

Review Article

Cancer stem cells of head and neck squamous cell carcinoma; distance towards clinical application; a systematic review of literature

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Abstract: Head and neck squamous cell carcinoma (HNSCC) is the major pathological type of head and neck cancer (HNC). The disease ranks sixth among the most common malignancies worldwide, with an increasing incidence rate yearly. Despite the development of therapy, the prognosis of HNSCC remains unsatisfactory, which may be attributed to the resistance to traditional radio-chemotherapy, relapse, and metastasis. To improve the diagnosis and treatment, the targeted therapy for HNSCC may be successful as that for some other tumors. Nanocarriers are the most effective system to deliver the anti-cancerous agent at the site of interest using passive or active targeting approaches. The system enhances the drug concentration in HCN target cells, increases retention, and reduces toxicity to normal cells. Among the different techniques in nanotechnology, quantum dots (QDs) possess multiple fluorescent colors emissions under single-source excitation and size-tunable light emission. Dendrimers are the most attractive nanocarriers, which possess the desired properties of drug retention, release, unaffected by the immune system, blood circulation time enhancing, and cells or organs specific targeting properties. In this review, we have discussed the up-to-date knowledge of the Cancer Stem Cells of Head and Neck Squamous Cell Carcinoma. Although a lot of data is available, still much more efforts remain to be made to improve the treatment of HNSCC.

Keywords: Cancer stem cells, head neck squamous cell carcinoma, target therapy, nanotechnology, quantum dots

Introduction

Head and neck cancer (HNC) is the malignancy arising from the epithelium of the upper digestive tract and upper respiratory tract, including the nasal cavity, paranasal sinus, pharynx, oral cavity, larynx, and cervical esophagus [1], as shown in **Figure 1**. Head and neck squamous cell carcinoma (HNSCC) are the major pathological type of HNC [2, 3]. Only 40-50% of people with HNSCC live for five years after being diagnosed [4]. The main causes of HNSCC are exposure to alcohol, betel quid products, and tobacco; where the risk factors include carcinogens, tobacco smoking, alcohol, human papillomavirus (HPV) infection, and genetic predisposition [5-7]. The major issue in HNSCC pathogenesis is carcinomas development in mucosal epithelium preneoplastic fields which are made up of genetically altered cells and

clonally similar to carcinoma [8]. On the tumours excised state, it may extend into the surgical margins, causing second primary tumors. The dissemination of HNSCC occurs in lymph nodes in the neck region. Here, the unravelling of molecular and biological process of HNSCC may be useful for better management and development of personalized therapies [9, 10]. The disease ranks the sixth most common malignancy worldwide, with an increasing incidence rate yearly [11]. Squamous cell carcinoma (SCC) accounts for over 90% of all head and neck malignancies [12, 13]. Despite various interdisciplinary therapy, HNSCC treatment is ineffective [14-16]. The theory of cancer stem cells (CSCs) is one of the milestones of cancer therapy. According to this theory, CSCs are a small subpopulation of tumor bulk with the abilities of self-renewal and differentiation into secondary heterotypic groups [17].

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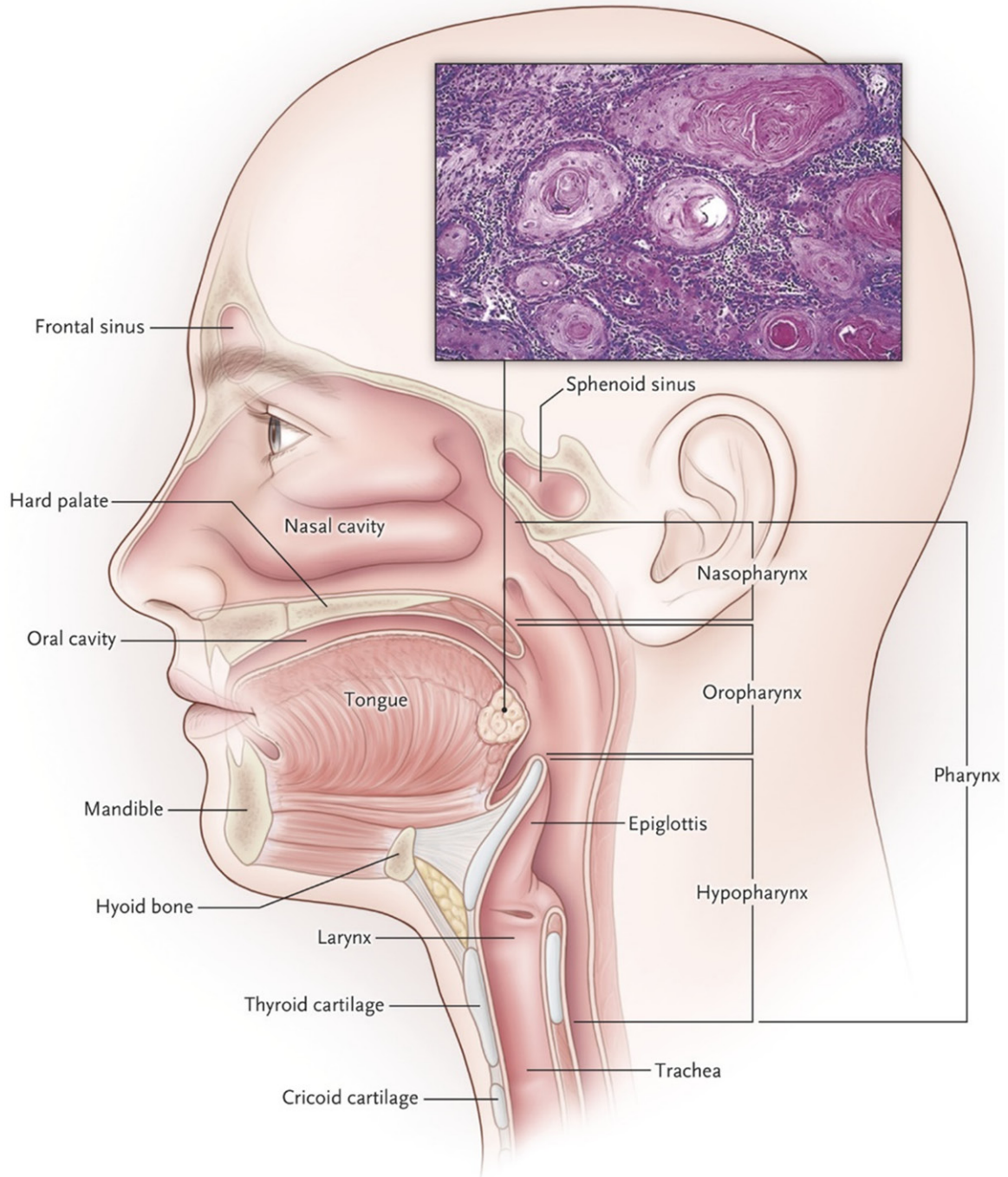


Figure 1. Major anatomical sites of HNSCC. Organs involved in HNC are the nasal cavity, paranasal sinus, pharynx, oral cavity, larynx, and cervical esophagus. With permission from Chow, 2020.

Eliminating CSCs may help to cure cancer. The Stemness Phenotype Model proposed a model and stated that CSCs have no specific subpopulation of tumors and cancer cells possess plasticity in CSCs and non-CSCs stemness that can interconvert into each other in different microenvironments. This model predicts a pure

CSC phenotype cancer cell to pure non-CSC [18-20]. The dissemination from primary tumors and seeding of new tumors from other distant body places involves an invasion-metastasis cascade. Carcinoma dissemination may occur by two mechanisms - single cell dissemination and collective dissemination of

tumor clusters. CSCs are believed to be the origin of a tumor and the source of metastasis. CSCs have more substantial potential for stemness, epithelial-to-mesenchymal transition (EMT), therapeutic resistance, and immune escape [21, 22]. Cancer-associated fibroblasts (CAFs) are major components of their microenvironment, performing numerous functions including remodeling, matrix deposition, signaling, and crosstalk with host immunity [23]. New markers and their molecular interactions may be identified for more tailored cancer therapies. Post-therapy relapse may be attributed to inadequate elimination of CSCs [24]. The stem-like cells have been isolated and identified from most solid tumors, including HNSCC. The cluster of differentiation (CD) molecules including CD44, CD133, and ALDH (Aldehyde dehydrogenase) have been identified as specific markers of CSCs in HNSCC. Side population (SP) cells and cells with the ability of sphere formation in a medium contained in EGF are also identified as stem-like cells of HNSCC [25]. The PI3K pathway in HNSCC shows significant activating genetic events which may be useful for further studies [26]. This will also unravel the molecular events behind it. In molecular biology, HNC is a manyfold complex process proceeding from carcinogen causing a single mutation to the dysregulation of many metabolic processes in signaling pathways where these events occur in different conditions and states. The molecular biology of HNS still needs much work to be recognized at every step and stage for a better understanding [27]. Many signaling pathways have been identified involved in HNC which may be a potential target and also inhibition of epidermal growth factor receptor (EGFR) for therapeutic strategy is one of the key targets.

The HNSCC microenvironment is permissive and seems more aggressive in nature, and the response to tumour stress and hypoxia by the immune system are still needed to be explored. Solving these phenomena may increase our knowledge of molecular biology for better management of HNSCC in future.

Potential regulators of HNSCC CSCs: potential targets and current bottleneck

We searched PUBMED's literature on HNSCC and CSCs and collected data from CSCs regulators (Tables S1, S2 and S3). These regula-

tors include: Phosphoinositide 3 kinase (PI3K) [28], mTOR signaling pathway [29], Hyaluronan (HA) [30], Snail [31], Human papillomavirus type 16 (HPV16) [32], Maternal embryonic leucine zipper kinase (MELK) [33], Renin-angiotensin system (RAS) [34], Short palate, lung and nasal epithelium clone 1 (SPLUNC1) and Mixed lineage leukemia-3 (MLL3) [35], X-linked inhibitor of apoptosis protein (XIAP) [36], Moloney murine leukemia virus insertion site 1 (BMI1) [37], Mitogen-activated protein kinases (p38 MAPK) [38], Wingless/Integrated (Wnt)/ β -catenin [39], Sry-like high-mobility group box (SOX8) [40], Sry-like high-mobility group box (SOX2) [41], mitotic arrest deficient 1 (RARS-MAD1L1) [42], LIN28 proteins [43], heat shock protein 90 (HSP90) [44], 5T4 (an N-glycosylated transmembrane protein whose gene is found on chromosome 6q14-15) [45], c-Met (a proto-oncogene) [46, 47], metastasis-associated colon cancer-1 (MACC1) [48], The Hippo-TAZ [49], Oct-4 [50], RXR α [51], epidermal growth factor receptor (EGFR) [52, 53], Notch1 [54, 55], Disruptor of telomeric silencing 1 (DOT1L) [56], Nucleotide-binding domain (NOD)-like receptor protein 3 inflammasome (NLRP3) [57], Tumor necrosis factor receptor-associated factor 6 (TRAF6) [58], glucose-regulated protein 78 (GRP78) [59], S100 Calcium Binding Protein A4 (S100A4) [60], RhoC (a member Rho family of GTPases) [61], Glycogen synthase kinase-3 beta (GSK3 β) [62], c-Fos (a proto-oncogene) [63], G9a (also called EHMT2 or KMT1C, is a major euchromatic methyltransferase) [64], histone deacetylase (HDAC) [65], Senescence-associated secretory phenotype (SASP) [66], PinX1 (a potent telomerase regulator) [67], Sialyl Lewis X (sLeX) [68], fucosylation [69], CD200 [70], casein kinase 2 (CK2, a constitutively active Ser/Thr protein kinase) [71], CD10 [72], SDF-1 α /CXCR4 (Stromal cell-derived factor-1) [73], Smad ubiquitination regulatory factor (SMURF1) [74], Procollagen-lysine, 2-oxoglutarate 5-dioxygenase 2 (PLOD2) [75], GLI family zinc finger 3 (GLI3); a mediator of genetic diseases [76], Zinc finger E-box binding homeobox 1 (ZEB1/ZEB2) [77], Inhibitor of binding/differentiation 2 (Id2) [78], Bone morphogenetic protein 4 (BMP4) [79], Interferon-stimulated gene 15 (ISG15) [80], ecotropic viral integration site 1 (EVI1) [81], Signal transducer and activator of transcription 3 (STAT3) [82], S-phase kinase associated protein 2 (Skp2) [83], Latent membrane protein 2A (LMP2A)

[84], Olfactomedin-4 (OLFM4) [85], topoisomerases [86], JARID1B (also known as PLU-1, is a Retinoblastoma-Binding Protein 2 (RBP2) Homolog, and a member of the jumonji, AT rich interactive domain (JARID) family with H3K4 demethylase activity) [87], Slug (an epithelial mesenchymal transition master gene) [88], CC-chemokine receptors (CCL21/CCR7) [89], Metastasis-associated Protein 3 (MTA3) [90], CMTM6 (belongs to the CKLF-like MARVEL transmembrane domain-containing family; CMTM1-8) [91], phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit alpha (PIK3CA) [92], Secreted Frizzled-related Protein 1 (SFRP1) [93], Nuclear factor-erythroid factor 2-related factor 2 (Nrf2) [94], Glioma-associated oncogene homologue 1 (GliA1) [95], Cd47-Signal Regulatory Protein α (CD47-SIRP α) [96], Wolf-Hirschhorn syndrome candidate 1 (WHSC1) [97], Zinc finger and SCAN domain containing 4 (ZSCAN4) [98], cystine transporter (xCT) [99], Mediator Complex Subunit 28 (RCOR1/MED28) [100], Wnt Family Member 5A (WNT5A) [101], the estrogen-regulated anterior gradient 2 (AGR2) [102], Cathepsins [103], cytochrome (CYP1B1) [104], signaling promotes regenerative proliferation (VAV2) [105], Tetraspanin 1 (TSPAN1) [106], Tropomyosin-related tyrosine kinase B (TrkB) [107], phosphoglycerate kinase (PGK1) [108], insulin-like growth factor 1 (IGF-1) [109], super-enhancers (SEs) [110], HOXA10-AS (a regulator of homeobox A10) [111], hydroxysteroid 17- β dehydrogenase 7, hydroxysteroid 17- β dehydrogenase 7 (HSD17B7) [112], Prostaglandin E₂ [113]. Aberrant miRNAs (lower level) miR-204 [114], miR-34a [115], microRNA-200c [116], MiR-520b [117], microRNA let-7a [118], Let-7c [119], miR-34a [120] and Long noncoding RNA-Pvt1 (LncRNA-Pvt1) [121], Long intergenic non-protein coding RNA, p53 induced transcript (LINC-PINT) [122], Hematopoietic cell-specific Lyn substrate-associated protein X-1 (miR-125a/HAX-1) [123], LINC00963 [124], miR-495 (low) [125], CCNG2 (miR-1246/CCNG2) [126] could regulate HNSCC CSCs. The niche associated factors; hypoxia [127], interleukin-6 (IL-6) [128], IL-4 [129], IL-1 β [130], EGF [131], TGF- β [132], tumor associated markers (TAM) markers [133], cancer-associated fibroblast [134] also promote the stemness of HNSCC CSCs. In addition, chewing tobacco [135], arecoline-exposure [136], nicotine [137], and cigarette smoke [138] could induce and activate

malignant phenotypes and stemness. It may suggest that HNSCC patients need a lifelong ban on tobacco and arecoline.

From data (Tables S1, S2 and S3), we have summarized the following two basic pieces of information 1). Most studies used HNSCC cell lines or available squamous cell carcinoma cell lines of a particular organ as in vitro research subjects; 2). Most studies have applied CSCs markers (CD44, ALDH, CD133), common CSCs related factors, SP traits, and sphere formation to identify HNSCC CSCs. However, in solid tumors, CSCs may not express a single marker or even none. In addition, CSCs isolated from cancer cell lines are not representative of solid tumors surrounded by niches. Although we have thoroughly reviewed the potential regulators of HNSCC CSCs, it remains unclear which pathway is dominant for HNSCC CSCs. In addition, CSCs may evolve through genetic instability leading to dynamic expression of markers and regulators. In a recent study, Salazar-García et al. [139] performed whole-exome sequencing to analyze the germinal line, tumor cells, and CSC ALDH⁺ samples from different HNSCC patients. They found that the difference in genes of oncogenic pathways. Therefore, exploring new ideas and novel strategies is essential to target HNSCC CSCs effectively.

Key molecules involved in the transcription of cancer stem cell genes and premetastatic genes in cancer stem cells

The distinct CSCs population in cancers is different. However, in HNSCC the specific CSCs population are CD133, CD44, CD98, ALDH, CD200, ALDH, CD44, GRP78 and BMI1 [140]. Similarly, CSC various abnormal extrinsic and intrinsic signals including niche, and mutations are involved in pathways deregulation that leading to their maintenance.

The Wnt/ β -catenin canonical pathway is initiated when Wnt ligand binds to Frizzled receptor and 5/6 coreceptors protein. This causes the recruitment of Axin and disheveled which prevents the degradation to protection of β -catenin. Free β -catenin complexes with TCF/LEF (T-cell/lymphoid enhancer factors) and regulate the Wnt target stem cell genes. Phosphorylation of CD133 also results to AKT activation and the NF- κ B pathway, leading to stemness genes activation. Hypoxia inducible

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factor-1- α (HIF1 α) and c-Myc (myelocytomatosis) are involved in increase of glycolysis and prevent the oxidative phosphorylation in CSCs. Activation of pyruvate dehydrogenase kinases (PDK1-3) is brought by HIF1 α , to stop pyruvate dehydrogenase (PDH), leading to oxidative phosphorylation inhibition. C-Myc has a role in activation of the hexokinase, Glucose transporter -1 receptor, and phosphofructokinase which are in favor of glycolysis [140].

Moreover, the activities of CSCs are governed by numerous pluripotent transcription factors. Some of these factors are Oct4, Sox2, Nanog, Kruppel-like factor 4 (KLF4) and Myc. Besides these pluripotent transcription factors, several intracellular signaling pathways, such as Wnt, NF- κ B, Notch, Hedgehog, JAK-STAT, TGF/SMAD, and PPAR along with external factors like vascular niches, hypoxia, tumor-associated macrophages, cancer-associated fibroblasts or mesenchymal stem cells, extracellular matrix, and exosomes play vital roles in the regulation of CSCs [141]. A favorable microenvironment is required for cancer development. Some studies showed intercellular communications are mediated by microvesicles (MVs) released by cells. Large MVs are produced by the tumor cells and released in the circulation and other biological fluids [142, 143]. These MVs have the pleiotropic effect, enabling their involvement in cancer development, progression, and premetastatic niche formation [144]. Normal stem cells are an important source of MVs that may act as paracrine mediators of genetic information through horizontal transfer [145-148]. The transmembrane glycoprotein CD44, a common stem cell niche component, integrates signaling in normal stem cells, cancer stem cells, and (pre)metastatic niches [149]. The biological markers of stem cells in pancreatic CSCs (Pan-CSC) include CD44v6, c-Met, Tspan8, alpha6beta4, C-X-C-chemokine receptor 4 (CXCR4), CD133, epithelial cellular-adhesion factor (EpCAM) and claudin7 [150], breast stem cancer cells such as ALDH1A1 (aldehyde-dehydrogenase 1 family member-A1) [151, 152], and brain CSCs such as CD133 and in HNSCC such as ALDH⁺, CD44⁺, CD166⁺ [153-155] have been investigated. Cancer stem celllike phenotype and the difference in the gene expression pattern responsible for diverse biological roles in HNSCC has been studied recently. Hypoxia-regulated genes have been

shown to be helpful for the prediction of radiotherapy responses in HNSCC patients [156]. The biological role of polycomb (PcG) genes *Bmi1* and *TERT* in tumorigenesis in human stromal cells which were immortalized as the HNSCC model studied recently. It was found that *Bmi1* was predominantly expressed in early head and neck squamous cell carcinoma, indicating that the PcG is essential in early cancer development [157]. The role of the p53-p16/RB pathway in HNSCC can be focused on interpreting the early carcinogenesis of HNSCC for better management. Besides, m6A modulators have been emerging cancer progression molecular mechanisms [158].

Stem cells regulators

CSCs have the ability of self-renewal, that may lead them to tumorigenesis [159]. The symmetrical division of CSCs divide them into two CSCs or, in other hand into one CSC and one daughter cell [160]. The symmetrical splitting manner of CSCs expand them to excessively increase cell growth, resulting in formation of tumor [161]. Normal stem cells and CSCs follow same regulatory signaling pathways, for example the Wnt/ β -catenin [162], Sonic Hedgehog (Hh) [163], and Notch pathways. These pathways are involved in the self-renewal process [164]. Besides, other signaling molecules, for example PTEN and the polycomb family, are involved in the regulation of CSCs cycle [165].

Several factors are involved in the regulation of stem cells. These might be transcription factors which play a basic role in the proliferation of stem cells. Reprogramming of somatic stem cells can be carried out to generate iPSC by increasing the expression of the transcription factors such as Octamer transcription factor4 (Oct4), Sox2, Nanog, KLF4, and Myc [166, 167]. Besides, SRY and Oct4 are considered as potential differentiation therapy targets in stem cells. The over-expression of Sox2 transcription factor in basal-like breast cancer may be supportive to characterize the cell phenotypes of poorly differentiated/stem cells [168]. The deletion p53 and Myc synergizes to induce proliferation and tumorigenesis in hepatocytes [169]. Besides the loss of p53, Bcl-2 and BMI-1 overexpression and deletion of p19ARF also shown the regulation of Myc in CSCs survival and proliferation [170]. In addition, the perturbation of Myc results in

Hepatocellular carcinoma stem cells differentiating into hepatocytes and biliary duct cells resulting the formation of bile duct structures [171].

CSCs have multipotential characteristics. They can also differentiate into other cell types in addition to their self-renewal capabilities. Bonnet and Dick [172] showed that CD34⁺/CD38⁻ LSCs (Leukemia stem cells) were able to differentiate and proliferate in severe combined immunodeficient mice. CSCs isolated from the brain of patients are positive for the markers CD133 and nestin [173]. CSCs from the breast indicated varied expression patterns of surface biomarkers. Some of these surface biomarkers are CD44⁺, CD24⁻, SP, and ALDH⁺ [174-176]. CD271⁻ or CD271⁺ melanoma stem cells were able to generate tumors in SCID mice [177].

Initial steps on targeting CSCs

Post-therapy recurrence and metastasis are related to residual CSCs. Therefore, effectively eliminating the subpopulation CSCs is essential in anti-cancer activities [178]. Therefore, much effort is needed to target HNSCC CSCs in preclinical studies.

Small molecular target drugs

Some authors reported that some molecular target drugs could eliminate HNSCC CSCs in vitro (Table S2). EGFR is one of the four members of the HER tyrosine kinase (RTK) receptor family, composed of EGFR/HER1/erbB1, HER2/erbB2, HER3/erbB3, and HER4/erbB4. The receptors of RTK, such as epidermal growth factor (EGF) and transforming growth factor alpha (TGF- α), can activate intracellular signaling pathways that control growth, differentiation, survival, and invasion [179]. The overexpression of EGFR is a frequent molecular alteration associated with aggressiveness, resistance to treatment, and poor clinical outcomes in HNSCC [180]. Therefore, the EGFR-targeted drug cetuximab has been recommended as a clinical combination with dihydropyrimidine dehydrogenase (DDP)-based chemotherapy as an anti-HNSCC treatment in NCCN guidelines. However, resistance to chemotherapy still exists. Recently, it's been reported that combining Cetuximab and Erlotinib could induce CSCs differentiation and transit

EMT-CSCs back to the epithelial phenotype, which sensitizes treatment and restricts local invasion and metastasis [181]. More recently, Roy S et al. [182] found that Afatinib, the second generation of FDA-approved pan-EGFR inhibitor, could inhibit the growth of HNSCC CSCs in vitro and in vivo by inducing severe apoptosis and an uncommon weak protective autophagic response preferentially in stem-like HNSCC cells [183]. However, CSCs usually have low EGFR expression and overexpress the anti-apoptotic Bcl-2 protein [184]. Anti-apoptosis members of the Bcl-2 family are associated with a poor prognosis of HNSCC. ABT-737 is a well-characterized BH3 mimetic that prevents binding ligands to anti-apoptotic Bcl-2 family protein and indirectly activates the pro-apoptotic Bcl-2 family members. Marion Gilormini et al. [185] suggested that ABT-737, alone or in synergism with radiation, can efficiently eliminate stem-like quiescent HNSCC cells in vitro and synergistically inhibit the growth of xenograft tumours. Another group observed the combined effect of ABT-199, Bcl-2 inhibitor, cetuximab, and radiation as anti-CSCs [186]. The combination significantly inhibited proliferation, invasion/migration, and resistance to apoptosis of HNSCC CSCs in vitro and strongly reduced the tumor growth and increased in vivo survival without side effects. Like EGFR, Lin28, an essential RNA-binding protein, also plays a critical role in regulating the balance between stemness and differentiation in embryonic stem cells (ESC) [187]. Chen and coauthors [188] showed that the combination of C1632 (Lin28 inhibitor) and metformin (anti-CSCs hypoglycemic medication) exerts synergistic anti-tumor effects in OSCC cell lines and xenograft tumor growth. Bruton's tyrosine kinase (BTK), a cytoplasmic non-receptor tyrosine kinase, is upstream of the phosphoinositide 3-kinase (PI3K-AKT) pathway, phospholipase-C, protein kinase-C, and NF- κ B, performing many functions, including cellular differentiation, proliferation, and adhesion to innate and adaptive immune responses [189]. Although BTK is mostly involved in the hematologic tumor, it is expressed aberrantly in concurrent chemoradiotherapy (CCRT) resistant OSCC tissues, correlated with stemness and EMT factors, and influences survival rate. The Ibrutinib, a first-class BTK inhibitor, reduced CSCs number and increased the DDP sensitivity of OSCC SP-derived cells [190]. Glycogen

synthase kinase 3 β (GSK3 β) controls the shift from EMT-CSCs to CSCs-epi. Hideo et al. demonstrated that GSK3 β inhibition induced mesenchymal-to-epithelial transition (MET) from CD44 (high)/ESA (low) cells to CD44 (high)/ESA (high) cells and pre-existing CD44 (high)/ESA (high) cells to differentiate. The CD44 (high)/ESA (low) cells overexpressed dihydropyrimidine dehydrogenase (DPD), a factor affecting the therapeutic sensitivity to 5-FU. Combination of both DPD inhibitor, 5-chloro-2,4-dihydropyridine (CDHP) and GSK3 β inhibitors markedly enhanced 5-FU-induced apoptosis of CD44 (high)/ESA (low) cells [191]. In addition, the Wnt/ β -catenin signal is another CSCs regulating pathway. Tankyrases are members of the poly (ADP-ribose) polymerase (PARP) family proteins, which serve as regulators of the canonical Wnt/ β -catenin signaling [192]. XAV-939, a small molecule of tankyrase inhibitor, reduced CSCs-mediated chemoresistance in DDP-resistant HNSCC cell lines combined with DDP via DNA damage [193]. Similarly, a recent study identified LF3, a 4-thioureido-benzenesulfonamide derivative, as a potent and specific inhibitor of activated Wnt/ β -catenin signals [194]. In this study, the self-renewal capacity of head neck CSCs was blocked by LF3, as examined by sphere formation. Beside, secreted frizzled-related protein 4 (sFRP4) is one of five members of the sFRP family and a naturally extracellular inhibitor of Wnt signaling [195]. Warrior's group showed that sFRP4 decreased the expression of CSCs markers (CD44 and ALDH) and inhibited proliferation, EMT, and enhanced chemosensitivity of HNSCC CSCs [196]. Moreover, histone deacetylases (HDACs) regulate several genes involved in cancer initiation and aggressiveness [197]. Similarly, Royal jelly acid showed suppression in HCC tumorigenicity that inhibited H3 histone lactylation targeted H3K9la and H3K14la sites [198]. In addition, transcriptome analysis of transgenic mice models predicted several oncogenes in brain [199], lungs [200].

Studies demonstrated that HDACs inhibitors (HDACi), suberoylanilide hydroxamic acid (SAHA), and trichostatin A (TSA), inhibited the stemness of HNSCC CSCs, and enhanced the DDP sensitivity, which may be attributed to reduced NANOG and Survivin expression [201, 202]. Similarly, valproic acid (VPA), another HDACi, inhibited the self-renewal abilities of

HNSCC CSCs with decreased expression of CSCs markers, such as Oct4, Sox2, and CD44, and enhanced sensitivity to DDP via reducing ABCC2, six and inducing apoptosis. The VPA combined with DDP attenuated xenograft tumor growth [203]. Entinostat, another HDACi, could induce cycle arrest (G0/G1 phase), tumor apoptosis and increase in ROS production, and significant reductions in HNSCC CSCs [204]. In addition, cancer cells have a super capacity to ROS scavenger via redox enzyme. Therefore, combining dimethyl fumarate (DMF), a GSH (glutathione) inhibitor, and Buthionine sulfoximine (BSO), a GSH synthesis inhibitor, could sensitize HNSCC CSCs to radiotherapy. It suggests that reduced antioxidant capacity may be a striking strategy to target CSCs [205].

Another reason for the radio-resistance of HNSCC CSCs is an extended G2/M arrest phase. Therefore, UCN-01, a checkpoint kinase (Chk1) inhibitor, and all-trans retinoic acid (ATRA), an inducer of differentiation, combined with irradiation drastically decreased the surviving fraction of HNSCC CSCs [206]. MEDI-5117 is an IL-6 inhibitor. Finkel and coauthors found that low-dose MEDI5117 antibodies decreased the CSCs fraction in three low-passage patient-derived xenografts (PDX) models of HNSCC [207]. They conducted a clinical trial in which MEDI5117 prevented tumor recurrence when used in the adjuvant setting. BMI-1, downstream of IL-6, is a CSCs-related factor. Jia et al. [208] demonstrated BMI-1⁺ CSCs contributed to the failure of PD-1 and DDP treatment in an HNSCC mouse model. PTC209 plus PD-1 inhibitor could eliminate CSCs via recruiting CD8⁺ T cells and prevent the progression and relapse of HNSCC. Meanwhile, an inhibitor of the IL-6R/BMI-1 axis, Tocilizumab, could target HNSCC CSCs via reversing DDP-induced self-renewal and chemoresistance in DDP-resistant HNSCC cells [209]. COX-2 is an inducible enzyme that triggers the biosynthesis of prostaglandins. Celecoxib, a COX-2 inhibitor, inhibited RNA expression of stemness-related genes and sphere formation in HNSCC cell lines [210].

Compounds extracted from natural herbs

Although targeted molecular drugs have shown the ability of anti-HNSCC CSCs experimentally, resistance is a challenging problem clinically.

These causes include activating mutations in the target itself and activating various compensatory pathways and EMT as a major mechanism of resistance [211]. Emerging evidence demonstrates that pure compounds extracted from natural herbs or plants exhibit features of multi-targets, anti-CSCs, and less toxicity. *Ovatodiolide (OV)*, a bioactive chemical substance purified from *Anisomeles indica* (L.) Kuntze (Labiatae) could inhibit NPC tumor sphere formation, attenuate NPC stem cell tumorigenicity, and enhance the sensitivity via reducing the expression of p-FAK, p-PXN, F-actin, slug proteins, SOX2, OCT4, and JAK-STAT signaling pathway [212]. In addition, *Epigallocatechin-3-gallate (EGCG)* is active polyphenolic catechin purified from *green tea*. Lee et al. examined the anti-tumor effect of EGCG on HNSCC CSCs. They demonstrated that EGCG inhibits the self-renewal capacity of HNSC CSCs via inhibition of stem cell markers, such as Oct4, Sox2, Nanog, CD44, ATP-binding cassette subfamily-G member-2 (ABCG2), and Notch signaling [213].

Quercetin is a polyphenolic flavonoid compound in nuts, teas, vegetables, herbs, and people's daily diets [214]. Chang et al. found that Quercetin could reduce the stemness of HNSCC CSCs through the decreased expression of Twist, N-cadherin, and Vimentin [215]. Besides, Deng's laboratory identified a new gamboge derivative compound 2 (C2). In their study, C2 treatment reduced colony formation of HNSCC CSCs and inhibited expression of CSCs markers (CD49f, CD133, and CD44) more significantly than DDP with less toxicity. The inhibition effect of C2 on CSCs was attributed to targeting Ki-67, phosphor-EGFR, CD49f, and CD133 [216]. *Cucurbitacin I* is a natural triterpenoid isolated from the *Cucurbitaceae* family plants and other plant types. Chen et al. found that *Cucurbitacin I* could inhibit the proliferation, tumor aggressiveness, and stemness signatures and induce apoptosis, differentiation, and radiosensitivity of HNSCC CSCs via suppression of STAT3, Janus-activated kinase 2 (JAK2), Bcl-2, Bcl-xL, and survivin [217].

Chang et al. screened for active components and discovered YMGKI-1 and YMGKI-2 from *Antrodia cinnamomea* Mycelia (ACM) natural products. They demonstrated that both components inhibited stemness, decreased expres-

sion of CSC markers, and promoted radiosensitivity of HNSCC CSCs by downregulating the activated autophagic signaling pathways, STAT3 and Src [218, 219].

In addition, *Plumbagin* (5-hydroxy-2-methyl-1,4-naphthoquinone, PLB) is a small molecular compound derived from the root of *Plumbago zeylanica* L, *Juglans regia*, *Juglans cinerea*, and *Juglans nigra*, with a variety of pharmacological activities. Recently Pan et al. [220] reported that PLB-induced apoptosis inhibited EMT and stemness and promoted MET via mediating multiple targets on tongue squamous cell carcinoma (TSCC) cell line SCC25 cells. In addition, translation inhibition could disrupt stemness properties [220]. SVC112 is a synthetic derivative of the cyclic hexapeptide bouvardin, a plant-derived translation elongation inhibitor with less toxicity. Keysar et al. demonstrated that SVC112 inhibits tumorsphere growth and enhances radiosensitivity in vitro by suppressing Myc, Cyclin D1, Myc, and Sox2. SVC112 alone and with radiation inhibits the growth of tumours and CSC in vivo [221]. Besides this, *Lovastatin (LV)* is a natural lipophilic statin derived from *Monascus* or *Aspergillus*-fermented rice and *Dioscorea*. In Peng et al.'s study, LV inhibited proliferation and self-renewal and induced apoptosis and cell cycle arrest of NPC CSCs. LV could also synergistically enhance the sensitivity of NPC CSCs to chemotherapy and photodynamic therapy [222]. *Tetrandrine* is a bis-benzylisoquinoline alkaloid isolated from *Stephania tetrandra* and other related species of *Menispermaceae*. Cui et al. demonstrated that tetrandrine inhibited the cell viability and proliferation of CD133 in Hep-2 cells by impacting the cell cycle and enhancing cell apoptosis via upregulating Bax and caspase-3 and downregulating Bcl-2 [223]. *Isoliquiritigenin (ISL)* is a natural flavonoid compound derived from the natural herb *licorice root (licorice)* with significant anti-tumor ability. Hu et al. showed that ISL was more cytotoxic to OSCC CSCs and hindered self-renewal by reducing ALDH1 enzymatic activity and CD44 positivity in OSCC-CSCs. ISL also enhanced sensitivity to DDP via inhibiting ABCG2. Finally, they demonstrated that the anti-CSCs ability of ISL was ascribed to regulating the protein expression of mRNA and membrane GRP78 [224]. *Sulforaphane (SF)* is an isothiocyanate isolated from *broccoli*.

Elkashty et al. found that SF-combined treatments inhibited the colony formation of HNSCC CSC and in vivo tumor progression with potential mechanisms including the stimulation of caspase-dependent apoptotic pathway, inhibition of SHH pathway, and decreased expression of SOX2 and OCT4 [225]. *Curcumin* is a bioactive polyphenolic compound identified in turmeric with significant anti-tumor ability. However, the low solubility in aqueous media, poor bioavailability, and pharmacokinetic profiles limit its therapeutic potential. Therefore, several different formulations have been produced. Recently Basak et al. found that Curcumin-difluorinated (CDF), a synthetic analog of curcumin, was packaged in liposomes and used to evaluate the growth inhibition of DDP-resistant HNSCC cell lines. Treatment with liposomal CDF resulted in a statistically significant tumor growth inhibition in nude mice xenograft and a reduction in the expression of CD44, indicating an inhibitory effect of liposomal CDF on CSCs [226]. In addition, curcumin and metformin combination could prevent 4-nitro quinoline-1-oxide (4NQO) induced oral carcinogenesis in a mice model through an overall downregulation of CSC markers [227]. *Isoorientin* (3',4',5,7-tetrahydroxy-6-C-glucopyranosyl flavone) is a C-glycosyl flavone extracted from *Aspalathus linearis* and several other plant species. In Liu et al.'s study, Isoorientin could significantly reduce the expression of p-STAT3, Wnt/ β -catenin, p-GSK3, and downstream effectors transcription factor-T cell factor 1 (TCF1) and LEF1, enhance DDP toxicity, and inhibit the tumorigenicity and growth of OSCC all in all attributing owing to targeting OSCC-SC-mediated stemness [228]. *Apigenin* (4',5,7-trihydroxyflavone) is one of the most studied phenolics abundant in fruits and vegetables. The compound could significantly down-regulated expressions of CSCs markers, CD44, NANOG, and CD105 of HNSCC cells and reduce the number of cells expressing CSCs markers under hypoxia [229]. Resveratrol (trans-3,5,4'-trihydroxystilbene) is a phytoalexin initially found in *Polygonum cuspidatum*. In Hu et al.'s study, resveratrol reduced the activity of CSCs markers (ALDH1 and CD44) and CSCs-related gene expressions (Oct4, Nanog, and Nestin) in HNC-CSC and regulated EMT-related markers in vitro and in vivo, which may lead to a valuable clinical therapeutics combining with conventional chemotherapy modalities for HNC [230].

Silibinin is a flavonolignan extracted from the fruit and seeds of *Milk thistle*. It is well-known for its hepatoprotective and anti-carcinogenic effect on various experimental cancer models. Chang et al. showed that Silibinin exerted an inhibitory influence on invasion, stemness, EMT, and anti-apoptosis ability of HNC-CSCs via activation of miR-494-inhibiting Bmi1/ADAM10 expression [231]. Recently in an in vitro study, *Propolis* could reduce CSCs numbers and decrease CSCs markers specifically [232]. The effects of the compounds mentioned above are summarized in [Table S3](#).

Immunotherapy

In addition to specific formulation, Immunotherapy targeting CSCs provides another promising perspective. Liao T et al. [233] tested responses against putative HNSCC CSCs by an alloantigen-specific model system in vitro. Although CSC populations were less sensitive to major histocompatibility complex (MHC) class I-restricted alloantigen-specific CD8⁺ CTL lysis, IFN- γ pretreatment upregulates molecules essential for antigen processing and presentation, leading to over-proportionally enhanced lysis of CSC-enriched spheroid culture-derived cells (SDC). Moreover, the subset of ALDH^{high} CSCs presented more sensitivity toward CD8⁺ CTL killing than the ALDH^{low} SDC. The in vitro experiment by Liao T et al. suggested that Immunotherapy targeting ALDH⁺ CSCs may be a promising approach. In a preclinical study, researchers induced and expanded human leukocyte antigens (HLA-A2) restricted, ALDH1A1 peptide-specific CD8⁺ T cells by *in vitro* stimulation of CD8⁺ T cells isolated from peripheral blood from regular HLA-A2⁺ donors. These HLA-A2-restricted, ALDH1A1 peptide-specific CD8⁺ T cells recognized and eliminated ALDH^{bright} cells, specifically in vitro and in vivo. They showed that the adoptive transfer of ALDH1A1-specific CD8⁺ T cells inhibited the growth of primary and metastatic tumors in xenografts [234].

Novel radiotherapy

Surgery and radiation are the mainstream in HNSCC therapy. HNSCC CSCs resistant traditional photon radiation and EGFR inhibitors; moreover, IR can activate EMT and the CSC phenotype [235]. However, in recent in vitro research, carbon ion irradiation effectively

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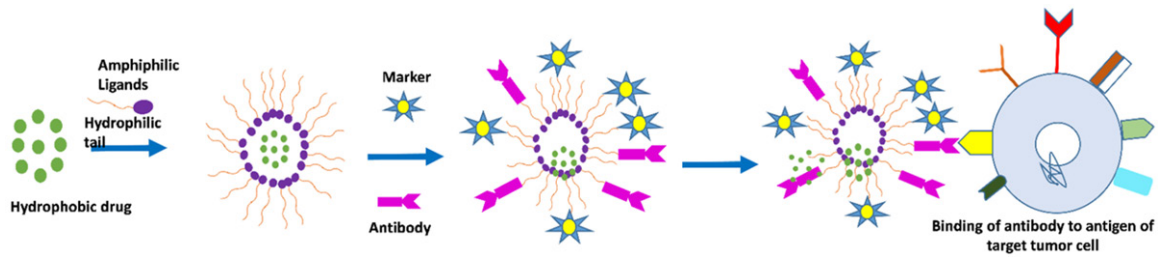


Figure 2. Overview of targeted drug delivery. The drug molecules (green) are surrounded by amphiphilic ligands with a hydrophilic tail. Attached to the tail of this amphiphilic ligand are markers and antibodies. These antibodies bind with surface-specific receptors on tumor cells in the drug and are released there to initiate apoptosis in HNC.

reduced migration/invasion of HNSCC CSCs and non-CSCs alone or combined with cetuximab [236]. More recently, a study showed that daily photobiomodulation with 3 J/cm² suppressed cellular viability and that 6 J/cm² decreased the number of spheres of OSCC cell lines and the expression of the CSC-related gene BMI1 [237]. Yu et al. demonstrated that topical 5-aminolevulinic acid-mediated photodynamic therapy (ALA-PDT) inhibited the ALDH1 activity of HNSCC cells, eliminated self-renewal capacity, CD44 positivity, stemness signatures, and enhanced chemosensitivity in HNSCC CSCs [238, 239].

Potential strategies

Given the therapeutic options above-mentioned, personalized treatment may be a good one to solve the problems. We can identify primary CSCs markers and regulators and then take adequate measures to target CSCs in individual HNSCCs. Based on individualized treatment: 1) Local CSC-targeted DDSs. It may be a promising approach to apply peritumoral injections using pure natural compounds with multi-targets and less toxicity as anti-CSCs formulations, such as nanoparticles, and in combination with radiation. Local CSC-targeted DDSs are better than conventional IVs. It could withstand the barrier of the niche, ensure anti-CSC agents arrive at the targeted CSCs, and can be taken up by CSCs with enhanced permeability and retention effect (EPR). 2) Immunotherapy targeting CSCs in individual HNSCC treatment. 3) The biology of CSCs depends on the niche; double targeting cancer stemness and the niche, such as hypoxia, microcirculation, or immune status, is a possible approach. Chinese traditional medicine (TCM) has its principle of the human environment, for instance, the Yin-Yang theory. According to the theory, illness is due to the imbalance of two opposing forces of

energy, Yin and Yang. In addition, the compounds mentioned above extracted from Chinese traditional herbs (TCH) exhibit anti-CSC effects. Pure combined compounds with fewer toxicities like Cocktail therapy, may have a promising prospect, and TCH carried by novel DDSs may be another promising strategy. In general, the anti-CSCs strategy seems a promising therapeutic option, although there is still a long way to go for successful clinical application.

Targeted drug delivery in HNSCC

Although the drug monomer presents an anti-CSCs effect, targeting CSCs requires unique drug delivery systems (DDSs) formulation, as shown in **Figure 2**. Su et al. [240] synthesized and characterized anti-CD44 antibody-coated superparamagnetic iron oxide nanoparticles (SPIONPs). The exploration of the formulation was dependent on the mechanism of hyperthermia therapy. The CD44-SPIONPs (superparamagnetic iron oxide nanoparticles) target CD44⁺ HNSCC CSCs by endocytosis, which could generate heat through magnetic vector and physical rotation under an alternating magnetic field to kill CSCs. After the AMF treatment, CD44-SPIONPs induced CSCs to undergo programmed death with an inhibitory ratio of 33.43%, significantly inhibiting the growth of grafted Cal-27 tumors in mice. In addition, Miyano et al. [241] used cyclic Arg-Gly-Asp (cRGD) peptide, an HNSCC CSCs marker specific binding to integrin $\alpha_v\beta_3$, on micellar nanomedicines incorporating cisplatin (cRGD-installed DDP/m). The cRGD-installed DDP/m showed significant antitumor activity against primary HNSCC xenograft tumors, with the rapid accumulation of the metastatic lymph nodes leading to prolonged mice survival.

RNAi therapies remain unsatisfactory due to delivery limitations by many factors, such as easy degradation by enzymes. Lo et al. [242] provided a feasible non-viral gene delivery method, cationic polyurethane-short branch polyethyleneimine (PU-PEI)-based delivery of nuclear localization signal (NLS) pre-conjugated dsDNA encoding siRNAs. In their study, co-administrated PU-PEI vehicles containing NLS-pre-conjugated dsDNA encoding either siEZH2 or siOct4 remarkably achieved gene silencing, which led to diminished CSC-like properties, suppression of EMT, enhanced radiosensitivity, and prevention of metastasis in HNSCC. Meanwhile, nano micelles could load multiple agents to target different subpopulations. Recently, Zhu et al. developed salinomycin (SAL)-loaded poly (ethylene glycol), 2000-di-stearoyl phosphatidyl-ethanolamine (DSPE-PEG)-methotrexate (MTX) nano micelles (M-SAL-MTX), for SAL targeted CD133⁺ CSCs and MTX could kill non-CSCs. In their study, M-SAL-MTX effectively accumulated in tumor tissues compared with a single treatment of SAL or MTX; therefore, M-SAL-MTX exhibits significant anti-CSCs and anti-non-CSCs in vivo [243].

In addition, peritumoral injections are available in HNSCC. Hyaluronic acid (HA) is a particular ligand for the CD44 surface receptors. Peritumoral injections of cisplatin conjugated to nanoscopic (25-100 nM) particles of HA (HA-cisplatin) provide superior antitumor efficacy and CSCs targeting compared to conventional IV cisplatin therapy in a laryngeal cancer xenograft model with less toxicity [244]. It may be useful in nanoparticles targeting CSCs markers with a specific route of administration may be a promising strategy to target CSCs.

Recent advances in cancer genetics, sequencing, and their role in therapeutic efforts have led to precision medicine. Precision medicine is mainly based on the genetic, environmental, and lifestyle characteristics that can lead to identifying the therapy for individual patients. Although this approach is very effective in oncology, some issues are still there, including drug resistance and toxicities. Drug delivery systems have enabled the modulation of pharmacological parameters, including stability, pharmacokinetics, absorption, and exposure to tumors and healthy tissues.

Nanomedicine is very helpful in targeted drug delivery, decreasing the drug toxicity to non-

target cells compared to non-carrier drugs [245]. Currently, different types of nanoparticles (NPs) have been reported and approved by US Food and Drug Administration [246] for cancer diagnosis and treatment. These are organic NPs (polymer, dendrimer, ferritin, and micelles) and inorganic (Q dots, silver iron oxide, gold). The NPs technologies have greatly improved controlled drug releases and enhanced the targeting of drug delivery to specific tissues [247-249]. These advantages of NPs drug delivery systems are improving the current treatments and paving the way for new therapy options. To cover the significant issues of medicines, including solubility and bioavailability, long time circulation, and unwanted toxicity to neighboring healthy tissues, phytomedicine integration into nano vehicle is a valuable and productive choice to enhance its biological effects and overcome the physiological barriers [250, 251].

Nanotechnology-based targeted drug delivery system for HNC therapy is alternatives treatments that maximize the efficacy and offer good efficacy to the problems compared to conventional therapies. A targeted drug delivery system reduces the rate of delivery failures and cell death and minimizes multidrug resistance. These properties of NPs are promising in HNC treatment because to reach the target, the therapeutic targets need to cross biological barriers, including the blood-brain barrier, which is a significant obstacle and reduces drug delivery to the brain [252]. To overcome the shortcomings of conventional methods in HNC management, using nanocarriers as diagnostic and therapeutic agents has improved efficacy and safety. The NPs guide anti-cancerous drugs to the target cells, increasing the concentrations of drugs in the intracellular environment of the target cells and reducing the toxicity to normal cells. They attach specific receptors to the target cells on the surfaces and are internalized by endocytosis.

Targeting through nanocarriers

Nanocarriers are the most effective system to deliver the anti-cancerous agent at the site of interest using passive or active targeting approaches. The system enhances the drug concentration in HCN target cells, increases retention, and reduces toxicity to normal cells [253, 254]. They are targeting through nanocarriers

Cancer stem cells of head and neck squamous cell carcinoma

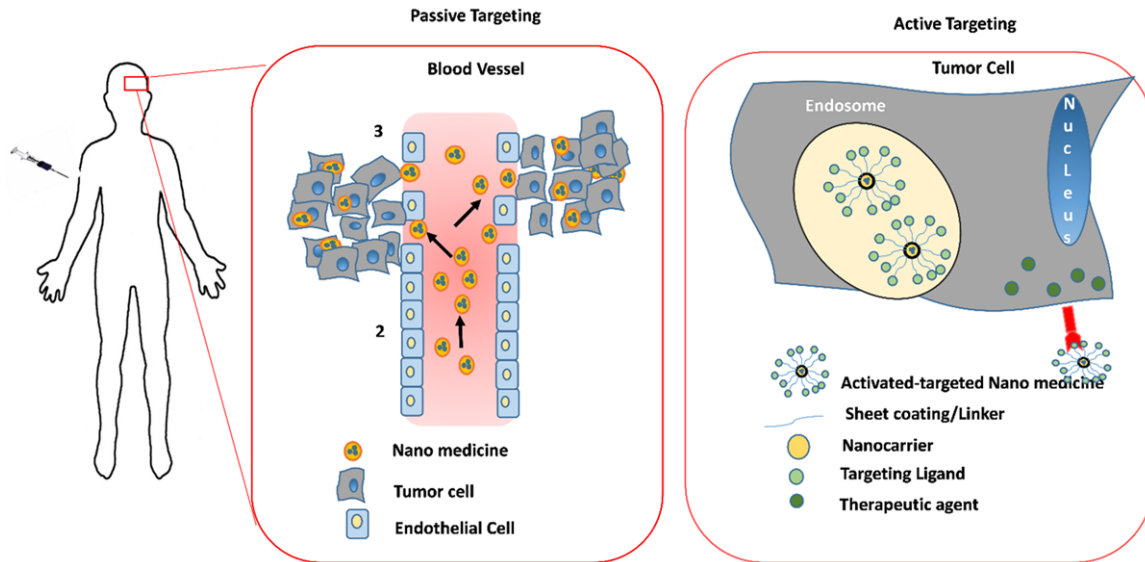


Figure 3. Flowchart mechanism of active and passive targeting.

involving active and passive approaches. Active targeting consists of using ligands/drugs to the site of interest in HNC, while passive targeting is without any stimulus or ligand to the target site.

Passive targeting

Passive targeting involves the systemic administration of nanocarriers, which tend to accumulate selectively at the desired location as a result of the enhanced permeability and retention (EPR) phenomenon [255]. The EPR may be influenced by different tumor microenvironment factors (TME), including vasculature, stage, macrophages, interstitial, and lymphatics fluid pressure [256, 257]. Thus, the anatomy and physiological conditions of the target are essential for passive targeting. The blood vessels are produced in high quantities in tumoral tissues promoting rapid growth that allows nanocarriers to be easily retained in tumor tissues of HNC [257].

Active targeting

Active targeting involves the specificity and designing of the nanocarrier to attach to the target site [255]. All NPs exhibit a good conjugation capability with target ligands, including antibodies, sugars, nucleic acids, peptides, vitamins, and other small molecules. The nanocarrier is conjugated with a molecule with a

good binding affinity to attach firmly to the target tissue [258]. Nanoparticles are attached to targeting ligands and given a reasonable degree of tumor specificity. During the tumor diagnosis/treatment, these drugs/ligands specifically bind to the NP's target and interact with the target cells receptors (which are tumor markers), and endocytosis carries the ligand inside the target cells [259]. In active targeting of HNC, the target cells must have high expressed markers compared to healthy tissues. A flowchart of active and passive targeting is provided in **Figure 3**.

Challenges in active targeting

The critical challenge is selecting a targeting agent to minimize the toxicity to surrounding healthy tissues. Physical/triggered targeting is also a type of active targeting that depends on the usage of internal (pH, enzymes, redox potential) or external (temperature, UV light, ultrasound) stimuli to potentiate the nanocarrier to the site of interest for releasing the drugs molecules [260, 261]. In one of its kind, the magnetic nanocarriers are driven to the target site using external stimuli of the magnetic field.

Nanocarriers for drug delivery

In this segment, our attention is directed towards the most auspicious nanocarriers intended for drug delivery in the treatment of

Cancer stem cells of head and neck squamous cell carcinoma

head and neck cancer (HNC). These include lipid-based, polymer-based, and metallic-based nanocarriers.

Lipid-based nanocarriers

In cancer-targeted drug delivery systems, lipid-based nanocarriers are commonly phospholipids possessing unique properties and self-organization in an aqueous environment to form organized shapes and structures. Lipids may form liposomes, micelles, or bilayers. Micelles and liposomes are the two most commonly used for drug delivery.

Micelles have a hydrophobic core and hydrocarbonated tails surrounded by polar heads. Micelles are formed by amphipathic molecules containing a polar group and a tail with only one hydrocarbon [262, 263]. Concerning HNSCC tumor studies, it has been reported that microRNA-107 is downregulated compared to healthy cells. To deliver the pre-miR-107 successfully, a cationic lipid nanoparticle was developed, consisting of DDAB (dimethyl octadecyl ammonium bromide), cholesterol, and α -Tocopheryl polyethylene glycol 1000 succinate [264].

Liposomes have been used extensively as nanocarriers in cancer therapy. Each liposome consists of a phospholipid bilayer and an aqueous inner cavity, which can encapsulate different types of polar molecules [265, 266]. The nano-formulations are in clinical trials; some have been marketed to treat HNC. Liposomes are an ideal system of nanoencapsulation to carry drugs, overcome pharmacokinetics problems, stability in vivo, and toxicity to healthy tissues. One important example is the encapsulation of curcumin in liposomes.

Curcumin is a component of *Curcuma longa*, possessing the potential property of antibiotic, anti-inflammatory, and antioxidant agent, and it also demonstrated good anticancer activity [267].

Polymer-based nanocarriers

These nanocarriers can be produced from synthetic or natural polymers [268, 269], which are biocompatible, stable, possess toxicity, have no side effects, and are entirely metabolized in the human body. In a previous study,

multifunctional polymer-based nanoparticles (Linear dendritic mPEG-BMA4) for targeted delivery of saracatinib (kinase inhibitor) into HNC cells in vivo. Compared with free drugs, the polymeric nanoparticles loaded with saracatinib demonstrated good anticancer activity [270]. Chen et al. reported an injectable, biodegradable polymer, cisplatin, for the human HNSCC treatment. This polymer exhibited a well released (80%) of cisplatin and was significantly involved in tumor suppression compared to free cisplatin [271].

Folate-targeted treatment with methotrexate (MTX) is commonly considered in HNSCC; however, its severe side effects are very severe [272]. Dendrimer-targeted delivery can minimize toxicity and enhance drug efficacy. Ward et al. used cell lines with null, intermediate, and high expression folate receptors and evaluated in vivo efficacy of G5 poly-amidoamine dendrimer-based targeted treatment. The targeted system was more effective against cell lines with high folate receptor expression with increased effectiveness compared to free MTX and control [273].

Metallic nano-carrier

Metallic-based nanocarriers were also found effective in HNC therapy. Zhang et al. used superparamagnetic nanoparticles as a novel targeted drug delivery system for HNCs therapy. A biocompatible mesoporous Fe_3O_4 NPs attached with superparamagnetic polyacrylic acid was developed. These mesoporous Fe_3O_4 NPs delivered bleomycin to the tumor tissue, starting its slow release and the tumor cells apoptosis. The drug also showed significantly reduced side effects of bleomycin to healthy cells. This new approach provided potential applications of mesoporous Fe_3O_4 NPs in HNC treatment using simple technologies, fewer side effects, and more efficacy [274].

Applications of quantum dots (QDs) in HNC

The current applications of magnetic resonance imaging (MRI), X-ray, ultrasound, and radionuclide imaging, to detect and diagnose tumors have limitations. These techniques are less sensitive in detecting malignant cells when small in number, unable to detect biomarkers specific to cancer cells' surface and exhibit hazardous effects to different levels. Thus, in

investigating advanced and novel approaches with fewer dangerous effects and high sensitivity, specificity is of prime importance and urgently required.

QDs, also known as “artificial atoms”, is today’s most attractive topic in nanobiology. Several researchers are interested in using QDs in cancer diagnostics [275, 276] because; they have excellent resistance to photo-bleaching; secondly, they have good optical properties with superior fluorescence intensity; thirdly, QDs possess multiple fluorescent colors emission under single-source excitation and size-tunable light emission. Furthermore, during the synthesis process, the wavelengths emitted could be tuned and controlled precisely by size and shape. This property is beneficial in performing nanometer resolution and co-localization of multicolor QDs using confocal microscopy. This is also important in reducing the slices of tissue that must be cut for biomarker observation [275-277].

Application of dendrimer nanoparticles in HNC

Dendrimers are synthetic polymers playing an important role in drug discovery and carrier systems [278]. They are “smart” nanocarriers in medicine with multifunctional that can be used in targeted drug delivery of one or more agents selectively to tumor cells with more safety and also to intracellular gene-specific targeting [279, 280]. Dendrimers with nano polymeric designs have been considered a highly specific class delivery system for drugs and genes [281, 282]. Over the past decade, gene therapy has been used in clinical trials. Although there are some drug and gene delivery concepts, including liposomes, viral vectors, cationic polymers, gold, and magnetic nanoparticles [283], however, dendrimers are the most attractive nowadays for their good safety and specificity to the target site [284]. Dendrimers which are 1-100 nm in size, are globular macromolecules consisting of a central core domain, a hyper-branched mantle domain, and a domain of corona with exterior reactive functional groups [285]. These are perfect (spherical) molecules as nanocarriers with specific properties for cell-specific targeting. Dendrimers may be of different kinds, including melamine, poly(propylene imine) (PPI), poly-amidoamine (PAMAM), poly(ethylene glycol), poly(glycerol-co-succinic

acid), poly-l-lysine (PLL), triazine, poly(glycerol), poly[2,2-bis(hydroxymethyl) propionic acid], (PEG), and citric acid-based ones [286, 287]. PAMAM and PPI vectors have been extensively examined for medical use [288, 289]. Both have amine-terminated end and pH-dependent drug release properties, making them most suitable for HNC treatment. The dendrimer’s ‘back folding’ or collapse on itself is the most attractive property of dendrimers due to the tertiary amine groups deprotonated at elevated pH [288]. The dendrimer scan traverse several barriers using active and passive tumor targeting.

Recently a poly-amidoamine generation 4 (G4) dendrimer and fluorescently labeled for gene delivery and folic acid-decorated conjugates in HNSCC-targeted have been reported [290]. The G4 dendrimer delivery system is conjugated with folic acid (FA) and has the properties of targeting the moiety of HNSCC. In HNSCC cells, complexing this G4 dendrimer with siRNA or plasmid significantly increases the knock-down system’s gene transfection or efficiency. In HNSCC, the G4-FA vector exhibited excellent tumor targeting capability, biocompatibility, sustained retention, and high uptake in a gene therapy approach.

Dendrimers have been synthetically engineered with nanodevices in nanocarrier drug delivery systems. The terminal moieties are responsible for dendrimers’ biological effect and global efficiency. Dendrimers in classical drugs overcome the physicochemical limitations, including solubility, stability, specificity, biodistribution, and therapeutic efficiency, **Figure 4**. They have the property to reach the right targets by immune clearance, penetration into cells, and interactions in off-target [291]. All the dendrimers have the desired properties of drug retention, release of the therapeutic agent, unaffacting by the immune system, blood circulation time enhancing, and cells or organs specific targeting [292]. An overview is provided in **Figure 5**.

Different intervention plans in HNC patients

Significant physical and psychological morbidity have been experienced by the HNC patients during radiotherapy (XRT) that not only resulted in the interruption of the treatment, but also quality of life. Intensive radiotherapy (XRT) is carried out in these patients either alone or in

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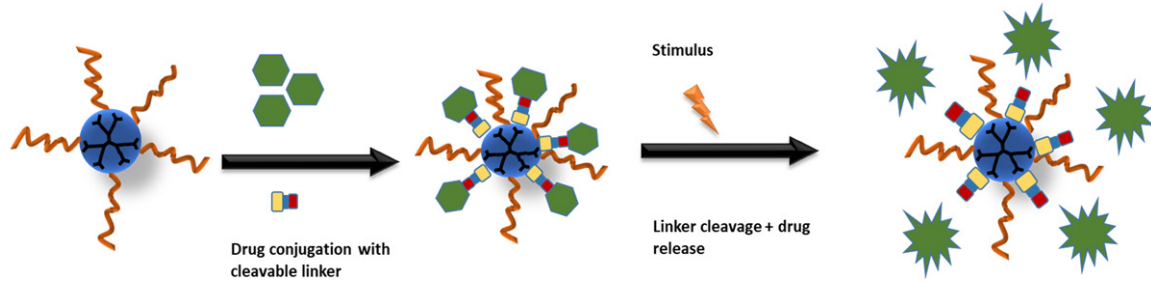


Figure 4. Synthesis of dendrimers and drug conjugation.

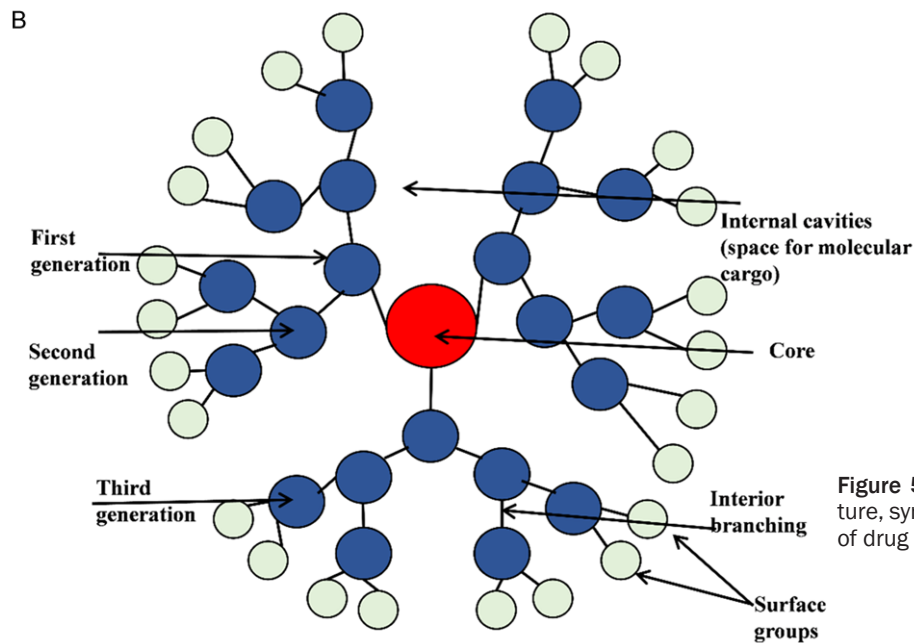
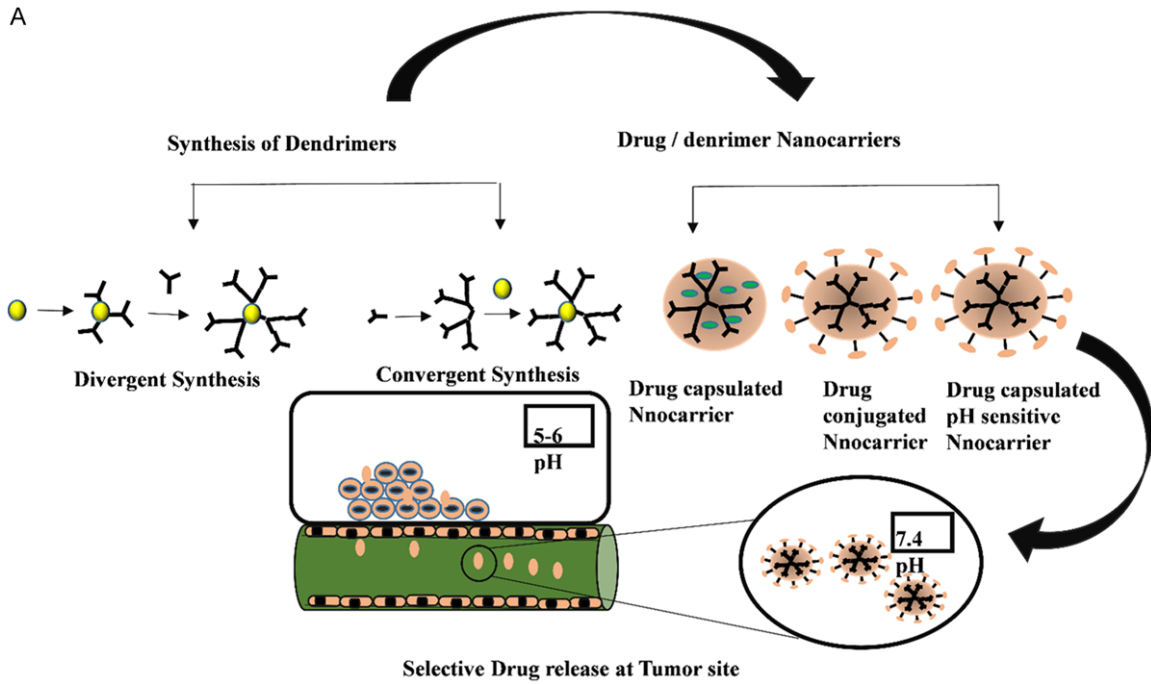


Figure 5. A. Dendrimer's structure, synthesis, and mechanism of drug release. B. Dendrimer.

Cancer stem cells of head and neck squamous cell carcinoma

combination of other treatments [293]. Different interventions have been applied for cancers prevention and control. However, for HNC survivors the interventions so far, may not address the general health and cancer related needs. A study developed a couple-based intervention called “Spouses coping with the Head and neck Radiation Experience” abbreviated as SHARE, was delivered through phone. This intervention supports psychoeducation that encourages self-management and teaches strategies to improve teamwork and coping. The study evaluated couples on self-management and coordination of care and support at the start of XRT to control/alleviate symptom burden (physical and psychological) and improve both partner’s adjustment. The results of the study supported the feasibility, acceptability, and preliminary efficacy of SHARE [294]. The successful treatment of HNCs can be based on the TNM (tumor, node, metastasis) staging system. A study based on Polish patients found concordance between clinical and pathological T and N stages in patients with HNCs. It was found that there is a moderate agreement between the clinical and pathological stages for stage T, while substantial agreement was found for stage N [295].

Focused on improving quality of life (QOL) and/or mood in HNC patients, a team of researchers reviewed the available literature and targeted the types of interventions such as educational, psychosocial, physical, and psychological symptom management, mindfulness, pharmacologic, exercise, and telemedicine. Preliminary feasibility and acceptability with some positive impacts on QOL and/or mood were found in HNC patients [296]. PRO-ACTIVE trial intervention in HNC patients suggested useful modifications in telehealth [297].

A more recent study investigation the impact of cognitive behavioral intervention on HNC and treatment on eating and talking, and health-related quality of life of survivorship [298]. This study reports that impact treatment can be particularly distressing and markedly changed activities among survivors. Engaging in therapeutic approaches to manage distress in treatment time may influence quality of life and mood of survivorship phase.

Another study applied Navigation for Disparities and Untimely Radiation Therapy, a multi-

level intervention to evaluate feasibility, preliminary efficacy, and acceptability in HNC. They found that the potential postoperative radiation therapy has been improved [299].

A previous study also applied the telehealth intervention in HNC treatment. It was observed that the telehealth intervention is also feasible and acceptable for better management of treatment for HNC [300].

Exercise has also been regarded as a potential intervention in prevention of different types of cancer. During the investigation regular exercise was also found to be a promising intervention in HNC patients. The patients were active participants in a six-week supervised exercise intervention in HNC treatment [301].

Conclusion

HNC has been ranked sixth among the most common cancer worldwide, and its occurrence is still increasing in the future. Due to the failure of HNC treatment, there is an urgent need to design innovative techniques for better management of HNC. Using the advanced approaches of NPs, drug concentration may be increased at the target site. There are several studies available who have evaluated HNSCC patients’ tumors or tissues, but the outcomes of the clinical data are not satisfactory. To establish clinically significant CSC markers in the head and neck regions, the primary barrier is that this region is secondary to the convention of amassing malignancies from different upper aerodigestive regions with diverse embryological and biological features. As a result, there is little definitive data about clinical implications of CSCs within HNSCC, the primary exception being prognostic value. Another reason might be due to no single biomarker for CSCs in HNSCC is available. In this context, nanomedicine emerged as an alternative and potential approach to using nanocarriers, which the body’s immune system could not sense. NPs have the potential to improve treatment efficiency without harming normal cells. Moreover, nanocarriers are essential to combat tumor resistance in various targeting strategies. Nanotechnology may shift the management of HNC through theragnostic approaches, simultaneously allowing diagnosis and therapy. QDs in HNC diagnostics may be useful as they have good optical properties and superior fluo-

rescence intensity. Very limited information is available about the applications of dendrimers in HNC therapy. The contribution of dendrimers for targeted drug delivery in HNC may be more useful. However, for better management of HNC, the NPs' biocompatibility, toxicity, and long-term implications need further trials and understanding before it is applied on a large scale.

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Disclosure of conflict of interest

None.

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References

- [1] Chow LQM. Head and neck cancer. *N Engl J Med* 2020; 382: 60-72.
- [2] Qian X, Nie X, Wollenberg B, Sudhoff H, Kaufmann AM and Albers AE. Heterogeneity of head and neck squamous cell carcinoma stem cells. *Adv Exp Med Biol* 2019; 1139: 23-40.
- [3] Heft Neal ME, Brenner JC, Prince MEP and Chinn SB. Advancement in cancer stem cell biology and precision medicine-review article head and neck cancer stem cell plasticity and the tumor microenvironment. *Front Cell Dev Biol* 2022; 9: 660210.
- [4] Johnson DE, Burtness B, Leemans CR, Lui VWY, Bauman JE and Grandis JR. Head and neck squamous cell carcinoma. *Nat Rev Dis Primers* 2020; 6: 92.
- [5] Sun Z, Sun X, Chen Z, Du J and Wu Y. Head and neck squamous cell carcinoma: risk factors, molecular alterations, immunology and peptide vaccines. *Int J Pept Res Ther* 2022; 28: 19.
- [6] Dong L, Sun Q, Song F, Song X, Lu C, Li Y and Song X. Identification and verification of eight cancer-associated fibroblasts related genes as a prognostic signature for head and neck squamous cell carcinoma. *Heliyon* 2023; 9: e14003.
- [7] Solomon B, Young RJ and Rischin D. Head and neck squamous cell carcinoma: genomics and emerging biomarkers for immunomodulatory cancer treatments. *Semin Cancer Biol* 2018; 52: 228-240.
- [8] Chen SMY, Krinsky AL, Woolaver RA, Wang X, Chen Z and Wang JH. Tumor immune microenvironment in head and neck cancers. *Mol Carcinog* 2020; 59: 766-774.
- [9] Leemans CR, Braakhuis BJ and Brakenhoff RH. The molecular biology of head and neck cancer. *Nat Rev Cancer* 2011; 11: 9-22.
- [10] Chen D and Wang CY. Targeting cancer stem cells in squamous cell carcinoma. *Precis Clin Med* 2019; 2: 152-165.
- [11] Patel JJ, Levy DA, Nguyen SA, Knochelmann HM and Day TA. Impact of PD-L1 expression and human papillomavirus status in anti-PD1/PDL1 immunotherapy for head and neck squamous cell carcinoma-systematic review and meta-analysis. *Head Neck* 2020; 42: 774-786.
- [12] Chi AC, Day TA and Neville BW. Oral cavity and oropharyngeal squamous cell carcinoma-an update. *CA Cancer J Clin* 2015; 65: 401-421.
- [13] Baniebrahimi G, Mir F and Khanmohammadi R. Cancer stem cells and oral cancer: insights into molecular mechanisms and therapeutic approaches. *Cancer Cell Int* 2020; 20: 113.
- [14] Mesía R, Henke M, Fortin A, Minn H, Yunes Ancona AC, Cmelak A, Markowitz AB, Hotte SJ, Singh S, Chan AT, Merlano MC, Skladowski K, Zhang A, Oliner KS, VanderWalde A and Giralt J. Chemoradiotherapy with or without panitumumab in patients with unresected, locally advanced squamous-cell carcinoma of the head and neck (CONCERT-1): a randomised, controlled, open-label phase 2 trial. *Lancet Oncol* 2015; 16: 208-220.
- [15] Bonner JA, Harari PM, Giralt J, Cohen RB, Jones CU, Sur RK, Raben D, Baselga J, Spencer SA, Zhu J, Youssoufian H, Rowinsky EK and Ang KK. Radiotherapy plus cetuximab for locoregionally advanced head and neck cancer: 5-year survival data from a phase 3 randomised trial, and relation between cetuximab-induced rash and survival. *Lancet Oncol* 2010; 11: 21-28.
- [16] Gomez KE, Wu F, Keysar SB, Morton JJ, Miller B, Chimed TS, Le PN, Nieto C, Chowdhury FN, Tyagi A, Lyons TR, Young CD, Zhou H, Somerset HL, Wang XJ and Jimeno A. Cancer cell CD44 mediates macrophage/monocyte-driven regulation of head and neck cancer stem cells. *Cancer Res* 2020; 80: 4185-4198.
- [17] Tu SM, Moran C, Norton W and Zacharias NM. Stem cell theory of cancer: origin of metastasis and sub-clonality. *Semin Diagn Pathol* 2023; 40: 63-68.

Cancer stem cells of head and neck squamous cell carcinoma

- [18] Kaushik V, Kulkarni Y, Felix K, Azad N, Iyer AKV and Yakisich JS. Alternative models of cancer stem cells: the stemness phenotype model, 10 years later. *World J Stem Cells* 2021; 13: 934-943.
- [19] Lambert AW, Pattabiraman DR and Weinberg RA. Emerging biological principles of metastasis. *Cell* 2017; 168: 670-691.
- [20] Kumazoe M, Takai M, Bae J, Hiroi S, Huang Y, Takamatsu K, Won Y, Yamashita M, Hidaka S, Yamashita S, Yamada S, Murata M, Tsukamoto S and Tachibana H. FOXO3 is essential for CD44 expression in pancreatic cancer cells. *Oncogene* 2017; 36: 2643-2654.
- [21] Aramini B, Masciale V, Arienti C, Dominici M, Stella F, Martinelli G and Fabbri F. Cancer stem cells (CSCs), circulating tumor cells (CTCs) and Their interplay with cancer associated fibroblasts (CAFs): a new world of targets and treatments. *Cancers (Basel)* 2022; 14: 2408.
- [22] Yu SS and Cirillo N. The molecular markers of cancer stem cells in head and neck tumors. *J Cell Physiol* 2020; 235: 65-73.
- [23] Yang D, Liu J, Qian H and Zhuang Q. Cancer-associated fibroblasts: from basic science to anticancer therapy. *Exp Mol Med* 2023; 55: 1322-1332.
- [24] Wolmarans E, Boy SC, Nel S, Mercier AE and Pepper MS. Cancer stem cells in head and neck carcinomas: identification and possible therapeutic implications. *Adv Exp Med Biol* 2018; 1083: 89-102.
- [25] Azizi E, Nagrath S, Kozminsky M and Wicha MS. Cancer stem cells and circulating tumor cells: molecular markers, isolation techniques, and clinical implications. *Circulating Tumor Cells*. Springer; 2016. pp. 75-97.
- [26] He Y, Sun MM, Zhang GG, Yang J, Chen KS, Xu WW and Li B. Targeting PI3K/Akt signal transduction for cancer therapy. *Signal Transduct Target Ther* 2021; 6: 425.
- [27] Jiang AM, Ren MD, Liu N, Gao H, Wang JJ, Zheng XQ, Fu X, Liang X, Ruan ZP, Tian T and Yao Y. Tumor mutation burden, immune cell infiltration, and construction of immune-related genes prognostic model in head and neck cancer. *Int J Med Sci* 2021; 18: 226-238.
- [28] Keysar SB, Le PN, Miller B, Jackson BC, Eagles JR, Nieto C, Kim J, Tang B, Glogowska MJ, Morton JJ, Padilla-Just N, Gomez K, Warnock E, Reisinger J, Arcaroli JJ, Messersmith WA, Wakefield LM, Gao D, Tan AC, Serracino H, Vasiliou V, Roop DR, Wang XJ and Jimeno A. Regulation of head and neck squamous cancer stem cells by PI3K and SOX2. *J Natl Cancer Inst* 2016; 109: djw189.
- [29] Yang C, Zhang Y, Zhang Y, Zhang Z, Peng J, Li Z, Han L, You Q, Chen X, Rao X, Zhu Y and Liao Z. Downregulation of cancer stem cell properties via mTOR signaling pathway inhibition by rapamycin in nasopharyngeal carcinoma. *Int J Oncol* 2015; 47: 909-917.
- [30] Bourguignon LYW, Earle C and Shiina M. Activation of matrix hyaluronan-mediated CD44 signaling, epigenetic regulation and chemoresistance in head and neck cancer stem cells. *Int J Mol Sci* 2017; 18: 1849.
- [31] Peng S, Wu C, Sun W, Liu D, Luo M, Su B, Zhang L, Mei Q and Hu G. Snail-mediated cancer stem cell-like phenotype in human CNE2 nasopharyngeal carcinoma cell. *Head Neck* 2018; 40: 485-497.
- [32] Hufbauer M, Maltseva M, Meinrath J, Lechner A, Beutner D, Huebbers CU and Akgul B. HPV16 increases the number of migratory cancer stem cells and modulates their miRNA expression profile in oropharyngeal cancer. *Int J Cancer* 2018; 143: 1426-1439.
- [33] Ren L, Deng B, Saloura V, Park JH and Nakamura Y. MELK inhibition targets cancer stem cells through downregulation of SOX2 expression in head and neck cancer cells. *Oncol Rep* 2019; 41: 2540-2548.
- [34] Siljee S, Buchanan O, Brasch HD, Bockett N, Patel J, Paterson E, Purdie GL, Davis PF, Itintang T and Tan ST. Cancer stem cells in metastatic head and neck cutaneous squamous cell carcinoma express components of the renin-angiotensin system. *Cells* 2021; 10: 243.
- [35] Bian S, Wang Z, Chen Y and Li R. SPLUNC1 and MLL3 regulate cancer stem cells in nasopharyngeal carcinoma. *J BUON* 2019; 24: 1700-1705.
- [36] Ji J, Yu Y, Li ZL, Chen MY, Deng R, Huang X, Wang GF, Zhang MX, Yang Q, Ravichandran S, Feng GK, Xu XL, Yang CL, Qiu MZ, Jiao L, Yang D and Zhu XF. XIAP limits autophagic degradation of Sox2 and is a therapeutic target in nasopharyngeal carcinoma stem cells. *Theranostics* 2018; 8: 1494-1510.
- [37] Chen D, Wu M, Li Y, Chang I, Yuan Q, Ekimyan-Salvo M, Deng P, Yu B, Yu Y, Dong J, Szymanski JM, Ramadoss S, Li J and Wang CY. Targeting BMI1(+) cancer stem cells overcomes chemoresistance and inhibits metastases in squamous cell carcinoma. *Cell Stem Cell* 2017; 20: 621-634, e626.
- [38] Roy S, Roy S, Kar M, Padhi S, Saha A, Anuja K and Banerjee B. Role of p38 MAPK in disease relapse and therapeutic resistance by maintenance of cancer stem cells in head and neck squamous cell carcinoma. *J Oral Pathol Med* 2018; 47: 492-501.
- [39] Lee SH, Koo BS, Kim JM, Huang S, Rho YS, Bae WJ, Kang HJ, Kim YS, Moon JH and Lim YC. Wnt/beta-catenin signalling maintains self-renewal and tumorigenicity of head and neck squamous cell carcinoma stem-like cells by activating Oct4. *J Pathol* 2014; 234: 99-107.

Cancer stem cells of head and neck squamous cell carcinoma

- [40] Xie SL, Fan S, Zhang SY, Chen WX, Li QX, Pan GK, Zhang HQ, Wang WW, Weng B, Zhang Z, Li JS and Lin ZY. SOX8 regulates cancer stem-like properties and cisplatin-induced EMT in tongue squamous cell carcinoma by acting on the Wnt/beta-catenin pathway. *Int J Cancer* 2018; 142: 1252-1265.
- [41] Lee SH, Oh SY, Do SI, Lee HJ, Kang HJ, Rho YS, Bae WJ and Lim YC. SOX2 regulates self-renewal and tumorigenicity of stem-like cells of head and neck squamous cell carcinoma. *Br J Cancer* 2014; 111: 2122-2130.
- [42] Zhong Q, Liu ZH, Lin ZR, Hu ZD, Yuan L, Liu YM, Zhou AJ, Xu LH, Hu LJ, Wang ZF, Guan XY, Hao JJ, Lui VWY, Guo L, Mai HQ, Chen MY, Han F, Xia YF, Grandis JR, Zhang X and Zeng MS. The RARS-MAD1L1 fusion gene induces cancer stem cell-like properties and therapeutic resistance in nasopharyngeal carcinoma. *Clin Cancer Res* 2018; 24: 659-673.
- [43] Chien CS, Wang ML, Chu PY, Chang YL, Liu WH, Yu CC, Lan YT, Huang PI, Lee YY, Chen YW, Lo WL and Chiou SH. Lin28B/Let-7 regulates expression of Oct4 and Sox2 and reprograms oral squamous cell carcinoma cells to a stem-like state. *Cancer Res* 2015; 75: 2553-2565.
- [44] Subramanian C, Kovatch KJ, Sim MW, Wang G, Prince ME, Carey TE, Davis R, Blagg BSJ and Cohen MS. Novel C-terminal heat shock protein 90 inhibitors (KU711 and Ku757) are effective in targeting head and neck squamous cell carcinoma cancer stem cells. *Neoplasia* 2017; 19: 1003-1011.
- [45] Kerk SA, Finkel KA, Pearson AT, Warner KA, Zhang Z, Nor F, Wagner VP, Vargas PA, Wicha MS, Hurt EM, Hollingsworth RE, Tice DA and Nor JE. 5T4-targeted therapy ablates cancer stem cells and prevents recurrence of head and neck squamous cell carcinoma. *Clin Cancer Res* 2017; 23: 2516-2527.
- [46] Lim YC, Kang HJ and Moon JH. C-Met pathway promotes self-renewal and tumorigenicity of head and neck squamous cell carcinoma stem-like cell. *Oral Oncol* 2014; 50: 633-639.
- [47] Sun S, Liu S, Duan SZ, Zhang L, Zhou H, Hu Y, Zhou X, Shi C, Zhou R and Zhang Z. Targeting the c-Met/FZD8 signaling axis eliminates patient-derived cancer stem-like cells in head and neck squamous carcinomas. *Cancer Res* 2014; 74: 7546-7559.
- [48] Evran E, Sahin H, Akbas K, Cigdem S and Gunduz E. Investigation of MACC1 gene expression in head and neck cancer and cancer stem cells. *Clin Invest Med* 2016; 39: 27506.
- [49] Li J, Li Z, Wu Y, Wang Y, Wang D, Zhang W, Yuan H, Ye J, Song X, Yang J, Jiang H and Cheng J. The Hippo effector TAZ promotes cancer stemness by transcriptional activation of SOX2 in head neck squamous cell carcinoma. *Cell Death Dis* 2019; 10: 603.
- [50] Nathansen J, Lukiyanchuk V, Hein L, Stolte MI, Borgmann K, Lock S, Kurth I, Baumann M, Krause M, Linge A and Dubrovskaya A. Oct4 confers stemness and radioresistance to head and neck squamous cell carcinoma by regulating the homologous recombination factors PSMC3IP and RAD54L. *Oncogene* 2021; 40: 4214-4228.
- [51] Jiang P, Xu C, Zhou M, Zhou H, Dong W, Wu X, Chen A and Feng Q. RXRalpha-enriched cancer stem cell-like properties triggered by CDDP in head and neck squamous cell carcinoma (HNSCC). *Carcinogenesis* 2018; 39: 252-262.
- [52] Leong HS, Chong FT, Sew PH, Lau DP, Wong BH, Teh BT, Tan DS and Iyer NG. Targeting cancer stem cell plasticity through modulation of epidermal growth factor and insulin-like growth factor receptor signaling in head and neck squamous cell cancer. *Stem Cells Transl Med* 2014; 3: 1055-1065.
- [53] Lv XX, Zheng XY, Yu JJ, Ma HR, Hua C and Gao RT. EGFR enhances the stemness and progression of oral cancer through inhibiting autophagic degradation of SOX2. *Cancer Med* 2020; 9: 1131-1140.
- [54] Zhao ZL, Zhang L, Huang CF, Ma SR, Bu LL, Liu JF, Yu GT, Liu B, Gutkind JS, Kulkarni AB, Zhang WF and Sun ZJ. NOTCH1 inhibition enhances the efficacy of conventional chemotherapeutic agents by targeting head neck cancer stem cell. *Sci Rep* 2016; 6: 24704.
- [55] Upadhyay P, Nair S, Kaur E, Aich J, Dani P, Sethunath V, Gardi N, Chandrani P, Godbole M, Sonawane K, Prasad R, Kannan S, Agarwal B, Kane S, Gupta S, Dutt S and Dutt A. Notch pathway activation is essential for maintenance of stem-like cells in early tongue cancer. *Oncotarget* 2016; 7: 50437-50449.
- [56] Bourguignon LY, Wong G and Shiina M. Up-regulation of histone methyltransferase, DOT1L, by matrix hyaluronan promotes microRNA-10 expression leading to tumor cell invasion and chemoresistance in cancer stem cells from head and neck squamous cell carcinoma. *J Biol Chem* 2016; 291: 10571-10585.
- [57] Huang CF, Chen L, Li YC, Wu L, Yu GT, Zhang WF and Sun ZJ. NLRP3 inflammasome activation promotes inflammation-induced carcinogenesis in head and neck squamous cell carcinoma. *J Exp Clin Cancer Res* 2017; 36: 116.
- [58] Chen L, Li YC, Wu L, Yu GT, Zhang WF, Huang CF and Sun ZJ. TRAF6 regulates tumour metastasis through EMT and CSC phenotypes in head and neck squamous cell carcinoma. *J Cell Mol Med* 2018; 22: 1337-1349.
- [59] Chiu CC, Lee LY, Li YC, Chen YJ, Lu YC, Li YL, Wang HM, Chang JT and Cheng AJ. Grp78 as a therapeutic target for refractory head-neck cancer with CD24(-)CD44(+) stemness phenotype. *Cancer Gene Ther* 2013; 20: 606-615.

Cancer stem cells of head and neck squamous cell carcinoma

- [60] Cheng LH, Hung KF, Huang TF, Hsieh HP, Wang SY, Huang CY and Lo JF. Attenuation of cancer-initiating cells stemness properties by abrogating S100A4 calcium binding ability in head and neck cancers. *Oncotarget* 2016; 7: 78946-78957.
- [61] slam M, Sharma S and Teknos TN. RhoC regulates cancer stem cells in head and neck squamous cell carcinoma by overexpressing IL-6 and phosphorylation of STAT3. *PLoS One* 2014; 9: e88527.
- [62] Shigeishi H, Biddle A, Gammon L, Emich H, Rodini CO, Gemenetzidis E, Fazil B, Sugiyama M, Kamata N and Mackenzie IC. Maintenance of stem cell self-renewal in head and neck cancers requires actions of GSK3beta influenced by CD44 and RHAMM. *Stem Cells* 2013; 31: 2073-2083.
- [63] Muhammad N, Bhattacharya S, Steele R, Phillips N and Ray RB. Involvement of c-Fos in the promotion of cancer stem-like cell properties in head and neck squamous cell carcinoma. *Clin Cancer Res* 2017; 23: 3120-3128.
- [64] Liu S, Ye D, Guo W, Yu W, He Y, Hu J, Wang Y, Zhang L, Liao Y, Song H, Zhong S, Xu D, Yin H, Sun B, Wang X, Liu J, Wu Y, Zhou BP, Zhang Z and Deng J. G9a is essential for EMT-mediated metastasis and maintenance of cancer stem cell-like characters in head and neck squamous cell carcinoma. *Oncotarget* 2015; 6: 6887-6901.
- [65] Giudice FS, Pinto DS Jr, Nor JE, Squarize CH and Castilho RM. Inhibition of histone deacetylase impacts cancer stem cells and induces epithelial-mesenchyme transition of head and neck cancer. *PLoS One* 2013; 8: e58672.
- [66] Lagunas AM, Francis M, Maniar NB, Nikolova G, Wu J and Crowe DL. Paracrine interaction of cancer stem cell populations is regulated by the senescence-associated secretory phenotype (SASP). *Mol Cancer Res* 2019; 17: 1480-1492.
- [67] Yu C, Chen F, Wang X, Cai Z, Yang M, Zhong Q, Feng J, Li J, Shen C and Wen Z. Pin2 telomeric repeat factor 1-interacting telomerase inhibitor 1 (PinX1) inhibits nasopharyngeal cancer cell stemness: implication for cancer progression and therapeutic targeting. *J Exp Clin Cancer Res* 2020; 39: 31.
- [68] Czerwinski MJ, Desiderio V, Shkeir O, Papagerakis P, Lapadatescu MC, Owen JH, Athanasiou-Papaefthymiou M, Zheng L, Papaccio G, Prince ME and Papagerakis S. In vitro evaluation of sialyl Lewis X relationship with head and neck cancer stem cells. *Otolaryngol Head Neck Surg* 2013; 149: 97-104.
- [69] Desiderio V, Papagerakis P, Tirino V, Zheng L, Matossian M, Prince ME, Paino F, Mele L, Papaccio F, Montella R, Papaccio G and Papagerakis S. Increased fucosylation has a pivotal role in invasive and metastatic properties of head and neck cancer stem cells. *Oncotarget* 2015; 6: 71-84.
- [70] Jung YS, Vermeer PD, Vermeer DW, Lee SJ, Goh AR, Ahn HJ and Lee JH. CD200: association with cancer stem cell features and response to chemoradiation in head and neck squamous cell carcinoma. *Head Neck* 2015; 37: 327-335.
- [71] Lu H, Yan C, Quan XX, Yang X, Zhang J, Bian Y, Chen Z and Van Waes C. CK2 phosphorylates and inhibits TAp73 tumor suppressor function to promote expression of cancer stem cell genes and phenotype in head and neck cancer. *Neoplasia* 2014; 16: 789-800.
- [72] Fukusumi T, Ishii H, Konno M, Yasui T, Nakahara S, Takenaka Y, Yamamoto Y, Nishikawa S, Kano Y, Ogawa H, Hasegawa S, Hamabe A, Haraguchi N, Doki Y, Mori M and Inohara H. CD10 as a novel marker of therapeutic resistance and cancer stem cells in head and neck squamous cell carcinoma. *Br J Cancer* 2014; 111: 506-514.
- [73] Faber A, Goessler UR, Hoermann K, Schultz JD, Umbreit C and Stern-Straeter J. SDF-1-CXCR4 axis: cell trafficking in the cancer stem cell niche of head and neck squamous cell carcinoma. *Oncol Rep* 2013; 29: 2325-2331.
- [74] Khammanivong A, Gopalakrishnan R and Dickerson EB. SMURF1 silencing diminishes a CD44-high cancer stem cell-like population in head and neck squamous cell carcinoma. *Mol Cancer* 2014; 13: 260.
- [75] Sheng X, Li Y, Li Y, Liu W, Lu Z, Zhan J, Xu M, Chen L, Luo X, Cai G and Zhang S. PLOD2 contributes to drug resistance in laryngeal cancer by promoting cancer stem cell-like characteristics. *BMC Cancer* 2019; 19: 840.
- [76] Rodrigues MFSD, Miguita L, De Andrade NP, Heguedusch D, Rodini CO, Moyses RA, Toporcov TN, Gama RR, Tajara EE and Nunes FD. GLI3 knockdown decreases stemness, cell proliferation and invasion in oral squamous cell carcinoma. *Int J Oncol* 2018; 53: 2458-2472.
- [77] Chu PY, Hu FW, Yu CC, Tsai LL, Yu CH, Wu BC, Chen YW, Huang PI and Lo WL. Epithelial-mesenchymal transition transcription factor ZEB1/ZEB2 co-expression predicts poor prognosis and maintains tumor-initiating properties in head and neck cancer. *Oral Oncol* 2013; 49: 34-41.
- [78] Bae WJ, Koo BS, Lee SH, Kim JM, Rho YS, Lim JY, Moon JH, Cho JH and Lim YC. Inhibitor of DNA binding 2 is a novel therapeutic target for stemness of head and neck squamous cell carcinoma. *Br J Cancer* 2017; 117: 1810-1818.
- [79] Qiao B, Johnson NW, Chen X, Li R, Tao Q and Gao J. Disclosure of a stem cell phenotype in

Cancer stem cells of head and neck squamous cell carcinoma

- an oral squamous cell carcinoma cell line induced by BMP-4 via an epithelial-mesenchymal transition. *Oncol Rep* 2011; 26: 455-461.
- [80] Chen RH, Du Y, Han P, Wang HB, Liang FY, Feng GK, Zhou AJ, Cai MY, Zhong Q, Zeng MS and Huang XM. ISG15 predicts poor prognosis and promotes cancer stem cell phenotype in nasopharyngeal carcinoma. *Oncotarget* 2016; 7: 16910-16922.
- [81] Lu Y, Liang Y, Zheng X, Deng X, Huang W and Zhang G. EVI1 promotes epithelial-to-mesenchymal transition, cancer stem cell features and chemo-/radioresistance in nasopharyngeal carcinoma. *J Exp Clin Cancer Res* 2019; 38: 82.
- [82] Bu LL, Zhao ZL, Liu JF, Ma SR, Huang CF, Liu B, Zhang WF and Sun ZJ. STAT3 blockade enhances the efficacy of conventional chemotherapeutic agents by eradicating head neck stemloid cancer cell. *Oncotarget* 2015; 6: 41944-41958.
- [83] Wang J, Huang Y, Guan Z, Zhang JL, Su HK, Zhang W, Yue CF, Yan M, Guan S and Liu QQ. E3-ligase Skp2 predicts poor prognosis and maintains cancer stem cell pool in nasopharyngeal carcinoma. *Oncotarget* 2014; 5: 5591-5601.
- [84] Kong QL, Hu LJ, Cao JY, Huang YJ, Xu LH, Liang Y, Xiong D, Guan S, Guo BH, Mai HQ, Chen QY, Zhang X, Li MZ, Shao JY, Qian CN, Xia YF, Song LB, Zeng YX and Zeng MS. Epstein-Barr virus-encoded LMP2A induces an epithelial-mesenchymal transition and increases the number of side population stem-like cancer cells in nasopharyngeal carcinoma. *PLoS Pathog* 2010; 6: e1000940.
- [85] Suzuki T, Yamazaki H, Honda K, Ryo E, Kaneko A, Ota Y and Mori T. Altered DNA methylation is associated with aberrant stemness gene expression in early-stage HNSCC. *Int J Oncol* 2019; 55: 915-924.
- [86] Oliveira-Costa JP, Oliveira LR, da Silveira GG, Soave DF, Soares FA and Ribeiro-Silva A. Topoisomerase expression in oral squamous cell carcinoma: relationship with cancer stem cells profiles and lymph node metastasis. *J Oral Pathol Med* 2012; 41: 762-768.
- [87] Lin CS, Lin YC, Adebayo BO, Wu A, Chen JH, Peng YJ, Cheng MF, Lee WH, Hsiao M, Chao TY and Yeh CT. Silencing JARID1B suppresses oncogenicity, stemness and increases radiation sensitivity in human oral carcinoma. *Cancer Lett* 2015; 368: 36-45.
- [88] Moon JH, Lee SH, Koo BS, Kim JM, Huang S, Cho JH, Eun YG, Shin HA and Lim YC. Slug is a novel molecular target for head and neck squamous cell carcinoma stem-like cells. *Oral Oncol* 2020; 111: 104948.
- [89] Chen Y, Shao Z, Jiang E, Zhou X, Wang L, Wang H, Luo X, Chen Q, Liu K and Shang Z. CCL21/CCR7 interaction promotes EMT and enhances the stemness of OSCC via a JAK2/STAT3 signaling pathway. *J Cell Physiol* 2020; 235: 5995-6009.
- [90] Yao Z, Du L, Xu M, Li K, Guo H, Ye G, Zhang D, Coppes RP and Zhang H. MTA3-SOX2 module regulates cancer stemness and contributes to clinical outcomes of tongue carcinoma. *Front Oncol* 2019; 9: 816.
- [91] Chen L, Yang QC, Li YC, Yang LL, Liu JF, Li H, Xiao Y, Bu LL, Zhang WF and Sun ZJ. Targeting CMTM6 suppresses stem cell-like properties and enhances antitumor immunity in head and neck squamous cell carcinoma. *Cancer Immunol Res* 2020; 8: 179-191.
- [92] Chen X, Cao Y, Sedhom W, Lu L, Liu Y, Wang H, Oka M, Bornstein S, Said S, Song J and Lu SL. Distinct roles of PIK3CA in the enrichment and maintenance of cancer stem cells in head and neck squamous cell carcinoma. *Mol Oncol* 2020; 14: 139-158.
- [93] Sunkara RR, Sarate RM, Setia P, Shah S, Gupta S, Chaturvedi P, Gera P and Waghmare SK. SFRP1 in skin tumor initiation and cancer stem cell regulation with potential implications in epithelial cancers. *Stem Cell Reports* 2020; 14: 271-284.
- [94] Lu BC, Li J, Yu WF, Zhang GZ, Wang HM and Ma HM. Elevated expression of Nrf2 mediates multidrug resistance in CD133(+) head and neck squamous cell carcinoma stem cells. *Oncol Lett* 2016; 12: 4333-4338.
- [95] Essid N, Chambard JC and Elgaaied AB. Induction of epithelial-mesenchymal transition (EMT) and Gli1 expression in head and neck squamous cell carcinoma (HNSCC) spheroid cultures. *Bosn J Basic Med Sci* 2018; 18: 336-346.
- [96] Pai S, Bamodu OA, Lin YK, Lin CS, Chu PY, Chien MH, Wang LS, Hsiao M, Yeh CT and Tsai JT. CD47-SIRPalpha signaling induces epithelial-mesenchymal transition and cancer stemness and links to a poor prognosis in patients with oral squamous cell carcinoma. *Cells* 2019; 8: 1658.
- [97] Saloura V, Vougiouklakis T, Bao R, Kim S, Baek S, Zewde M, Bernard B, Burkitt K, Nigam N, Izumchenko E, Dohmae N, Hamamoto R and Nakamura Y. WHSC1 monomethylates histone H1 and induces stem-cell like features in squamous cell carcinoma of the head and neck. *Neoplasia* 2020; 22: 283-293.
- [98] Portney BA, Arad M, Gupta A, Brown RA, Khatri R, Lin PN, Hebert AM, Angster KH, Silipino LE, Meltzer WA, Taylor RJ and Zalzman M. ZSCAN4 facilitates chromatin remodeling and promotes the cancer stem cell phenotype. *Oncogene* 2020; 39: 4970-4982.
- [99] Yoshikawa M, Tsuchihashi K, Ishimoto T, Yae T, Motohara T, Sugihara E, Onishi N, Masuko T,

Cancer stem cells of head and neck squamous cell carcinoma

- Yoshizawa K, Kawashiri S, Mukai M, Asoda S, Kawana H, Nakagawa T, Saya H and Nagano O. xCT inhibition depletes CD44v-expressing tumor cells that are resistant to EGFR-targeted therapy in head and neck squamous cell carcinoma. *Cancer Res* 2013; 73: 1855-1866.
- [100] Xiang Z, Zhou S, Liang S, Zhang G and Tan Y. RCOR1 directly binds to MED28 and weakens its inducing effect on cancer stem cell-like activity of oral cavity squamous cell carcinoma cells. *J Oral Pathol Med* 2020; 49: 741-750.
- [101] Qin L, Yin YT, Zheng FJ, Peng LX, Yang CF, Bao YN, Liang YY, Li XJ, Xiang YQ, Sun R, Li AH, Zou RH, Pei XQ, Huang BJ, Kang TB, Liao DF, Zeng YX, Williams BO and Qian CN. WNT5A promotes stemness characteristics in nasopharyngeal carcinoma cells leading to metastasis and tumorigenesis. *Oncotarget* 2015; 6: 10239-10252.
- [102] Ma SR, Wang WM, Huang CF, Zhang WF and Sun ZJ. Anterior gradient protein 2 expression in high grade head and neck squamous cell carcinoma correlated with cancer stem cell and epithelial mesenchymal transition. *Oncotarget* 2015; 6: 8807-8821.
- [103] Featherston T, Marsh RW, van Schaijik B, Brasch HD, Tan ST and Itinteang T. Expression and localization of cathepsins B, D, and G in two cancer stem cell subpopulations in moderately differentiated oral tongue squamous cell carcinoma. *Front Med (Lausanne)* 2017; 4: 100.
- [104] Morvan VL, Richard E, Cadars M, Fessart D, Broca-Brisson L, Auzanneau C, Pasquies A, Modesto A, Lusque A, Mathoulin-Pelissier S, Lansiaux A and Robert J. Cytochrome P450 1B1 polymorphism drives cancer cell stemness and patient outcome in head-and-neck carcinoma. *Br J Cancer* 2020; 123: 772-784.
- [105] Lorenzo-Martin LF, Menacho-Marquez M and Bustelo XR. Drug vulnerabilities and disease prognosis linked to the stem cell-like gene expression program triggered by the RHO GTPase activator VAV2 in hyperplastic keratinocytes and head and neck cancer. *Cancers (Basel)* 2020; 12: 2498.
- [106] Garcia-Mayea Y, Mir C, Carballo L, Castellvi J, Temprana-Salvador J, Lorente J, Benavente S, Garcia-Pedrero JM, Allonca E, Rodrigo JP and LLeonart ME. TSPAN1: a novel protein involved in head and neck squamous cell carcinoma chemoresistance. *Cancers (Basel)* 2020; 12: 3269.
- [107] Hu X, Zou W, Liu D, Qin G and Jiang L. The down-regulation of TrkB alleviates the malignant biological behavior and cancer stem-like property of laryngeal cancer. *Cancer Manag Res* 2020; 12: 6865-6875.
- [108] Zhang Y, Cai H, Liao Y, Zhu Y, Wang F and Hou J. Activation of PGK1 under hypoxic conditions promotes glycolysis and increases stem cell-like properties and the epithelial-mesenchymal transition in oral squamous cell carcinoma cells via the AKT signalling pathway. *Int J Oncol* 2020; 57: 743-755.
- [109] Ferreira Mendes JM, de Faro Valverde L, Torres Andion Vidal M, Paredes BD, Coelho P, Allahdadi KJ, Coletta RD, Souza BSF and Rocha CAG. Effects of IGF-1 on proliferation, angiogenesis, tumor stem cell populations and activation of AKT and hedgehog pathways in oral squamous cell carcinoma. *Int J Mol Sci* 2020; 21: 6487.
- [110] Zhang M, Hoyle RG, Ma Z, Sun B, Cai W, Cai H, Xie N, Zhang Y, Hou J, Liu X, Chen D, Kellogg GE, Harada H, Sun Y, Wang C and Li J. FOSL1 promotes metastasis of head and neck squamous cell carcinoma through super-enhancer-driven transcription program. *Mol Ther* 2021; 29: 2583-2600.
- [111] Wang D. Promotive effects of HOXA10 antisense RNA on the stemness of oral squamous cell carcinoma stem cells through a microRNA-29a/MCL-1/phosphatidylinositol 3-kinase/protein kinase B axis. *Arch Oral Biol* 2021; 126: 105114.
- [112] Xu X, Tassone B, Ostano P, Katarkar A, Proust T, Joseph JM, Riganti C, Chiorino G, Kutalik Z, Lefort K and Dotto GP. HSD17B7 gene in self-renewal and oncogenicity of keratinocytes from Black versus White populations. *EMBO Mol Med* 2021; 13: e14133.
- [113] Shigeishi H, Hashikata M, Yokoyama S, Sakuma M, Murozumi H, Kato H, Rahman MZ, Seino S, Ishioka Y, Ohta K, Takechi M and Sugiyama M. CD44(high)/ESA(low) squamous cell carcinoma cell-derived prostaglandin E2 confers resistance to 5-fluorouracil-induced apoptosis in CD44(high)/ESA(high) cells. *Int J Clin Exp Pathol* 2018; 11: 2356-2363.
- [114] Yu CC, Chen PN, Peng CY, Yu CH and Chou MY. Suppression of miR-204 enables oral squamous cell carcinomas to promote cancer stemness, EMT traits, and lymph node metastasis. *Oncotarget* 2016; 7: 20180-20192.
- [115] Sun Z, Hu W, Xu J, Kaufmann AM and Albers AE. MicroRNA-34a regulates epithelial-mesenchymal transition and cancer stem cell phenotype of head and neck squamous cell carcinoma in vitro. *Int J Oncol* 2015; 47: 1339-1350.
- [116] Lo WL, Yu CC, Chiou GY, Chen YW, Huang PI, Chien CS, Tseng LM, Chu PY, Lu KH, Chang KW, Kao SY and Chiou SH. MicroRNA-200c attenuates tumour growth and metastasis of presumptive head and neck squamous cell carcinoma stem cells. *J Pathol* 2011; 223: 482-495.

Cancer stem cells of head and neck squamous cell carcinoma

- [117] Lu YC, Cheng AJ, Lee LY, You GR, Li YL, Chen HY and Chang JT. MiR-520b as a novel molecular target for suppressing stemness phenotype of head-neck cancer by inhibiting CD44. *Sci Rep* 2017; 7: 2042.
- [118] Yu CC, Chen YW, Chiou GY, Tsai LL, Huang PI, Chang CY, Tseng LM, Chiou SH, Yen SH, Chou MY, Chu PY and Lo WL. MicroRNA let-7a represses chemoresistance and tumorigenicity in head and neck cancer via stem-like properties ablation. *Oral Oncol* 2011; 47: 202-210.
- [119] Peng CY, Wang TY, Lee SS, Hsieh PL, Liao YW, Tsai LL, Fang CY, Yu CC and Hsieh CS. Let-7c restores radiosensitivity and chemosensitivity and impairs stemness in oral cancer cells through inhibiting interleukin-8. *J Oral Pathol Med* 2018; 47: 590-597.
- [120] Zhou YM, Yao YL, Liu W, Shen XM, Shi LJ and Wu L. MicroRNA-134 inhibits tumor stem cell migration and invasion in oral squamous cell carcinomas via downregulation of PI3K-Akt signaling pathway by inhibiting LAMC2 expression. *Cancer Biomark* 2020; 29: 51-67.
- [121] Cui M, Chang Y, Fang QG, Du W, Wu JF, Wang JH, Liu ST and Luo SX. Non-coding RNA Pvt1 promotes cancer stem cell-like traits in nasopharyngeal cancer via inhibiting miR-1207. *Pathol Oncol Res* 2019; 25: 1411-1422.
- [122] Yuan Z, Xiu C, Liu D, Zhou G, Yang H, Pei R, Ding C, Cui X, Sun J and Song K. Long noncoding RNA LINC-PINT regulates laryngeal carcinoma cell stemness and chemoresistance through miR-425-5p/PTCH1/SHH axis. *J Cell Physiol* 2019; 234: 23111-23122.
- [123] Liu J, Tang Q, Li S and Yang X. Inhibition of HAX-1 by miR-125a reverses cisplatin resistance in laryngeal cancer stem cells. *Oncotarget* 2016; 7: 86446-86456.
- [124] Lee SP, Hsieh PL, Fang CY, Chu PM, Liao YW, Yu CH, Yu CC and Tsai LL. LINC00963 promotes cancer stemness, metastasis, and drug resistance in head and neck carcinomas via ABCB5 regulation. *Cancers (Basel)* 2020; 12: 1073.
- [125] You X, Zhou Z, Chen W, Wei X, Zhou H and Luo W. MicroRNA-495 confers inhibitory effects on cancer stem cells in oral squamous cell carcinoma through the HOXC6-mediated TGF-beta signaling pathway. *Stem Cell Res Ther* 2020; 11: 117.
- [126] Lin SS, Peng CY, Liao YW, Chou MY, Hsieh PL and Yu CC. miR-1246 Targets CCNG2 to enhance cancer stemness and chemoresistance in oral carcinomas. *Cancers (Basel)* 2018; 10: 272.
- [127] Wu CP, Du HD, Gong HL, Li DW, Tao L, Tian J and Zhou L. Hypoxia promotes stem-like properties of laryngeal cancer cell lines by increasing the CD133+ stem cell fraction. *Int J Oncol* 2014; 44: 1652-1660.
- [128] Krishnamurthy S, Warner KA, Dong Z, Imai A, Nor C, Ward BB, Helman JI, Taichman RS, Bellile EL, McCauley LK, Polverini PJ, Prince ME, Wicha MS and Nor JE. Endothelial interleukin-6 defines the tumorigenic potential of primary human cancer stem cells. *Stem Cells* 2014; 32: 2845-2857.
- [129] Guan GF, Tang XX, Zhang DJ, Zheng Y, Yu DJ, Zhao Y, Lu YQ and Zhu L. Constitutive secretion of Interleukin-4 dictates CD133+ side population cells to resist drug treatment and cell death. *J BUON* 2015; 20: 1350-1359.
- [130] Lu L, Wang P, Zou Y, Zha Z, Huang H, Guan M, Wu Y and Liu G. IL-1beta promotes stemness of tumor cells by activating smad/ID1 signaling pathway. *Int J Med Sci* 2020; 17: 1257-1268.
- [131] Xu Q, Zhang Q, Ishida Y, Hajjar S, Tang X, Shi H, Dang CV and Le AD. EGF induces epithelial-mesenchymal transition and cancer stem-like cell properties in human oral cancer cells via promoting Warburg effect. *Oncotarget* 2017; 8: 9557-9571.
- [132] Bae WJ, Lee SH, Rho YS, Koo BS and Lim YC. Transforming growth factor beta1 enhances stemness of head and neck squamous cell carcinoma cells through activation of Wnt signaling. *Oncol Lett* 2016; 12: 5315-5320.
- [133] He KF, Zhang L, Huang CF, Ma SR, Wang YF, Wang WM, Zhao ZL, Liu B, Zhao YF, Zhang WF and Sun ZJ. CD163+ tumor-associated macrophages correlated with poor prognosis and cancer stem cells in oral squamous cell carcinoma. *Biomed Res Int* 2014; 2014: 838632.
- [134] Yu B, Wu K, Wang X, Zhang J, Wang L, Jiang Y, Zhu X, Chen W and Yan M. Periostin secreted by cancer-associated fibroblasts promotes cancer stemness in head and neck cancer by activating protein tyrosine kinase 7. *Cell Death Dis* 2018; 9: 1082.
- [135] Datta KK, Patil S, Patel K, Babu N, Raja R, Nanjappa V, Mangalaparthy KK, Dhaka B, Rajagopalan P, Deolankar SC, Kannan R, Kumar P, Prasad TSK, Mathur PP, Kumari A, Manoharan M, Coral K, Murugan S, Sidransky D, Gupta R, Gupta R, Khanna-Gupta A, Chatterjee A and Gowda H. Chronic exposure to chewing tobacco induces metabolic reprogramming and cancer stem cell-like properties in esophageal epithelial cells. *Cells* 2019; 8: 949.
- [136] Wang TY, Peng CY, Lee SS, Chou MY, Yu CC and Chang YC. Acquisition cancer stemness, mesenchymal transdifferentiation, and chemoresistance properties by chronic exposure of oral epithelial cells to arecoline. *Oncotarget* 2016; 7: 84072-84081.
- [137] Yu MA, Kiang A, Wang-Rodriguez J, Rahimy E, Haas M, Yu V, Ellies LG, Chen J, Fan JB, Brumund KT, Weisman RA and Ongkeko WM. Nico-

Cancer stem cells of head and neck squamous cell carcinoma

- tine promotes acquisition of stem cell and epithelial-to-mesenchymal properties in head and neck squamous cell carcinoma. *PLoS One* 2012; 7: e51967.
- [138] An Y, Kiang A, Lopez JP, Kuo SZ, Yu MA, Abhold EL, Chen JS, Wang-Rodriguez J and Ongkeko WM. Cigarette smoke promotes drug resistance and expansion of cancer stem cell-like side population. *PLoS One* 2012; 7: e47919.
- [139] Salazar-Garcia L, Perez-Sayans M, Garcia-Garcia A, Carracedo A, Cruz R, Lozano A, Sobrino B and Barros F. Whole exome sequencing approach to analysis of the origin of cancer stem cells in patients with head and neck squamous cell carcinoma. *J Oral Pathol Med* 2018; 47: 938-944.
- [140] Nimmakayala RK, Batra SK and Ponnusamy MP. Unraveling the journey of cancer stem cells from origin to metastasis. *Biochim Biophys Acta Rev Cancer* 2019; 1871: 50-63.
- [141] Yang L, Shi P, Zhao G, Xu J, Peng W, Zhang J, Zhang G, Wang X, Dong Z, Chen F and Cui H. Targeting cancer stem cell pathways for cancer therapy. *Signal Transduct Target Ther* 2020; 5: 8.
- [142] Muralidharan-Chari V, Clancy JW, Sedgwick A and D'Souza-Schorey C. Microvesicles: mediators of extracellular communication during cancer progression. *J Cell Sci* 2010; 123: 1603-1611.
- [143] van Doormaal FF, Kleinjan A, Di Nisio M, Büller HR and Nieuwland R. Cell-derived microvesicles and cancer. *Neth J Med* 2009; 67: 266-273.
- [144] Castellana D, Zobairi F, Martinez MC, Panaro MA, Mitolo V, Freyssinet JM and Kunzelmann C. Membrane microvesicles as actors in the establishment of a favorable prostatic tumoral niche: a role for activated fibroblasts and CX-3CL1-CX3CR1 axis. *Cancer Res* 2009; 69: 785-793.
- [145] Ratajczak J, Miekus K, Kucia M, Zhang J, Reca R, Dvorak P and Ratajczak MZ. Embryonic stem cell-derived microvesicles reprogram hematopoietic progenitors: evidence for horizontal transfer of mRNA and protein delivery. *Leukemia* 2006; 20: 847-856.
- [146] Deregibus MC, Cantaluppi V, Calogero R, Lo Iacono M, Tetta C, Biancone L, Bruno S, Bussolati B and Camussi G. Endothelial progenitor cell derived microvesicles activate an angiogenic program in endothelial cells by a horizontal transfer of mRNA. *Blood* 2007; 110: 2440-2448.
- [147] Quesenberry PJ, Dooner MS and Aliotta JM. Stem cell plasticity revisited: the continuum marrow model and phenotypic changes mediated by microvesicles. *Exp Hematol* 2010; 38: 581-592.
- [148] Grange C, Tapparo M, Collino F, Vitillo L, Damasco C, Deregibus MC, Tetta C, Bussolati B and Camussi G. Microvesicles released from human renal cancer stem cells stimulate angiogenesis and formation of lung premetastatic niche. *Cancer Res* 2011; 71: 5346-5356.
- [149] Williams K, Motiani K, Giridhar PV and Kasper S. CD44 integrates signaling in normal stem cell, cancer stem cell and (pre)metastatic niches. *Exp Biol Med (Maywood)* 2013; 238: 324-338.
- [150] Heiler S, Wang Z and Zöller M. Pancreatic cancer stem cell markers and exosomes - the incentive push. *World J Gastroenterol* 2016; 22: 5971-6007.
- [151] Mao XD, Wei X, Xu T, Li TP and Liu KS. Research progress in breast cancer stem cells: characterization and future perspectives. *Am J Cancer Res* 2022; 12: 3208-3222.
- [152] Liu C, Qiang J, Deng Q, Xia J, Deng L, Zhou L, Wang D, He X, Liu Y, Zhao B, Lv J, Yu Z, Lei QY, Shao ZM, Zhang XY, Zhang L and Liu S. ALDH1A1 activity in tumor-initiating cells remodels myeloid-derived suppressor cells to promote breast cancer progression. *Cancer Res* 2021; 81: 5919-5934.
- [153] Krishnamurthy S and Nör JE. Head and neck cancer stem cells. *J Dent Res* 2012; 91: 334-340.
- [154] Baumann M and Krause M. CD44: a cancer stem cell-related biomarker with predictive potential for radiotherapy. *Clin Cancer Res* 2010; 16: 5091-5093.
- [155] Yan M, Yang X, Wang L, Clark D, Zuo H, Ye D, Chen W and Zhang P. Plasma membrane proteomics of tumor spheres identify CD166 as a novel marker for cancer stem-like cells in head and neck squamous cell carcinoma. *Mol Cell Proteomics* 2013; 12: 3271-3284.
- [156] Wiehac E, Matic N, Ali A and Roberg K. Hypoxia induces radioresistance, epithelial-mesenchymal transition, cancer stem cell-like phenotype and changes in genes possessing multiple biological functions in head and neck squamous cell carcinoma. *Oncol Rep* 2022; 47: 58.
- [157] Mori T. Involvement of the p53-p16/RB pathway control mechanism in early-stage carcinogenesis in head and neck squamous cell carcinoma. *Pathol Int* 2022; 72: 577-588.
- [158] Liu S, Li Q, Chen K, Zhang Q, Li G, Zhuo L, Zhai B, Sui X, Hu X and Xie T. The emerging molecular mechanism of m6A modulators in tumorigenesis and cancer progression. *Biomed Pharmacother* 2020; 127: 110098.
- [159] Bjerkvig R, Tysnes BB, Aboody KS, Najbauer J and Terzis AJ. Opinion: the origin of the cancer stem cell: current controversies and new insights. *Nat Rev Cancer* 2005; 5: 899-904.

Cancer stem cells of head and neck squamous cell carcinoma

- [160] Matsui W, Huff CA, Wang Q, Malehorn MT, Barber J, Tanhehco Y, Smith BD, Civin CI and Jones RJ. Characterization of clonogenic multiple myeloma cells. *Blood* 2004; 103: 2332-2336.
- [161] Hill RP. Identifying cancer stem cells in solid tumors: case not proven. *Cancer Res* 2006; 66: 1891-1895; discussion 1890.
- [162] Huntly BJ and Gilliland DG. Leukaemia stem cells and the evolution of cancer-stem-cell research. *Nat Rev Cancer* 2005; 5: 311-321.
- [163] Li C, Heidt DG, Dalerba P, Burant CF, Zhang L, Adsay V, Wicha M, Clarke MF and Simeone DM. Identification of pancreatic cancer stem cells. *Cancer Res* 2007; 67: 1030-1037.
- [164] Wang W, Osenbroch P, Skinnis R, Esbensen Y, Bjørås M and Eide L. Mitochondrial DNA integrity is essential for mitochondrial maturation during differentiation of neural stem cells. *Stem Cells* 2010; 28: 2195-2204.
- [165] Pardal R, Clarke MF and Morrison SJ. Applying the principles of stem-cell biology to cancer. *Nat Rev Cancer* 2003; 3: 895-902.
- [166] Takahashi K and Yamanaka S. Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell* 2006; 126: 663-676.
- [167] Maherali N, Sridharan R, Xie W, Utikal J, Eminli S, Arnold K, Stadtfeld M, Yachechko R, Tchiew J, Jaenisch R, Plath K and Hochedlinger K. Directly reprogrammed fibroblasts show global epigenetic remodeling and widespread tissue contribution. *Cell Stem Cell* 2007; 1: 55-70.
- [168] Rodriguez-Pinilla SM, Sarrio D, Moreno-Bueno G, Rodriguez-Gil Y, Martinez MA, Hernandez L, Hardisson D, Reis-Filho JS and Palacios J. Sox2: a possible driver of the basal-like phenotype in sporadic breast cancer. *Mod Pathol* 2007; 20: 474-481.
- [169] Beer S, Zetterberg A, Ihrie RA, McTaggart RA, Yang Q, Bradon N, Arvanitis C, Attardi LD, Feng S, Ruebner B, Cardiff RD and Felsher DW. Developmental context determines latency of MYC-induced tumorigenesis. *PLoS Biol* 2004; 2: e332.
- [170] Jacobs JJ, Scheijen B, Voncken JW, Kieboom K, Berns A and van Lohuizen M. Bmi-1 collaborates with c-Myc in tumorigenesis by inhibiting c-Myc-induced apoptosis via INK4a/ARF. *Genes Dev* 1999; 13: 2678-2690.
- [171] Shachaf CM, Kopelman AM, Arvanitis C, Karlsson A, Beer S, Mandl S, Bachmann MH, Borowsky AD, Ruebner B, Cardiff RD, Yang Q, Bishop JM, Contag CH and Felsher DW. MYC inactivation uncovers pluripotent differentiation and tumour dormancy in hepatocellular cancer. *Nature* 2004; 431: 1112-1117.
- [172] Bonnet D and Dick JE. Human acute myeloid leukemia is organized as a hierarchy that originates from a primitive hematopoietic cell. *Nat Med* 1997; 3: 730-737.
- [173] Hemmati HD, Nakano I, Lazareff JA, Masterman-Smith M, Geschwind DH, Bronner-Fraser M and Kornblum HI. Cancerous stem cells can arise from pediatric brain tumors. *Proc Natl Acad Sci U S A* 2003; 100: 15178-15183.
- [174] Al-Hajj M, Wicha MS, Benito-Hernandez A, Morrison SJ and Clarke MF. Prospective identification of tumorigenic breast cancer cells. *Proc Natl Acad Sci U S A* 2003; 100: 3983-3988.
- [175] Ginestier C, Hur MH, Charafe-Jauffret E, Monville F, Dutcher J, Brown M, Jacquemier J, Viens P, Kleer CG, Liu S, Schott A, Hayes D, Birnbaum D, Wicha MS and Dontu G. ALDH1 is a marker of normal and malignant human mammary stem cells and a predictor of poor clinical outcome. *Cell Stem Cell* 2007; 1: 555-567.
- [176] Hirschmann-Jax C, Foster AE, Wulf GG, Nuchtern JG, Jax TW, Gobel U, Goodell MA and Brenner MK. A distinct "side population" of cells with high drug efflux capacity in human tumor cells. *Proc Natl Acad Sci U S A* 2004; 101: 14228-14233.
- [177] Quintana E, Shackleton M, Foster HR, Fullen DR, Sabel MS, Johnson TM and Morrison SJ. Phenotypic heterogeneity among tumorigenic melanoma cells from patients that is reversible and not hierarchically organized. *Cancer Cell* 2010; 18: 510-523.
- [178] Ayob AZ and Ramasamy TS. Cancer stem cells as key drivers of tumour progression. *J Biomed Sci* 2018; 25: 20.
- [179] Sigismund S, Avanzato D and Lanzetti L. Emerging functions of the EGFR in cancer. *Mol Oncol* 2018; 12: 3-20.
- [180] Alshahafi EN, Thavaraj S, Sarvestani N, Novoplansky O, Elkabets M, Ayaz B, Tavassoli M and Legends MF. EGFR overexpression increases radiotherapy response in HPV-positive head and neck cancer through inhibition of DNA damage repair and HPV E6 downregulation. *Cancer Lett* 2021; 498: 80-97.
- [181] Setubal Destro Rodrigues MF, Gammon L, Rahman MM, Biddle A, Nunes FD and Mackenzie IC. Effects of cetuximab and Erlotinib on the behaviour of cancer stem cells in head and neck squamous cell carcinoma. *Oncotarget* 2018; 9: 13488-13500.
- [182] Macha MA, Rachagani S, Qazi AK, Jahan R, Gupta S, Patel A, Seshacharyulu P, Lin C, Li S, Wang S, Verma V, Kishida S, Kishida M, Nakamura N, Kibe T, Lydiatt WM, Smith RB, Ganti AK, Jones DT, Batra SK and Jain M. Afatinib radiosensitizes head and neck squamous cell carcinoma cells by targeting cancer stem cells. *Oncotarget* 2017; 8: 20961-20973.
- [183] Liu X, Suo H, Zhou S, Hou Z, Bu M, Liu X and Xu W. Afatinib induces pro-survival autophagy and increases sensitivity to apoptosis in stem-like HNSCC cells. *Cell Death Dis* 2021; 12: 728.

Cancer stem cells of head and neck squamous cell carcinoma

- [184] Guy JB, Espenel S, Louati S, Gauthier A, Garcia MA, Vial N, Malesys C, Ardail D, Alphonse G, Wozny AS, Rodriguez-Lafrasse C and Magne N. Combining radiation to EGFR and Bcl-2 blockade: a new approach to target cancer stem cells in head and neck squamous cell carcinoma. *J Cancer Res Clin Oncol* 2021; 147: 1905-1916.
- [185] Gilormini M, Malesys C, Armandy E, Manas P, Guy JB, Magne N, Rodriguez-Lafrasse C and Ardail D. Preferential targeting of cancer stem cells in the radiosensitizing effect of ABT-737 on HNSCC. *Oncotarget* 2016; 7: 16731-16744.
- [186] Guy J, Espenel S, Vallard A, Méry B, Rancoule C, Wozny A, Simonet S, Beuve M, Alphonse G and Ardail D. Targeting cancer stem cells in HNSCC: synergic effect of cetuximab and ABT-199 in combination with photon radiation. *Int J Radiat Oncol Biol Phys* 2017; 99: E593.
- [187] Shyh-Chang N and Daley GQ. Lin28: primal regulator of growth and metabolism in stem cells. *Cell Stem Cell* 2013; 12: 395-406.
- [188] Chen H, Sa G, Li L, He S and Wu T. In vitro and in vivo synergistic anti-tumor effect of LIN28 inhibitor and metformin in oral squamous cell carcinoma. *Eur J Pharmacol* 2021; 891: 173757.
- [189] Burger JA. Bruton tyrosine kinase inhibitors: present and future. *Cancer J* 2019; 25: 386-393.
- [190] Liu SC, Wu YC, Huang CM, Hsieh MS, Huang TY, Huang CS, Hsu TN, Huang MS, Lee WH, Yeh CT and Lin CS. Inhibition of Bruton's tyrosine kinase as a therapeutic strategy for chemoresistant oral squamous cell carcinoma and potential suppression of cancer stemness. *Oncogenesis* 2021; 10: 20.
- [191] Shigeishi H, Biddle A, Gammon L, Rodini CO, Yamasaki M, Seino S, Sugiyama M, Takechi M and Mackenzie IC. Elevation in 5-FU-induced apoptosis in head and neck cancer stem cells by a combination of CDHP and GSK3beta inhibitors. *J Oral Pathol Med* 2015; 44: 201-207.
- [192] Huang J, Qu Q, Guo Y, Xiang Y and Feng D. Tankyrases/beta-catenin signaling pathway as an anti-proliferation and anti-metastatic target in hepatocarcinoma cell lines. *J Cancer* 2020; 11: 432-440.
- [193] Roy S, Roy S, Kar M, Chakraborty A, Kumar A, Delogu F, Asthana S, Hande MP and Banerjee B. Combined treatment with cisplatin and the tankyrase inhibitor XAV-939 increases cytotoxicity, abrogates cancer-stem-like cell phenotype and increases chemosensitivity of head-and-neck squamous-cell carcinoma cells. *Mutat Res Genet Toxicol Environ Mutagen* 2019; 846: 503084.
- [194] Fang L, Zhu Q, Neuenschwander M, Specker E, Wulf-Goldenberg A, Weis WI, von Kries JP and Birchmeier W. A small-molecule antagonist of the beta-catenin/TCF4 interaction blocks the self-renewal of cancer stem cells and suppresses tumorigenesis. *Cancer Res* 2016; 76: 891-901.
- [195] Bukhari SA, Yasmin A, Zahoor MA, Mustafa G, Sarfraz I and Rasul A. Secreted frizzled-related protein 4 and its implication in obesity and type-2 diabetes. *IUBMB Life* 2019; 71: 1701-1710.
- [196] Warriar S, Bhuvanlakshmi G, Arfuso F, Rajan G, Millward M and Dharmarajan A. Cancer stem-like cells from head and neck cancers are chemosensitized by the Wnt antagonist, sFRP4, by inducing apoptosis, decreasing stemness, drug resistance and epithelial to mesenchymal transition. *Cancer Gene Ther* 2014; 21: 381-388.
- [197] Milazzo G, Mercatelli D, Di Muzio G, Triboli L, De Rosa P, Perini G and Giorgi FM. Histone deacetylases (HDACs): evolution, specificity, role in transcriptional complexes, and pharmacological actionability. *Genes (Basel)* 2020; 11: 556.
- [198] Xu H, Li L, Wang S, Wang Z, Qu L, Wang C and Xu K. Royal jelly acid suppresses hepatocellular carcinoma tumorigenicity by inhibiting H3 histone lactylation at H3K9la and H3K14la sites. *Phytomedicine* 2023; 118: 154940.
- [199] Xu Z, Bahadar N, Zhang Y, Tan S, Wang Z, Ren B, Liu S, Dai H, Zheng Y and Han B. Transcriptomic profiling of mice brain under Bex3 regulation. *Turk J Biol* 2021; 46: 57-68.
- [200] Bahadar N, Ullah H, Adlat S, Kumar Sah R, Zun Zaw Myint M, Mar Oo Z, Binta Bah F, Hayel Nagi F, Htoo H, Ud Din A, Feng X and Zheng Y. Analyzing differentially expressed genes and pathways of Bex2-deficient mouse lung via RNA-Seq. *Turk J Biol* 2021; 45: 588-598.
- [201] Chikamatsu K, Ishii H, Murata T, Sakakura K, Shino M, Toyoda M, Takahashi K and Masuyama K. Alteration of cancer stem cell-like phenotype by histone deacetylase inhibitors in squamous cell carcinoma of the head and neck. *Cancer Sci* 2013; 104: 1468-1475.
- [202] Kumar B, Yadav A, Lang JC, Teknos TN and Kumar P. Suberoylanilide hydroxamic acid (SAHA) reverses chemoresistance in head and neck cancer cells by targeting cancer stem cells via the downregulation of nanog. *Genes Cancer* 2015; 6: 169-181.
- [203] Lee SH, Nam HJ, Kang HJ, Samuels TL, Johnston N and Lim YC. Valproic acid suppresses the self-renewal and proliferation of head and neck cancer stem cells. *Oncol Rep* 2015; 34: 2065-2071.
- [204] Marques AEM, do Nascimento Filho CHV, Marinho Bezerra TM, Guerra ENS, Castilho RM and Squarize CH. Entinostat is a novel therapeutic agent to treat oral squamous cell carcinoma. *J Oral Pathol Med* 2020; 49: 771-779.

Cancer stem cells of head and neck squamous cell carcinoma

- [205] Boivin A, Hanot M, Malesys C, Maalouf M, Rousson R, Rodriguez-Lafrasse C and Ardail D. Transient alteration of cellular redox buffering before irradiation triggers apoptosis in head and neck carcinoma stem and non-stem cells. *PLoS One* 2011; 6: e14558.
- [206] Bertrand G, Maalouf M, Boivin A, Battiston-Montagne P, Beuve M, Levy A, Jalade P, Fournier C, Ardail D, Magne N, Alphonse G and Rodriguez-Lafrasse C. Targeting head and neck cancer stem cells to overcome resistance to photon and carbon ion radiation. *Stem Cell Rev Rep* 2014; 10: 114-126.
- [207] Finkel KA, Warner KA, Kerk S, Bradford CR, McLean SA, Prince ME, Zhong H, Hurt EM, Hollingsworth RE, Wicha MS, Tice DA and Nor JE. IL-6 inhibition with MEDI5117 decreases the fraction of head and neck cancer stem cells and prevents tumor recurrence. *Neoplasia* 2016; 18: 273-281.
- [208] Jia L, Zhang W and Wang CY. BMI1 inhibition eliminates residual cancer stem cells after PD1 blockade and activates antitumor immunity to prevent metastasis and relapse. *Cell Stem Cell* 2020; 27: 238-253, e236.
- [209] Herzog AE, Warner KA, Zhang Z, Bellile E, Bhagat MA, Castilho RM, Wolf GT, Polverini PJ, Pearson AT and Nor JE. The IL-6R and Bmi-1 axis controls self-renewal and chemoresistance of head and neck cancer stem cells. *Cell Death Dis* 2021; 12: 988.
- [210] Saito S, Ozawa H, Imanishi Y, Sekimizu M, Watanabe Y, Ito F, Ikari Y, Nakahara N, Kameyama K and Ogawa K. Cyclooxygenase-2 expression is associated with chemoresistance through cancer stemness property in hypopharyngeal carcinoma. *Oncol Lett* 2021; 22: 533.
- [211] Liu Q, Yu S, Zhao W, Qin S, Chu Q and Wu K. EGFR-TKIs resistance via EGFR-independent signaling pathways. *Mol Cancer* 2018; 17: 53.
- [212] Liu SC, Huang CM, Bamodu OA, Lin CS, Liu BL, Tzeng YM, Tsai JT, Lee WH and Chen TM. Ovatodiolide suppresses nasopharyngeal cancer by targeting stem cell-like population, inducing apoptosis, inhibiting EMT and dysregulating JAK/STAT signaling pathway. *Phytomedicine* 2019; 56: 269-278.
- [213] Lee SH, Nam HJ, Kang HJ, Kwon HW and Lim YC. Epigallocatechin-3-gallate attenuates head and neck cancer stem cell traits through suppression of notch pathway. *Eur J Cancer* 2013; 49: 3210-3218.
- [214] Derosa G, Maffioli P, D'Angelo A and Di Pierro F. A role for quercetin in coronavirus disease 2019 (COVID-19). *Phytother Res* 2021; 35: 1230-1236.
- [215] Chang WW, Hu FW, Yu CC, Wang HH, Feng HP, Lan C, Tsai LL and Chang YC. Quercetin in elimination of tumor initiating stem-like and mesenchymal transformation property in head and neck cancer. *Head Neck* 2013; 35: 413-419.
- [216] Deng R, Wang X, Liu Y, Yan M, Hanada S, Xu Q, Zhang J, Han Z, Chen W and Zhang P. A new gamboge derivative compound 2 inhibits cancer stem-like cells via suppressing EGFR tyrosine phosphorylation in head and neck squamous cell carcinoma. *J Cell Mol Med* 2013; 17: 1422-1433.
- [217] Chen YW, Chen KH, Huang PI, Chen YC, Chiou GY, Lo WL, Tseng LM, Hsu HS, Chang KW and Chiou SH. Cucurbitacin I suppressed stem-like property and enhanced radiation-induced apoptosis in head and neck squamous carcinoma-derived CD44(+)/ALDH1(+) cells. *Mol Cancer Ther* 2010; 9: 2879-2892.
- [218] Chang CW, Chen CC, Wu MJ, Chen YS, Chen CC, Sheu SJ, Lin TW, Chou SH, Lin SC, Liu CJ, Lee TC, Huang CY and Lo JF. Active component of *antrodia cinnamomea* mycelia targeting head and neck cancer initiating cells through exaggerated autophagic cell death. *Evid Based Complement Alternat Med* 2013; 2013: 946451.
- [219] Chang CW, Chen YS, Chen CC, Chan IO, Chen CC, Sheu SJ, Lin TW, Chou SH, Liu CJ, Lee TC and Lo JF. Targeting cancer initiating cells by promoting cell differentiation and restoring chemosensitivity via dual inactivation of STAT3 and src activity using an active component of *antrodia cinnamomea* mycelia. *Oncotarget* 2016; 7: 73016-73031.
- [220] Pan ST, Qin Y, Zhou ZW, He ZX, Zhang X, Yang T, Yang YX, Wang D, Zhou SF and Qiu JX. Plumbagin suppresses epithelial to mesenchymal transition and stemness via inhibiting Nrf2-mediated signaling pathway in human tongue squamous cell carcinoma cells. *Drug Des Dev Ther* 2015; 9: 5511-5551.
- [221] Keysar SB, Gomes N, Miller B, Jackson BC, Le PN, Morton JJ, Reisinger J, Chimed TS, Gomez KE, Nieto C, Frederick B, Pronk GJ, Somerset HL, Tan AC, Wang XJ, Raben D, Su TT and Jimeno A. Inhibiting translation elongation with SVC112 suppresses cancer stem cells and inhibits growth in head and neck squamous carcinoma. *Cancer Res* 2020; 80: 1183-1198.
- [222] Peng Y, He G, Tang D, Xiong L, Wen Y, Miao X, Hong Z, Yao H, Chen C, Yan S, Lu L, Yang Y, Li Q and Deng X. Lovastatin inhibits cancer stem cells and sensitizes to chemo- and photodynamic therapy in nasopharyngeal carcinoma. *J Cancer* 2017; 8: 1655-1664.
- [223] Cui X, Xiao D and Wang X. Inhibition of laryngeal cancer stem cells by tetrandrine. *Anticancer Drugs* 2019; 30: 886-891.
- [224] Hu FW, Yu CC, Hsieh PL, Liao YW, Lu MY and Chu PM. Targeting oral cancer stemness and

Cancer stem cells of head and neck squamous cell carcinoma

- chemoresistance by isoliquiritigenin-mediated GRP78 regulation. *Oncotarget* 2017; 8: 93912-93923.
- [225] Elkashty OA and Tran SD. Broccoli extract increases drug-mediated cytotoxicity towards cancer stem cells of head and neck squamous cell carcinoma. *Br J Cancer* 2020; 123: 1395-1403.
- [226] Basak SK, Zinabadi A, Wu AW, Venkatesan N, Duarte VM, Kang JJ, Dalgard CL, Srivastava M, Sarkar FH, Wang MB and Srivatsan ES. Liposome encapsulated curcumin-difluorinated (CDF) inhibits the growth of cisplatin resistant head and neck cancer stem cells. *Oncotarget* 2015; 6: 18504-18517.
- [227] Siddappa G, Kulsum S, Ravindra DR, Kumar VV, Raju N, Raghavan N, Sudheendra HV, Sharma A, Sunny SP, Jacob T, Kuruvilla BT, Benny M, Antony B, Seshadri M, Lakshminarayan P, Hicks W Jr, Suresh A and Kuriakose MA. Curcumin and metformin-mediated chemoprevention of oral cancer is associated with inhibition of cancer stem cells. *Mol Carcinog* 2017; 56: 2446-2460.
- [228] Liu SC, Huang CS, Huang CM, Hsieh MS, Huang MS, Fong IH, Yeh CT and Lin CC. Isoorientin inhibits epithelial-to-mesenchymal properties and cancer stem-cell-like features in oral squamous cell carcinoma by blocking Wnt/beta-catenin/STAT3 axis. *Toxicol Appl Pharmacol* 2021; 424: 115581.
- [229] Ketkaew Y, Osathanon T, Pavasant P and Soompon S. Apigenin inhibited hypoxia induced stem cell marker expression in a head and neck squamous cell carcinoma cell line. *Arch Oral Biol* 2017; 74: 69-74.
- [230] Hu FW, Tsai LL, Yu CH, Chen PN, Chou MY and Yu CC. Impairment of tumor-initiating stem-like property and reversal of epithelial-mesenchymal transdifferentiation in head and neck cancer by resveratrol treatment. *Mol Nutr Food Res* 2012; 56: 1247-1258.
- [231] Chang YC, Jan CI, Peng CY, Lai YC, Hu FW and Yu CC. Activation of microRNA-494-targeting Bmi1 and ADAM10 by silibinin ablates cancer stemness and predicts favourable prognostic value in head and neck squamous cell carcinomas. *Oncotarget* 2015; 6: 24002-24016.
- [232] Nor F, Nor C, Bento LW, Zhang Z, Bretz WA and Nor JE. Propolis reduces the stemness of head and neck squamous cell carcinoma. *Arch Oral Biol* 2021; 125: 105087.
- [233] Liao T, Kaufmann AM, Qian X, Sangvatanakul V, Chen C, Kube T, Zhang G and Albers AE. Susceptibility to cytotoxic T cell lysis of cancer stem cells derived from cervical and head and neck tumor cell lines. *J Cancer Res Clin Oncol* 2013; 139: 159-170.
- [234] Visus C, Wang Y, Lozano-Leon A, Ferris RL, Silver S, Szczepanski MJ, Brand RE, Ferrone CR, Whiteside TL, Ferrone S, DeLeo AB and Wang X. Targeting ALDH(bright) human carcinoma-initiating cells with ALDH1A1-specific CD8(+) T cells. *Clin Cancer Res* 2011; 17: 6174-6184.
- [235] Su Z, Li G, Liu C, Ren S, Tian Y, Liu Y and Qiu Y. Ionizing radiation promotes advanced malignant traits in nasopharyngeal carcinoma via activation of epithelial-mesenchymal transition and the cancer stem cell phenotype. *Oncol Rep* 2016; 36: 72-78.
- [236] Moncharmont C, Guy JB, Wozny AS, Gilormini M, Battiston-Montagne P, Ardail D, Beuve M, Alphonse G, Simoens X, Rancoule C, Rodriguez-Lafrasse C and Magne N. Carbon ion irradiation withstands cancer stem cells' migration/invasion process in head and neck squamous cell carcinoma (HNSCC). *Oncotarget* 2016; 7: 47738-47749.
- [237] Ibarra AMC, Garcia MP, Ferreira M, de Fatima Teixeira da Silva D, Pavani C, Mesquita-Ferrari RA, Fernandes KPS, Nunes FD and Rodrigues MFSD. Effects of photobiomodulation on cellular viability and cancer stem cell phenotype in oral squamous cell carcinoma. *Lasers Med Sci* 2021; 36: 681-690.
- [238] Fang CY, Chen PY, Ho DC, Tsai LL, Hsieh PL, Lu MY, Yu CC and Yu CH. miR-145 mediates the anti-cancer stemness effect of photodynamic therapy with 5-aminolevulinic acid (ALA) in oral cancer cells. *J Formos Med Assoc* 2018; 117: 738-742.
- [239] Yu CH and Yu CC. Photodynamic therapy with 5-aminolevulinic acid (ALA) impairs tumor initiating and chemo-resistance property in head and neck cancer-derived cancer stem cells. *PLoS One* 2014; 9: e87129.
- [240] Su Z, Liu D, Chen L, Zhang J, Ru L, Chen Z, Gao Z and Wang X. CD44-targeted magnetic nanoparticles kill head and neck squamous cell carcinoma stem cells in an alternating magnetic field. *Int J Nanomedicine* 2019; 14: 7549-7560.
- [241] Miyano K, Cabral H, Miura Y, Matsumoto Y, Mochida Y, Kinoh H, Iwata C, Nagano O, Saya H, Nishiyama N, Kataoka K and Yamasoba T. cRGD peptide installation on cisplatin-loaded nanomedicines enhances efficacy against locally advanced head and neck squamous cell carcinoma bearing cancer stem-like cells. *J Control Release* 2017; 261: 275-286.
- [242] Lo WL, Chien Y, Chiou GY, Tseng LM, Hsu HS, Chang YL, Lu KH, Chien CS, Wang ML, Chen YW, Huang PI, Hu FW, Yu CC, Chu PY and Chiou SH. Nuclear localization signal-enhanced RNA interference of EZH2 and Oct4 in the eradication of head and neck squamous cell carcinoma.

Cancer stem cells of head and neck squamous cell carcinoma

- ma-derived cancer stem cells. *Biomaterials* 2012; 33: 3693-3709.
- [243] Zhu M, Chen S, Hua L, Zhang C, Chen M, Chen D, Dong Y, Zhang Y, Li M, Song X, Chen H and Zheng H. Self-targeted salinomycin-loaded DSPE-PEG-methotrexate nanomicelles for targeting both head and neck squamous cell carcinoma cancer cells and cancer stem cells. *Nanomedicine (Lond)* 2017; 12: 295-315.
- [244] Sim MW, Grogan PT, Subramanian C, Bradford CR, Carey TE, Forrest ML, Prince ME and Cohen MS. Effects of peritumoral nanoconjugated cisplatin on laryngeal cancer stem cells. *Laryngoscope* 2016; 126: E184-E190.
- [245] Salvati A and Poelstra K. Drug targeting and nanomedicine: lessons learned from liver targeting and opportunities for drug innovation. *Pharmaceutics* 2022; 14: 217.
- [246] Zhang Y, Chan HF and Leong KW. Advanced materials and processing for drug delivery: the past and the future. *Adv Drug Deliv Rev* 2013; 65: 104-120.
- [247] Zhao Z, Ukidve A, Kim J and Mitragotri S. Targeting strategies for tissue-specific drug delivery. *Cell* 2020; 181: 151-167.
- [248] Mura S, Nicolas J and Couvreur P. Stimuli-responsive nanocarriers for drug delivery. *Nat Mater* 2013; 12: 991-1003.
- [249] Wang AZ, Langer R and Farokhzad OC. Nanoparticle delivery of cancer drugs. *Annu Rev Med* 2012; 63: 185-198.
- [250] Thomas OS and Weber W. Overcoming physiological barriers to nanoparticle delivery-are we there yet? *Front Bioeng Biotechnol* 2019; 7: 415.
- [251] Karthikeyan A, Senthil N and Min T. Nanocurcumin: a promising candidate for therapeutic applications. *Front Pharmacol* 2020; 11: 487.
- [252] Dinda SC and Pattnaik G. Nanobiotechnology-based drug delivery in brain targeting. *Curr Pharm Biotechnol* 2013; 14: 1264-1274.
- [253] Ibrahim H. Nanotechnology and its applications to medicine: an over view. *QJM: An International Journal of Medicine* 2020; 113: hcaa060.008.
- [254] Cho K, Wang X, Nie S, Chen Z and Shin DM. Therapeutic nanoparticles for drug delivery in cancer. *Clin Cancer Res* 2008; 14: 1310-1316.
- [255] Zhao Y, Chen H, Chen X, Hollett G, Gu Z, Wu J and Liu X. Targeted nanoparticles for head and neck cancers: overview and perspectives. *Wiley Interdiscip Rev Nanomed Nanobiotechnol* 2017; 9.
- [256] Inagaki FF, Furusawa A, Choyke PL and Kobayashi H. Enhanced nanodrug delivery in tumors after near-infrared photodynamic therapy. *Nanophotonics* 2019; 8: 1673-1688.
- [257] Wojtynek NE and Mohs AM. Image-guided tumor surgery: the emerging role of nanotechnology. *Wiley Interdiscip Rev Nanomed Nanobiotechnol* 2020; 12: e1624.
- [258] Zhang W, Taheri-Ledari R, Ganjali F, Afruzi FH, Hajizadeh Z, Saeidrad M, Qazi FS, Kashtiaray A, Sehat SS, Hamblin MR and Maleki A. Nanoscale Bioconjugates: a review of the structural attributes of drug-loaded nanocarrier conjugates for selective cancer therapy. *Heliyon* 2022; 8: e09577.
- [259] Bertrand N, Wu J, Xu X, Kamaly N and Farokhzad OC. Cancer nanotechnology: the impact of passive and active targeting in the era of modern cancer biology. *Adv Drug Deliv Rev* 2014; 66: 2-25.
- [260] Pawar PV, Domb AJ and Kumar N. Systemic targeting systems-EPR effect, ligand targeting systems. *Focal Controlled Drug Delivery*. Springer; 2014. pp. 61-91.
- [261] Kudr J, Haddad Y, Richtera L, Heger Z, Cernak M, Adam V and Zitka O. Magnetic nanoparticles: from design and synthesis to real world applications. *Nanomaterials (Basel)* 2017; 7: 243.
- [262] Hirsjarvi S, Passirani C and Benoit JP. Passive and active tumour targeting with nanocarriers. *Curr Drug Discov Technol* 2011; 8: 188-196.
- [263] Maibaum L, Dinner AR and Chandler D. Micelle formation and the hydrophobic effect. *J Phys Chem B* 2004; 108: 6778-6781.
- [264] Piao L, Zhang M, Datta J, Xie X, Su T, Li H, Teknos TN and Pan Q. Lipid-based nanoparticle delivery of Pre-miR-107 inhibits the tumorigenicity of head and neck squamous cell carcinoma. *Mol Ther* 2012; 20: 1261-1269.
- [265] Puri A, Loomis K, Smith B, Lee JH, Yavlovich A, Heldman E and Blumenthal R. Lipid-based nanoparticles as pharmaceutical drug carriers: from concepts to clinic. *Crit Rev Ther Drug Carrier Syst* 2009; 26: 523-80.
- [266] Sheikhholeslami B, Lam NW, Dua K and Haghi M. Exploring the impact of physicochemical properties of liposomal formulations on their in vivo fate. *Life Sci* 2022; 300: 120574.
- [267] Shah M, Murad W, Mubin S, Ullah O, Rehman NU and Rahman MH. Multiple health benefits of curcumin and its therapeutic potential. *Environ Sci Pollut Res Int* 2022; 29: 43732-43744.
- [268] Mahendiran B, Azeez NA, Muthusamy S and Krishnakumar GS. Polymer-based bionanomaterials for targeted drug delivery. *Fundamentals of Bionanomaterials*. Elsevier; 2022. pp. 241-271.
- [269] Lan JS, Liu L, Zeng RF, Qin YH, Hou JW, Xie SS, Yue S, Yang J, Ho RJY, Ding Y and Zhang T. Tumor-specific carrier-free nanodrugs with GSH depletion and enhanced ROS generation for endogenous synergistic anti-tumor by a chemotherapy-photodynamic therapy. *Chemical Engineering Journal* 2021; 407: 127212.

Cancer stem cells of head and neck squamous cell carcinoma

- [270] Lang L, Shay C, Xiong Y, Thakkar P, Chemmalakuzhy R, Wang X and Teng Y. Combating head and neck cancer metastases by targeting Src using multifunctional nanoparticle-based saccatinib. *J Hematol Oncol* 2018; 11: 85.
- [271] Chen FA, Kuriakose MA, Zhou MX, DeLacure MD and Dunn RL. Biodegradable polymer-mediated intratumoral delivery of cisplatin for treatment of human head and neck squamous cell carcinoma in a chimeric mouse model. *Head Neck* 2003; 25: 554-560.
- [272] Argiris A, Karamouzis MV, Raben D and Ferris RL. Head and neck cancer. *Lancet* 2008; 371: 1695-1709.
- [273] Ward BB, Dunham T, Majoros IJ and Baker JR Jr. Targeted dendrimer chemotherapy in an animal model for head and neck squamous cell carcinoma. *J Oral Maxillofac Surg* 2011; 69: 2452-2459.
- [274] Zhang Z, Zhuang L, Lin Y, Yan M, Lv J, Li X, Lin H, Zhu P, Lin Q and Xu Y. Novel drug delivery system based on hollow mesoporous magnetic nanoparticles for head and neck cancers-targeted therapy in vitro and in vivo. *Am J Cancer Res* 2020; 10: 350-364.
- [275] Luo K, Chen H, Zhou Q, Yan Z, Su Z and Li K. Corrigendum to "A facile one step solvothermal controllable synthesis of FeS(2) quantum dots with multiple color emission for the visual detection of aconitine" [*Spectrochim. Acta A Mol. Biomol. Spectrosc.* 240 (2020) 118563]. *Spectrochim Acta A Mol Biomol Spectrosc* 2021; 247: 119046.
- [276] Moniruzzaman M, Anantha Lakshmi B, Kim S and Kim J. Preparation of shape-specific (trilateral and quadrilateral) carbon quantum dots towards multiple color emission. *Nanoscale* 2020; 12: 11947-11959.
- [277] Akerman ME, Chan WCW, Laakkonen P, Bhatia SN and Ruoslahti E. Nanocrystal targeting in vivo. *Proc Natl Acad Sci U S A* 2002; 99: 12617-12621.
- [278] Abbasi E, Aval SF, Akbarzadeh A, Milani M, Nasrabadi HT, Joo SW, Hanifehpour Y, Nejati-Koshki K and Pashaei-Asl R. Dendrimers: synthesis, applications, and properties. *Nanoscale Res Lett* 2014; 9: 247.
- [279] Hossen S, Hossain MK, Basher MK, Mia MNH, Rahman MT and Uddin MJ. Smart nanocarrier-based drug delivery systems for cancer therapy and toxicity studies: a review. *J Adv Res* 2018; 15: 1-18.
- [280] Nan W and Mingyu W. The Effect of sevoflurane on the proliferation, epithelial-mesenchymal transition (EMT) and apoptosis in human breast cancer cells. *J Biol Regul Homeost Agents* 2022; 36: 583-592.
- [281] Palmerston Mendes L, Pan J and Torchilin VP. Dendrimers as nanocarriers for nucleic acid and drug delivery in cancer therapy. *Molecules* 2017; 22: 1401.
- [282] Mittal P, Saharan A, Verma R, Altalbawy FMA, Alfaidi MA, Batiha GE, Akter W, Gautam RK, Uddin MS and Rahman MS. Dendrimers: a new race of pharmaceutical nanocarriers. *Biomed Res Int* 2021; 2021: 8844030.
- [283] Moraes Silva S, Tavallaie R, Sandiford L, Tilley RD and Gooding JJ. Gold coated magnetic nanoparticles: from preparation to surface modification for analytical and biomedical applications. *Chem Commun (Camb)* 2016; 52: 7528-7540.
- [284] Ambekar RS, Choudhary M and Kandasubramanian B. Recent advances in dendrimer-based nanopatform for cancer treatment: a review. *European Polymer Journal* 2020; 126: 109546.
- [285] Tomalia DA. Birth of a new macromolecular architecture: dendrimers as quantized building blocks for nanoscale synthetic polymer chemistry. *Prog Polym Sci* 2005; 30: 294-324.
- [286] Bhadra D, Bhadra S, Jain S and Jain NK. A PEGylated dendritic nanoparticulate carrier of fluorouracil. *Int J Pharm* 2003; 257: 111-124.
- [287] Tripathi PK and Tripathi S. 6 - Dendrimers for anticancer drug delivery. In: Chauhan A, Kulkarni H, editors. *Pharmaceutical Applications of Dendrimers*. Elsevier; 2020. pp. 131-150.
- [288] Kesharwani P, Jain K and Jain NK. Dendrimer as nanocarrier for drug delivery. *Prog Polym Sci* 2014; 39: 268-307.
- [289] Kannan RM, Nance E, Kannan S and Tomalia DA. Emerging concepts in dendrimer-based nanomedicine: from design principles to clinical applications. *J Intern Med* 2014; 276: 579-617.
- [290] Xu L and Yang H. Folate-decorated polyamidoamine dendrimer nanoparticles for head and neck cancer gene therapy. *RNA Interference and Cancer Therapy*. Springer; 2019. pp. 393-408.
- [291] Mignani S, El Brahm N, Eloy L, Poupon J, Nicolas V, Steinmetz A, El Kazzouli S, Bousmina MM, Blanchard-Desce M, Caminade AM, Majoral JP and Cresteil T. Anticancer copper (II) phosphorus dendrimers are potent proapoptotic Bax activators. *Eur J Med Chem* 2017; 132: 142-156.
- [292] Needham D and Dewhirst MW. The development and testing of a new temperature-sensitive drug delivery system for the treatment of solid tumors. *Adv Drug Deliv Rev* 2001; 53: 285-305.
- [293] Pfister DG, Spencer S, Adelstein D, Adkins D, Anzai Y, Brizel DM, Bruce JY, Busse PM, Caudell JJ, Cmelak AJ, Colevas AD, Eisele DW, Fenton M, Foote RL, Galloway T, Gillison ML, Haddad RI, Hicks WL, Hitchcock YJ, Jimeno A, Leizman

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- D, Maghami E, Mell LK, Mittal BB, Pinto HA, Ridge JA, Rocco JW, Rodriguez CP, Shah JP, Weber RS, Weinstein G, Witek M, Worden F, Yom SS, Zhen W, Burns JL and Darlow SD. Head and neck cancers, version 2.2020, NCCN clinical practice guidelines in oncology. *J Natl Compr Canc Netw* 2020; 18: 873-898.
- [294] Badr H, Herbert K, Chhabria K, Sandulache VC, Chiao EY and Wagner T. Self-management intervention for head and neck cancer couples: results of a randomized pilot trial. *Cancer* 2019; 125: 1176-1184.
- [295] Chloupek A, Kania J and Jurkiewicz D. Concordance between clinical and pathological T and N stages in polish patients with head and neck cancers. *Diagnostics (Basel)* 2023; 13: 2202.
- [296] Senchak JJ, Fang CY and Bauman JR. Interventions to improve quality of life (QOL) and/or mood in patients with head and neck cancer (HNC): a review of the evidence. *Cancers Head Neck* 2019; 4: 2.
- [297] Khan MM, Manduchi B, Rodriguez V, Fitch MI, Barbon CEA, McMillan H, Hutcheson KA and Martino R. Exploring patient experiences with a telehealth approach for the PRO-ACTIVE trial intervention in head and neck cancer patients. *BMC Health Serv Res* 2022; 22: 1218.
- [298] Thilges S, Mumby P, Sinacore J, Clark J and Czerlanis C. Implementing a cognitive behavioral intervention for patients with head and neck cancer. *Support Care Cancer* 2023; 31: 476.
- [299] Graboyes EM, Sterba KR, Li H, Warren GW, Alberg AJ, Calhoun EA, Nussenbaum B, McCay J, Marsh CH, Osazuwa-Peters N, Neskey DM, Kaczmar JM, Sharma AK, Harper J, Day TA and Hughes-Halbert C. Development and evaluation of a navigation-based, multilevel intervention to improve the delivery of timely, guideline-adherent adjuvant therapy for patients with head and neck cancer. *JCO Oncol Pract* 2021; 17: e1512-e1523.
- [300] Head BA, Keeney C, Studts JL, Khayat M, Bumpous J and Pfeifer M. Feasibility and acceptance of a telehealth intervention to promote symptom management during treatment for head and neck cancer. *J Support Oncol* 2011; 9: e1-e11.
- [301] McNeely ML. Exercise as a promising intervention in head & neck cancer patients. *Indian J Med Res* 2013; 137: 451-453.

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Table S1. Chemical agents and their targets in CSCs of HNC

Cetuximab and Erlotinib	EGFR	Decrease percentage of CSCs, induce differentiation.	[1]
Afatinib	EGFR	Afatinib alone or with radiotherapy decreased CSC population.	[2]
	EGFR	Inducing severe apoptosis and an uncommon weak protective autophagic response preferentially in stem-like HNSCC cells.	[3]
ABT-737	Bcl-2	Alone or in combination with radiation, can efficiently eliminate CSCs.	[4]
cetuximab plus ABT-199 with fractional irradiation	EGFR/Bcl-2	The combination significantly inhibited proliferation, invasion/migration, and resistance to apoptosis of HNSCC CSCs in vitro and strongly reduced the tumor growth and increased in vivo survival without side effects.	[5]
C1632 and metformin	Lin28	The combined treatment exerts synergistic anti-tumor effects in OSCC cell lines and xenograft tumor growth in vivo.	[6]
Ibrutinib	BTK	Reduced CSCs number and increase DDP sensitivity of OSCC SP-derived cells.	[7]
CDHP and GSK3 β inhibitors	GSK3 β	Markedly enhanced 5-FU-induced apoptosis of CD44(high)/ESA(low) cells.	[8]
cisplatin and XAV-939	tankyrase	Synergistically abrogate chemoresistance by increasing DNA damage.	[9]
LF3	Wnt/ β -catenin signals	The self-renewal capacity of CSCs was blocked by LF3, as examined by sphere formation.	[10]
sFRP4	Wnt signaling	Decreased expression of CSCs markers (CD44 and ALDH), and inhibit proliferation, EMT and enhanced chemosensitivity of HNSCC CSCs.	[11]
SAHA and TSA	HDAC	Inhibit CSCs marker expression and change stemness genes.	[12]
SAHA	HDAC	Significantly decreased tumorsphere formation of DDP resistant cell lines.	[13]
VPA	HDAC	Inhibit self-renewal, CSC marker expression and potentiated the cytotoxic effect of cisplatin.	[14]
Entinostat	HDAC	Induce cycle arrest (G0/G1 phase), tumor apoptosis and increase in ROS production and significant reductions in CSCs.	[15]
DMF and BSO	GSH	Transient GSH depletion triggered radiation-induced cell death in CSCs.	[16]
UCN-01 and ATRA	Chk1	Decreased the surviving fraction of SQ20B-CSCs after photon irradiation and carbon ions.	[17]
MEDI5117	IL-6	Low dose MEDI5117 decrease CSCs fraction in 3 low-passage patient-derived xenograft (PDX) models of HNSCC, prevented tumor recurrence in a clinical trial.	[18]
PTC209	BMI-1	Enable immune checkpoint blockade to inhibit metastatic tumor growth and prevent tumor relapse by activating cell-intrinsic immunity, in addition to eliminating CSCs.	[19]
celecoxib	COX-2	Suppress messenger RNA expression of stemness-related genes and sphere formation.	[20]

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Table S2. Natural products and targets in HNSCC

Compounds of natural herbs	Target point	Effects	Ref
Ovatodioliide (ova)	p-FAK, p-PXN, F-actin, Slug proteins, SOX2, OCT4 and JAK-STAT signaling pathway	Ova inhibited HNSCC tumorsphere formation and attenuated HNSCC stem cell tumorigenicity, inhibited tumor growth.	[21]
Epigallocatechin-3-gallate (EGCG)	Oct4, Sox2, Nanog, CD44, ABCG2 and Notch signaling	EGCG inhibits the self-renewal capacity of HNSC CSCs by suppressing their sphere forming capacity through suppression of Notch pathway.	[22]
quercetin	Twist, N-cadherin, and vimentin	Reduce self-renewal property and stemness signatures expression in head and neck cancer-derived sphere cells.	[23]
A new gamboge derivative compound 2 (C2)	CD49f, CD133, CD44, Ki-67, phosphor-EGFR, CD49f and CD133	Effectively suppresses the growth of CSCs and the formation of tumor spheres.	[24]
Cucurbitacin I	STAT3, JAK2, Bcl-2, Bcl-xL, and survivin	Can effectively inhibit the expression of p-STAT3 and capacities for tumorigenicity, sphere formation, and radio resistance in HNSCC-CD44(+) ALDH1(+).	[25]
YMGKI-1 and YMGKI-2	STAT3 and Src, phosphor-mTOR, HER2, phosphor-EGFR, Phosphatidylinositol 3-kinases (PI3K), phosphor-p44/42 MAPK (Thr202/Tyr204), and phosphor-AMPK	Inhibited stemness (specifically to HNSCC CSCs), decreased expression of CSC markers and promoted radiosensitivity of HNSCC CSCs.	[26, 27]
Plumbagin	Multiple targets (seen in ref)	Inhibit stemness, EMT and induce MET.	[28]
SVC112	Myc, Cyclin D1, Myc and Sox2. SVC112	Inhibits tumor sphere growth in vitro, decrease CSCs number in vivo.	[29]
Lovastatin	CD44, SOX2, c-Myc, CBFb, and Snail	Inhibit CSCs properties and enhance sensitivity of CSCs to therapy.	[30]
Tetrandrine	upregulating Bax and caspase-3 and downregulating Bcl-2	Inhibited the viability of CD133 Hep-2 cells.	[31]
Isoliquiritigenin	ALDH1, CD44, ABCG2, GRP78	Hinder self-renewal, decrease activity of ALDH1 and CD44.	[32]
sulforaphane	SHH, SOX2 and OCT4	SF inhibit HNSCC-CSC viability alone or combined with CIS or 5-FU.	[33]
Liposome encapsulated curcumin-difluorinated (CDF)	CD44	Liposome encapsulated CDF inhibit CD44 ^{hi} CSCs growth in vitro and vivo.	[34]
Curcumin and metformin		Inhibited the migratory and self-renewal properties of CSCs.	[35]
Isorientin	p-STAT3, Wnt/ β -catenin, p-GSK3, TCF1/TCF7 and LEF1	Targeting OSCC-SC-mediated stemness via blocking Wnt/ β -catenin/STAT3 axis.	[36]
Apigenin	CD44, NANOG, and CD105	Down-regulate expressions of CSCs markers, CD44, NANOG, and CD105 of HNSCC cells and the number of cells expressing CSCs markers under hypoxia.	[37]
Resveratrol	ALDH1, CD44, Oct4, Nanog, and Nestin	Reduced CSCs properties in vitro and in vivo via regulating CSCs markers, stemness-related gene signatures and EMT markers.	[38]
silibinin	miR-494-inhibiting Bmi1/ADAM10 expression	Reduced CSCs stemness via activation of miR-494-inhibiting Bmi1/ADAM10 expression.	[39]
Propolis		Reduce CSCs numbers and decrease CSCs markers specifically.	[40]

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Table S3. HNSCC markers

Factor	Sample	Identification of HNSCC CSCs	Pathways	Effects	Ref
PI3K	cells derived from HNSCC patient-derived xenografts (PDXs)	by marker (ALDH and CD44)	PI3K upregulates SOX2, SOX2 activate ALDH1A1 and induce CDH1.	CSCs (HPV-positive and HPV-negative) are resistant to standard therapy but are particularly susceptible to PI3K inhibition.	[41]
	Human OSCC cell lines	GO-like OSCC cells		Regulate transition of JARID1B+ CSCs to GO-like cells.	[42]
Hyaluronan (HA)			HA/CD44 signaling.	Stimulate a variety of CSC functions including self-renewal, clone formation and differentiation.	[43]
	Tumor-derived HSC-3 cell line	By marker (ALDH and CD44)	Stimulates the CD44v3 (an HA receptor) interaction with Oct4-Sox2-Nanog leading to both a complex formation and the nuclear translocation of three CSC transcription factors. Suppression of several epigenetic regulators (AOF1/AOF2 and DNMT1) and the up-regulation of several survival proteins (cIAP-1, cIAP-2, and XIAP) leading to self-renewal, clonal formation, and cisplatin resistance by miR-302.	The acquisition of cancer stem cell properties, including self-renewal, clonal formation, and chemotherapy resistance in HA-CD44v3-activated head and neck cancer.	[44]
Snail	HNSCC cell lines	by marker (ALDH and CD44)		Snail-induced EMT gained CSC-like phenotype and was associated with increased chemoresistance.	[45]
	HNSCC cell lines	by marker (ALDH and CD44)/sphere formation	snail.	Induce EMT, chemoresistance and invasive ability and maintained the CSC-like phenotype.	[46]
	Tongue squamous cell carcinoma cell lines	Colony-forming assay/by CSC marker		Induce EMT and promote CSC-like traits.	[47]
	NPC cell lines	by marker CD44 and CD133, sphere formation		Snail mediated a CSC-like phenotype.	[48]
HPV16	HPV-negative OSCC cell lines	Sphere formation/by marker ALDH1+	HPV16/miR-181a/d axis.	Increase self-renewal and enhance CSC-related factor expression.	[49]
	HNSCC cell lines	by marker (CD44 ^{high} and EpCAM ^{low})	miR-1281 and miR3194-5p.	HPV16-E6E7 lead to an increase of the migratory CSCs.	[50]
	Human HPV-negative and -positive HNSCC cell lines	Cells with low proteasome activity (expressing the C-terminal degon of murine ODC (cODC) fused to a green fluorescent protein)		RT can dedifferentiate HNSCC cells into CSCs; and radiation-induced dedifferentiation depends on the HPV status of the tumor.	[51]
MELK	HNSCC cell lines	sphere formation	MELK inhibition downregulates SOX2.	MELK plays a key role in CSCs property through the regulation of SOX2.	[52]
Renin-angiotensin system (RAS)	MDHNSCC tissue samples and MDHNSCC-derived primary cell lines	by stemness-related gene expression		RAS are expressed by the oct4+ and SOX2+ cells within the tumor nests (TNs) and the peritumoral stroma (PTS).	[53]
	buccal SCC tissue samples and -derived primary cell lines	by stemness-related gene expression		RAS are expressed by the oct4+ and SALL4+ in buccal SCC within the tumor nests (TNs) and the peritumoral stroma (PTS).	[54]
	MDOTSCC tissue samples	IHC staining for stemness-related gene expression		expression of components of the RAS by the CSC subpopulations within MDOTSCC.	[55]
	MHNCSCC tissue samples and MHNCSCC-derived primary cell lines	SOX2+ cells as CSCs		Expression of components of the RAS by the CSC subpopulations within MHNCSCC.	[56]

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SPLUNC1 and MLL3	NPC mouse model using C666-1 cells	by SOX2 expression	Downregulated expression of SPLUNC1, increased expression of MLL3.	Downregulated expression of SPLUNC1, increased expression of MLL3 is corrected with stage of NPC.	[57]
XIAP	Human NPC S-18 and S-26 cell lines	SP cells; CD44+ cells; by stemness-related gene expression	Blocked autophagic degradation of Sox2 by inhibiting ERK1 activation in CSCs, positively correlated with Sox2 expression.	Maintain CSCs, antagonist of IAPs combined with DDP/5-FU exerted antitumor effect on CSCs.	[58]
BMI1	HNSCC induced from mouse model, HNSCC cell lines	BMI+ HNSCC cells	BMI increased AP-1 activities.	BMI1+ CSCs drives invasive growth and metastasis of HNSCC, Inhibiting AP-1 or BMI1 sensitized tumors to DDP, and eliminated lymph node metastases.	[59]
p38 MAPK	HNSCC tissue samples, laryngeal Hep-2 cell lines	by marker CD133		BMI1 was required to maintain the proliferative capacity of laryngeal CD133 CSCs.	[60]
	HNSCC tissue samples, cell lines	by CSC marker SOX2, Oct4, Klf4, c-Myc and CD44		p38 MAPK activation may play a role in therapeutic resistance and disease relapse in HNSCC by maintenance of CSCs phenotype.	[61]
Wnt/ β -catenin	HNSCC cell lines, primary HNSCC tissues	by marker (ALDH and CD44)/sphere formation	β -catenin directly regulates Oct4 transcription in HNSCC stem-like cells.	Maintains self-renewal and tumorigenicity of head and neck squamous cell carcinoma stem-like cells.	[62]
SOX8	Cisplatin-resistant tongue squamous cell carcinoma (TSCC) cells lines	Sphere formation/by stemness-related gene expression	SOX8 could bind to the promoter region of FZD7 and activated FZD7-mediated Wnt/ β -catenin pathway.	Induce chemoresistance, stem-like properties and EMT.	[63]
SOX2	HNSCC cell lines, Primary sphere cells from surgical specimens from HNSCC patients	by CSC marker/sphere formation/by stemness-related gene expression	Induce ABCG2, snail, cyclin B1 expression.	SOX2 enhanced stemness and enhance chemoresistance of CSCs.	[64]
RARS-MAD1L1	NPC cell lines	SP cells, by stemness-related gene expression	RARS-MAD1L1 interacted with AIMP2, which resulted in activation of FUBP1/c-Myc pathway.	RARS-MAD1L1 might contribute to tumorigenesis, CSC-like properties, and therapeutic resistance.	[65]
LIN28B	Dissociated cells derived from the samples of OSCC patients	by marker (ALDH and CD44)/sphere formation	ARID3B and HMGA2 as direct targets of LIN28B/Let7, mediating Oct4 and Sox2 expression by direct regulation of Oct4 and Sox2 promoter activity, respectively.	Lin28B(HIGH)/Let7(LOW) regulates key cancer stem-like properties in oral squamous cancers.	[66]
HSP90	HNSCC cell lines	by marker (ALDH and CD44)/sphere formation		HSP90 inhibitor decrease CSC stemness.	[67]
5T4	HNSCC cell lines	by marker (ALDH and CD44)		5T4 is more highly expressed in head and neck CSC, inhibitor caused a significant reduction in the CSC fraction.	[68]
c-Met	Cell lines from HNSCC tumor tissue	by marker (ALDH and CD44)/sphere formation/SP cells/by stemness-related gene expression		c-met upregulate ALDH1 and ABCG2 activity and associated with stemness as well as chemoresistance.	[69]
	Cell lines were obtained at the primary surgery before any treatment	Mouse HNSCC xenograft model/sphere formation	Increased expression of self-renewal pathways.	c-Met could serve as a novel single marker for CSCs at least in HNSCC.	[70]
	Single-cell suspensions derived from primary specimens	by CSC marker (c-Met and CD44)	Due to downregulation of Wnt/ β -catenin signaling in HN-CSC and that the Wnt pathway receptor FZD8 was essential for interactions of c-Met and Wnt/ β -catenin signaling in HN-CSC.	Silencing c-Met is sufficient to suppress sphere formation, tumor initiation, and metastatic properties of HN-CSC.	[71]
MACC1	HNSCC cell lines	by marker ALDH1		MACC-1 gene overexpress in CSCs compared with cancer cells.	[72]

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The Hippo-TAZ	HNSCC cell lines	by marker (CD133 and CD44)/stemness-related gene expression	SOX2 as a putative downstream target of TAZ.	Promote CSCs maintenance and tumorigenicity in HNSCC.	[73]
	OSCC cell lines	the colony formation, tumorsphere assays, by marker CD44 and CD133	TAZ-TEADs binding and subsequent transcriptional activation of EMT mediators and pluripotency factors are presumably responsible for TAZ-mediated EMT and non-CSCs-to-CSCs conversion.	TAZ is required for oral CSCs self-renewal and maintenance and endow non-CSCs with CSCs-like traits.	[74]
Oct-4	Cell lines from HNSCC tumor tissue	by CSC marker		Oct-4a is a stemness marker and oct-4a+ cells have ability of chemoresistance.	[75]
	HNSCC cell lines	by marker (ALDH and CD44)/sphere formation/SP cells	Regulate the cell cycle checkpoint kinases CHK1 and WEE1 and homologous recombination (HR) repair genes PSMC3IP and RAD54L.	Loss and overexpression of Oct4 may lead to tumor cell radio sensitization, Oct4-regulated genes contribute to the HNSCC radio resistance by modulating the DNA repair and CSC properties.	[76]
RXR α	hep-2 and fadu cells	by marker (CD133 and CD44)/sphere formation/SP cells/by stemness-related gene expression		Overexpression of RXR α was able to expand the CSC-like properties in HNSCC cells.	[77]
IGF-R and EGFR	Cell lines were obtained at the primary surgery before any treatment	Sphere formation/by marker (ALD)		Using specific inhibitors against EGFR and IGF-1R reduced stem cell fractions drastically.	[78]
EGFR	Tongue epithelial squamous cell carcinoma cell line	Sphere formation	SOX2 was a binding partner and substrate of EGFR.	EGFR plays an important role in the development of cancer stem cells.	[79]
	HNSCC cell lines	by marker (CD44)/sphere formation/by stemness-related gene expression	The induction of CD44, BMI-1, Oct-4, NANOG, CXCR4, and SDF-1.	EGFR plays critical roles in the survival, maintenance, and function of cancer stem cells.	[80]
Notch1	HNSCC cell lines	by CSC marker/sphere formation/by stemness-related gene expression	Notch1 acted upstream of canonical Wnt signaling in HNSCC cells and regulate CSCs markers as well as ABC transporters.	Notch1 signaling contributes to stemness.	[81]
	HNSCC cell lines CAL27 and FaDu, Human HNSCC tissue	by self-renewal-related markers, CD44, BMI1, SOX2, ALDH1, and Slug/sphere formation/by marker (CD44 CD133)		NOTCH1 is associated with CSCs in HNSCC tissue and NOTCH1 inhibition delays tumorigenesis and effectively reduces CSC self-renewal of nude mice HNSCC xenograft. Furthermore, chemotherapeutically combined NOTCH1 inhibitor synergistically attenuated chemotherapy-enriched CSC population in vitro and in vivo, which provides the possibility to effectively eliminate head and neck CSCs.	[82]
	TSCC cell lines	by marker (ALDH1, CD133 and CD44)/sphere formation		NOTCH1 is required for stemness of TSCC tumor cells.	[83]
DOT1L	Tumor-derived HSC-3 cell line (isolated from human squamous carcinoma cells of the mouth)	by marker (ALDH and CD44)	Histone methyltransferase, DOT1L-associated epigenetic changes induced by HA play pivotal roles in miR-10 production leading to up-regulation of RhoGTPase and survival proteins.	Acquisition of cancer stem cell properties including self-renewal, tumor cell invasion, and chemotherapy resistance.	[84]
NLRP3 inflammasome	SCCHN cell lines	by marker (BMI1, ALDH and CD44)/sphere formation/by stemness-related gene expression	Activated by LPS and ATP and upregulate of BMI1, ALDH1 and CD44.	CSCs self-renewal activation in SCCHN.	[85]
TRAF6	HNSCC cell lines	by EMT marker (Vimentin and Slug) and CSC marker (CD44, ALDH1, KLF4, SOX2 and AGR2)/sphere formation	Associated with CD44, ALDH1, KLF4 and SOX2 expression, the anchor-dependent colony formation number and sphere number formed.	TRAF6 plays a role in EMT phenotypes, the generation and maintenance of CSCs in SCCHN.	[86]

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GRP78	HNSCC cell lines	by marker (CD133, ALDH and Cripto-1)/sphere formation/SP/by stemness-related gene expression		Induce self-renewal ability, side population cells and expression of stemness genes.	[87]
	HNSCC cell lines	CD24(-) CD44(+) cells	Regulating CSC related factors (Oct-4 and Slug) and EMT related factors (CK18 and involucrin).	Grp78 regulated the conversion of CD24(-) CD44(+) cells, a characteristic of HNC stem cells.	[88]
S100A4	HNSCC cell lines	Sphere formation	Mediated by repressing p53 and subsequently activating the Nanog expression.	Both the calcium-binding ability and the C-terminal region of S100A4 are important for HN-CICs to sustain its stemness property and malignancy.	[89]
	HNSCC cell lines	Sphere formation/SP cells	Induce Notch2 and PI3K (phosphoinositide 3-kinase)/pAKT.	Maintaining the stemness properties and tumorigenicity of CSCs.	[90]
RhoC	HNSCC cell lines derived from patients with T2N0 of floor of the mouth and T3N1 of base of the tongue respectively	by marker (ALDH and CD44)/sphere formation	by overexpressing IL-6 and phosphorylation of STAT3.	Regulation and activation of stem cell transcription factors.	[91]
GSK3 β	SCC cell lines	Sphere formation/by marker CD44 and ESA	Regulate Oct4, Sox2, and Nanog and reversely regulate Calgranulin B and Involucrin.	Maintenance of stem cell self-renewal.	[92]
c-Fos	HNSCC cell lines	Sphere formation	Increased the expression of pERK and cyclin D1.	Enhanced the epithelial-mesenchymal transition (EMT) state and expression of CSC markers (Nanog, c-Myc, Sox2, and Notch1).	[93]
G9a	HNSCC cell lines	by marker (CD44)/sphere formation	Interacts with Snail and mediates Snail-induced transcriptional repression of E-cadherin and EMT, through methylation of histone H3 lysine-9 (H3K9).	EMT-mediated metastasis and maintenance of cancer stem cell-like characters.	[94]
HDAC	HNSCC cell lines	by marker (ALDH)/sphere formation		Inhibition of HDAC may constitute a novel strategy to disrupt the population of CSC in head and neck tumors to create a homogeneous population of cancer cells with biologically defined signatures and predictable behavior.	[95]
SASP	Mouse model	by CSC marker	The effects of chemokines were primarily mediated by PI3K signaling.	The telomere DDR regulates senescence-associated paracrine interactions between cancer stem cell populations, dramatically affecting tumor progression and metastasis.	[96]
sLeX	Primary and metastatic HNSCC cell lines	by marker (ALDH and CD44)/sphere formation		sLeX relationship with CSC in HNSCC.	[97]
fucosylation	HNSCC cell lines	sphere formation	Fucosyl-transferases FUT3 and FUT6 highly up-regulated, increased sLex expression and increased adhesion by shear flow assays in orospheres.	Fucosylation function to spheres formation, fucosylation is of paramount importance in the invasion and metastatic process of CSCs.	[98]
CD200	HNSCC tumor cell lines	by stemness-related gene expression (shh and BMI1)	CD200 was diversely expressed and consistently associated with expression of Bmi-1 and Shh. Overexpression of CD200 induced Bmi-1 and Shh.	CD200 was related to CSC features.	[99]
CK2	HNSCC tumor cell lines	SP cells/by stemness-related gene expression (Nanog, Oct4 and Sox2)/sphere formation	CK2 is associated with stem cell gene.	Aberrant CK2 signaling inhibits TAp73 to promote the expression of CSC genes and phenotype.	[100]
CD10	HNSCC tumor cell lines	by marker (ALDH, CD133 and CD44)/sphere formation/by stemness-related gene expression (OCT3/4)	CD10-positive subpopulation expressed the CSC marker OCT3/4 at a higher level, CD10-positive subpopulation was more refractory to cisplatin, fluorouracil and radiation.	CD10 is associated with therapeutic resistance and CSC-like properties of HNSCC.	[101]

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SDF-1 α /CXCR4	HNSCC tissue	by marker CD44		SDF-1-CXCR4 axis may play an important role in the cancer stem cell theory of HNSCC.	[102]
PinX1	Nasopharyngeal cancer cell line CNE2	by CSC marker (CD133)	Possibly through TRF1, Mad1/c-Myc, and p53-mediated pathways.	PinX1 downregulates telomerase activity in CD133 ⁺ CSCs, inhibits its proliferation, migration, and invasion, and induces apoptosis.	[103]
	Nasopharyngeal cancer cell line CNE2	by CSC marker (CD133)	by regulating the P53/miR-200b-mediated transcriptional suppression of Snail1, Twist11, and Zeb1.	Overexpression of PinX1 and P53 could inhibit nasopharyngeal CD133 ⁺ CSC proliferation, migration, and invasion.	[104]
SMURF1	HNSCC cell lines	Sphere formation/by marker (ALDH and CD44)	Inhibition of BMP signaling potentiates the long-term survival of HNSCC CSCs.	The maintenance and progression of CSCs.	[105]
PLOD2	FaDu and Hep2 cell line	Sphere formation/by marker (CD133 and CD44)/SP cell	Activate Wnt pathway.	PLOD2 contributes to the CSC-like properties of Hep-2 and FaDu cells.	[106]
GLI3	Tongue squamous cell carcinoma cell lines	by marker CD44	<i>GLI3</i> gene silencing resulted in a significant decrease in <i>CD44</i> , <i>BMI1</i> , <i>POU5F1</i> (<i>OCT4</i>) and <i>SNAI2</i> (<i>SLUG</i>), IVL and S100A9 (Calgranulin B).	GLI3 contributes to OSCC stemness and malignant behavior.	[107]
ZEB1/ZEB2	Cell lines from HNSCC tumor tissue	by marker CD133		CSC-like properties, including self-renewal ability, the expression of stemness markers, and drug resistance.	[108]
Id2	HNSCC cell lines	Sphere formation/by marker (CD44)/by stemness-related gene expression		Id2 associated with stemness of HNSCC cells.	[109]
BMP4	OSCC cell lines	by EMT and stemness-related gene expression (E-cadherin, Snail, Slug, Bmi-1, CD44, hTERT)	Regulation of CSC markers.	BMP-4 (bone morphogenetic protein-4) could induce epithelial-mesenchymal transition (EMT) with acquisition of stem cell-like phenotypes in a cell-culture model.	[110]
ISG15	NPC cells lines	The colony formation, tumor sphere assays, by stemness-related gene expression (BMI1, c-MYC, NANOG, and KLF4)	Expression levels of pluripotency-associated genes, including BMI1, c-MYC, NANOG, and KLF4 increased.	ISG15 promotes CSC phenotype and radiation and chemotherapy resistance in NPC.	[111]
EVI1	NPC cell lines	SP, sphere formation, by marker PKH26 ⁺ and ALDH1 ⁺ , by stemness-related gene expression (SOX2, Nanog, c-Myc, and Oct4)	EVI1, snail, and HDAC1 formed a co-repressor complex to repress E-cadherin expression; EVI1 directly bound at β -catenin promoter and activated its expression.	β -catenin mediated EVI1's function on cancer stem cells (CSCs) properties: the sphere formation ability and the pluripotent transcription factors (SOX2, Nanog, c-Myc).	[112]
STAT3	HNSCC cell lines	by self-renewal-related markers, ALDH1, CD44, OCT4 and SOX2/sphere formation		p-stat3 corrected with CSC property.	[113]
Skp2	NPC cell lines AND NPC specimens	by marker ALDH1 and sphere formation		Knockdown of Skp2 partially reduced cell proliferation, promoted cellular senescence, and decreased the population of stem cell like aldehyde dehydrogenase 1 positive NPC cells as well as their self-renewal ability.	[114]
LMP2A	NPC cell lines AND NPC specimens	by stemness-related gene expression (BCG2, Bmi-1, Nanog, and SOX2)/SP cells/sphere formation	Regulate Akt activity.	LMP2A induces EMT-like cellular marker alteration and strongly up-regulates the cancer stem cell-like population in NPC.	[115]
OLFM4	HNSCC cell lines	OLFM4 is a stem cell-related gene	Regulated by aberrant DNA methylation.	The aberrant stemness gene OLFM4 expression caused by altered DNA methylation appeared to regulate early-stage HNSCC characteristics.	[116]

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topoisomerases	OSCCs tissue	by marker CD44 and CD24		All topoisomerases correlate with OSCC CSCs.	[117]
mTOR signaling pathway	NPC cell lines	by marker CD44, sox2 and OCT4	Downregulating the expression of CD44, SOX2 and MMP-2.	mTOR signaling plays significant roles both in maintaining NPC CSCs and in cancer progression.	[118]
JARID1B	Human OSCC cell lines	Sphere formation		JARID1B knockdown inhibit CSCs activity and reduced mRNA levels of NQO1, KEAP1, NRF2, FOXO1, FOXO3, KLF4, OCT4, CD133, and Nanog with radiotherapy.	[119]
Slug	HNSCC cell lines	Sphere formation/by marker CD44 and ALDH/SP cells/by stemness-related gene expression		Slug expression corrects with self-renewal capacity, stemness-associated gene expression, and cisplatin chemoresistance.	[120]
CCL21/CCR7	OSCC cell lines	by stemness-related gene expression (CD133, CD44, ALDH1A1, OCT4, BMI1, BCG2, Bmi-1, Nanog, and SOX2)/sphere formation	By activating the JAK2/STAT3 signaling pathway.	CCL21/CCR7 axis regulated EMT progress and promoted the stemness of OSCC.	[121]
MTA3	TSCC cell lines	by marker ALDH1	Negatively correlated with SOX2.	MTA3 is capable of repressing TSCC CSC properties and tumor growth.	[122]
CMTM6	HNSCC cell lines	by marker ALDH1, CD44 and BMI1, and sphere formation	A significant positive correlation between expression of CMTM6 and EMT- and CSC-related genes, immune checkpoint components.	CMTM6 regulates stemness, EMT, and T-cell dysfunction.	[123]
PIK3CA	HNSCC cell lines	by marker CD44, CD24low and ALDH1/SP/sphere formation	Activate Ephs, TRKs and the c-Kit pathway.	Promotes CSC population.	[124]
	a mouse model	CSCs derived from HNSCC of mouse		Hyper-activation of PIK3CA and loss of p53 in stem cells lead to the spontaneous development of multilineage of tumors with immunosuppressive TME.	[125]
SFRP1	Cell lines from tumor of mouse skin	by marker	Loss of SFRP1 Expression Leads to Upregulation of SOX-2.	Sfrp1 loss results in early tumor initiation and CSC regulation.	[126]
Nrf2	Cell culture from primary HNSCC samples	by marker CD133 and SP	Nrf2 upregulate ABCG2.	Nrf2 mediated drug resistance in HNSCC CSCs.	[127]
Gli1	Tongue squamous cell carcinoma line CAL33	Sphere formation		A CSCs marker.	[128]
CD47-SIRP α	OSCC cell lines	Sphere formation/by stemness-related gene expression CD47, OCT4, SOX2, CD133, and c-MYC	Associated with regulation of CD133, SOX2, OCT4, c-Myc, vimentin, Slug, Snail, E-cadherin and N-cadherin.	Promoted the generation of CSCs and malignant OSCC phenotypes.	[129]
WHSC1	HNSCC cell lines	Sphere formation	WHSC1-mediated H1.4K85 mono-methylation induces transcriptional activation of OCT4.	Enhance stemness features in HNSCC cells.	[130]
ZSCAN4	HNSCC cell lines	by marker CD44 and ALDH1/sphere formation/by stemness-related gene expression (OCT3/4, NANOG, KLF4, BMI1 and SOX2)	ZSCAN4 leads to a functional histone 3 hyperacetylation at the promoters of OCT3/4 and NANOG.	Form tumorspheres and tumor growth.	[131]
xCT	Tissue samples, HNSCC cell lines	by marker CD44	Xct mediated control of redox status in CD44v-expressing cancer cells.	xCT-targeted therapy may deplete CD44v-expressing undifferentiated HNSCC cells and concurrently sensitize the remaining differentiating cells to available treatments including EGFR-targeted therapy.	[132]
RCOR1/MED28	ocsc cell lines		MED28 correlate with CSC-related properties and induce expression of CSCs markers (CD44, KLF4, NANOG, and OCT4).	The effect of MED28 could be abrogated by RCOR1 via direct interaction.	[133]

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WNT5A	The NPC line CNE-2 and its clones S18, S22, and S26	by marker CD44 ^{high} CD24 ^{low} /SP cells/by EMT marker Vimentin	Activate protein kinase C (PKC) signaling.	Promoted EMT in NPC cells, induced the accumulation of CD24-CD44 ⁺ cells and side population.	[134]
AGR2	HNSCC cell line	Sphere formation/by stemness-related gene expression (Nanog, Sox2 and OCT4)/by EMT marker (Slug and Snail)	AGR2 was remarkably correlated with Survivin, Cyclin D1, ALDH1, Sox2, Oct4, and Slug in HNSCC tissue.	AGR2 is involved in EMT and self-renewal of CSC.	[135]
Cathepsins	Tissue samples	by stemness-related gene expression OCT4		Cathepsins B and D were localized to CSCs within the tumor nests, while cathepsin B was localized to the CSCs within the peri-tumoral stroma, and cathepsin G was localized to the tryptase ⁺ phenotypic mast cells within the peri-tumoral stroma.	[136]
CYP1B1	HNSCC cell lines	by stemness-related gene expression (OCT4, NANOG, KLF4, BMI1, ALDHA1)/by marker ALDH		CYP1B1 is crucial for stemness.	[137]
VAV2	hnSCC (tongue-located) patient-derived cells	by stemness-related gene expression		VAV2-regulated stem cell-like (SCL) program does harbor a number of cell cycle- and signaling-related kinases, VAV2-regulated SCL gene signature is associated with poor hnSCC patient prognosis.	[138]
TSPAN1	HNSCC cell lines		Correlated with EMT features and SRC activation.	Increase in resistant cells, and is associated with proliferation, resistance, metastasis, survival of resistant cells.	[139]
TrkB	Laryngeal cancer cell lines Hep-2, TU177, TU686, and AMC-HN-8	Sphere formation/by marker (CD44)/by stemness-related gene expression (SOX2 and OCT4)	miR-10a-5p Regulated TrkB Expression by Interacting with 3'-UTR of BDNF.	TrkB induce cancer stem cell-like property, miR-10a-5p inversely regulate TrkB expression.	[140]
PGK1	Patients and tissue samples, OSCC tumor cell lines	Sphere formation/by stemness-related gene expression (Sox2, Oct4 and Nanog)	Through the AKT signaling pathway.	PGK1 expression and glycolysis whilst activating the characteristics of oral cancer stem cells and EMT under hypoxia.	[141]
IGF-1	OSCC cell line SCC-4	Sphere formation/by marker CD44	IGF-1-mediated regulation of AKT and HH pathways.	IGF-1 exerts pro-tumorigenic effects by stimulating SCC-4 cell proliferation, migration, invasion and stemness.	[142]
super-enhancers (SEs)	HNSCC cell lines	by marker CD44, EpCAM and ALDH/sphere formation	FOSL1, BRD4 recruit mediators to establish SEs at a cohort of cancer stemness and pro-metastatic genes.	FOSL1 and BET inhibitors disrupting SEs inhibit CSCs and eliminate CSCs, furtherly inhibit tumor growth and metastasis.	[143, 144]
HOXA10-AS	OSCC cell lines	by marker CD133, CD44/sphere formation	Through the miR-29a/MCL-1/PI3K/AKT axis.	HOXA10-AS enhances the stem cell property of OSCC stem cells.	[145]
HSD17B7	Patients' samples and HNSCC cell lines	Sphere formation	High expression in many genes of the signature related to cellular respiration, electron transport chain, and mitochondrial organization and biogenesis.	Involve in stem self-renewal and tumorigenicity of primary keratinocytes (HKCs) and SCC cells.	[146]
PGE2	HNSCC cell line	by marker CD44 and ESA	PGE2-induced NR4A2 expression.	CD44 ^{high} /ESA ^{low} CSCs produce PGE2 to compromise 5-FU induced apoptosis to CD44 ^{high} /ESA ^{high} cells.	[147]
Cancer associated fibroblast/wnt	Cell lines from HNSCC tumor tissue	by marker (ALDH and CD44)/sphere formation	Cancer associated fibroblast activate and regulate wnt signaling within cancer cells.	Wnt in cancer cells and the tumor epithelial-stromal boundary is essential in CSCs property.	[148]
cancer associated fibroblast/PTK7	HNSCC cell lines	by CSCs marker/sphere formation	CAF-derived POSTN activate PTK7-Wnt/ β -Catenin signaling activation.	Promote cancer stemness.	[149]

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Cancer-Associated Fibroblasts (CAFs)	HNSCC cell line	Sphere formation/by stemness-related gene expression ALDH1, NANOG, SOX2, OCT4, ABCG2, CD44, CD133, and Nestin	Factors secreted by CAFs activate EGFR, IGFR, and PDGFR Signaling.	Induced the expression of stemness-related genes and sustain cancer stem properties.	[150]
hypoxia	Human laryngeal cancer cell lines, Hep-2 and AMC-HN-8	by marker CD133	Upregulated stem-related gene expression.	Upgrade the stem-like biological properties of laryngeal cancer cell lines by increasing the CD133+ stem cell fraction.	[151]
TAM markers	OSCC tissue	by marker (SOX2, ALDH1, and CD44)		TAM markers are associated with cancer stem cell marker and OSCC overall survival.	[152]
IL-6	HNSCC cell lines	by marker (ALDH and CD44)		Endothelial cell-secreted IL-6 induce HNSCC CSCs migration, EMT and maintain CSCs.	[153]
IL-6	Low-passage patient-derived xenograft (PDX) models of HNSCC			IL-6 enhances the survival and tumorigenic potential of head and neck cancer stem cells.	[36]
IL-6	Single-cell suspensions derived from primary specimens	by marker (ALDH and CD44)/sphere formation	IL-6 induces STAT3 phosphorylation.	Endothelial cell secreted-IL-6 signaling promotes self-renewal and survival of human primary head and neck cancer stem-like cells.	[154]
IL-4	Cell culture from primary HNSCC samples	by marker CD133 and SP cells		Autocrine IL-4 from CD133 SP cells promote multidrug and apoptosis resistance.	[155]
IL-1 β	Mouse SCC cell lines	Sphere formation/by CSC marker/by stemness-related gene expression	Activate Smad/ID1 signal pathway.	Promote the stem-like capabilities of HNSCC cells.	[156]
EGF	HNCSS cell lines	by CD44 and EMT regulators and stemness-related gene expression	EGFR/PI3K/HIF-1 α axis-orchestrated glycolysis.	EGF induces EMT and enrichment of CSCs.	[157]
TGF β	HNSCC K3 CSCs	The CSC properties of the cell line have been validated	TGF β 1 induce Oct4, Sox2, ABCG2, Twist, Snail, Slug and Wnt/ β -catenin signaling.	Increase EMT, self-renewal capacity and chemoresistance to cisplatin of HNSCC CSCs.	[158]
Chewing Tobacco	Het1A, a non-neoplastic and non-transformed epithelial cell line from the human esophagus	by marker (ICAM1 and CD44)		Chronic Exposure to Tobacco Extract lead to elevated expression of several ESCC cancer stem cell markers.	[159]
Arecoline-exposure	Gingival epithelial and FaDu OSCC cell lines	by marker (ALDH and CD44)/sphere formation/by stemness-related gene expression	Down-regulation of miR-145.	Chronic arecoline exposure induces malignant phenotype with the acquisition of cancer stemness/EMT, and oncogenicity.	[160]
Nicotine	HNSCC cell lines	Sphere formation/by stemness-related gene expression (CD44, Bmi1, Oct4, Nanog)	by repressing E-cadherin expression and led to the induction of stem cell markers Oct-4, Nanog, CD44 and BMI-1, the upregulation of miR-9, a repressor of E-cadherin, and the downregulation of miR-101, a repressor of EZH2.	Regulating cancer stem cell characteristics.	[161]
Cigarette smoke	HNSCC cell lines	SP cells	via Akt-mediated regulation of ABCG2 activity.	Increased the size of the side population (SP).	[162]
lower level of miR-204	HNCSS cell lines	ALDH1+, SP, and sphere forming OSCC cells/sphere formation	miR-204 binds on their 3'UTR-regions of Slug and Sox4 for an inhibitive role.	Down-regulation of miR-204 significantly increased cancer stemness and the lymph nodes incidence of orthotopic animal models.	[163]
miR34a	HNSCC cell lines	by marker (ALDH)/sphere formation	MiR-34a reversely regulate EMT- and CSCs related transcription factors.	Upregulation of miR 34a significantly inhibited the capability for EMT formation of CSC-phenotype and functionally reduced clonogenic and invasive capacity.	[164]
MicroRNA-200c	Cell lines from HNSCC tumor tissue	by marker (ALDH and CD44)	by reducing the expression of BMI1/ZEB1, Snail and N-cadherin.	Negatively modulates the expression of BMI1 but also significantly inhibits the metastatic capability of epithelial-mesenchymal transitions.	[165]

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MiR-520b	HNSCC cell lines	Sphere formation	Inhibit CD44 and multiple stemness regulators expression.	Suppressed spheroid cell formation in vitro, restrained tumorigenesis in vivo.	[166]
MicroRNA let-7a (low expression)	Cell lines from HNSCC tumor tissue	by marker ALDH1	let-7a inversely regulate CSCs related gene.	Plays a role in eliminating the putative HNSCC CSC population.	[167]
Let-7c	OSCC cell lines	by marker (ALDH1 and CD44)/sphere formation	let-7c-IL-8 pathways.	Ectopic expression of let-7c in CSCs down-regulated the stemness hallmarks and the radio/chemoresistance.	[168]
lncRNA-PVT1	NPC cell lines	Sphere formation/by marker (CD24 and CD44)/by stemness-related gene expression (Oct4, c-Myc, SOX2, ALDH1)	via inhibiting miR-1207 and activating the PI3K/AKT signal pathway.	Pvt1 promotes cancer stem cell-like properties in NPC cells.	[169]
LINC-PINT (low expression)	Human type-2 epithelial cells (Hep-2)	Sphere formation	miR-425-5p/PTCH1/SHH axis.	Downregulation of LINC-PINT was associated with increased laryngeal cancer stemness and chemoresistance to cisplatin.	[170]
miR-125a/HAX-1	Laryngeal Hep-2 cell lines	by CSC marker CD133	miR-125a inhibit HAX-1.	Low miR-125a expression lead to Hep-2 CSCs chemoresistance.	[171]
LINC00963	OSCC cell lines	by marker CD44 and ALDH1 and sphere formation	Positively correlated with the expression of the cancer stemness markers (Sox2 and CD44) and drug resistance markers (ABCG2 and ABCB5).	The downregulation of LINC00963 inhibited CSC hallmarks: elf-renewal, invasion and colony formation ability.	[172]
miR-495 (low)	Cell lines from OSCC tumor tissue	by marker (CD133 and CD44)/sphere formation	miR-495 could inhibit the activation of the TGF- β signaling pathway and HOXC6.	miR-495 may suppress HOXC6 to inhibit EMT, proliferation, migration, and invasion while promoting apoptosis of CSCs in OSCC by inhibiting the TGF- β signaling pathway.	[173]
miR-1246/CCNG2	HNSCC cell lines	by marker CD44, cd133 and ALDH1 and sphere formation	Repression of CCNG2 and induction of ABCG2.	Induce stemness, modulate chemoresistance.	[174]
MicroRNA-134	OSCC cell lines	by marker CD44, CD133	MicroRNA-134 mediate LAMC2 downregulation to suppressing PI3K-Akt signaling pathway.	Inhibit migration and invasion of OSCC CSCs.	[175]

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Supplementary References

- [1] Setubal Destro Rodrigues MF, Gammon L, Rahman MM, Biddle A, Nunes FD and Mackenzie IC. Effects of cetuximab and Erlotinib on the behaviour of cancer stem cells in head and neck squamous cell carcinoma. *Oncotarget* 2018; 9: 13488-13500.
- [2] Macha MA, Rachagani S, Qazi AK, Jahan R, Gupta S, Patel A, Seshacharyulu P, Lin C, Li S, Wang S, Verma V, Kishida S, Kishida M, Nakamura N, Kibe T, Lydiatt WM, Smith RB, Ganti AK, Jones DT, Batra SK and Jain M. Afatinib radiosensitizes head and neck squamous cell carcinoma cells by targeting cancer stem cells. *Oncotarget* 2017; 8: 20961-20973.
- [3] Liu X, Suo H, Zhou S, Hou Z, Bu M, Liu X and Xu W. Afatinib induces pro-survival autophagy and increases sensitivity to apoptosis in stem-like HNSCC cells. *Cell Death Dis* 2021; 12: 728.
- [4] Gilormini M, Malesys C, Armandy E, Manas P, Guy JB, Magne N, Rodriguez-Lafrasse C and Ardail D. Preferential targeting of cancer stem cells in the radiosensitizing effect of ABT-737 on HNSCC. *Oncotarget* 2016; 7: 16731-16744.
- [5] Guy JB, Espenel S, Louati S, Gauthier A, Garcia MA, Vial N, Malesys C, Ardail D, Alphonse G, Wozny AS, Rodriguez-Lafrasse C and Magne N. Combining radiation to EGFR and Bcl-2 blockade: a new approach to target cancer stem cells in head and neck squamous cell carcinoma. *J Cancer Res Clin Oncol* 2021; 147: 1905-1916.
- [6] Chen H, Sa G, Li L, He S and Wu T. In vitro and in vivo synergistic anti-tumor effect of LIN28 inhibitor and metformin in oral squamous cell carcinoma. *Eur J Pharmacol* 2021; 891: 173757.
- [7] Liu SC, Wu YC, Huang CM, Hsieh MS, Huang TY, Huang CS, Hsu TN, Huang MS, Lee WH, Yeh CT and Lin CS. Inhibition of Bruton's tyrosine kinase as a therapeutic strategy for chemoresistant oral squamous cell carcinoma and potential suppression of cancer stemness. *Oncogenesis* 2021; 10: 20.
- [8] Shigeishi H, Biddle A, Gammon L, Rodini CO, Yamasaki M, Seino S, Sugiyama M, Takechi M and Mackenzie IC. Elevation in 5-FU-induced apoptosis in head and neck cancer stem cells by a combination of CDHP and GSK-3beta inhibitors. *J Oral Pathol Med* 2015; 44: 201-207.
- [9] Roy S, Roy S, Kar M, Chakraborty A, Kumar A, Delogu F, Asthana S, Hande MP and Banerjee B. Combined treatment with cisplatin and the tankyrase inhibitor XAV-939 increases cytotoxicity, abrogates cancer-stem-like cell phenotype and increases chemosensitivity of head-and-neck squamous-cell carcinoma cells. *Mutat Res Genet Toxicol Environ Mutagen* 2019; 846: 503084.
- [10] Fang L, Zhu Q, Neuenschwander M, Specker E, Wulf-Goldenberg A, Weis WI, von Kries JP and Birchmeier W. A small-molecule antagonist of the beta-catenin/TCF4 interaction blocks the self-renewal of cancer stem cells and suppresses tumorigenesis. *Cancer Res* 2016; 76: 891-901.
- [11] Warriar S, Bhuvanlakshmi G, Arfuso F, Rajan G, Millward M and Dharmarajan A. Cancer stem-like cells from head and neck cancers are chemosensitized by the Wnt antagonist, sFRP4, by inducing apoptosis, decreasing stemness, drug resistance and epithelial to mesenchymal transition. *Cancer Gene Ther* 2014; 21: 381-388.
- [12] Chikamatsu K, Ishii H, Murata T, Sakakura K, Shino M, Toyoda M, Takahashi K and Masuyama K. Alteration of cancer stem cell-like phenotype by histone deacetylase inhibitors in squamous cell carcinoma of the head and neck. *Cancer Sci* 2013; 104: 1468-1475.
- [13] Kumar B, Yadav A, Lang JC, Teknos TN and Kumar P. Suberoylanilide hydroxamic acid (SAHA) reverses chemoresistance in head and neck cancer cells by targeting cancer stem cells via the downregulation of nanog. *Genes Cancer* 2015; 6: 169-181.
- [14] Lee SH, Nam HJ, Kang HJ, Samuels TL, Johnston N and Lim YC. Valproic acid suppresses the self-renewal and proliferation of head and neck cancer stem cells. *Oncol Rep* 2015; 34: 2065-2071.
- [15] Marques AEM, do Nascimento Filho CHV, Marinho Bezerra TM, Guerra ENS, Castilho RM and Squarize CH. Entinostat is a novel therapeutic agent to treat oral squamous cell carcinoma. *J Oral Pathol Med* 2020; 49: 771-779.
- [16] Boivin A, Hanot M, Malesys C, Maalouf M, Rousson R, Rodriguez-Lafrasse C and Ardail D. Transient alteration of cellular redox buffering before irradiation triggers apoptosis in head and neck carcinoma stem and non-stem cells. *PLoS One* 2011; 6: e14558.
- [17] Bertrand G, Maalouf M, Boivin A, Battiston-Montagne P, Beuve M, Levy A, Jalade P, Fournier C, Ardail D, Magne N, Alphonse G and Rodriguez-Lafrasse C. Targeting head and neck cancer stem cells to overcome resistance to photon and carbon ion radiation. *Stem Cell Rev Rep* 2014; 10: 114-126.
- [18] Finkel KA, Warner KA, Kerk S, Bradford CR, McLean SA, Prince ME, Zhong H, Hurt EM, Hollingsworth RE, Wicha MS, Tice DA and Nor JE. IL-6 Inhibition With MEDI5117 Decreases The fraction of head and neck cancer stem cells and prevents tumor recurrence. *Neoplasia* 2016; 18: 273-281.
- [19] Jia L, Zhang W and Wang CY. BMI1 inhibition eliminates residual cancer stem cells after PD1 blockade and activates antitumor immunity to prevent metastasis and relapse. *Cell Stem Cell* 2020; 27: 238-253, e236.

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- [20] Saito S, Ozawa H, Imanishi Y, Sekimizu M, Watanabe Y, Ito F, Ikari Y, Nakahara N, Kameyama K and Ogawa K. Cyclooxygenase-2 expression is associated with chemoresistance through cancer stemness property in hypopharyngeal carcinoma. *Oncol Lett* 2021; 22: 533.
- [21] Liu SC, Huang CM, Bamodu OA, Lin CS, Liu BL, Tzeng YM, Tsai JT, Lee WH and Chen TM. Ovatodiolide suppresses nasopharyngeal cancer by targeting stem cell-like population, inducing apoptosis, inhibiting EMT and dysregulating JAK/STAT signaling pathway. *Phytomedicine* 2019; 56: 269-278.
- [22] Lee SH, Nam HJ, Kang HJ, Kwon HW and Lim YC. Epigallocatechin-3-gallate attenuates head and neck cancer stem cell traits through suppression of Notch pathway. *Eur J Cancer* 2013; 49: 3210-3218.
- [23] Chang WW, Hu FW, Yu CC, Wang HH, Feng HP, Lan C, Tsai LL and Chang YC. Quercetin in elimination of tumor initiating stem-like and mesenchymal transformation property in head and neck cancer. *Head Neck* 2013; 35: 413-419.
- [24] Deng R, Wang X, Liu Y, Yan M, Hanada S, Xu Q, Zhang J, Han Z, Chen W and Zhang P. A new gamboge derivative compound 2 inhibits cancer stem-like cells via suppressing EGFR tyrosine phosphorylation in head and neck squamous cell carcinoma. *J Cell Mol Med* 2013; 17: 1422-1433.
- [25] Chen YW, Chen KH, Huang PI, Chen YC, Chiou GY, Lo WL, Tseng LM, Hsu HS, Chang KW and Chiou SH. Cucurbitacin I suppressed stem-like property and enhanced radiation-induced apoptosis in head and neck squamous carcinoma-derived CD44(+)ALDH1(+) cells. *Mol Cancer Ther* 2010; 9: 2879-2892.
- [26] Chang CW, Chen CC, Wu MJ, Chen YS, Chen CC, Sheu SJ, Lin TW, Chou SH, Lin SC, Liu CJ, Lee TC, Huang CY and Lo JF. Active component of *antrodia cinnamomea* mycelia targeting head and neck cancer initiating cells through exaggerated autophagic cell death. *Evid Based Complement Alternat Med* 2013; 2013: 946451.
- [27] Chang CW, Chen YS, Chen CC, Chan IO, Chen CC, Sheu SJ, Lin TW, Chou SH, Liu CJ, Lee TC and Lo JF. Targeting cancer initiating cells by promoting cell differentiation and restoring chemosensitivity via dual inactivation of STAT3 and src activity using an active component of *antrodia cinnamomea* mycelia. *Oncotarget* 2016; 7: 73016-73031.
- [28] Pan ST, Qin Y, Zhou ZW, He ZX, Zhang X, Yang T, Yang YX, Wang D, Zhou SF and Qiu JX. Plumbagin suppresses epithelial to mesenchymal transition and stemness via inhibiting Nrf2-mediated signaling pathway in human tongue squamous cell carcinoma cells. *Drug Des Devel Ther* 2015; 9: 5511-5551.
- [29] Keysar SB, Gomes N, Miller B, Jackson BC, Le PN, Morton JJ, Reisinger J, Chimed TS, Gomez KE, Nieto C, Frederick B, Pronk GJ, Somerset HL, Tan AC, Wang XJ, Raben D, Su TT and Jimeno A. Inhibiting translation elongation with SVC112 suppresses cancer stem cells and inhibits growth in head and neck squamous carcinoma. *Cancer Res* 2020; 80: 1183-1198.
- [30] Peng Y, He G, Tang D, Xiong L, Wen Y, Miao X, Hong Z, Yao H, Chen C, Yan S, Lu L, Yang Y, Li Q and Deng X. Lovastatin inhibits cancer stem cells and sensitizes to chemo- and photodynamic therapy in nasopharyngeal carcinoma. *J Cancer* 2017; 8: 1655-1664.
- [31] Cui X, Xiao D and Wang X. Inhibition of laryngeal cancer stem cells by tetrandrine. *Anticancer Drugs* 2019; 30: 886-891.
- [32] Hu FW, Yu CC, Hsieh PL, Liao YW, Lu MY and Chu PM. Targeting oral cancer stemness and chemoresistance by isoliquiritigenin-mediated GRP78 regulation. *Oncotarget* 2017; 8: 93912-93923.
- [33] Elkashty OA and Tran SD. Broccoli extract increases drug-mediated cytotoxicity towards cancer stem cells of head and neck squamous cell carcinoma. *Br J Cancer* 2020; 123: 1395-1403.
- [34] Basak SK, Zinabadi A, Wu AW, Venkatesan N, Duarte VM, Kang JJ, Dalgard CL, Srivastava M, Sarkar FH, Wang MB and Srivatsan ES. Liposome encapsulated curcumin-difluorinated (CDF) inhibits the growth of cisplatin resistant head and neck cancer stem cells. *Oncotarget* 2015; 6: 18504-18517.
- [35] Siddappa G, Kulsum S, Ravindra DR, Kumar VV, Raju N, Raghavan N, Sudheendra HV, Sharma A, Sunny SP, Jacob T, Kuruvilla BT, Benny M, Antony B, Seshadri M, Lakshminarayan P, Hicks W Jr, Suresh A and Kuriakose MA. Curcumin and metformin-mediated chemoprevention of oral cancer is associated with inhibition of cancer stem cells. *Mol Carcinog* 2017; 56: 2446-2460.
- [36] Liu SC, Huang CS, Huang CM, Hsieh MS, Huang MS, Fong IH, Yeh CT and Lin CC. Isoorientin inhibits epithelial-to-mesenchymal properties and cancer stem-cell-like features in oral squamous cell carcinoma by blocking Wnt/beta-catenin/STAT3 axis. *Toxicol Appl Pharmacol* 2021; 424: 115581.
- [37] Ketkaew Y, Osathanon T, Pavasant P and Soompon S. Apigenin inhibited hypoxia induced stem cell marker expression in a head and neck squamous cell carcinoma cell line. *Arch Oral Biol* 2017; 74: 69-74.
- [38] Hu FW, Tsai LL, Yu CH, Chen PN, Chou MY and Yu CC. Impairment of tumor-initiating stem-like property and reversal of epithelial-mesenchymal transdifferentiation in head and neck cancer by resveratrol treatment. *Mol Nutr Food Res* 2012; 56: 1247-1258.
- [39] Chang YC, Jan CI, Peng CY, Lai YC, Hu FW and Yu CC. Activation of microRNA-494-targeting Bmi1 and ADAM10 by silibinin ablates cancer stemness and predicts favourable prognostic value in head and neck squamous cell carcinomas. *Oncotarget* 2015; 6: 24002-24016.

Cancer stem cells of head and neck squamous cell carcinoma

- [40] Nor F, Nor C, Bento LW, Zhang Z, Bretz WA and Nor JE. Propolis reduces the stemness of head and neck squamous cell carcinoma. *Arch Oral Biol* 2021; 125: 105087.
- [41] Keysar SB, Le PN, Miller B, Jackson BC, Eagles JR, Nieto C, Kim J, Tang B, Glogowska MJ, Morton JJ, Padilla-Just N, Gomez K, Warnock E, Reisinger J, Arcaroli JJ, Messersmith WA, Wakefield LM, Gao D, Tan AC, Serracino H, Vasiliou V, Roop DR, Wang XJ and Jimeno A. Regulation of head and neck squamous cancer stem cells by PI3K and SOX2. *J Natl Cancer Inst* 2016; 109: djw189.
- [42] Facompre ND, Harmeyer KM, Sole X, Kabraji S, Belden Z, Sahu V, Whelan K, Tanaka K, Weinstein GS, Montone KT, Roesch A, Gimotty PA, Herlyn M, Rustgi AK, Nakagawa H, Ramaswamy S and Basu D. JARID1B enables transit between distinct states of the stem-like cell population in oral cancers. *Cancer Res* 2016; 76: 5538-5549.
- [43] Bourguignon LYW, Earle C and Shiina M. Activation of matrix hyaluronan-mediated CD44 signaling, epigenetic regulation and chemoresistance in head and neck cancer stem cells. *Int J Mol Sci* 2017; 18: 1849.
- [44] Bourguignon LY, Wong G, Earle C and Chen L. Hyaluronan-CD44v3 interaction with Oct4-Sox2-Nanog promotes miR-302 expression leading to self-renewal, clonal formation, and cisplatin resistance in cancer stem cells from head and neck squamous cell carcinoma. *J Biol Chem* 2012; 287: 32800-32824.
- [45] Masui T, Ota I, Yook JI, Mikami S, Yane K, Yamanaka T and Hosoi H. Snail-induced epithelial-mesenchymal transition promotes cancer stem cell-like phenotype in head and neck cancer cells. *Int J Oncol* 2014; 44: 693-699.
- [46] Ota I, Masui T, Kurihara M, Yook JI, Mikami S, Kimura T, Shimada K, Konishi N, Yane K, Yamanaka T and Kitahara T. Snail-induced EMT promotes cancer stem cell-like properties in head and neck cancer cells. *Oncol Rep* 2016; 35: 261-266.
- [47] Zhu LF, Hu Y, Yang CC, Xu XH, Ning TY, Wang ZL, Ye JH and Liu LK. Snail overexpression induces an epithelial to mesenchymal transition and cancer stem cell-like properties in SCC9 cells. *Lab Invest* 2012; 92: 744-752.
- [48] Peng S, Wu C, Sun W, Liu D, Luo M, Su B, Zhang L, Mei Q and Hu G. Snail-mediated cancer stem cell-like phenotype in human CNE2 nasopharyngeal carcinoma cell. *Head Neck* 2018; 40: 485-497.
- [49] Lee SH, Lee CR, Rigas NK, Kim RH, Kang MK, Park NH and Shin KH. Human papillomavirus 16 (HPV16) enhances tumor growth and cancer stemness of HPV-negative oral/oropharyngeal squamous cell carcinoma cells via miR-181 regulation. *Papillomavirus Res* 2015; 1: 116-125.
- [50] Hufbauer M, Maltseva M, Meinrath J, Lechner A, Beutner D, Huebbers CU and Akgul B. HPV16 increases the number of migratory cancer stem cells and modulates their miRNA expression profile in oropharyngeal cancer. *Int J Cancer* 2018; 143: 1426-1439.
- [51] Vlashi E, Chen AM, Boyrie S, Yu G, Nguyen A, Brower PA, Hess CB and Pajonk F. Radiation-Induced dedifferentiation of head and neck cancer cells into cancer stem cells depends on human papillomavirus status. *Int J Radiat Oncol Biol Phys* 2016; 94: 1198-1206.
- [52] Ren L, Deng B, Saloura V, Park JH and Nakamura Y. MELK inhibition targets cancer stem cells through down-regulation of SOX2 expression in head and neck cancer cells. *Oncol Rep* 2019; 41: 2540-2548.
- [53] Nallaiah S, Lee VMY, Brasch HD, de Jongh J, Schaijck BV, Marsh R, Tan ST and Itinteang T. Cancer stem cells within moderately differentiated head and neck cutaneous squamous cell carcinoma express components of the renin-angiotensin system. *J Plast Reconstr Aesthet Surg* 2019; 72: 1484-1493.
- [54] Featherston T, Yu HH, Dunne JC, Chibnall AM, Brasch HD, Davis PF, Tan ST and Itinteang T. Cancer stem cells in moderately differentiated buccal mucosal squamous cell carcinoma express components of the renin-angiotensin system. *Front Surg* 2016; 3: 52.
- [55] Itinteang T, Dunne JC, Chibnall AM, Brasch HD, Davis PF and Tan ST. Cancer stem cells in moderately differentiated oral tongue squamous cell carcinoma express components of the renin-angiotensin system. *J Clin Pathol* 2016; 69: 942-945.
- [56] Siljee S, Buchanan O, Brasch HD, Bockett N, Patel J, Paterson E, Purdie GL, Davis PF, Itinteang T and Tan ST. Cancer stem cells in metastatic head and neck cutaneous squamous cell carcinoma express components of the renin-angiotensin system. *Cells* 2021; 10: 243.
- [57] Bian S, Wang Z, Chen Y and Li R. SPLUNC1 and MLL3 regulate cancer stem cells in nasopharyngeal carcinoma. *J BUON* 2019; 24: 1700-1705.
- [58] Ji J, Yu Y, Li ZL, Chen MY, Deng R, Huang X, Wang GF, Zhang MX, Yang Q, Ravichandran S, Feng GK, Xu XL, Yang CL, Qiu MZ, Jiao L, Yang D and Zhu XF. XIAP limits autophagic degradation of Sox2 and is a therapeutic target in nasopharyngeal carcinoma stem cells. *Theranostics* 2018; 8: 1494-1510.
- [59] Chen D, Wu M, Li Y, Chang I, Yuan Q, Ekimyan-Salvo M, Deng P, Yu B, Yu Y, Dong J, Szymanski JM, Ramadoss S, Li J and Wang CY. Targeting BMI1(+) cancer stem cells overcomes chemoresistance and inhibits metastases in squamous cell carcinoma. *Cell Stem Cell* 2017; 20: 621-634, e626.
- [60] Chen H, Zhou L, Dou T, Wan G, Tang H and Tian J. BMI1'S maintenance of the proliferative capacity of laryngeal cancer stem cells. *Head Neck* 2011; 33: 1115-1125.

Cancer stem cells of head and neck squamous cell carcinoma

- [61] Roy S, Roy S, Kar M, Padhi S, Saha A, Anuja K and Banerjee B. Role of p38 MAPK in disease relapse and therapeutic resistance by maintenance of cancer stem cells in head and neck squamous cell carcinoma. *J Oral Pathol Med* 2018; 47: 492-501.
- [62] Lee SH, Koo BS, Kim JM, Huang S, Rho YS, Bae WJ, Kang HJ, Kim YS, Moon JH and Lim YC. Wnt/beta-catenin signalling maintains self-renewal and tumourigenicity of head and neck squamous cell carcinoma stem-like cells by activating Oct4. *J Pathol* 2014; 234: 99-107.
- [63] Xie SL, Fan S, Zhang SY, Chen WX, Li QX, Pan GK, Zhang HQ, Wang WW, Weng B, Zhang Z, Li JS and Lin ZY. SOX8 regulates cancer stem-like properties and cisplatin-induced EMT in tongue squamous cell carcinoma by acting on the Wnt/beta-catenin pathway. *Int J Cancer* 2018; 142: 1252-1265.
- [64] Lee SH, Oh SY, Do SI, Lee HJ, Kang HJ, Rho YS, Bae WJ and Lim YC. SOX2 regulates self-renewal and tumorigenicity of stem-like cells of head and neck squamous cell carcinoma. *Br J Cancer* 2014; 111: 2122-2130.
- [65] Zhong Q, Liu ZH, Lin ZR, Hu ZD, Yuan L, Liu YM, Zhou AJ, Xu LH, Hu LJ, Wang ZF, Guan XY, Hao JJ, Lui VWY, Guo L, Mai HQ, Chen MY, Han F, Xia YF, Grandis JR, Zhang X and Zeng MS. The RARS-MAD1L1 fusion gene induces cancer stem cell-like properties and therapeutic resistance in nasopharyngeal carcinoma. *Clin Cancer Res* 2018; 24: 659-673.
- [66] Chien CS, Wang ML, Chu PY, Chang YL, Liu WH, Yu CC, Lan YT, Huang PI, Lee YY, Chen YW, Lo WL and Chiou SH. Lin28B/Let-7 regulates expression of Oct4 and Sox2 and reprograms oral squamous cell carcinoma cells to a stem-like state. *Cancer Res* 2015; 75: 2553-2565.
- [67] Subramanian C, Kovatch KJ, Sim MW, Wang G, Prince ME, Carey TE, Davis R, Blagg BSJ and Cohen MS. Novel C-terminal heat shock protein 90 inhibitors (KU711 and Ku757) are effective in targeting head and neck squamous cell carcinoma cancer stem cells. *Neoplasia* 2017; 19: 1003-1011.
- [68] Kerk SA, Finkel KA, Pearson AT, Warner KA, Zhang Z, Nor F, Wagner VP, Vargas PA, Wicha MS, Hurt EM, Hollingsworth RE, Tice DA and Nor JE. 5T4-targeted therapy ablates cancer stem cells and prevents recurrence of head and neck squamous cell carcinoma. *Clin Cancer Res* 2017; 23: 2516-2527.
- [69] Lim YC, Kang HJ and Moon JH. C-Met pathway promotes self-renewal and tumorigenicity of head and neck squamous cell carcinoma stem-like cell. *Oral Oncol* 2014; 50: 633-639.
- [70] Sun S and Wang Z. Head neck squamous cell carcinoma c-Met(+) cells display cancer stem cell properties and are responsible for cisplatin-resistance and metastasis. *Int J Cancer* 2011; 129: 2337-2348.
- [71] Sun S, Liu S, Duan SZ, Zhang L, Zhou H, Hu Y, Zhou X, Shi C, Zhou R and Zhang Z. Targeting the c-Met/FZD8 signaling axis eliminates patient-derived cancer stem-like cells in head and neck squamous carcinomas. *Cancer Res* 2014; 74: 7546-7559.
- [72] Evran E, Sahin H, Akbas K, Cigdem S and Gunduz E. Investigation of MACC1 gene expression in head and neck cancer and cancer stem cells. *Clin Invest Med* 2016; 39: 27506.
- [73] Li J, Li Z, Wu Y, Wang Y, Wang D, Zhang W, Yuan H, Ye J, Song X, Yang J, Jiang H and Cheng J. The Hippo effector TAZ promotes cancer stemness by transcriptional activation of SOX2 in head neck squamous cell carcinoma. *Cell Death Dis* 2019; 10: 603.
- [74] Li Z, Wang Y, Zhu Y, Yuan C, Wang D, Zhang W, Qi B, Qiu J, Song X, Ye J, Wu H, Jiang H, Liu L, Zhang Y, Song LN, Yang J and Cheng J. The Hippo transducer TAZ promotes epithelial to mesenchymal transition and cancer stem cell maintenance in oral cancer. *Mol Oncol* 2015; 9: 1091-1105.
- [75] Reers S, Pfannerstill AC, Maushagen R, Pries R and Wollenberg B. Stem cell profiling in head and neck cancer reveals an Oct-4 expressing subpopulation with properties of chemoresistance. *Oral Oncol* 2014; 50: 155-162.
- [76] Nathansen J, Lukiyanchuk V, Hein L, Stolte MI, Borgmann K, Lock S, Kurth I, Baumann M, Krause M, Linge A and Dubrovskaya A. Oct4 confers stemness and radioresistance to head and neck squamous cell carcinoma by regulating the homologous recombination factors PSMC3IP and RAD54L. *Oncogene* 2021; 40: 4214-4228.
- [77] Jiang P, Xu C, Zhou M, Zhou H, Dong W, Wu X, Chen A and Feng Q. RXRalpha-enriched cancer stem cell-like properties triggered by CDDP in head and neck squamous cell carcinoma (HNSCC). *Carcinogenesis* 2018; 39: 252-262.
- [78] Leong HS, Chong FT, Sew PH, Lau DP, Wong BH, Teh BT, Tan DS and Iyer NG. Targeting cancer stem cell plasticity through modulation of epidermal growth factor and insulin-like growth factor receptor signaling in head and neck squamous cell cancer. *Stem Cells Transl Med* 2014; 3: 1055-1065.
- [79] Lv XX, Zheng XY, Yu JJ, Ma HR, Hua C and Gao RT. EGFR enhances the stemness and progression of oral cancer through inhibiting autophagic degradation of SOX2. *Cancer Med* 2020; 9: 1131-1140.
- [80] Abhold EL, Kiang A, Rahimy E, Kuo SZ, Wang-Rodriguez J, Lopez JP, Blair KJ, Yu MA, Haas M, Brumund KT, Altuna X, Patel A, Weisman RA and Ongkeko WM. EGFR kinase promotes acquisition of stem cell-like properties: a potential therapeutic target in head and neck squamous cell carcinoma stem cells. *PLoS One* 2012; 7: e32459.
- [81] Lee SH, Do SI, Lee HJ, Kang HJ, Koo BS and Lim YC. Notch1 signaling contributes to stemness in head and neck squamous cell carcinoma. *Lab Invest* 2016; 96: 508-516.

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- [82] Zhao ZL, Zhang L, Huang CF, Ma SR, Bu LL, Liu JF, Yu GT, Liu B, Gutkind JS, Kulkarni AB, Zhang WF and Sun ZJ. NOTCH1 inhibition enhances the efficacy of conventional chemotherapeutic agents by targeting head neck cancer stem cell. *Sci Rep* 2016; 6: 24704.
- [83] Upadhyay P, Nair S, Kaur E, Aich J, Dani P, Sethunath V, Gardi N, Chandrani P, Godbole M, Sonawane K, Prasad R, Kannan S, Agarwal B, Kane S, Gupta S, Dutt S and Dutt A. Notch pathway activation is essential for maintenance of stem-like cells in early tongue cancer. *Oncotarget* 2016; 7: 50437-50449.
- [84] Bourguignon LY, Wong G and Shiina M. Up-regulation of Histone Methyltransferase, DOT1L, by matrix hyaluronan promotes microRNA-10 expression leading to tumor cell invasion and chemoresistance in cancer stem cells from head and neck squamous cell carcinoma. *J Biol Chem* 2016; 291: 10571-10585.
- [85] Huang CF, Chen L, Li YC, Wu L, Yu GT, Zhang WF and Sun ZJ. NLRP3 inflammasome activation promotes inflammation-induced carcinogenesis in head and neck squamous cell carcinoma. *J Exp Clin Cancer Res* 2017; 36: 116.
- [86] Chen L, Li YC, Wu L, Yu GT, Zhang WF, Huang CF and Sun ZJ. TRAF6 regulates tumour metastasis through EMT and CSC phenotypes in head and neck squamous cell carcinoma. *J Cell Mol Med* 2018; 22: 1337-1349.
- [87] Wu MJ, Jan CI, Tsay YG, Yu YH, Huang CY, Lin SC, Liu CJ, Chen YS, Lo JF and Yu CC. Elimination of head and neck cancer initiating cells through targeting glucose regulated protein78 signaling. *Mol Cancer* 2010; 9: 283.
- [88] Chiu CC, Lee LY, Li YC, Chen YJ, Lu YC, Li YL, Wang HM, Chang JT and Cheng AJ. Grp78 as a therapeutic target for refractory head-neck cancer with CD24(-)CD44(+) stemness phenotype. *Cancer Gene Ther* 2013; 20: 606-615.
- [89] Cheng LH, Hung KF, Huang TF, Hsieh HP, Wang SY, Huang CY and Lo JF. Attenuation of cancer-initiating cells stemness properties by abrogating S100A4 calcium binding ability in head and neck cancers. *Oncotarget* 2016; 7: 78946-78957.
- [90] Lo JF, Yu CC, Chiou SH, Huang CY, Jan CI, Lin SC, Liu CJ, Hu WY and Yu YH. The epithelial-mesenchymal transition mediator S100A4 maintains cancer-initiating cells in head and neck cancers. *Cancer Res* 2011; 71: 1912-1923.
- [91] Islam M, Sharma S and Teknos TN. RhoC regulates cancer stem cells in head and neck squamous cell carcinoma by overexpressing IL-6 and phosphorylation of STAT3. *PLoS One* 2014; 9: e88527.
- [92] Shigeishi H, Biddle A, Gammon L, Emich H, Rodini CO, Gemenetzidis E, Fazil B, Sugiyama M, Kamata N and Mackenzie IC. Maintenance of stem cell self-renewal in head and neck cancers requires actions of GSK3beta influenced by CD44 and RHAMM. *Stem Cells* 2013; 31: 2073-2083.
- [93] Muhammad N, Bhattacharya S, Steele R, Phillips N and Ray RB. Involvement of c-Fos in the promotion of cancer stem-like cell properties in head and neck squamous cell carcinoma. *Clin Cancer Res* 2017; 23: 3120-3128.
- [94] Liu S, Ye D, Guo W, Yu W, He Y, Hu J, Wang Y, Zhang L, Liao Y, Song H, Zhong S, Xu D, Yin H, Sun B, Wang X, Liu J, Wu Y, Zhou BP, Zhang Z and Deng J. G9a is essential for EMT-mediated metastasis and maintenance of cancer stem cell-like characters in head and neck squamous cell carcinoma. *Oncotarget* 2015; 6: 6887-6901.
- [95] Giudice FS, Pinto DS Jr, Nor JE, Squarize CH and Castilho RM. Inhibition of histone deacetylase impacts cancer stem cells and induces epithelial-mesenchyme transition of head and neck cancer. *PLoS One* 2013; 8: e58672.
- [96] Lagunas AM, Francis M, Maniar NB, Nikolova G, Wu J and Crowe DL. Paracrine interaction of cancer stem cell populations is regulated by the senescence-associated secretory phenotype (SASP). *Mol Cancer Res* 2019; 17: 1480-1492.
- [97] Czerwinski MJ, Desiderio V, Shkeir O, Papagerakis P, Lapadatescu MC, Owen JH, Athanassiou-Papaefthymiou M, Zheng L, Papaccio G, Prince ME and Papagerakis S. In vitro evaluation of sialyl Lewis X relationship with head and neck cancer stem cells. *Otolaryngol Head Neck Surg* 2013; 149: 97-104.
- [98] Desiderio V, Papagerakis P, Tirino V, Zheng L, Matossian M, Prince ME, Paino F, Mele L, Papaccio F, Montella R, Papaccio G and Papagerakis S. Increased fucosylation has a pivotal role in invasive and metastatic properties of head and neck cancer stem cells. *Oncotarget* 2015; 6: 71-84.
- [99] Jung YS, Vermeer PD, Vermeer DW, Lee SJ, Goh AR, Ahn HJ and Lee JH. CD200: association with cancer stem cell features and response to chemoradiation in head and neck squamous cell carcinoma. *Head Neck* 2015; 37: 327-335.
- [100] Lu H, Yan C, Quan XX, Yang X, Zhang J, Bian Y, Chen Z and Van Waes C. CK2 phosphorylates and inhibits TAp73 tumor suppressor function to promote expression of cancer stem cell genes and phenotype in head and neck cancer. *Neoplasia* 2014; 16: 789-800.
- [101] Fukusumi T, Ishii H, Konno M, Yasui T, Nakahara S, Takenaka Y, Yamamoto Y, Nishikawa S, Kano Y, Ogawa H, Hasegawa S, Hamabe A, Haraguchi N, Doki Y, Mori M and Inohara H. CD10 as a novel marker of therapeutic resistance and cancer stem cells in head and neck squamous cell carcinoma. *Br J Cancer* 2014; 111: 506-514.

Cancer stem cells of head and neck squamous cell carcinoma

- [102] Faber A, Goessler UR, Hoermann K, Schultz JD, Umbreit C and Stern-Straeter J. SDF-1-CXCR4 axis: cell trafficking in the cancer stem cell niche of head and neck squamous cell carcinoma. *Oncol Rep* 2013; 29: 2325-2331.
- [103] Shen C, Chen F, Wang H, Li G, Yu C, Wang X and Wen Z. The Pinx1 gene downregulates telomerase and inhibits proliferation of CD133+ cancer stem cells isolated from a nasopharyngeal carcinoma cell line by regulating Trfs and Mad1/C-Myc/p53 pathways. *Cell Physiol Biochem* 2018; 49: 282-294.
- [104] Yu C, Chen F, Wang X, Cai Z, Yang M, Zhong Q, Feng J, Li J, Shen C and Wen Z. Pin2 telomeric repeat factor 1-interacting telomerase inhibitor 1 (PinX1) inhibits nasopharyngeal cancer cell stemness: implication for cancer progression and therapeutic targeting. *J Exp Clin Cancer Res* 2020; 39: 31.
- [105] Khammanivong A, Gopalakrishnan R and Dickerson EB. SMURF1 silencing diminishes a CD44-high cancer stem cell-like population in head and neck squamous cell carcinoma. *Mol Cancer* 2014; 13: 260.
- [106] Sheng X, Li Y, Li Y, Liu W, Lu Z, Zhan J, Xu M, Chen L, Luo X, Cai G and Zhang S. PLOD2 contributes to drug resistance in laryngeal cancer by promoting cancer stem cell-like characteristics. *BMC Cancer* 2019; 19: 840.
- [107] Rodrigues MFSD, Miguita L, De Andrade NP, Heguedusch D, Rodini CO, Moyses RA, Toporcov TN, Gama RR, Tajara EE and Nunes FD. GLI3 knockdown decreases stemness, cell proliferation and invasion in oral squamous cell carcinoma. *Int J Oncol* 2018; 53: 2458-2472.
- [108] Chu PY, Hu FW, Yu CC, Tsai LL, Yu CH, Wu BC, Chen YW, Huang PI and Lo WL. Epithelial-mesenchymal transition transcription factor ZEB1/ZEB2 co-expression predicts poor prognosis and maintains tumor-initiating properties in head and neck cancer. *Oral Oncol* 2013; 49: 34-41.
- [109] Bae WJ, Koo BS, Lee SH, Kim JM, Rho YS, Lim JY, Moon JH, Cho JH and Lim YC. Inhibitor of DNA binding 2 is a novel therapeutic target for stemness of head and neck squamous cell carcinoma. *Br J Cancer* 2017; 117: 1810-1818.
- [110] Qiao B, Johnson NW, Chen X, Li R, Tao Q and Gao J. Disclosure of a stem cell phenotype in an oral squamous cell carcinoma cell line induced by BMP-4 via an epithelial-mesenchymal transition. *Oncol Rep* 2011; 26: 455-461.
- [111] Chen RH, Du Y, Han P, Wang HB, Liang FY, Feng GK, Zhou AJ, Cai MY, Zhong Q, Zeng MS and Huang XM. ISG15 predicts poor prognosis and promotes cancer stem cell phenotype in nasopharyngeal carcinoma. *Oncotarget* 2016; 7: 16910-16922.
- [112] Lu Y, Liang Y, Zheng X, Deng X, Huang W and Zhang G. EVI1 promotes epithelial-to-mesenchymal transition, cancer stem cell features and chemo-/radioresistance in nasopharyngeal carcinoma. *J Exp Clin Cancer Res* 2019; 38: 82.
- [113] Bu LL, Zhao ZL, Liu JF, Ma SR, Huang CF, Liu B, Zhang WF and Sun ZJ. STAT3 blockade enhances the efficacy of conventional chemotherapeutic agents by eradicating head neck stemloid cancer cell. *Oncotarget* 2015; 6: 41944-41958.
- [114] Wang J, Huang Y, Guan Z, Zhang JL, Su HK, Zhang W, Yue CF, Yan M, Guan S and Liu QQ. E3-ligase Skp2 predicts poor prognosis and maintains cancer stem cell pool in nasopharyngeal carcinoma. *Oncotarget* 2014; 5: 5591-5601.
- [115] Kong QL, Hu LJ, Cao JY, Huang YJ, Xu LH, Liang Y, Xiong D, Guan S, Guo BH, Mai HQ, Chen QY, Zhang X, Li MZ, Shao JY, Qian CN, Xia YF, Song LB, Zeng YX and Zeng MS. Epstein-Barr virus-encoded LMP2A induces an epithelial-mesenchymal transition and increases the number of side population stem-like cancer cells in nasopharyngeal carcinoma. *PLoS Pathog* 2010; 6: e1000940.
- [116] Suzuki T, Yamazaki H, Honda K, Ryo E, Kaneko A, Ota Y and Mori T. Altered DNA methylation is associated with aberrant stemness gene expression in early-stage HNSCC. *Int J Oncol* 2019; 55: 915-924.
- [117] Oliveira-Costa JP, Oliveira LR, da Silveira GG, Soave DF, Soares FA and Ribeiro-Silva A. Topoisomerase expression in oral squamous cell carcinoma: relationship with cancer stem cells profiles and lymph node metastasis. *J Oral Pathol Med* 2012; 41: 762-768.
- [118] Yang C, Zhang Y, Zhang Y, Zhang Z, Peng J, Li Z, Han L, You Q, Chen X, Rao X, Zhu Y and Liao Z. Downregulation of cancer stem cell properties via mTOR signaling pathway inhibition by rapamycin in nasopharyngeal carcinoma. *Int J Oncol* 2015; 47: 909-917.
- [119] Lin CS, Lin YC, Adebayo BO, Wu A, Chen JH, Peng YJ, Cheng MF, Lee WH, Hsiao M, Chao TY and Yeh CT. Silencing JARID1B suppresses oncogenicity, stemness and increases radiation sensitivity in human oral carcinoma. *Cancer Lett* 2015; 368: 36-45.
- [120] Moon JH, Lee SH, Koo BS, Kim JM, Huang S, Cho JH, Eun YG, Shin HA and Lim YC. Slug is a novel molecular target for head and neck squamous cell carcinoma stem-like cells. *Oral Oncol* 2020; 111: 104948.
- [121] Chen Y, Shao Z, Jiang E, Zhou X, Wang L, Wang H, Luo X, Chen Q, Liu K and Shang Z. CCL21/CCR7 interaction promotes EMT and enhances the stemness of OSCC via a JAK2/STAT3 signaling pathway. *J Cell Physiol* 2020; 235: 5995-6009.
- [122] Yao Z, Du L, Xu M, Li K, Guo H, Ye G, Zhang D, Coppes RP and Zhang H. MTA3-SOX2 module regulates cancer stemness and contributes to clinical outcomes of tongue carcinoma. *Front Oncol* 2019; 9: 816.

Cancer stem cells of head and neck squamous cell carcinoma

- [123] Chen L, Yang QC, Li YC, Yang LL, Liu JF, Li H, Xiao Y, Bu LL, Zhang WF and Sun ZJ. Targeting CMTM6 suppresses stem cell-like properties and enhances antitumor immunity in head and neck squamous cell carcinoma. *Cancer Immunol Res* 2020; 8: 179-191.
- [124] Chen X, Cao Y, Sedhom W, Lu L, Liu Y, Wang H, Oka M, Bornstein S, Said S, Song J and Lu SL. Distinct roles of PIK3CA in the enrichment and maintenance of cancer stem cells in head and neck squamous cell carcinoma. *Mol Oncol* 2020; 14: 139-158.
- [125] Chen SMY, Li B, Nicklawsky AG, Krinsky AL, Brunetti T, Woolaver RA, Wang X, Chen Z, Young CD, Gao D, Wang XJ and Wang JH. Deletion of p53 and hyper-activation of PIK3CA in keratin-15(+) stem cells lead to the development of spontaneous squamous cell carcinoma. *Int J Mol Sci* 2020; 21: 6585.
- [126] Sunkara RR, Sarate RM, Setia P, Shah S, Gupta S, Chaturvedi P, Gera P and Waghmare SK. SFRP1 in skin tumor initiation and cancer stem cell regulation with potential implications in epithelial cancers. *Stem Cell Reports* 2020; 14: 271-284.
- [127] Lu BC, Li J, Yu WF, Zhang GZ, Wang HM and Ma HM. Elevated expression of Nrf2 mediates multidrug resistance in CD133(+) head and neck squamous cell carcinoma stem cells. *Oncol Lett* 2016; 12: 4333-4338.
- [128] Essid N, Chambard JC and Elgaaied AB. Induction of epithelial-mesenchymal transition (EMT) and Gli1 expression in head and neck squamous cell carcinoma (HNSCC) spheroid cultures. *Bosn J Basic Med Sci* 2018; 18: 336-346.
- [129] Pai S, Bamodu OA, Lin YK, Lin CS, Chu PY, Chien MH, Wang LS, Hsiao M, Yeh CT and Tsai JT. CD47-SIRPalpha signaling induces epithelial-mesenchymal transition and cancer stemness and links to a poor prognosis in patients with oral squamous cell carcinoma. *Cells* 2019; 8: 1658.
- [130] Saloura V, Vougiouklakis T, Bao R, Kim S, Baek S, Zewde M, Bernard B, Burkitt K, Nigam N, Izumchenko E, Dohmae N, Hamamoto R and Nakamura Y. WHSC1 monomethylates histone H1 and induces stem-cell like features in squamous cell carcinoma of the head and neck. *Neoplasia* 2020; 22: 283-293.
- [131] Portney BA, Arad M, Gupta A, Brown RA, Khatri R, Lin PN, Hebert AM, Angster KH, Silipino LE, Meltzer WA, Taylor RJ and Zalzman M. ZSCAN4 facilitates chromatin remodeling and promotes the cancer stem cell phenotype. *Oncogene* 2020; 39: 4970-4982.
- [132] Yoshikawa M, Tsuchihashi K, Ishimoto T, Yae T, Motohara T, Sugihara E, Onishi N, Masuko T, Yoshizawa K, Kawashiri S, Mukai M, Asoda S, Kawana H, Nakagawa T, Saya H and Nagano O. xCT inhibition depletes CD44v-expressing tumor cells that are resistant to EGFR-targeted therapy in head and neck squamous cell carcinoma. *Cancer Res* 2013; 73: 1855-1866.
- [133] Xiang Z, Zhou S, Liang S, Zhang G and Tan Y. RCOR1 directly binds to MED28 and weakens its inducing effect on cancer stem cell-like activity of oral cavity squamous cell carcinoma cells. *J Oral Pathol Med* 2020; 49: 741-750.
- [134] Qin L, Yin YT, Zheng FJ, Peng LX, Yang CF, Bao YN, Liang YY, Li XJ, Xiang YQ, Sun R, Li AH, Zou RH, Pei XQ, Huang BJ, Kang TB, Liao DF, Zeng YX, Williams BO and Qian CN. WNT5A promotes stemness characteristics in nasopharyngeal carcinoma cells leading to metastasis and tumorigenesis. *Oncotarget* 2015; 6: 10239-10252.
- [135] Ma SR, Wang WM, Huang CF, Zhang WF and Sun ZJ. Anterior gradient protein 2 expression in high grade head and neck squamous cell carcinoma correlated with cancer stem cell and epithelial mesenchymal transition. *Oncotarget* 2015; 6: 8807-8821.
- [136] Featherston T, Marsh RW, van Schaijik B, Brasch HD, Tan ST and Itinteang T. Expression and localization of cathepsins B, D, and G in two cancer stem cell subpopulations in moderately differentiated oral tongue squamous cell carcinoma. *Front Med (Lausanne)* 2017; 4: 100.
- [137] Morvan VL, Richard E, Cadars M, Fessart D, Broca-Brisson L, Auzanneau C, Pasquies A, Modesto A, Lusque A, Mathoulin-Pelissier S, Lansiaux A and Robert J. Cytochrome P450 1B1 polymorphism drives cancer cell stemness and patient outcome in head-and-neck carcinoma. *Br J Cancer* 2020; 123: 772-784.
- [138] Lorenzo-Martin LF, Menacho-Marquez M and Bustelo XR. Drug vulnerabilities and disease prognosis linked to the stem cell-like gene expression program triggered by the RHO GTPase activator VAV2 in hyperplastic keratinocytes and head and neck cancer. *Cancers (Basel)* 2020; 12: 2498.
- [139] Garcia-Mayea Y, Mir C, Carballo L, Castellvi J, Temprana-Salvador J, Lorente J, Benavente S, Garcia-Pedrero JM, Allonca E, Rodrigo JP and ME LL. TSPAN1: a novel protein involved in head and neck squamous cell carcinoma chemoresistance. *Cancers (Basel)* 2020; 12: 3269.
- [140] Hu X, Zou W, Liu D, Qin G and Jiang L. The down-regulation of TrkB alleviates the malignant biological behavior and cancer stem-like property of laryngeal cancer. *Cancer Manag Res* 2020; 12: 6865-6875.
- [141] Zhang Y, Cai H, Liao Y, Zhu Y, Wang F and Hou J. Activation of PGK1 under hypoxic conditions promotes glycolysis and increases stem cell-like properties and the epithelial-mesenchymal transition in oral squamous cell carcinoma cells via the AKT signalling pathway. *Int J Oncol* 2020; 57: 743-755.
- [142] Ferreira Mendes JM, de Faro Valverde L, Torres Andion Vidal M, Paredes BD, Coelho P, Allahdadi KJ, Coletta RD, Souza BSF and Rocha CAG. Effects of IGF-1 on proliferation, angiogenesis, tumor stem cell populations and activation of AKT and hedgehog pathways in oral squamous cell carcinoma. *Int J Mol Sci* 2020; 21: 6487.

Cancer stem cells of head and neck squamous cell carcinoma

- [143] Zhang M, Hoyle RG, Ma Z, Sun B, Cai W, Cai H, Xie N, Zhang Y, Hou J, Liu X, Chen D, Kellogg GE, Harada H, Sun Y, Wang C and Li J. FOSL1 promotes metastasis of head and neck squamous cell carcinoma through super-enhancer-driven transcription program. *Mol Ther* 2021; 29: 2583-2600.
- [144] Dong J, Li J, Li Y, Ma Z, Yu Y and Wang CY. Transcriptional super-enhancers control cancer stemness and metastasis genes in squamous cell carcinoma. *Nat Commun* 2021; 12: 3974.
- [145] Wang D. Promotive effects of HOXA10 antisense RNA on the stemness of oral squamous cell carcinoma stem cells through a microRNA-29a/MCL-1/phosphatidylinositol 3-kinase/protein kinase B axis. *Arch Oral Biol* 2021; 126: 105114.
- [146] Xu X, Tassone B, Ostano P, Katarkar A, Proust T, Joseph JM, Riganti C, Chiorino G, Kutalik Z, Lefort K and Dotto GP. HSD17B7 gene in self-renewal and oncogenicity of keratinocytes from Black versus White populations. *EMBO Mol Med* 2021; 13: e14133.
- [147] Shigeishi H, Hashikata M, Yokoyama S, Sakuma M, Murozumi H, Kato H, Rahman MZ, Seino S, Ishioka Y, Ohta K, Takechi M and Sugiyama M. CD44(high)/ESA(low) squamous cell carcinoma cell-derived prostaglandin E2 confers resistance to 5-fluorouracil-induced apoptosis in CD44(high)/ESA(high) cells. *Int J Clin Exp Pathol* 2018; 11: 2356-2363.
- [148] Le PN, Keysar SB, Miller B, Eagles JR, Chimed TS, Reisinger J, Gomez KE, Nieto C, Jackson BC, Somerset HL, Morton JJ, Wang XJ and Jimeno A. Wnt signaling dynamics in head and neck squamous cell cancer tumor-stroma interactions. *Mol Carcinog* 2019; 58: 398-410.
- [149] Yu B, Wu K, Wang X, Zhang J, Wang L, Jiang Y, Zhu X, Chen W and Yan M. Periostin secreted by cancer-associated fibroblasts promotes cancer stemness in head and neck cancer by activating protein tyrosine kinase 7. *Cell Death Dis* 2018; 9: 1082.
- [150] Alvarez-Teijeiro S, Garcia-Inclan C, Villaronga MA, Casado P, Hermida-Prado F, Granda-Diaz R, Rodrigo JP, Calvo F, Del-Rio-Ibiate N, Gandarillas A, Moris F, Hermsen M, Cutillas P and Garcia-Pedrero JM. Factors secreted by cancer-associated fibroblasts that sustain cancer stem properties in head and neck squamous carcinoma cells as potential therapeutic targets. *Cancers (Basel)* 2018; 10: 334.
- [151] Wu CP, Du HD, Gong HL, Li DW, Tao L, Tian J and Zhou L. Hypoxia promotes stem-like properties of laryngeal cancer cell lines by increasing the CD133+ stem cell fraction. *Int J Oncol* 2014; 44: 1652-1660.
- [152] He KF, Zhang L, Huang CF, Ma SR, Wang YF, Wang WM, Zhao ZL, Liu B, Zhao YF, Zhang WF and Sun ZJ. CD163+ tumor-associated macrophages correlated with poor prognosis and cancer stem cells in oral squamous cell carcinoma. *Biomed Res Int* 2014; 2014: 838632.
- [153] Kim HS, Chen YC, Nor F, Warner KA, Andrews A, Wagner VP, Zhang Z, Zhang Z, Martins MD, Pearson AT, Yoon E and Nor JE. Endothelial-derived interleukin-6 induces cancer stem cell motility by generating a chemotactic gradient towards blood vessels. *Oncotarget* 2017; 8: 100339-100352.
- [154] Krishnamurthy S, Warner KA, Dong Z, Imai A, Nor C, Ward BB, Helman JI, Taichman RS, Bellile EL, McCauley LK, Polverini PJ, Prince ME, Wicha MS and Nor JE. Endothelial interleukin-6 defines the tumorigenic potential of primary human cancer stem cells. *Stem Cells* 2014; 32: 2845-2857.
- [155] Guan GF, Tang XX, Zhang DJ, Zheng Y, Yu DJ, Zhao Y, Lu YQ and Zhu L. Constitutive secretion of Interleukin-4 dictates CD133+ side population cells to resist drug treatment and cell death. *J BUON* 2015; 20: 1350-1359.
- [156] Lu L, Wang P, Zou Y, Zha Z, Huang H, Guan M, Wu Y and Liu G. IL-1beta promotes stemness of tumor cells by activating smad/ID1 signaling pathway. *Int J Med Sci* 2020; 17: 1257-1268.
- [157] Xu Q, Zhang Q, Ishida Y, Hajjar S, Tang X, Shi H, Dang CV and Le AD. EGF induces epithelial-mesenchymal transition and cancer stem-like cell properties in human oral cancer cells via promoting Warburg effect. *Oncotarget* 2017; 8: 9557-9571.
- [158] Bae WJ, Lee SH, Rho YS, Koo BS and Lim YC. Transforming growth factor beta1 enhances stemness of head and neck squamous cell carcinoma cells through activation of Wnt signaling. *Oncol Lett* 2016; 12: 5315-5320.
- [159] Datta KK, Patil S, Patel K, Babu N, Raja R, Nanjappa V, Mangalparthi KK, Dhaka B, Rajagopalan P, Deolankar SC, Kannan R, Kumar P, Prasad TSK, Mathur PP, Kumari A, Manoharan M, Coral K, Murugan S, Sidransky D, Gupta R, Gupta R, Khanna-Gupta A, Chatterjee A and Gowda H. Chronic exposure to chewing tobacco induces metabolic reprogramming and cancer stem cell-like properties in esophageal epithelial cells. *Cells* 2019; 8: 949.
- [160] Wang TY, Peng CY, Lee SS, Chou MY, Yu CC and Chang YC. Acquisition cancer stemness, mesenchymal trans-differentiation, and chemoresistance properties by chronic exposure of oral epithelial cells to arecoline. *Oncotarget* 2016; 7: 84072-84081.
- [161] Yu MA, Kiang A, Wang-Rodriguez J, Rahimy E, Haas M, Yu V, Ellies LG, Chen J, Fan JB, Brumund KT, Weisman RA and Ongkeko WM. Nicotine promotes acquisition of stem cell and epithelial-to-mesenchymal properties in head and neck squamous cell carcinoma. *PLoS One* 2012; 7: e51967.
- [162] An Y, Kiang A, Lopez JP, Kuo SZ, Yu MA, Abhold EL, Chen JS, Wang-Rodriguez J and Ongkeko WM. Cigarette smoke promotes drug resistance and expansion of cancer stem cell-like side population. *PLoS One* 2012; 7: e47919.

Cancer stem cells of head and neck squamous cell carcinoma

- [163] Yu CC, Chen PN, Peng CY, Yu CH and Chou MY. Suppression of miR-204 enables oral squamous cell carcinomas to promote cancer stemness, EMT traits, and lymph node metastasis. *Oncotarget* 2016; 7: 20180-20192.
- [164] Sun Z, Hu W, Xu J, Kaufmann AM and Albers AE. MicroRNA-34a regulates epithelial-mesenchymal transition and cancer stem cell phenotype of head and neck squamous cell carcinoma in vitro. *Int J Oncol* 2015; 47: 1339-1350.
- [165] Lo WL, Yu CC, Chiou GY, Chen YW, Huang PI, Chien CS, Tseng LM, Chu PY, Lu KH, Chang KW, Kao SY and Chiou SH. MicroRNA-200c attenuates tumour growth and metastasis of presumptive head and neck squamous cell carcinoma stem cells. *J Pathol* 2011; 223: 482-495.
- [166] Lu YC, Cheng AJ, Lee LY, You GR, Li YL, Chen HY and Chang JT. MiR-520b as a novel molecular target for suppressing stemness phenotype of head-neck cancer by inhibiting CD44. *Sci Rep* 2017; 7: 2042.
- [167] Yu CC, Chen YW, Chiou GY, Tsai LL, Huang PI, Chang CY, Tseng LM, Chiou SH, Yen SH, Chou MY, Chu PY and Lo WL. MicroRNA let-7a represses chemoresistance and tumourigenicity in head and neck cancer via stem-like properties ablation. *Oral Oncol* 2011; 47: 202-210.
- [168] Peng CY, Wang TY, Lee SS, Hsieh PL, Liao YW, Tsai LL, Fang CY, Yu CC and Hsieh CS. Let-7c restores radiosensitivity and chemosensitivity and impairs stemness in oral cancer cells through inhibiting interleukin-8. *J Oral Pathol Med* 2018; 47: 590-597.
- [169] Cui M, Chang Y, Fang QG, Du W, Wu JF, Wang JH, Liu ST and Luo SX. Non-coding RNA Pvt1 promotes cancer stem cell-like traits in nasopharyngeal cancer via inhibiting miR-1207. *Pathol Oncol Res* 2019; 25: 1411-1422.
- [170] Yuan Z, Xiu C, Liu D, Zhou G, Yang H, Pei R, Ding C, Cui X, Sun J and Song K. Long noncoding RNA LINC-PINT regulates laryngeal carcinoma cell stemness and chemoresistance through miR-425-5p/PTCH1/SHH axis. *J Cell Physiol* 2019; 234: 23111-23122.
- [171] Liu J, Tang Q, Li S and Yang X. Inhibition of HAX-1 by miR-125a reverses cisplatin resistance in laryngeal cancer stem cells. *Oncotarget* 2016; 7: 86446-86456.
- [172] Lee SP, Hsieh PL, Fang CY, Chu PM, Liao YW, Yu CH, Yu CC and Tsai LL. LINC00963 promotes cancer stemness, metastasis, and drug resistance in head and neck carcinomas via ABCB5 regulation. *Cancers (Basel)* 2020; 12: 1073.
- [173] You X, Zhou Z, Chen W, Wei X, Zhou H and Luo W. MicroRNA-495 confers inhibitory effects on cancer stem cells in oral squamous cell carcinoma through the HOXC6-mediated TGF-beta signaling pathway. *Stem Cell Res Ther* 2020; 11: 117.
- [174] Lin SS, Peng CY, Liao YW, Chou MY, Hsieh PL and Yu CC. miR-1246 targets CCNG2 to enhance cancer stemness and chemoresistance in oral carcinomas. *Cancers (Basel)* 2018; 10: 272.
- [175] Zhou YM, Yao YL, Liu W, Shen XM, Shi LJ and Wu L. MicroRNA-134 inhibits tumor stem cell migration and invasion in oral squamous cell carcinomas via downregulation of PI3K-Akt signaling pathway by inhibiting LAMC2 expression. *Cancer Biomark* 2020; 29: 51-67.