

Review Article

Progress and prospects of the combination of BMI1-targeted therapy and immunotherapy in cervical cancer

Yingying Chen^{1,2}, Shiyu Liu^{1,2}, Xia Yin^{1,2}

¹Department of Obstetrics and Gynecology, West China Second Hospital, Sichuan University, Chengdu, Sichuan, P. R. China; ²Key Laboratory of Birth Defects and Related Diseases of Women and Children, Ministry of Education, Sichuan University, Chengdu, Sichuan, P. R. China

Received September 12, 2024; Accepted January 14, 2025; Epub January 15, 2025; Published January 30, 2025

Abstract: Cervical cancer is one of the most prevalent gynecologic malignancies, posing a significant threat to women's health and survival. Despite advancements in early screening and diagnosis, which have led to cervical cancer being termed a "preventable" cancer, treatment options for advanced and recurrent cervical cancer remain limited. Consequently, identifying new therapeutic targets and treatments is crucial for advancing the research and management of cervical cancer. In recent years, targeted therapy and immunotherapy have become focal points in oncology research, offering new avenues and directions for the treatment of cancer. Preclinical studies have demonstrated that targeting BMI1 can inhibit cervical cancer progression, while immunotherapy has advanced to phase III clinical trials, showing promising results. To date, there have been no reports on the combination of BMI1-targeted therapy and immunotherapy in cervical cancer. This review, therefore, elucidates the current state of research and explores the potential and perspectives of combining targeted therapy with immunotherapy for cervical cancer.

Keywords: BMI1, cervical cancer, targeted therapy, immunotherapy, PTC596

Introduction

Cervical cancer, the most common malignant gynaecological cancer, is a major global health problem and a major cause of disability adjusted life years [1]. It mainly affects women in developing countries, and studies have found it to be associated with persistent infections with high-risk human papillomavirus (HPV) types. The prevention and early diagnosis and treatment of cervical cancer has developed rapidly due to the popularity of cervical cancer screening technology and the attention paid to the disease and the use of HPV vaccine nowadays [2]. The stage of cervical cancer at diagnosis strongly influences treatment approaches and outcomes for cervical cancer patients. According to statistics, the 5-year overall survival rate for all stages combined is 67%: 91% for early stage, 60% for locally advanced disease, and 19% for metastatic disease [3]. Therefore, the treatment of advanced cervical cancer and recurrent and metastatic cervical cancer has

become a key and difficult issue in the treatment of cervical cancer.

With the recent approval of a PD-1 blocking antibody for recurrent or metastatic disease, immunotherapy offers a new method for cancer treatment. Clinical studies on cervical cancer have been conducted since 2015. The notable KEYNOTE-158 phase II clinical trial investigated the use of pembrolizumab as a monotherapy in cervical cancer [4]. Based on the results of this study, the FDA approved pembrolizumab for the treatment of cervical cancer [4]. With more and more in-depth research on immunotherapy for cervical cancer, immunotherapy has also become a second-line treatment strategy for replaced cervical cancer.

Recent years, more and more researches demonstrated that BMI1 significantly overexpressed in many cancers including cervical cancer. Targeting BMI1 in a variety of ways, including induction of autophagy-mediated necrosis, inhi-

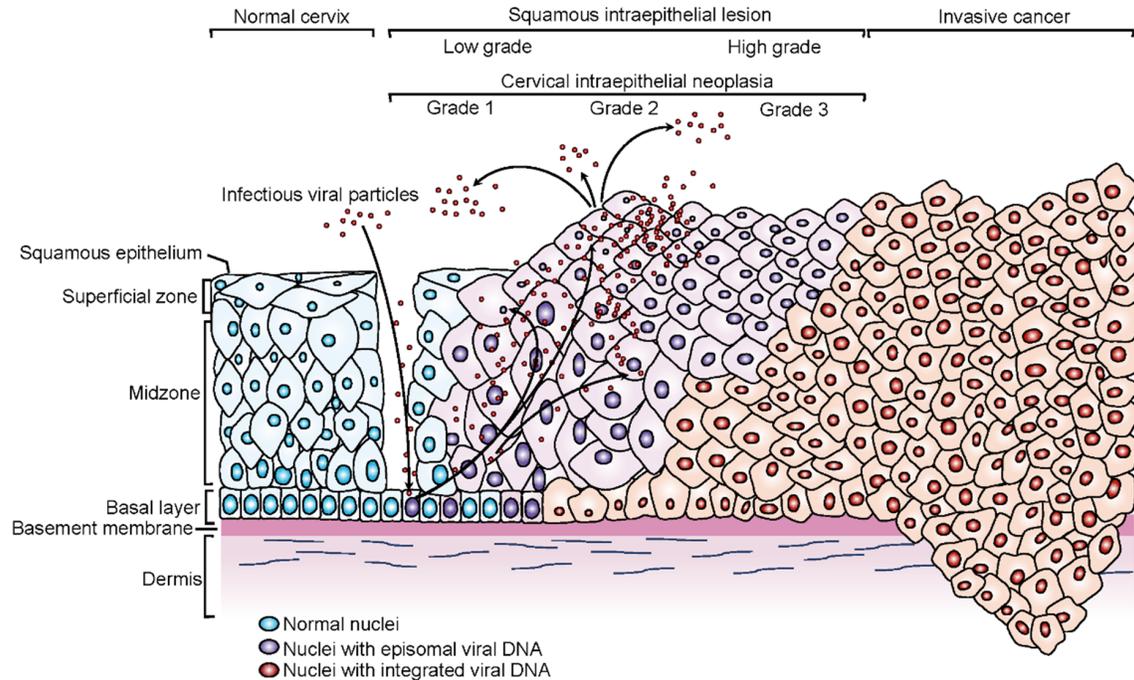


Figure 1. Schematic diagram of the pathogenesis of cervical cancer. Persistent infection of normal cervical epithelium with HPV causes the cervical epithelium to overexpress HPV oncoproteins E6 and E7, gradually leading to malignant transformation of the cervical epithelium.

bition of epithelial-mesenchymal transition and regulation of tumor immunoenvironment, dramatically reduces cancer proliferation and metastasis [5]. BMI1 expression levels are closely correlated with the histological grading of cervical cancer, and the inhibition of cervical cancer cell proliferation, colony formation, and lymph node metastasis [6]. And in other cancer research, BMI1 can enhance the infiltration and activity of CD8+ T cells in the tumor micro-environment, and improve the immunotherapeutic efficacy of PD-1 inhibitors, thereby preventing immune escape [7].

This review explores the potential synergistic effects of BMI1 inhibition and immunotherapy, with the goal of addressing existing therapeutic limitations. Addressing the intricate tumor microenvironment and resistance mechanisms of cervical cancer, this combined approach could provide more effective and sustainable treatment solutions for patients.

Current status of cervical cancer

Cervical cancer is the most prevalent gynecologic malignancy, ranking fourth among all female cancers, posing a significant threat to

women's health and lives [8]. Over 500,000 women are diagnosed with cervical cancer annually, with over 300,000 deaths occurring globally each year [9]. Most cervical cancers are linked to persistent infections with high-risk HPV types, such as 16 and 18. HPV screening and vaccination programs have become effective strategies for cervical cancer prevention [10]. Persistent high-risk HPV infection causes cervical epithelial cells to overexpress the oncoproteins E6 and E7, which inhibit the tumor suppressors p53 and Rb, respectively, leading to the malignant transformation of cervical epithelium [11] (**Figure 1**). These oncoproteins also inhibit apoptosis, destabilize the genome, prevent telomere shortening, promote angiogenesis, and facilitate the invasion and metastasis of cervical cancer [12]. Squamous cell carcinoma and adenocarcinoma are the most common histological subtypes of cervical cancer, comprising approximately 70% and 25% of cases, respectively [13]. Despite advances in prevention, screening, diagnosis, and treatment over the past decade, significant regional and global disparities persist in cervical cancer treatment outcomes. These disparities prompted the International Gynecologic Cancer

BMI1-targeted therapy and immunotherapy in cervical cancer

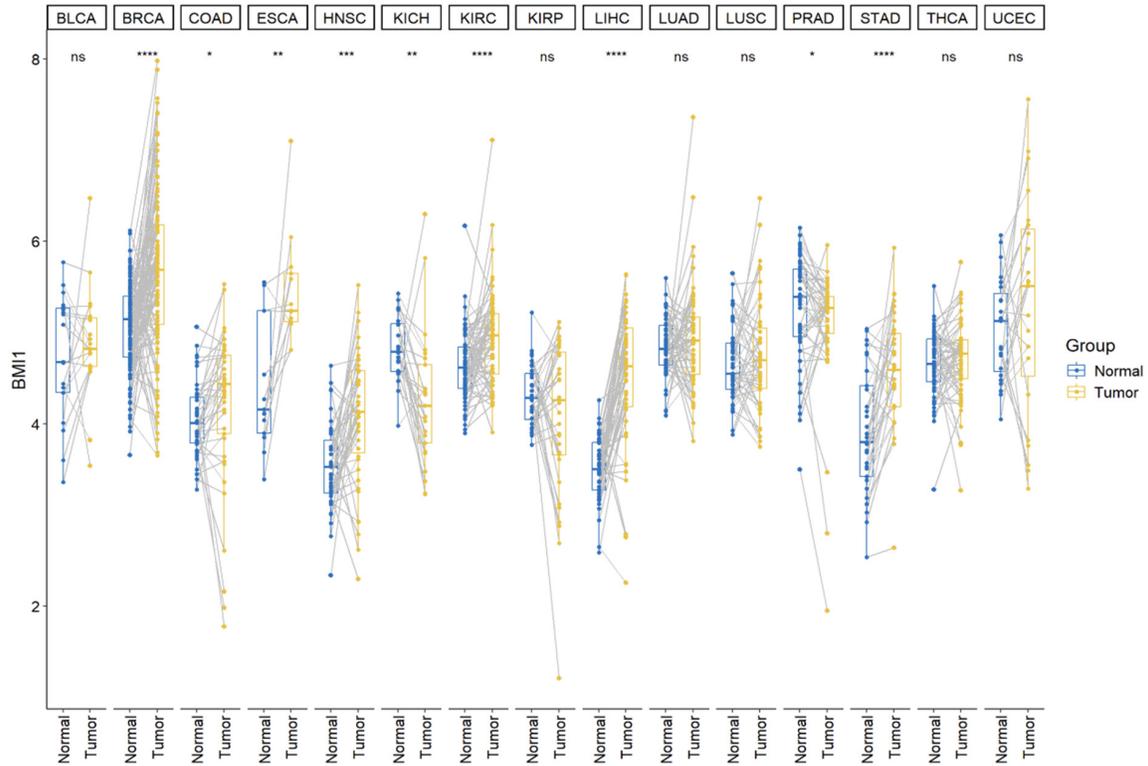


Figure 2. The expression of BMI1 in paired samples of cervical tissue from public databases (TCGA and GTEx).

Society to publish evidence-based management guidelines aimed at improving patient care quality [14]. The treatment of advanced and recurrent cervical cancer remains clinically challenging, prompting researchers to explore new therapeutic approaches.

Association between BMI1 and cervical cancer

Overview of BMI1

The B lymphoma Mo-MLV insertion region 1 homolog (BMI1) gene was first identified in 1991 by the Netherlands Cancer Institute in lymphoma cells. It is a core member of the Polycomb gene family (PcG) with oncogenic properties and is considered a proto-oncogene that collaborates with c-myc to drive cell transformation and tumor formation [15]. Research indicates that PcG proteins are pivotal in the epigenetic modification of chromatin and stem cell self-renewal, governing cell fate decisions, cancer development, and acting as crucial transcriptional repressors [16]. BMI1, a member of the Polycomb repressive complex 1 (PRC1) [17], has recently been recognized as a survival

factor for tissue and cancer stem cells [18], influencing the expression of genes involved in cell growth, proliferation, apoptosis, senescence, and DNA repair [19-21].

Relationship between BMI1 and cervical cancer

BMI1 is reported to be highly expressed in various cancers, including glioma [22], head and neck tumors [23], non-small cell lung cancer [24], gastric cancer [25], prostate cancer [26], ovarian cancer [27] and cervical cancer [28]. Analysis of BMI1 expression in paired samples of cervical tissue from public databases (TCGA and GTEx) indicated that BMI1 expression levels were significantly higher in tumor tissue than in normal tissue (**Figure 2**). Although the expression level of BMI1 is not the highest in cervical cancer, it is higher than the most other tumors (**Figure 2**). High levels of BMI1 have been shown to promote cervical cancer development by binding to the E-Box region of the Sox2 promoter, leading to upregulation of Sox2, a tumor stem cell-associated transcription factor [28]. BMI1 expression levels are closely correlated with the histological grading of cervical

BMI1-targeted therapy and immunotherapy in cervical cancer

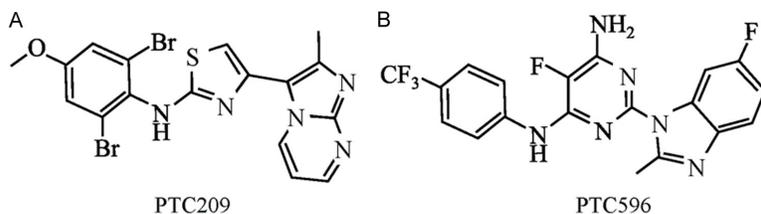


Figure 3. Molecular structure diagrams of BMI1 inhibitors. A. Molecular structure diagram of the first-generation inhibitor PTC209; B. Molecular structure diagram of the second generation inhibitor PTC596.

cancer, and the inhibition of cervical cancer cell proliferation, colony formation, and lymph node metastasis [6]. With the down-regulation of BMI1 expression level, the expression level of STAT3 and pSTAT3 was significantly decreased, which in turn reduced the expression of N-cadherin and Vimentin, leading to a reduction in the occurrence of epithelial mesenchymal transition in cervical cells from the transcriptional level [29]. Targeting BMI1-positive squamous cell carcinoma tumor stem cells has been shown to improve tumor resistance and inhibit tumor growth [23], suggesting that BMI1 could serve as a potential therapeutic target for cervical cancer treatment [30].

BMI1 inhibitor

There are several BMI1 inhibitors used in many researches, such as PTC209, PTC-209 HBr, PTC208 and PTC596. PTC-209 is a first-generation BMI1 inhibitor and is widely used in many oncology studies. PTC-209 HBr is the hydrobromide salt of PTC-209, a potent and selective BMI-1 inhibitor [31], and its function is same to PTC209. PTC-028 is a biologically orally active compound capable of reducing BMI1 levels through post-translational modification. Treatment with PTC-028 selectively inhibited cancer cells without affecting normal cells in clonogenic growth assays and cell viability assays. PTC-028 removes intracellular highly phosphorylated BMI-1 and leads to a transient decrease in ATP levels and impaired mitochondrial redox homeostasis, which further strengthens the caspase-dependent apoptotic response [32]. PTC596, also called unesbulin, is a second-generation BMI-1 inhibitor that accelerates the degradation of BMI-1. PTC596 is now in Phase II clinical trials as it is more biologically safe. In this review, we focus on the PTC209 and PTC596.

PTC209

In 2013, a research team developed the first-generation BMI1-selective inhibitor, PTC209, using high-throughput screening through a proprietary drug discovery platform technology called Gene Expression Modulation by Small-Molecules (GEMS) [19] (The molecular structure of

PTC209 is shown in **Figure 3A**). PTC-209 inhibits UTR-mediated expression of reporter genes and endogenous BMI-1 in human colorectal HCT116 and human fibrosarcoma HT1080 tumor cells. And PTC-209 reduces the growth of rectal tumor cells dependent on BMI-1. In addition, PTC-209 damages tumor initiating cells by inhibiting their irreversible growth [19]. Subsequent studies have reported that PTC209 was effective in preclinical trials against various tumors, including colorectal cancer [33], breast cancer, bile duct cancer [34], glioma [35], prostate cancer [26], lung cancer [36], head and neck tumors [23], and cervical cancer [37]. Additionally, evidence suggests that PTC209 induces cell cycle arrest and apoptosis in cervical cancer cell lines [37].

PTC596

Compared to PTC209, PTC596, a second-generation BMI1 inhibitor (**Figure 3B**), is cell-permeable and promotes the degradation of BMI1 protein at nanomolar concentrations (as opposed to micromolar concentrations for PTC209) [38]. Furthermore, PTC209 has not yet entered clinical trials due to limited efficacy and poor pharmacokinetic properties, whereas PTC596, with a promising safety profile, has advanced to early clinical trials [39] (ClinicalTrials.gov Identifiers: NCT02404480, NCT03206645, NCT03605550, NCT03761095) (**Table 1**). Therefore, PTC596 shows greater effectiveness than PTC209 in cancer treatment.

PTC596 is a novel small-molecule inhibitor of BMI1, and recent studies have demonstrated its ability to inhibit cancer cell proliferation and promote apoptosis both in vivo and in vitro [40]. PTC596 effectively reduces the function, activity, and quantity of BMI1 through phosphorylation, accelerates BMI1 protein degradation, and subsequently inhibits BMI1-mediated sig-

Table 1. Ongoing clinical trials of PTC596

Clinical Trial Title	ClinicalTrials.gov Identifiers	Study population	Number of participants	Clinical Trial Phase	Treatment options
PTC596 in combination with dacarbazine for advanced smooth muscle sarcoma (LMS) participant study	NCT03761095	Locally recurrent, unresectable, or metastatic recurrent/refractory smooth muscle sarcoma	41	Phase 1B	PTC596 + Dacarbazine
Phase 1b study of PTC596 in children with newly diagnosed diffuse pontine glioma and high-grade glioma	NCT03605550	Diffuse pontine glioma and high-grade glioma	54	Phase 1B	PTC596 + Radiotherapy
Study of PTC596 in patients with advanced smooth muscle sarcoma	NCT05269355	Advanced smooth muscle sarcoma	345	Phase 2/3	PTC596 + Dacarbazine
PTC596 for patients with advanced solid tumors	NCT02404480	Advanced solid tumors	31	Phase 1	PTC596
PTC596 for women with ovarian cancer receiving neoadjuvant chemotherapy	NCT03206645	High-grade plasmacytoid ovarian cancer	27	Phase 1B	PTC596 + Carboplatin + Paclitaxel

naling pathways to reduce tumor growth in vitro, PTC596 preferentially targets chemotherapy-resistant tumor stem cells [38]. Studies on PTC596's inhibition of BMI1 function have so far been limited to lymphoma, acute leukemia, ovarian cancer, and glioma, but it has entered clinical trials for the treatment of advanced solid tumors, including advanced ovarian cancer. In phase I clinical trials, PTC596 was well tolerated by patients, with manageable gastrointestinal side effects (ClinicalTrials.gov Identifiers: NCT03206645). Recent studies report that PTC596 inhibits BMI1 expression in cervical cancer and promotes Mcl-1 degradation and apoptosis in HeLa (cervical cancer) and Caki (kidney cancer) cells by downregulating DUB3 levels, thereby inhibiting tumor progression [41].

BMI1 and tumor microenvironment

Tumor microenvironment

Definition and components of tumor microenvironment: The tumor microenvironment (TME) is a critical component of tumor development. The tumor microenvironment consists of not only the tumor cells but also the surrounding components, such as stromal cells, vascular endothelial cells, and associated immune cells, including tumor-associated fibroblasts, T cells, NK cells, macrophages, and B cells [42] (**Figure 4**).

Despite the complexity of the tumor immune microenvironment, the advent of single-cell sequencing technology has enabled a detailed dissection of its nature [43]. Understanding the

fundamental role of the tumor microenvironment in cancer evolution has shifted the perspective from a tumor cell-centered view to the concept of a complex tumor ecosystem that supports tumor growth and metastasis [44].

Immune cells in the tumor microenvironment: Tumor-associated immune cells are generally classified into two categories: anti-tumor immune cells and pro-tumor immune cells, each playing distinct roles at various stages of tumor progression.

Anti-tumor immune cells primarily include effector T cells (CD8+ cytotoxic T cells and CD4+ effector T cells), Natural Killer (NK) cells, Dendritic cells (DCs), M1-polarized macrophages, and N1-polarized neutrophils, while tumor-promoting immune cells mainly comprise regulatory T cells (Tregs) and myeloid-derived suppressor cells (MDSCs) [45]. This discussion focuses on CD8+ cytotoxic T cells (CTLs). CD8+ CTLs have long been recognized as the primary lymphocyte subpopulation responsible for killing cancer cells presenting Major Histocompatibility Complex Class I (MHC-I) molecules [46]. Upon antigen presentation by DCs, CD8+ T cells can be induced to differentiate into cytotoxic effector CD8+ T cells [47]. Guided by chemokines secreted by DCs, such as CXCL9 and CXCL10, activated CTLs migrate into the inflammatory environment through the expression of CXCR3 [48, 49]. The interaction between ligands on DCs (CD70 and CD80-CD86) and receptors on CD8+ T cells (CD27 and CD28) is considered a critical step in CD8+ T cell activation [47].

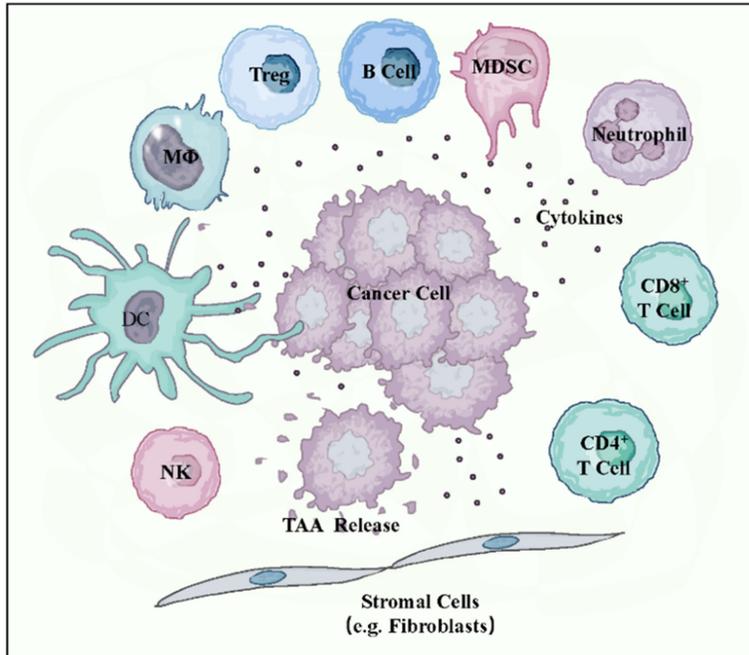


Figure 4. Tumor microenvironment pattern map. TAA: Tumor Associated Antigens; DCs: Dendritic cells; NK: Natural Killer. The tumor microenvironment includes immune cells, stromal cells, endothelial cells, and tumor-associated fibroblasts in addition to tumor cells.

Immune checkpoints: The activation and regulation of CTLs require two signals: one from the T-cell receptor (TCR) and the other from receptors known as immune checkpoints (ICs) [50]. Immune checkpoints are categorized into two types: suppressive checkpoints (e.g., CTLA-4 [51], PD-1 [52], TIM-3 [53], LAG-3 [54], TIGIT [55] and CD96 [56]) and stimulatory immune checkpoints (e.g., ICOS [57], OX-40 [58], 4-1BB [59], GITR [60], CD27 [61], HVEM [62], CD40L [63]).

Further research has revealed that in certain tumors, cancer cells inhibit CTLs by expressing programmed death-ligand 1 (PD-L1), which interacts with the inhibitory checkpoint receptor programmed death receptor 1 (PD-1), a key mechanism of immune escape in cancer [64]. Additionally, within the tumor microenvironment (TME), persistent antigenic and inflammatory responses can drive CTLs into a state of “exhaustion”, resulting in T-cell dysfunction and promoting tumor development [65].

Programmed death receptors and programmed death ligands: Programmed cell death protein 1 (PD-1; also known as CD279) is a co-suppressor receptor expressed on the surface of acti-

vated T cells following antigen stimulation [66]. PD-1 interacts with two ligands: programmed death-ligand 1 (PD-L1; also known as CD-274) and programmed death-ligand 2 (PD-L2; also known as CD273). PD-L1 is typically found in tumor cells and tumor stroma, where it is highly expressed. Its binding to PD-1 alters T cell activity through multiple pathways, inhibiting T cell proliferation, survival, cytokine production, and other effector functions [67], thereby suppressing antitumor immunity. Currently, antibodies targeting the PD-L1-PD-1 axis are under evaluation in over 1000 clinical trials and have been approved by the Food and Drug Administration (FDA) for the treatment of melanoma, non-small cell lung cancer (NSCLC), renal

cell carcinoma (RCC), Hodgkin’s lymphoma, bladder cancer, head and neck squamous cell carcinoma (HNSCC), Merkel cell carcinoma, and microsatellite instability-high (MSI-H) or mismatch repair-deficient (dMMR) solid tumors [68].

BMI1 and tumor immune microenvironment

With in-depth studies of the tumor microenvironment, BMI1 has been found to regulate the immune microenvironment of tumors and play a significant role, and involved in the regulations of many immune cells, such as T cell. In multiple myeloma (MM), BMI1 regulates the pro-myeloma characteristics of tumor-associated macrophages (TAMs) [69]. BMI1 has also been reported to inhibit IL-10 expression in macrophages during the acute phase response induced by lipopolysaccharide (LPS) [70]. In pancreatic cancer, BMI1 inhibits antitumor immunity by reducing NK cell-mediated killing through the suppression of MHC class I chain-related protein A/B (MICA/B) expression. Additionally, the study showed that BMI1 suppresses GATA2 in pancreatic cancer cells, leading to the downregulation of MICA/B expression and ultimately promoting immune escape [71].

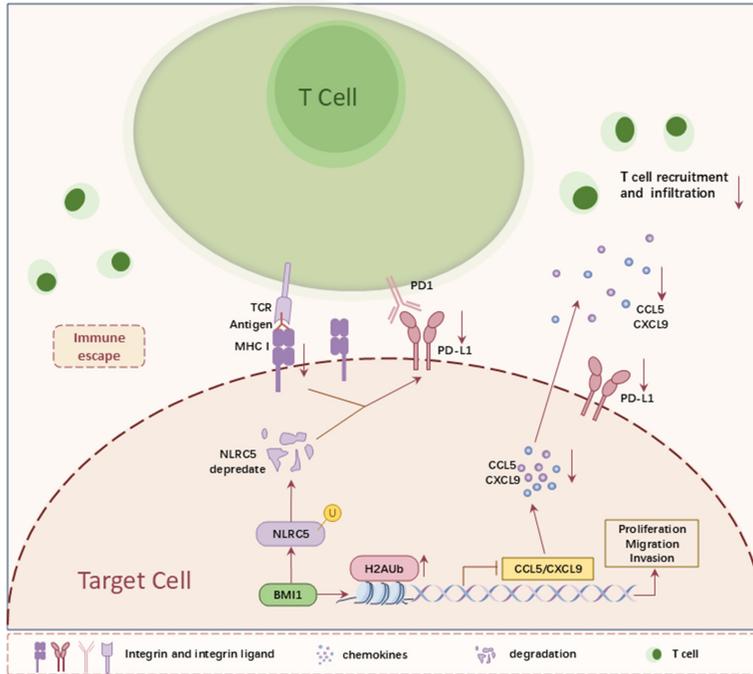


Figure 5. The diagram related to the potential mechanism of BMI1 regulating T cell function.

BMI1 is known to mediate gene silencing via H2AUb and to repress the expression of genes [72]. As the **Figure 5** showed that a research indicated that BMI1 inhibitors reduced the expression of H2AUb, thereby decreasing the transcriptional levels of CCL5 and CXCL9, further reducing the recruitment process of CD8+ T lymphocytes in cholangiocarcinoma (CCA) [73]. Furthermore, BMI1 maintains antigen-specific CD8+ T-cell clonal expansion during chronic viral infection, and BMI1 deficiency promotes senescence [74]. And another research indicated that BMI1 regulates the clonal expansion of T cells in the hepatocellular carcinoma (HCC) microenvironment by proteomic analysis of TIL fractions in two murine HCC models, followed by flow cytometry analysis [75]. In head and neck tumors, BMI1 inhibitors can target cancer stem cells (CSCs), enhance the infiltration and activity of CD8+ T cells in the tumor microenvironment, and improve the immunotherapeutic efficacy of PD-1 inhibitors, thereby preventing immune escape [7]. In addition, BMI1 induces ubiquitination and protein degradation of NLRCS and suppresses HLA class I expression, and consequently inhibit T cell activation and increased PD-1/PD-L1 levels in the co-culture system, which potentially helps immune escape in non-small cell lung cancer (NSCLC) [76]. Therefore, BMI1 plays a regula-

tory role in the tumor immune microenvironment and contributes to tumorigenesis.

Current status of immune microenvironment and immunotherapy for cervical cancer

Characteristics of the immune microenvironment in cervical cancer

Cervical cancer cells evade the immune system by inducing an immunosuppressive state within their microenvironment [77, 78]. High-risk subtypes of human papillomavirus (hr-HPV), such as types 16 and 18, are more likely to persist and integrate into the host genome, resulting in the overexpression of oncoproteins E6 and E7 [79]. These oncoproteins downregulate key pathways that disrupt

natural immune responses, including interferon production and the cGAS/STING pathway, and inhibit HPV antigen presentation via MHC class I molecules [80]. Additionally, HPV oncoproteins prevent regulatory transcription factor 9 (IRF9) from binding to phosphorylated STAT1 and STAT2, and inhibit STAT1 and STAT2 phosphorylation by interacting with tyrosine kinase 2 (TYK2), which in turn inhibits the IFN- α/β receptor (IFNAR) pathway, disrupting the positive feedback loop in virus-infected cells [81].

Intensive research on the immune system has identified the Jak/STAT signaling pathway as a critical communication hub [82], playing a pivotal role in cancer progression. In June 2018, the US FDA approved pembrolizumab, the first immunotherapy drug for the treatment of advanced and recurrent cervical cancer, based on the findings of the KEYNOTE-158 phase II clinical trial [4], marking significant progress in cervical cancer immunotherapy. In acquired immunity, signaling pathways such as Jak/STAT, PI3K, MAPK, and NF- κ B are reported to be involved in the induction of PD-L1 expression by IFN- γ [83, 84]. The Jak/STAT signaling pathway is essential for PD-L1 expression and contributes to drug resistance [85] and as reported, it not only upregulates PD-L1 but also

influences tumor antigen expression [86]. Jak1 is crucial for IFN- γ -mediated immune responses and MHC class I/II expression, while Jak2 promotes IFN- γ -induced STAT5 phosphorylation and PD-L1 expression, thereby suppressing antitumor immunity [86]. In pancreatic cancer, gemcitabine upregulates PD-L1 expression via the Jak/STAT signaling pathway, leading to chemoresistance [87].

Tumor immunotherapy

Tumor immunotherapy treats cancer by enhancing the body's immune capacity to eliminate malignant cells, representing a major breakthrough in tumor research. Currently, immunotherapy is clinically employed to treat various cancers, including melanoma [88], non-small cell lung cancer [89], renal cell carcinoma [90] and triple-negative breast cancer [91], with notable efficacy, though the precise mechanisms remain unclear. Historically, immunotherapy has encompassed lysovirus therapy [92], cancer vaccine therapy [93], cytokine therapy [94], relay immune cell therapy [95] and immune checkpoint inhibitor therapy [96]. Adoptive cell transfer therapy and immune checkpoint inhibitor therapy have gained prominence in immunotherapy due to their superior clinical efficacy [43], with their success demonstrating that immune cells, particularly T cells, can effectively eliminate tumor cells. Immune cells form the cellular foundation of immunotherapy, making the understanding of immune infiltration in the tumor microenvironment crucial for enhancing immunotherapy responsiveness and developing new therapeutic strategies [97]. T cells have emerged as the focus of tumor immunotherapy research due to their potent tumor-killing capabilities [98]. T cell function is initiated by the binding of the T cell receptor (TCR) to major histocompatibility complex (MHC) molecules or human leukocyte antigens (HLAs) presenting tumor antigen peptides [99].

Immune checkpoints are molecules within co-suppressive signaling pathways between immune cells and target cells that maintain immune tolerance, but cancer cells often exploit these pathways to evade immune surveillance [88, 100]. The interaction between PD-1 and PD-L1 is the co-suppressive mechanism most commonly responsible for immune

escape [101]. PD-1/PD-L1-based immune checkpoint blockade therapies have demonstrated significant efficacy across various tumors, but the emergence of drug resistance has posed a major challenge in immunotherapy, attributed to T cell dysfunction, impaired antigen recognition, and T cell exhaustion [102]. Consequently, many researchers have proposed combination strategies in immunotherapy to address these challenges. The combined application of radiotherapy and immunotherapy has demonstrated significant advantages in the clinical treatment of tumors. Immunotherapy can reduce radiotherapy resistance in non-small cell tumors, while radiotherapy can enhance immune infiltration and significantly improve patient prognosis [103].

Current status of research on immunotherapy for cervical cancer

Clinical studies on cervical cancer have been conducted since 2015. The notable KEYNOTE-158 phase II clinical trial investigated the use of pembrolizumab as a monotherapy in cervical cancer [4]. Based on the results of this study, the FDA approved pembrolizumab for the treatment of cervical cancer [4]. Another notable study is CheckMate-358, which investigated the use of nivolumab as monotherapy in cervical cancer patients [104]. These studies demonstrated that pembrolizumab and nivolumab exhibit antitumor activity in advanced/recurrent cervical cancer. Pembrolizumab's efficacy may be associated with PD-L1 expression as a potential predictive marker, whereas for nivolumab, positive PD-L1 expression was not a predictor of efficacy. Further studies with larger sample sizes are needed to validate these findings [105].

To expand and advance the indications, the pivotal phase III clinical trial KEYNOTE-826 evaluated the efficacy of pembrolizumab in combination with chemotherapy, with or without bevacizumab, for the first-line treatment of cervical cancer, regardless of PD-L1 expression status [106] (Table 2). The results, presented at the European Society of Medical Oncology (ESMO) Congress in 2021, marked the first phase III trial with positive outcomes for both progression-free survival (PFS) and overall survival (OS) as dual endpoints in first-line PD-1/PD-L1 treatment for cervical cancer, indepen-

BMI1-targeted therapy and immunotherapy in cervical cancer

Table 2. Ongoing phase III clinical trials with immune checkpoint inhibitors for the treatment of cervical cancer

Clinical Trial Title	ClinicalTrials.gov Identifiers	Study population	Number of participants	Clinical Trial Phase	Treatment options
Efficacy and safety of BCD-100 (anti-PD-1) in combination with platinum-based chemotherapy with or without bevacizumab as first-line treatment in subjects with advanced cervical cancer (FERMATA)	NCT03912415	Advanced cervical cancer	316	Phase III	Paclitaxel + cisplatin (or carboplatin)/ bevacizumab/BCD-100 (anti-PD1)
Efficacy and safety study of pablizumab (MK-3475) in combination with chemotherapy versus placebo in the first-line treatment of women with persistent, recurrent or metastatic cervical cancer (MK-3475- 826/KEYNOTE-826)	NCT03635567	Recurrent or metastatic cervical cancer	600	Phase III	Paclitaxel + Cisplatin (or Carboplatin)/ Bevacizumab/Pembrolizumab
Platinum chemotherapy plus paclitaxel with bevacizumab and atezumab for metastatic carcinoma of the cervix	NCT03556839	Recurrent or metastatic cervical cancer	404	Phase III	Paclitaxel + cisplatin/bevacizumab/ atezumab

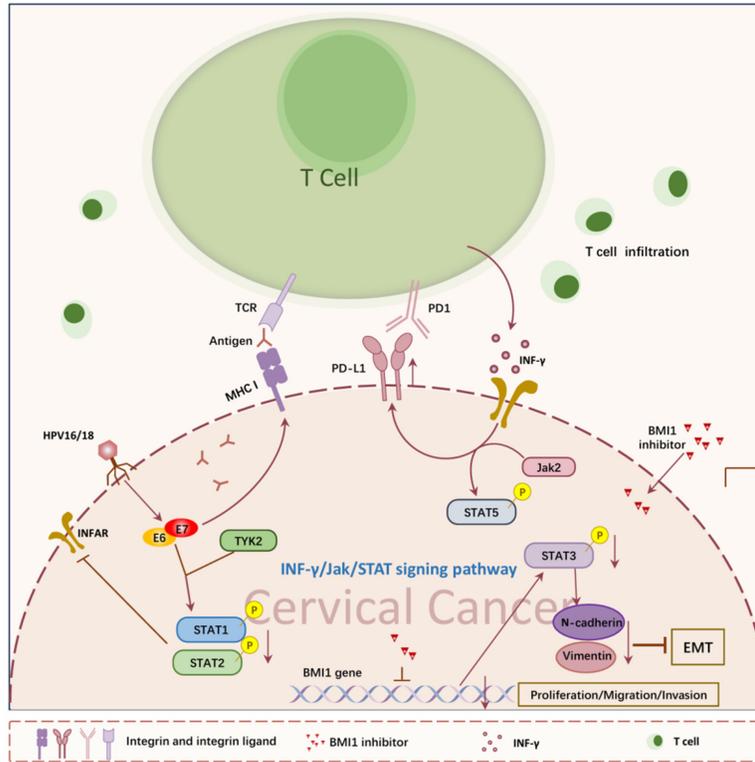


Figure 6. The diagram related to the potential mechanism of BMI1 inhibitor combining immunotherapy in cervical cancer.

dent of PD-L1 expression status [106]. The KEYNOTE-826 study demonstrated that pembrolizumab combined with chemotherapy, with or without bevacizumab, significantly improved OS and PFS in first-line patients with advanced metastatic cervical cancer, offering a reliable safety profile compared to placebo, regardless of PD-L1 status [106]. This regimen is anticipated to become a new standard of care for the first-line treatment of advanced recurrent and metastatic cervical cancer, marking a new chapter in the clinical management of this patient population.

Summary and outlook

Cervical cancer is the most common malignant tumor of the female reproductive system, with an increasing incidence rate, particularly among younger women, posing a significant threat to women’s health and life. Clinically, managing advanced and recurrent cervical cancer remains challenging, as conventional treatments like surgery, radiotherapy, and chemotherapy are often ineffective. Thus, explor-

ing new therapeutic targets and treatments for cervical cancer is crucial.

With advancements in molecular biology and genomics, targeted therapies and immunotherapies have become prominent research areas in cervical cancer treatment. Targeted therapy, often referred to as a “biological missile”, specifically binds to defined oncogenic sites at the molecular level, killing tumor cells while sparing normal tissues, thereby improving efficacy and reducing toxic side effects [107]. Immunotherapy is a therapeutic approach that stimulates the host’s immune system to actively or passively produce anti-tumor immunity through in vitro intervention. It has been used with remarkable success to treat various tumors, including melanoma, non-small cell lung cancer,

renal cell carcinoma, ovarian cancer, and cervical cancer [108]. Targeting the BMI1 protein has been shown to inhibit the progression of cervical cancer, and immunotherapy for cervical cancer has also entered phase III clinical trials with promising results. And some researches showed that BMI1 inhibitor can influence the Jak/STAT signaling pathway, which is critical pathway in cervical cancer immunoenvironment (Figure 6). Therefore, researching targeted therapy and immunotherapy methods can provide valuable data and insights for treating cervical cancer, particularly advanced and recurrent cases, and offer practical opportunities for prolonging patient survival and improving quality of life.

Disclosure of conflict of interest

None.

Address correspondence to: Dr. Xia Yin, Department of Obstetrics and Gynecology, West China Second Hospital, Sichuan University, Renmin South Road, Chengdu 610041, Sichuan, P. R. China. Tel: +86-

BMI1-targeted therapy and immunotherapy in cervical cancer

13980079129; Fax: +86-02885570644; E-mail: xiadashazi@foxmail.com

References

- [1] D’Oria O, Corrado G, Laganà AS, Chiantera V, Vizza E and Giannini A. New advances in cervical cancer: from bench to bedside. *Int J Environ Res Public Health* 2022; 19: 7094.
- [2] Elakkiya R, Subramaniaswamy V, Vijayakumar V and Mahanti A. Cervical cancer diagnostics healthcare system using hybrid object detection adversarial networks. *IEEE J Biomed Health Inform* 2022; 26: 1464-1471.
- [3] Siegel RL, Giaquinto AN and Jemal A. Cancer statistics, 2024. *CA Cancer J Clin* 2024; 74: 12-49.
- [4] Chung HC, Ros W, Delord JP, Perets R, Italiano A, Shapira-Frommer R, Manzuk L, Piha-Paul SA, Xu L, Zeigenfuss S, Pruitt SK and Leary A. Efficacy and safety of pembrolizumab in previously treated advanced cervical cancer: results from the phase II KEYNOTE-158 study. *J Clin Oncol* 2019; 37: 1470-1478.
- [5] Ye K, Chen QW, Sun YF, Lin JA and Xu JH. Loss of BMI-1 dampens migration and EMT of colorectal cancer in inflammatory microenvironment through TLR4/MD-2/MyD88-mediated NF- κ B signaling. *J Cell Biochem* 2018; 119: 1922-1930.
- [6] Jiang Y, Su B, Meng X, Liu C, Liu B, Liu D, Fan Y and Yang H. Effect of siRNA-mediated silencing of Bmi-1 gene expression on HeLa cells. *Cancer Sci* 2010; 101: 379-386.
- [7] Jia L, Zhang W and Wang CY. BMI1 inhibition eliminates residual cancer stem cells after PD1 blockade and activates antitumor immunity to prevent metastasis and relapse. *Cell Stem Cell* 2020; 27: 238-253, e236.
- [8] Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA and Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2018; 68: 394-424.
- [9] Cohen PA, Jhingran A, Oaknin A and Denny L. Cervical cancer. *Lancet* 2019; 393: 169-182.
- [10] Crosbie EJ, Einstein MH, Franceschi S and Kitchener HC. Human papillomavirus and cervical cancer. *Lancet* 2013; 382: 889-899.
- [11] Roden RBS and Stern PL. Opportunities and challenges for human papillomavirus vaccination in cancer. *Nat Rev Cancer* 2018; 18: 240-254.
- [12] Pal A and Kundu R. Human papillomavirus E6 and E7: the cervical cancer hallmarks and targets for therapy. *Front Microbiol* 2020; 10: 3116.
- [13] Small W Jr, Bacon MA, Bajaj A, Chuang LT, Fisher BJ, Harkenrider MM, Jhingran A, Kitchener HC, Mileskin LR, Viswanathan AN and Gaffney DK. Cervical cancer: a global health crisis. *Cancer* 2017; 123: 2404-2412.
- [14] Cibula D, Pötter R, Planchamp F, Avall-Lundqvist E, Fischerova D, Haie Meder C, Köhler C, Landoni F, Lax S, Lindegaard JC, Mahantshetty U, Mathevet P, McCluggage WG, McCormack M, Naik R, Nout R, Pignata S, Ponce J, Querleu D, Raspagliesi F, Rodolakis A, Tamussino K, Wimberger P and Raspollini MR. The European Society of Gynaecological Oncology/European Society for Radiotherapy and Oncology/European Society of Pathology guidelines for the management of patients with cervical cancer. *Radiother Oncol* 2018; 127: 404-416.
- [15] van Lohuizen M, Frasch M, Wientjens E and Berns A. Sequence similarity between the mammalian bmi-1 proto-oncogene and the Drosophila regulatory genes Psc and Su(z)2. *Nature* 1991; 353: 353-355.
- [16] Smith RA, Andrews KS, Brooks D, Fedewa SA, Manassaram-Baptiste D, Saslow D, Brawley OW and Wender RC. Cancer screening in the United States, 2018: a review of current American Cancer Society guidelines and current issues in cancer screening. *CA Cancer J Clin* 2018; 68: 297-316.
- [17] Jacobs JJ, Kieboom K, Marino S, DePinho RA and van Lohuizen M. The oncogene and Polycomb-group gene bmi-1 regulates cell proliferation and senescence through the ink4a locus. *Nature* 1999; 397: 164-168.
- [18] Liu S, Dontu G, Mantle ID, Patel S, Ahn NS, Jackson KW, Suri P and Wicha MS. Hedgehog signaling and Bmi-1 regulate self-renewal of normal and malignant human mammary stem cells. *Cancer Res* 2006; 66: 6063-6071.
- [19] Kreso A, van Galen P, Pedley NM, Lima-Fernandes E, Frelin C, Davis T, Cao L, Baiazitov R, Du W, Sydorenko N, Moon YC, Gibson L, Wang Y, Leung C, Iscove NN, Arrowsmith CH, Szentgyorgyi E, Gallinger S, Dick JE and O’Brien CA. Self-renewal as a therapeutic target in human colorectal cancer. *Nat Med* 2014; 20: 29-36.
- [20] M JR and S V. BMI1 and PTEN are key determinants of breast cancer therapy: a plausible therapeutic target in breast cancer. *Gene* 2018; 678: 302-311.
- [21] McGinty RK, Henrici RC and Tan S. Crystal structure of the PRC1 ubiquitylation module bound to the nucleosome. *Nature* 2014; 514: 591-596.
- [22] Jin X, Kim LJY, Wu Q, Wallace LC, Prager BC, Sanvoranart T, Gimple RC, Wang X, Mack SC, Miller TE, Huang P, Valentim CL, Zhou QG, Barnholtz-Sloan JS, Bao S, Sloan AE and Rich JN. Targeting glioma stem cells through combined BMI1 and EZH2 inhibition. *Nat Med* 2017; 23: 1352-1361.

BMI1-targeted therapy and immunotherapy in cervical cancer

- [23] Chen D, Wu M, Li Y, Chang I, Yuan Q, Ekimyan-Salvo M, Deng P, Yu B, Yu Y, Dong J, Szymanski JM, Ramadoss S, Li J and Wang CY. Targeting BMI1(+) cancer stem cells overcomes chemoresistance and inhibits metastases in squamous cell carcinoma. *Cell Stem Cell* 2017; 20: 621-634, e626.
- [24] Crunkhorn S. Cancer: BMI1 inhibition reverses lung cancer. *Nat Rev Drug Discov* 2016; 15: 678.
- [25] Li Y, Tian Z, Tan Y, Lian G, Chen S, Chen S, Li J, Li X, Huang K and Chen Y. Bmi-1-induced miR-27a and miR-155 promote tumor metastasis and chemoresistance by targeting RKIP in gastric cancer. *Mol Cancer* 2020; 19: 109.
- [26] Yoo YA, Vatapalli R, Lysy B, Mok H, Desouki MM and Abdulkadir SA. The role of castration-resistant Bmi1+Sox2+ cells in driving recurrence in prostate cancer. *J Natl Cancer Inst* 2019; 111: 311-321.
- [27] Zhao Q, Qian Q, Cao D, Yang J, Gui T and Shen K. Role of BMI1 in epithelial ovarian cancer: investigated via the CRISPR/Cas9 system and RNA sequencing. *J Ovarian Res* 2018; 11: 31.
- [28] Xu R, Chen L and Yang WT. Aberrantly elevated Bmi1 promotes cervical cancer tumorigenicity and tumor sphere formation via enhanced transcriptional regulation of Sox2 genes. *Oncol Rep* 2019; 42: 688-696.
- [29] Sun X, Xu H, Dai T, Xie L, Zhao Q, Hao X, Sun Y, Wang X, Jiang N and Sang M. Alantolactone inhibits cervical cancer progression by downregulating BMI1. *Sci Rep* 2021; 11: 9251.
- [30] Rong X, Gao W, Yang X and Guo J. Downregulation of hsa_circ_0007534 restricts the proliferation and invasion of cervical cancer through regulating miR-498/BMI-1 signaling. *Life Sci* 2019; 235: 116785.
- [31] Xu LB, Qin YF, Su L, Huang C, Xu Q, Zhang R, Shi XD, Sun R, Chen J, Song Z, Jiang X, Shang L, Xiao G, Kong X, Liu C and Wong PP. Cathepsin-facilitated invasion of BMI1-high hepatocellular carcinoma cells drives bile duct tumor thrombi formation. *Nat Commun* 2023; 14: 7033.
- [32] Dey A, Xiong X, Crim A, Dwivedi SKD, Mustafi SB, Mukherjee P, Cao L, Sydorenko N, Baiazitov R, Moon YC, Dumble M, Davis T and Bhat-tacharya R. Evaluating the mechanism and therapeutic potential of PTC-028, a novel inhibitor of BMI-1 function in ovarian cancer. *Mol Cancer Ther* 2018; 17: 39-49.
- [33] Xu J, Zhang Y, Xu J, Wang M, Liu G, Wang J, Zhao X, Qi Y, Shi J, Cheng K, Li Y, Qi S and Nie G. Reversing tumor stemness via orally targeted nanoparticles achieves efficient colon cancer treatment. *Biomaterials* 2019; 216: 119247.
- [34] Mayr C, Neureiter D, Wagner A, Pichler M and Kiesslich T. The role of polycomb repressive complexes in biliary tract cancer. *Expert Opin Ther Targets* 2015; 19: 363-375.
- [35] Kong Y, Ai C, Dong F, Xia X, Zhao X, Yang C, Kang C, Zhou Y, Zhao Q, Sun X and Wu X. Targeting of BMI-1 with PTC-209 inhibits glioblastoma development. *Cell Cycle* 2018; 17: 1199-1211.
- [36] Yong KJ, Basseres DS, Welner RS, Zhang WC, Yang H, Yan B, Alberich-Jorda M, Zhang J, de Figueiredo-Pontes LL, Battelli C, Hetherington CJ, Ye M, Zhang H, Maroni G, O'Brien K, Magli MC, Borczuk AC, Varticovski L, Kocher O, Zhang P, Moon YC, Sydorenko N, Cao L, Davis TW, Thakkar BM, Soo RA, Iwama A, Lim B, Halmos B, Neuberger D, Tenen DG and Levantini E. Targeted BMI1 inhibition impairs tumor growth in lung adenocarcinomas with low CEBP α expression. *Sci Transl Med* 2016; 8: 350ra104.
- [37] Li J, Vangundy Z and Poi M. PTC209, a specific inhibitor of BMI1, promotes cell cycle arrest and apoptosis in cervical cancer cell lines. *Anticancer Res* 2020; 40: 133-141.
- [38] Nishida Y, Maeda A, Kim MJ, Cao L, Kubota Y, Ishizawa J, AlRawi A, Kato Y, Iwama A, Fujisawa M, Matsue K, Weetall M, Dumble M, Andreeff M, Davis TW, Branstrom A, Kimura S and Kojima K. The novel BMI-1 inhibitor PTC596 downregulates MCL-1 and induces p53-independent mitochondrial apoptosis in acute myeloid leukemia progenitor cells. *Blood Cancer J* 2017; 7: e527.
- [39] Bolomsky A, Muller J, Stangelberger K, Lejeune M, Duray E, Breid H, Vrancken L, Pfeiffer C, Hübl W, Willheim M, Weetall M, Branstrom A, Zojer N, Caers J and Ludwig H. The anti-mitotic agents PTC-028 and PTC596 display potent activity in pre-clinical models of multiple myeloma but challenge the role of BMI-1 as an essential tumour gene. *Br J Haematol* 2020; 190: 877-890.
- [40] Maeda A, Nishida Y, Weetall M, Cao L, Branstrom A, Ishizawa J, Nii T, Schober WD, Abe Y, Matsue K, Yoshimura M, Kimura S and Kojima K. Targeting of BMI-1 expression by the novel small molecule PTC596 in mantle cell lymphoma. *Oncotarget* 2018; 9: 28547-28560.
- [41] Wu K, Woo SM, Seo SU and Kwon TK. Inhibition of BMI-1 induces apoptosis through downregulation of DUB3-mediated Mcl-1 stabilization. *Int J Mol Sci* 2021; 22: 10107.
- [42] Hinshaw DC and Shevde LA. The tumor microenvironment innately modulates cancer progression. *Cancer Res* 2019; 79: 4557-4566.
- [43] Zhang Y and Zhang Z. The history and advances in cancer immunotherapy: understanding the characteristics of tumor-infiltrating immune cells and their therapeutic implications. *Cell Mol Immunol* 2020; 17: 807-821.
- [44] Pitt JM, Marabelle A, Eggermont A, Soria JC, Kroemer G and Zitvogel L. Targeting the tumor

BMI1-targeted therapy and immunotherapy in cervical cancer

- microenvironment: removing obstruction to anticancer immune responses and immunotherapy. *Ann Oncol* 2016; 27: 1482-1492.
- [45] Lei X, Lei Y, Li JK, Du WX, Li RG, Yang J, Li J, Li F and Tan HB. Immune cells within the tumor microenvironment: biological functions and roles in cancer immunotherapy. *Cancer Lett* 2020; 470: 126-133.
- [46] Tanaka H, Yoshizawa H, Yamaguchi Y, Ito K, Kagamu H, Suzuki E, Gejyo F, Hamada H and Arakawa M. Successful adoptive immunotherapy of murine poorly immunogenic tumor with specific effector cells generated from gene-modified tumor-primed lymph node cells. *J Immunol* 1999; 162: 3574-3582.
- [47] Farhood B, Najafi M and Mortezaee K. CD8(+) cytotoxic T lymphocytes in cancer immunotherapy: a review. *J Cell Physiol* 2019; 234: 8509-8521.
- [48] Mikucki ME, Fisher DT, Matsuzaki J, Skitzki JJ, Gaulin NB, Muhitch JB, Ku AW, Frelinger JG, Odunsi K, Gajewski TF, Luster AD and Evans SS. Non-redundant requirement for CXCR3 signalling during tumoricidal T-cell trafficking across tumour vascular checkpoints. *Nat Commun* 2015; 6: 7458.
- [49] Spranger S, Dai D, Horton B and Gajewski TF. Tumor-residing Batf3 dendritic cells are required for effector T cell trafficking and adoptive T cell therapy. *Cancer Cell* 2017; 31: 711-723, e714.
- [50] Watts TH and DeBenedette MA. T cell co-stimulatory molecules other than CD28. *Curr Opin Immunol* 1999; 11: 286-293.
- [51] Leach DR, Krummel MF and Allison JP. Enhancement of antitumor immunity by CTLA-4 blockade. *Science* 1996; 271: 1734-1736.
- [52] Freeman GJ, Long AJ, Iwai Y, Bourque K, Chernova T, Nishimura H, Fitz LJ, Malenkovich N, Okazaki T, Byrne MC, Horton HF, Fouser L, Carter L, Ling V, Bowman MR, Carreno BM, Collins M, Wood CR and Honjo T. Engagement of the PD-1 immunoinhibitory receptor by a novel B7 family member leads to negative regulation of lymphocyte activation. *J Exp Med* 2000; 192: 1027-1034.
- [53] Das M, Zhu C and Kuchroo VK. Tim-3 and its role in regulating anti-tumor immunity. *Immunol Rev* 2017; 276: 97-111.
- [54] Wang J, Sanmamed MF, Datar I, Su TT, Ji L, Sun J, Chen L, Chen Y, Zhu G, Yin W, Zheng L, Zhou T, Badri T, Yao S, Zhu S, Boto A, Sznol M, Melerio I, Vignali DAA, Schalper K and Chen L. Fibrinogen-like protein 1 is a major immune inhibitory ligand of LAG-3. *Cell* 2019; 176: 334-347, e312.
- [55] Anderson AC, Joller N and Kuchroo VK. Lag-3, Tim-3, and TIGIT: co-inhibitory receptors with specialized functions in immune regulation. *Immunity* 2016; 44: 989-1004.
- [56] Dougall WC, Kurtulus S, Smyth MJ and Anderson AC. TIGIT and CD96: new checkpoint receptor targets for cancer immunotherapy. *Immunol Rev* 2017; 276: 112-120.
- [57] Dong C, Juedes AE, Temann UA, Shresta S, Allison JP, Ruddle NH and Flavell RA. ICOS co-stimulatory receptor is essential for T-cell activation and function. *Nature* 2001; 409: 97-101.
- [58] Kaleeba JA, Offner H, Vandenbark AA, Lublinski A and Weinberg AD. The OX-40 receptor provides a potent co-stimulatory signal capable of inducing encephalitogenicity in myelin-specific CD4+ T cells. *Int Immunol* 1998; 10: 453-461.
- [59] Kwon B, Lee HW and Kwon BS. New insights into the role of 4-1BB in immune responses: beyond CD8+ T cells. *Trends Immunol* 2002; 23: 378-380.
- [60] Knee DA, Hewes B and Brogdon JL. Rationale for anti-GITR cancer immunotherapy. *Eur J Cancer* 2016; 67: 1-10.
- [61] van de Ven K and Borst J. Targeting the T-cell co-stimulatory CD27/CD70 pathway in cancer immunotherapy: rationale and potential. *Immunotherapy* 2015; 7: 655-667.
- [62] Ware CF. Targeting the LIGHT-HVEM pathway. *Adv Exp Med Biol* 2009; 647: 146-155.
- [63] Hollenbaugh D, Grosmaire LS, Kullas CD, Chalupny NJ, Braesch-Andersen S, Noelle RJ, Stamenkovic I, Ledbetter JA and Aruffo A. The human T cell antigen gp39, a member of the TNF gene family, is a ligand for the CD40 receptor: expression of a soluble form of gp39 with B cell co-stimulatory activity. *EMBO J* 1992; 11: 4313-4321.
- [64] Iwai Y, Ishida M, Tanaka Y, Okazaki T, Honjo T and Minato N. Involvement of PD-L1 on tumor cells in the escape from host immune system and tumor immunotherapy by PD-L1 blockade. *Proc Natl Acad Sci U S A* 2002; 99: 12293-12297.
- [65] Pauken KE and Wherry EJ. SnapShot: T cell exhaustion. *Cell* 2015; 163: 1038-1038, e1031.
- [66] Ishida Y, Agata Y, Shibahara K and Honjo T. Induced expression of PD-1, a novel member of the immunoglobulin gene superfamily, upon programmed cell death. *EMBO J* 1992; 11: 3887-3895.
- [67] Butte MJ, Keir ME, Phamduy TB, Sharpe AH and Freeman GJ. Programmed death-1 ligand 1 interacts specifically with the B7-1 costimulatory molecule to inhibit T cell responses. *Immunity* 2007; 27: 111-122.
- [68] Sun C, Mezzadra R and Schumacher TN. Regulation and function of the PD-L1 checkpoint. *Immunity* 2018; 48: 434-452.

BMI1-targeted therapy and immunotherapy in cervical cancer

- [69] Zhang D, Huang J, Wang F, Ding H, Cui Y, Yang Y, Xu J, Luo H, Gao Y, Pan L, Wu Y, Gong Y, Xie L, Liu Z, Qu Y, Zhang L, Liu W, Zhang W, Zhao S, Yi Q, Niu T and Zheng Y. BMI1 regulates multiple myeloma-associated macrophage's promyeloma functions. *Cell Death Dis* 2021; 12: 495.
- [70] Sienerth AR, Scheuermann C, Galmiche A, Rapp UR and Becker M. Polycomb group protein Bmi1 negatively regulates IL-10 expression in activated macrophages. *Immunol Cell Biol* 2011; 89: 812-816.
- [71] Duan Q, Li H, Gao C, Zhao H, Wu S, Wu H, Wang C, Shen Q and Yin T. High glucose promotes pancreatic cancer cells to escape from immune surveillance via AMPK-Bmi1-GATA2-MICA/B pathway. *J Exp Clin Cancer Res* 2019; 38: 192.
- [72] Blackledge NP, Rose NR and Klose RJ. Targeting Polycomb systems to regulate gene expression: modifications to a complex story. *Nat Rev Mol Cell Biol* 2015; 16: 643-649.
- [73] Liu Z, Hu C, Zheng L, Liu J, Li K, Li X, Wang Y, Mu W, Chen T, Shi A, Qiu B, Zhang X, Zhang Z and Xu Y. BMI1 promotes cholangiocarcinoma progression and correlates with antitumor immunity in an exosome-dependent manner. *Cell Mol Life Sci* 2022; 79: 469.
- [74] Heffner M and Fearon DT. Loss of T cell receptor-induced Bmi-1 in the KLRG1(+) senescent CD8(+) T lymphocyte. *Proc Natl Acad Sci U S A* 2007; 104: 13414-13419.
- [75] Wang S, Xu N, Wang J, Chen Y, Li W, Chen H, Shen C, Xu C, Wei X, Lu D, Qiu N, Zheng S, Wei Q and Xu X. BMI1-induced CD127+KLRG1+ memory T cells enhance the efficacy of liver cancer immunotherapy. *Cancer Lett* 2023; 571: 216336.
- [76] Lu ZH, Tu GJ, Fu SL, Shang K, Peng SJ, Chen L and Gu XJ. BMI1 induces ubiquitination and protein degradation of Nod-like receptor family CARD domain containing 5 and suppresses human leukocyte antigen class I expression to induce immune escape in non-small cell lung cancer. *Kaohsiung J Med Sci* 2022; 38: 1190-1202.
- [77] Kobayashi A, Weinberg V, Darragh T and Smith-McCune K. Evolving immunosuppressive microenvironment during human cervical carcinogenesis. *Mucosal Immunol* 2008; 1: 412-420.
- [78] Piersma SJ. Immunosuppressive tumor microenvironment in cervical cancer patients. *Cancer Microenviron* 2011; 4: 361-375.
- [79] Wendel Naumann R and Leath CA 3rd. Advances in immunotherapy for cervical cancer. *Curr Opin Oncol* 2020; 32: 481-487.
- [80] Zhou C, Tuong ZK and Frazer IH. Papillomavirus immune evasion strategies target the infected cell and the local immune system. *Front Oncol* 2019; 9: 682.
- [81] Yuan Y, Cai X, Shen F and Ma F. HPV post-infection microenvironment and cervical cancer. *Cancer Lett* 2021; 497: 243-254.
- [82] Villarino AV, Kanno Y and O'Shea JJ. Mechanisms and consequences of Jak-STAT signaling in the immune system. *Nat Immunol* 2017; 18: 374-384.
- [83] Mimura K, Kua LF, Shiraishi K, Kee Siang L, Shabbir A, Komachi M, Suzuki Y, Nakano T, Yong WP, So J and Kono K. Inhibition of mitogen-activated protein kinase pathway can induce upregulation of human leukocyte antigen class I without PD-L1-upregulation in contrast to interferon- γ treatment. *Cancer Sci* 2014; 105: 1236-1244.
- [84] Lee SK, Seo SH, Kim BS, Kim CD, Lee JH, Kang JS, Maeng PJ and Lim JS. IFN- γ regulates the expression of B7-H1 in dermal fibroblast cells. *J Dermatol Sci* 2005; 40: 95-103.
- [85] Chen G, Huang AC, Zhang W, Zhang G, Wu M, Xu W, Yu Z, Yang J, Wang B, Sun H, Xia H, Man Q, Zhong W, Antelo LF, Wu B, Xiong X, Liu X, Guan L, Li T, Liu S, Yang R, Lu Y, Dong L, McGettigan S, Somasundaram R, Radhakrishnan R, Mills G, Lu Y, Kim J, Chen YH, Dong H, Zhao Y, Karakousis GC, Mitchell TC, Schuchter LM, Herlyn M, Wherry EJ, Xu X and Guo W. Exosomal PD-L1 contributes to immunosuppression and is associated with anti-PD-1 response. *Nature* 2018; 560: 382-386.
- [86] Luo N, Formisano L, Gonzalez-Ericsson PI, Sanchez V, Dean PT, Opalenik SR, Sanders ME, Cook RS, Arteaga CL, Johnson DB and Balko JM. Melanoma response to anti-PD-L1 immunotherapy requires JAK1 signaling, but not JAK2. *Oncoimmunology* 2018; 7: e1438106.
- [87] Doi T, Ishikawa T, Okayama T, Oka K, Mizushima K, Yasuda T, Sakamoto N, Katada K, Kamada K, Uchiyama K, Handa O, Takagi T, Naito Y and Itoh Y. The JAK/STAT pathway is involved in the upregulation of PD-L1 expression in pancreatic cancer cell lines. *Oncol Rep* 2017; 37: 1545-1554.
- [88] Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, Gonzalez R, Robert C, Schadendorf D, Hassel JC, Akerley W, van den Eertwegh AJ, Lutzky J, Lorigan P, Vaubel JM, Linette GP, Hogg D, Ottensmeier CH, Lebbé C, Peschel C, Quirt I, Clark JI, Wolchok JD, Weber JS, Tian J, Yellin MJ, Nichol GM, Hoos A and Urba WJ. Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med* 2010; 363: 711-723.
- [89] Gandhi L, Rodríguez-Abreu D, Gadgeel S, Esteban E, Felip E, De Angelis F, Domine M, Clingan P, Hochmair MJ, Powell SF, Cheng SY, Bischoff HG, Peled N, Grossi F, Jennens RR, Reck M, Hui

BMI1-targeted therapy and immunotherapy in cervical cancer

- R, Garon EB, Boyer M, Rubio-Viqueira B, Novello S, Kurata T, Gray JE, Vida J, Wei Z, Yang J, Raftopoulos H, Pietanza MC and Garassino MC; KEYNOTE-189 Investigators. Pembrolizumab plus chemotherapy in metastatic non-small-cell lung cancer. *N Engl J Med* 2018; 378: 2078-2092.
- [90] Motzer RJ, Escudier B, McDermott DF, George S, Hammers HJ, Srinivas S, Tsykodi SS, Sosman JA, Procopio G, Plimack ER, Castellano D, Choueiri TK, Gurney H, Donskov F, Bono P, Wagstaff J, Gauler TC, Ueda T, Tomita Y, Schutz FA, Kollmannsberger C, Larkin J, Ravaud A, Simon JS, Xu LA, Waxman IM and Sharma P; CheckMate 025 Investigators. Nivolumab versus everolimus in advanced renal-cell carcinoma. *N Engl J Med* 2015; 373: 1803-1813.
- [91] Schmid P, Adams S, Rugo HS, Schneeweiss A, Barrios CH, Iwata H, Diéras V, Hegg R, Im SA, Shaw Wright G, Henschel V, Molinero L, Chui SY, Funke R, Husain A, Winer EP, Loi S and Emens LA; IMpassion130 Trial Investigators. Atezolizumab and nab-paclitaxel in advanced triple-negative breast cancer. *N Engl J Med* 2018; 379: 2108-2121.
- [92] Orange M, Reuter U and Hobohm U. Coley's lessons remembered: augmenting mistletoe therapy. *Integr Cancer Ther* 2016; 15: 502-511.
- [93] Mastelic-Gavillet B, Balint K, Boudousquie C, Gannon PO and Kandalaf LE. Personalized dendritic cell vaccines-recent breakthroughs and encouraging clinical results. *Front Immunol* 2019; 10: 766.
- [94] Berraondo P, Sanmamed MF, Ochoa MC, Etxeberria I, Aznar MA, Pérez-Gracia JL, Rodríguez-Ruiz ME, Ponz-Sarvisé M, Castañón E and Melero I. Cytokines in clinical cancer immunotherapy. *Br J Cancer* 2019; 120: 6-15.
- [95] Rapoport AP, Stadtmauer EA, Binder-Scholl GK, Golubeva O, Vogl DT, Lacey SF, Badros AZ, Garfall A, Weiss B, Finklestein J, Kulikovskaya I, Sinha SK, Kronsberg S, Gupta M, Bond S, Melchiori L, Brewer JE, Bennett AD, Gerry AB, Pumphrey NJ, Williams D, Tayton-Martin HK, Ribeiro L, Holdich T, Yanovich S, Hardy N, Yared J, Kerr N, Philip S, Westphal S, Siegel DL, Levine BL, Jakobsen BK, Kalos M and June CH. NY-ESO-1-specific TCR-engineered T cells mediate sustained antigen-specific antitumor effects in myeloma. *Nat Med* 2015; 21: 914-921.
- [96] Hellmann MD, Ciuleanu TE, Pluzanski A, Lee JS, Otterson GA, Audigier-Valette C, Minenza E, Linardou H, Burgers S, Salman P, Borghaei H, Ramalingam SS, Brahmer J, Reck M, O'Byrne KJ, Geese WJ, Green G, Chang H, Szustakowski J, Bhagavatheeswaran P, Healey D, Fu Y, Nathan F and Paz-Ares L. Nivolumab plus ipilimumab in lung cancer with a high tumor mutational burden. *N Engl J Med* 2018; 378: 2093-2104.
- [97] Balkwill FR, Capasso M and Hagemann T. The tumor microenvironment at a glance. *J Cell Sci* 2012; 125: 5591-5596.
- [98] Galon J, Costes A, Sanchez-Cabo F, Kirilovsky A, Mlecnik B, Lagorce-Pagès C, Tosolini M, Camus M, Berger A, Wind P, Zinzindohoué F, Bruneval P, Cugnenc PH, Trajanoski Z, Fridman WH and Pagès F. Type, density, and location of immune cells within human colorectal tumors predict clinical outcome. *Science* 2006; 313: 1960-1964.
- [99] Clambey ET, Davenport B, Kappler JW, Marrack P and Homann D. Molecules in medicine mini review: the $\alpha\beta$ T cell receptor. *J Mol Med (Berl)* 2014; 92: 735-741.
- [100] Chen L and Flies DB. Molecular mechanisms of T cell co-stimulation and co-inhibition. *Nat Rev Immunol* 2013; 13: 227-242.
- [101] Zhang J, Fang W, Qin T, Yang Y, Hong S, Liang W, Ma Y, Zhao H, Huang Y, Xue C, Huang P, Hu Z, Zhao Y and Zhang L. Co-expression of PD-1 and PD-L1 predicts poor outcome in nasopharyngeal carcinoma. *Med Oncol* 2015; 32: 86.
- [102] Ren D, Hua Y, Yu B, Ye X, He Z, Li C, Wang J, Mo Y, Wei X, Chen Y, Zhou Y, Liao Q, Wang H, Xiang B, Zhou M, Li X, Li G, Li Y, Zeng Z and Xiong W. Predictive biomarkers and mechanisms underlying resistance to PD1/PD-L1 blockade cancer immunotherapy. *Mol Cancer* 2020; 19: 19.
- [103] Kordbacheh T, Honeychurch J, Blackhall F, Faivre-Finn C and Illidge T. Radiotherapy and anti-PD-1/PD-L1 combinations in lung cancer: building better translational research platforms. *Ann Oncol* 2018; 29: 301-310.
- [104] Naumann RW, Hollebecque A, Meyer T, Devlin MJ, Oaknin A, Kerger J, López-Picazo JM, Machiels JP, Delord JP, Evans TRJ, Boni V, Calvo E, Topalian SL, Chen T, Soumaoro I, Li B, Gu J, Zwirter R and Moore KN. Safety and efficacy of nivolumab monotherapy in recurrent or metastatic cervical, vaginal, or vulvar carcinoma: results from the phase I/II CheckMate 358 trial. *J Clin Oncol* 2019; 37: 2825-2834.
- [105] Kagabu M, Nagasawa T, Sato C, Fukagawa Y, Kawamura H, Tomabechi H, Takemoto S, Shoji T and Baba T. Immunotherapy for uterine cervical cancer using checkpoint inhibitors: future directions. *Int J Mol Sci* 2020; 21: 2335.
- [106] Colombo N, Dubot C, Lorusso D, Caceres MV, Hasegawa K, Shapira-Frommer R, Tewari KS, Salman P, Hoyos Usta E, Yañez E, Gümüş M, Olivera Hurtado de Mendoza M, Samouëlian V, Castonguay V, Arkhipov A, Toker S, Li K, Keefe SM and Monk BJ; KEYNOTE-826 Investigators. Pembrolizumab for persistent, recurrent, or

BMI1-targeted therapy and immunotherapy in cervical cancer

- metastatic cervical cancer. *N Engl J Med* 2021; 385: 1856-1867.
- [107] Ojima I. Guided molecular missiles for tumor-targeting chemotherapy--case studies using the second-generation taxoids as warheads. *Acc Chem Res* 2008; 41: 108-119.
- [108] Galluzzi L, Humeau J, Buqué A, Zitvogel L and Kroemer G. Immunostimulation with chemotherapy in the era of immune checkpoint inhibitors. *Nat Rev Clin Oncol* 2020; 17: 725-741.