernier J and Vermorken JB. Phase II study on gemcitabine in recurrent and/or metastatic adenoid cystic carcinoma of the head and neck (EORTC 24982). Eur J Cancer 2008; 44: 2542-2545.
- [19] Schramm VL Jr, Srodes C and Myers EN. Cisplatin therapy for adenoid cystic carcinoma. Arch Otolaryngol 1981; 107: 739-741.
- [20] Licitra L, Marchini S, Spinazzè S, Rossi A, Rocca A, Grandi C and Molinari R. Cisplatin in advanced salivary gland carcinoma. A phase II study of 25 patients. Cancer 1991; 68: 1874-1877.
- [21] de Haan LD, De Mulder PH, Vermorken JB, Schornagel JH, Vermey A and Verweij J. Cisplatin-based chemotherapy in advanced adenoid cystic carcinoma of the head and neck. Head Neck 1992; 14: 273-277.
- [22] Tannock IF and Sutherland DJ. Chemotherapy for adenocystic carcinoma. Cancer 1980; 46: 452-454.
- [23] Verweij J, de Mulder PH, de Graeff A, Vermorken JB, Wildiers J, Kerger J, Schornagel J, Cognetti F, Kirkpatrick A, Sahmoud T and Lefebvre JL. Phase II study on mitoxantrone in adenoid cystic carcinomas of the head and neck. Ann Oncol 1996; 7: 867-869.
- [24] Mattox DE, Von Hoff DD and Balcerzak SP. Southwest oncology group study of mitoxantrone for treatment of patients with advanced adenoid cystic carcinoma of the head and neck. Invest New Drugs 1990; 8: 105-107.
- [25] Vermorken JB, Verweij J, de Mulder PH, Cognetti F, Clavel M, Rodenhuis S, Kirkpatrick A and Snow GB. Epirubicin in patients with advanced or recurrent adenoid cystic carcinoma of the head and neck: a phase II study of the EORTC head and neck cancer cooperative group. Ann Oncol 1993; 4: 785-788.

- [26] Belani CP, Eisenberger MA and Gray WC. Preliminary experience with chemotherapy in advanced salivary gland neoplasms. Med Pediatr Oncol 1988; 16: 197-202.
- [27] Dreyfuss AI, Clark JR, Fallon BG, Posner MR, Norris CM and Miller D. Cyclophosphamide, doxorubicin, and cisplatin combination chemotherapy for advanced carcinomas of salivary gland origin. Cancer 1987; 60: 2869-2872.
- [28] Creagan ET, Woods JE, Rubin J and Schaid DJ. Cisplatin-based chemotherapy for neoplasms arising from salivary glands and contiguous structures in the head and neck. Cancer 1988; 62: 2313-2319.
- [29] Licitra L, Cavina R, Grandi C, Di Palma S, Guzzo M, Demicheli R and Molinari R. Cisplatin, doxorubicin and cyclophosphamide in advanced salivary gland carcinoma. A phase II trial of 22 patients. Ann Oncol 1996; 7: 640-642.
- [30] Dimery IW, Legha SS, Shirinian M and Waun Ki Hong. Fluorouracil, doxorubicin, cyclophosphamide, and cisplatin combination chemotherapy in advanced or recurrent salivary gland carcinoma. J Clin Oncol 1990; 8: 1056-1062.
- [31] Triozzi PL, Brantley A, Fisher S, Cole TB, Crocker I and Huang AT. 5-fluorouracil, cyclophosphamide, and vincristine for adenoid cystic carcinoma of the head and neck. Cancer 1987; 59: 887-890.
- [32] Nakano K, Sato Y, Sasaki T, Shimbashi W, Fukushima H, Yonekawa H, Mitani H, Kawabata K and Takahashi S. Combination chemotherapy of carboplatin and paclitaxel for advanced/ metastatic salivary gland carcinoma patients: differences in responses by different pathological diagnoses. Acta Otolaryngol 2016; 136: 948-951.
- [33] Airoldi M, Fornari G, Pedani F, Marchionatti S, Gabriele P, Succo G and Bumma C. Paclitaxel and carboplatin for recurrent salivary gland malignancies. Anticancer Res 2000; 20: 3781-3783.
- [34] Airoldi M, Pedani F, Succo G, Gabriele AM, Ragona R, Marchionatti S and Bumma C. Phase II randomized trial comparing vinorelbine versus vinorelbine plus cisplatin in patients with recurrent salivary gland malignancies. Cancer 2001; 91: 541-547.
- [35] Airoldi M, Garzaro M, Pedani F, Ostellino O, Succo G, Riva G, Sensini M, Naqe N, Bellini E, Raimondo L and Pecorari G. Cisplatin + vinorelbine treatment of recurrent or metastatic salivary gland malignancies (RMSGM): a final report on 60 cases. Am J Clin Oncol 2017; 40: 86-90.
- [36] Hong MH, Kim CG, Koh YW, Choi EC, Kim J, Yoon SO, Kim HR and Cho BC. Efficacy and safety of vinorelbine plus cisplatin chemotherapy for patients with recurrent and/or meta-

static salivary gland cancer of the head and neck. Head Neck 2018; 40: 55-62.

- [37] Hill ME, Constenla DO, A'Hern RP, Henk JM, Rhys-Evans P, Breach N, Archer D and Gore ME. Cisplatin and 5-fluorouracil for symptom control in advanced salivary adenoid cystic carcinoma. Oral Oncol 1997; 33: 275-278.
- [38] Edwards PC, Bhuiya T and Kelsch RD. C-kit expression in the salivary gland neoplasms adenoid cystic carcinoma, polymorphous lowgrade adenocarcinoma, and monomorphic adenoma. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2003; 95: 586-593.
- [39] Pfeffer MR, Talmi Y, Catane R, Symon Z, Yosepovitch A and Levitt M. A phase II study of Imatinib for advanced adenoid cystic carcinoma of head and neck salivary glands. Oral Oncol 2007; 43: 33-36.
- [40] Bahl A, Panda NK, Elangovan A, Bakshi J, Verma R, Mohindra S, Gupta R, Oinam AS, Kaur S, Vashishta RK and Ghoshal S. Evaluation of multimodality management of adenoid cystic carcinoma of the head and neck. Indian J Otolaryngol Head Neck Surg 2019; 71 Suppl 1: 628-632.
- [41] Hotte SJ, Winquist EW, Lamont E, MacKenzie M, Vokes E, Chen EX, Brown S, Pond GR, Murgo A and Siu LL. Imatinib mesylate in patients with adenoid cystic cancers of the salivary glands expressing c-kit: a princess margaret hospital phase II consortium study. J Clin Oncol 2005; 23: 585-590.
- [42] Wong SJ, Karrison T, Hayes DN, Kies MS, Cullen KJ, Tanvetyanon T, Argiris A, Takebe N, Lim D, Saba NF, Worden FP, Gilbert J, Lenz HJ, Razak AR, Roberts JD, Vokes EE and Cohen EE. Phase II trial of dasatinib for recurrent or metastatic c-KIT expressing adenoid cystic carcinoma and for nonadenoid cystic malignant salivary tumors. Ann Oncol 2016; 27: 318-323.
- [43] Ghosal N, Mais K, Shenjere P, Julyan P, Hastings D, Ward T, Ryder WD, Bruce I, Homer J and Slevin NJ. Phase II study of cisplatin and imatinib in advanced salivary adenoid cystic carcinoma. Br J Oral Maxillofac Surg 2011; 49: 510-515.
- [44] Vered M, Braunstein E and Buchner A. Immunohistochemical study of epidermal growth factor receptor in adenoid cystic carcinoma of salivary gland origin. Head Neck 2002; 24: 632-636.
- [45] Jakob JA, Kies MS, Glisson BS, Kupferman ME, Liu DD, Lee JJ, El-Naggar AK, Gonzalez-Angulo AM and Blumenschein GR. Phase II study of gefitinib in patients with advanced salivary gland cancers. Head Neck 2015; 37: 644-649.
- [46] Locati LD, Bossi P, Perrone F, Potepan P, Crippa F, Mariani L, Casieri P, Orsenigo M, Losa M, Bergamini C, Liberatoscioli C, Quattrone P,

Calderone RG, Rinaldi G, Pilotti S and Licitra L. Cetuximab in recurrent and/or metastatic salivary gland carcinomas: a phase II study. Oral Oncol 2009; 45: 574-578.

- [47] Agulnik M, Cohen EW, Cohen RB, Chen EX, Vokes EE, Hotte SJ, Winquist E, Laurie S, Hayes DN, Dancey JE, Brown S, Pond GR, Lorimer I, Daneshmand M, Ho J, Tsao MS and Siu LL. Phase II study of lapatinib in recurrent or metastatic epidermal growth factor receptor and/or erbB2 expressing adenoid cystic carcinoma and non-adenoid cystic carcinoma malignant tumors of the salivary glands. J Clin Oncol 2007; 25: 3978-3984.
- [48] Zhang J, Peng B and Chen X. Expressions of nuclear factor κB, inducible nitric oxide synthese, and vascular endothelial growth factor in adenoid cystic carcinoma of salivary glands: correlations with the angiogenesis and clinical outcome. Clin Cancer Res 2005; 11: 7334-7343.
- [49] Persson M, Andrén Y, Mark J, Horlings HM, Persson F and Stenman G. Recurrent fusion of MYB and NFIB transcription factor genes in carcinomas of the breast and head and neck. Proc Natl Acad Sci U S A 2009; 106: 18740-18744.
- [50] Chen C, Choudhury S, Wangsa D, Lescott CJ, Wilkins DJ, Sripadhan P, Liu X, Wangsa D, Ried T, Moskaluk C, Wick MJ, Glasgow E, Schlegel R and Agarwal S. A multiplex preclinical model for adenoid cystic carcinoma of the salivary gland identifies regorafenib as a potential therapeutic drug. Sci Rep 2017; 7: 11410.
- [51] Ho AL, Sherman EJ, Baxi SS, Haque S, Ni A, Antonescu CR, Katabi N, Morris LG, Chan TA and Pfister DG. Phase II study of regorafenib in progressive, recurrent/metastatic adenoid cystic carcinoma. J Clin Oncol 2016; 34: 6096-6096.
- [52] Chau NG, Hotte SJ, Chen EX, Chin SF, Turner S, Wang L and Siu LL. A phase II study of sunitinib in recurrent and/or metastatic adenoid cystic carcinoma (ACC) of the salivary glands: current progress and challenges in evaluating molecularly targeted agents in ACC. Ann Oncol 2012; 23: 1562-1570.
- [53] Kim Y, Lee SJ, Lee JY, Lee SH, Sun JM, Park K, An HJ, Cho JY, Kang EJ, Lee HY, Kim J, Keam B, Kim HR, Lee KE, Choi MY, Lee KH and Ahn MJ. Clinical trial of nintedanib in patients with recurrent or metastatic salivary gland cancer of the head and neck: a multicenter phase 2 study (Korean Cancer Study Group HN14-01). Cancer 2017; 123: 1958-1964.
- [54] Locati LD, Galbiati D, Calareso G, Alfieri S, Singer S, Cavalieri S, Bergamini C, Bossi P, Orlandi E, Resteghini C, Platini F, Granata R, Quattrone P, Mancinelli M, Mariani L, Lo Vullo

S and Licitra LF. Patients with adenoid cystic carcinomas of the salivary glands treated with lenvatinib: activity and quality of life. Cancer 2020; 126: 1888-1894.

- [55] Tchekmedyian V, Sherman EJ, Dunn L, Tran C, Baxi S, Katabi N, Antonescu CR, Ostrovnaya I, Haque SS, Pfister DG and Ho AL. Phase II study of lenvatinib in patients with progressive, recurrent or metastatic adenoid cystic carcinoma. J Clin Oncol 2019; 37: 1529-1537.
- [56] Ho AL, Dunn L, Sherman EJ, Fury MG, Baxi SS, Chandramohan R, Dogan S, Morris LG, Cullen GD, Haque S, Sima CS, Ni A, Antonescu CR, Katabi N and Pfister DG. A phase II study of axitinib (AG-013736) in patients with incurable adenoid cystic carcinoma. Ann Oncol 2016; 27: 1902-1908.
- [57] Locati LD, Cavalieri S, Bergamini C, Resteghini C, Alfieri S, Calareso G, Bossi P, Perrone F, Tamborini E, Quattrone P, Granata R, Galbiati D, Platini F, Orlandi E, Mariani L and Licitra L. Phase II trial with axitinib in recurrent and/or metastatic salivary gland cancers of the upper aerodigestive tract. Head Neck 2019; 41: 3670-3676.
- [58] Thomson DJ, Silva P, Denton K, Bonington S, Mak SK, Swindell R, Homer J, Sykes AJ, Lee LW, Yap BK and Slevin NJ. Phase II trial of sorafenib in advanced salivary adenoid cystic carcinoma of the head and neck. Head Neck 2015; 37: 182-187.
- [59] Locati LD, Perrone F, Cortelazzi B, Bergamini C, Bossi P, Civelli E, Morosi C, Lo Vullo S, Imbimbo M, Quattrone P, Dagrada GP, Granata R, Resteghini C, Mirabile A, Alfieri S, Orlandi E, Mariani L, Saibene G, Pilotti S and Licitra L. A phase II study of sorafenib in recurrent and/or metastatic salivary gland carcinomas: translational analyses and clinical impact. Eur J Cancer 2016; 69: 158-165.
- [60] Nör JE, Christensen J, Mooney DJ and Polverini PJ. Vascular endothelial growth factor (VEGF)mediated angiogenesis is associated with enhanced endothelial cell survival and induction of Bcl-2 expression. Am J Pathol 1999; 154: 375-384.
- [61] Carlinfante G, Lazzaretti M, Ferrari S, Bianchi B and Crafa P. p53, bcl-2 and Ki-67 expression in adenoid cystic carcinoma of the palate. A clinico-pathologic study of 21 cases with long-term follow-up. Pathol Res Pract 2005; 200: 791-799.
- [62] Acasigua GA, Warner KA, Nör F, Helman J, Pearson AT, Fossati AC, Wang S and Nör JE. BH3-mimetic small molecule inhibits the growth and recurrence of adenoid cystic carcinoma. Oral Oncol 2015; 51: 839-847.
- [63] Myoken Y, Myoken Y, Okamoto T, Sato JD, Kan M, Mckeehan WL, Nakahara M and Takada K.

Immunohistochemical study of overexpression of fibroblast growth factor-1 (FGF-1), FGF-2, and FGF receptor-1 in human malignant salivary gland tumours. J Pathol 1996; 178: 429-436.

- [64] Miglarese MR, Halaban R and Gibson NW. Regulation of fibroblast growth factor 2 expression in melanoma cells by the c-MYB proto-oncoprotein. Cell Growth Differ 1997; 8: 1199-210.
- [65] Katoh M. Therapeutics targeting FGF signaling network in human diseases. Trends Pharmacol Sci 2016; 37: 1081-1096.
- [66] Doddapaneni R, Tao W, Naranjo A, Nikpoor N, Tse DT and Pelaez D. Fibroblast growth factor receptor 1 (FGFR1) as a therapeutic target in adenoid cystic carcinoma of the lacrimal gland. Oncotarget 2019; 10: 480-493.
- [67] Lee SH. In vivo target modulation and biological activity of CHIR-258, a multitargeted growth factor receptor kinase inhibitor, in colon cancer models. Clin Cancer Res 2005; 11: 3633-3641.
- [68] Keam B, Kim SB, Shin SH, Cho BC, Lee KW, Kim MK, Yun HJ, Lee SH, Yoon DH and Bang YJ. Phase 2 study of dovitinib in patients with metastatic or unresectable adenoid cystic carcinoma. Cancer 2015; 121: 2612-2617.
- [69] Dillon PM, Petroni GR, Horton BJ, Moskaluk CA, Fracasso PM, Douvas MG, Varhegyi N, Zaja-Milatovic S and Thomas CY. A phase II study of dovitinib in patients with recurrent or metastatic adenoid cystic carcinoma. Clin Cancer Res 2017; 23: 4138-4145.
- [70] Adams A, Warner K and Nör JE. Salivary gland cancer stem cells. Oral Oncol 2013; 49: 845-853.
- [71] Lawson DA, Bhakta NR, Kessenbrock K, Prummel KD, Yu Y, Takai K, Zhou A, Eyob H, Balakrishnan S, Wang CY, Yaswen P, Goga A and Werb Z. Single-cell analysis reveals a stem-cell program in human metastatic breast cancer cells. Nature 2015; 526: 131-135.
- [72] Cojoc M, Mäbert K, Muders MH and Dubrovska A. A role for cancer stem cells in therapy resistance: cellular and molecular mechanisms. Semin Cancer Biol 2015; 31: 16-27.
- [73] Sun S and Wang Z. ALDH high adenoid cystic carcinoma cells display cancer stem cell properties and are responsible for mediating metastasis. Biochem Biophys Res Commun 2010; 396: 843-848.
- [74] Keysar SB, Eagles JR, Miller B, Jackson BC, Chowdhury FN, Reisinger J, Chimed TS, Le PN, Morton JJ, Somerset HL, Varella-Garcia M, Tan AC, Song JI, Bowles DW, Reyland ME and Jimeno A. Salivary gland cancer patient-derived xenografts enable characterization of cancer stem cells and new gene events associated

with tumor progression. Clin Cancer Res 2018; 24: 2935-2943.

- [75] Fujita S and Ikeda T. Cancer stem-like cells in adenoid cystic carcinoma of salivary glands: relationship with morphogenesis of histological variants. J Oral Pathol Med 2012; 41: 207-213.
- [76] Nör F, Warner KA, Zhang Z, Acasigua GA, Pearson AT, Kerk SA, Helman JI, Sant'Ana Filho M, Wang S and Nör JE. Therapeutic inhibition of the MDM2-p53 interaction prevents recurrence of adenoid cystic carcinomas. Clin Cancer Res 2017; 23: 1036-1048.
- [77] Mitani Y, Liu B, Rao PH, Borra VJ, Zafereo M, Weber RS, Kies M, Lozano G, Futreal PA, Caulin C and El-Naggar AK. Novel MYBL1 gene rearrangements with recurrent MYBL1-NFIB fusions in salivary adenoid cystic carcinomas lacking t(6;9) translocations. Clin Cancer Res 2016; 22: 725-733.
- [78] Galoczova M, Coates P and Vojtesek B. STAT3, stem cells, cancer stem cells and p63. Cell Mol Biol Lett 2018; 23: 12.
- [79] West RB, Kong C, Clarke N, Gilks T, Lipsick JS, Cao H, Kwok S, Montgomery KD, Varma S and Le Q. MYB expression and translocation in adenoid cystic carcinomas and other salivary gland tumors with clinicopathologic correlation. Am J Surg Pathol 2011; 35: 92-99.
- [80] Andersson MK, Afshari MK, Andrén Y, Wick MJ and Stenman G. Targeting the oncogenic transcriptional regulator MYB in adenoid cystic carcinoma by inhibition of IGF1R/AKT signaling. J Natl Cancer Inst 2017; 109: 109.
- [81] Pattabiraman DR and Gonda TJ. Role and potential for therapeutic targeting of MYB in leukemia. Leukemia 2013; 27: 269-277.
- [82] Dai P, Akimaru H, Tanaka Y, Hou DX, Yasukawa T, Kanei-Ishii C, Takahashi T and Ishii S. CBP as a transcriptional coactivator of c-Myb. Genes Dev 1996; 10: 528-540.
- [83] Coulibaly A, Haas A, Steinmann S, Jakobs A, Schmidt TJ and Klempnauer KH. The natural anti-tumor compound Celastrol targets a Myb-C/ΕΒΡβ-p300 transcriptional module implicated in myeloid gene expression. PLoS One 2018; 13: e0190934.
- [84] Jiang Y, Gao R, Cao C, Forbes L, Li J, Freeberg S, Fredenburg KM, Justice JM, Silver NL, Wu L, Varma S, West R, Licht JD, Zajac-Kaye M, Kentsis A and Kaye FJ. MYB-activated models for testing therapeutic agents in adenoid cystic carcinoma. Oral Oncol 2019; 98: 147-155.
- [85] Ramaswamy K, Forbes L, Minuesa G, Gindin T, Brown F, Kharas MG, Krivtsov AV, Armstrong SA, Still E, De Stanchina E, Knoechel B, Koche R and Kentsis A. Peptidomimetic blockade of MYB in acute myeloid leukemia. Nat Commun 2018; 9: 110.

- [86] Yusenko MV, Trentmann A, Andersson MK, Ghani LA, Jakobs A, Arteaga Paz MF, Mikesch JH, Peter von Kries J, Stenman G and Klempnauer KH. Monensin, a novel potent MYB inhibitor, suppresses proliferation of acute myeloid leukemia and adenoid cystic carcinoma cells. Cancer Lett 2020; 479: 61-70.
- [87] Mandelbaum J, Shestopalov IA, Henderson RE, Chau NG, Knoechel B, Wick MJ and Zon LI. Zebrafish blastomere screen identifies retinoic acid suppression of MYB in adenoid cystic carcinoma. J Exp Med 2018; 215: 2673-2685.
- [88] Williams BB, Wall M, Miao RY, Williams B, Bertoncello I, Kershaw MH, Mantamadiotis T, Haber M, Norris MD, Gautam A, Darcy PK and Ramsay RG. Induction of T cell-mediated immunity using a c-Myb DNA vaccine in a mouse model of colon cancer. Cancer Immunol Immunother 2008; 57: 1635-1645.
- [89] Pham T, Carpinteri S, Sampurno S, Pereira L, Roth S, Narasimhan V, Darcy P, Desai J, Heriot AG and Ramsay RG. Novel vaccine targeting colonic adenoma: a pre-clinical model. J Gastrointest Surg 2019; 23: 626-633.
- [90] Cross RS, Malaterre J, Davenport AJ, Carpinteri S, Anderson RL, Darcy PK and Ramsay RG. Therapeutic DNA vaccination against colorectal cancer by targeting the MYB oncoprotein. Clin Transl Immunol 2015; 4: e30.
- [91] Pham T, Pereira L, Roth S, Galletta L, Link E, Akhurst T, Solomon B, Michael M, Darcy P, Sampurno S, Heriot A, Ramsay R and Desai J. First-in-human phase I clinical trial of a combined immune modulatory approach using Tet-MYB vaccine and anti-PD-1 antibody in patients with advanced solid cancer including colorectal or adenoid cystic carcinoma: the MYPHISMO study protocol (NCT03287427). Contemp Clin Trials Commun 2019; 16: 100409.
- [92] Calvo E, Soria JC, Ma WW, Wang T, Bahleda R, Tolcher AW, Gernhardt D, O'Connell J, Millham R, Giri N, Wick MJ, Adjei AA and Hidalgo M. A phase I clinical trial and independent patientderived xenograft study of combined targeted treatment with dacomitinib and figitumumab in advanced solid tumors. Clin Cancer Res 2017; 23: 1177-1185.
- [93] Mahadevan D, Sutton GR, Arteta-Bulos R, Bowden CJ, Miller PJ, Swart RE, Walker MS, Haluska P, Munster PN, Marshall J, Hamid O and Kurzrock R. Phase 1b study of safety, tolerability and efficacy of R1507, a monoclonal antibody to IGF-1R in combination with multiple standard oncology regimens in patients with advanced solid malignancies. Cancer Chemother Pharmacol 2014; 73: 467-473.
- [94] Lampreia FP, Carmelo JG and Anjos-Afonso F. Notch signaling in the regulation of hematopoi-

etic stem cell. Curr Stem Cell Rep 2017; 3: 202-209.

- [95] Panaccione A, Chang MT, Carbone BE, Guo Y, Moskaluk CA, Virk RK, Chiriboga L, Prasad ML, Judson B, Mehra S, Yarbrough WG and Ivanov SV. NOTCH1 and SOX10 are essential for proliferation and radiation resistance of cancer stem-like cells in adenoid cystic carcinoma. Clin Cancer Res 2016; 22: 2083-2095.
- [96] Stephens PJ, Davies HR, Mitani Y, Van Loo P, Shlien A, Tarpey PS, Papaemmanuil E, Cheverton A, Bignell GR, Butler AP, Gamble J, Gamble S, Hardy C, Hinton J, Jia M, Jayakumar A, Jones D, Latimer C, McLaren S, McBride DJ, Menzies A, Mudie L, Maddison M, Raine K, Nik-Zainal S, O'Meara S, Teague JW, Varela I, Wedge DC, Whitmore I, Lippman SM, McDermott U, Stratton MR, Campbell PJ, El-Naggar AK and Futreal PA. Whole exome sequencing of adenoid cystic carcinoma. J Clin Invest 2013; 123: 2965-2968.
- [97] Hanna GJ, Bae JE, Lorch JH, Schoenfeld JD, Margalit DN, Tishler RB, Haddad RI and Chau NG. Long-term outcomes and clinicogenomic correlates in recurrent, metastatic adenoid cystic carcinoma. Oral Oncol 2020; 106: 104690.
- [98] Ferrarotto R, Alpert G, Gluschnaider U, Rauch R, Mondshine A, Solomon O, Kramer B, Izumchenko E, Heymach J, Vergara-Silva A, Aster J and Davis M. Abstract 4885: AL101 mediated tumor inhibition in notch mutated ACC PDX models. Clin Res (Excluding Clin Trials) 2019; 79: 4885-4885.
- [99] Aung KL, El-Khoueiry AB, Gelmon K, Tran B, Bajaj G, He B, Chen T, Zhu L, Poojary S, Basak S, Qi Z, Spreafico A, Fischer BS and Desai J. A multi-arm phase I dose escalating study of an oral NOTCH inhibitor BMS-986115 in patients with advanced solid tumours. Invest New Drugs 2018; 36: 1026-1036.
- [100] Even C, Lassen U, Merchan J, Le Tourneau C, Soria JC, Ferte C, Ricci F, Diener JT, Yuen E, Smith C, Oakley GJ, Benhadji KA and Massard C. Safety and clinical activity of the Notch inhibitor, crenigacestat (LY3039478), in an open-label phase I trial expansion cohort of advanced or metastatic adenoid cystic carcinoma. Invest New Drugs 2020; 38: 402-409.
- [101] Ferrarotto R, Eckhardt G, Patnaik A, LoRusso P, Faoro L, Heymach JV, Kapoun AM, Xu L and Munster P. A phase I dose-escalation and dose-expansion study of brontictuzumab in subjects with selected solid tumors. Ann Oncol 2018; 29: 1561-1568.
- [102] Perez Garcia JM, Cortés J, Stathis A, Mous R, López-Miranda E, Azaro A, Genta S, Nuciforo P, Vivancos A, Ferrarotto R, Bertoni F, Rossi D, Spardy Burr N, Schönborn-Kellenberger O, Jor-

ga K, Beni L, Lehal R, Bauer M, Weber D and Garralda E. First-in-human phase 1-2A study of CB-103, an oral protein-protein interaction inhibitor targeting pan-NOTCH signalling in advanced solid tumors and blood malignancies. J Clin Oncol 2018; 36: TPS2619.

- [103] Kubbutat MH, Jones SN and Vousden KH. Regulation of p53 stability by Mdm2. Nature 1997; 387: 299-303.
- [104] Marine JC, Francoz S, Maetens M, Wahl G, Toledo F and Lozano G. Keeping p53 in check: essential and synergistic functions of Mdm2 and Mdm4. Cell Death Differ 2006; 13: 927-934.
- [105] Rodriguez-Ramirez C and Nör JE. p53 and cell fate: sensitizing head and neck cancer stem cells to chemotherapy. Crit Rev Oncog 2018; 23: 173-187.
- [106] Cicalese A, Bonizzi G, Pasi CE, Faretta M, Ronzoni S, Giulini B, Brisken C, Minucci S, Di Fiore PP and Pelicci PG. The tumor suppressor p53 regulates polarity of self-renewing divisions in mammary stem cells. Cell 2009; 138: 1083-1095.
- [107] Tao L, Roberts AL, Dunphy KA, Bigelow C, Yan H and Jerry DJ. Repression of mammary stem/ progenitor cells by p53 is mediated by notch and separable from apoptotic activity. Stem Cells 2011; 29: 119-127.
- [108] Lin T, Chao C, Saito S, Mazur SJ, Murphy ME, Appella E and Xu Y. p53 induces differentiation of mouse embryonic stem cells by suppressing Nanog expression. Nat Cell Biol 2005; 7: 165-171.
- [109] Gomes CC, Diniz MG, Orsine LA, Duarte AP, Fonseca-Silva T, Conn BI, de Marco L, Pereira CM and Gomez RS. Assessment of TP53 mutations in benign and malignant salivary gland neoplasms. PLoS One 2012; 7: e41261.
- [110] Warner KA, Nör F, Acasigua GA, Martins MD, Zhang Z, McLean SA, Spector ME, Chepeha DB, Helman J, Wick MJ, Moskaluk CA, Castilho RM, Pearson AT, Wang S and Nör JE. Targeting MDM2 for treatment of adenoid cystic carcinoma. Clin Cancer Res 2016; 22: 3550-3559.
- [111] Roy DM, Walsh LA and Chan TA. Driver mutations of cancer epigenomes. Protein Cell 2014; 5: 265-296.
- [112] Jin Q, Yu LR, Wang L, Zhang Z, Kasper LH, Lee JE, Wang C, Brindle PK, Dent SY and Ge K. Distinct roles of GCN5/PCAF-mediated H3K9ac and CBP/p300-mediated H3K18/27ac in nuclear receptor transactivation. EMBO J 2011; 30: 249-262.
- [113] Almeida LO, Guimarães DM, Martins MD, Martins MAT, Warner KA, Nör JE, Castilho RM and Squarize CH. Unlocking the chromatin of adenoid cystic carcinomas using HDAC inhibitors

sensitize cancer stem cells to cisplatin and induces tumor senescence. Stem Cell Res 2017; 21: 94-105.

- [114] Goncalves PH, Heilbrun LK, Barrett MT, Kummar S, Hansen AR, Siu LL, Piekarz RL, Sukari AW, Chao J, Pilat MJ, Smith DW, Casetta L, Boerner SA, Chen A, Lenkiewicz E, Malasi S and LoRusso PM. A phase 2 study of vorinostat in locally advanced, recurrent, or metastatic adenoid cystic carcinoma. Oncotarget 2017; 8: 32918-32929.
- [115] Chan T, Tse E and Kwong YL. Chidamide in the treatment of peripheral T-cell lymphoma. Onco Targets Ther 2017; 10: 347-352.
- [116] Shailesh H, Zakaria ZZ, Baiocchi R and Sif S. Protein arginine methyltransferase 5 (PRMT5) dysregulation in cancer. Oncotarget 2018; 9: 36705-36718.
- [117] Kumar B, Yadav A, Brown NV, Zhao S, Cipolla MJ, Wakely PE, Schmitt AC, Baiocchi RA, Teknos TN, Old M and Kumar P. Nuclear PRMT5, cyclin D1 and IL-6 are associated with poor outcome in oropharyngeal squamous cell carcinoma patients and is inversely associated with p16-status. Oncotarget 2017; 8: 14847-14859.
- [118] Wang Z, Kong J, Wu Y, Zhang J, Wang T, Li N, Fan J, Wang H, Zhang J and Ling R. PRMT5 determines the sensitivity to chemotherapeutics by governing stemness in breast cancer. Breast Cancer Res Treat 2018; 168: 531-542.
- [119] Siu LL, Rasco DW, Vinay SP, Romano PM, Menis J, Opdam FL, Heinhuis KM, Egger JL, Gorman SA, Parasrampuria R, Wang K, Kremer BE and Gounder MM. METEOR-1: a phase I study of GSK3326595, a first-in-class protein arginine methyltransferase 5 (PRMT5) inhibitor, in advanced solid tumours. Ann Oncol 2019; 30: v159.
- [120] Xia P and Xu XY. PI3K/Akt/mTOR signaling pathway in cancer stem cells: from basic research to clinical application. Am J Cancer Res 2015; 5: 1602-1609.
- [121] Branco KFCC, Ribeiro ALR, de Mendonça RP, de Jesus Viana Pinheiro J, da Silva Kataoka MS, Arnaud MVC and de Melo Alves Junior S. Abnormal activation of the Akt signaling pathway in adenoid cystic carcinoma. Eur Arch Otorhinolaryngology 2018; 275: 3039-3047.
- [122] Ouyang D, Liang L, Ke Z, Zheng G, Weng D, Yang W, Su Y and Liao G. Association between high expression of phosphorylated Akt and mammalian target of rapamycin and improved survival in salivary gland adenoid cystic carcinoma. Head Neck 2017; 39: 1145-1154.
- [123] Kim DW, Oh DY, Shin SH, Kang JH, Cho BC, Chung JS, Kim HJ, Park KU, Kwon JH, Han JY, Kim MJ and Bang YJ. A multicenter phase II study of everolimus in patients with progres-

sive unresectable adenoid cystic carcinoma. BMC Cancer 2014; 14: 795.

- [124] Hoover AC, Milhem MM, Anderson CM, Sun W, Smith BJ, Hoffman HT and Buatti JM. Efficacy of nelfinavir as monotherapy in refractory adenoid cystic carcinoma: results of a phase II clinical trial. Head Neck 2015; 37: 722-726.
- [125] Sultan M, Coyle KM, Vidovic D, Thomas ML, Gujar S and Marcato P. Hide-and-seek: the interplay between cancer stem cells and the immune system. Carcinogenesis 2017; 38: 107-118.
- [126] Lee Y, Shin JH, Longmire M, Wang H, Kohrt HE, Chang HY and Sunwoo JB. Cd44+ cells in head and neck squamous cell carcinoma suppress t-cell-mediated immunity by selective constitutive and inducible expression of PD-L1. Clin Cancer Res 2016; 22: 3571-3581.
- [127] Cohen RB, Delord JP, Doi T, Piha-Paul SA, Liu SV, Gilbert J, Algazi AP, Damian S, Hong RL, Le Tourneau C, Day D, Varga A, Elez E, Wallmark J, Saraf S, Thanigaimani P, Cheng J and Keam B. Pembrolizumab for the treatment of advanced salivary gland carcinoma. Am J Clin Oncol 2018; 41: 1083-1088.
- [128] Schoenfeld JD, Mahmood U, Chen YH, Mak RH, Lorch JH, Hanna GJ, Sridharan V, Bang A, Busse PM, Willers H, Mamon HJ, Yoo HJ, Pai SI, Wirth LJ, Haddad RI and Chau NG. A randomized phase II study of pembrolizumab with or without radiation in patients with recurrent or metastatic adenoid cystic carcinoma. J Clin Oncol 2019; 37: 6082.
- [129] Rodriguez CP, Wu QV, Voutsinas J, Fromm JR, Jiang X, Pillarisetty VG, Lee SM, Santana-Davila R, Goulart B, Baik CS, Chow LQM, Eaton K and Martins R. A phase II trial of pembrolizumab and vorinostat in recurrent metastatic head and neck squamous cell carcinomas and salivary gland cancer. Clin Cancer Res 2020; 26: 837-845.
- [130] Nakano T, Takizawa K, Uezato A, Taguchi K, Toh S and Masuda M. Prognostic value of programed death ligand-1 and ligand-2 co-expression in salivary gland carcinomas. Oral Oncol 2019; 90: 30-37.
- [131] Mosconi C, de Arruda JAA, de Farias ACR, Oliveira GAQ, de Paula HM, Fonseca FP, Mesquita RA, Silva TA, Mendonça EF and Batista AC. Immune microenvironment and evasion mechanisms in adenoid cystic carcinomas of salivary glands. Oral Oncol 2019; 88: 95-101.
- [132] Tanegashima T, Togashi Y, Azuma K, Kawahara A, Ideguchi K, Sugiyama D, Kinoshita F, Akiba J, Kashiwagi E, Takeuchi A, Irie T, Tatsugami K, Hoshino T, Eto M and Nishikawa H. Immune suppression by PD-L2 against spontaneous and treatment-related antitumor immunity. Clin Cancer Res 2019; 25: 4808-4819.

- [133] Wu D, Wang J, Cai Y, Ren M, Zhang Y, Shi F, Zhao F, He X, Pan M, Yan C and Dou J. Effect of targeted ovarian cancer immunotherapy using ovarian cancer stem cell vaccine. J Ovarian Res 2015; 8: 68.
- [134] Circelli L, Petrizzo A, Tagliamonte M, Heidenreich R, Tornesello ML, Buonaguro FM and Buonaguro L. Immunological effects of a novel RNA-based adjuvant in liver cancer patients. Cancer Immunol Immunother 2017; 66: 103-112.
- [135] Roers A, Hiller B and Hornung V. Recognition of endogenous nucleic acids by the innate immune system. Immunity 2016; 44: 739-754.
- [136] Heidenreich R, Jasny E, Kowalczyk A, Lutz J, Probst J, Baumhof P, Scheel B, Voss S, Kallen KJ and Fotin-Mleczek M. A novel RNA-based adjuvant combines strong immunostimulatory capacities with a favorable safety profile. Int J Cancer 2015; 137: 372-384.
- [137] Kyi C, Roudko V, Sabado R, Saenger Y, Loging W, Mandeli J, Thin TH, Lehrer D, Donovan M, Posner M, Misiukiewicz K, Greenbaum B, Salazar A, Friedlander P and Bhardwaj N. Therapeutic immune modulation against solid cancers with intratumoral poly-ICLC: a pilot trial. Clin Cancer Res 2018; 24: 4937-4948.
- [138] Mullins SR, Vasilakos J, Deschler K, Grigsby I, Ren S, Elder MJ, Dovedi SJ, Leishman AJ, Ryan P, Cooper Z, Elvecrog J, Herbst R, Kumar R, Tomai M and Wilkinson RW. Intratumoral immunotherapy with TLR7/8 agonist MEDI9197 modulates the tumor microenvironment leading to enhanced activity when combined with other immunotherapies. J Immunology Cancer 2019; 7: 244.
- [139] Eigentler T, Krauss J, Schreiber J, Weishaupt C, Terheyden P, Heinzerling L, Mohr P, Weide B, Gutzmer R, Becker JC, Kiecker F, Daehling A, Funkner F, Heidenreich R, Kays SK, Klinkhardt U, Scheel B, Schoenborn-Kellenberger O, Seibel T, Stosnach C, Strack T and Gnad-Vogt U. Abstract LB-021: intratumoral RNA-based TLR-7/-8 and RIG-I agonist CV8102 alone and in combination with anti-PD-1 in a Phase I doseescalation and expansion trial in patients with advanced solid tumors. Cancer Res 2019; 79: LB-021 LP-LB-021.

- [140] Barrett AW and Speight PM. Perineural invasion in adenoid cystic carcinoma of the salivary glands: a valid prognostic indicator? Oral Oncol 2009; 45: 936-940.
- [141] Mendenhall WM, Morris CG, Amdur RJ, Werning JW, Hinerman RW and Villaret DB. Radiotherapy alone or combined with surgery for adenoid cystic carcinoma of the head and neck. Head Neck 2004; 26: 154-162.
- [142] Kobayashi K, Ando M, Saito Y, Kondo K, Omura G, Shinozaki-Ushiku A, Fukayama M, Asakage T and Yamasoba T. Nerve growth factor signals as possible pathogenic biomarkers for perineural invasion in adenoid cystic carcinoma. Otolaryngol Neck Surg 2015; 153: 218-224.
- [143] Hao L, Xiao-lin N, Qi C, Yi-ping Y, Jia-quan L and Yan-ning L. Nerve growth factor and vascular endothelial growth factor: retrospective analysis of 63 patients with salivary adenoid cystic carcinoma. Int J Oral Sci 2010; 2: 35-44.
- [144] Chung V, Wang L, Fletcher MS, Massarelli E, Reckamp KL, Cristea MC, Prajapati N, Parikh A, Whiting RL, Wang M and Wu J. First-time inhuman study of VMD-928, an allosteric and irreversible TrkA selective inhibitor, in patients with solid tumors or lymphoma. J Clin Oncol 2019; 37: TPS3146.