Original Article

Exercise pre-conditioning prevents vascular toxicity caused by infusion of 5-fluorouracil in male rats

Stephen T Hammond^{1,2,3,4*}, Dryden R Baumfalk^{1,2,5*}, Andrew G Horn^{1,6}, Britton C Scheuermann^{1,2}, Olivia N Kunkel^{1,2}, Carl J Ade^{1,2,7}, Bradley J Behnke^{1,2}

¹Department of Kinesiology, Kansas State University, Manhattan, KS, USA; ²Johnson Cancer Research Center, Kansas State University, Manhattan, KS, USA; ³Department of Medicine, Medical College of Wisconsin, Milwaukee, WI, USA; ⁴Cardiovascular Research Center, Medical College of Wisconsin, Milwaukee, WI, USA; ⁵Department of Orthopaedic Surgery, Duke University, Durham, NC, USA; ⁶Cardiovascular Research and Training Institute, University of Utah, Salt Lake, UT, USA; ⁷Physicians Associates Studies, Kansas State University, Manhattan, KS, USA. *Equal contributors.

Received June 12, 2025; Accepted September 10, 2025; Epub September 25, 2025; Published September 30, 2025

Abstract: 5-fluorouracil (5-FU) is one of the most common chemotherapies used in cancer treatment yet is often associated with acute cardiotoxicity (e.g., angina, vasospasm). To date, countermeasures to prevent 5-FU cardiotoxicity are lacking. Therefore, we tested the hypothesis that short-term, moderate-intensity exercise completed before 5-FU administration would prevent 5-FU-induced alterations in vascular and cardiac function. Male Sprague-Dawley rats were randomized to sedentary control (SEDCON, n = 9), sedentary 5-FU (SED5FU, n = 10), exercise control (EXCON, n = 8) or exercise 5-FU (EX5FU, n = 8) groups. Rats remained sedentary or completed 4-days of treadmill running (20-25 min, 20 m/min, 5% grade) with the final bout ending ~90-min before treatment with a clinically relevant dose of 5-FU (50 mg/kg bolus + 265 mg/kg 2-hr infusion) or volume matched saline. Echocardiographic indices of left ventricular function and Doppler measurements of aortic pulse wave velocity (PWV) were completed at baseline (BL) and after the 2-hr (2-hr) infusion. 5-FU did not induce changes in left ventricular function. PWV increased from BL to 2-hr in SED5FU (BL: 396 ± 39 cm/s; 2-hr: 452 ± 54 cm/s; P = 0.002), but not SEDCON (BL: 417 ± 55 cm/s; 2-hr: 392 ± 64 cm/s; P = 0.35), EXCON (BL: 408 ± 35 cm/s; 2-hr: 410 ± 46 cm/s; P > 0.99), or EX5FU (BL: 398 ± 35 cm/s; 2-hr: 410 ± 46 cm/s; P > 0.99), or EX5FU (BL: 408 ± 35 cm/s; 2-hr: 410 ± 46 cm/s; P > 0.99), or EX5FU (BL: 408 ± 35 cm/s; 2-hr: 410 ± 46 cm/s; P > 0.99), or EX5FU (BL: 408 ± 35 cm/s; 2-hr: 410 ± 46 cm/s; P > 0.99), or EX5FU (BL: 408 ± 35 cm/s; 2-hr: 410 ± 46 cm/s; P > 0.99), or EX5FU (BL: 408 ± 35 cm/s; 2-hr: 410 ± 46 cm/s; P > 0.99), or EX5FU (BL: 408 ± 35 cm/s; 410 ± 46 cm/s; P > 0.99), or EX5FU (BL: 408 ± 35 cm/s; 410 ± 46 cm/s; P > 0.99), or EX5FU (BL: 408 ± 35 cm/s; 410 ± 46 cm/s; P > 0.99), or EX5FU (BL: 408 ± 35 cm/s; 410 ± 46 cm/s; P > 0.99), or EX5FU (BL: 408 ± 35 cm/s; 410 ± 46 cm/s; P > 0.99), or EX5FU (BL: 408 ± 35 cm/s; 410 ± 46 cm/s; 410 ± 466 cm/ 13 cm/s; 2-hr: $417 \pm 23 \text{ cm/s}$; P = 0.67). Additionally, PWV at the 2-hr time point was significantly higher in SED5FU compared to SEDCON (P = 0.002). These findings suggest that moderate-intensity exercise preconditioning may protect against 5-FU-induced alterations in arterial PWV—potentially mitigating early signs of cardiotoxicity. Future studies are warranted to identify the mechanisms of 5-FU-induced cardiotoxicity and the feasibility and efficacy of pre-treatment exercise regimens in human patients.

Keywords: Cardiotoxicity, cardio-oncology, exercise oncology, chemotherapy, pulse wave velocity, arterial stiffness

Introduction

5-fluorouracil (5-FU) chemotherapy has remained a cornerstone of numerous gastrointestinal, breast, and head/neck cancer treatment regimens since its inception in 1957 [1]. While this longevity speaks to its effectiveness as an anticancer agent, 5-FU has gained notoriety for its unfortunate association with treatment-induced cardiotoxicity in up to 19% of treated patients [2-7]. Reported side effects often arise during or within 72 hours of completing treatment [5], and include angina (i.e., chest pain), acute coronary syndromes at rest

or with exertion, ECG alterations indicative of myocardial ischemia, and in severe instances, heart failure and sudden cardiac death [2, 4, 5, 8-11]

Evidence of coronary spasm [12, 13], peripheral vascular constriction/dysfunction [14-16], and increased arterial stiffness [17] in 5-FU treated patients have led to the predominant theory that 5-FU cardiotoxicity primarily arises via a direct effect on the vasculature [5]. However, data from preclinical studies suggest both myocardial [18-21] and vascular [22-28] etiology—with findings potentially influenced by

how closely experimental protocols replicate clinical infusion regimens and incorporate clinically relevant measures of cardiac and vascular injury. Preclinical studies have historically administered 5-FU via intraperitoneal or bolus intravenous (IV) injections alone [19-21, 29]. This differs from clinical practice where 5-FU is often delivered via IV 5-FU bolus (400 mg/m²) followed by a continuous IV infusion of 5-FU (2,400 mg/m²) for 46-72 hours [30-32]. Considering that rates of 5-FU cardiotoxicity are higher following continuous infusion compared to bolus alone [33, 34], preclinical models which employ continuous infusion protocols similar to those used clinically may elucidate whether 5-FU primarily mediates its cardiotoxic effects via the vasculature, the heart, or both.

Given its importance as a first-line cancer treatment, strategies to alleviate or prevent 5-FU cardiotoxicity have been of recent interest. Pharmacological therapy with nitrates or calcium channel blockers have yielded mixed findings [16, 35-37], and these treatments are typically only administered after a patient experiences adverse treatment-related side effects. As such, therapies administered prior to or during treatment with the potential to prevent cardiotoxicity are greatly needed. One such strategy is the implementation of an exercise preconditioning regimen, as aerobic exercise has established cardioprotective effects on both the heart and vasculature [38]. Yet, while long-term exercise preconditioning (7-12 wks.) can effectively prevent chemotherapy-induced cardiotoxicity in rodents [39-42], such programs may not be clinically feasible in the timeframe between initial diagnosis and treatment onset-especially if the patient is recovering from surgical procedures. Conversely, shortterm exercise preconditioning programs can also offer cardioprotective benefits [43, 44] and could conceivably be implemented in the week preceding treatment. Studies in rodents have demonstrated 1-5 days of exercise preconditioning is sufficient to prevent cardiotoxicity caused by anthracycline chemotherapy [45-48]; however, the efficacy of exercise preconditioning as a means of preventing 5-FU cardiotoxicity remains untested. To address this gap, we developed a clinically relevant rat model of continuous 5-FU infusion and tested the hypothesis that acute aerobic exercise preconditioning would prevent signs of cardiac and vascular toxicity. We specifically evaluated the

impact of 5-FU on left ventricular function and arterial stiffness to determine whether exercise may attenuate early manifestations of 5-FU-induced cardiotoxicity. Given recent evidence that cancer may independently contribute to cardiovascular maladaptation [49-52], the present studies were conducted in tumor naïve rats to eliminate potential confounding effects of cancer and determine the direct impact of 5-FU on cardiovascular well-being.

Materials and methods

Animals

All procedures performed herein were approved by the Kansas State University Institutional Animal Care and Use Committee and conformed to the National Institutes of Health Guide for the Care and Use of Laboratory Animals. Male Sprague Dawley rats (n = 38, 6-8 mo. old, $\sim\!670$ g; Charles River; Wilmington, MA) were used for this study. All rodents were housed in a temperature-controlled facility (23 \pm 2°C) on a 12:12-h light-dark cycle with standard rat chow and water provided *ab libitum*.

Study design

The week before the experimental protocol, all rats were habituated to treadmill exercise on a custom-built motor-driven rodent treadmill (< 5 min/day at 15 m/min, 0% incline for three days). Each rat was then randomized to either exercise or sedentary groups and to receive treatment with 5-FU or volume-matched saline. This resulted in a total of four experimental groups: sedentary + saline control (SEDCON, n = 10), sedentary + 5-FU (SED5FU, n = 10), exercise + saline control (EXCON, n = 9), and exercise + 5-FU (EX5FU, n = 9). Three days before 5-FU or saline treatment, exercise animals (EXCON and EX5FU) began a 4-day acute exercise preconditioning program (Figure 1). Briefly, rats ran on a motorized treadmill at 20 m/min (5% grade) for 20-25 min/day, with the final exercise session commencing ~90-min before 5-FU or saline treatment (see 5-FU or Saline Administration). Based on the prior work of others, we estimate this exercise intensity to be ~65% VO₂max [53].

Surgical procedures

Approximately 30-min following completion of the final exercise bout, rats were first anesthe-

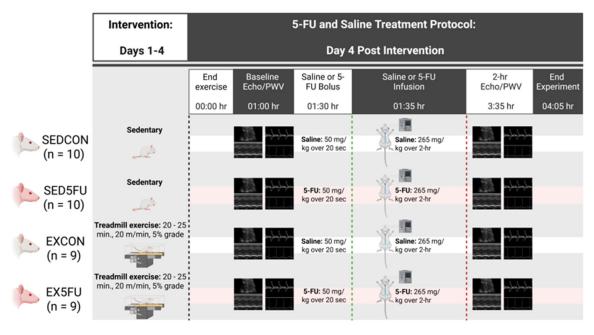


Figure 1. Study design. A timeline of events for the acute exercise preconditioning and 5-FU chemotherapy infusion protocol. Animals were randomized to sedentary or acute exercise preconditioning groups. Exercised animals completed four, once daily bouts of moderate intensity exercise on a motorized treadmill (20 m/min, 5% grade). Exercise preconditioning commenced three days prior to planned 5-FU/saline infusion with the fourth and final bout ending ~90 minutes prior to the start of the treatment. *In vivo* assessment of left ventricular function and aortic pulse wave velocity (PWV) were conducted immediately prior to and following completion of the 5-FU/saline infusion to assess the effect of 5-FU on cardiovascular function. Figure designed using Biorender.com.

tized with a 5% isoflurane-02 mixture (isoflurane vaporizer; Harvard Apparatus; Cambridge, MA) and subsequently maintained on 2.5% isofluorane-O2 on a heated surgical station (Rodent Surgical Monitor, Indus Instruments; Houston, TX) at 37 ± 1°C. An incision was made at the left ventral neck to expose the left jugular vein for catheterization (PE-50 Intra-Medic polyethylene tubing, Becton Dickson, Sparks, MD) to administer 5-FU or saline. A second catheter (PE-10 connected to PE-50) was placed in the caudal artery for continuous blood pressure monitoring. Following instrumentation, rats were transitioned to pentobarbital sodium anesthesia (20 mg/kg body wt) administered via the caudal artery (total volume per injection ~0.2 ml) while concentrations of isoflurane were decreased and subsequently discontinued over ~30 min. The level of anesthesia was regularly monitored via toe pinch and palpebral reflex, with pentobarbital anesthesia supplemented (~16 mg/kg/hr) as necessary for the remainder of the experiment. After completing the 2-hr infusion and associated cardiovascular measurements (see

below), all animals were euthanized via cardiac excision under deep plane anesthesia. A graphical depiction of the experimental timeline is presented in **Figure 1**.

5-FU and saline administration

To align our study with common clinical drug delivery methods, we designed a translational rat model of 5-FU-induced cardiotoxicity using allometric scaling [54]. Following the completion of surgical procedures and collection of baseline cardiovascular measurements (see below), a bolus dose of 5-FU (50 mg/kg) or volume-matched saline was delivered over ~20 seconds via a jugular vein catheter using a 10 mL syringe. The jugular vein catheter was then attached to a programmable infusion syringe pump (Pump 11 Elite; Harvard Apparatus; Holliston, MA) to initiate a 2-hour continuous infusion of 5-FU (265 mg/kg) or volumematched saline. These 5-FU doses were determined using the human bolus (400 mg/m²) and continuous infusion (2400 mg/m²) doses commonly used in 5-FU based regimens (e.g., FOLFOX, FOLFIRI) [30, 32].

Hemodynamic variables

Heart rate (HR) and mean arterial blood pressure (MAP) were continuously monitored over the duration of the protocol using a commercially available blood pressure monitor (Digi-Med BPA; Micro-Med; Louisville, KY) with analog output to a laboratory computer via a multifunction data acquisition device (NI USB 6211, National Instruments, Austin, TX). Data were sampled at 1,000 Hz and recorded for offline analysis. Measurements of HR and MAP were averaged over 8-10 minutes immediately before administration of the 5-FU/saline treatment (baseline, BL) and at the end of the 2-hour infusion (2-hr).

Echocardiographic evaluation of left ventricular function

Transthoracic echocardiography measures of left ventricular function were performed over three continuous cardiac cycles at BL and following the 2-hr continuous infusion (Logiq e; GE Medical Systems; Milwaukee, WI) as previously demonstrated by our group [49, 55-58]. Following removal of hair from the chest (Nair, Johnson & Johnson, New Brunswick, NJ), twodimensional guided M-mode images were collected at the level of the mitral leaflets in the parasternal short axis view using a 22-MHz linear transducer (GE L10-22-RS, GE Medical Systems). Images were stored on an offline storage device for future measurement of left ventricular dimensions and posterior wall thickness, which were subsequently used to estimate left ventricular end-diastolic (EDV) and end-systolic (ESV) volumes using the Teichhholz formula and to calculate fractional shortening (FS), stroke volume (SV), and cardiac output (Q) [59].

Doppler pulse wave velocity

Measurements of aortic pulse wave velocity (PWV)—an established marker of arterial stiffness [60]—were made at BL and following the 2-hour continuous infusion using a commercially available Doppler flow velocity system (Indus Instruments; Houston, TX). Rats were positioned supine, with external needle electrodes placed on each paw for ECG collection. Two Doppler probes connected to a pulsed Doppler system transceiver were used to obtain simultaneous Doppler spectrograms from the

descending and abdominal aorta. One probe was placed along the abdominal midline to capture abdominal aorta flow velocity and held in place by a micro-positioner. The second probe was placed to the right of the sternum near the base of the upper left limb to capture descending aorta velocity. Images were captured simultaneously, and time aligned with ECG for offline analyses using manufacturer software. The distance between probes (mm) was measured immediately following the obtainment of the image. PWV was calculated as follows: PWV = distance between probe tips (mm)/(time from R wave to the abdominal aorta - time from R wave to aortic arch) (ms) and reported in cm/s.

Data analysis

All statistical analyses were completed using a commercially available software package (GraphPad Prism 9.5.0; San Diego, Ca, USA). Echocardiographic and hemodynamic variables, as well as aortic pulse wave velocity, were assessed via two-way repeated measures analyses (mixed-model) with main effects for group (SEDCON, SED5FU, EXCON, EX5FU) and time (BL, 2-hours). Multiple comparisons were completed using Šídák's multiple comparisons test when significant interactions or main effects were present. Group differences in body mass were assessed using one-way ANOVA. All data are presented as mean ± standard deviation unless otherwise stated. Statistical significance was set at P < 0.05.

Results

A total of 38 rats were used in this study. Three rats died during the surgical procedures, and thus findings from a total sample of 35 rats (SEDCON: n = 9; SED5FU: n = 10, EXCON: n = 8; EX5FU: n = 8) are reported herein. There were no differences in body mass between the groups (SEDCON: 642 ± 41 g; SED5FU: 637 ± 87 g; EXCON: 728 ± 119 g; EX5FU: 685 ± 87 g; P = 0.13).

Hemodynamic variables

Mean values for hemodynamic variables are presented in **Table 1**. There was no significant group × time interaction for MAP (P = 0.99), HR (P = 0.78), or Q (P = 0.44). HR did not differ between the groups (P = 0.70), nor did it change significantly from BL to 2 hours (P > 0.99).

Acute exercise prevents 5-FU cardiotoxicity

Table 1. Mean data for echocardiographic and hemodynamic variables

	Sedentary Control (SEDCON) n = 9		Sedentary 5-FU (SED5FU) n = 10		Exercise Control (EXCON) n = 8		Exercise 5-FU (EX5FU) n = 8	
	BL	2-hr	BL	2-hr	BL	2-hr	BL	2-hr
ESV (mL)	0.07 ± 0.04	0.14 ± 0.08**	0.09 ± 0.04	0.16 ± 0.10**	0.10 ± 0.02	0.13 ± 0.05	0.10 ± 0.06	0.18 ± 0.05**
EDV (mL)	0.82 ± 0.12	0.88 ± 0.21	0.82 ± 0.20	0.96 ± 0.26	1.02 ± 0.15	0.97 ± 0.15	0.92 ± 0.19	1.03 ± 0.13
SV (mL/beat)	0.75 ± 0.11	0.75 ± 0.16	0.73 ± 0.17	0.80 ± 0.17	0.91 ± 0.13	0.83 ± 0.11	0.82 ± 0.15	0.84 ± 0.08
FS (%)	59.8 ± 8.5	50.8 ± 9.7†	55.5 ± 6.1	50.0 ± 8.1*	55.8 ± 2.8	50.9 ± 3.5	57.3 ± 8.9	45.9 ± 2.8†
MAP (mmHg)	78 ± 12	86 ± 21	81 ± 9	87 ± 10	76 ± 15	85 ± 24	76 ± 10	84 ± 23
HR (BPM)	323 ± 26	326 ± 28	321 ± 31	314 ± 29	316 ± 28	308 ± 33	319 ± 57	329 ± 14
Q (mL/min)	243.0 ± 49.0	245.6 ± 68.3	234.9 ± 61.0	250.7 ± 52.7	286.3 ± 33.3	256.7 ± 46.2	264.6 ± 70.9	274.6 ± 25.3

Abbreviations: BL, Baseline; ESV, End systolic volume; EDV, End diastolic volume; SV, Stroke volume; FS, Fractional shortening; MAP, Mean arterial pressure; HR, Heart rate; Q, Cardiac output. Data are presented as mean ± standard deviation. *P < 0.05 baseline to 2-hr; **P < 0.01 baseline to 2-hr; †P < 0.001 baseline to 2-hr.

Similarly, Q was not different between the groups (P = 0.36) and did not change over the course of the infusion (P = 0.98). MAP did not differ between groups (P = 0.90) but increased significantly over the course of the 2-hour infusion (P = 0.01). However, multiple comparisons did not reveal a significant increase in MAP for any of the individual groups (SEDCON: P = 0.17; SED5FU: P = 0.26; EXCON: P = 0.19; EX5FU: P = 0.19).

Echocardiographic measurements

Echocardiographic measurements were conducted to assess how 5-FU may directly impact myocardial function and the potential cardioprotective role of exercise preconditioning. Group means for individual echocardiographic variables are reported in **Table 1**. There were no significant group × time interactions for any of the left ventricular volumes or parameters (all P > 0.05) (**Figure 2A-D**). Similarly, no significant main effects for group were detected (all P > 0.05). There was no significant main effect for time on EDV (P = 0.06) (Figure 2B) or SV (P = 0.96) (Figure 2C); however, ESV (P < 0.001)(Figure 2A) and FS (P < 0.001) (Figure 2D) did change significantly over the course of the 2-hour infusion. Post hoc analyses revealed a significant increase in ESV for SEDCON (P = 0.003), SED5FU (P = 0.002), and EX5FU (P = 0.002), but not for EXCON (P = 0.15), over the course of the 2-hour infusion (Figure 2A). Similarly, FS was reduced from BL to 2 hours in SEDCON (P < 0.001), SED5FU (P = 0.01), and EX5FU (P < 0.001), but did not reach statistical significance in EXCON (P = 0.05) (Figure 2D). However, it is important to note that the changes in FS remained within the normal physiological range for healthy rats [55-58, 61], and the absence of a treatment-dependent pattern suggests these effects may reflect time-dependent alterations related to prolonged anesthesia rather than true myocardial depression.

Aortic pulse wave velocity (PWV)

PWV was assessed to gain insight into the potential effects 5-FU may have on the vasculature. A significant group \times time interaction was observed for PWV (P = 0.004). There were no significant differences in PWV between any of the groups at BL (all P > 0.05), however, SED5FU had a significantly greater PWV than SEDCON at the 2-hr time point (SED5FU: 452 \pm 54 cm/s;

SEDCON: 392 ± 64 cm/s; P = 0.03), indicating acute vascular dysfunction with 5-FU. No other group comparisons at the 2-hr timepoint reached statistical significance (all > 0.05). Additionally, PWV increased significantly from BL to 2-hr in SED5FU (BL: 396 ± 39 cm/s; 2-hr: 452 ± 54 cm/s; P < 0.001) but not in SEDCON (BL: 417 ± 55 cm/s; $2-hr: 392 \pm 64$ cm/s; P =0.10). Rats that completed four days of moderate-intensity treadmill exercise before 5-FU or saline infusion showed no significant change in PWV (EXCON: BL: 408 ± 35 cm/s; 2-hr: 410 ± 46 cm/s; P = 0.86; EX5FU: BL: 398 ± 13 cm/s; 2-hr: 417 ± 23 cm/s; P = 0.24) over the same time period suggesting a protective effect of exercise preconditioning on 5-FU-induced increases in PWV (Figure 3A). When PWV was compared as a percent change from BL, SED-5FU had a significantly greater change than SEDCON (SED5FU: 14.5 ± 10.9%; SEDCON: $-5.3 \pm 15.7\%$; P = 0.002) while no difference was observed between SED5FU and EXCON $(0.72 \pm 10.9\%; P = 0.06)$ or EX5FU $(4.87 \pm 10.9\%; P = 0.06)$ 6.7%; P = 0.27) (**Figure 3B**). Taken together, these findings support the hypothesis that pre-treatment exercise protects against early 5-FU-induced vascular toxicity.

Discussion

The primary objective of this study was to determine whether acute exercise preconditioning is sufficient to prevent signs of cardiotoxicity in rats exposed to a clinically relevant 5-FU chemotherapy dosing regimen (i.e., bolus + infusion). In agreement with others, our findings suggest that acute 5-FU administration predominantly results in alterations in vascular function. While 5-FU caused no apparent changes in echocardiographic indices of left ventricular function, aortic PWV increased over the 2-hr infusion in SED5FU while remaining unchanged in all other groups. This increase resulted in significantly higher PWV at the 2-hr time point in SED5FU compared to SEDCON despite the absence of differences in key hemodynamic variables that can influence PWV (e.g., MAP, SV, Q, etc.). Notably, this increase in PWV was not observed in rats treated with 5-FU following 4 consecutive days of moderateintensity treadmill running. We interpret these findings to suggest that vascular complications in rodents can arise following a single clinically relevant cycle of 5-FU (bolus + continuous infu-

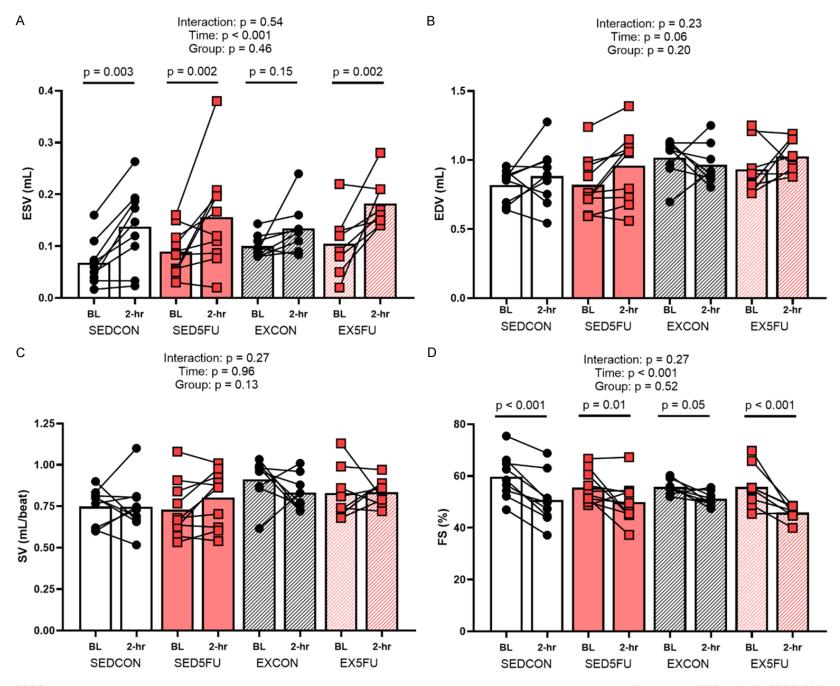


Figure 2. Effect of 5-FU infusion on left ventricular variables. A. A significant main effect for time was present for ESV. There was a significant increase in ESV from BL to 2-hr in all groups aside from EXCON. B. There was no significant change over time or difference between the groups for EDV over the course of the infusion. C. There was no significant change over time or difference between the groups for SV over the course of the infusion. D. There was a significant effect of time on FS. FS was significantly reduced in all groups aside from EXCON over the course of the 2-hr infusion. SEDCON n = 9, SED5FU n = 10, EXCON n = 8, EX5FU n = 8. Data analyzed using mixed effects model. Post hoc comparisons were completed using Šídák's multiple comparisons test when significant interactions or main effects were present. ESV n = 80 end systolic volume, EDV n = 81 end diastolic volume, SV n = 82 stroke volume, FS n = 83 fractional shortening.

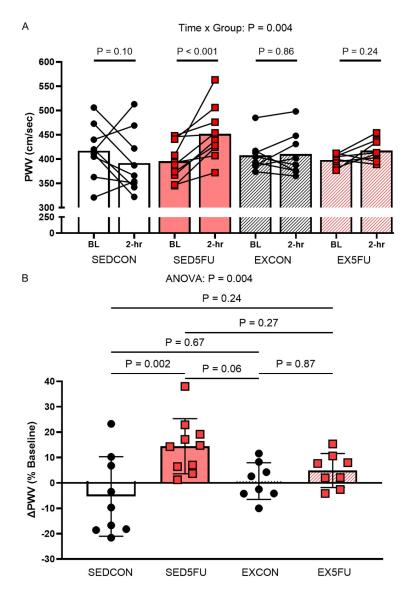


Figure 3. Effect of 5-FU infusion on aortic pulse wave velocity. A. PWV significantly increases from baseline (BL) to 2 hours (2-hr) in SED5FU but not in SEDCON, EXCON, or EX5FU, suggesting acute exercise may dampen the effect of 5FU on the vasculature. B. SED5FU had a significantly greater change in PWV from BL to the 2-hour time point than SEDCON. No other comparisons achieved statistical significance. Data were analyzed using a mixed effects model (post hoc comparisons made using Šídák's multiple comparisons test) (A) and one-way ANOVA (post hoc comparisons made using Tukey's multiple comparisons test) (B). SEDCON n = 9, SED5FU n = 10, EXCON n = 8, EX5FU = 8. PWV = pulse wave velocity.

sion) and that short-term aerobic preconditioning may be sufficient to prevent these adverse effects.

Despite continued advances in cancer care, 5-FU chemotherapy has remained a central component of numerous anticancer treatment regimens for over 60 years [62, 63]. However, a serious yet often underreported [64, 65] consequence of 5-FU is the onset of acute cardiotoxicities that arise during or shortly following cessation of treatment [5, 11, 66]. Heightened understanding of the mechanisms responsible for 5-FU cardiotoxicity coupled with identification of modalities to prevent its occurrence may help ensure patients can continue to receive 5-FU-based regimens without increased risk to cardiovascular health. While preclinical studies have played a significant role in growing the current understanding of how 5-FU impacts the cardiovascular system, prior works have exclusively employed IV bolus (20 sec. to 10 minutes) [20, 24, 26, 27, 67] or intraperitoneal [19, 21, 68, 69] dosing models. These models differ significantly from most common 5-FU-based regimens used in clinical practice where 5-FU is often delivered via IV bolus + continuous IV infusion over 46-72 hours [30-32, 70]. This prolonged exposure to 5-FU has been associated with greater treatment efficacy [71,

72] and heightened rates of cardiotoxicity compared to bolus injection alone [34]. Thus, we designed a preclinical dosing model incorporating a continuous infusion of 5-FU to align our study more closely with the delivery methods frequently used in clinical practice. While the present work was not specifically designed to assess the mechanisms of 5-FU cardiotoxicity, preclinical incorporation of a continuous infusion dosing model may provide a more translational approach to interrogate signaling pathways involved in the onset of 5-FU cardiotoxicity in the clinical setting.

In the present study, a single IV bolus + continuous infusion of 5-FU had no effect on left ventricular function compared to saline treatment. This is in agreement with the findings of Zhang and colleagues, who found no differences in FS between a control group and rats receiving 5 consecutive daily doses of either 25 or 50 mg/ kg of 5-FU via intraperitoneal injection [21]. However, these authors did report changes in cardiac mitochondrial function following 5-FU treatment, suggesting 5-FU may be responsible for subclinical cardiac alterations that do not rise to the level of overt cardiac dysfunction. Reductions in left ventricular function following 5-FU treatment are seemingly rare in largescale clinical studies but have been reported in several clinical case reports [8, 73, 74]. It is worth noting, however, that increases in ESV and reductions in FS were observed in each of our experimental groups over the course of the 2-hr infusion—with changes reaching statistical significance in 3 of 4 groups for both parameters. These alterations are seemingly independent of 5-FU and may be a consequence of prolonged use of pentobarbital anesthesia, as reductions in FS have been noted over the course of 3-hr of pentobarbital anesthesia in dogs [75]. Importantly, despite statistically significant reductions in FS, the values reported herein remain well above those seen in our rat model of heart failure (FS = \sim 20-26%) [55-58] and are consistent with previously published values in healthy rats [55-58, 61].

The most prevalent theory of 5-FU cardiotoxicity maintains that side effects (e.g., angina) are likely a consequence of a direct effect of 5-FU on the vasculature. In support of this notion, PWV was increased in SED5FU following the 2-hr infusion but remained unchanged

in SEDCON, EXCON, and EX5FU. This change in PWV occurred without changes in hemodynamic variables known to influence PWV (e.g., MAP, Q, etc.). SED5FU also presented with significantly higher PWV at the 2-hr time point and as a change from BL compared to SEDCON. Notably, PWV increased from BL to the 2-hr time point in all 10 SED5FU animals (average increase of 14.5%). Our group has previously demonstrated a relationship between anticancer therapy and arterial stiffness [76, 77], as well as an association between an index of arterial stiffness and cardiovascular/cancer mortality in some cancer populations [78]. These data highlight the importance of PWV/ arterial stiffness measurements throughout cancer survivorship. However, these measurements are typically made longitudinally (e.g., after completion of several chemotherapy cycles or in the years following treatment) rather than following a single chemotherapy cycle. To the best of our knowledge, others have yet to investigate changes in PWV following a single cycle of 5-FU. Visvikis and colleagues found a ~10% increase in the PWV of colorectal cancer patients after 6-12 cycles of 5-FU delivered via continuous IV infusion [17]. Interestingly, Groehs et al. found no significant change in PWV after 30 weekly bolus IV injections of 5-FU (Groehs et al. 2020) [79]. While the mechanisms responsible for 5-FU-induced vascular complications are not fully understood, they appear to involve a complex array of signaling events spanning the endothelial and smooth muscle layers. Our group and others have demonstrated impairments in endothelial function—an established contributor to increases in PWV and the onset of arterial stiffening [80]—in cancer patients following 5-FU treatment [14, 16]. In rodent models, acute inhibition of nitric oxide synthase is sufficient to increase PWV [81], suggesting that pathways involving nitric oxide signaling may contribute to the increased PWV seen in our study. Alternatively, preclinical studies have noted that 5-FU elicits a direct vasoconstrictor effect on aortic rings [28, 82] that seemingly involves endothelium-independent pathways involving protein kinase C [28]. Thus, future mechanistic studies aimed at illuminating the specific pathways in which 5-FU impairs vascular function remain warranted.

A critical novel finding of the present work was the protective effect of pre-treatment moderate-intensity exercise on 5-FU-induced changes in PWV. Indeed, the significant increase in PWV-apparent in SED5FU over the 2-hr infusion—was not reciprocated in EX5FU. Similarly, the absolute PWV was not different between EX5FU and SEDCON at the 2-hr time point, nor were there differences between the groups when comparing the change in PWV from BL. We interpret these findings to suggest that 4 bouts of exercise preconditioning may induce stimuli capable of preventing the adverse vascular consequences of 5-FU treatment. Acute exercise preconditioning has previously been demonstrated to prevent cardiotoxicity induced by anthracycline chemotherapy in rodents [45, 46, 48] and human patients [83] with a specific focus on left ventricular function. To our knowledge, like studies have yet to be completed prior to 5-FU treatment. Using a long-term (4-8 weeks) exercise preconditioning model, Hayward and colleagues demonstrated that aortic rings isolated from exercise trained rats (20-25 m/min, 5 d/w, 8-wks) exhibit improved acetylcholine-mediated dilation following preconstriction with 5-FU [82]. The authors posit this improvement in endothelium-dependent vasodilation may be related to increased aortic content of endothelial NO synthase [82]. While the findings of Hayward and colleagues indicate the potential benefits of long-term exercise preconditioning prior to 5-FU exposure, implementation of such regimens (i.e., 4-8 weeks) may not be feasible in the time between diagnosis and treatment onset - especially considering that minimization of this timeframe is critical for cancer survival outcomes [84, 85]. In a non-cancer cohort of patients experiencing rest angina, Morikawa and colleagues found that three days of aerobic interval training improved endothelial function and reduced the incidence of coronary spastic angina [43]. Given that 5-FU cardiotoxicity most frequently presents as angina during or shortly following the first cycle of treatment [5, 34], these findings underscore the potential benefit of acute exercise for preventing the primary symptom of 5-FU cardiotoxicity. Though the risk for cardiovascular events in the months following 5-FU completion is seemingly low [86], this is not to say that additional exercise prescription of longer durations following treatment completion would not be beneficial. Courneya and colleagues recently found significant reductions in relative risk for disease recurrence. new primary cancer, and disease-free survival in colorectal cancer patients who participated in a three-year exercise training program initiated following completion of adjuvant chemotherapy containing 5-FU (or its oral prodrug Capecitabine) [87]. While cardiovascular outcomes were not reported, these findings signify the benefit post treatment exercise can have on overall cancer survival. Future work focused on implementation of individualized exercise prescriptions that prevent adverse cardiovascular effects of the specific treatment while simultaneously improving cancer survival outcomes are warranted and have the potential to greatly improve quality of life across the cancer survivorship continuum.

Experimental considerations

Several experimental considerations are pertinent to interpreting the present study's findings. First, a key component of our work included using a novel pre-clinical dosing regimen designed to reflect 5-FU delivery methods used in clinical practice. To perform this protocol, animals remained under pentobarbital anesthesia for approximately 150-180 minutes. Such prolonged periods of pentobarbital anesthesia can result in alterations in cardiac parameters (as discussed above) that should be considered in the interpretation of our findings. However, pentobarbital has been demonstrated to have less impact on cardiovascular function than other commonly used anesthetics (e.g., isoflurane, ketamine/xylazine) [88]. Additionally, anesthetic use was the same across all groups to minimize the potential effect of anesthesia on group comparisons. We cannot fully discount a potential interaction between pentobarbital sodium and 5-FU; however, since each animal was treated the same, and there were no differences in MAP or HR across groups (Table 1), the use of anesthesia likely had minimal impact on the PWV results herein. Future work could consider use of both implantable infusion pumps and/or telemetry devices that would allow for continuous infusion and measurement of cardiovascular parameters under minimal anesthesia. Second, only male animals were used in the present study. While large-scale studies have found no difference in the incidence of 5-FU cardiotoxicity between men and women [3, 7], the onset of general cardiac and vascular pathology often differs between sexes. For example, women may have slightly higher angina incidence rates than men across the lifespan [89, 90] and menopausal transition can contribute to vasomotor changes and elevate cardiovascular risk [91]. Additionally, emerging evidence suggests potential differences in both molecular signaling and adaption to exercise in men and women [92]. This is an important and emerging topic of research that highlights the critical need to consider biological sex in future exercise oncology research. Future studies assessing the ability of exercise to mitigate 5-FU cardiotoxicity would benefit from the inclusion of both male and female rats to identify potential differences to either 5-FU treatment or the efficacy of exercise preconditioning. All rats used in the present study were free of cancer, as is common in most studies of chemotherapy-induced cardiotoxicity [19, 21, 24, 26, 27, 45-47]. While tumor naïve rodent models are key to isolate the effects of chemotherapy from those of the cancer itself, recent evidence suggests that the cancer alone [51] or in concert with various treatments [93] may increase the risk of future cardiovascular disease and mortality. Preclinical models also indicate that the tumor may contribute to cardiovascular pathology independent of cancer treatment [49, 50, 52]. Though it is possible that the underlying mechanisms of 5-FU cardiotoxicity may include complex interactions between 5-FU and the cancer itself, 5-FU has an established direct vasoconstrictor effect on isolated aortic rings [28, 82], and the acute nature of the toxicity, coupled with its disappearance upon cessation of 5-FU treatment suggest a direct effect of the drug independent of the tumor is likely. Nonetheless, inclusion of tumor-bearing animals in future studies may help illuminate a potential role of the tumor and further improve the translatability of preclinical models of 5-FU cardiotoxicity. Similarly, 5-FU is regularly delivered alongside several other chemotherapy drugs (e.g., leucovorin, oxaliplatin, bevacizumab, etc.) to improve cancer treatment efficacy. Some of these drugs have been suggested to increase the incidence of 5-FU-induced cardiotoxicity [33, 941. While the focus of the current study was to understand the impact of 5-FU alone, future works may seek to include additional treatments to better understand the cardiovascular

effects of specific 5-FU-based regimens (e.g., FOLFOX, FOLFIRINOX).

Conclusion

The present work sought to determine whether acute exercise preconditioning could prevent cardiotoxicity caused by a clinically relevant dose of 5-FU chemotherapy. Our findings demonstrate that a single cycle of 5-FU increases PWV (a clinical marker of aortic stiffness) but does not cause overt changes in left ventricular function in male rats. Importantly, exercise preconditioning was sufficient to prevent the 5-FU-induced increase in PWV. These findings indicate regimented moderate-intensity exercise in the days immediately before 5-FU treatment may mitigate the onset of 5-FU cardiotoxicity. Future studies designed to assess the mechanisms of 5-FU cardiotoxicity as well as the efficacy and feasibility of exercise preconditioning programs in human cancer patients remain critical to ensuring patients can continue to receive this effective treatment without detrimental effects to cardiovascular well-being.

Acknowledgements

This work was supported by funding from the Johnson Cancer Research Center awarded to C.J.A., B.J.B., S.T.H. and D.R.B. and the American College of Sports Medicine Foundation Doctoral Student Research Grant awarded to S.T.H. S.T.H. is supported by the National Institutes of Health Postdoctoral Fellowship Grant NIH T32HL34643 and the MCW Cardiovascular Research Center's A.O. Smith Fellowship Scholars Program. D.R.B is supported by the American Heart Association Grant award 25POST1373227. A.G.H. is supported by the National Institutes of Health Postdoctoral Fellowship Grant T32HL007576.

Disclosure of conflict of interest

None.

Address correspondence to: Dr. Stephen T Hammond, Department of Medicine, Cardiovascular Research Center, Medical College of Wisconsin, 8701 Watertown Plank Road, Milwaukee, WI 53226, USA. E-mail: sthammond@mcw.edu; st.hammond1@gmail.com; Dr. Dryden R Baumfalk, Department of Orthapaedic Surgery, Duke University, 2080 Duke

University Road, Durham, NC 27708, USA. Email: d.baumfalk@duke.edu

References

- [1] Heidelberger C, Chaudhuri NK, Danneberg P, Mooren D, Griesbach L, Duschinsky R, Schnitzer RJ, Pleven E and Scheiner J. Fluorinated pyrimidines, a new class of tumour-inhibitory compounds. Nature 1957; 179: 663-666.
- [2] Dyhl-Polk A, Schou M, Vistisen KK, Sillesen AS, Serup-Hansen E, Faber J, Klausen TW, Bojesen SE, Vaage-Nilsen M and Nielsen DL. Myocardial ischemia induced by 5-fluorouracil: a prospective electrocardiographic and cardiac biomarker study. Oncologist 2021; 26: e403e413.
- [3] Meyer CC, Calis KA, Burke LB, Walawander CA and Grasela TH. Symptomatic cardiotoxicity associated with 5-fluorouracil. Pharmacotherapy 1997; 17: 729-736.
- [4] Rezkalla S, Kloner RA, Ensley J, al-Sarraf M, Revels S, Olivenstein A, Bhasin S, Kerpel-Fronious S and Turi ZG. Continuous ambulatory ECG monitoring during fluorouracil therapy: a prospective study. J Clin Oncol 1989; 7: 509-514.
- [5] Sara JD, Kaur J, Khodadadi R, Rehman M, Lobo R, Chakrabarti S, Herrmann J, Lerman A and Grothey A. 5-fluorouracil and cardiotoxicity: a review. Ther Adv Med Oncol 2018; 10: 1758835918780140.
- [6] Wacker A, Lersch C, Scherpinski U, Reindl L and Seyfarth M. High incidence of angina pectoris in patients treated with 5-fluorouracil. A planned surveillance study with 102 patients. Oncology 2003; 65: 108-112.
- [7] Zafar A, Drobni ZD, Mosarla R, Alvi RM, Lei M, Lou UY, Raghu VK, Murphy SP, Jones-O'Connor M, Hartmann S, Gilman HK, Weekes CD, Clark JR, Clark J, Blaszkowsky L, Tavares E and Neilan TG. The incidence, risk factors, and outcomes with 5-fluorouracil-associated coronary vasospasm. JACC CardioOncol 2021; 3: 101-109.
- [8] Dechant C, Baur M, Bock R, Czejka M, Podczeck-Schweighofer A, Dittrich C and Christ G. Acute reversible heart failure caused by coronary vasoconstriction due to continuous 5-fluorouracil combination chemotherapy. Case Rep Oncol 2012; 5: 296-301.
- [9] Dyhl-Polk A, Vaage-Nilsen M, Schou M, Vistisen KK, Lund CM, Kumler T, Appel JM and Nielsen DL. Incidence and risk markers of 5-fluorouracil and capecitabine cardiotoxicity in patients with colorectal cancer. Acta Oncol 2020; 59: 475-483.
- [10] Lestuzzi C, Stolfo D, De Paoli A, Banzato A, Buonadonna A, Bidoli E, Tartuferi L, Viel E, De An-

- gelis G, Lonardi S, Innocente R, Berretta M, Bergamo F, Guglielmi A, Sinagra G and Herrmann J. Cardiotoxicity from capecitabine chemotherapy: prospective study of incidence at rest and during physical exercise. Oncologist 2022; 27: e158-e167.
- [11] Polk A, Vaage-Nilsen M, Vistisen K and Nielsen DL. Cardiotoxicity in cancer patients treated with 5-fluorouracil or capecitabine: a systematic review of incidence, manifestations and predisposing factors. Cancer Treat Rev 2013; 39: 974-984.
- [12] Luwaert RJ, Descamps O, Majois F, Chaudron JM and Beauduin M. Coronary artery spasm induced by 5-fluorouracil. Eur Heart J 1991; 12: 468-470.
- [13] Shoemaker LK, Arora U and Rocha Lima CM. 5-fluorouracil-induced coronary vasospasm. Cancer Control 2004; 11: 46-49.
- [14] Hammond ST, Baumfalk DR, Parr SK, Butenas ALE, Scheuermann BC, Turpin VG, Behnke BJ, Hashmi MH and Ade CJ. Impaired microvascular reactivity in patients treated with 5-fluorouracil chemotherapy regimens: potential role of endothelial dysfunction. Int J Cardiol Heart Vasc 2023; 49: 101300.
- [15] Salepci T, Seker M, Uyarel H, Gumus M, Bilici A, Ustaalioglu BB, Ozturk A, Sonmez B, Orcun A, Ozates M, Irmak R and Yaylaci M. 5-fluorouracil induces arterial vasoconstrictions but does not increase angiotensin II levels. Med Oncol 2010; 27: 416-420.
- [16] Sudhoff T, Enderle MD, Pahlke M, Petz C, Teschendorf C, Graeven U and Schmiegel W. 5-fluorouracil induces arterial vasocontractions. Ann Oncol 2004; 15: 661-664.
- [17] Visvikis A, Kyvelou SM, Pietri P, Georgakopoulos C, Manousou K, Tousoulis D, Stefanadis C, Vlachopoulos C and Pektasides D. Cardiotoxic profile and arterial stiffness of adjuvant chemotherapy for colorectal cancer. Cancer Manag Res 2020; 12: 1175-1185.
- [18] Focaccetti C, Bruno A, Magnani E, Bartolini D, Principi E, Dallaglio K, Bucci EO, Finzi G, Sessa F, Noonan DM and Albini A. Effects of 5-fluorouracil on morphology, cell cycle, proliferation, apoptosis, autophagy and ROS production in endothelial cells and cardiomyocytes. PLoS One 2015; 10: e0115686.
- [19] Muhammad RN, Sallam N and El-Abhar HS. Activated ROCK/Akt/eNOS and ET-1/ERK pathways in 5-fluorouracil-induced cardiotoxicity: modulation by simvastatin. Sci Rep 2020; 10: 14693.
- [20] Tsibiribi P, Bui-Xuan C, Bui-Xuan B, Lombard-Bohas C, Duperret S, Belkhiria M, Tabib A, Maujean G, Descotes J and Timour Q. Cardiac lesions induced by 5-fluorouracil in the rabbit. Hum Exp Toxicol 2006; 25: 305-309.

- [21] Zhang D and Ma J. Mitochondrial dynamics in rat heart induced by 5-fluorouracil. Med Sci Monit 2018; 24: 6666-6672.
- [22] Altieri P, Murialdo R, Barisione C, Lazzarini E, Garibaldi S, Fabbi P, Ruggeri C, Borile S, Carbone F, Armirotti A, Canepa M, Ballestrero A, Brunelli C, Montecucco F, Ameri P and Spallarossa P. 5-fluorouracil causes endothelial cell senescence: potential protective role of glucagon-like peptide 1. Br J Pharmacol 2017; 174: 3713-3726.
- [23] Cwikiel M, Eskilsson J, Albertsson M and Stavenow L. The influence of 5-fluorouracil and methotrexate on vascular endothelium. An experimental study using endothelial cells in the culture. Ann Oncol 1996; 7: 731-737.
- [24] Cwikiel M, Eskilsson J, Wieslander JB, Stjernquist U and Albertsson M. The appearance of endothelium in small arteries after treatment with 5-fluorouracil. An electron microscopic study of late effects in rabbits. Scanning Microsc 1996; 10: 805-818; discussion 819.
- [25] Gajalakshmi P, Priya MK, Pradeep T, Behera J, Muthumani K, Madhuwanti S, Saran U and Chatterjee S. Breast cancer drugs dampen vascular functions by interfering with nitric oxide signaling in endothelium. Toxicol Appl Pharmacol 2013; 269: 121-131.
- [26] Kinhult S, Albertsson M, Eskilsson J and Cwikiel M. Antithrombotic treatment in protection against thrombogenic effects of 5-fluorouracil on vascular endothelium: a scanning microscopy evaluation. Scanning 2001; 23: 1-8.
- [27] Kinhult S, Albertsson M, Eskilsson J and Cwikiel M. Effects of probucol on endothelial damage by 5-fluorouracil. Acta Oncol 2003; 42: 304-308.
- [28] Mosseri M, Fingert HJ, Varticovski L, Chokshi S and Isner JM. In vitro evidence that myocardial ischemia resulting from 5-fluorouracil chemotherapy is due to protein kinase C-mediated vasoconstriction of vascular smooth muscle. Cancer Res 1993; 53: 3028-3033.
- [29] Li Y, Zhang Y, Zhou X, Lei X, Li X and Wei L. Dynamic observation of 5-fluorouracil-induced myocardial injury and mitochondrial autophagy in aging rats. Exp Ther Med 2021; 22: 1451.
- [30] Cheeseman SL, Joel SP, Chester JD, Wilson G, Dent JT, Richards FJ and Seymour MT. A 'modified de Gramont' regimen of fluorouracil, alone and with oxaliplatin, for advanced colorectal cancer. Br J Cancer 2002; 87: 393-399.
- [31] Conroy T, Desseigne F, Ychou M, Bouche O, Guimbaud R, Becouarn Y, Adenis A, Raoul JL, Gourgou-Bourgade S, de la Fouchardiere C, Bennouna J, Bachet JB, Khemissa-Akouz F, Pere-Verge D, Delbaldo C, Assenat E, Chauffert

- B, Michel P, Montoto-Grillot C and Ducreux M; Groupe Tumeurs Digestives of Unicancer; PRODIGE Intergroup. FOLFIRINOX versus gemcitabine for metastatic pancreatic cancer. N Engl J Med 2011; 364: 1817-1825.
- [32] Petrelli F, Borgonovo K, Cabiddu M, Ghilardi M, Lonati V, Maspero F, Sauta MG, Beretta GD and Barni S. FOLFIRI-bevacizumab as first-line chemotherapy in 3500 patients with advanced colorectal cancer: a pooled analysis of 29 published trials. Clin Colorectal Cancer 2013; 12: 145-151.
- [33] Kosmas C, Kallistratos MS, Kopterides P, Syrios J, Skopelitis H, Mylonakis N, Karabelis A and Tsavaris N. Cardiotoxicity of fluoropyrimidines in different schedules of administration: a prospective study. J Cancer Res Clin Oncol 2008; 134: 75-82.
- [34] Saif MW, Shah MM and Shah AR. Fluoropyrimidine-associated cardiotoxicity: revisited. Expert Opin Drug Saf 2009; 8: 191-202.
- [35] Eskilsson J and Albertsson M. Failure of preventing 5-fluorouracil cardiotoxicity by prophylactic treatment with verapamil. Acta Oncol 1990; 29: 1001-1003.
- [36] Patel B, Kloner RA, Ensley J, Al-Sarraf M, Kish J and Wynne J. 5-fluorouracil cardiotoxicity: left ventricular dysfunction and effect of coronary vasodilators. Am J Med Sci 1987; 294: 238-243
- [37] Zafar A, Drobni ZD, Lei M, Gongora CA, Quinaglia T, Lou UY, Mosarla R, Murphy SP, Jones-O'Connor M, Mahmood A, Hartmann S, Gilman HK, Weekes CD, Nipp R, Clark JR, Clark JW, Blaszkowsky LS, Tavares E and Neilan TG. The efficacy and safety of cardio-protective therapy in patients with 5-FU (Fluorouracil)-associated coronary vasospasm. PLoS One 2022; 17: e0265767.
- [38] Tucker WJ, Fegers-Wustrow I, Halle M, Haykowsky MJ, Chung EH and Kovacic JC. Exercise for primary and secondary prevention of cardiovascular disease: JACC focus seminar 1/4. J Am Coll Cardiol 2022; 80: 1091-1106.
- [39] Chicco AJ, Schneider CM and Hayward R. Voluntary exercise protects against acute doxorubicin cardiotoxicity in the isolated perfused rat heart. Am J Physiol Regul Integr Comp Physiol 2005; 289: R424-R431.
- [40] Chicco AJ, Schneider CM and Hayward R. Exercise training attenuates acute doxorubicin-induced cardiac dysfunction. J Cardiovasc Pharmacol 2006; 47: 182-189.
- [41] Wang J, Liu S, Meng X, Zhao X, Wang T, Lei Z, Lehmann HI, Li G, Alcaide P, Bei Y and Xiao J. Exercise inhibits doxorubicin-induced cardiotoxicity via regulating B cells. Circ Res 2024; 134: 550-568.

- [42] Marques-Aleixo I, Santos-Alves E, Torrella JR, Oliveira PJ, Magalhães J and Ascensão A. Exercise and doxorubicin treatment modulate cardiac mitochondrial quality control signaling. Cardiovasc Toxicol 2018; 18: 43-55.
- [43] Morikawa Y, Mizuno Y, Harada E, Katoh D, Kashiwagi Y, Morita S, Yoshimura M, Uemura S, Saito Y and Yasue H. Aerobic interval exercise training in the afternoon reduces attacks of coronary spastic angina in conjunction with improvement in endothelial function, oxidative stress, and inflammation. Coron Artery Dis 2013; 24: 177-182.
- [44] Thijssen DHJ, Uthman L, Somani Y and van Royen N. Short-term exercise-induced protection of cardiovascular function and health: why and how fast does the heart benefit from exercise? J Physiol 2022; 600: 1339-1355.
- [45] Ascensao A, Lumini-Oliveira J, Machado NG, Ferreira RM, Goncalves IO, Moreira AC, Marques F, Sardao VA, Oliveira PJ and Magalhaes J. Acute exercise protects against calcium-induced cardiac mitochondrial permeabilitytransition pore opening in doxorubicin-treated rats. Clin Sci (Lond) 2011; 120: 37-49.
- [46] Kavazis AN, Smuder AJ, Min K, Tumer N and Powers SK. Short-term exercise training protects against doxorubicin-induced cardiac mitochondrial damage independent of HSP72. Am J Physiol Heart Circ Physiol 2010; 299: H1515-1524.
- [47] Kavazis AN, Smuder AJ and Powers SK. Effects of short-term endurance exercise training on acute doxorubicin-induced FoxO transcription in cardiac and skeletal muscle. J Appl Physiol (1985) 2014; 117: 223-230.
- [48] Wonders KY, Hydock DS, Schneider CM and Hayward R. Acute exercise protects against doxorubicin cardiotoxicity. Integr Cancer Ther 2008; 7: 147-154.
- [49] Baumfalk DR, Opoku-Acheampong AB, Caldwell JT, Ade CJ, Copp SW, Musch TI and Behnke BJ. Effects of prostate cancer and exercise training on left ventricular function and cardiac and skeletal muscle mass. J Appl Physiol (1985) 2019; 126: 668-680.
- [50] Esau PJ, Gittemeier EM, Opoku-Acheampong AB, Rollins KS, Baumfalk DR, Poole DC, Musch TI, Behnke BJ and Copp SW. Prostate cancer reduces endurance exercise capacity in association with reductions in cardiac and skeletal muscle mass in the rat. Am J Cancer Res 2017; 7: 2566-2576.
- [51] Guan T, Zhang H, Yang J, Lin W, Wang K, Su M, Peng W, Li Y, Lai Y and Liu C. Increased risk of cardiovascular death in breast cancer patients without chemotherapy or (and) radiotherapy: a large population-based study. Front Oncol 2020; 10: 619622.

- [52] Ogilvie LM, Delfinis LJ, Coyle-Asbil B, Vudatha V, Alshamali R, Garlisi B, Pereira M, Matuszewska K, Garibotti MC, Gandhi S, Brunt KR, Wood GA, Trevino JG, Perry CGR, Petrik J and Simpson JA. Cardiac atrophy, dysfunction, and metabolic impairments: a cancer-induced cardiomyopathy phenotype. Am J Pathol 2024; 194: 1823-1843.
- [53] Bedford TG, Tipton CM, Wilson NC, Oppliger RA and Gisolfi CV. Maximum oxygen consumption of rats and its changes with various experimental procedures. J Appl Physiol Respir Environ Exerc Physiol 1979; 47: 1278-1283.
- [54] Nair AB and Jacob S. A simple practice guide for dose conversion between animals and human. J Basic Clin Pharm 2016; 7: 27-31.
- [55] Craig JC, Colburn TD, Caldwell JT, Hirai DM, Tabuchi A, Baumfalk DR, Behnke BJ, Ade CJ, Musch TI and Poole DC. Central and peripheral factors mechanistically linked to exercise intolerance in heart failure with reduced ejection fraction. Am J Physiol Heart Circ Physiol 2019; 317: H434-H444.
- [56] Butenas ALE, Rollins KS, Williams AC, Parr SK, Hammond ST, Ade CJ, Hageman KS, Musch TI and Copp SW. Thromboxane A(2) receptors contribute to the exaggerated exercise pressor reflex in male rats with heart failure. Physiol Rep 2021; 9: e15052.
- [57] Butenas ALE, Rollins KS, Williams AC, Parr SK, Hammond ST, Ade CJ, Hageman KS, Musch TI and Copp SW. Exaggerated sympathetic and cardiovascular responses to dynamic mechanoreflex activation in rats with heart failure: role of endoperoxide 4 and thromboxane A(2) receptors. Auton Neurosci 2021; 232: 102784.
- [58] Butenas ALE, Rollins KS, Matney JE, Williams AC, Kleweno TE, Parr SK, Hammond ST, Ade CJ, Hageman KS, Musch TI and Copp SW. No effect of endoperoxide 4 or thromboxane A(2) receptor blockade on static mechanoreflex activation in rats with heart failure. Exp Physiol 2020; 105: 1840-1854.
- [59] Teichholz LE, Kreulen T, Herman MV and Gorlin R. Problems in echocardiographic volume determinations: echocardiographic-angiographic correlations in the presence of absence of asynergy. Am J Cardiol 1976; 37: 7-11.
- [60] Laurent S, Cockcroft J, Van Bortel L, Boutouyrie P, Giannattasio C, Hayoz D, Pannier B, Vlachopoulos C, Wilkinson I and Struijker-Boudier H; European Network for Non-invasive Investigation of Large Arteries. Expert consensus document on arterial stiffness: methodological issues and clinical applications. Eur Heart J 2006; 27: 2588-2605.
- [61] Pawlush DG, Moore RL, Musch TI and Davidson WR Jr. Echocardiographic evaluation of

- size, function, and mass of normal and hypertrophied rat ventricles. J Appl Physiol (1985) 1993; 74: 2598-2605.
- [62] Ansfield FJ, Schroeder JM and Curreri AR. Five years clinical experience with 5-fluorouracil. JAMA 1962; 181: 295-299.
- [63] Gustavsson B, Carlsson G, Machover D, Petrelli N, Roth A, Schmoll HJ, Tveit KM and Gibson F. A review of the evolution of systemic chemotherapy in the management of colorectal cancer. Clin Colorectal Cancer 2015; 14: 1-10.
- [64] Lestuzzi C, Vaccher E, Talamini R, Lleshi A, Meneguzzo N, Viel E, Scalone S, Tartuferi L, Buonadonna A, Ejiofor L and Schmoll HJ. Effort myocardial ischemia during chemotherapy with 5-fluorouracil: an underestimated risk. Ann Oncol 2014; 25: 1059-1064.
- [65] Lestuzzi C, Tartuferi L, Viel E, Buonadonna A, Vaccher E and Berretta M. Fluoropyrimidine-associated cardiotoxicity: probably not so rare as it seems. Oncologist 2020; 25: e1254.
- [66] Polk A, Vistisen K, Vaage-Nilsen M and Nielsen DL. A systematic review of the pathophysiology of 5-fluorouracil-induced cardiotoxicity. BMC Pharmacol Toxicol 2014; 15: 47.
- [67] Matsubara I, Kamiya J and Imai S. Cardiotoxic effects of 5-fluorouracil in the guinea pig. Jpn J Pharmacol 1980; 30: 871-879.
- [68] Barary M, Hosseinzadeh R, Kazemi S, Liang JJ, Mansoori R, Sio TT, Hosseini M and Moghadamnia AA. The effect of propolis on 5-fluorouracil-induced cardiac toxicity in rats. Sci Rep 2022; 12: 8661.
- [69] Durak I, Karaayvaz M, Kavutcu M, Cimen MY, Kacmaz M, Buyukkocak S and Ozturk HS. Reduced antioxidant defense capacity in myocardial tissue from guinea pigs treated with 5-fluorouracil. J Toxicol Environ Health A 2000; 59: 585-589.
- [70] Akdeniz N, Kaplan MA, Uncu D, Inanc M, Kaya S, Dane F, Kucukoner M, Demirci A, Bilici M, Durnali AG, Koral L, Sendur MAN, Erol C, Turkmen E, Olmez OF, Acikgoz O, Lacin S, Sahinli H, Urakci Z and Isikdogan A. The comparison of FOLFOX regimens with different doses of 5-FU for the adjuvant treatment of colorectal cancer: a multicenter study. Int J Colorectal Dis 2021; 36: 1311-1319.
- [71] Amorim LC and Peixoto RD. Should we still be using bolus 5-FU prior to infusional regimens in gastrointestinal cancers? A practical review. Int Cancer Conf J 2022; 11: 2-5.
- [72] de Gramont A, Bosset JF, Milan C, Rougier P, Bouche O, Etienne PL, Morvan F, Louvet C, Guillot T, Francois E and Bedenne L. Randomized trial comparing monthly low-dose leucovorin and fluorouracil bolus with bimonthly highdose leucovorin and fluorouracil bolus plus

- continuous infusion for advanced colorectal cancer: a French intergroup study. J Clin Oncol 1997; 15: 808-815.
- [73] Allison JD, Tanavin T, Yang Y, Birnbaum G and Khalid U. Various manifestations of 5-fluorouracil cardiotoxicity: a multicenter case series and review of literature. Cardiovasc Toxicol 2020; 20: 437-442.
- [74] Chaudary S, Song SY and Jaski BE. Profound, yet reversible, heart failure secondary to 5-fluorouracil. Am J Med 1988; 85: 454-456.
- [75] Lang RM, Marcus RH, Neumann A, Janzen D, Hansen D, Fujii AM and Borow KM. A timecourse study of the effects of pentobarbital, fentanyl, and morphine chloralose on myocardial mechanics. J Appl Physiol (1985) 1992; 73: 143-150.
- [76] Frye JN, Sutterfield SL, Caldwell JT, Behnke BJ, Copp SW, Banister HR and Ade CJ. Vascular and autonomic changes in adult cancer patients receiving anticancer chemotherapy. J Appl Physiol (1985) 2018; 125: 198-204.
- [77] Parr SK, Liang J, Schadler KL, Gilchrist SC, Steele CC and Ade CJ. Anticancer therapy-related increases in arterial stiffness: a systematic review and meta-analysis. J Am Heart Assoc 2020; 9: e015598.
- [78] Parr SK, Steele CC, Hammond ST, Turpin VRG and Ade CJ. Arterial stiffness is associated with cardiovascular and cancer mortality in cancer patients: insight from NHANESIII. Int J Cardiol Hypertens 2021; 9: 100085.
- [79] Groehs RV, Negrao MV, Hajjar LA, Jordao CP, Carvalho BP, Toschi-Dias E, Andrade AC, Hodas FP, Alves MJNN, Sarmento AO, Testa L, Hoff PMG, Negrao CE and Filho RK. Adjuvant treatment with 5-fluorouracil and oxaliplatin does not influence cardiac function, neurovascular control, and physical capacity in patients with colon cancer. Oncologist 2021; 26: e907.
- [80] Van den Bergh G, Opdebeeck B, D'Haese PC and Verhulst A. The vicious cycle of arterial stiffness and arterial media calcification. Trends Mol Med 2019; 25: 1133-1146.
- [81] Fitch RM, Vergona R, Sullivan ME and Wang YX. Nitric oxide synthase inhibition increases aortic stiffness measured by pulse wave velocity in rats. Cardiovasc Res 2001; 51: 351-358.
- [82] Hayward R, Ruangthai R, Schneider CM, Hyslop RM, Strange R and Westerlind KC. Training enhances vascular relaxation after chemotherapy-induced vasoconstriction. Med Sci Sports Exerc 2004; 36: 428-434.
- [83] Kirkham AA, Shave RE, Bland KA, Bovard JM, Eves ND, Gelmon KA, McKenzie DC, Virani SA, Stohr EJ, Warburton DER and Campbell KL. Protective effects of acute exercise prior to doxorubicin on cardiac function of breast can-

Acute exercise prevents 5-FU cardiotoxicity

- cer patients: a proof-of-concept RCT. Int J Cardiol 2017; 245: 263-270.
- [84] Ungvari Z, Fekete M, Fekete JT, Lehoczki A, Buda A, Munkácsy G, Varga P, Ungvari A and Győrffy B. Treatment delay significantly increases mortality in colorectal cancer: a metaanalysis. Geroscience 2025; 47: 5337-5353.
- [85] Tevis SE, Kohlnhofer BM, Stringfield S, Foley EF, Harms BA, Heise CP and Kennedy GD. Postoperative complications in patients with rectal cancer are associated with delays in chemotherapy that lead to worse disease-free and overall survival. Dis Colon Rectum 2013; 56: 1339-1348.
- [86] Shanmuganathan JWD, Kragholm K, Tayal B, Polcwiartek C, Poulsen LØ, El-Galaly TC, Fosbøl EL, D'Souza M, Gislason G, Køber L, Schou M, Nielsen D, Søgaard P, Torp-Pedersen CT, Mamas MA and Freeman P. Risk for myocardial infarction following 5-fluorouracil treatment in patients with gastrointestinal cancer: a nationwide registry-based study. JACC CardioOncol 2021; 3: 725-733.
- [87] Courneya KS, Vardy JL, O'Callaghan CJ, Gill S, Friedenreich CM, Wong RKS, Dhillon HM, Coyle V, Chua NS, Jonker DJ, Beale PJ, Haider K, Tang PA, Bonaventura T, Wong R, Lim HJ, Burge ME, Hubay S, Sanatani M, Campbell KL, Arthuso FZ, Turner J, Meyer RM, Brundage M, O'Brien P, Tu D and Booth CM; CHALLENGE Investigators. Structured exercise after adjuvant chemotherapy for colon cancer. N Engl J Med 2025; 393: 13-25.
- [88] Stein AB, Tiwari S, Thomas P, Hunt G, Levent C, Stoddard MF, Tang XL, Bolli R and Dawn B. Effects of anesthesia on echocardiographic assessment of left ventricular structure and function in rats. Basic Res Cardiol 2007; 102: 28-41.

- [89] Hemingway H, Langenberg C, Damant J, Frost C, Pyörälä K and Barrett-Connor E. Prevalence of angina in women versus men: a systematic review and meta-analysis of international variations across 31 countries. Circulation 2008; 117: 1526-1536.
- [90] Ji H, Kwan AC, Chen MT, Ouyang D, Ebinger JE, Bell SP, Niiranen TJ, Bello NA and Cheng S. Sex differences in myocardial and vascular aging. Circ Res 2022; 130: 566-577.
- [91] El Khoudary SR, Aggarwal B, Beckie TM, Hodis HN, Johnson AE, Langer RD, Limacher MC, Manson JE, Stefanick ML and Allison MA; American Heart Association Prevention Science Committee of the Council on Epidemiology and Prevention; and Council on Cardiovascular and Stroke Nursing. Menopause transition and cardiovascular disease risk: implications for timing of early prevention: a scientific statement from the american heart association. Circulation 2020; 142: e506-e532.
- [92] Ansdell P, Thomas K, Hicks KM, Hunter SK, Howatson G and Goodall S. Physiological sex differences affect the integrative response to exercise: acute and chronic implications. Exp Physiol 2020; 105: 2007-2021.
- [93] Chi K, Luo Z, Zhao H, Li Y, Liang Y, Xiao Z, He Y, Zhang H, Ma Z, Zeng L, Zhou R, Feng M, Li W, Rao H and Yi M. The impact of tumor characteristics on cardiovascular disease death in breast cancer patients with CT or RT: a population-based study. Front Cardiovasc Med 2023; 10: 1149633.
- [94] Abdel-Rahman O. 5-fluorouracil-related cardiotoxicity; findings from five randomized studies of 5-fluorouracil-based regimens in metastatic colorectal cancer. Clin Colorectal Cancer 2019; 18: 58-63.