

Review Article

Progress and challenges of multidrug resistance proteins in diseases

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Received June 9, 2022; Accepted October 9, 2022; Epub October 15, 2022; Published October 30, 2022

Abstract: Chemotherapy remains the first choice for patients with advanced cancers when other treatments are ineffective. Multidrug resistance (MDR) is an unavoidable factor that negatively affects the effectiveness of cancer chemotherapy drugs. Researchers are trying to reduce MDR, improve the effectiveness of chemotherapeutic drugs, and alleviate patient suffering to positively contribute to disease treatment. MDR also occurs in inflammation and genetic disorders, which increases the difficulty of clinically beneficial treatments. The ATP-binding cassette (ABC) is an active transporter that plays an important role in the barrier and secretory functions of many normal cells. As the C subfamily in the ABC family, multidrug resistance proteins (MRPs/ABCCs) export a variety of antitumour drugs and are expressed in a variety of cancers. The present review summarises the role of MRPs in cancer and other diseases and recent research progress of MRP inhibitors to better examine the mechanism and function of MRPs, and establish a good relationship with clinical treatment.

Keywords: Multidrug resistance protein, ATP-binding cassette, cancer, hereditary diseases

Introduction

Chemotherapy is a primary method in advanced cancer treatment, and the emergence of multidrug resistance (MDR) has become a major obstacle to tumour treatment. MDR describes the resistance of tumour cells to different drugs with multiple chemical structures, multiple mechanisms of action and multiple targets, and it is classified as intrinsic or acquired resistance [1, 2]. MDR is characterized by reduced intracellular concentrations of chemotherapeutic drugs and limited efficacy [3, 4]. MDR is a pathophysiological phenomenon within tumour cells involving multiple mechanisms, including tumour factors, host factors and tumour-host interactions [5, 6]. Other mechanisms of MDR are related to the activation of detoxification systems, cell proliferation, activation of DNA repair mechanisms, increased drug excretion or impaired drug uptake, loss of pro-apoptotic factors in nuclear factor kappa B (NF- κ B) factors, or overexpression of anti-apoptotic factors [7-10] (**Figure 1**). The phosphatidylinositol 3-kinase (PI3K)/pro-

tein kinase B (AKT) pathway is involved in the regulation of multiple targets, such as the expression of the ATP-binding cassette (ABC) transporters and NF- κ B. It is also a key link in the regulation of MDR and manifests in breast, ovarian and hepatocellular carcinoma (HCC) [11, 12]. Lysosomes also promote hydrophobic anticancer drugs by promoting weak alkaline efflux, and ABC transporters likely localize to lysosomes involved in multidrug resistance [13].

However, one of the common mechanisms of MDR is the promotion of energy-dependent efflux of multiple hydrophobic cytotoxic drugs, and ABC transporters play an important role in this process [14, 15]. Overexpression of the ABC transporter uses the energy provided by adenosine triphosphate (ATP) hydrolysis to expel a variety of cytotoxic drugs from cancer cells, which results in significantly reduced efficacy and the promotion of MDR [16, 17], and it leads to a decrease in the cellular uptake of drugs and an increase in drug pumping, which result in a decrease in intracellular drug con-

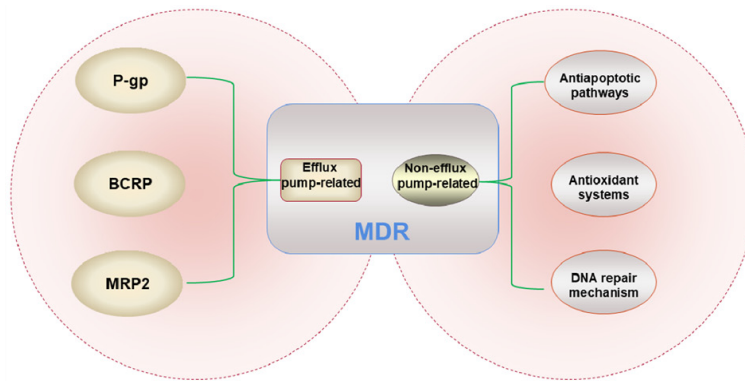


Figure 1. The mechanism of multidrug resistance. MDR are divided into two main categories, namely efflux pump-related and non-ex efflux pump-related. ABC transporter proteins are the main class of identified drug efflux pumps, mainly including P-gp, MRP2 and BCRP. While the main non-ex efflux pump modalities of MDR are antiapoptotic pathways, antioxidant systems and DNA repair mechanisms.

centration and the failure to achieve the desired effect [18, 19]. As mentioned in our previous review, the ABC transporter is activated by ATP and regulates lipid metabolism at the plasma membrane by regulating the import and export of transmembrane substances involved in atherosclerosis, metabolic diseases, atherosclerosis, coronary artery disease and skin diseases [20].

As a subfamily of ABCs, Multidrug resistance proteins (MRPs) mediate MDR in cancer by secreting various chemotherapeutic drugs or their metabolites from tumour cells [21-23]. MRPs are energy-dependent “drug pump” transmembrane glycoproteins with a broad-spectrum of substrates. MRPs non-directionally exclude intracellular drugs and prevent target-to-target interactions. Drug combinations are the direct cause of multidrug resistance in tumours [24]. Unlike other proteins of the ABC family, MRPs have unique advantages in acquiring multidrug resistance [25]. MRPs are associated with many cancers, but the exact mechanisms are not clear. We reviewed the specific mechanisms of MRP activity in various cancers and summarised the mechanisms of MRP action in other diseases and inhibitors linked to MRPs to highlight the importance of MRPs and provide novel suggestions for clinical treatment.

MRPs belong to the family of ABC transporters

Azad et al. [26] cloned an MRP in 1992 from an H69/AR-resistant cell line and overexpressed it

in doxorubicin-resistant H69-AR cells. It is a resistance-related gene functionally similar to P-glycoprotein (P-gp). The MRP family has 13 members, nine of which are associated with tumour resistance: MRP1/ABCC1, MRP2/ABCC2, MRP3/ABCC3, MRP4/ABCC4, MRP5/ABCC5, MRP6/ABCC6, MRP7/ABCC10, MRP8/ABCC11 and MRP9/ABCC12. These nine MRPs use ATP for energy and the transport of endogenous substances and chemotherapeutic drugs. The structure is similar to P-gp, with two transmembrane regions and two ATP-binding domains.

Members of the MRP family are divided into two categories. MRP4, MRP5, MRP8, and MRP9 with typical ABC structures (ABCC4, ABCC5, ABCC11, and ABCC12) are “short-term” MRPs, and MRP1, MRP2, MRP3, MRP6, and MRP7 (ABCC1, ABCC2, ABCC3, ABCC6 and ABCC10) are called “long-term” MRPs [27]. The MRP family is widely distributed with localisation to the apical and/or basolateral membranes of hepatocytes, enterocytes, renal proximal tubular cells and blood-brain barrier endothelial cells, and these proteins are expressed to varying degrees in the liver, kidney, intestine, brain and other tissues [28].

The structure, localisation and family of MRPs

Related information on the structure, localisation and family of MRP1-9 is summarised below and in **Table 1** and **Figure 2**. The molecular structures of MRP1-9 are shown in **Figure 3**, and protein sequences from the unified protein database are described in detail in the [Supplementary Materials](#).

MRP1 was originally found in the human lung cancer cell lines H69 and HL60 [29, 30]. The gene is located on the first chromosome at band 13.1 of the long arm of human chromosome 16. The molecular weight of the MRP1 protein is 190 kDa, and it contains 1531 amino acids and consists of 3 TMDs, 2 NBDs and 1 N-terminal intracellular junction region (LO) (arranged as TMD0-LO-TMD1-NBD1-TMD2-NBD2). NBD participates in the binding and hydrolysis of ATP to provide energy for transport and functions as an antineoplastic drug outflow

The role of MRPs in diseases

Table 1. The information of the MRP family

MRPs	Amino add sequence length	Chromosome location	Related diseases	Ref
MRP1 (ABCC1)	1531	16p13.12	Hepatocellular, esophagus cancer, non-small cell lung cancer, Ovarian cancer	[30, 61-64, 71-73, 111-113]
MRP2 (ABCC2)	1545	10q24.2	Colorectal cancer, Durbin-Johnson syndrome, Hepatocellular carcinoma, Acute myeloid leukemia	[39, 71, 72, 95-97, 104, 115-117]
MRP3 (ABCC3)	1527	17q21.33	Hepatocellular carcinoma, Ovarian cancer, Adult acute lymphoblastic leukemia	[39, 71, 72, 104, 111-113]
MRP4 (ABCC4)	1325	13q32.1	Prostatic cancer, Neuroblastoma, Acute myeloid leukemia, Pancreatic cancer, Breast cancer, Ovarian cancer	[39, 76, 79, 104, 107, 111-113]
MRP5 (ABCC5)	1437	3q27.1	Lung cancer, Colorectal cancer, Breast cancer, Pancreatic cancer	[61-64, 76, 77, 79, 95-97, 107]
MRP6 (ABCC6)	1503	16p13.12	Pseudo-polychordoma	[118]
MRP7 (ABCC10)	1492	6p21.1	Salivary gland carcinomas	[58]
MRP8 (ABCC11)	1382	16q12.1	Breast cancer, Ovarian cancer, Pancreatic cancer	[76, 77, 79, 107, 111-113]
MRP9 (ABCC12)	1356	16q12.1	Lung cancer, Colorectal cancer	[61-64, 92, 93]

Abbreviations: MRP, Multidrug Resistance Protein; ABC, ATP-Binding Cassette.

pump. TMD is responsible for substrate identification, binding and transport [31]. MRP1 is extensively visualised in various tissues, such as lung, testis, kidney, bone and heart muscle, placenta and macrophages. It is also located in blood-tissue barriers, such as the basement membrane of choroid plexus cells in the blood-cerebrospinal fluid barrier and the trophoblast membrane of bronchial epithelial cells and placental apical commissures [32]. Cells overexpressing MRP1 exhibit low levels of resistance to lycopene, colchicine and vincristine, and moderate resistance to anthracyclines (e.g. etoposide) and vincristine (e.g. pergolide). MRP1 transports a variety of endogenous compounds, such as heavy metal anions, glutathione, glutathione-bound leukotrienes, glucuronic acid and its sulphate conjugates [33]. Because of the dependence of this protein on ATP, glutathione stimulates the transport of various unbound hydrophobic substrates and certain glucosinolate and sulphate conjugates that are resistant to natural product drugs such as periwinkle and epithelial phytotoxins [34].

MRP2 was originally found in human and rat hepatocytes, and it is also called cholinergic MRP (cMRP)/cMOAT. The gene is found on chromosome 10q24, and the protein consists of a total of 1545 amino acids with three MSDs and two NBDs. It is localised to the proximal tubular apical membrane of the kidney, gallbladder epithelial cells, small intestinal epithelial cells,

colonic epithelial cells and pulmonary epithelial cells, which is different from other members of the MRP family [22]. Some studies showed that MRP2 led to functional deficiency due to natural mutations, which is associated with Doberman syndrome and associated hyperbilirubinemia, and MRP3 was also involved [35]. Translocation of bilirubin and glucose from the liver to the bile is one of the physiological functions of MRP2. Excretion and protection are the main functions of MRP2 [36]. MRP2 regulates the pharmacokinetics of many drugs by transporting some chemotherapeutic agents, such as antibiotics and heavy metals, and various compounds, specifically lipids bound to glutathione, glucuronide and sulphate, which play large roles in detoxification and chemoprotection [37].

MRP3/MOAT-D is located on chromosome 17q21.3, and it is expressed in the adrenals, small intestine, duodenum, colon, pancreas and gallbladder. There is a high level of MRP3 expression in the liver, small intestine and colon but a low level in the lung, spleen, kidney, stomach and tonsils [38, 39]. MRP3 is an organic ion transporter that transports MRP1 and MRP2 and transport lipid anti-ions such as bile acid and glucagon, but it has poor affinity for GSH-binding compounds [40]. MRP3 is also responsible for the transport of curcumin-O-glucosamine, which is the main metabolite of curcumin in plasma and faeces, from hepato-

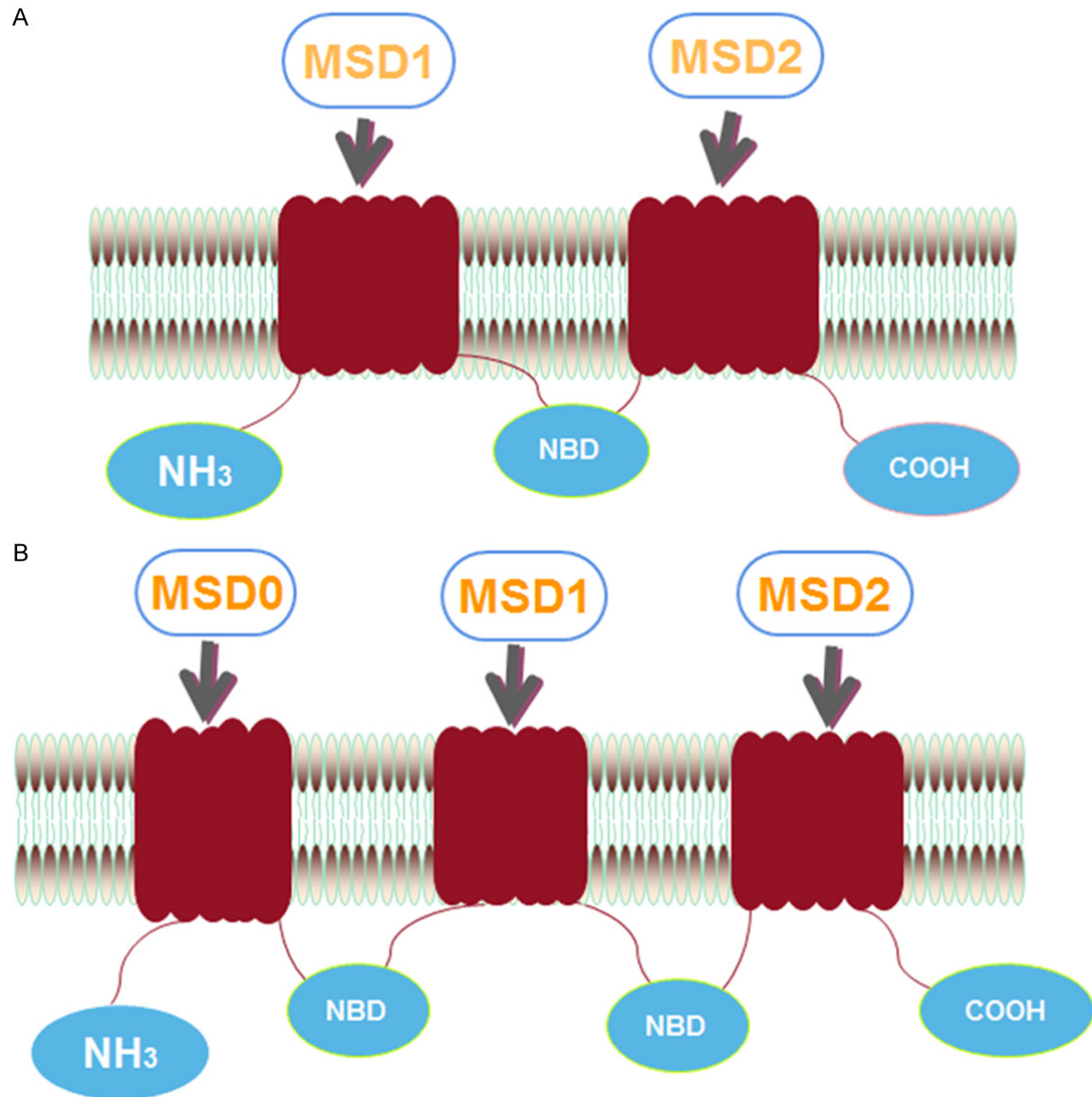


Figure 2. The structure diagram of multidrug resistance protein. A. MRPs are characterized by common structural features: multiple transmembrane (TM) α helices arranged in membrane-spanning domains (MSDs), and intracellular nucleotide-binding domains (NBDs) for ATP binding and hydrolysis. B. "Short-term" MRPs: The basic structure of MRP4, MRP5, MRP8, MRP9. The short MRP is a tetrameric structural domain arrangement consisting of two MSDs and two NBDs. The tetrameric structure of two MSDs and two NBDs is commonly found in the ABC superfamily. "Long-term" MRPs: The basic structure of MRP1, MRP2, MRP3, MRP6, MRP7. The long MRP is an additional NH₂-proximal MSD0 outside the four-structure domain arrangement consisting of two MSDs and two NBDs. Abbreviations: MRP, Multidrug Resistance Protein; COOH, Carboxyl Groups; NH₃, Amino Groups; MSDs, Membrane-Spanning Domains; NBDs, Intracellular Nucleotide-Binding Domains.

cytes into the bloodstream, and its transformation by glucosylation in enterocytes and hepatocytes [41].

MRP4/MOAT-B was found in a T lymphocyte line in 1996. It encodes 1325 amino acids, and the gene is positioned on the long arm of chro-

mosome 13q32.1. Structurally, MRP4 consists of a typical ABC transport core with two TMDs and two NBDs [42]. MRP4 is concentrated in diverse structures, such as the lung, muscle, pancreas, kidney, prostate, bladder, ovary and testis [42, 43]. MRP4 is a prostatic excretion pump that regulates drug trafficking in the pros-

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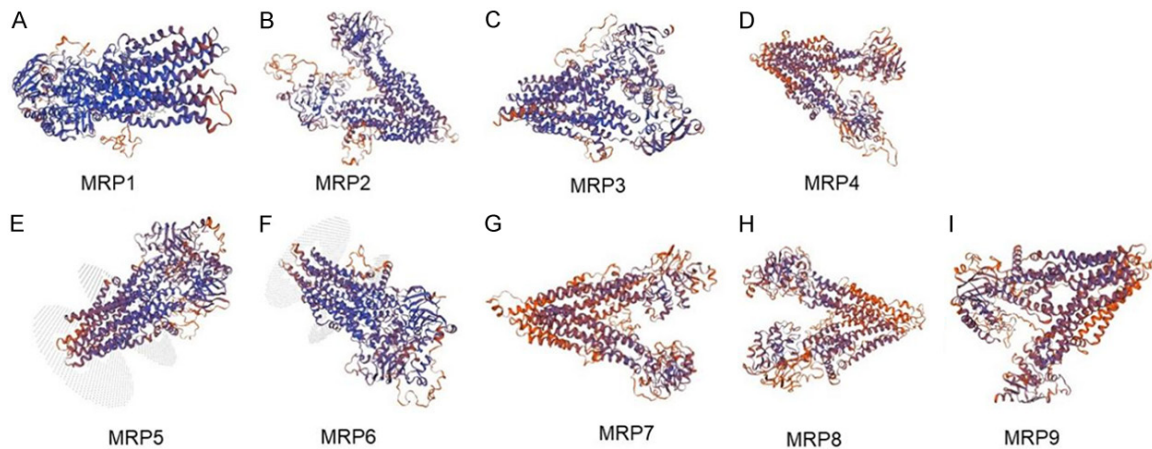


Figure 3. Molecular structures of nine multidrug resistance protein isoforms. It shows the protein fold structure of MRP1-9. A. MRP1; B. MRP2; C. MRP3; D. MRP4; E. MRP5; F. MRP6; G. MRP7; H. MRP8; I. MRP9. Among them, the ligands of MRP1 structure are ADENOSINE-5'-DIPHOSPHATE, ADENOSINE-5'-TRIPHOSPHATE, CHOLESTEROL, MAGNESIUM ION. The ligands of the MRP5 structure are MAGNESIUM ION, and the ligands do not appear in other types of structure diagrams. All the structural diagrams are drawn on the mapping software SWISS-MODEL. The protein sequence of MRP1-9 is input on the software to export the structural diagram.

tate [44]. MRP4 may expel a range of endogenous and heterogeneous organic ionized chemicals from cells, which is different from other C family members. MRP4 regulates the pharmacology and kinetics of several medications as a transporter, including antiviral, antineoplastic, and diuretic drugs [45]. In addition to its action as a drug transporter and renal excretion, MRP4 is also critical in the transmission of cellular signalling molecules, and it is closely linked to active cellular and body activities and physiological processes [46, 47]. MRP4 is also a component in the pathogenesis of metabolic diseases such as obesity and diabetes [48].

The MRP5/MOAT-C gene is located on chromosome 3q27, and the protein contains 1437 amino acids. Some studies found that MRP5 was expressed in a broad range of tissues with highest expression in the heart, brain, lung and skeletal muscle. It is localized in smooth muscle sarcomeres, cardiac myocytes endothelial cells, and cardiomyocytes in the heart. MRP5 is also obviously expressed in the adrenal glands [49].

MRP6, also known as MOAT-E, is situated on chromosome 16, it and encodes a single polypeptide chain of 1503 amino acids with a central MRP structure (with two transmembrane structural domains TMD1 and TMD2 and two cytoplasmic nucleotide-binding structural domains NBD1 and NBD2) [50-52]. MRP6 is pri-

marily located in the liver and kidney, but it is expressed at low or undetectable levels in other tissues, such as the duodenum, colon, brain and salivary glands. MRP6 regulates several key enzymes to produce inorganic pyrophosphate (PPI, an effective inhibitor of ectopic calcification) and prevent its degradation to regulate extracellular purines that participate in ectopic calcification. MRP6 deficiency promoted ectopic calcification and aggravated atherosclerosis [53].

MRP7 was discovered in 2001, and it is located on the 6p12 chromosome. It encodes a protein of 1636 amino acids. It is a 171-kDa protein containing three membrane span domains (MSDs) and two NBDs [54]. MRP7 is most highly expressed in the pancreas, but it is also expressed in the liver, placenta, lungs, kidneys, brain, ovaries, lymph nodes, spleen, heart and colon, and shows low levels in the spleen, stomach, kidneys, heart and brain [55]. MRP7 is a lipophilic ion transporter implicated in the transport of drugs and other endogenous molecules, and it pumps endogenous and heterogeneous biological substrates from the cytoplasm into the extracellular environment [56]. Phosphorylated MRP7 contributes to the dynamic balance of ions and fluids in healthy humans by conducting chloride and bicarbonate ions to the parietal lamellae of epithelial cells within the primary conducting airways and submucosal glands [57, 58].

The role of MRPs in diseases

Table 2. The role of MRPs in cancers

MRPs	Disease	Impact	Mechanism	Ref
MRP1	Non-small cell lung cancer	Aggravate	SLAMF3↑→MRP1↓→Reducing drug resistance	[75]
	Colon cancer	Aggravate	hypoxia↑→HIF-1α↓→MRP1↓→drug resistance↓	[90]
	Renal cell carcinoma	Aggravate	miR-210-3p↓→MRP1↑→drug resistance↑	[84]
MRP1, MRP4	Ovarian cancer	Aggravate	Eicosanes derived from AA↑→SKOV3-R↑→cisplatin-resistance↑activity of cisplatin transfer↑	[108-113]
MRP2	Colorectal cancer	Aggravate	MRP2↑→LTB4↑→circosanooids↑	[95-97]
	Durbin-Johnson syndrome	Aggravate	MRP2↓→CB↑→Inflammatory cytokine release↑→Extracellular regulated kinase↓→MRP2↓	[115-117]
	Hepatocellular carcinoma	Aggravate	CircMRP2↑→miR-665↑→MRP2↑	[118]
	Non-small cell lung cancer	Attenuate	MRP2↑→activation of PARP↑→activation of caspase-3↑→cisplatin-resistance↑	[69]
MRP4	Acute myeloid leukemia	Aggravate	MRP4↓→cAMP↑→Proliferation and differentiation of AML cells↑	[104]
	Pancreatic cancer	Aggravate	MRP4↑→the growth of Pancreatic-3↑→EMT↑	[76]
	Breast cancer	Aggravate	MRP4↑→PGE2↑	[107]
MRP7	Endometrial cancer	Aggravate	NEAT1↑→miR-98↓→MRP7↑→drug resistance↑	[82]

Abbreviations: MRP1, Multidrug Resistance Protein 1; MRP2, Multidrug Resistance Protein 2; MRP4, Multidrug Resistance Protein 4; MRP6, Multidrug Resistance Protein 6; LTB4, Leukotriene B4; CB, Conjugated Bilirubin; PARP, Poly (ADP-Ribose) Polymerase; AA, Arachidonic Acid; CAMP, Cyclic Adenosine Monophosphate; EMT, Epithelial-Mesenchymal Transition; PGE2, Prostaglandin E2; IL-1, IL-6, IL-8, Interleukin; NF-kappa B, Nuclear Factor Kappa B; SLAMF3, A Member Of The Signaling Lymphocyte-Activating Molecule Family Of Receptors; HIF-1α, Hypoxia-Inducible Factor; NEAT1, Inhibitors Of MiR-98.

Similar to other members of the family, MRP8 is a complete transport protein with two nucleotide binding regions and 12 transmembrane regions. It has high homology with MRP5 and its gene locus is on chromosome 16q12.1. It is expressed at moderate levels in normal breast and testis tissues and at very low levels in the liver, brain and placenta [59]. The transmembrane protein structure of MRP8 is similar to MRP4 and MRP5, and it possesses just two transmembrane structural domains. MRP8 is poorly expressed in all human tissues other than lung, foetal tissue, kidney, spleen, colon and brain [60].

The MRP9 sequence is similar to MRP8 and was obtained from a cDNA library of human adult liver via cloning. It is detected in gene records of human tissues and other organs, such as the liver, kidney and lung [61-64]. MRP9 sits next to MRP8 on chromosome 16q12.1. MRP9 is represented by a single ATP-binding domain, but the two MSDs each have four transmembrane domains. MRP9 is highly expressed in testis and breast tissues [65].

Mechanisms of MRPs in diseases

MRPs and cancer

Many recent studies showed that MRPs played important roles in cancers. We summarise the

related studies of MRPs in cancer below and in **Table 2** and **Figure 5**.

Gastric adenocarcinoma: Mechanisms of chemoresistance are important for the failure of traditional Chinese in the treatment of gastric adenocarcinoma (GAC), which promotes GAC multidrug resistance [66]. Abdulla et al. [67] found MRP1, MRP3, MRP4, MRP5 were significantly expressed in GAC, which may be involved in chemoresistance of GAC. Through the development of the GAC-derived cell line AGS animal model, it was discovered that MRP1 and MRP4 inhibition, as well as the MRP inhibitors diclofenac and probenecid, dramatically improved the responsiveness of AGS cells to sorafenib and docetaxel, which enabled anti-cancer medication in GAC.

Non-small cell lung cancer: Qi et al. [68] found that overexpression of MRP1 promoted multidrug resistance in non-small cell lung cancer (NSCLC). Using the anticancer drug etoposide (VP-16), they found that it exacerbated ROS activation of HIF-1α, switched cellular metabolism to glycolysis in NSCLC, and increased lactate in the extracellular environment, which increased the expression of MRP1. They indicated that the development of MRP1 transporter inhibitors will play an active role in the clinical treatment of NSCLC in the future. Cisplatin resistance is a major obstacle to cisplatin treat-

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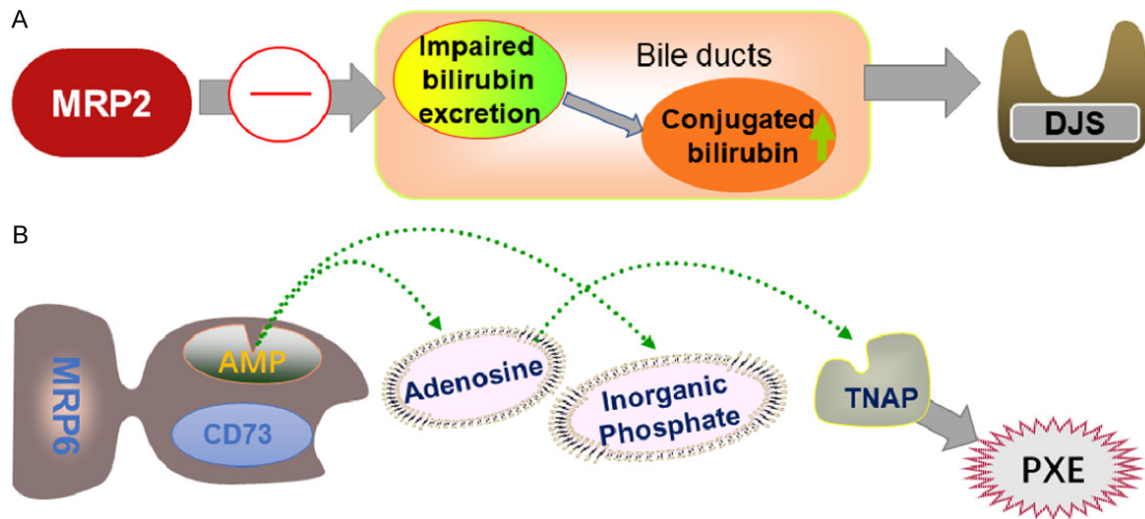


Figure 4. Mechanism of action of multidrug resistance proteins in hereditary diseases. A. Impaired function of MRP2 leads to DJS by impairing bilirubin, which is one of the pathogenic mechanisms of DJS, leading to cholestasis; B. MRP6 secretes AMP and CD73 act together downstream to degrade AMP to adenosine and inorganic phosphate. Adenosine inhibits TNAP expression and activity, thereby regulating PPI. Abbreviations: DJS, Dubin-Johnson Syndrome; PXE, Pseudoxyloma Elasticum.

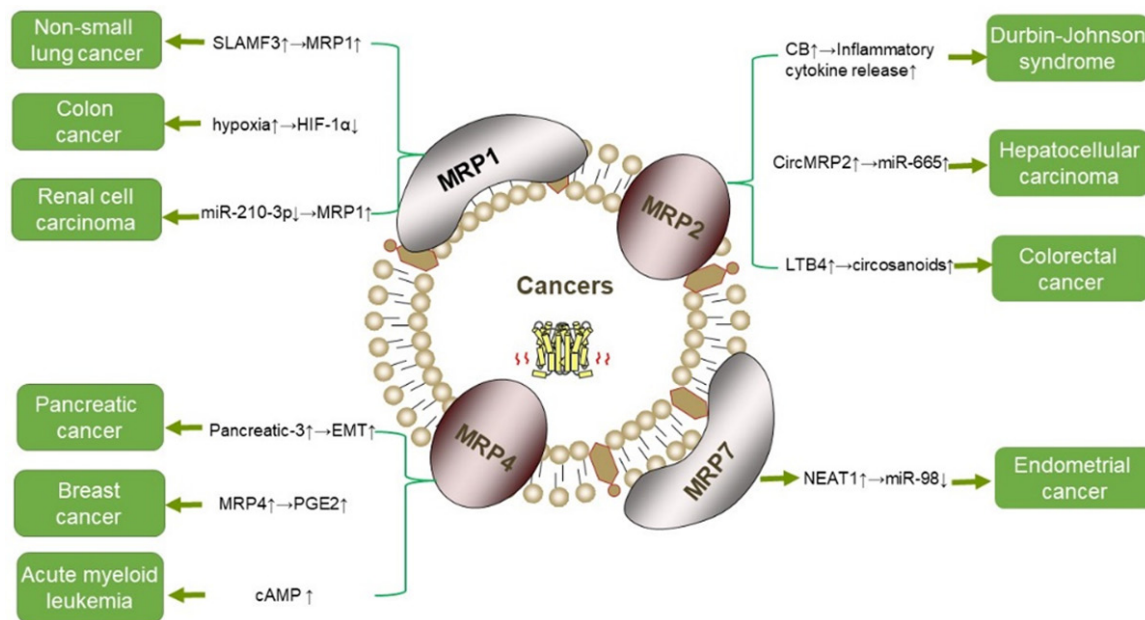


Figure 5. Mechanism of multidrug resistance proteins-mediated cancer. Multidrug resistance is one of the major obstacles leading to clinical treatment of cancer, and multidrug resistance proteins-mediated multidrug resistance plays an important role.

ment of NSCLC. Chen et al. [69] found that reducing MRP2 expression was an important strategy to increase the sensitivity of NSCLC to cisplatin. The cleavage of PARP in cisplatin-resistant NSCLC cells is an important indicator of apoptosis and the activation of caspase-3 apoptotic signalling, and MRP2 knockdown ac-

tivated PARP and promoted apoptosis in these cells.

Hepatocellular carcinoma: Fouquet et al. [70] found that MRP1 was highly expressed in tumour tissues of HCC, and overexpression of the signalling lymphocyte activating molecule

family member 3 (SLAMF3) in HCC cells specifically reduced MRP1, which reduced multidrug resistance in HCC and facilitated the treatment of HCC [71-73]. SLAMF3 participates in the cell cycle in an RB/PLK-1-dependent manner in HCC cells by regulating the ERK1/2, JNK and mTOR pathways [74]. In addition, the expression of MRP2 also increased, mainly through the upregulation of CircMRP2 and miR-665 [118]. The overexpression of SLAMF3 in HCC may be due to the regulation of cell proliferation and apoptosis and the enhancement of treatment efficacy, but further research is needed to determine the exact mechanism by which SLAMF3 and MRP1 reduce tumour growth [75].

Pancreatic cancer: Pancreatic cancer (PC) is a highly fatal cancer with few symptoms in its early stages. It is normally considered terminal when it is diagnosed and has a poor prognosis. High chemotherapy resistance is one of the characteristics of PDAC and MRPs play a regulatory role in the chemoresistance [76].

Guo et al. [77] established a PDAC mouse model and found that the combination of gemcitabine and rhodopsin reduced the expression levels of P-gp, MRP1 and MRP5, which were beneficial in inhibiting tumour growth and resistance to gemcitabine in mice. However, the exact mechanism is not clear. MRP3 is also considered a therapeutic target for pancreatic cancer. Emodin may promote a reduction of multidrug resistance in PDAC by inhibiting the levels of P-gp, MRP1 and MRP5, but the specific mechanisms are not clear. MRP3 is also considered a therapeutic target for pancreatic cancer. Mutant p53 regulates MRP3 via miR-34C while advancing pancreatic cancer via bioactive lipid lysophosphatidylinositol. Patient survival rate is favourably linked with low expression of the tumour suppressor miR-34C with lower expression levels being linked to lower survival rates. P53 is a cancer suppressor gene. A decrease in proliferation of pancreatic cancer cells in vitro and in vivo was achieved by knocking down ABCC3, which inhibited key regulators of PDAC progression, the STAT3 and HIF1 α signalling pathways [78].

Sahores et al. [76] found that the overexpression of MRP4 accelerated the development of BxPC-3 clonal cell lines in mice, which altered the expression of epithelial mesenchymal tran-

sition (EMT) signalling markers and promoted PDAC cell growth. The level of MRP4 expression negatively correlated with the prognosis of PDAC in PDAC patients. Therefore, MRP4 has become a target of gene therapy for PDAC. The expression of MRP4 was higher in PDAC samples, and MRP4 was linked to a more invasive PDAC phenotype. MRP4 is the main protein leading to cAMP efflux, and its inhibition or silencing abolished cell proliferation by activating the cAMP/Epac/Rap1 signalling pathway. The anti-proliferative effect of MRP4 was restored by cAMP, and the cAMP pathway is often activated in pancreatic cancer [79].

Osteosarcoma: He et al. [80] established the human osteosarcoma cell line MG63/DOX that is resistant to adriamycin (ADM) and compared it with the primary cell line GSF-0686. The results showed that the expression of MRP4 in MG63/DOX cells was significantly higher than MG63 cells, which suggests that overexpressed MRP4 may be involved in the ADM resistance of osteosarcoma and promote the drug resistance of osteosarcoma cells. However, the relationship between MRP4 and ADM is not clear.

Endometrial cancer: Chemoresistance is a major factor in the poor prognosis of endometrial cancer (EC) [81]. MRP7 is significantly expressed in EC tissues and associated with poor EC prognosis. Huang et al. [82] found that the tumour suppressor miR-98 inhibited the expression of MRP7, and facilitated resistance to the oncotherapy drug paclitaxel. An inhibitor of miR-98, NEAT1, increased paclitaxel resistance in EC cells by decreasing the expression of miR-98 to upregulate the level of MRP7.

Renal cell carcinoma: Chen et al. [83] showed that miR-210-3p was associated with various cancer pathologies and negatively regulated MRP1. Li et al. [84] showed that the downregulation of miR-210-3p in renal cell carcinoma (RCC) cells increased the expression level of MRP1, which increased multidrug resistance in RCC, but the specific mechanism has not been explained.

Colorectal cancer: Colorectal cancer (CRC) is one of the most common malignancies worldwide [85-87]. CRC is thought to develop from polyps that gradually develop into invasive tumours [88]. High expression of MRP1 posi-

tively correlated with the prognosis of CRC [89]. Lv et al. showed that hypoxia inhibited and reduced HIF-1 α , which induced MRP1, and facilitated a reduction of drug resistance in colon cancer. MRP1 was a downstream target gene of HIF-1 α , which provided a mechanistic basis for the treatment of HIF-1 α -mediated CRC [90].

Inflammation is a risk factor for CRC, and intestinal inflammation and ABC transporter proteins create a link between environmental and potential intestinal carcinogenesis [91-93]. Andersen et al. [94] found that the early events of high expression of the MRP2 gene and low expression of the ABCG2 gene occurred in the CRC-cancer sequence. MRP2 participates in the early process of CRC by transporting LTB4 or MRP2 to transport other eicosanes, and LTB4 and eicosanes promote inflammatory and cancer inflammatory signalling pathways [95-97].

Cholangiocarcinoma: Cholangiocarcinoma (CCA) is a primary liver cancer, and the main obstacle for treatment is the increased resistance of MRP-induced tumours to chemotherapy [98]. Yang et al. [99] found that the expression of MRP5 and MRP6 was significantly upregulated after exposure to the anticancer drug gemcitabine in CCA liver cell lines, which supports the development of MRP6 and MRP5 inhibitors as a treatment for CCA.

Acute myelogenous leukaemia: Acute myelogenous leukaemia (AML) is one of the most common heterogeneous haematological malignancies worldwide [100]. The occurrence and development of AML is due to the disturbance of haematopoietic stem cell differentiation and the excessive proliferation of myeloid progenitor cell clones. Intracellular cAMP is a signalling molecule that regulates cell proliferation, differentiation and apoptosis, and plays an important role in AML [101-103]. Gonzalez et al. [104] induced MRP4 expression and increased cAMP efflux in AML by histamine treatment, and the combination of histamine and MRP inhibitors promoted a reduction in AML cell proliferation in favour of AML treatment. MRP4 regulates the proliferation and differentiation of AML cells by regulating the level of cAMP in AML cell lines.

Breast cancer: Breast cancer (BC), especially metastatic breast cancer, is a leading contribu-

tor to morbidity and mortality in women with related diseases [105, 106]. Kochel et al. [107] demonstrated that overexpression of MRP4 was associated with breast cancer. Overexpression of MRP4 increased prostaglandins (PGE2), which are major eicosanoid products in tumours and are detrimental to the prognosis of breast cancer.

Ovarian cancer: Arachidonic acid (AA) metabolism plays an important role in advanced ovarian cancer [108, 109]. Zhang et al. [110] showed that overexpression of MRP1 and MRP4 in cisplatin (DDP)-resistant SKOV3 (SKOV3-R) cells increased the production of AA-derived eicosanoids, which had transduction effects on the drug sensitivity of ovarian cancer DDP capacity to the detriment of ovarian cancer treatment [111-113].

MRPs and hereditary diseases

Durbin-Johnson syndrome: Durbin-Johnson syndrome (DJS) is a hereditary autochthonous recessive disorder featuring conjugated hyperbilirubinemia, which is an increased concentration of bilirubin glucoside in the blood, impaired secretion of anion conjugates of bile in hepatocytes and dark pigmentation of hepatocytes that causes a dark blue or black colouration of the liver in affected individuals [114, 115]. Mutations in the MRP2 gene lead to impaired expression and targeting of MRP2 protein on the cholinesterase membrane, which lead to Durbin-John syndrome that is characterized by conjugated hyperbilirubinemia and defective excretion of organic ions. It is characterized by mutations in the MRP2 gene and impaired MRP2 function, which lead to impaired bilirubin excretion and elevated conjugated bilirubin that result in further cholestasis. Bilirubin is the mechanism by which DJS occurs, and MRP2 is an important driver of bile flow. Downregulated MRP2 expression leads to an excessive release of inflammatory cytokines, which inhibit extracellular regulated kinases in hepatocytes, and a downregulation of MRP2. This vicious cycle exacerbates liver injury [115-117].

Pseudopolychordoma elasticity: Pseudoxyloma elasticum (PXE) is a hereditary infectious disease of the connective tissue [118, 119]. Connective tissue disorder and elastic fibre abnormality are the early manifestations of PXE [120], which is characterized by progressive dystrophy and mineralisation of elasticfi-

bles, including calcification of elastic structures in the skin. Patients with PXE often have skin lesions, vision loss, intermittent coagulation and myocardial infarction [121-123]. Several studies suggested that PXE was caused by mutations in the MRP6 gene, but the process is complex and involves many uncertainties. MRP6 may be associated with the release of ATP from cells, and MRP6 is involved in ATP metabolism and inhibits ectopic calcification. MRP6 deficiency can promote ectopic calcification. MRP6 secretes AMP (adenosine 5-monophosphate), which is converted from ATP by ENPP1 (nucleotide pyrophosphatase/phosphodiesterase) and CD73 (an exo-5-nucleotide) acts together downstream to degrade AMP into adenosine and inorganic phosphate. Adenosine inhibits the expression and activity of TNAP to regulate PPI (**Figure 4**). PPI is an effective calcification inhibitor and a protective endocrine factor that is missing in PXE. TNAP is a tissue nonspecific alkaline phosphatase that degrades pyrophosphate [124-127].

MRPs and other diseases

Acute liver injury: Liver injury is associated with endoplasmic reticulum (ER) stress and NF- κ B signalling, and MRP2 is regulated by NF- κ B [128, 129]. Huang et al. found significant ER stress, increased MRP2 expression and activation of NF- κ B in a carbon tetrachloride (CCl₄)-induced acute liver injury model and a Tg-induced ER stress model in mice. The down-regulation of MRP2 expression in the liver increased ER stress and promoted hepatocyte apoptosis and liver injury in mice with CCl₄-induced acute liver injury. The mechanism of increased MRP2 expression in the liver is the activation of NF- κ B signalling after ER stress, which alleviates ER stress and acute liver injury [51, 128].

Atopic dermatitis: Atopic dermatitis (AD) is an immune skin disease mediated by T cells [130, 131]. Lee et al. established Jurkat cells and mouse CD4 T-cell models with AD, and found that MRP1 physically bound to kamanol in activated T cells, in which reduced the expression and phosphorylation of JNK in the treatment of atopic dermatitis [132].

Prostate inflammation: Prostate inflammation (PI) is often accompanied by symptoms of the lower urinary tract, and the urethra plays an

important role in the urinary cycle [133]. Alexandre et al. [134] used histological and cGMP level analyses and found that PI dysfunction decreased urethral contractility and the expression of MRP5, which may modulate inflammatory processes, increased intracellular cGMP (guanosine cyclophosphate) accumulation in urethral tissues, and significantly relaxed smooth muscle, which may adversely affect urethral tissues.

Kawasaki disease: Kawasaki disease (KD) was first reported in 1967 [135]. It is a systemic vasculitis syndrome that primarily affects coronary vasculitis. It is an acute self-limited vasculitis and acute febrile disease of unknown cause [136, 137]. It typically occurs in children under 5 years of age and as a feature of hyperhaemoglobinaemia [138]. KD may cause coronary artery aneurysms, ischaemic heart disease, myocardial infarction, and death [139-141]. Intravenous immunoglobulin (IVIG) is the first choice for the treatment of KD, but McCrindle found IVIG resistance such as recurrence or persistent fever after treatment [142]. MRP4 is a mediator of prostaglandin E1 and prostaglandin E2 efflux, and an MRP4 single nucleotide polymorphism may influence Kawasaki disease. Because the triggering factor of PGE2 expression in the acute phase of Kawasaki disease is IVIG, there is a certain correlation between the change in plasma PGE2 levels and IVIG resistance. However, there is uncertainty about the role of MRP4 single nucleotide polymorphisms in IVIG resistance and other roles for MRP4 in the pathogenesis of KD.

Ulcerative colitis: Ulcerative colitis (UC) is a type of chronic colitis that belongs to inflammatory bowel diseases, with clinical manifestations comprised of rectal bleeding, persistent diarrhoea and belly pain [143]. Several studies showed that the MRP7/CFTR gene and protein were involved in the occurrence and development of UC. Marco et al. [120] used MRP7/CFTR knockout mice and discovered that CFTR downregulation was responsible for increased expression of inflammatory interleukin 6 (IL-6), tumour necrosis factor- α (TNF- α) and NF- κ B in the mouse intestine, which resulted in an inflammatory response. NF- κ B is a key inflammatory regulator that is activated by a variety of stimuli, including bacterial components (such as lipopolysaccharide), proinflammatory cytokines (such as TNF- α and IL-1), viruses and DNA

proliferators. The activation of transcriptional NF- κ B promotes an increase in ILs, such as IL-1, IL-6 and IL-8, and participates in the inflammatory response [144].

Progress of MRP inhibitors

Meloxicam

COX inhibitors reduce multidrug resistance by inhibiting the expression of ATP-binding cassette transporters, which is beneficial to the anticancer drug doxorubicin for the treatment of tumours [145]. Chen et al. [146] revealed the role of a specific COX-2 inhibitor, meloxicam, by developing a human A549 lung cancer model. A549 cells had increased accumulation of a substrate of intracellular MRP, doxorubicin, which helped inhibit the development of lung cancer by reducing the expression levels of MRP1 and MRP4.

Lapatinib

Lapatinib inhibits the phosphorylation of EGFR, AKT and P38 in breast cancer globulin, reduces the expression of MRP1, and promotes the toxicity of tumour-globulin cells [147]. Kwon et al. [148] found that blocking the PI3K/AKT and p38/MAPK signalling pathways reduced MRP1 levels in doxorubicin-treated tumour spheroids. These results indicated that lapatinib suppresses adriamycin-induced MRP1 expression by inhibiting the PI3K/AKT and p38MAPK signalling pathways, which may be an effective agent for the treatment of breast cancer.

HOXA10

Multidrug resistance is a major obstacle in the treatment of chronic myeloid leukaemia (CML) with the anticancer drug doxorubicin [149]. HOXA10 is a member of the HOX gene family and it is frequently expressed in CML [150]. Ying et al. [151] found that knockdown of HOXA10 inhibited the expression of P-gp and MRP1, which indicated that HOXA10 was a potential target for reducing multidrug resistance in CML.

Conclusion and future

Multidrug resistance is increasingly recognised as a major obstacle to cancer treatment. Multidrug resistance proteins are involved in

the occurrence and development of various cancers. As the C subfamily of the ABC family, MRPs play an important role in the multidrug resistance of various cancers and are closely related to the occurrence and development of other systemic diseases. EBP50 knockdown in HepG2 cells, the respective substrates of MRP and breast cancer resistance protein (BCRP), reduced the activities of P-gp and MRP, but not BCRP. These results suggest that EBP50 regulates ET activity. MRP-1 is implicated in multidrug resistance and was described as prognostic in high-risk patients with soft-tissue sarcoma. The expression of MRP-1 is elevated in colorectal adenocarcinoma tissues, and patients with negative expression of MRP-1 have a better prognosis. Therefore, MRP-1 may be a reference indicator for clinical diagnosis and prognosis. Other multidrug resistance proteins also play irreplaceable roles in various cancers.

The development of MRP inhibitors is expected to improve the efficacy of anticancer drugs and be beneficial to the treatment of other diseases. However, to overcome MDR in cancer, a gap between in vitro/in vivo research of MRPs modulators and therapeutic applications remains, and the discovery of specific MRP inhibitors is currently lacking. Therefore, to overcome chemoresistance in clinical practice, very powerful and selective MRP inhibitors and sizable MDR cancer cohorts with thorough clinicopathological data are needed. The combined use of MRP inhibitors and anticancer drugs is expected to become the development trend in cancer treatment.

Although ABC transporter proteins play an important role in the development of multidrug-resistant cancers, efforts to target these transporter proteins have not been successful. Future strategies to combat multidrug resistance are likely going to include the following: (1) the use of bioinformatics for customized treatment; (2) continuous research on new pathways relating ABC transporter proteins to diseases, including MDR; and (3) accelerating anticancer drug development platforms through the use of high-throughput and the packaging of new or current chemotherapeutic agents as nano-drugs to boost medication delivery to drug-resistant cancer cells. By using novel medicines and cutting-edge technologies as tactics for

attacking ABC-transporting proteins and multi-drug resistance, such as nanoparticles, monoclonal antibodies, and gene technologies, anti-cancer drug delivery to resistant cells may be increased. Combination therapies that maximize chemosensitivity should also be further investigated. These methods may hold the solution to the growing issue of malignancies with medication resistance.

We should also investigate the cause of multi-drug resistance from the MRP family to understand the mechanism of cancer, genetics and other diseases from various aspects and whether the MRP family is related to other causes that affect the disease. Given these unanswered questions, we have much work to do in the future.

Acknowledgements

This work was supported by the National Natural Science Foundation of China (81873076, 81973725) and the Hundred Talents Program from Shanghai University of Traditional Chinese Medicine.

Disclosure of conflict of interest

None.

Abbreviations

AA, Arachidonic Acid; ABC, ATP-Binding Cassette; AD, Atopic Dermatitis; ADM, Adriamycin; AML, Acute Myelogenous Leukemia; BC, Breast Cancer; CCA, Cholangiocarcinoma; cMRP, Cholinergic MRP; CML, Chronic Myeloid Leukemia; CRC, Colorectal Cancer; DJS, Durbin-Johnson Syndrome; EC, Endometrial Cancer; EMT, Epithelial Mesenchymal Transformation; ER, Endoplasmic Reticulum; GAC, Gastric Adenocarcinoma; HCC, Hepatocellular Carcinoma; IL-6, Inflammatory Interleukin 6; IVIG, Intravenous Immunoglobulin; KD, Kawasaki Disease; MDR, Multidrug Resistance; MRPs/ABCCs, Multidrug Resistance Proteins; MSD, Membrane Span Domains; NF- κ B, Nuclear Factor KappaB; NSCLC, Non-Small Cell Lung Cancer; PC, Pancreatic Cancer; P-gp, P-Glycoprotein; PGE₂, Prostaglandin; PI, Prostate Inflammation; PXE, Pseudoxyloma Elasticum; RCC, Renal Cell Carcinoma; SLAMF3, Signaling Lymphocyte Activation Molecule Family Member 3; TNF- α , Tumor Necrosis Factor- α ; UC, Ulcerative Colitis.

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References

- [1] Lepeltier E, Rijo P, Rizzolio F, Popovtzer R, Petrikaite V, Assaraf YG and Passirani C. Nanomedicine to target multidrug resistant tumors. *Drug Resist Updat* 2020; 52: 100704.
- [2] Li Y and Xu X. Nanomedicine solutions to intricate physiological-pathological barriers and molecular mechanisms of tumor multidrug resistance. *J Control Release* 2020; 323: 483-501.
- [3] Wang JQ, Yang Y, Cai CY, Teng QX, Cui Q, Lin J, Assaraf YG and Chen ZS. Multidrug resistance proteins (MRPs): structure, function and the overcoming of cancer multidrug resistance. *Drug Resist Updat* 2021; 54: 100743.
- [4] Wei X, Song M, Li W, Huang J, Yang G and Wang Y. Multifunctional nanoplateforms co-delivering combinatorial dual-drug for eliminating cancer multidrug resistance. *Theranostics* 2021; 11: 6334-6354.
- [5] Zhou XW, Xia YZ, Zhang YL, Luo JG, Han C, Zhang H, Zhang C, Yang L and Kong LY. Tomentodione M sensitizes multidrug resistant cancer cells by decreasing P-glycoprotein via inhibition of p38 MAPK signaling. *Oncotarget* 2017; 8: 101965-101983.
- [6] Assaraf YG, Brozovic A, Gonçalves AC, Jurkovicova D, Linē A, Machuqueiro M, Saponara S, Sarmiento-Ribeiro AB, Xavier CPR and Vasconcelos MH. The multi-factorial nature of clinical multidrug resistance in cancer. *Drug Resist Updat* 2019; 46: 100645.
- [7] Poma P, Labbozzetta M, McCubrey JA, Ramarosandratana AV, Sajeve M, Zito P and Notarbartolo M. Antitumor mechanism of the essential oils from two succulent plants in multidrug resistance leukemia cell. *Pharmaceuticals (Basel)* 2019; 12: 124.
- [8] Huang WJ, Ruan S, Wen F, Lu XN, Gu SP, Chen XX, Liu M and Shu P. Multidrug resistance of gastric cancer: the mechanisms and chinese medicine reversal agents. *Cancer Manag Res* 2020; 12: 12385-12394.

- [9] Bukowski K, Kciuk M and Kontek R. Mechanisms of multidrug resistance in cancer chemotherapy. *Int J Mol Sci* 2020; 21: 3233.
- [10] Vaidya FU, Sufiyan Chhipa A, Mishra V, Gupta VK, Rawat SG, Kumar A and Pathak C. Molecular and cellular paradigms of multidrug resistance in cancer. *Cancer Rep (Hoboken)* 2020; e1291.
- [11] Rahmani F, Ziaemehr A, Shahidsales S, Gharib M, Khazaei M, Ferns GA, Ryzhikov M, Avan A and Hassanian SM. Role of regulatory miRNAs of the PI3K/AKT/mTOR signaling in the pathogenesis of hepatocellular carcinoma. *J Cell Physiol* 2020; 235: 4146-4152.
- [12] Liu R, Chen Y, Liu G, Li C, Song Y, Cao Z, Li W, Hu J, Lu C and Liu Y. PI3K/AKT pathway as a key link modulates the multidrug resistance of cancers. *Cell Death Dis* 2020; 11: 797.
- [13] Dallavalle S, Dobričić V, Lazzarato L, Gazzano E, Machuqueiro M, Pajeva I, Tsakovska I, Zidar N and Fruttero R. Improvement of conventional anti-cancer drugs as new tools against multidrug resistant tumors. *Drug Resist Updat* 2020; 50: 100682.
- [14] Bar-Zeev M, Livney YD and Assaraf YG. Targeted nanomedicine for cancer therapeutics: towards precision medicine overcoming drug resistance. *Drug Resist Updat* 2017; 31: 15-30.
- [15] Zhang YK, Zhang GN, Wang YJ, Patel BA, Talele TT, Yang DH and Chen ZS. Bafetinib (INNO-406) reverses multidrug resistance by inhibiting the efflux function of ABCB1 and ABCG2 transporters. *Sci Rep* 2016; 6: 25694.
- [16] Dei S, Coronello M, Bartolucci G, Manetti D, Romanelli MN, Udomtanakunchai C, Salerno M and Teodori E. Design and synthesis of new potent N,N-bis(arylalkyl)piperazine derivatives as multidrug resistance (MDR) reversing agents. *Eur J Med Chem* 2018; 147: 7-20.
- [17] Chang L, Hou Y, Zhu L, Wang Z, Chen G, Shu C and Liu Y. Veliparib overcomes multidrug resistance in liver cancer cells. *Biochem Biophys Res Commun* 2020; 521: 596-602.
- [18] Beretta GL, Cassinelli G, Pennati M, Zuco V and Gatti L. Overcoming ABC transporter-mediated multidrug resistance: the dual role of tyrosine kinase inhibitors as multitargeting agents. *Eur J Med Chem* 2017; 142: 271-289.
- [19] Mansoori B, Mohammadi A, Davudian S, Shirjang S and Baradaran B. The different mechanisms of cancer drug resistance: a brief review. *Adv Pharm Bull* 2017; 7: 339-348.
- [20] Ye Z, Lu Y and Wu T. The impact of ATP-binding cassette transporters on metabolic diseases. *Nutr Metab (Lond)* 2020; 17: 61.
- [21] Muriithi W, Macharia LW, Heming CP, Echevarria JL, Nyachio A, Filho PN and Neto VM. ABC transporters and the hallmarks of cancer: roles in cancer aggressiveness beyond multidrug resistance. *Cancer Biol Med* 2020; 17: 253-269.
- [22] Mohammad IS, He W and Yin L. Understanding of human ATP binding cassette superfamily and novel multidrug resistance modulators to overcome MDR. *Biomed Pharmacother* 2018; 100: 335-348.
- [23] Briz O, Perez-Silva L, Al-Abdulla R, Abete L, Reviejo M, Romero MR and Marin JGG. What "the cancer genome atlas" database tells us about the role of ATP-binding cassette (ABC) proteins in chemoresistance to anticancer drugs. *Expert Opin Drug Metab Toxicol* 2019; 15: 577-593.
- [24] Xiao H, Zheng Y, Ma L, Tian L and Sun Q. Clinically-relevant ABC transporter for anti-cancer drug resistance. *Front Pharmacol* 2021; 12: 648407.
- [25] Zhang YK, Wang YJ, Gupta P and Chen ZS. Multidrug resistance proteins (mrps) and cancer therapy. *AAPS J* 2015; 17: 802-812.
- [26] Campia I, Buondonno I, Castella B, Rolando B, Kopecka J, Gazzano E, Ghigo D and Riganti C. An autocrine cytokine/JAK/STAT-signaling induces kynurenine synthesis in multidrug resistant human cancer cells. *PLoS One* 2015; 10: e0126159.
- [27] Slot AJ, Molinski SV and Cole SP. Mammalian multidrug-resistance proteins (MRPs). *Essays Biochem* 2011; 50: 179-207.
- [28] Nethathe B, Abera A and Naidoo V. Expression and phylogeny of multidrug resistance protein 2 and 4 in African white backed vulture (*gyps africanus*). *PeerJ* 2020; 8: e10422.
- [29] Conseil G and Cole SPC. The first cytoplasmic loop in the core structure of the ABCC1 (multidrug resistance protein 1; MRP1) transporter contains multiple amino acids essential for its expression. *Int J Mol Sci* 2021; 22: 9710.
- [30] Gao M, Liu T, Li J, Guan Q, Wang H, Yan S, Li Z, Zuo D, Zhang W and Wu Y. YAN, a novel microtubule inhibitor, inhibits P-gp and MRP1 function and induces mitotic slippage followed by apoptosis in multidrug-resistant A549/Taxol cells. *Toxicol In Vitro* 2020; 69: 104971.
- [31] Gökirmak T, Campanale JP, Reitzel AM, Shipp LE, Moy GW and Hamdoun A. Functional diversification of sea urchin ABCC1 (MRP1) by alternative splicing. *Am J Physiol Cell Physiol* 2016; 310: C911-920.
- [32] Chen ZS and Tiwari AK. Multidrug resistance proteins (MRPs/ABCCs) in cancer chemotherapy and genetic diseases. *FEBS J* 2011; 278: 3226-3245.
- [33] Johnson ZL and Chen J. ATP binding enables substrate release from multidrug resistance protein 1. *Cell* 2018; 172: 81-89, e10.
- [34] Szöllősi D, Rose-Sperling D, Hellmich UA and Stockner T. Comparison of mechanistic trans-

- port cycle models of ABC exporters. *Biochim Biophys Acta Biomembr* 2018; 1860: 818-832.
- [35] Ben Saad A, Bruneau A, Mareux E, Lapalus M, Delaunay JL, Gonzales E, Jacquemin E, Aït-Slimane T and Falguières T. Molecular regulation of canalicular abc transporters. *Int J Mol Sci* 2021; 22: 2113.
- [36] Tátrai P and Krajcsi P. Prediction of drug-induced hyperbilirubinemia by in vitro testing. *Pharmaceutics* 2020; 12: 755.
- [37] Wu L, Zhang W, Jia S, Zhao X, Zhou D, Xu A, Duan W, Wu Z, Li H, Zheng S, Nan Y, Jia J, Huang J and Ou X. Mutation analysis of the ABCC2 gene in Chinese patients with Dubin-Johnson syndrome. *Exp Ther Med* 2018; 16: 4201-4206.
- [38] De Vocht T, Buyck C, Deferm N, Qi B, Van Brantegem P, van Vlijmen H, Snoeys J, Hoebe E, Vermeulen A and Annaert P. Identification of novel inhibitors of rat Mrp3. *Eur J Pharm Sci* 2021; 162: 105813.
- [39] van der Kolk DM, de Vries EG, Noordhoek L, van den Berg E, van der Pol MA, Müller M and Vellenga E. Activity and expression of the multidrug resistance proteins P-glycoprotein, MRP1, MRP2, MRP3 and MRP5 in de novo and relapsed acute myeloid leukemia. *Leukemia* 2001; 15: 1544-1553.
- [40] Singh H, Lata S, Choudhari R and Dhole TN. Prevalence of ABCC3-1767G/A polymorphism among patients with antiretroviral-associated hepatotoxicity. *Mol Genet Genomic Med* 2020; 8: e1124.
- [41] Jia YM, Zhu T, Zhou H, Ji JZ, Tai T and Xie HG. Multidrug resistance-associated protein 3 is responsible for the efflux transport of curcumin glucuronide from hepatocytes to the blood. *Drug Metab Dispos* 2020; 48: 966-971.
- [42] Chen Y, Yuan X, Xiao Z, Jin H, Zhang L and Liu Z. Discovery of novel multidrug resistance protein 4 (MRP4) inhibitors as active agents reducing resistance to anticancer drug 6-Mercaptopurine (6-MP) by structure and ligand-based virtual screening. *PLoS One* 2018; 13: e0205175.
- [43] Yuan B, Yoshino Y, Fukushima H, Markova S, Takagi N, Toyoda H and Kroetz DL. Multidrug resistance-associated protein 4 is a determinant of arsenite resistance. *Oncol Rep* 2016; 35: 147-154.
- [44] Huang H, Li J, Shen J, Lin L, Wu X, Xiang S, Li Y, Xu Y, Zhao Q, Zhao Y, Kaboli PJ, Li M, Li X, Wang W, Wen Q and Xiao Z. Increased ABCC4 expression induced by ERR α leads to docetaxel resistance via efflux of docetaxel in prostate cancer. *Front Oncol* 2020; 10: 1474.
- [45] Angelis I, Moussis V, Tsoukatos DC and Tsikaris V. Multidrug resistance protein 4 (MRP4/ABCC4): a suspected efflux transporter for human's platelet activation. *Protein Pept Lett* 2021; 28: 983-995.
- [46] Wen J, Luo J, Huang W, Tang J, Zhou H and Zhang W. The pharmacological and physiological role of multidrug-resistant protein 4. *J Pharmacol Exp Ther* 2015; 354: 358-375.
- [47] Lu X, Long Y, Li X, Zhang L, Li Q, Wen H, Zhong S and Cui Z. Generation of knockout and transgenic zebrafish to characterize Abcc4 functions in detoxification and efflux of lead. *Int J Mol Sci* 2021; 22: 2054.
- [48] Donepudi AC, Lee Y, Lee JY, Schuetz JD and Manautou JE. Multidrug resistance-associated protein 4 (Mrp4) is a novel genetic factor in the pathogenesis of obesity and diabetes. *FASEB J* 2021; 35: e21304.
- [49] Gameiro M, Silva R, Rocha-Pereira C, Carmo H, Carvalho F, Bastos ML and Remião F. Cellular models and in vitro assays for the screening of modulators of P-gp, MRP1 and BCRP. *Molecules* 2017; 22: 600.
- [50] Favre G, Laurain A, Aranyi T, Szeri F, Fulop K, Le Saux O, Duranton C, Kauffenstein G, Martin L and Lefthérotis G. The ABCC6 transporter: a new player in biomineralization. *Int J Mol Sci* 2017; 18: 1941.
- [51] Moitra K, Garcia S, Jaldin M, Etoundi C, Cooper D, Roland A, Dixon P, Reyes S, Turan S, Terry S and Dean M. ABCC6 and pseudoxanthoma elasticum: the face of a rare disease from genetics to advocacy. *Int J Mol Sci* 2017; 18: 1488.
- [52] Ostuni A, Castiglione Morelli MA, Cuvillo F, Bavoso A and Bisaccia F. Structural characterization of the LO cytoplasmic loop of human multidrug resistance protein 6 (MRP6). *Biochim Biophys Acta Biomembr* 2019; 1861: 380-386.
- [53] Brampton C, Pomozi V, Chen LH, Apana A, McCurdy S, Zoll J, Boisvert WA, Lambert G, Henrion D, Blanchard S, Kuo S, Lefthérotis G, Martin L and Le Saux O. ABCC6 deficiency promotes dyslipidemia and atherosclerosis. *Sci Rep* 2021; 11: 3881.
- [54] Kathawala RJ, Wang YJ, Shukla S, Zhang YK, Alqahtani S, Kaddoumi A, Ambudkar SV, Ashby CR Jr and Chen ZS. ATP-binding cassette subfamily B member 1 (ABCB1) and subfamily C member 10 (ABCC10) are not primary resistance factors for cabazitaxel. *Chin J Cancer* 2015; 34: 115-120.
- [55] Wang JQ, Wu ZX, Yang Y, Li JS, Yang DH, Fan YF and Chen ZS. Establishment and characterization of a novel multidrug resistant human ovarian cancer cell line with heterogenous MRP7 overexpression. *Front Oncol* 2021; 11: 731260.
- [56] Dai F, Yoo WG, Lee JY, Lu Y, Pak JH, Sohn WM and Hong SJ. Molecular and structural characteristics of multidrug resistance-associated

- protein 7 in Chinese liver fluke *clonorchis sinensis*. *Parasitol Res* 2017; 116: 953-962.
- [57] Nguyen JP, Kim Y, Cao Q and Hirota JA. Interactions between ABCC4/MRP4 and ABCC7/CFTR in human airway epithelial cells in lung health and disease. *Int J Biochem Cell Biol* 2021; 133: 105936.
- [58] Naramoto H, Uematsu T, Uchihashi T, Doto R, Matsuura T, Usui Y, Uematsu S, Li X, Takahashi M, Yamaoka M and Furusawa K. Multidrug resistance-associated protein 7 expression is involved in cross-resistance to docetaxel in salivary gland adenocarcinoma cell lines. *Int J Oncol* 2007; 30: 393-401.
- [59] Toyoda Y, Gomi T, Nakagawa H, Nagakura M and Ishikawa T. Diagnosis of human axillary osmidrosis by genotyping of the human ABCC11 gene: clinical practice and basic scientific evidence. *Biomed Res Int* 2016; 2016: 7670483.
- [60] Tun-Yhong W, Chinpaisal C, Pamonsinlapatham P and Kaewkitichai S. Tenofovir disoproxil fumarate is a new substrate of ATP-binding cassette subfamily C member 11. *Antimicrob Agents Chemother* 2017; 61: e01725-16.
- [61] Yu H, Pang Z, Li G and Gu T. Bioinformatics analysis of differentially expressed miRNAs in non-small cell lung cancer. *J Clin Lab Anal* 2021; 35: e23588.
- [62] Wang H, Chi CH, Zhang Y, Shi B, Jia R and Wang BJ. Effects of histone deacetylase inhibitors on ATP-binding cassette transporters in lung cancer A549 and colorectal cancer HCT116 cells. *Oncol Lett* 2019; 18: 63-71.
- [63] Hamamoto J, Yasuda H, Aizawa K, Nishino M, Nukaga S, Hirano T, Kawada I, Naoki K, Bet-suyaku T and Soejima K. Non-small cell lung cancer PC-9 cells exhibit increased sensitivity to gemcitabine and vinorelbine upon acquiring resistance to EGFR-tyrosine kinase inhibitors. *Oncol Lett* 2017; 14: 3559-3565.
- [64] Zhang L, Huang P, Huang C, Jiang L, Lu Z and Wang P. Varied clinical significance of ATP-binding cassette C sub-family members for lung adenocarcinoma. *Medicine (Baltimore)* 2021; 100: e25246.
- [65] Roy U, Barber P, Tse-Dinh YC, Batrakova EV, Mondal D and Nair M. Role of MRP transporters in regulating antimicrobial drug inefficacy and oxidative stress-induced pathogenesis during HIV-1 and TB infections. *Front Microbiol* 2015; 6: 948.
- [66] Zhao X, Wu P, Liu D, Li C, Xue L, Liu Z, Zhu M, Yang J, Chen Z, Li Y and She Y. An immunity-associated lncRNA signature for predicting prognosis in gastric adenocarcinoma. *J Healthc Eng* 2022; 2022: 3035073.
- [67] Al-Abdulla R, Perez-Silva L, Lozano E, Macias RIR, Herraes E, Abad M, Segues N, Bujanda L, Briz O and Marin JJG. Sensitizing gastric adenocarcinoma to chemotherapy by pharmacological manipulation of drug transporters. *Biochem Pharmacol* 2020; 171: 113682.
- [68] Dong Q, Zhou C, Ren H, Zhang Z, Cheng F, Xiong Z, Chen C, Yang J, Gao J, Zhang Y, Xu L, Fang J, Cao Y, Wei H and Wu Z. Lactate-induced MRP1 expression contributes to metabolism-based etoposide resistance in non-small cell lung cancer cells. *Cell Commun Signal* 2020; 18: 167.
- [69] Chen Y, Zhou H, Yang S and Su D. Increased ABCC2 expression predicts cisplatin resistance in non-small cell lung cancer. *Cell Biochem Funct* 2021; 39: 277-286.
- [70] Fouquet G, Marie C, Collet L, Vilpoux C, Ouled-Haddou H, Nguyen-Khac E, Bayry J, Naassila M, Marcq I and Bouhlal H. Rescuing SLAMF3 expression restores sorafenib response in hepatocellular carcinoma cells through the induction of mesenchymal-to-epithelial transition. *Cancers (Basel)* 2022; 14: 910.
- [71] Huang W, Huang F and Feng C. CircFoxo3 promotes adriamycin resistance through regulation of miR-199a-5p/ATP binding cassette subfamily C member 1 Axis in hepatocellular carcinoma. *Onco Targets Ther* 2020; 13: 5113-5122.
- [72] Satyananda V, Oshi M, Tokumaru Y, Maiti A, Hait N, Matsuyama R, Endo I and Takabe K. Sphingosine 1-phosphate (S1P) produced by sphingosine kinase 1 (SphK1) and exported via ABCC1 is related to hepatocellular carcinoma (HCC) progression. *Am J Cancer Res* 2021; 11: 4394-4407.
- [73] Cao Z, Liang N, Yang H and Li S. Visfatin mediates doxorubicin resistance in human non-small-cell lung cancer via Akt-mediated up-regulation of ABCC1. *Cell Prolif* 2017; 50: e12366.
- [74] Li D, Xiong W, Wang Y, Feng J, He Y, Du J, Wang J, Yang M, Zeng H, Yang YG, Wu N, Chen S and Dong Z. SLAMF3 and SLAMF4 are immune checkpoints that constrain macrophage phagocytosis of hematopoietic tumors. *Sci Immunol* 2022; 7: eabj5501.
- [75] Fouquet G, Debuysscher V, Ouled-Haddou H, Eugenio MS, Demey B, Singh AR, Ossart C, Al Bagami M, Regimbeau JM, Nguyen-Khac E, Naassila M, Marcq I and Bouhlal H. Hepatocyte SLAMF3 reduced specifically the multi-drugs resistance protein MRP-1 and increases HCC cells sensitization to anti-cancer drugs. *Oncotarget* 2016; 7: 32493-32503.
- [76] Sahoies A, Carozzo A, May M, Gomez N, Di Siervi N, De Sousa Serro M, Yaneff A, Rodriguez-Gonzalez A, Abba M, Shayo C and Davio C. Multidrug transporter MRP4/ABCC4 as a key determinant of pancreatic cancer aggressiveness. *Sci Rep* 2020; 10: 14217.

- [77] Guo H, Liu F, Yang S and Xue T. Emodin alleviates gemcitabine resistance in pancreatic cancer by inhibiting MDR1/P-glycoprotein and MRPs expression. *Oncol Lett* 2020; 20: 167.
- [78] Adamska A, Ferro R, Lattanzio R, Capone E, Domenichini A, Damiani V, Chiorino G, Akkaya BG, Linton KJ, De Laurenzi V, Sala G and Falasca M. ABCC3 is a novel target for the treatment of pancreatic cancer. *Adv Biol Regul* 2019; 73: 100634.
- [79] Carozzo A, Yaneff A, Gómez N, Di Siervi N, Sahores A, Diez F, Attorresi AI, Rodríguez-González Á, Monczor F, Fernández N, Abba M, Shayo C and Davio C. Identification of MRP4/ABCC4 as a target for reducing the proliferation of pancreatic ductal adenocarcinoma cells by modulating the cAMP efflux. *Mol Pharmacol* 2019; 96: 13-25.
- [80] He Z, Hu B, Tang L, Zheng S, Sun Y, Sheng Z, Yao Y and Lin F. The overexpression of MRP4 is related to multidrug resistance in osteosarcoma cells. *J Cancer Res Ther* 2015; 11: 18-23.
- [81] Salinas-Vera YM, Gallardo-Rincón D, Ruíz-García E, Silva-Cázares MB, de la Peña-Cruz CS and López-Camarillo C. The role of hypoxia in endometrial cancer. *Curr Pharm Biotechnol* 2022; 23: 221-234.
- [82] Huang W, Zhang J, Dong B, Chen H, Shao L and Li X. A novel miR-98 negatively regulates the resistance of endometrial cancer cells to paclitaxel by suppressing ABCC10/MRP-7. *Front Oncol* 2021; 11: 809410.
- [83] Chen M, Li D, Gong N, Wu H, Su C, Xie C, Xiang H, Lin C and Li X. MiR-133b down-regulates ABCC1 and enhances the sensitivity of CRC to anti-tumor drugs. *Oncotarget* 2017; 8: 52983-52994.
- [84] Li S, Yang J, Wang J, Gao W, Ding Y, Ding Y and Jia Z. Down-regulation of miR-210-3p encourages chemotherapy resistance of renal cell carcinoma via modulating ABCC1. *Cell Biosci* 2018; 8: 9.
- [85] Hong Z, Tang P, Liu B, Ran C, Yuan C, Zhang Y, Lu Y, Duan X, Yang Y and Wu H. Ferroptosis-related genes for overall survival prediction in patients with colorectal cancer can be inhibited by gallic acid. *Int J Biol Sci* 2021; 17: 942-956.
- [86] Ji L, Chen S, Gu L and Zhang X. Exploration of potential roles of M6A regulators in colorectal cancer prognosis. *Front Oncol* 2020; 10: 768.
- [87] Tian T, Bi H, Liu Y, Li G, Zhang Y, Cao L, Hu F, Zhao Y and Yuan H. Copy number variation of ubiquitin-specific proteases genes in blood leukocytes and colorectal cancer. *Cancer Biol Ther* 2020; 21: 637-646.
- [88] Xu H, Ma Y, Zhang J, Gu J, Jing X, Lu S, Fu S and Huo J. Identification and verification of core genes in colorectal cancer. *Biomed Res Int* 2020; 2020: 8082697.
- [89] Yang J, Song P and Zhou G. A study on the correlations of MRP-1 expression with the pathogenesis and prognosis of colorectal cancer. *J BUON* 2019; 24: 84-90.
- [90] Lv Y, Zhao S, Han J, Zheng L, Yang Z and Zhao L. Hypoxia-inducible factor-1 α induces multidrug resistance protein in colon cancer. *Onco Targets Ther* 2015; 8: 1941-1948.
- [91] Andersen V, Svenningsen K, Knudsen LA, Hansen AK, Holmskov U, Stensballe A and Vogel U. Novel understanding of ABC transporters ABCB1/MDR/P-glycoprotein, ABCC2/MRP2, and ABCG2/BCRP in colorectal pathophysiology. *World J Gastroenterol* 2015; 21: 11862-11876.
- [92] He X, Ma J, Zhang M, Cui J and Yang H. Circ_0007031 enhances tumor progression and promotes 5-fluorouracil resistance in colorectal cancer through regulating miR-133b/ABCC5 axis. *Cancer Biomark* 2020; 29: 531-542.
- [93] Zhang L, Li B, Zhang B, Zhang H and Suo J. miR-361 enhances sensitivity to 5-fluorouracil by targeting the FOXM1-ABCC5/10 signaling pathway in colorectal cancer. *Oncol Lett* 2019; 18: 4064-4073.
- [94] Andersen V, Vogel LK, Kopp TI, Sæbø M, Nonboe AW, Hamfjord J, Kure EH and Vogel U. High ABCC2 and low ABCG2 gene expression are early events in the colorectal adenoma-carcinoma sequence. *PLoS One* 2015; 10: e0119255.
- [95] Tian T, Bi H, Zhang D, Liu Y, Sun H, Jia C, Zheng T, Huang H, Fu J, Zhu L and Zhao Y. Methylation of three genes encoded by X chromosome in blood leukocytes and colorectal cancer risk. *Cancer Med* 2021; 10: 4964-4976.
- [96] Wong MC, Ding H, Wang J, Chan PS and Huang J. Prevalence and risk factors of colorectal cancer in Asia. *Intest Res* 2019; 17: 317-329.
- [97] Kang X, Zhang R, Kwong TN, Lui RN, Wu WK, Sung JJ, Yu J and Wong SH. Serrated neoplasia in the colorectum: gut microbiota and molecular pathways. *Gut Microbes* 2021; 13: 1-12.
- [98] Chen Q, Wang H, Li Z, Li F, Liang L, Zou Y, Shen H, Li J, Xia Y, Cheng Z, Yang T, Wang K and Shen F. Circular RNA ACTN4 promotes intrahepatic cholangiocarcinoma progression by recruiting YBX1 to initiate FZD7 transcription. *J Hepatol* 2022; 76: 135-147.
- [99] Yang J, Sontag D, Gong Y and Minuk GY. Enhanced gemcitabine cytotoxicity with knock-down of multidrug resistance protein genes in human cholangiocarcinoma cell lines. *J Gastroenterol Hepatol* 2021; 36: 1103-1109.
- [100] Jayavelu AK, Wolf S, Buettner F, Alexe G, Häupl B, Comoglio F, Schneider C, Doebele C, Fuhrmann DC, Wagner S, Donato E, Andresen C, Wilke AC, Zindel A, Jahn D, Splettstoesser B,

- Plessmann U, Münch S, Abou-El-Ardat K, Makowka P, Acker F, Enssle JC, Cremer A, Schnütgen F, Kurrle N, Chapuy B, Löber J, Hartmann S, Wild PJ, Wittig I, Hübschmann D, Kaderali L, Cox J, Brüne B, Röhlig C, Thiede C, Steffen B, Bornhäuser M, Trumpp A, Urlaub H, Stegmaier K, Serve H, Mann M and Oellerich T. The proteogenomic subtypes of acute myeloid leukemia. *Cancer Cell* 2022; 40: 301-317, e312.
- [101] Miyamoto K and Minami Y. Cutting edge molecular therapy for acute myeloid leukemia. *Int J Mol Sci* 2020; 21: 5114.
- [102] Handschuh L, Wojciechowski P, Kazmierczak M and Lewandowski K. Transcript-level dysregulation of BCL2 family genes in acute myeloblastic leukemia. *Cancers (Basel)* 2021; 13: 3175.
- [103] Lu X, Han L, Busquets J, Collins M, Lodi A, Marszałek JR, Konopleva M and Tiziani S. The combined treatment with the FLT3-inhibitor AC220 and the complex I inhibitor IACS-010759 synergistically depletes Wt- and FLT3-mutated acute myeloid leukemia cells. *Front Oncol* 2021; 11: 686765.
- [104] Rodríguez González A, Sahores A, Díaz-Nebreda A, Yaneff A, Di Siervi N, Gómez N, Monczor F, Fernández N, Davio C and Shayo C. MRP4/ABCC4 expression is regulated by histamine in acute myeloid leukemia cells, determining cAMP efflux. *FEBS J* 2021; 288: 229-243.
- [105] Fisusi FA and Akala EO. Drug combinations in breast cancer therapy. *Pharm Nanotechnol* 2019; 7: 3-23.
- [106] Lin Q, Chen X, Meng F, Ogawa K, Li M, Song R, Zhang S, Zhang Z, Kong X, Xu Q, He F, Liu D, Bai X, Sun B, Hung MC, Liu L, Wands JR and Dong X. Multi-organ metastasis as destination for breast cancer cells guided by biomechanical architecture. *Am J Cancer Res* 2021; 11: 2537-2567.
- [107] Kochel TJ, Reader JC, Ma X, Kundu N and Fulton AM. Multiple drug resistance-associated protein (MRP4) exports prostaglandin E2 (PGE2) and contributes to metastasis in basal/triple negative breast cancer. *Oncotarget* 2017; 8: 6540-6554.
- [108] Dietze R, Hammoud MK, Gómez-Serrano M, Unger A, Bieringer T, Finkernagel F, Sokol AM, Nist A, Stiewe T, Reinartz S, Ponath V, Preußner C, von Strandmann EP, Müller-Brüsselbach S, Graumann J and Müller R. Phosphoproteomics identify arachidonic-acid-regulated signal transduction pathways modulating macrophage functions with implications for ovarian cancer. *Theranostics* 2021; 11: 1377-1395.
- [109] Yang J, Zaman MM, Vlasakov I, Roy R, Huang L, Martin CR, Freedman SD, Serhan CN and Moses MA. Adipocytes promote ovarian cancer chemoresistance. *Sci Rep* 2019; 9: 13316.
- [110] Zhang Q, Yan G, Lei J, Chen Y, Wang T, Gong J, Zhou Y, Zhao H, Chen H, Zhou Y, Wu L, Zhang J, Zhang X, Wang J and Li Y. The SP1-12LOX axis promotes chemoresistance and metastasis of ovarian cancer. *Mol Med* 2020; 26: 39.
- [111] Sedláková I, Laco J, Caltová K, Červinka M, Tošner J, Řezáč A and Špaček J. Clinical significance of the resistance proteins LRP, Pgp, MRP1, MRP3, and MRP5 in epithelial ovarian cancer. *Int J Gynecol Cancer* 2015; 25: 236-243.
- [112] Zhang H, Hu L, Cheng M, Wang Q, Hu X and Chen Q. The Hedgehog signaling pathway promotes chemotherapy resistance via multidrug resistance protein 1 in ovarian cancer. *Oncol Rep* 2020; 44: 2610-2620.
- [113] Sedláková I, Laco J, Tošner J and Špaček J. [Prognostic significance of Pgp, MRP1, and MRP3 in ovarian cancer patients]. *Ceska Gynekol* 2015; 80: 405-413.
- [114] Wu L, Li Y, Song Y, Zhou D, Jia S, Xu A, Zhang W, You H, Jia J, Huang J and Ou X. A recurrent ABCC2 p.G693R mutation resulting in loss of function of MRP2 and hyperbilirubinemia in Dubin-Johnson syndrome in China. *Orphanet J Rare Dis* 2020; 15: 74.
- [115] Wu H, Zhao XK and Zhu JJ. Clinical characteristics and ABCC2 genotype in Dubin-Johnson syndrome: a case report and review of the literature. *World J Clin Cases* 2021; 9: 878-885.
- [116] You SJ, Sun YX, Zhang J, He Q, Wu XM and Hu Y. Literature review and report of three cases of Dubin-Johnson syndrome related to ABCC2 gene mutations in children. *Am J Transl Res* 2021; 13: 4477-4488.
- [117] Meng LL, Qiu JW, Lin WX and Song YZ. Clinical features and ABCC2 genotypic analysis of an infant with Dubin-Johnson syndrome. *Zhongguo Dang Dai Er Ke Za Zhi* 2019; 21: 64-70.
- [118] Borst P, Váradi A and van de Wetering K. PXE, a mysterious inborn error clarified. *Trends Biochem Sci* 2019; 44: 125-140.
- [119] Link SL and Nayak RP. Review of rapid advances in cystic fibrosis. *Mo Med* 2020; 117: 548-554.
- [120] Li Q, van de Wetering K and Uitto J. Pseudoxanthoma elasticum as a paradigm of heritable ectopic mineralization disorders: pathomechanisms and treatment development. *Am J Pathol* 2019; 189: 216-225.
- [121] Shimada BK, Pomozi V, Zoll J, Kuo S, Martin L and Le Saux O. ABCC6, pyrophosphate and ectopic calcification: therapeutic solutions. *Int J Mol Sci* 2021; 22: 4555.
- [122] Czimer D, Porok K, Csete D, Gyüre Z, Lavró V, Fülöp K, Chen Z, Gyergyák H, Tusnády GE, Burgess SM, Mócsai A, Váradi A and Varga M. A new zebrafish model for pseudoxanthoma elasticum. *Front Cell Dev Biol* 2021; 9: 628699.

- [123] Luo H, Li Q, Cao Y and Uitto J. Therapeutics development for pseudoxanthoma elasticum and related ectopic mineralization disorders: update 2020. *J Clin Med* 2020; 10: 114.
- [124] Szeri F, Corradi V, Niaziorimi F, Donnelly S, Conseil G, Cole SPC, Tieleman DP and van de Wetering K. Mutagenic analysis of the putative ABCC6 substrate-binding cavity using a new homology model. *Int J Mol Sci* 2021; 22: 6910.
- [125] Ziegler SG, Ferreira CR, MacFarlane EG, Riddle RC, Tomlinson RE, Chew EY, Martin L, Ma CT, Sergienko E, Pinkerton AB, Millán JL, Gahl WA and Dietz HC. Ectopic calcification in pseudoxanthoma elasticum responds to inhibition of tissue-nonspecific alkaline phosphatase. *Sci Transl Med* 2017; 9: eaa1669.
- [126] Ibold B, Tiemann J, Faust I, Ceglarek U, Dittrich J, Gorgels T, Bergen AAB, Vanakker O, Van Gils M, Knabbe C and Hendig D. Genetic deletion of Abcc6 disturbs cholesterol homeostasis in mice. *Sci Rep* 2021; 11: 2137.
- [127] Bisaccia F, Koshal P, Abruzzese V, Castiglione Morelli MA and Ostuni A. Structural and functional characterization of the ABCC6 transporter in hepatic cells: role on PXE, cancer therapy and drug resistance. *Int J Mol Sci* 2021; 22: 2858.
- [128] Huang WG, Wang J, Liu YJ, Wang HX, Zhou SZ, Chen H, Yang FW, Li Y, Yi Y and He YH. Endoplasmic reticulum stress increases multidrug-resistance protein 2 expression and mitigates acute liver injury. *Curr Mol Med* 2020; 20: 548-557.
- [129] Chen J, Wu H, Tang X and Chen L. 4-phenylbutyrate protects against rifampin-induced liver injury via regulating MRP2 ubiquitination through inhibiting endoplasmic reticulum stress. *Bioengineered* 2022; 13: 2866-2877.
- [130] Bieber T. Atopic dermatitis: an expanding therapeutic pipeline for a complex disease. *Nat Rev Drug Discov* 2022; 21: 21-40.
- [131] Narla S, Silverberg JI and Simpson EL. Management of inadequate response and adverse effects to dupilumab in atopic dermatitis. *J Am Acad Dermatol* 2022; 86: 628-636.
- [132] Lee HS and Jeong GS. Therapeutic effect of kaempferol on atopic dermatitis by attenuation of T cell activity via interaction with multidrug resistance-associated protein 1. *Br J Pharmacol* 2021; 178: 1772-1788.
- [133] Funahashi Y, Takahashi R, Mizoguchi S, Suzuki T, Takaoka E, Ni J, Wang Z, DeFranco DB, de Groat WC, Tyagi P and Yoshimura N. Bladder overactivity and afferent hyperexcitability induced by prostate-to-bladder cross-sensitization in rats with prostatic inflammation. *J Physiol* 2019; 597: 2063-2078.
- [134] Alexandre EC, Cao N, Mizoguchi S, Saito T, Kurobe M, Gotoh D, Okorie M, Igarashi T, Antunes E and Yoshimura N. Urethral dysfunction in a rat model of chemically induced prostatic inflammation: potential involvement of the MRP5 pump. *Am J Physiol Renal Physiol* 2020; 318: F754-F762.
- [135] Rife E and Gedalia A. Kawasaki disease: an update. *Curr Rheumatol Rep* 2020; 22: 75.
- [136] Tirelli F, Marrani E, Giani T and Cimaz R. One year in review: Kawasaki disease. *Curr Opin Rheumatol* 2020; 32: 15-20.
- [137] Menikou S, Langford PR and Levin M. Kawasaki disease: the role of immune complexes revisited. *Front Immunol* 2019; 10: 1156.
- [138] Sakurai Y. Autoimmune aspects of Kawasaki disease. *J Investig Allergol Clin Immunol* 2019; 29: 251-261.
- [139] Soni PR, Noval Rivas M and Ardit M. A comprehensive update on kawasaki disease vasculitis and myocarditis. *Curr Rheumatol Rep* 2020; 22: 6.
- [140] Zhao QM, Chu C, Wu L, Liang XC, Sun SN, He L, Zhao L, Wang F, Huang GY, Niu C and Liu F. Systemic artery aneurysms and Kawasaki disease. *Pediatrics* 2019; 144: e20192254.
- [141] Liu X, Zhou K, Hua Y, Wu M, Liu L, Shao S and Wang C. Neurological involvement in Kawasaki disease: a retrospective study. *Pediatr Rheumatol Online J* 2020; 18: 61.
- [142] Wu S, Liao Y, Sun Y, Zhang CY, Zhang QY, Yan H, Qi JG, Liu XQ, Chen YH, Wang YL, Li XY, Jin HF and Du JB. Prediction of intravenous immunoglobulin resistance in Kawasaki disease in children. *World J Pediatr* 2020; 16: 607-613.
- [143] Luo C and Zhang H. The role of proinflammatory pathways in the pathogenesis of colitis-associated colorectal cancer. *Mediators Inflamm* 2017; 2017: 5126048.
- [144] Villeda-Ramírez MA, Meza-Guillen D, Barreto-Zúñiga R and Yamamoto-Furusho JK. ABCC7/CFTR expression is associated with the clinical course of ulcerative colitis patients. *Gastroenterol Res Pract* 2021; 2021: 5536563.
- [145] Cui J and Jia J. Natural COX-2 inhibitors as promising anti-inflammatory agents: an update. *Curr Med Chem* 2021; 28: 3622-3646.
- [146] Chen SF, Zhang ZY and Zhang JL. Meloxicam increases intracellular accumulation of doxorubicin via downregulation of multidrug resistance-associated protein 1 (MRP1) in A549 cells. *Genet Mol Res* 2015; 14: 14548-14560.
- [147] Chun SY, Kwon YS, Nam KS and Kim S. Lapatinib enhances the cytotoxic effects of doxorubicin in MCF-7 tumorspheres by inhibiting the drug efflux function of ABC transporters. *Biomed Pharmacother* 2015; 72: 37-43.
- [148] Kwon YS, Chun SY, Nam KS and Kim S. Lapatinib sensitizes quiescent MDA-MB-231 breast cancer cells to doxorubicin by inhibiting the expression of multidrug resistance-associated protein-1. *Oncol Rep* 2015; 34: 884-890.

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- [149] Wang H, Jia XH, Chen JR, Yi YJ, Wang JY, Li YJ and Xie SY. HOXB4 knockdown reverses multidrug resistance of human myelogenous leukemia K562/ADM cells by downregulating P-gp, MRP1 and BCRP expression via PI3K/Akt signaling pathway. *Int J Oncol* 2016; 49: 2529-2537.
- [150] Song YP, Xian P, Luo H, Dai JY, Bai Y, Li Y and Tang XL. Comprehensive landscape of HOXA2, HOXA9, and HOXA10 as potential biomarkers for predicting progression and prognosis in prostate cancer. *J Immunol Res* 2022; 2022: 5740971.
- [151] Yi YJ, Jia XH, Wang JY, Li YJ, Wang H and Xie SY. Knockdown of HOXA10 reverses the multidrug resistance of human chronic myelogenous leukemia K562/ADM cells by downregulating P-gp and MRP-1. *Int J Mol Med* 2016; 37: 1405-1411.

Supplementary Materials

The protein sequences of human MRP1-9 from the Unified Protein Data Bank are described as follows.

1. Multidrug resistance-associated protein 1 • Homo sapiens (Human)
2. ATP-binding cassette sub-family C member 2 • Homo sapiens (Human)
3. ATP-binding cassette sub-family C member 3 • Homo sapiens (Human)
4. ATP-binding cassette sub-family C member 4 • Homo sapiens (Human)
5. ATP-binding cassette sub-family C member 5 • Homo sapiens (Human)
6. ATP-binding cassette sub-family C member 6 • Homo sapiens (Human)
7. ATP-binding cassette sub-family C member 10 • Homo sapiens (Human)
8. ATP-binding cassette sub-family C member 11 • Homo sapiens (Human)
9. ATP-binding cassette sub-family C member 12 • Homo sapiens (Human)

Multidrug resistance-associated protein 1 • Homo sapiens (Human) amino acid sequence:

>sp|P33527|MRP1_HUMAN Multidrug resistance-associated protein 1 OS=Homo sapiens OX=9606
GN=ABCC1 PE=1 SV=3

MALRGFCSADGSDPLWDWNVTWNTSNPDFTKCFQNTVLVWVPCFYLWACFPFYFLYLSRH
DRGYIQMTPLNKTALGFLWIVCWADLFYSFWERSRGIFLAPVFLVSPTLLGITMLLA
TFLIQLERRKGVQSSGIMLTFWLVALVCALILRSKIMTALKEDAQVDLFRDITFYVYFS
LLLIQLVLSCFSDRSPLFSETIHDPNPCPESSASFLSRITFWWITGLIVRGYRQPLEGSD
LWSLNKEDTSEQVVPVLVKNWKKCAKTRKQPVKVVYSSKDPAQPKESSKVDANEEVEAL
IVKSPQKEWNPSLFKVLYKTFGPYFLMSFFFKAIHDLMMFSGPQILKLLIKFVNDTKAPD
WQGYFYTVLLFVTACLQTLVLHQYFHICFVSGMRIKTAVIGAVYRKALVITNSARKSSTV
GEIVNLMSVDAQRFMDLATYINMIWSAPLQVILALYLLWNLGPSVLAGVAVMVLMPVN
AVMAMKTKTYQVAHMKSKDNRIKLMNEILNGIKVLKLYAWELAFKDKVLAIRQEELKVLK
KSAYLSAVGTFTWVCTPFLVALCTFAVYVTIDENNILDAQTAFVSLALFNILRFPLNILP
MVISSIVQASVSLKRLRIFLSHEELEPDSEIERRPVKDGGGTNSITVRNATFTWARS DPPT
LNGITFSIPEGALVAVVGQVCGKSSLLSALLAEMDKVEGHVAIKGSVAYVPQQAWIQND
SLRENILFGCQLEEPYRSVIQACALLPDLEILPSGDRTEIGEKGVNLSGGQKQQRVSLAR
AVYSNADIYLFDDPLSAVDAHVGKHIFENVIGPKGMLKNKTRILVTHSMSYLPQVDVIIV
MSGGKISEMGSYQELLARDGAFAEFLRTYASTEQEQAEEENGVTGVSGPGKEAKQMENG M
LVTD SAGKQLQRQLSSSSSYSGDISRHHNSTAELQKAEAKKEETWKLMEADKAQTGQVKL

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SVYWDYMKAIGLFISFLSIFLFCMNHVSALASNYWLSLWTDPIVNGTQEHTKVRLSVYG
ALGISQGIAVFGYSMAVSIGGILASRCLHVDLLHSILRSPMSFFERTPSGNLVNRFSKEL
DTVDSMIPEVIKMFMGSLFNVIGACIVILLATPIAAIIPPLGLIYFFVQRFYVASSRQL
KRLESVSRSPVYSHFNETLLGVSVIRAFEEQERFIHQSDLKVDENQKAYYPSIVANRWLA
VRLECVGNCIVLFAALFAVISRHLSAGLVGLSVSYSLQVTTYLNWLVRMSSEMETNIVA
VERLKEYSETEKEAPWQIQETAPPSSWPQVGRVEFRNYCLRYREDLDFVLRHINVTINGG
EKVGIVGRTGAGKSSLTGLFRINESAEGEIIGDGINIAKIGLHDLRFKITIIPQDPVLF
SGSLRMNLDPFQSQYSDEEVWTSLELAHLKDFVSALPDKLDHECAEGGENLSVGQRQLVCL
ARALLRKTILVLDEATAAVDLETDLIQSTIRTQFEDCTVLTIAHRLNTIMDYTRVIVL
DKGEIQEYGAPSDLLQQRGLFYMAKDAGLV

Human MRP1:

Seq Identity

90.52%

Template

6uy0.1. A Multidrug resistance-associated protein 1

Cryo-EM structure of wild-type bovine multidrug resistance protein 1 (MRP1) under active turnover conditions

ATP-binding cassette sub-family C member 2 • Homo sapiens (Human) amino acid sequence:

>sp|Q92887|MRP2_HUMAN ATP-binding cassette sub-family C member 2 OS=Homo sapiens OX=9606
GN=ABCC2 PE=1 SV=3

MLEKFCNSTFWNSSFLDSPEADPLCFEQTVLVWIPLGYLWLLAPWQLLHVYKSRTKRSS
TTKLYLAKQVFGVFLILAAIELALVLTEDSGQATVPAVRYTNPSLYLGTWLLVLLIQYS
RQWCVCQKNSWFLSLFWILSILCGTFQFQTLIRTLQGDNSNLAYSCLFFISYGFQILILI
FSAFSENNESSNNPSSIASFLSSITYSWYDSIILKGYKRPLTEDVWEVDEEMKTKTLVS
KFETHMKRELQKARRALQRRQEKSSQQNSGARLPGLNKNQSQSQDALVLEDVEKKKKKSG
TKKDVPKSWLMKALFKTFYMVLLKSFLLKLVNDIFTFVSPQLLKLLISFASDRDTYLVIG
YLCAILLFTAALIQSFCLQCYFQLCFKLGVKVRTAIMASVYKKALTLSNLARKEYTVGET
VNLMSVDAQKLM DVTNFMHMLWSSVLQIVLSIFFLWRELGPSVLAVGVMMVLVIPINAIL
STKSKTIQVKNMKNKDKRLKIMNEILSGIKILKYFAWEPSFRDQVQNLRRKKELKNLLAFS
QLQCVVIFVFQLPVLVSVVTFVYVLVDSNNILDAQKAFSTITLNFILRFPLSMLPMMI
SSMLQASVSTERLEKYLGGDDLDTSAIRHDCNFDKAMQFSEASFTWEHDSEATVRDVNLD

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IMAGQLVAVIGPVGSGKSSLSISAMLGEMENVHGHITIKGTTAYVPQQSWIQNGTIKDNL
FGTEFNEKRYQQVLEACALLPDLEMLPGGDLAIEGEGKINLSGGQKQRISLARATYQNLD
IYLLDDPLSAVDAHVGKHIFNKVLGPNGLLKGGKTRLLVTHSMHFLPQVDEIVVLGNGTIV
EKGSYSALLAKKGEFAKNLKTFLRHTGPEEEATVHDGSEEEEDDYGLISSVEEIPEDAAS
ITMRRENSFRRTLRSRRSRNGRHLKSLRNSLKTRNVNSLKEDEELVKGQKLIKKEFIETG
KVKFSIYLEYLQAIGLFSIFFIILAFVMNSVAFIGSNLWLSAWTSDSKIFNSTDYPASQR
DMRVGVYGALGLAQGIFVFIAHFWSAFGFVHASNILHKQLLNNILRAPMRFFDTTPTGRI
VNRFAGDISTVDDTL PQSLRSWITCFLGIISTLVMICMATPVFTIIVIPLGIIYVSQMF
YVSTSRQLRRLDSVTRSPIYSHFSETVSGLPVIRAFEHQQRFLKHNEVRIDTNQKCVFSW
ITSNRWLAI RLELVGNLTVFFSALMMVIYRDTLSGDTVGFVLSNALNITQTLNWLVRMTS
EIETNIVAVERITEYTKVENEAPWVTDKRPPPDWPSKGKIQFNQYQVRYRPELDLVL RGI
TCDIGSMEKIGVVGRGTGAGKSSLTNCLFRILEAAGGQIIDGVDIASIGLHDLREKLTII
PQDPILFSGSLRMNLDPFNNYSDEEIWKALELAHLKSFVASLQLGLSHEVTEAGGNLSIG
QRQLLCLGRALLRKSKILVLDEATAAVDLETDNLIQTTIQNEFAHCTVITIAHRLHTIMD
SDKVMVL DNGKIIECGSPEELLQIPGPFYFMAKEAGIENVNSTKF

Human MRP2:

Seq Identity

51.02%

Template

5uj9.1. A bovine multidrug resistance protein 1 (MRP1), Multidrug resistance-associated protein 1

Cryo-EM structure of bovine multidrug resistance protein 1 (MRP1)

ATP-binding cassette sub-family C member 3 • Homo sapiens (Human) amino acid sequence:

>sp|O15438|MRP3_HUMAN ATP-binding cassette sub-family C member 3 OS=Homo sapiens OX=9606
GN=ABCC3 PE=1 SV=3

MDALCGSGELGSKFWDSNLSVHTENPDLT PCFQNSLLAWVPCIYLWVALPCYLLYLRHHC
RGYIILSHLSKLMVLGVLLWCVSWADLFYSFHGLVHGRAPAPVFFVTPLVVGVTMLLAT
LLIQYERLQGVQSSGVLIIFWFLCVVCAIVPFRSKILLAKAEGEISDPFRFTTFYIH FAL
VLSALILACFREKPPFFSAKNVDPNPYPETSAGFLSRLFFWWFTKMAIYGYRH PLEEKDL
WSLKEEDRSQM VVQQLLEAWRKQEKQTARHKASAAPGKNASGEDEVLLGARPRPRKPSFL
KALLATFGSSFLISACFKLIQDLLSFINPQLLSILIRFISNPMAPSWWGFLVAGLMFLCS
MMQSLILQHYYHYIFVTGVKFRGTGIMGVYRKALVITNSVKRASTVGEIVNLMSVDAQRF

The role of MRPs in diseases

MDLAPFLNLLWSAPLQIILAIYFLWQNLGPSVLGVAFMVLLIPLNGAVAVKMRAFQVKQ
MKLKDSRIKLMSEILNGIKVLKLYAWEPSFLKQVEGIRQGELQLLRTAAYLHTTTTFTWM
CSPFLVTLITLWVYVYVDPNNVLDAEKAFVSVSLFNILRLPLNMLPQLISNLTQASVSLK
RIQQFLSQEELDPQSVERKTISPGYAITIHSGTFTWAQDLPTLHSLDIQVPKGALVAVV
GPVGCCKSSLVSALLGEMEKLEGKVHMKGSVAYVPQQAWIQNCTLQENVLFGKALNPKRY
QQTLEACALLADLEMLPGGDQTEIGEKGINLSGGQRQRVSLARAVYSDADIFLLDDPLSA
VDSHVAKHIFDHVIGPEGVLGKTRVLVTHGISFLPQTDFIIVLADGQVSEMGYPALLQ
RNGSFANFLCNYPDEDDQGHLEDSWTALEGAEDKEALLIEDTLNHTDLTDNDPVTYVVQ
KQFMRQLSALSSDGEGQGRPVPRRHLPSEKVQVTEAKADGALTQEEKAAIGTVELSVFW
DYAKAVGLCTTLAICLLYVGQSAAGANVWLSAWTNDAMADSRQNNTSLRLGVYAALGI
LQGFLVMLAAMAMAAGGIQAARVLHQALLHNKIRSPQSFFDTTPSGRILNCFSKDIYVVD
EVLAPVILMLLNSFFNAISTLVVIMASTPLFTVVILPLAVLYTLVQRFYAATSRQLKRLE
SVSRSPIYSHFSETVTGASVIRAYNRSRDFEISDTKVDANQRSCYPYIISNRWLSIGVE
FVGNCVVLFAALFAVIGRSSLNPGVLGLSVSYSLQVTFALNWMIRMMMSDLESNIVAVERV
KEYSKTETEAPWVVEGSRPPEGWPPRGEVEFRNYSVRYRPGDLVLRDLSLHVHGGEKVG
IVGRTGAGKSSMTLCLFRILEAAKGEIRIDGLNVADIGLHDLRSQLTIIQDPILFSGTL
RMNLDPFGSYSEEDIWWALELSHLHTFVSSQPAGLDFQCSEGGENLSVGQRQLVCLARAL
LRKSRLVLDEATAAIDLETDNLIQATIRTQFDTCTVLTIAHRLNTIMDYTRVLVLDKGV
VAEFDSPANLIAARGIFYGMARDAGLA

Human MRP3:

Seq Identity

57.22%

Template

5uj9.1. A bovine multidrug resistance protein 1 (MRP1), Multidrug resistance-associated protein 1

Cryo-EM structure of bovine multidrug resistance protein 1 (MRP1)

ATP-binding cassette sub-family C member 4 • Homo sapiens (Human) amino acid sequence:

>sp|O15439|MRP4_HUMAN ATP-binding cassette sub-family C member 4 OS=Homo sapiens OX=9606
GN=ABCC4 PE=1 SV=3

MLPVYQEVKPNPLQDANLCSRFFFWLNLPLFKIGHKRRLEEDDMYSVLPEDRSQHLGEEL
QGFWDKVELRAENDAQKPSLTRAIIKCYWKSYLEVLGIFTLIEESAKVIQPIFLGKIINYF
ENYDPMDSVALNTAYAYATVLTFTLILAILHHLYFYHVQCAGMRLRVAMCHMIYRKALR

The role of MRPs in diseases

LSNMAMGKTTTGQIVNLLSNDVNKFDQVTVFLHFLWAGPLQAIQAVTALLWMEIGISCLAG
MAVLIILLPLQSCFGKLFSSLRSKTATFTDARITMNEVITGIRIIKMYAWEKSFSNLIT
NLRKKEISKILRSSCLRGMNLASFFSASKIIVFTFTTYVLLGSVITASRVFVAVTLYGA
VRLTVTLFFPSAIERVSEAIVSIRRIQTFLLLDEISQRNRQLPSDGKKMVHVQDFTAFWD
KASEPTLQGLSFTVRPGELLAVVGPVGAGKSSLLSAVLGELAPSHGLVSVHGRIAYVSQ
QPWVFSGTLRSNILFGKKYKERYEKVIKACALKKDLQLEDGDLTVIGDRGTTLSGGQK
ARVNLARAVYQDADIYLLDDPLSAVDAEVSRLHFLCICQILHEKITILVTHQLQYLKAA
SQILILKDGKMOVQKGTYTEFLKSGIDFGSLLKKDNEESEQPPVPGTPTLRNRTFSESSVW
SQQSSRPSLKDGALESQDTENVPVTLSEENRSEKVGFGAYKNYFRAGAHWIVFIFLILL
NTAAQVAYVLQDWWSYWANKQSMNLNVTVNGGGNVTEKLDLNWYLGISGLTVATVLFGI
ARSLLVFYVLNSSQTLHNKMFESILKAPVLFDRNPGRILNRFSKDIGHLDDLLPLTF
LDFIQTLLQVGVVSVAVAVIPWIAIPLVPLGIIFLRRYFLETSRDVKRLESTTRSPV
FSLHSSSLQGLWTIRAYKAEERCQELFDAHQDLHSEAWFLFTTSRWFVRLDAICAMFV
IIVAFGSLILAKTLTAGVGLALSALTLMGMFQWCVRQSAEVENMMISVERVIEYTDLE
KEAPWEYQKRPPPAWPHEGVIIFDNVNFMYSPPGGLVLKHLTALIKSQEKVGIVGRTGAG
KSSLISALFRLSEPEGKIWDKILTTEIGLHDLRKKMSIIPQEPVLTGTMRKNLDPFNE
HTDEELWNALQEVQLKETIEDLPGKMDTELAESGSNFSVGQRQLVCLARAILRKNQILII
DEATANVDPRTDELIIQKKIREKFAHCTVLTIAHRLNTIISDKIMVLDSGRLKEYDEPYV
LLQNKESLFYKMOVQQLGKAEAAALTETAKQVYFKRNYPHIGHTDHMTNTSNGQPSTLTI
FETAL

Human MRP4:

Seq Identity

39.97%

Template

7m69.1. A Metal resistance protein YCF1

E1435Q Ycf1 mutant in inward-facing wide conformation

ATP-binding cassette sub-family C member 5 • Homo sapiens (Human) amino acid sequence:

>sp|O15440|MRP5_HUMAN ATP-binding cassette sub-family C member 5 OS=Homo sapiens OX=9606
GN=ABCC5 PE=1 SV=2

MKDIDIGKEYIIPSPGYRSVRERTSTSGTHRDREDSKFRRTTRPLECQDALETAARAEGLS
LDASMHSQRLRLDEEHPKGYHHGLSALKPIRTTSKHQHPVDNAGLFSCMTFSWLSSLAR

The role of MRPs in diseases

VAHKKGELSMEDVWSLSKHESSDVNCRRRLERLWQEELNEVGPDAASLRRVWVFCRTRLI
LSIVCLMITQLAGFSGPAFMVKHLLEYTQATESNLQYSLLLVLGLLLTEIVRSWSLALTW
ALNYRTGVRLRGAILTMAFKKILKLKNIKEKSLGELINICSNDGQRMFEAAVGSLLAGG
PVVAILGMIYNVILGPTGFLGSAFIFYPAMMFASRLTAYFRRKCVAATDERVQKMNE
VLTYIKFIKMYAWVKAFSQSVQKIREEEERRILEKAGYFQSITVGVAPIVVVIASVVTFSV
HMTLGFDLTAAQFTVVTVFNMTFALKVTPFSVKSLSEASVAVDRFKSLFLMEEVHMIK
NKPASPHIKIEMKNATLAWDSSHSSIQNSPKLTPKMKKDKRASRGKKEKVRQLQRTEHQA
VLAEQKGHLLLDSDERPSPEEEEGKHIHLGHLRLQRTLHSIDLEIQEGKLVGICGSGVSGG
KTSLSAILGQMTLLEGSIAISGTFAYVAQQAWILNATLRDNILFGKEYDEERYNSVLNS
CCLRPDLAILPSSDLTEIGERGANLSGGQRQRISLARALYSDRSIYILDDPLSALDAHVG
NHIFNSAIRKHLKSKTVLFVTHQLQYLVDCEVIFMKEGCITERGTHEELMNLNGDYATI
FNNLLLGETPPVEINSKKETSGSQKKSQDKGPKTGSVKKEKAVKPEEGQLVQLEEKQGS
VPWSVYGVIYAAGGPLAFLVIMALFMLNVGSTAFSTWWLSYWIKQSGNNTTVTRGNETS
VSDSMKDNPHMQYYASIYALSMAVMLILKAIRGVVFKGTLRASSRLHDELFRILRSPM
KFFDTPPTGRILNRFKDMDEVDVRLPFQAEMFIQNVILVFFCVGMIAGVFPWFLVAVGP
LVILFVLHIVSRVLIRELKRLDNITQSPFLSHITSSIQGLATIHAYNKGQEFLEHRYQEL
LDDNQAPFFLFTCAMRWLAVRLDLISIALITTTGLMIVLMHGQIPPAYAGLAISYAVQLT
GLFQFTVRLASETEARFTSVERINHYIKTSLSEAPARIKNKAPSPDWPQEGEVTFENAEM
RYRENPLVLKKSFTIKPKEKIGIVGRTGSGKSSLGMALFRLVELSGGCIKIDGVRISD
IGLADLRSLKLSIIPQEPVLFSGTVRSNLDPFNQYTEDQIWDALERTHMKECIAQLPLKLE
SEVMENGDNFSVGERQLLCIARALLRHCKILILDEATAAMDTELDLLIQETIREAFADCT
MLTIAHRLHTVLGSDRIMVLAQGQVVEFDTPSVLLSNDSSRFYAMFAAAENKVAVKG

Human MRP5:

Seq Identity

36.60%

Template

6uy0.1. A Multidrug resistance-associated protein 1

Cryo-EM structure of wild-type bovine multidrug resistance protein 1 (MRP1) under active turnover conditions

ATP-binding cassette sub-family C member 6-Homo sapiens (Human) amino acid sequence:

>sp|O95255|MRP6_HUMAN ATP-binding cassette sub-family C member 6 OS=Homo sapiens OX=9606
GN=ABCC6 PE=1 SV=2

The role of MRPs in diseases

MAAPAEP CAGQG VWNQTEPEPAAT SLLSLCFLRTAGVWVPPMYLWVLGP IYLLFIHHHGR
GYLRMSPLFKAKMVLGFALIVLCTSSVAVALWKIQQGTPEAPEFLIHPTVWLT TMSFAVF
LIHTERKKGVQSSGVLF GYWLLCFVLPATNAAQQASGAGFQSDPVRHLSTYLCLSLVVAQ
FVLSCLADQPPFFPEDPQQSNPCPETGA AFPSKATFWWVSGLVWRGYRRPLRPKDLWSLG
RENSSEELVSRLEKEWMRNRSAARRHNKAI AFKRKGGSGMKAPETEPFLRQEGSQWRPLL
KAIWQVFHSTFLLGTLSLIISDVFRFTVPKLLSLFLEFIGDPKPPAWKGYLLAVLMFLSA
CLQTLFEQQNMYRLKVLQMRLRSAITGLVYRKVLALSSGSRKASAVGDVVNLVSV DVQRL
TESVLYLNGLWLPLVWIVVCFVYLWQLLGPSALTAIAVFLSLLPLNFFISKRNHHQEEQ
MRQKDSRARLTSSILRNSKTIKFHGWEGAFLDRVLGIRGQELGALRTSGLLFSVSLVSFQ
VSTFLVALVFAVHTLVAENAMNAEKAFVTLTVLNILNKAQAFLPFSIHSVLQARVSFDR
LVTFLCLEEVDPGVVDSSSSGSAAGKDCITIHSATFAWSQESPPCLHRINLTPQGCLLA
VVGPGVAGKSSLLSALLGELSKVEGFVSIEGAVAYVPQEAWVQNTSVVENVCFGQELDPP
WLERVLEACALQPDVDSFPEGIHTSIGEQGMNLSGGQKQRLSLARAVYRKA AVYLLDDPL
AALDAHVGQHVFNQVIGPGGLLQGTT RILVTHALHILPQADWIIVLANGAIAEMGSYQEL
LQRKGALMCLLDQARQPGDRGEGETEPTSTKDPRGTSAGRRPELRRERSIKSVPEKDRT
TSEAQTEVPLDDPDRA GWPAGKDSIQYGRVKATVHLAYLRAVGTPCLCLYALFLCQQVA
SFCRGYWL SLWADDP AVGGQQTQAALRGGIFGLLGCLQAIGLFASMAAVLLGGARASRL
FQRLWDVVRSPISFFERTPIGHLLNRFSKETDTVDVDIPDKLRSLLMYA FGLLEVSLVV
AVATPLATVAILPLFLLYAGFQSLYVVSSCQLRRLESASYSSVCSHMAETFQGSTVVRAF
RTQAPFVAQNNARVDESQRISFPRLVADRWLAANVELLGNGLVFAAATCAVLSKAHLSAG
LVGFSVSAALQVTQTLQWVVRNWDLENSIVSVERMQDYAWTPKEAPWRLPTCAAQPPWP
QGGQIEFRDFGLRYP ELPLAVQGVSFKIHAGEKVGIVGRTGAGKSSLASGLLRLQEAAE
GGIWIDGVPIAHVGLHTLRSRISIIPQDPILFPGSLRMNLDLLQEHSDEAIWAALET VQL
KALVASLPGQLQYKCADRGEDLSVGQKQLLCLARALLRKTQILILDEATAAVDPGT ELQM
QAMLGSWFAQCTVLLIAHRLRSVMDCARVLVMDKGQVAESGSPAQLLAQKGLFYRLAQES
GLV

Human MRP6:

Seq Identity

47.64%

Template

The role of MRPs in diseases

6uy0.1. A Multidrug resistance-associated protein 1

Cryo-EM structure of wild-type bovine multidrug resistance protein 1 (MRP1) under active turnover conditions

ATP-binding cassette sub-family C member 10 • Homosapiens (Human) amino acid sequence:

>sp|Q5T3U5|MRP7_HUMANATP-binding cassette sub-family C member 10 OS=Homo sapiens OX=9606 GN=ABCC10 PE=1 SV=1

MERLLAQLCGSSAAWPLPLWEGDTTGHCFTQLVLSALPHALLAVLSACYLGTPRSPDYIL
PCSPGWRLRLAASFLLSVFPLLDLLPVALPPGAGPGPIGLEVLACGVA AVAWISHSLALW
VLAHSPHGH SRGPLALALVALLPAPALVLTVLWHCQRGTLLPPLLPGPMARCLLLILQLA
ALLAYALGWAAPGGPREPWAQEPLLPEDQEPEVAEDGESWLSRFSYAWLAPLLARGACGE
LRQPQDICRLPHRLQPTYLARVFQAHWQEGARLWRALYGAFGRCYLALGLLKLVGTMLGF
SGPLLLSLLVGFLFEEGQEPLSHGLLYALGLAGGAVLGAVLQNYGYEVYKVTLQARGAVL
NILYCKALQLGPSRPPTGEALNLLGTDSERLNLFAGSFHEAWGLPLQLAITLYLLYQQVG
VAFVGGILALLLLVPVNKVIATRIMASNQEMLQHKDARVKLVTELLSGIRVIKFCGWEQA
LGARVEACRARELGRLRVIKYLDAA CVYLWAALPVVISIVIFITYVLMGHQLTATKVFTA
LALVRMLILPLNFPWVINGLLEAKVSLDRIQLFLDLPNHN PQAYSPDPPAEPSTVLEL
HGALFSWDPVGTSLETFISHLEVKKGMLVGIVGKVGCGKSSLLAAIAGELHRLRGHVAVR
GLSKGFGLATQEPWIQFATIRDNILFGKTFDAQLYKEVLEACALNDDLSILPAGDQTEVG
EKGVTLSGGQRARIALARAVYQEKELYLLDDPLAAVDADVANHLLHRCILGMLS YTT RLL
CTHRTEYLERADAVLLMEAGRLIRAGPPSEILPLVQAVPKAWAENGQESDSATAQSVQNP
EKTKEGLEEEQSTSGRLLQEESKKEGAVALHVYQAYWKAVGQGLALAILFSLLLMQATRN
AADWWLSHWISQLKAENSSQEAQPSTSPASMGLFSPQLLL FSPGNLYIPVFPLPKAAPNG
SSDIRFYLT VYATIAGVNSLCTLLRAVLFAAGTLQAAATLHRRL LHRVLMAPVTFFNATP
TGRILNRFSSDVACADDSL PFILNILLANAAGLLGLLAVLGSGLPWLLLLLPPLSIMYYH
VQRHYRASSRELRRLGSLTSLPLYSHLADTLAGLSVLRATGATYRFEEENLRLELNQRC
QFATSATMQWLDIRLQLMGAAV VSAIAGIALVHQQGLANPGLVGLSLSYALSLTGLLSG
LVSSFTQTEAMLVSVERLEEYTCDL PQEPQGQPLQLGTGWLTQGGVEFQDVVLAYRPGLP
NALDGVTFVCVQPGEKLGIVGRTGSGKSSLLLV LFRLLPSSGRVLLDGVDTSQLELAQLR
SQLAIIPQEPFLFSGTVRENLD PQGLHKDRALWQALKQCHLSEVITSMGGLDGELGEGGR
SLSLGQRQLLCLARALLTDAKILCIDEATASVDQKTDQLLQQTICKRFANKTVLTIAHRL
NTILNSDRVLLV LQAGRVVELDSPATLRNQPHSLFQQLLQSSQQGV PASLGGP

The role of MRPs in diseases

Human MRP7:

Seq Identity

33.71%

Template

7mpe.1. A Metal resistance protein YCF1

Cryo-EM structure of the yeast cadmium factor 1 protein (Ycf1p)

ATP-binding cassette sub-family C member 11 • Homo sapiens (Human) aminoacid sequence:

>sp|Q96J66|MRP8_HUMAN ATP-binding cassette sub-family C member 11 OS=Homo sapiens OX=9606
GN=ABCC11 PE=1 SV=1

MTRKRTYWVPNSSGGLVNRGIDIGDDMVSGLIYKTYTLQDGPWSQQERNPEAPGRAAVPP
WGKYDAALRTMIPFRPKPRFPAPQPLDNAGLFSYLTVSWLTPMLIQSLRSRLDENTIPPL
SVHDASDKNVQRLHRLWEEEVSRRGIEKASVLLVMLRFQRTLIFDALLGICFCIASVLG
PILIIPKILEYSEEQLGNVVHGVGLCFALFLSECVKSLSFSSSWIINQRTAIRFRAAVSS
FAFEKLIQFKSVIHITSGEAISFFTGDVNYLFEGVCYGPLVLITCASLVICSISSYFIIG
YTAFIAILCYLLVFPLAVFMTRMAVKAQHHTSEVSDQRIRVTSEVLTCKLIKMYTWEKP
FAKIIEDLRRKERKLEKCGLVQSLTSITLFIPTVATAVWVLIHTSLKCLKLTASMAFSM
LASLNLLRLSVFFVPIAVKGLTNSKSAVMRFKKFFLQESPVFYVQTLQDPSKALVFEEAT
LSWQQTCPGIVNGALELERNGHASEGMTRPRDALGPEEEGNSLGPELHKINLVVSKGMML
GVCNGTSGSKSSLLSAILEEMHLEGSVGVQGSLAYVPQQAWIVSGNIRENILMGGAYDK
ARYLQVLHCCSLNRDLELLPFGDMTEIGERGLNLSGGQKQRISLARAVYSDRQIYLLDDP
LSAVDAHVGKHIFEECIKKTLRGKTVVLVTHQLQYLEFCGQIILLENGKICENGTHSELM
QKKGKYAQLIQKMHKEATSDMLQDTAKIAEKPKVESQALATSLEESLNGNAVPEHQLTQE
EEMEEGSLSWRVYHHYIQAAGGYMVSCIIFFFVVLIVFLTIFSFWWLSYWLEQSGTNSS
RESNGTMADLGNIADNPQLSFYQLVYGLNALLICVGCSSGIFTKVTRKASTALHNKLF
NKVFRCPMSFFDTIPIGRLLNCFAGDLEQLDQLLPIFSEQFLVLSLMVIAVLLIVSVLSP
YILLMGAIIMVICFIYMMFKAIGVFKRLENYSRSPLFSHILNSLQGLSSIHVYGKTED
FISQFKRLTDAQNNYLLLFLSSTRWMALRLEIMTNLVTLAVALFVAFGISSTPYSFKVMA
VNIVLQLASSFQATARIGLETEAQFTAVERILQYMKMCVSEAPLHMEGTSCPQGWPHGE
IIFQDYHMKYRDNTPTVLHGINLTIRGHEVVGIVGRTGSGKSSLGMALFRLVEPMAGRIL
IDGVDCISIGLEDLSKLSVIPQDPVLLSGTIRFNLDPFDRHTDQQIWDALERTFLTAKI
SKFPPKKLHTDVVENGGNFSVGERQLLCIARAVLRNSKIILIDEATASIDMETDTLIQRTI

The role of MRPs in diseases

REAFQGCTVLVIAHRVTTVLNCDHILVMGNGKVVEFDRPEVLRKKPGSLFAALMATATSS

LR

Human MRP8:

Seq Identity

34.41%

Template

7mpe.1. A Metal resistance protein YCF1

Cryo-EM structure of the yeast cadmium factor 1 protein (Ycf1p)

ATP-binding cassette sub-family C member 12 • Homo sapiens (Human) amino acid sequence:

>sp|Q96J65|MRP9_HUMAN ATP-binding cassette sub-family C member 12 OS=Homo sapiens
OX=9606 GN=ABCC12 PE=1 SV=2

MVGEGPYLISDLQGRRRRSFAERYDPSLKTMIIPVRPCARLAPNPVDDAGLLSFATFSWL
TPVMVKGYRQRLTVDLPLSTYDSSDTNAKRFRVLWDEEVARGPEKASLSHVVWKFOR
TRVLMDIVANILCIIMAAIGPVILIHQILQQTERTSGKVWVGIGLCIALFATEFTKVFFW
ALAWAINYRTAIRLKVALSTLVFENLVSFKTLTHISVGEVLNILSSDSYSLFEALFCPL
PATIPILMVFCAAYAFFILGPTALIGISVYVIFIPVQMFMAKLNSAFRRSAILVTDKRVQ
TMNEFLTICIRLIKMYAWEKSFTNTIQDIRRRERKLEKAGFVQSGNSALAPIVSTIAIVL
TLSCHILLRRKLTAPVAFSVIAMFNVMKFSIAILPFSIKAMAEANVSLRRMKKILIDKSP
PSYITQPEDPDTVLLLANATLTWEHEASRKSTPKKLQNQKRHLCKKQRSEAYERSPPAK
GATGP EEGQSDSLKSVLHSISFVVRKGKILGICGNVSGSKSSLLAALLGQMQQLQKGVVAVN
GTLAYVSQQAWIFHGNVRENILFGEKYDHQRYQHTVRVCGLQKDLSNLPYGDLTEIGERG
LNLGGQRQRISLARAVYSRQLYLLDDPLSAVDHVGKHFEECIKKTLRGKTVVLVTH
QLQFLESCDEVILLEDGEICEKGTHKELMEERGRYAKLIHNLRLGLQFKDPEHLYNAAMVE
AFKESPAEREEDAGIIVLAPGNEKDEGKESETGSEFVDTKVPEHQLIQTESPQEGTVTWK
TYHTYIKASGGYLLSLFTVFLFLLMIGSAAFSNWWLGLWLDKGSRMTCGPQGNRTMCEVG
AVLADIGQHVVYQWVYTASMVFLVFGVTKG FVFTKTTLMASSSLHDTVFDKILKSPMSFF
DTTPTGRLMNRFSKDMDEL DVRLPFHAENFLQQFFMVVFILVILAAVFAVLLVVASLAV
GFFILLRIFHRGVQELKKVENVSRSPWFTHITSSMQGLGIIHAYGKKESCITYHLLYFNC
ALRWFALRMDVLMNLTFTVALLVTLFS SISTSSKGLSLSYIIQLSGLLQVCVRTGTET
QAKFTSVELLREYISTCVPECTHPLKVGTCPKDWPSRGEITFRDYQMRYRDNTPLVLDL
NLNIQSGQTVGIVGRTGSGKSSLGMA LFRLVEPASGTIFIDEVDICILSLEDLRTKLTVI

The role of MRPs in diseases

PQDPVLFVGTVRYNLDPFESHTDEMLWQVLERTFMRDTIMKLPEKLQAEVTENGENFSVG

ERQLLCVARALLRNSKIILLDEATASMSKDTLQNTIKDAFKGCTVLTIAHRLNTVLN

CDHVLVMENGKVIEFDKPEVLAIEKPDSEAFAMLLAAEVRL

Human MRP9:

Seq Identity

38.04%

Template

5uj9.1. A bovine multidrug resistance protein 1 (MRP1), Multidrug resistance-associated protein 1

Cryo-EM structure of bovine multidrug resistance protein 1 (MRP1)