Original Article Development and validation of prognostic m6A-related IncRNA and mRNA model in thyroid cancer

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Received April 19, 2022; Accepted June 25, 2022; Epub July 15, 2022; Published July 30, 2022

Abstract: Although N6-methyladenosine (m6A) regulators and IncRNAs influence the carcinogenesis of thyroid cancer (THCA), the association between m6A-related IncRNAs and THCA remains unexplored. Therefore, we have developed and validated a prognostic model based on m6A-related IncRNAs and mRNAs in THCA. Data from the Cancer Genome Atlas were used to analyze the expression and prognostic characteristics of m6A-related IncRNAs and mRNAs in THCA. Univariate Cox regression analysis was used to screen out independent prognostic factors, while Lasso Cox regression was performed to construct m6A-related IncRNA and mRNA models. The correlation between the prognostic models and gene mutation, immune cell infiltration, tumor microenvironment score, tumor mutational burden, and microsatellite instability were assessed. The prognostic models showed excellent accuracy in predicting the prognosis of patients with THCA. Our study established an m6A-related nomogram capable of predicting the prognosis of patients with THCA. In addition, the hub IncRNAs and mRNAs provide insight into improving the prognosis of THCA. These findings can improve our understanding of m6A modifications in THCA and the prognosis and treatment strategies of THCA.

Keywords: N6-methyladenosine, endocrine, cancer, long non-coding RNA, tumor, nomogram

Introduction

The incidence of thyroid cancer, the most common type of endocrine cancer, has significantly increased over the past 10 years, thereby posing a significant public health threat [1-3]. Miller [4] has studied the incidence of tumors in adolescents and young adults in the United States and found that overall cancer incidence in that population has increased during 2007-2016, largely attributed to thyroid cancer, which rose by approximately 3% annually among those aged 20-39 years and 4% among those aged 15-19 years.

Thyroid cancer can be divided into different pathological types [5, 6], the most common which is papillary thyroid carcinoma derived from thyroid follicular epithelial cells and accounts for 80% of all reported cases. Meanwhile, the other pathological types include follicular thyroid cancer, poorly differentiated thyroid cancer, and undifferentiated thyroid cancer [6]. Although thyroid cancer generally exhibits indolent behavior and has favorable prognosis, approximately 20-30% of all patients with THCA experience recurrence or distant metastasis following the initial treatment [5], some patients do not respond well to conventional treatment, leading to poor prognosis [5].

N6-methyladenosine (m6A) modification is the most common post-transcriptional modification of messenger RNA (mRNA) and non-coding RNA that plays a vital role in RNA maturation, stability, translation, export, and decay [7, 8]. To date, m6A modifications have been identified in more than 7,600 genes and 300 non-coding RNAs in mammals [7]. As a reversible dynamic RNA epigenetic process, the molecular components of m6A RNA methylation involve intracellular methyltransferase, demethylase, and signal trans-

ducers [9], which regulate gene expression and interact with various biological functions, such as RNA splicing, export, stability, translation, and ncRNA biogenesis [10]. Evidence has shown that m6A affects several important life processes including cell division and proliferation, focal death, and apoptosis. Abnormal m6A modification plays a key role in the occurrence and development of a variety of tumors including THCA [8]. Song et al. [11] demonstrated that β -catenin represses miR455-3p to stimulate m6A modification of HSF1 mRNA and promote its translation in colorectal cancer. However, data on the prognostic value of m6Arelated genes in THCA remain limited.

Long non-coding RNA (IncRNA) is a non-proteincoding RNA transcript of more than 200 nucleotides [12, 13]; with the development of highthroughput sequencing technology, more than 20,000 IncRNAs have been studied [12]. Long non-coding RNAs are involved in various processes such as transcription, splicing, and translation and have a significant influence in various human diseases, including cancer [14-17]. They are abnormally expressed in various types of tumors and play a key regulatory role [17], and play a significant role in various biological events such as cell cycle, differentiation, proliferation, apoptosis, metastasis, and invasion of various tumor cells [15]. Previous studies have explored the IncRNA expression profile of THCA and demonstrated their potential use as biomarkers for the prognosis and diagnosis of THCA [18-20]. Notably, IncRNA modified by m6A can affect the stability of the transcript and gene expression, resulting in abnormal regulation, which in turn affects the occurrence and development of tumors [21, 22]. Yang et al. [23] have reported that METTL14 suppresses proliferation and metastasis of colorectal cancer by downregulating oncogenic IncRNA Xinactive specific transcript. However, the effect of m6A-modified IncRNA on the prognosis of THCA remains poorly understood.

In this study, we explore the correlation between m6A-related IncRNA and mRNA and the clinicopathological characteristics of THCA. A prognostic model of m6A-related IncRNA and m6A-related mRNA was constructed. The independent prognostic factors obtained via multivariate Cox analysis were integrated into a nomogram to determine the probability of predicting the 1-, 3-, and 5-year progression-free survival in patients with thyroid cancer. The calibration curve was used to evaluate the predictive effect of the nomogram. Furthermore, we assessed the correlation between the prognostic models of m6A-IncRNA and m6A-mRNA with gene mutations, tumor microenvironment, and clinicopathological characteristics. The results of this study provides new insights into the potential mechanism underlying THCA development and facilitate clinical prognosis prediction.

Methods

Data sources and preprocessing

In this study, 23 m6A methylation-related regulators were obtained from previously published studies [24-27]. First, the Cancer Genome Atlas (TCGA) somatic mutation data were downloaded using the TCGAbiolinks R package [28] and visualized using the maftoolR package. The count sequencing data of tumor tissues from 502 patients with THCA and 58 matched control tissues were downloaded from the Genomic Data Commons Data Portal (https://portal. gdc.cancer.gov/). The corresponding clinical data were obtained from the University of California Santa Cruz (UCSC) Xena platform (http://genome.ucsc.edu). By integrating clinical data with sequencing data and excluding samples with progression-free-survival of shorter than one month, we included a total of 497 patients with THCA. The caret (classification and regression training) package in R was used to divide the datasets of 497 patients with THCA into training set and validation set at a 6:4 ratio for subsequent model construction. To ensure the comparability of gene expression between different samples, RNA sequencing data were standardized to TPM (transcripts per kilobase million) format data. Based on the annotation data of the human genome, the genes of the sequencing data were divided into protein-coding and IncRNAs genes. The Hmisc package in R was used to analyze the correlation between the m6A gene and IncRNA and other mRNA molecules. The absolute value of the correlation coefficient > 0.8 (0.6) and P value < 0.05 were used as the screening criteria for m6A-related mRNAs (m6A-related Inc-RNAs). The TPM datasets of TCGA and GTEx were downloaded from the UCSC Xena platform for subsequent pan-cancer analysis of gene expression. Finally, the merged dataset contained a total of 33 types of tumors and 31 types of normal tissue sequencing expression profiles.

Construction and validation of prognostic model based on m6A-related IncRNAs and m6A regulators

In this study, data from 298 patients with THCA were used as the internal training set for constructing the prognostic risk model. Univariate Cox regression analysis was used to identify m6A-related IncRNA and mRNA closely related to the prognosis of progression-free survival (P value < 0.05 was used as the screening index). Then, the glmnet package in R was used to perform Lasso Cox regression analysis to identify the genes associated with progression-free survival in patients with THCA [29]. To determine the appropriate lambda value and lasso (least absolute shrinkage and selection operator) correlation coefficient, 1,000 repetitions of 10-fold cross-validation lasso regression analysis were performed on the training set to determine the minimum lambda value as the criterion for screening the most suitable gene set. The risk score of each THCA patient is based on the following algorithm:

$$Risk \ Score = \sum_{i}^{n} (Exp_i Coef_i) \tag{1}$$

where Exp, is the expression value of each IncRNA or mRNA, and Coef, represents the regression coefficient of the corresponding IncRNA or mRNAs. Based on the correlation between the risk score in the training set and the patient's progression-free survival, we used the Sur_cutpoint function in the Survminer package to calculate the best cut-off that divides the population into high and low risk groups [30]. The minimum number of samples not below 40% of the total number of samples in the dataset was set. The Kaplan-Meier survival curve combined with the log-rank test in the survival package in R was used to distinguish the prognostic difference between high and low risk groups. In the training set, validation set, and the entire dataset, the prediction accuracy of the IncRNA or mRNA prognostic risk model was evaluated using the receiver operating characteristic (ROC) curve.

Interaction network construction

The interaction between m6A-related mRNAs and m6A molecules was visualized using the

Cytoscape software (version 3.7.2). The CytoHubba plug-in was used to calculate the maximum correlation standard value of the nodes in the interaction network [31]. The top 25 hub genes in the network were obtained, and the Pearson's correlation coefficient between m6A-mRNA and m6A-IncRNA molecules was calculated. The absolute value of the correlation coefficient > 0.85 and *P* value < 0.05 were used as the screening criteria for the correlation between mRNA and IncRNA. Cytoscape software was then used to visualize the mRNA-IncRNA interaction.

Gene enrichment analysis, gene set enrichment analysis, and gene set variation analysis

To evaluate the biological processes associated with m6A-related mRNA molecules and mRNAs in the IncRNA-mRNA network, we used the clusterProfiler package to perform gene ontology (GO) or Kyoto Encyclopedia of Genes and Genomes (KEGG) enrichment analysis [32]. The entry screening criteria were AdjP.value < 0.05 and q.value < 0.05, with the Benjamini-Hochberg used as a P value correction method. The clusterProfiler package was also used to perform gene set enrichment analysis on the high- and low-risk subgroups of the mRNA prognostic model in the entire TCGA dataset to identify the enrichment pathways closely associated with the survival of the two risk groups. The parameters used in the gene set enrichment analysis were as follows: the seed was 2,020, the number of calculations was 1,000, the number of genes contained in each gene set was at least 10, and the number of genes contained at most was 500; the Benjamini-Hochberg used as a P value correction method. The reference gene set was derived from the c2.cp.v7.2.symbols.gmt file, while the screening criteria for significant enrichment were set to P < 0.05 with a false discovery rate of less than 0.20. The gene set variation analysis (GSVA) was performed on the molecular expression profile of m6A-related mRNAs through the GSVA package in R [33]. First, the enriched pathways were quantitatively scored, and the difference between the tumor and normal tissues was analyzed. Furthermore, pathways significantly enriched between the two groups were obtained. The selection conditions of the enrichment pathway were P value < 0.05 and |logFC| > 1.5. The reference enriched

gene set was obtained from the c2.cp. v7.4.symbols.gmt file.

Identification of independent prognostic factors

To identify the prognostic factors independently related to progression-free survival, we included the mRNA risk score, age, gender, and tumor grade to perform univariate and multivariate Cox regression analyses. P < 0.1 was used as the screening condition for independent prognostic factors.

Construction and validation of the novel prognostic nomogram

The independent prognostic factors obtained using multivariate Cox analysis were integrated into a nomogram to determine the probability of predicting the 1-, 3-, and 5-year progressionfree survival of patients with THCA. The calibration curve was used to evaluate the prediction effect of the nomogram. To confirm reliability, decision curve analysis was used to compare nomograms with tumor grades and risk scores.

Assessment of tumor microenvironment

In this study, the ESTIMATE algorithm in the estimate package in R was used to calculate the immune and interstitial scores of patients with thyroid cancer [34]. The CIBERSORT algorithm was used to estimate the percentage of 22 immune cell subtypes in each tumor sample [35]. The number of replacements in the algorithm was 100, while no interquartile standardization was performed. The difference in the immune infiltration between the high-risk and low-risk groups was estimated using this result.

Statistical analysis

All analyses were performed using R statistical software (Version 4.0.5). Continuous variables were expressed as mean \pm standard deviation. The Wilcoxon rank sum test was used to compare the two groups. The Kruskal-Wallis test method was used to compare three or more groups. The comparison of progression-free survival between the two groups was performed via Kaplan-Meier analysis combined with the log-rank test. P < 0.05 indicated significant difference.

Results

Expression profile of m6A RNA methylation regulatory gene in THCA

m6A regulatory genes play an important role in the progression of malignant tumors: hence. we explored the differences in the expression of 23 m6A methylation-related genes between THCA tumor tissues and normal tissues. As shown in Figure 1A, 1B, the expression of m6A methylation genes was significantly different between tumor and normal tissues, with 19 m6A regulators differentially expressed between tumor and normal tissues. Except for ELAVL1 and IGF2BP2 genes, which are highly expressed in tumor tissues, the remaining 17 m6A genes were significantly low in tumor tissues, including all Erasers (ALKBH5 and FTO), 7 Writers (CBLL1, KIAA1429, METL14, METL3, RBM15, WTAP and ZC3H13), and 8 Readers (HNRNPA2B1, IGF2BP1, IGF2BP3, LRPPRC, YTHDC1, YTHDC2, YTHDF1, YTHDF3). These findings indicate that m6A RNA methylation regulators play an important role in THCA.

Identification of m6A-related IncRNAs and mRNAs in THCA

To identify potential m6A-related IncRNA and mRNA molecules, we calculated Pearson's correlation coefficient between m6A methylation regulatory genes and the expression of IncRNA and mRNA molecules. In this study, based on the screening criteria of the absolute value of the correlation coefficient > 0.6 and P value < 0.05, 2,253 pairs of m6A regulatory gene-IncRNA correlations were obtained, including 771 IncRNAs. In addition, based on the screening criteria of the absolute value of the correlation coefficient > 0.8 and the *P* value < 0.05, a total of 5,531 m6A regulator-mRNA correlation pairs, including 1,898 mRNAs, was obtained. Figure 1C illustrates the interaction network of m6A regulators and related mRNAs. The red nodes are m6A RNA methylation regulatory genes, while the blue nodes are m6A-related mRNAs. The top 25 hub genes of the maximum correlation standard ranking were obtained from the interaction network diagram through Cytohubba plug-in of Cytoscape, including 8 non-m6A genes (Figure 1D). Figure S1A shows a heat map of the differential expression of eight genes in tumor and normal tissues. We



Figure 1. Expression of m6A regulatory genes between the thyroid cancer group and adjacent tissues, and the construction of the molecular interaction network between m6A gene and its related mRNAs. A. Heat map of the expression of 23 m6A regulatory genes in thyroid cancer and adjacent tissues; B. Expression of 23 m6A regulatory genes between thyroid cancer and adjacent tissues; C. Interaction network between m6A regulatory genes and their related mRNAs; D. 25 hub genes in the interaction network between m6A regulatory genes and their related mRNAs; red nodes represent m6A regulatory genes, and blue nodes represent m6A-mRNAs molecules.

| Characteristics | Train (n = 298) | Test (n = 199) | P-value |
|--|-----------------|----------------|---------|
| Age (mean (sd)) | 45.84±15.46 | 49.54±16.16 | 0.12 |
| Gender (%) | | | 0.77 |
| Female | 219 (73.49) | 143 (71.86) | |
| Male | 79 (26.51) | 56 (28.14) | |
| T_category (%) | | | 0.80 |
| T1 | 101 (33.89) | 69 (34.67) | |
| T2 | 96 (32.21) | 67 (33.67) | |
| T3 | 84 (28.19) | 56 (28.14) | |
| Τ4 | 16 (5.37) | 6 (3.02) | |
| ТХ | 1 (0.34) | 1 (0.5) | |
| N_category (%) | | | 0.66 |
| NO | 127 (42.62) | 93 (46.73) | |
| N1 | 141 (47.32) | 87 (43.72) | |
| NX | 30 (10.07) | 19 (9.55) | |
| M_category (%) | | | 0.92 |
| MO | 5 (1.68) | 4 (2.01) | |
| M1 | 126 (42.28) | 81 (40.7) | |
| MX | 167 (56.04) | 114 (57.29) | |
| UICC_stage (%) | | | 0.35 |
| I | 177 (59.4) | 103 (51.76) | |
| II | 31 (10.4) | 21 (10.55) | |
| III | 61 (20.47) | 50 (25.13) | |
| IV | 29 (9.73) | 25 (12.56) | |
| Papillary Carcinoma Type (%) | | | 0.12 |
| Classical/usual | 5 (1.68) | 4 (2.01) | |
| Follicular (\geq 99 follicular patterned) | 219 (73.49) | 133 (66.83) | |
| Tall Cell (\geq 50 tall cell features) | 59 (19.8) | 41 (20.6) | |
| Other, specify | 15 (5.03) | 21 (10.55) | |

 Table 1. Patient characteristics of LncRNA prognostic model in the TCGA training and validation datasets

further compared the expression levels of these 8 genes in 33 tumor tissues and their corresponding normal tissues and showed that the expression of eight genes was different between the tumor and normal tissues (Figure S1B-I).

Construction and validation of prognostic model based on m6A-related IncRNAs

The complete prognostic data from 497 patients with THCA were divided into training (n = 298) and validation sets (n = 199) (**Table 1**) at a 6:4 ratio. Using single-factor Cox regression and lasso Cox regression analysis methods, a prognostic model of 20 IncRNA molecules was constructed on the training set (**Figure 2A, 2B**). For each patient in the training set, validation set, and the entire dataset, the risk score was calculated using equation 2: AC000123.4* 0.48053 + LINC00271* - 0.39758 + LINC01012*0.23698 + ARHGAP5-AS1* -0.44110 + CTD-2541M15.1*0.14856 + CTD-3157E16.2* - 0.63425 + PRKAR2A - AS1* 0.60911 + PSMG3 - AS1*0.28977 + RP1 -12G14.7* - 0.07105 + RP1 - 153G14.4* -0.48361 + RP11 - 196G18.22*0.41636 + RP11 - 216B9.6* - 0.88477 + RP11 -240G22.5*2.63020 + RP11 - 313D6.3* 0.14101 + RP11 - 337C18.9*0.10700 + RP11 - 359B12.2*1.57736 + RP11-680F8.3* -0.51279 + RP4 - 568C11.4* - 0.57081 + SSSCA1 - AS1*0.81404 + XXbac -B444P24.13* - 1.26468 [2].

Using the survival data of the training set and the surv_cutpoint function of the survminer package, -5.935 was set as the cut-off value to distinguish between the high-risk group (n = 132) and the low-risk group (n = 166). This cut-



Figure 2. Construction and verification of m6A-IncRNA prognostic model. A. Lasso regression coefficients of 81 genes in the THCA training set; B. In lasso regression, based on the minimum lambda value, the most suitable gene set for constructing the model is screened; C-E. Kaplan-Meier survival analysis of the m6A-IncRNA prognostic model in the training set, validation set, and the entire dataset; F-H. ROC curve of m6A-IncRNA prognostic model in training set, validation set, and entire dataset; I-L. Correlation analysis between m6A-IncRNA prognostic model and tumor grade, age, sex and pathological categories.

off value was also used as the distinguishing value between the high and low risk groups of

the validation set and the entire dataset. According to the Kaplan-Meier analysis (**Figure**

2C-E), the prognosis of the low-risk group was significantly better than that of the high-risk group (P < 0.05). To further evaluate the prognosis accuracy of the IncRNA model, the area under the ROC curve (AUC value) in the training. validation, and the entire dataset was calculated for 1-, 3- and 5-year progression-free survival predictions, which resulted in 0.99, 0.58, 0.63; 0.94, 0.66, 0.74; 0.94, 0.74, 0.82, respectively (Figure 2F-H). Therefore, the IncRNA model accurately predicted the prognosis of THCA patients. The correlations between risk score and tumor grade, age, gender, and papillary tumor subtypes of thyroid cancer were displayed as box plots to evaluate the relationship between IncRNA models and clinical indicators. The results showed no significant correlation between the risk score and these indicators (P > 0.05) (Figure 2I-L).

Correlation analysis of molecules in IncRNA model and enrichment analysis of m6A-related mRNAs

A significant positive correlation among 20 IncRNAs was observed in this study (Figure **3A**). Potential signal pathways or functions related to m6A-mRNAs were enriched with GO and KEGG through the clusterProfiler package. The m6A-related mRNA molecules were mainly enriched in RNA splicing and covalent chromatin modification in biological processes. In terms of cell components, they were mainly enriched in the nuclear speck and chromatin; in terms of molecular functions, they were enriched in ubiquitin-like protein transferase activity and ubiquitin-protein transferase activity; in terms of molecular pathways, they were enriched in spliceosome and ubiguitin mediated proteolysis (Figure 3B, 3C; Table 3). The GSVA algorithm was used to perform functional annotations on m6A-related mRNAs, which showed significantly different enrichment pathways between the groups. Tumor samples were enriched in the olfactory signaling pathway and NOS1 pathway, whereas normal samples were enriched in sialic acid metabolism and response to metal ion pathways (Figure 3D).

Interaction network between m6A-related IncRNAs and mRNAs

The Pearson's correlation coefficient was used to evaluate the correlation between the expression of IncRNAs and mRNAs. The absolute value of the correlation coefficient > 0.85 and P value < 0.05 were considered the screening criteria for the correlation between molecules. A total of 595 pairs of IncRNA-mRNA interactions were obtained, including 37 IncRNAs and 440 mRNAs. To further explore the function of IncRNAs in this network, we conducted GO enrichment analysis on related mRNAs and showed that the biological process is mainly enriched in ubiqutin-like protein transferase activity and histone binding; in cell components, it is enriched in nuclear speck and spindle; in molecular function, it is enriched in RNA splicing (**Figure 4; Table 4**).

Construction and validation of prognostic model based on m6A-related mRNAs

The complete prognostic data from 497 patients with THCA were divided to training set (n = 298) and validation set (n = 199) (**Table 2**) at a 6:4 ratio. Using single factor Cox regression and lasso Cox regression analyses, a prognostic model of 12 mRNA molecules was constructed on the training set (**Figure 5A, 5B**). For each patient in the training set, validation set, and entire dataset, the risk score was calculated using the following risk formula:

BRD8*1.77078 + CCDC82* - 0.65499 + EPG5*0.89343 + KHNYN*0.19688 + LCMT2* - 0.15309 + MFAP1* - 0.00118 + SNIP1* -0.78969 + SRRM2*0.17337 + STX16*0.00351 + TAF3* - 0.41327 + ZBTB49* - 0.94961 + ZFAND4* - 0.44280.

Using the survival data of the training set and the surv_cutpoint function of the survminer package, 2.7461 was used as the cut-off value to distinguish between the high-risk (n = 172) and low-risk groups (n = 126). This cut-off value was also used to distinguish between the high and low risk groups of the validation set and the entire dataset (Figure 5D-F). The results of the Kaplan-Meier curve analysis showed that the prognosis of the low-risk group was significantly better than that of the high-risk group (P < 0.05). We also calculated the AUC value of the training, validation, and the entire dataset for the 1-, 3- and 5-year progression-free survival prediction, which showed 0.84, 0.69, 0.74; 0.77, 0.69, 0.72; 0.78, 0.83, 0.75, respectively (Figure 5G-L). The mRNA model accurately predicted the prognosis of patients with THCA. The correlation between the mRNA prognostic model and clinical characteristics was evaluated using a box plot to show the difference in the



Figure 3. Functional enrichment analysis of m6A-related mRNAs. A. Correlation between m6A-IncRNA prognostic model molecules; B. GO enrichment analysis results of m6A-related mRNAs (m6A-mRNAs); BP, Biological Process; CC, Cellular Components; MF, Molecular Function; C. KEGG enrichment analysis of m6A-related mRNAs, the red dots represent related genes in the KEGG enrichment pathway; D. GSVA analysis of m6A-related mRNAs.

| Characteristics | Train (n = 298) | Test (n = 199) | P-value |
|--|-----------------|----------------|---------|
| Age (mean (sd)) | 48.08±16.1 | 46.19±15.38 | 0.30 |
| Gender (%) | | | 1.00 |
| Female | 81 (27.18) | 54 (27.14) | |
| Male | 217 (72.82) | 145 (72.86) | |
| T_category (%) | | | 0.19 |
| T1 | 108 (36.24) | 62 (31.16) | |
| T2 | 88 (29.53) | 75 (37.69) | |
| ТЗ | 84 (28.19) | 56 (28.14) | |
| Τ4 | 17 (5.7) | 5 (2.51) | |
| ТХ | 1 (0.34) | 1 (0.5) | |
| N_category (%) | | | 0.65 |
| NO | 32 (10.74) | 17 (8.54) | |
| N1 | 133 (44.63) | 95 (47.74) | |
| NX | 133 (44.63) | 87 (43.72) | |
| M_category (%) | | | 0.26 |
| MO | 161 (54.03) | 120 (60.3) | |
| M1 | 130 (43.62) | 77 (38.69) | |
| MX | 7 (2.35) | 2 (1.01) | |
| UICC_stage (%) | | | 0.45 |
| I | 162 (54.36) | 118 (59.3) | |
| 11 | 71 (23.83) | 40 (20.1) | |
| 111 | 29 (9.73) | 23 (11.56) | |
| IV | 36 (12.08) | 18 (9.05) | |
| Papillary Carcinoma Type (%) | | | 0.10 |
| Classical/usual | 6 (2.01) | 3 (1.51) | |
| Follicular (\geq 99 follicular patterned) | 221 (74.16) | 131 (65.83) | |
| Tall Cell (≥ 50 tall cell features) | 49 (16.44) | 51 (25.63) | |
| Other, specify | 22 (7.38) | 14 (7.04) | |

Table 2. Patient characteristics of mRNA prognostic model in the TCGA training and validation datasets

risk scores between different tumor grades, age, and gender. The risk score of the highgrade tumors was significantly higher than that of low-grade tumors, although no significant correlation between the risk score and age and gender was observed (P > 0.05) (Figure 5J-L).

Biological characteristics of prognostic models

The correlation between the prognostic model of m6A-IncRNA and m6A-mRNA and the tumor microenvironment and clinicopathological characteristics was explored. The entire TCGA dataset was compared using the risk model scores; the distinguishing value of high and low risk scores was the same as that reported in section 3.6. **Figure 6A-D** shows the overall landscape of gene mutations in the high and low mRNA (IncRNA) risk scores; the rate of BRAF mutations in the high-risk group was significantly higher (lower) than that in the low-risk group.

Immune infiltration plays an important role in tumor formation and progression, and successful clinical applications of immunosuppressants also highlight the role of intratumor immune infiltration in tumor treatment. There were significant differences in CD8+ T cells, regulatory T cells (Tregs), γδ-T cells, MO macrophages, and activated dendritic cells in the high- and lowrisk groups of the mRNA model (P < 0.05) (Figure 6E). Furthermore, the high- and low-risk populations of the IncRNA model showed significant differences in Naïve CD4+ T cells, follicular T helper

cells, $\gamma \delta$ -T cells, monocytes, activated mast cells, and eosinophils (P < 0.05) (**Figure 6F**). The tumor microenvironment score can also reflect the degree of infiltration of immune cells and interstitial cells in tumor tissues. The results showed that the mesenchymal score of the high-mRNA risk group was significantly lower than that of the low-risk group, whereas no difference in the immune score between the two groups was observed (**Figure 6G**). This shows that the intratumoral stroma content of the tumor tissues of the high-mRNAs risk score group is significantly lower than that of the lowrisk group. However, no significant correlation between the IncRNA risk score and the intersti-

| GO Category | Term | Description | Count | Adjust P |
|-------------|------------|--|-------|----------|
| BP | GO:0008380 | RNA splicing | 146 | 7.46E-39 |
| BP | GO:0016569 | covalent chromatin modification | 135 | 8.01E-32 |
| BP | GO:0016570 | histone modification | 132 | 5.21E-32 |
| BP | GO:0000375 | RNA splicing, via transesterification reactions | 122 | 1.81E-33 |
| BP | G0:0000377 | RNA splicing, via transesterification reactions with bulged adenosine as nucleophile | 121 | 1.86E-33 |
| BP | GO:0000398 | mRNA splicing, via spliceosome | 121 | 1.86E-33 |
| BP | GO:0018205 | peptidyl-lysine modification | 108 | 1.36E-23 |
| BP | GO:0051169 | nuclear transport | 106 | 1.56E-27 |
| BP | GO:0006913 | nucleocytoplasmic transport | 105 | 2.47E-27 |
| BP | GO:0051052 | regulation of DNA metabolic process | 99 | 3.42E-16 |
| MF | GO:0019787 | ubiquitin-like protein transferase activity | 99 | 1.48E-15 |
| MF | GO:0004842 | ubiquitin-protein transferase activity | 94 | 3.17E-15 |
| MF | GO:0140098 | catalytic activity, acting on RNA | 79 | 8.27E-09 |
| MF | GO:0003713 | transcription coactivator activity | 69 | 9.39E-09 |
| MF | GO:0042393 | histone binding | 68 | 1.18E-18 |
| MF | GO:0017016 | Ras GTPase binding | 65 | 0.00229 |
| MF | G0:0004674 | protein serine/threonine kinase activity | 65 | 0.00388 |
| MF | GO:0031267 | small GTPase binding | 65 | 0.00471 |
| MF | GO:0016887 | ATPase activity | 61 | 0.00111 |
| MF | G0:0004386 | helicase activity | 53 | 1.27E-1 |
| CC | GO:0016607 | nuclear speck | 116 | 1.24E-29 |
| CC | GO:0000790 | nuclear chromatin | 91 | 4.78E-1 |
| CC | GO:0005635 | nuclear envelope | 86 | 8.49E-10 |
| CC | G0:0098687 | chromosomal region | 81 | 2.72E-14 |
| CC | GO:0005681 | spliceosomal complex | 67 | 1.46E-22 |
| CC | GO:0005819 | spindle | 67 | 1.77E-08 |
| CC | GO:0000151 | ubiquitin ligase complex | 59 | 7.90E-09 |
| СС | GO:0031965 | nuclear membrane | 59 | 4.92E-08 |
| CC | GO:0035770 | ribonucleoprotein granule | 58 | 1.79E-1 |
| CC | G0:0036464 | cytoplasmic ribonucleoprotein granule | 53 | 9.51E-1 |
| KEGG | hsa05168 | Herpes simplex virus 1 infection | 117 | 5.36E-22 |
| KEGG | hsa03040 | Spliceosome | 50 | 3.51E-1 |
| KEGG | hsa03013 | RNA transport | 43 | 1.37E-07 |
| KEGG | hsa05132 | Salmonella infection | 38 | 0.01660 |
| KEGG | hsa04120 | Ubiquitin mediated proteolysis | 37 | 4.37E-08 |

Table 3. GO and KEGG enrichment analysis of m6A-related mRNAs

tial score and immune score of the tumor microenvironment was observed (P > 0.05) (**Figure 6J**). Both the mutational burden and instability of microsatellites in tumors were closely associated with the infiltration of immune cells in tumor tissues. Several studies have shown that the higher the mutation load in tumor tissue and the more unstable the microsatellite instability, the stronger the immunoreactivity in tumor tissue, the higher the probability of successful application of immunosuppressive agents. Patients with high mRNA

risk scores have significantly higher tumor mutation burdens (Figure 6H). However, there was no significant difference in tumor mutation burden between the IncRNA high and low risk groups (Figure 6K). The mRNA and IncRNA risk scores were not correlated with microsatellite instability (Figure 6I, 6L).

Identification of independent prognostic factors

In the TCGA training set, single-factor and multifactor Cox regression analyses were used to



Figure 4. Interaction network between m6A-related IncRNA and m6A-related mRNA. A. Interaction network between m6A-related IncRNAs and m6A-related mRNAs; B. GO enrichment results of mRNAs in the interaction network; BP, Biological Process; CC, Cellular Components; MF, Molecular Function.

| GO Category | Term | Description | Count | Adjust P |
|-------------|------------|--|-------|----------|
| BP | GO:0008380 | RNA splicing | 40 | 3.37E-10 |
| BP | GO:0016569 | covalent chromatin modification | 37 | 7.33E-09 |
| BP | GO:0016570 | histone modification | 36 | 7.33E-09 |
| BP | GO:0000377 | RNA splicing, via transesterification reactions with bulged adenosine as nucleophile | 34 | 2.01E-09 |
| BP | GO:0000398 | mRNA splicing, via spliceosome | 34 | 2.01E-09 |
| BP | GO:0000375 | RNA splicing, via transesterification reactions | 34 | 2.01E-09 |
| BP | GO:0051052 | regulation of DNA metabolic process | 29 | 7.03E-06 |
| BP | GO:0007059 | chromosome segregation | 28 | 1.19E-07 |
| BP | GO:0000209 | protein polyubiquitination | 27 | 2.32E-07 |
| MF | GO:0019787 | ubiquitin-like protein transferase activity | 36 | 2.32E-09 |
| MF | G0:0004842 | ubiquitin-protein transferase activity | 33 | 1.75E-08 |
| MF | GO:0004674 | protein serine/threonine kinase activity | 21 | 0.024184 |
| MF | G0:0042393 | histone binding | 20 | 4.95E-06 |
| MF | GO:0061659 | ubiquitin-like protein ligase activity | 20 | 4.76E-05 |
| MF | GO:0003713 | transcription coactivator activity | 20 | 0.002593 |
| MF | GO:0061630 | ubiquitin protein ligase activity | 19 | 8.85E-05 |
| MF | GO:0004386 | helicase activity | 15 | 0.000341 |
| MF | G0:0042623 | ATPase activity, coupled | 15 | 0.038239 |
| MF | GO:0008234 | cysteine-type peptidase activity | 14 | 0.004058 |
| CC | GO:0016607 | nuclear speck | 40 | 2.99E-14 |
| CC | GO:0005819 | spindle | 28 | 1.10E-07 |
| CC | G0:0098687 | chromosomal region | 28 | 1.10E-07 |
| CC | GO:0000790 | nuclear chromatin | 27 | 1.52E-06 |
| CC | GO:0005635 | nuclear envelope | 25 | 0.000289 |
| CC | GO:0000775 | chromosome, centromeric region | 20 | 3.07E-07 |
| CC | GO:0000793 | condensed chromosome | 19 | 9.01E-06 |
| CC | GO:0000151 | ubiquitin ligase complex | 19 | 0.000189 |
| CC | GO:0031965 | nuclear membrane | 19 | 0.000289 |
| CC | GO:0061695 | transferase complex, transferring phosphorus-containing groups | 18 | 0.000189 |

 Table 4. GO enrichment analysis of mRNAs within mRNA-IncRNA interaction network

determine whether the mRNA prognosis model risk score, age, gender, and tumor grade were independent prognostic factors. Both risk score and tumor grade were independent prognostic factors of the training set (**Table 5**) (risk score, HR (hazard ratio): 2.86; 95% CI (confidence interval): 1.97-4.13; P < 0.001; tumor grade, HR: 1.35; 95% CI: 0.98-1.86, P < 0.1).

Construction and validation of prognostic nomogram

The risk score and tumor grade data based on 12 mRNAs were independent prognostic factors for patients in the THCA training set. The nomogram constructed by integrating these two factors was used to predict the 1-, 3-, and 5-year progression-free survival (**Figure 7A**). The correction curve of the training set showed

that the nomogram's prediction of the 1- and 5-year progression-free survival was significantly better than the 3-year progression-free survival (**Figure 7B**). The nomogram showed high prognostic accuracy in the validation set and the entire dataset (**Figure 7C**, **7D**). According to the decision curve analysis curve (<u>Figure S2</u>), the nomogram could provide the highest net benefit compared to the tumor grade and risk score.

Construction and validation of IncRNA combined mRNA molecular model

A total of 497 thyroid cancer patients with complete prognostic data were included in this study, and they were divided into training set (N = 298) and validation set (N = 199) according to the ratio of 6:4 (**Table 2**). A prognostic model



Figure 5. Construction and verification of the prognostic model of m6A-related mRNAs. A. Lasso regression coefficients of 27 genes in the THCA training set; B. In lasso regression, based on the minimum lambda value, the most suitable gene set for constructing the model was screened; C. Gene set enrichment analysis results in the high-risk group of the m6A-related mRNAs prognostic model; D-F. Kaplan-Meier survival analysis of the prognostic model of m6A-related mRNAs in the training set, validation set, and the entire dataset; G-I. The ROC curve of the prognostic model of m6A-related mRNAs in the training set, validation set, and the entire dataset; J-L. Correlation analysis between the prognostic model of m6A-related mRNAs and tumor grade, age, and sex.

containing 12 IncRNA molecules was constructed in the training set using univariate Cox regression and LASSO Cox regression analysis (Figure 8A, 8B). For each patient in the train-



Figure 6. Relationship between m6A-mRNA and m6A-lncRNA prognostic models and gene mutation, tumor microenvironmental immune infiltration, tumor mutation burden (TMB), and microsatellite instability (MSI). A. Panoramic view of mutations in high-risk groups of mRNA risk model; B. Panoramic view of mutations in the low-risk population of the mRNA risk model; C. Panoramic view of mutations in high-risk groups of the lncRNA risk model; D. Panoramic view of mutations in the low-risk population of the lncRNA risk model; E. Correlation between the high and low scores of the mRNA risk model and the infiltration of immune cells in the tumor tissue; F. Correlation between the high and low scores of the lncRNA risk model and the infiltration between the high and low scores of the mRNA risk model and the immune score and interstitial score; H. Correlation between the high and low scores of the mRNA risk model and the incRNA risk model and the incRNA risk model and the immune score and interstitial score; K. Correlation between the high and low scores of the mRNA risk model and the immune score and interstitial score; K. Correlation between the high and low scores of the lncRNA risk model and TMB; L. Correlation between the high and low scores of the lncRNA risk model and TMB; L. Correlation between the high and low scores of the lncRNA risk model and MSI.

| Table 5. Univariate and multivariate Cox regression to identify independent prognostic predictor in the |
|---|
| TCGA training cohort |

| | Progression-free survival in TCGA Training Cohort | | | | |
|-------------------------|---|------------------|-----------------------|------------------|---------|
| Characteristics | Number of Univariate Analysis | | Multivariate Analysis | | |
| | patients | HR (95% CI) | P-Value | HR (95% CI) | P-Value |
| Age (≥ 65 vs < 65) | 298 | 1.61 (0.69-3.75) | 0.266 | | |
| Gender (Male vs Female) | 298 | 1.41 (0.66-2.99) | 0.374 | | |
| Risk.Score | 298 | 3.34 (2.37-4.71) | < 0.001 | 2.86 (1.97-4.13) | < 0.001 |
| UICC stage | 298 | 1.76 (1.3-2.39) | < 0.001 | 1.35 (0.98-1.86) | 0.062 |

| A | Points | 0 10 20 30 40 50 60 70 80 90 100 |
|---|-----------------|---|
| | Risk Score | 0 0.5 1 1.5 2 2.5 3 3.5 4 4.5 5 5.5 6 6.5 |
| | UICC_stage | |
| | Total Points | 0 10 20 30 40 50 60 70 80 90 100 120 |
| | 1-year survival | |

Figure 7. Construction and verification of nomogram. A. Nomogram constructed on the TCGA training set; the 1-4 of UICC_ stage represent Stages I-IV, respectively; B. Nomograms as corrected graphs of 1, 3, and 5 years in the training set; C. Calibration chart of the nomogram for 1, 3, and 5 years in the validation set; D. Nomogram as the calibration chart of 1, 3, and 5 years in the validation chart of 1, 3, and 5 years in the entire dataset.



09

0.7 0.5 0.3 0.1

0.9

0.7 0.5 0.3

ing set, validation set and the entire dataset, the risk score is calculated based on the following risk formula: LENG8-AS1* 0.45213 - LINC00271*0.42282 - LINC01257* 0.59266 + RP1-283E3.8*0.36323 - RP11-141M1.4*0.05587 - RP11-175019.4*0.16684 + RP11-196G18.22*0.46581 - RP11-216-B9.6*0.79439 - RP11-251G23.5*0.50873 + RP11-33B1.4*0.129 - 777023.2*0.02021 -RP5-991G20.1*0.00003. Using the survival data from the training set and the surv_cutpoint function of the "survminer" package, we

3-year survival



Figure 8. Construction and validation of IncRNA combined mRNA molecular model. A. Lasso regression coefficients of 26 genes in the THCA training set; B. Screen out the optimal gene set for building the model based on the minimum lambda value in LASSO regression; C-E. Kalan-Meier survival analysis of m6A-IncRNA combined mRNA prognostic model in training set, validation set and the whole dataset; F-H. Time-ROC curves of m6A-IncRNA combined mRNA prognosis model in training set, validation set and the whole dataset; I-L. Correlation analysis of m6A-IncRNA combined mRNA prognosis model and tumor grade, age, gender and pathological grade.

define -0.6314 as the cutoff value to distinguish between high risk (N = 149) and low risk

(N = 149) groups, which will also serve as validation Discriminatory value between high and

low risk groups for the set and the entire dataset. According to the Kaplan-Meier curve analysis results in Figure 8C-E, the prognosis of the low-risk group was significantly better than that of the high-risk group (P < 0.05). In addition, to further evaluate the accuracy of the IncRNA combined mRNA model for prognosis prediction, we calculated the area under the ROC curve (AUC). The AUCs for 1-year, 3-year and 5-year progression-free survival were 0.80 vs 0.78 vs 0.78; 0.81 vs 0.68 vs 0.76; 0.79 vs 0.88 vs 0.77 in the training set, validation set, and the entire dataset, respectively. It can be seen that the IncRNA combined mRNA model has very good accuracy in predicting the prognosis of thyroid cancer patients (Figure 8F-H). We further evaluated the correlation between the IncRNA combined mRNA model and clinicopathological factors. In this study, the differences in risk scores among different tumor grades, different ages, different genders, and different pathological subtypes were shown in box plots. The results showed that the risk scores of high-grade tumors were significantly higher than those of low-grade tumors, and different histopathological subtypes There was also a significant difference in the risk score between the two groups, but there was no significant correlation between the risk score and age and gender (P > 0.05) (Figure 8I-L).

Discussion

The incident of thyroid cancer has a rapidly increased, thereby posing significant clinical, economic, and psychological burdens [1, 2]. Overall, the prognosis of THCA is relatively good [5]; however, some patients whose tumors are highly aggressive and more likely to recur [6] often undergo a second operation, which may lead to post-surgical complications such as recurrent laryngeal nerve and parathyroid injuries [36-38]. In addition, postoperative patients require thyroid stimulating hormone suppression therapy, which can lead to potential side effects such as osteoporosis and atrial fibrillation [39, 40]. Therefore, it is essential to better identify and evaluate the prognosis of patients with THCA.

Traditional clinicopathological parameters such as tumor-node-metastasis staging can predict the mortality associated with THCA, yet it is difficult to accurately estimate the risk of recurrence [41-43]. The American Thyroid Association recurrence risk stratification can predict the recurrence risk of thyroid cancer, although its accuracy, which does not reflect the biological progress of papillary thyroid carcinoma, requires further improvement [44, 45]. Accurate prediction of the prognosis of patients with THCA will help to better identify patients with high risk of recurrence. In addition, extensive surgery, I¹³¹ treatment, and thyroid stimulating hormone suppression therapy for low-risk patients will be avoided, which will improve the quality of life of patients.

Previous studies have reported that the expression of m6A regulators and IncRNA varies among different types of tumors, thereby playing a key regulatory role [7, 10, 46] and affecting the prognosis of cancer patients. In this study, most m6A RNA methylation regulatory genes underwent expression changes in THCA; therefore, we analyzed the correlation between m6A-related IncRNA and mRNA and the clinicopathological characteristics of THCA, hence constructed a risk model for m6A-related IncRNA and m6A-related mRNA molecules. In addition, these prognostic genes may play an important role in the progression of papillary thyroid carcinoma and represent potential targets for suppressing recurrence and metastasis.

In this study, complete prognostic data from 497 patients with THCA were analyzed using single-factor Cox regression and lasso Cox regression analyses to construct a prognostic model of 20 IncRNAs, among which LINCOO-271 is associated with thyroid cancer [47-50]. Buishand et al. [48] reported that adrenocortical tumors have a distinct, long, non-coding RNA expression profile and LINC00271 is downregulated in malignancy. Ma et al. [50] showed that LINC00271 expression was significantly downregulated in papillary thyroid carcinoma versus adjacent normal tissues (P < 0.001). LINC00271 was also found to be an independent risk factor for extrathyroidal extension, lymph node metastasis, advanced tumor stage III/IV, and recurrence in multivariate analyses [50]. The IncRNA ARHGAP5-AS1 in the prediction model constructed in this study is associated with breast and gastric cancers [51, 52].

The role of the other IncRNAs in tumor development has not been reported. Compared to the previous prognostic models of THCA [53, 54], the prognostic model developed in this study is novel and has potential clinical applications. Our findings will help to identify potential prognostic IncRNAs and mRNAs regulated by m6A, providing new insights into evaluating the risk of THCA patients.

The nomogram of this study combined with m6A-related IncRNA and mRNA prediction models and clinicopathological parameters can provide clinicians with a convenient and accurate method to assess the prognosis of patients with THCA. The graphical scoring system is e patient friendly and helps with treatment planning to achieve personalized.

This study has some limitations. The main source of clinical information for our dataset is the TCGA database where most patients were from either Europe or the United States; thus, caution should be exercised when extending the results to the patients from other regions. The establishment and verification of the nomogram are also based on the TCGA database; therefore, in future studies, external datasets should be used to verify clinical information and gene expression data. Finally, our prognostic model and nomogram must be validated in prospective studies.

In conclusion, we revealed the expression and prognostic value of m6A-related IncRNAs and mRNAs in THCA. Our study established an m6A-related tumor prognosis prediction nomogram, which can reliably predict the prognosis of patients with THCA. These findings will expand our understanding of m6A modification in THCA and provide insights into the prognosis and treatment strategies of THCA.

Acknowledgements

This work was supported by grants from the Science and technology Research Special project of Sichuan provincial Administration of Traditional Chinese Medicine (Nos. 2021ZD011) and Ren-song Yue's TCM studio.

Disclosure of conflict of interest

None.

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m6A-related IncRNA/mRNA thyroid cancer model





Figure S1. Difference in the expression of eight key m6A-mRNA molecules in pan-cancer tumor tissues. A. Heat map display of the expression levels of eight key m6A-mRNA molecules between thyroid cancer and adjacent tissues; B-I. Results of differential expression of HNRNPK, BCLAF1, DDX42, THRAP3, WDR82, ADNP, CDK13, and CUL3 molecules between 33 pan-cancer tumor tissues and corresponding 31 normal tissues; *P < 0.05; **P < 0.01; ***P < 0.001.



2



Figure S2. DCA curve of the nomogram model in the TCGA training set, validation set, and the entire TCGA dataset. A-C. DCA images of 1-, 3-, and 5-year prognostic prediction results in the training set; D-F. DCA images of 1-, 3-, and 5-year prognosis prediction results in the validation set; G-I. DCA images of 1-, 3-, and 5-year prognostic prediction results in the entire dataset.