Brief Communication High PIVKA-II level and ASAP score predict 1-year risk of hepatocellular carcinoma in non-cirrhotic chronic hepatitis B patients

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Received February 28, 2023; Accepted April 21, 2023; Epub June 15, 2023; Published June 30, 2023

Abstract: Protein induced by Vitamin K absence or antagonists-II (PIVKA-II) is a diagnostic marker of hepatocellular carcinoma (HCC). We aimed to investigate the predictive role of PIVKA-II and ASAP score for development of HCC in 1 year among untreated patients of chronic hepatitis B (CHB). We conducted this case-control study to include untreated CHB patients followed at the National Taiwan University Hospital and grouped into HCC and matched non-HCC groups. Their archived serum samples were assayed for PIVKA-II levels 1 year before HCC, at HCC or their last serum sample. A total of 69 HCC cases and 102 non-HCC controls were recruited. Baseline PIVKA-II level was significantly higher in the HCC group than in the control group and it could predict HCC development in 1 year with an area under the receiver operating characteristic curve of 0.76. Multivariable analysis adjusting age, sex, liver function and alpha-fetoprotein level showed that baseline PIVKA-II ≥31 mAU/mL (vs. <31 mAU/mL) increased 12.5-fold risk (95% CI: 4.9-31.7) of HCC in 1 year, and even in patients with normal alpha-fetoprotein levels. The ASAP score, a combination of age, sex, alpha-fetoprotein and PIVKA-II, increases the predictability for HCC in 1 year. We concluded that both high PIVKA-II level and ASAP score may predict HCC development in 1 year in untreated CHB patients, especially in patients with normal alpha-fetoprotein level.

Keywords: Protein induced by vitamin K absence or antagonists-II, Des-gamma-carboxy prothrombin, DCP, liver cancer, HCC, biomarker, tumor marker

Introduction

According to the World Health Organization, HCC is the fifth most common malignancy worldwide and the second most cancer-related death [1]. Chronic Hepatitis B (CHB) is a major cause of HCC and contributes to approximately 50% of HCC [2]. Several potent antiviral therapies have been developed and effectively reduce the risk of HCC development [3]; however, there are still many patients who do not fulfill the criteria to start antiviral therapies or do not have the access to antiviral agents. As prognosis is highly related to the stage of HCC, early detection of HCC is vital to improve the overall survival [4]. HCC surveillance to detect earlystage HCC and to reduce mortality is recommended by current guidelines [5-8]. All current guidelines recommended regular ultrasonographic examination every 6 months in at risk patients [5-9]. Ultrasound alone yielded a sensitivity of 63%, which was not very effective for detecting early-HCC [10]. Alpha-fetoprotein (AFP) had been suggested as a surveillance option combined with ultrasonography by the Asian Pacific Association for the Study of the Liver (APASL), American Association for the Study of Liver Diseases (AASLD), Korean Liver Cancer Association (KLCA) and the Japan Society of Hepatology (JSH) guideline [5-8]; however, AFP was less specific for HCC in case of active hepatitis [11], and also showed poor sensitivity for small HCC [12, 13].

PIVKA-II, also known as Des-gamma-carboxy prothrombin, was first reported in 1984 that elevated in patients with HCC and might serve as a diagnostic tumor marker [14]. PIVKA-II shows greater diagnostic accuracy than AFP in early stage HCC in a systemic review [15]; however, the generalizability and accuracy remained to be validated [15]. PIVKA-II had been used as a complementary marker to AFP to increase the diagnostic rate (such as the GALAD score and ASAP model) [16, 17]. PIVKA-II, AFP and AFP-L3 had been recommended by the JSH consensus for HCC surveillance [6].

The role of PIVKA-II level to predict the development of HCC remains limited. Our previous study targeting on patients of CHB-related cirrhosis on antiviral therapy, we found PIVKA-II and AFP at time of virological suppression may predict the development of subsequent HCC [18]. In this study, we would like to investigate the predictive role of PIVKA-II for HCC in treatment-naive CHB patients.

Materials and methods

Patient enrollment

This was a retrospective case-control study conducted in National Taiwan University Hospital, a tertiary medical center in Taipei, Taiwan. Patients with treatment-naïve chronic hepatitis B (CHB) without cirrhosis previously included in the Elucidation of Risk Factors for Disease Control or Advancement in Taiwanese Hepatitis B Carriers (ERADICATE-B) cohort were screened [19]. HCC surveillance was performed by abdominal ultrasonography and AFP level at least once every six months. HCC was diagnosed by either histology or two typical dynamic images by computed tomography or magnetic resonance imaging, with an AFP level of \geq 200 ng/mL before 2010 and one typical dynamic imaging study after 2010 according to the AASLD guidelines [20]. These patients were categorized into HCC and non-HCC groups.

Because we planned to investigate PIVKA-II level as a predictive marker, we included patients with stored serum in the -20°C freezer at the hepatitis research center of the National

Taiwan University Hospital. For the HCC group, we included patients with stored serum at 1 year before the diagnosis of HCC (as baseline) and at the time of HCC. We screened age and sex matched patients without HCC, and included those with stored serum at least 1 year before (as baseline) their last follow-up in our hospital to confirm their absence of HCC at the end of follow-up. Baseline PIVKA-II levels were measured to predict the development of HCC in 1 year. People with incomplete clinical data, unavailable stored sera, a history of HCC, and patients who received vitamin K or warfarin, which might interfere the levels of PIVKA-II were excluded. The clinical characteristics of patients were collected, including age, sex, platelet, total bilirubin, AST, ALT, HBeAg, HBV DNA, and AFP.

This study was approved by the Institutional Review Board of the National Taiwan University Hospital (200909047R) and conformed to the ethical principles for medical research involving human subjects of the Declaration of Helsinki updated in 2013. All patients provided written informed consents before enrollment.

Measurement of serum PIVKA-II

Serum PIVKA-II concentrations were analyzed by the ARCHITECT i2000SR immunoassay analyzer (Abbott Laboratories, North Chicago, IL) through the manufacturer's instructions. The ARCHITECT PIVKA-II assay is a two-step chemiluminescent microparticle immunoassay for the quantitative measurement of PIVKA-II in human serum or plasma. The PIVKA-II present in the sample binds to the anti-PIVKA-II coated microparticles in the capture step. After washing, the analyte particles were mixed with acridinium-labeled anti-prothrombin conjugate to create a reaction mixture. The resulting chemiluminescent reaction is measured as relative light units (RLUs) and the final results are expressed as arbitrary units (mAU/mL) for the quantitative measurement. The precision of the assay was below 10% CV, with a measuring range from 20 to 30000 mAU/mL [21]. All clinical data, including the presence of HCC, were blinded to laboratory technicians to avoid bias.

Calculation of ASAP score

The ASAP score was calculated as In (p/(1-p)) =In -7.57711770 + 0.04666357 * age -

Parameters	Ν	Non-HCC	HCC	P-value
Ν		102	69	
Age, year		56 (48-63)	56 (49-63)	0.819
sex				
Male		80 (78)	53 (77)	0.803
Female		22 (22)	16 (23)	
HBeAg	35			
Negative		16 (94)	13 (72)	0.086
Positive		1(6)	5 (28)	
HBV DNA, log ₁₀ IU/mL	42	1.3 (1.3-2.9)	4.6 (2.8-5.3)	<.001
AST, U/L	159	23 (19-27)	38 (28-51)	<.001
ALT, U/L	167	23 (16-31)	38 (27-54)	<.001
Platelet, k/uL	76	197 (172-218)	120 (76-176)	0.003
Total bilirubin, mg/dL	89	0.9 (0.7-1.0)	0.8 (0.6-1.0)	0.594
AFP, ng/mL	148	2.7 (2.2-4.4)	11 (5.4-20)	<.001
<20		96 (98)	40 (80)	<.001
≥20		2 (2)	10 (20)	
PIVKA-II, mAU/mL	171	24 (20-30)	36 (26-71)	0.013
<31		84 (82)	24 (35)	<.001
≥31		18 (18)	45 (65)	

Table 1. Baseline (1-year before outcome date) characteristics of enrolled patients

Note: Data are expressed as median (interquartile range) or number (percentage). Abbreviations: AST: aspartate transferase, ALT: alanine aminotransferase, AFP: alpha-fetoprotein, PIVKA-II: Prothrombin induced by Vitamin K Absence of Antagonist-II, HCC: hepatocellular carcinoma.

0.57611693 * sex + 0.42243533 * In (AFP) + 1.10518910 * In (PIVKA-II), which had been used to detect HCC among patients with hepatitis B [17].

Statistical analysis

Continuous data are reported as median (interquartile range), and differences between groups were compared by Student's t-test, or paired t-test as appropriate. Categorical data are reported as numbers (percentages), and differences between groups were evaluated by chi-square test as appropriate.

The receiver operating characteristic (ROC) curve was used to explore the optimal cut-off value (by maximizing the Youden Index) for the PIVKA-II and ASAP score value for predicting occurrence of HCC in 1 year. The univariate and multivariable logistic regression analysis were used to identify predictors for HCC. The statistical analyses were performed by using STATA (version 16; Stata Corp, College Station, TX, USA) and all tests were 2-sided and a *p* value

< 0.05 was considered statistically significant.

Result

Baseline characteristics

Totally, 171 non-cirrhotic, untreated HBV carriers were collected into analysis, including 69 HCC cases and 102 non-HCC controls. The median age was 55 years and 78% were men (Table 1). The baseline median PIVKA-II (36 vs. 24 mAU/ mL, P=0.013), and AFP (11 vs. 2.7 ng/mL, P<0.001) were significantly higher in the HCC group than in the control group. The AST (38 vs. 23 U/L, P<0.001), ALT (38 vs. 23 U/L, P<0.001), and HBVDNA (4.6 vs. 1.3 log₁₀ IU/mL, P<0.001) were also significantly higher in the HCC group compared to the control group. Of these HCC patients, 80% had AFP level <20 ng/mL, while 65% had PIVKA-II ≥31 mAU/mL, which indicates PIVKA-II might serve as a supplementary predictor of HCC for those with normal AFP level.

PIVKA-II predicts HCC development in 1 year

In the HCC group, the median PIVKA-II level increased significantly until the development of HCC (from 36 to 44 mAU/mL, P=0.029), which confirmed that PIVKA-II as the tumor marker of HCC and might be used for HCC surveillance. We further explored whether we can use the baseline PIVKA-II to predict HCC development in 1 year. The receiver operating characteristic (ROC) curve was plotted for PIVKA-II in predicting HCC and the area under the ROC curve (AUROC) was 0.76 (95% confidence interval [CI]: 0.68-0.94) (Figure 1). The optimal cutoff value for PIVKA-II was 31 mAU/mL with a sensitivity of 65% and specificity of 83% to predict HCC development in 1 year (Supplementary Tables 1, 2).

AFP \geq 20 ng/mL or PIVKA-II \geq 31 mAU/mL predict HCC development in 1 year

We further evaluated the predictors of HCC including the PIVKA-II level. Univariate analysis showed that baseline PIVKA-II \geq 31 mAU/mL,



Figure 1. The receiver operating characteristic (ROC) curve of PIVKA-II for predicting HCC in 1 year.

AFP \geq 20 ng/mL, and high ALT level were significantly associated with the development of HCC in 1 year (**Table 2**). Multivariable logistic regression analysis after controlled age, sex, ALT and AFP levels showed baseline PIVKA-II \geq 31 mAU/mL increased 12.5-fold risk of HCC (vs. <31 mAU/mL, OR: 12.5, 95% CI: 4.9-31.7) in 1 year.

PIVKA-II level \geq 31 mAU/mL predicts HCC in patients with normal AFP level <20 ng/mL

In our treatment-naïve CHB cohort, 80% of patients have normal AFP level 1 year before their HCC development. In order to investigate whether PIVKA-II helps to predict HCC in these patients, 136 patients (AFP <20 ng/mL) were used for subgroup analysis, including 40 HCC cases and 96 non-HCC controls. The baseline PIVKA-II level was predictive of 1-year HCC with an AUROC of 0.80 (95% CI: 0.72-0.89) (Figure 2). We further investigate the risk predictor for HCC in this subgroup of patients. Univariate analysis showed that baseline PIVKA-II ≥31 mAU/mL and higher ALT levels were significantly associated with the development of HCC in 1 year (Table 3). Multivariable logistic regression analysis after adjustment of confounding factors showed baseline PIVKA-II ≥31 mAU/mL increased 12.4-fold risk of HCC in 1 year (vs. <31 mAU/mL, OR: 12.4, 95% CI: 4.7-32.7).

High ASAP score predicts HCC in 1 year

Furthermore, we investigated the predictive performance of PIVKA-II in combination with

AFP level and the ASAP score was evaluated. The ASAP scores were significantly higher in the HCC group than in the control group (0.48 vs. 0.25, P<0.001). The ROC curve was plotted for ASAP score in predicting HCC and the area under the ROC curve (AUROC) was 0.90 (95% CI: 0.85-0.95) (Figure 3) and 0.89 (95% CI: 0.83-0.95) in subgroup analysis of AFP level <20 ng/mL (Supplementary Figure 1). The optimal cutoff is 0.31 yielded by Youden Index to predict of 1-year HCC, with a sensitivity of 92.5% and a specificity of 71.7% (Supplementary Tables 3, 4). In our cohort, 60% patients with ASAP score ≥ 0.31

developed HCC in 1 year, compare with 4% in those with ASAP score <0.31 (**Figure 4**). Multivariable logistic regression analysis after adjustment of confounding factors showed baseline ASAP score >0.31 increased 111-fold risk of HCC (OR: 111.16, 95% CI: 21.4-577.4) (**Table 4**).

Discussion

In our study, we found that among untreated non-cirrhotic chronic hepatitis B patients, PIV-KA-II \geq 31 mAU/mL was associated with 12.5-fold risk of developing HCC in 1 year, which could be a potential predictor for future HCC development. Notably, patients with normal AFP level (<20 ng/mL), PIVKA-II \geq 31 mAU/mL were still associated with 12.4-fold increased risk of HCC in 1 year. The combination of PIVKA-II and AFP into the ASAP score can predict subsequent HCC in 1 year with improved accuracy.

Surveillance in high-risk populations is an effective measure for HCC early detection, because we can identify early-stage HCC for curative therapy. Asian chronic non-cirrhotic HBV carriers for females >50 years have a 0.3-0.6% HCC risk per year and for males >40 years have a 0.4-0.6% HCC risk per year [6]. Therefore, a surveillance strategy for HCC is recommended. The efficacy of HCC surveillance every 6 months had been demonstrated in a large prospective randomized controlled trial in China [22]. Current guidelines suggest

Davamatara	Univariate		Multivariable		
Parameters	OR (95% CI)	P value	Adjusted OR (95% CI)	P value	
Age (1 year increase)	1.00 (0.96-1.03)	0.753	1.04 (1.00-1.09)	0.071	
Male, sex	0.91 (0.44-1.09)	0.803	0.67 (0.24-1.86)	0.443	
ALT (1 U/L increase)	1.02 (1.01-1.04)	0.003	1.01 (1.00-1.03)	0.056	
AFP ≥20 vs. AFP <20	12.00 (2.52-57.24)	0.002	21.36 (3.58-127.50)	0.001	
PIVKA-II ≥31 vs. PIVKA-II <31	8.75 (4.30-17.80)	<.001	12.53 (4.95-31.73)	<.001	

Table 2. Univariate and multivariable analysis for the predictors of HCC in 1 year

Abbreviations: ALT: alanine aminotransferase, AFP: alpha-fetoprotein, PIVKA-II: Prothrombin induced by Vitamin K Absence of Antagonist-II, HCC: hepatocellular carcinoma, OR: odds ratio, CI: Confidence interval.



Figure 2. The receiver operating characteristic (ROC) curve of PIVKA-II for predicting HCC in 1 year in patients with AFP <20 ng/mL.

6-month abdominal ultrasonography as the standard protocol for HCC surveillance. However, for patients with obesity, liver atrophy, or marked cirrhosis, it is difficult to find small HCC by ultrasonography. Additional biomarker is therefore clinically needed.

AFP has already been widely used as a biomarker for HCC. Currently, ultrasonography with or without AFP has been recommended by APASL and AASLD for high-risk patients [5, 6]. However, for HCC size \leq 3 cm, serum AFP level was frequently normal [13], which makes it insensitive for early detection of HCC.

PIVKA-II increases in malignant hepatocyte during vitamin K deficiency and supposed to reflect liver function under warfarin treatment first in 1963 [23]. In liver tissue of HCC, impaired vitamin-K dependent carboxylation pathway for prothrombin precursor leads to overproduction of prothrombin precursor with des-γ-carboxy-

lation, and thus increased the serum PIVKA-II level [24]. The PIVKA-II levels significantly increased in HCC patients; therefore, it may serve as a diagnostic marker for HCC [25, 26]. PIVKA-II level is associated with portal vein invasion and advanced tumor stage [27]. PIVKA-II is not correlated with AFP [14], therefore, PIV-KA-II might be able to complement the predictive role of AFP. PIVKA-II together with AFP reflect different mechanisms of hepatocarcinogenesis. The JSH guideline suggested AFP, AFP-L3, and PIVKA-II combine with ultrasound for high-risk group in their regular surveillance [28].

Currently, PIVKA-II has been considered as promising predictive biomarker for HCC in different clinical scenarios, including predicting prognosis of HCC patients [29], at time of anti-HBV therapy induced virological remission in patients of CHB-related cirrhosis [18], and to predict the complete responses of trans-arterial chemoembolization in patients of HCC [30]. Several studies supported that the combination of PIVKA-II and AFP had better diagnostic efficacy in HCC [31]. Most recent studies on PIVKA-II have concentrated on its diagnostic accuracy of HCC or its predictive role in HCC progression [32-34]. Few have focused on the capability of PIVKA-II in predict HCC before clinical diagnosis on a high-risk population. A casecontrol study with 189 CHB patients in 2016 indicated the benefit of combination of AFP and PIVKA-II for diagnosis and early detection of CHB-related HCC [35]. However, the predictive role of PIVKA-II 1 year before HCC had an

Deremetere	Univariate		Multivariable		
Parameters	OR (95% CI)	P value	Adjusted OR (95% CI)	P value	
Age (1 year increase)	1.01 (0.97-1.05)	0.745	1.04 (0.99-1.09)	0.131	
Male, sex	0.89 (0.38-2.11)	0.794	0.61 (0.21-1.79)	0.373	
ALT (1 U/L increase)	1.03 (1.01-1.05)	0.006	1.03 (1.01-1.05)	0.005	
PIVKA-II ≥31 vs. PIVKA-II <31	10.39 (4.43-24.35)	<.001	12.45 (4.74-32.66)	<.001	

Table 3. Univariate and multivariable analysis for predictors of HCC in 1 year in patients with AFP <20 $\,$ ng/mL

Abbreviations: ALT: alanine aminotransferase, PIVKA-II: Prothrombin induced by Vitamin K Absence of Antagonist-II, HCC: hepatocellular carcinoma, OR: odds ratio, CI: Confidence interval.



Figure 3. The receiver operating characteristic (ROC) curve of ASAP score for predicting HCC in 1 year.



Figure 4. Using ASAP score 0.31 for HCC prediction in 1 year.

AUROC of 0.54 only and there were no suggested cut-off PIVKA-II values for HCC prediction. Serial AFP and PIVKA-II levels measured before HCC have the potential for HCC prediction, while PIVKA-II has a better AUROC than AFP before diagnosis [15]. Our study indicated the predictive role of PIVKA-II 1 year before HCC had an AUROC of 0.76 (95% confidence interval [CI]: 0.68-0.94) with an optimal cutoff value for PIVKA-II was 31 mAU/mL.

The GALAD score (gender, age, AFP-L3, AFP, and DCP) had been introduced for detecting HCC in chronic liver disease patients with an AUROC greater than 0.90 irrespective of HCC stages [16], and had been validated in several case-control cohorts [36, 37], however, AFP-L3 is not available in many countries, which limits its application. The simplified ASAP score, has been proposed to discriminate HCC

from chronic HBV, HBV related cirrhosis, benign hepatic tumors, and healthy control participants from 11 hospitals in China [17]. The addition of AFP-L3 did not increase the diagnostic accuracy for HCC in both 2198 patients in the training cohort and 727 patients in the validation cohort. A cut-off value of 0.5256 of the ASAP score had been recommended as a diagnostic marker [17]. In another case-control study included 168 HCV-HCC patients and a control group of 193 HCV-infected patients, the ASAP score had higher AUROC compared to the GALAD score (0.917 vs. 0.894, P=0.057) in detect any stage of HCC, both in the overall and cirrhotic groups [38]. According to the AUROC result, the ASAP score was significantly better than the GALAD score in detection of early-stage (BCLC stage 0-A) HCC (P<0.05), suggesting the potential of ASAP score in early detection of HCC in patients of chronic hepatitis C [38]. The ASAP score had been explored in

Developmenter	Univariate		Multivariable		
Parameters	OR (95% CI)	P value	Adjusted OR (95% CI)	P value	
Age (1 year increase)	1.00 (0.96-1.03)	0.753	0.91 (0.85-0.97)	0.003	
Male, sex	0.91 (0.44-1.09)	0.803	0.36 (0.10-1.34)	0.129	
ALT (1 U/L increase)	1.02 (1.01-1.04)	0.003	1.01 (0.99-1.03)	0.185	
ASAP ≥0.31 vs. ASAP <0.31	33.05 (9.35-116.88)	<.001	111.16 (21.40-577.44)	<.001	

Table 4. Univariate and multivariable analysis for the predictors of HCC in 1 year

Abbreviations: ALT: alanine aminotransferase, HCC: hepatocellular carcinoma, OR: odds ratio, CI: Confidence interval.

a cohort of 1012 patients for HCC surveillance. It should be calibrated according to different etiology of liver disease for individual risk stratification [39]. Therefore, we investigated the predictive role of ASAP score in CHB related HCC, and confirmed baseline ASAP score >0.31 increased 111-fold risk of HCC after adjustment of confounding factors. Our research provides the first evidence to apply the ASAP score as an HCC prediction tool in non-cirrhotic HBV carriers.

Our study identifies the predictive role of PIVKA-II in treatment-naïve non-cirrhotic HBV patients, which represents a large population of highrisk patients in Asia. We identify a cut-off 31.0 mAU/mL of PIVKA-II, which is not high, because it is derived from non-cirrhotic patients, and for prediction of HCC 1 year before, rather than for diagnostic purposes (cut-off value: 37.5-45.0 mAU/mL) [33, 34]. Our data validate that combination of AFP and PIVKA-II increase sensitivity without decreasing specificity, although the cost-effectiveness remains to be assessed [6]. Besides, using ASAP score increased the AUROC up to 0.90 for predicting HCC in 1 year, which highlights combining PIVKA-II and AFP increased the predictive power and potential to early detect HCC in non-cirrhotic HBV carriers. Therefore, we suggest when patients have normal AFP during HCC surveillance, PIVKA-II may be measured and abnormal PIVKA-II level warns the risk of HCC. The high PVIKA-II or ASAP level reminds the physician to closely monitor HCC in at-risk patients, and may detect HCC at an early stage.

There are a few limitations of this study. First, the small sample size from a single center and the lack of external validation reduced our statistical power; thus, further studies with larger case numbers are required for confirmation of our results. Second, missing data and selection bias were unavoidable in this retrospective study, which limited the accuracy and analytic aspects relatively. The applicability of our results needs to be verified in different ethnicity and populations.

In conclusion, both high PIVKA II level and ASAP score are promising to predict 1-year HCC risk in untreated non-cirrhotic CHB patients. Further prospective studies are needed to validate these findings.

Acknowledgements

We thank Miss Wan-Ting Yang, Shih-Wan Chou, Pei-Ying Chiang, and Pei-Ying Yang, for their administrative and statistical assistance, and the staff of the Department of Medical Research, National Taiwan University Hospital for the Integrated Medical Database (NTUH-iMD). We thank the 7th Core Laboratory of the Department of Medical Research, National Taiwan University Hospital for technical assistance. This work was supported by grants from the Ministry of Science and Technology, Taiwan (grant numbers MOST 109-2326-B-002-012-MY3, MOST 110-2326-B-400-004, MOST 110-2628-B-002-041), Ministry of Health and Welfare (MOHW111-TDU-B-221-114003), National Taiwan University Hospital (grant numbers VN-112-13, 112-TMU151, 112-S0245), and the Liver Disease Prevention & Treatment Research Foundation, Taiwan. The PIVKA-II assays were supported by Abbott. The funders have no roles in study design; data collection, analysis, interpretation of data, manuscript preparation or the decision to submit the paper for publication.

Disclosure of conflict of interest

None.

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References

- [1] Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A and Bray F. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2021; 71: 209-249.
- [2] Maucort-Boulch D, de Martel C, Franceschi S and Plummer M. Fraction and incidence of liver cancer attributable to hepatitis B and C viruses worldwide. Int J Cancer 2018; 142: 2471-2477.
- [3] Su TH, Hu TH, Chen CY, Huang YH, Chuang WL, Lin CC, Wang CC, Su WW, Chen MY, Peng CY, Chien RN, Huang YW, Wang HY, Lin CL, Yang SS, Chen TM, Mo LR, Hsu SJ, Tseng KC, Hsieh TY, Suk FM, Hu CT, Bair MJ, Liang CC, Lei YC, Tseng TC, Chen CL and Kao JH; C-TEAM study group and the Taiwan Liver Diseases Consortium. Four-year entecavir therapy reduces hepatocellular carcinoma, cirrhotic events and mortality in chronic hepatitis B patients. Liver Int 2016; 36: 1755-1764.
- [4] Singal AG, Pillai A and Tiro J. Early detection, curative treatment, and survival rates for hepatocellular carcinoma surveillance in patients with cirrhosis: a meta-analysis. PLoS Med 2014; 11: e1001624.
- [5] Marrero JA, Kulik LM, Sirlin CB, Zhu AX, Finn RS, Abecassis MM, Roberts LR and Heimbach JK. Diagnosis, staging, and management of hepatocellular carcinoma: 2018 practice guidance by the American Association for the Study of Liver Diseases. Hepatology 2018; 68: 723-750.
- [6] Omata M, Cheng AL, Kokudo N, Kudo M, Lee JM, Jia J, Tateishi R, Han KH, Chawla YK, Shiina S, Jafri W, Payawal DA, Ohki T, Ogasawara S, Chen PJ, Lesmana CRA, Lesmana LA, Gani RA, Obi S, Dokmeci AK and Sarin SK. Asia-Pacific clinical practice guidelines on the management of hepatocellular carcinoma: a 2017 update. Hepatol Int 2017; 11: 317-370.
- [7] European Association for the Study of the Liver. Electronic address: easloffice@easloffice. eu; European Association for the Study of the Liver. EASL clinical practice guidelines: management of hepatocellular carcinoma. J Hepatol 2018; 69: 182-236.
- [8] Korean Liver Cancer Association; National Cancer Center. 2018 Korean Liver Cancer As-

sociation-national cancer center Korea practice guidelines for the management of hepatocellular carcinoma. Gut Liver 2019; 13: 227-299.

- [9] Shao YY, Wang SY and Lin SM; Diagnosis Group; Systemic Therapy Group. Management consensus guideline for hepatocellular carcinoma: 2020 update on surveillance, diagnosis, and systemic treatment by the Taiwan Liver Cancer Association and the Gastroenterological Society of Taiwan. J Formos Med Assoc 2021; 120: 1051-1060.
- [10] Singal A, Volk ML, Waljee A, Salgia R, Higgins P, Rogers MA and Marrero JA. Meta-analysis: surveillance with ultrasound for early-stage hepatocellular carcinoma in patients with cirrhosis. Aliment Pharmacol Ther 2009; 30: 37-47.
- [11] Di Bisceglie AM, Sterling RK, Chung RT, Everhart JE, Dienstag JL, Bonkovsky HL, Wright EC, Everson GT, Lindsay KL, Lok AS, Lee WM, Morgan TR, Ghany MG and Gretch DR; HALT-C Trial Group. Serum alpha-fetoprotein levels in patients with advanced hepatitis C: results from the HALT-C trial. J Hepatol 2005; 43: 434-441.
- [12] Tateishi R, Yoshida H, Matsuyama Y, Mine N, Kondo Y and Omata M. Diagnostic accuracy of tumor markers for hepatocellular carcinoma: a systematic review. Hepatol Int 2008; 2: 17-30.
- [13] Chen DS, Sung JL, Sheu JC, Lai MY, How SW, Hsu HC, Lee CS and Wei TC. Serum alpha-fetoprotein in the early stage of human hepatocellular carcinoma. Gastroenterology 1984; 86: 1404-1409.
- [14] Liebman HA, Furie BC, Tong MJ, Blanchard RA, Lo KJ, Lee SD, Coleman MS and Furie B. Des-γcarboxy (abnormal) prothrombin as a serum marker of primary hepatocellular carcinoma. N Engl J Med 1984; 310: 1427-1431.
- [15] Li C, Zhang Z, Zhang P and Liu J. Diagnostic accuracy of Des-gamma-carboxy prothrombin versus α -fetoprotein for hepatocellular carcinoma: a systematic review. Hepatol Res 2014; 44: E11-25.
- [16] Johnson PJ, Pirrie SJ, Cox TF, Berhane S, Teng M, Palmer D, Morse J, Hull D, Patman G, Kagebayashi C, Hussain S, Graham J, Reeves H and Satomura S. The detection of hepatocellular carcinoma using a prospectively developed and validated model based on serological biomarkers. Cancer Epidemiol Biomarkers Prev 2014; 23: 144-153.
- [17] Yang T, Xing H, Wang G, Wang N, Liu M, Yan C, Li H, Wei L, Li S, Fan Z, Shi M, Chen W, Cai S, Pawlik TM, Soh A, Beshiri A, Lau WY, Wu M, Zheng Y and Shen F. A novel online calculator based on serum biomarkers to detect hepatocellular carcinoma among patients with hepatitis B. Clin Chem 2019; 65: 1543-1553.
- [18] Su TH, Peng CY, Chang SH, Tseng TC, Liu CJ, Chen CL, Liu CH, Yang HC, Chen PJ and Kao JH.

Serum PIVKA-II and alpha-fetoprotein at virological remission predicts hepatocellular carcinoma in chronic hepatitis B related cirrhosis. J Formos Med Assoc 2022; 121: 703-711.

- [19] Tseng TC, Liu CJ, Yang HC, Su TH, Wang CC, Chen CL, Kuo SF, Liu CH, Chen PJ, Chen DS and Kao JH. High levels of hepatitis B surface antigen increase risk of hepatocellular carcinoma in patients with low HBV load. Gastroenterology 2012; 142: 1140-1149, e3; quiz e13-4.
- [20] Bruix J and Sherman M; American Association for the Study of Liver Diseases. Management of hepatocellular carcinoma: an update. Hepatology 2011; 53: 1020-1022.
- [21] Fujita K, Kinukawa H, Ohno K, Ito Y, Saegusa H and Yoshimura T. Development and evaluation of analytical performance of a fully automated chemiluminescent immunoassay for protein induced by vitamin K absence or antagonist II. Clin Biochem 2015; 48: 1330-1336.
- [22] Zhang BH, Yang BH and Tang ZY. Randomized controlled trial of screening for hepatocellular carcinoma. J Cancer Res Clin Oncol 2004; 130: 417-422.
- [23] Hemker HC, Veltkamp JJ, Hensen A and Loeliger EA. Nature of prothrombin biosynthesis: preprothrombinaemia in vitamin K-deficiency. Nature 1963; 200: 589-590.
- [24] Ono M, Ohta H, Ohhira M, Sekiya C and Namiki M. Measurement of immunoreactive prothrombin precursor and vitamin-K-dependent gamma-carboxylation in human hepatocellular carcinoma tissues: decreased carboxylation of prothrombin precursor as a cause of Desgamma-carboxyprothrombin synthesis. Tumour Biol 1990; 11: 319-326.
- [25] Marrero JA, Su GL, Wei W, Emick D, Conjeevaram HS, Fontana RJ and Lok AS. Des-gamma carboxyprothrombin can differentiate hepatocellular carcinoma from nonmalignant chronic liver disease in american patients. Hepatology 2003; 37: 1114-1121.
- [26] Yu R, Tan Z, Xiang X, Dan Y and Deng G. Effectiveness of PIVKA-II in the detection of hepatocellular carcinoma based on real-world clinical data. BMC Cancer 2017; 17: 608.
- [27] Koike Y, Shiratori Y, Sato S, Obi S, Teratani T, Imamura M, Yoshida H, Shiina S and Omata M. Des-gamma-carboxy prothrombin as a useful predisposing factor for the development of portal venous invasion in patients with hepatocellular carcinoma: a prospective analysis of 227 patients. Cancer 2001; 91: 561-569.
- [28] Kokudo N, Takemura N, Hasegawa K, Takayama T, Kubo S, Shimada M, Nagano H, Hatano E, Izumi N, Kaneko S, Kudo M, Iijima H, Genda T, Tateishi R, Torimura T, Igaki H, Kobayashi S, Sakurai H, Murakami T, Watadani T and Matsuyama Y. Clinical practice guidelines for hepa-

tocellular carcinoma: the Japan Society of Hepatology 2017 (4th JSH-HCC guidelines) 2019 update. Hepatol Res 2019; 49: 1109-1113.

- [29] Omata M, Lesmana LA, Tateishi R, Chen PJ, Lin SM, Yoshida H, Kudo M, Lee JM, Choi BI, Poon RT, Shiina S, Cheng AL, Jia JD, Obi S, Han KH, Jafri W, Chow P, Lim SG, Chawla YK, Budihusodo U, Gani RA, Lesmana CR, Putranto TA, Liaw YF and Sarin SK. Asian Pacific Association for the study of the liver consensus recommendations on hepatocellular carcinoma. Hepatol Int 2010; 4: 439-474.
- [30] Wang SY, Su TH, Chen BB, Liu CJ, Liu CH, Yang HC, Tseng TC, Chen PJ and Kao JH. Prothrombin induced by vitamin K absence or antagonist-II (PIVKA-II) predicts complete responses of transarterial chemoembolization for hepatocellular carcinoma. J Formos Med Assoc 2022; 121: 1579-1587.
- [31] Yang Y, Li G, Lu Z, Liu Y, Kong J and Liu J. Progression of prothrombin induced by vitamin K absence-II in hepatocellular carcinoma. Front Oncol 2021; 11: 726213.
- [32] Feng H, Li B, Li Z, Wei Q and Ren L. PIVKA-II serves as a potential biomarker that complements AFP for the diagnosis of hepatocellular carcinoma. BMC Cancer 2021; 21: 401.
- [33] Seo SI, Kim HS, Kim WJ, Shin WG, Kim DJ, Kim KH, Jang MK, Lee JH, Kim JS, Kim HY, Kim DJ, Lee MS and Park CK. Diagnostic value of PIV-KA-II and alpha-fetoprotein in hepatitis B virusassociated hepatocellular carcinoma. World J Gastroenterol 2015; 21: 3928-3935.
- [34] Ji J, Liu L, Jiang F, Wen X, Zhang Y, Li S, Lou J, Wang Y, Liu N, Guo Q, Jia Y and Gao C. The clinical application of PIVKA-II in hepatocellular carcinoma and chronic liver diseases: a multi-center study in China. J Clin Lab Anal 2021; 35: e24013.
- [35] Yu R, Xiang X, Tan Z, Zhou Y, Wang H and Deng G. Efficacy of PIVKA-II in prediction and early detection of hepatocellular carcinoma: a nested case-control study in Chinese patients. Sci Rep 2016; 6: 35050.
- [36] Best J, Bechmann LP, Sowa JP, Sydor S, Dechêne A, Pflanz K, Bedreli S, Schotten C, Geier A, Berg T, Fischer J, Vogel A, Bantel H, Weinmann A, Schattenberg JM, Huber Y, Wege H, von Felden J, Schulze K, Bettinger D, Thimme R, Sinner F, Schütte K, Weiss KH, Toyoda H, Yasuda S, Kumada T, Berhane S, Wichert M, Heider D, Gerken G, Johnson P and Canbay A. GALAD score detects early hepatocellular carcinoma in an international cohort of patients with nonalcoholic steatohepatitis. Clin Gastroenterol Hepatol 2020; 18: 728-735, e4.
- [37] Yang JD, Addissie BD, Mara KC, Harmsen WS, Dai J, Zhang N, Wongjarupong N, Ali HM, Ali

HA, Hassan FA, Lavu S, Cvinar JL, Giama NH, Moser CD, Miyabe K, Allotey LK, Algeciras-Schimnich A, Theobald JP, Ward MM, Nguyen MH, Befeler AS, Reddy KR, Schwartz M, Harnois DM, Yamada H, Srivastava S, Rinaudo JA, Gores GJ, Feng Z, Marrero JA and Roberts LR. GALAD score for hepatocellular carcinoma detection in comparison with liver ultrasound and proposal of GALADUS score. Cancer Epidemiol Biomarkers Prev 2019; 28: 531-538.

- [38] Liu SY, Li C, Sun LY, Guan MC, Gu LH, Yin DX, Yao LQ, Liang L, Wang MD, Xing H, Zhu H, Pawlik TM, Lau WY, Shen F, Tong XM and Yang T. ASAP score versus GALAD score for detection of hepatitis C-related hepatocellular carcinoma: a multicenter case-control analysis. Front Oncol 2022; 12: 1018396.
- [39] Li B, Zhao Y, Cai W, Ming A and Li H. Validation and update of a multivariable prediction model for the identification and management of patients at risk for hepatocellular carcinoma. Clin Proteomics 2021; 18: 21.

Cut-off value	Sensitivity (%)	Specificity (%)	Youden Index
20	88.41	22.55	0.1096
25	79.71	54.90	0.3461
31	65.22	81.37	0.4659
35	55.07	88.24	0.4331
40	46.38	92.16	0.3854
50	33.33	98.04	0.3137

Supplementary	/ Table 1. The	cut-off values	for PIVKA-II for	predicting HCC in	1 vear
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N=171, AUROC=0.76 (95% CI: 0.68-0.84).

Supplementary Table 2. The cut-off values for PIVKA-II for predicting HCC in 1 year in patients with AFP <20 ng/mL

Cut-off value	Sensitivity (%)	Specificity (%)	Youden Index
15	100.00	3.13	0.0313
20	95.00	22.92	0.1792
25	82.5	56.25	0.3875
31	67.50	82.29	0.4979
35	57.50	88.54	0.4604
40	45.00	92.71	0.3771
50	32.50	96.88	0.2938

N=136, AUROC=0.80 (95% CI: 0.72-0.89).



Supplementary Figure 1. The receiver operating characteristic curve of ASAP score for predicting HCC in 1 year in patients with AFP <20 ng/mL.

PIVKA-II and ASAP score predicts 1-year HCC

Cut-off value	Sensitivity (%)	Specificity (%)	Youden Index
0.25	100.00	48.91	0.4891
0.28	97.50	58.70	0.5620
0.31	92.50	71.74	0.6424
0.40	77.50	81.52	0.5902
0.50	45.00	94.57	0.3957
0.60	30.00	100.00	0.3000

Supplementary	Table 3.	The cut-off	values for	ASAP score	e for predicting	g HCC in 1	year
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N=132, AUROC=0.90 (95% CI: 0.85-0.95).

Supplementary Table 4.	The cut-off values for ASAP score for predicting HCC in 1 year in patients
with AFP <20 ng/mL	

Cut-off point	Sensitivity (%)	Specificity (%)	Youden Index
0.25	100.00	50.00	0.5000
0.28	96.67	58.89	0.5556
0.31	90.00	72.22	0.6222
0.40	76.67	82.22	0.5889
0.50	40.00	94.44	0.3444
0.60	26.67	100.00	0.2667

N=120, AUROC=0.89 (95% CI: 0.83, 0.95).