

## Review Article

# COL10A1 beyond skeletal development: a hypertrophic chondrocyte-specific collagen emerging as a potential biomarker and tumor microenvironment regulator in solid cancers

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**Abstract:** Type X collagen (COL10A1) is an extracellular matrix protein primarily expressed by hypertrophic chondrocytes and is essential for endochondral ossification and skeletal mineralization. Although traditionally regarded as cartilage-specific, recent studies have reported its re-expression in a range of solid tumors. This shift in expression pattern suggests that COL10A1 may have functions beyond development, particularly within the tumor microenvironment (TME). Elevated COL10A1 levels have been reported in cancers such as breast, gastric, colorectal, and lung cancers, where both tissue expression and circulating levels are frequently associated with advanced disease stage and poor clinical outcomes. These observations support COL10A1 as a potential diagnostic and prognostic biomarker. Beyond its clinical associations, accumulating evidence indicates that COL10A1 actively contributes to tumor progression. It is involved in extracellular matrix remodeling, angiogenesis, epithelial-mesenchymal transition (EMT), and alterations in immune cell infiltration, and possibly immunotherapy response. In addition, its enrichment in specific subsets of cancer-associated fibroblasts (CAFs) highlights its essential role in tumor-stroma interactions. At the mechanistic level, COL10A1 has been linked to multiple oncogenic signaling pathways, including TGF- $\beta$ 1/SOX9, as well as downstream DDR2/FAK and ITGB1/PI3K/AKT pathways, which may collectively promote tumor invasion, metastasis, and therapy resistance. However, its biological functions are not yet fully defined, and tumor heterogeneity continues to complicate its clinical application. Further studies are needed to clarify how COL10A1-functions within the tumor microenvironment and to determine its clinical value as a biomarker and a potential therapeutic target in solid cancers.

**Keywords:** COL10A1, cancer-associated fibroblasts, biomarker, tumor microenvironment

## Introduction

The extracellular matrix (ECM) is a key component of the tumor microenvironment, consisting of a complex network of structural and functional macromolecules [1]. Rather than

acting merely as a static scaffold for cell adhesion, the ECM functions as a dynamic and biologically active system that influences essential cellular behaviors such as proliferation, differentiation, and migration [2]. It is composed of collagens, glycoproteins, proteoglycans, and

various polysaccharides, which together provide both mechanical support and signal transduction [3]. Under normal physiological conditions, ECM homeostasis is tightly regulated, maintaining a balance between synthesis and degradation to support tissue structure and development [4]. However, in cancer, this balance is disrupted. The ECM undergoes extensive remodeling, which is conducive to tumor initiation, progression, and therapeutic resistance [5]. These changes may involve alterations in composition and structure, abnormal collagen cross-linking, redistribution of matrix components, and increased matrix stiffness, all of which collectively reshape the tumor microenvironment [6]. This remodeling not only promotes tumor cell proliferation, invasion, and metastasis [5], but also forms a physical and biochemical barrier that hinders drug delivery and limits immune cell infiltration, ultimately reducing treatment efficacy [5, 7].

Collagen is one of the most abundant proteins in the human body and is the main structural component of the extracellular matrix [8, 9]. To date, 28 types of collagen have been identified. They all share a characteristic triple-helical structure composed of three  $\alpha$  chains arranged in a right-handed configuration [10, 11]. These  $\alpha$  chains are defined by repeating glycine-X-Y motifs, where X and Y are often proline and hydroxyproline, and these residues play a critical role in stabilizing the triple-helical structure [10, 12].

In addition to its well-established structural role in maintaining tissue integrity and mechanical properties, collagen is also involved in cell signal transduction. Through interactions with cell surface receptors, it can influence processes such as cell proliferation, differentiation, and intracellular signaling pathways [9, 13, 14].

Collagens are typically classified into several categories, including fibril-forming, basement membrane-associated, short-chain, and FACIT (fibril-associated collagens with interrupted triple helices) types [11]. These subclasses differ in both structure and function. Fibril-forming collagens constitute the primary tensile framework of the ECM, while FACIT collagens regulate fiber organization. Other collagen types contribute to specialized matrix structures, such as

basement membranes, which are essential for maintaining tissue structure and function [15-18].

Type X collagen is a short-chain, network-structured type of collagen encoded by the *COL10A1* gene [11]. Under physiological conditions, its expression is strictly limited and serves as a marker of hypertrophic chondrocytes during endochondral ossification. During this process, COL10A1 forms a specialized ECM that supports vascular invasion and subsequent bone formation [19]. Although sequencing data indicate relatively high expression in certain non-pathological tissues (such as the gallbladder and ovaries), its levels in most normal tissues remain relatively low [20]. In contrast, COL10A1 is widely overexpressed in various solid tumors [21], suggesting its involvement in oncogenic processes. Evidence from bioinformatic analyses, functional studies, and animal models indicates that COL10A1 functions as a potent oncogene [22]. Its upregulation is not only a consequence of tumor formation but is increasingly regarded as an active driver of tumor malignant progression. Therefore, given its unique expression pattern between normal and diseased states, COL10A1 represents a promising biomarker for cancer diagnosis. This review summarizes the current understanding of COL10A1 in cancer, focusing on its expression patterns, molecular mechanisms, and role in tumor microenvironment remodeling.

### **The biology and pathobiology of type X collagen**

#### *Molecular structure and physiological function*

Type X collagen is a homotrimer composed of three identical  $\alpha 1$  (X) chains, which are encoded by the *COL10A1* gene [23]. Its structure consists of a short central triple-helical collagenous domain, with an N-terminal NC2 domain on one side and a spherical C-terminal NC1 domain on the other [24]. The highly conserved NC1 domain is crucial for initiating and stabilizing trimer assembly [25, 26]. This collagen is specifically produced by hypertrophic chondrocytes in the growth plate [19]. In the process of ECM remodeling, the intact trimeric NC1 domain is secreted into the circulation. This fragment, referred to as the colla-

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**Table 1.** MCDS-related pathogenic mutations in the COL10A1 gene

Mutation position	Variant type	Molecular consequence	Ref.
52	G>A (p.Gly18Arg)	Missense variant	[167]
1293-1296	dup (p.Pro433fs)	Frameshift variant	/
1772	G>T (p.Cys591Phe)	Missense variant	/
1790	A>G (p.Tyr597Cys)	Missense variant	[168]
1833	G>A (p.Trp611Ter)	Nonsense	/
1841	T>C (p.Leu614Pro)	Missense variant	[169]
1853-1866	del (p.Gly618fs)	Frameshift variant	/
1857-1869	del (p.Val621fs)	Frameshift variant	[40]
1858-1859	del (p.Pro620fs)	Frameshift variant	[170]
1859	del (p.Pro620fs)	Frameshift variant	[171]
1884	C>G (p.Tyr628Ter)	Nonsense	[170]
1900	del (p.Asp634fs)	Frameshift variant	/
1945	C>T (p.Gln649Ter)	Nonsense	/
1951-1952	dup (p.Trp651fs)	Frameshift variant	/
1952-153	insA (p.Trp651Ter)	Nonsense	/
1989	C>A (p.Tyr663Ter)	Nonsense	[172]
1989	C>G (p.Tyr663Ter)	Nonsense	[44]

gen X marker (CXM), is a serum biomarker used to evaluate bone growth rate in children [27, 28].

Type X collagen plays a critical role in endochondral skeletal development. It is involved in matrix formation, proteoglycan accumulation, and initiation of mineralization [29-31]. During this process, mesenchymal cells differentiate into chondrocytes, which undergo hypertrophy after proliferation [32]. At this stage, chondrocytes secrete large amounts of type X collagen [33]. Mechanistically, collagen X regulates calcification by binding to matrix vesicles and promoting calcium influx [34, 35]. It also forms a pericellular network that gathers key components (including matrix vesicles and proteoglycans) in the hypertrophic zone [36]. This spatial organization ensures proper initiation and control of mineralization, which is essential for normal ossification [36].

The importance of these functions is supported by genetic models. Early collagen X-null mice exhibited minimal phenotypic abnormalities [37], which might be due to compensatory mechanisms. However, other models demonstrate its importance. Collagen X deficient mice show abnormal matrix distribution and impaired trabecular bone formation [36]. Transgenic

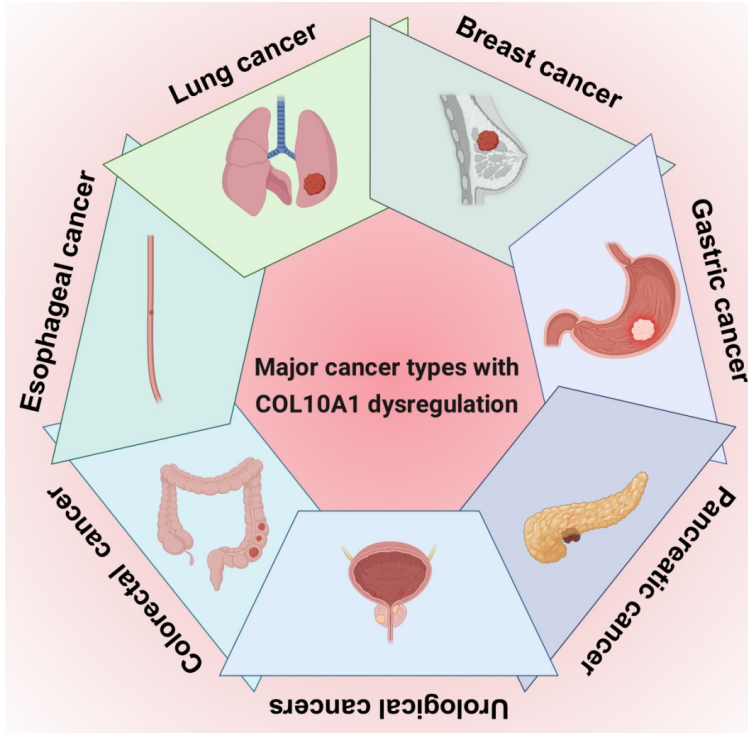
mice expressing mutant collagen X develop skeletal deformities, including growth plate compression and reduced bone formation [38]. Moreover, COL10A1 knockdown in mesenchymal stem cells disrupts ECM synthesis during cartilage differentiation and significantly impairs bone formation in transplantation models [39]. In summary, these studies emphasize the essential role of collagen X in the process of cartilage differentiation and endochondral ossification.

### *Dysregulation and associated diseases*

Metaphyseal chondrodysplasia, Schmid type (MCDS), is an autosomal dominant skeletal disorder caused by COL10A1 mutations [40, 41]. Patients

typically present with short stature, coxa vara, genu varum, and a waddling gait [40, 42]. Most mutations are located in the NC1 region, which is critical for trimerization and stability of collagen X [26, 43]. These mutations, including missense, nonsense, and frameshift variants, often produce misfolded  $\alpha$  chains that accumulate in hypertrophic chondrocytes (Table 1) [44]. This accumulation induces endoplasmic reticulum (ER) stress and activates the unfolded protein response [45], leading to impaired chondrocyte differentiation and degradation of mutant proteins, ultimately resulting in functional haploinsufficiency [46]. In some cases, missense mutations exert dominant-negative effects by incorporating into collagen trimers and disrupting wild-type chain function [47]. These mechanisms disrupt growth plate organization and endochondral ossification, resulting in the characteristic abnormalities of MCDS [48]. Recently, the FDA-approved antiepileptic drug carbamazepine (CBZ) has emerged as a potential therapeutic option for MCDS [49]. Studies show that CBZ promotes clearance of misfolded proteins and alleviates ER stress in model systems, and it has progressed to clinical trials [50-52].

Elucidating the regulatory mechanisms of COL10A1 expression is essential for under-



**Figure 1.** Major cancer types characterized by COL10A1 dysregulation. COL10A1 expression is aberrantly elevated across multiple human malignancies, including breast, gastric, pancreatic, urological, colorectal, esophageal, and lung cancers.

standing chondrocyte hypertrophy. Under physiological conditions, COL10A1 transcription is tightly regulated by multiple factors. HIF-2 $\alpha$  directly binds to hypoxia response elements in its promoter to activate transcription [53], while RUNX2 promotes expression through interaction with cis-regulatory elements [54]. Additional transcription factors, including DLX5, MEF2A, DDX5, and the inhibitory regulator TBX5, also participate in its regulation [55, 56]. Notably, SOX9 suppresses RUNX2 expression and inhibits the transition from proliferative to hypertrophic chondrocytes [57, 58]. However, in the hypertrophic zone, SOX9 cooperates with MEF2C to promote COL10A1 expression [57]. MicroRNAs also contribute to this regulation: miR-218 targets RUNX2, MEF2C, and COL10A1 to inhibit hypertrophic differentiation [59], while miR-26 suppresses COL10A1 production and maintains cartilage matrix stability [60].

COL10A1 and its fragments also have important roles in bone development and joint diseases. The collagen X marker, which corresponds to the trimeric NC1 domain, is re-

leased into circulation during endochondral ossification [61, 62]. CXM serves as a real-time indicator of bone growth and height velocity in children [28, 61], and abnormal levels reflect impaired ossification in conditions such as osteogenesis imperfecta, skeletal dysplasia, achondroplasia, and fracture healing [61, 63-66].

Osteoarthritis (OA) is a chronic degenerative joint disease characterized by progressive cartilage degeneration, leading to pain, stiffness, and functional impairment [67-69]. OA chondrocytes undergo hypertrophic transformation, marked by increased collagen X expression and reduced cartilage-specific markers [70, 71]. This process triggers matrix degradation, pathological calcification, and vascular invasion [72, 73]. Additionally, a collagen X neo-epitope (Col10neo) has been identified as a potential diagnostic biomarker for knee OA [74].

The C-Col10 assay further shows that serum collagen X levels are elevated in early OA and correlate with cartilage degeneration [75].

Targeting aberrant chondrocyte hypertrophy has gained attention as a potential therapeutic approach for osteoarthritis. A number of regulatory pathways involved in this process have been identified. For example, undercarboxylated osteocalcin (ucOCN) has been shown to suppress hypertrophic markers such as COL10A1 and MMP13, while promoting autophagy through the GPRC6A/HIF-1 $\alpha$  pathway [76]. Similarly, the neuronal guidance molecule Sema3A can inhibit hypertrophic changes via PI3K signaling [77].

Emerging evidence also highlights a role for extracellular vesicles in modulating disease progression. Exosomal miR-26b-5p has been reported to reduce chondrocyte hypertrophy and influence macrophage polarization [78]. In parallel, targeting key transcription factors such as DLX5 and RUNX2 can attenuate OA

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**Table 2.** COL10A1 expression and its biological roles in breast and gynecologic tumors

Cancer type	Sample type	Exp.	Research content	Ref.
Breast cancer	Tissue	↑	The upregulated COL10A1 promotes the progression of BRCA and is associated with poor prognosis and disease stage.	[92, 93, 173]
		↑	COL10A1 promotes the malignant progression of BRCA by upregulating the expression of P4HB.	[174]
		↑	COL10A1 promotes the progression of TNBC through the Wnt/ $\beta$ -catenin signaling pathway. Its high expression leads to poorer outcomes in terms of OS and RFS.	[113]
		↑	Increased expression of COL10A1 and low levels of TILs are associated with poor pathological response in ER+/HER2+ breast tumors.	[127]
		↑	COL10A1 increased from pure DCIS to DCIS of DCIS/IBC mixed tumors.	[175]
		↑	Tumors with high expression of COL10A1 may possess higher EMT ability and metastasis potential.	[176]
		↑	COL10A1 regulates EMT and BRCA brain metastasis.	[146]
		↑	The specific expression of COL10A1 in tumors and its localization in blood vessels.	[21]
		↑	COL10A1 directly interacts with ITGB1, activating the PI3K/AKT signaling pathway, and promoting the growth and metastasis of TNBC.	[162]
		/	LncRNA HAGLROS binds to miR-135-3p, thereby alleviating its repressive effect on COL10A1 expression.	[122]
		/	COL10A1 is correlated with LINC01614 and the small molecule inhibitions of TGF- $\beta$ or FAK signaling suppress LINC01614 expression.	[177]
		/	Urchin-Like magnetic nanoparticles loaded with COL10A1 siRNA and stannic for TNBC therapy.	[178]
		Blood	↑	The combination containing COL10A1 can distinguish between BC patients and benign diseases.
Cervical cancer	Tissue	↑	COL10A1 promotes the proliferation, invasion and EMT of cervical cancer by activating the TGF- $\beta$ /SMAD signaling pathway.	[155]

Abbreviations: BRCA, breast cancer; DCIS, ductal carcinoma in situ; IBC, invasive breast cancer; OS, overall survival; RFS, recurrence-free survival; TNBC, triple-negative breast cancer.

progression, at least in part by downregulating COL10A1 expression [79, 80].

Additional signaling axes have also been implicated. Inhibition of the AMPK- $\beta$ -catenin-Runx2 pathway has been shown to reduce downstream matrix metalloproteinase activity [81]. Furthermore, studies examining the role of leptin in age-related OA, along with the identification of aging-associated biomarkers, provide new perspectives on disease mechanisms and potential therapeutic targets [82, 83].

### Dual-platform biomarker: from tissue to liquid biopsy

As the global burden of cancer continues to increase, there is a growing need for methods that enable early detection and real-time disease monitoring [84]. Although tissue biopsy

remains the standard approach for histological diagnosis, its invasive nature and susceptibility to sampling bias limit its suitability for repeated assessment [85].

Liquid biopsy has emerged as an alternative strategy, allowing the detection of tumor-derived components from small volumes of body fluids and enabling longitudinal monitoring [86, 87]. While COL10A1 was originally studied in the context of cartilage development, accumulating evidence indicates that it is upregulated across a range of malignancies (**Figure 1; Tables 2-5**) [22, 88-90]. Notably, COL10A1 can also be detected in circulation, supporting its potential use as a non-invasive serum biomarker [89, 91].

The presence of COL10A1 in both tumor tissues and blood samples suggests that it may

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**Table 3.** COL10A1 expression and its biological roles in digestive system tumors

Cancer type	Sample type	Exp.	Research content	Ref.
Gastric cancer	Tissue	↑	COL10A1 is related to the staging and prognosis of GC.	[95, 96]
		↑	COL10A1 is associated with GC metastasis and reduced survival period and the TGF-β1-SOX9 axis induces the expression of COL10A1.	[114]
		↑	Overexpression of COL10A1 promotes the invasiveness of GC through EMT.	[145]
		↑	Knockdown of COL10A1 inhibits the proliferation and migration of GC cells while promoting their apoptosis.	[179]
		↑	COL10A1 is identified as a tumor-specific ECM protein through the decellularization technology.	[135]
		/	The N110T and H165R mutants of SOX9 promote tumor progression by enhancing the binding affinity of the COL10A1-SOX9 complex.	[180]
		/	COL10A1 is a potential mRNA vaccine candidate for STAD.	[181]
	/	COL10A1 is one of the hub genes in the circRNA-miRNA-mRNA regulatory network and overexpression of COL10A1 is associated with a lower OS rate in STAD.	[182]	
	Blood	↑	Elevated plasma levels of COL10A1 are associated with poor survival in patients.	[89]
		↑	High expression of COL10A1 is associated with gastrointestinal tumorigenesis and can be detected in the blood.	[183]
↑		The inclusion of COL10A1 in a panel of biomarkers enhances the early diagnosis of gastric cancer and improves the distinction between benign and malignant gastric diseases.	[109]	
Colorectal cancer	Tissue	↑	High expression of COL10A1 is an independent risk factor for prognosis and overall survival in CRC patients.	[98]
		↑	COL10A1 is a potential diagnostic biomarker associated with deficient mismatch repair and immune infiltration in colon cancer.	[125]
		↑	The abundance of COL10A1 in CRC tissues predicts metastatic and immunogenic properties and it may mediate the interaction between cancer cells and TME.	[99]
		/	COL10A1 is upregulated in FAP-positive CAFs.	[184]
		/	The COL10A1+Fib subpopulation is associated with the progression of colorectal cancer and poor prognosis of patients.	[118]
	/	SYNPO2L promotes the secretion of COL10A1 and the infiltration of tumor-associated fibroblasts, thereby facilitating EMT.	[185]	
	Blood	↑	Serum COL10A1 serve as a potential diagnostic candidate for detecting adenomatous lesions and colorectal neoplasms.	[106]
Pancreatic cancer	Tissue	↑	Patients with high COL10A1 expression exhibits worse RFS and OS.	[107]
		↑	COL10A1 promotes pancreatic adenocarcinoma tumorigenesis by regulating CD276.	[100]
Esophageal cancer	Tissue	↑	Serum COL10A1 has high sensitivity and specificity for the diagnosis of PDAC.	[107]
		↑	COL10A1 is a potential diagnostic and prognostic biomarker in ESCC.	[186]
		↑	The potential of COL10A1 as a molecular biomarker for early diagnosis of ESCC.	[187]
/	COL10A1Var1 is a novel and recurrent transcript variant associated with Esophageal Adenocarcinoma, and it has potential tumor-promoting functions.	[188]		
Hepatocellular carcinoma	Tissue	↑	COL10A1 serves as a diagnostic biomarker for differentiating MASLD from MASL-HCC.	[189]
Cholangiocarcinoma	Tissue	↓	COL10A1 is a down-regulated differentially expressed gene.	[190]

Abbreviations: CRC, colorectal cancer; ESCC, esophageal squamous cell carcinoma; GC, gastric cancer; HCC, hepatocellular carcinoma; MASLD, metabolic dysfunction-associated steatotic liver disease; PDAC, pancreatic ductal adenocarcinoma; STAD, stomach adenocarcinoma.

provide clinically relevant information on tumor progression and could assist in guiding patient.

### *Prognostic indicator across multiple tumors*

COL10A1 is highly expressed in various cancers and is associated with poor prognosis. In

breast cancer (**Table 2**), its expression is particularly elevated in aggressive subtypes, including invasive lobular and ductal carcinoma [21, 92]. Tissue microarray-based immunohistochemistry further demonstrates that high COL10A1 expression correlates with advanced stage and unfavorable outcomes [92, 93]. In

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**Table 4.** COL10A1 expression and its biological roles in tumors of respiratory and urinary systems

Cancer type	Sample type	Exp.	Research content	Ref.
Lung Cancer	Tissue	↑	Potential diagnostic value of COL10A1 in IPF and NSCLC.	[191]
		↑	miR-384 reduces the level of COL10A1, thereby inhibiting cell proliferation and promoting apoptosis and autophagy in NSCLC cells.	[192]
		↑	COL10A1 is identified as a key hub gene linking ECM remodeling and tumor progression across COPD, LUAD, and LUSC.	[193]
		↑	The COL10A1/DDR2/FAK axis regulates the proliferation and metastasis of LUAD cells.	[101]
		/	COL10A1 secreted by CAFs promotes LUSC cell proliferation and suppresses apoptosis induced by oxidative stress through METTL3-mediated m <sup>6</sup> A modification of its mRNA, thereby accelerating Tumor growth.	[194]
	Blood	↑	COL10A1 levels are elevated in lung cancer patients compared to healthy heavy smokers.	[91]
Nasopharyngeal carcinoma	Tissue	↑	COL10A1 may serve as a prognostic and diagnostic marker for nasopharyngeal carcinoma.	[195]
Prostate Cancer	Tissue	↑	COL10A1 is related to tumor prognosis and tumor prediction.	[102, 123]
		↑	COL10A1 directly interacts with INHBA and activates the PI3K/AKT signaling pathway.	[164]
		/	COL10A1 may be involved in predicting bone metastasis of prostate cancer.	[196]
		/	COL10A1 is involved in predicting the biochemical recurrence of prostate cancer patients.	[197]
Bladder cancer	Tissue	↑	COL10A1 is upregulated in the BLCA samples, and increased expression of COL10A1 is associated with a lower overall survival rate.	[100]
		↑	COL10A1 is one of the markers for preoperative prediction of lymph node metastasis in BLCA.	[198]

Abbreviations: BLCA, bladder cancer; COPD, chronic obstructive pulmonary disease; IPF, idiopathic pulmonary fibrosis; LUAD, lung adenocarcinoma; LUSC, lung squamous cell carcinoma; NSCLC, non-small cell lung cancer; OS, overall survival.

basal cell carcinoma (BCC), COL10A1 is not uniformly expressed across all high-risk subtypes but is enriched in the stroma of sclerotic/infiltrative and basosquamous variants [94], suggesting its value in subtype classification and prognosis.

Similar patterns are observed in digestive and urologic malignancies. In gastric cancer (**Table 3**), COL10A1 expression is higher in tumor tissues than in adjacent normal tissues [95] and is associated with lymph node metastasis and reduced overall survival [89, 95]. It is also linked to immune infiltration within the tumor microenvironment [96, 97]. In colorectal cancer, elevated COL10A1 predicts poorer overall and disease-free survival [98, 99]. Likewise, in bladder cancer, its upregulation is associated with shorter survival, supporting its role as a prognostic biomarker [100].

COL10A1 also shows prognostic significance in other adenocarcinomas. In lung adenocarcinoma (**Table 4**), its expression is elevated and

positively correlates with lymph node metastasis, serving as an independent predictor of poor prognosis [101]. In prostate cancer, COL10A1 expression is increased and associated with clinical features, demonstrating both diagnostic and prognostic value [102]. In pancreatic cancer, COL10A1 overexpression correlates with advanced clinicopathological features, supporting its potential as an early diagnostic and prognostic marker [103]. Additionally, COL10A1 is upregulated in laryngeal squamous cell carcinoma and oral cancer (**Table 5**) [104, 105].

### *Serum biomarker in tumors*

Circulating COL10A1 has gained attention as a potential biomarker across multiple cancer types. Its presence in blood, together with its association with disease progression, makes it an attractive candidate for minimally invasive and repeatable monitoring. In gastric cancer, elevated circulating COL10A1 levels have been linked to poorer survival outcomes, suggesting

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**Table 5.** COL10A1 expression and its biological roles in other carcinomas and sarcomas

Cancer type	Sample type	Exp.	Research content	Ref.
Basal cell carcinoma	Tissue	↑	COL10A1 is a marker for high-risk basal cell carcinoma.	[94]
Laryngeal squamous cell carcinoma	Tissue	↑	COL10A1 may serve as a potential drug target for Laryngeal squamous cell carcinoma.	[104]
Oral cancer	Tissue	↑	The exosomes derived from hBMSCs contain miR-101-3p, which targets COL10A1 and inhibits the progression of oral cancer.	[105]
Thyroid carcinoma	Tissue	/	COL10A1 may serve as a novel candidate diagnostic marker for TCPTC.	[199]
Osteosarcoma	Tissue	/	COL10A1 is a potential procollagen substrate of PCOLCE and is positively correlated with TWIST1, a key molecule in metastasis.	[200]
		/	LncRNA TTTY14 may influence the development of osteosarcoma by co-expressing with COL10A1.	[201]
Chondrosarcoma	Tissue	/	COL10A1 expression is higher in both Chondrosarcoma tissue and 3D cultures than in 2D cultures.	[202]
		/	Depsipeptide inhibits the growth of chondrosarcoma cells by inducing cell cycle arrest and/or apoptosis. Long-term treatment with low-dose depsipeptide leads to differentiation into a hypertrophic phenotype, characterized by increased COL10A1 expression in chondrosarcoma cells.	[203]

Abbreviations: TCPTC, tall cell variant papillary thyroid carcinoma.

potential value for both early detection and prognostic assessment [89]. A similar trend has been reported in colorectal disease, where increased serum COL10A1 can distinguish patients with colon cancer or adenomas from cancer-free individuals, supporting its role in early diagnosis [106].

In pancreatic ductal adenocarcinoma, COL10A1 overexpression has been observed in both tumor tissues and plasma, with higher levels correlating with disease progression and reduced survival, indicating both diagnostic and prognostic relevance [107]. In non-small cell lung cancer, plasma COL10A1 concentrations are elevated compared with those in healthy heavy smokers [91], although current evidence does not show a clear relationship with clinical stage or survival [91].

Despite these findings, the diagnostic performance of individual circulating biomarkers remains limited. To improve accuracy, combined biomarker approaches have been explored. For instance, integrating COL10A1 with COL11A1 and COMP significantly enhances the ability to distinguish breast cancer from benign breast conditions [108]. Similarly, multi-marker models incorporating COL10A1 have shown improved performance in early detec-

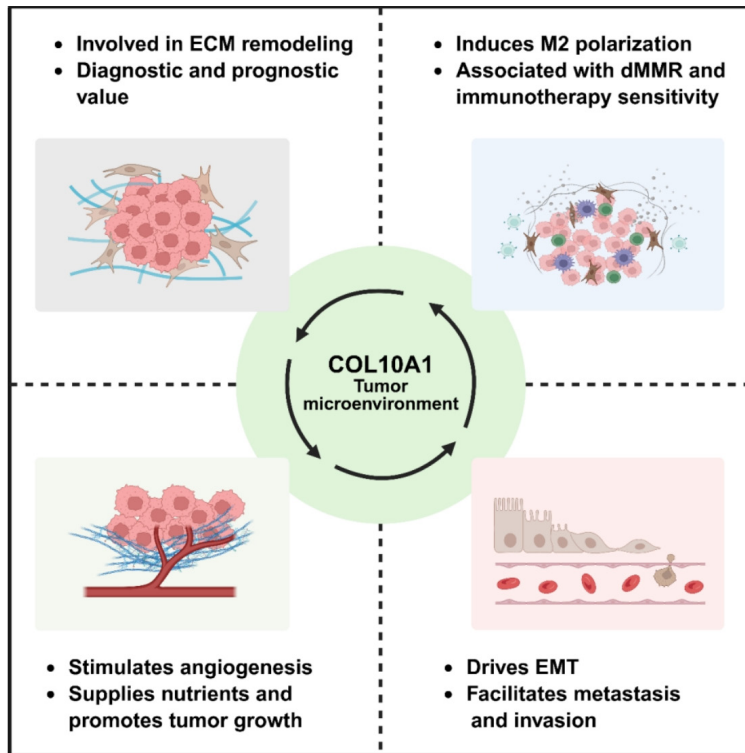
tion and in differentiating malignant from benign tumors [109].

The consistent elevation of circulating COL10A1 across different malignancies suggests that common underlying processes may be involved. Increased tumor expression, together with active extracellular matrix remodeling, likely contributes to the release of collagen X fragments into the bloodstream. In this context, circulating COL10A1 may serve as an indirect indicator of ECM turnover in cancer, analogous to its role in skeletal development.

### The emerging role of COL10A1 in the tumor microenvironment

The tumor microenvironment (TME) regulates cancer development, therapeutic response, and patient prognosis through complex interactions between cells and molecules [110, 111]. The extracellular matrix has evolved from a passive scaffold into an active regulator of tumor progression, engaging in bidirectional crosstalk with malignant and stromal cells [112]. In this context, short-chain collagen type X has become an important functional component of the TME. In various cancers, the abnormal expression of COL10A1 is not only a consequence of tumor growth but also a driving factor

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**Figure 2.** The emerging role of COL10A1 in the tumor microenvironment. COL10A1 participates in extracellular matrix (ECM) remodeling and functions as a dual biomarker in both tissue and serum. It promotes tumor progression by inducing epithelial-mesenchymal transition (EMT), angiogenesis, metastasis, and M2 macrophage polarization.

for malignancy. COL10A1 promotes a tumor-supportive environment by regulating cancer-associated fibroblasts (CAFs), immune cell infiltration, and ECM architecture [99, 101]. In addition, it enhances tumor progression by inducing epithelial-mesenchymal transition (EMT) and stimulating angiogenesis (Figure 2) [113, 114].

### *Cancer-associated fibroblasts*

Cancer-associated fibroblasts (CAFs) are a heterogeneous population that regulates tumor growth, invasion, and therapeutic resistance [115, 116]. Multi-omics analyses have identified a distinct CAF subpopulation characterized by COL10A1 expression [90, 117]. These "COL10A1-positive fibroblasts" are associated with colorectal cancer progression and poor prognosis [118]. Mechanistically, COL10A1 directly induces EMT in tumor cells and promotes macrophage polarization toward the M2 phenotype through activation of the CD18/JAK1/STAT3 axis [118]. Importantly, M2 ma-

crophages further enhance COL10A1 expression in fibroblasts via the TGF- $\beta$ /RUNX2 pathway, forming a positive feedback loop that sustains tumor progression [118].

The role of COL10A1 in CAF biology extends beyond gastrointestinal cancers. In basal cell carcinoma, COL10A1 is highly expressed in the stroma of high-risk subtypes and is associated with a specific subset of ECM-producing fibroblasts adjacent to invasive tumors [94]. This CAF population exhibits strong ECM remodeling activity, suggesting that COL10A1 may serve as a potential therapeutic target in high-risk BCC [94].

Similarly, in breast cancer, particularly the triple-negative subtype, COL10A1 is part of a CAF-associated gene signature. The CAF+ subtype, enriched with COL10A1, is associated with worse overall survival and an immunosuppressive

TME [117]. In pancreatic ductal adenocarcinoma (PDAC), COL10A1 is mainly expressed in myofibroblast CAFs (myCAFs) [90], which are characterized by high  $\alpha$ -SMA and low interleukin-6 (IL-6) expression and are linked to metastasis and immunosuppression [119]. This non-canonical collagen profile is deregulated across multiple cancers and is frequently associated with poor prognosis [90].

### *Immune microenvironment*

The immune landscape of the tumor microenvironment (TME) is a key determinant of tumor progression, although many tumors develop mechanisms to evade immune surveillance and dampen cytotoxic responses [120]. Interactions between cancer-associated fibroblasts (CAFs) and immune cells contribute to the establishment of an immunosuppressive milieu [121]. In this context, COL10A1 expression has been linked to specific patterns of immune infiltration, particularly the polarization of macrophages toward the M2 phenotype [102, 118].

Experimental evidence indicates that COL10A1-positive fibroblasts can promote the polarization of M2-type macrophages, which in turn may reinforce COL10A1 expression through a feedback mechanism within the microenvironment [118]. Additional regulatory pathways have also been implicated. For instance, the lncRNA HAGLROS has been shown to promote the development of breast cancer through the miR-135b-3p/COL10A1 axis while simultaneously promoting M2 polarization, although the exact contribution of COL10A1 remains to be fully defined [122].

Similar correlations have been observed in different types of tumors. For instance, in prostate cancer, elevated COL10A1 expression is associated with increased infiltration of M2-type macrophages, indicating its conserved role in shaping an immunosuppressive environment [102, 123]. In bladder cancer, COL10A1 expression is associated with broader immune cell infiltration and may influence immune cell recruitment and function [100]. A prognostic model based on macrophages has shown predictive value for patient outcomes [124]. In gastric cancer, COL10A1 levels correlate with infiltration by multiple immune cell populations, including CD8<sup>+</sup> T cells, cytotoxic lymphocytes, regulatory T cells, and natural killer cells [96].

The potential role of COL10A1 in the response to immunotherapy is receiving increasing attention. In colorectal cancer, its expression has been associated with immune infiltration and deficient mismatch repair status [125], a feature linked to responsiveness to immune checkpoint blockade therapy [125, 126]. Some studies also suggest that higher COL10A1 levels may be associated with increased sensitivity to anti-PD-1 therapy [125]. In contrast, in ER<sup>+</sup>/HER2<sup>+</sup> breast cancer, elevated COL10A1 expression combined with low levels of tumor-infiltrating lymphocytes (TILs) has been associated with a reduced response to neoadjuvant treatment [127]. Pan-cancer analyses further indicate associations between COL10A1 and genomic features such as tumor mutational burden (TMB) and microsatellite instability (MSI) [123].

### *Extracellular matrix remodeling*

In addition to providing structural support, the extracellular matrix also influences tumor

behavior through both biochemical signaling and mechanical properties [128, 129]. The continuous remodeling of the ECM is a significant feature of tumor progression and plays a central role in facilitating metastasis, making it an attractive therapeutic target [130]. In this setting, COL10A1 is considered an active contributor to ECM reorganization.

In lung adenocarcinoma (LUAD), elevated COL10A1 expression has been linked to ECM remodeling, which is achieved by activating the COL10A1/DDR2/FAK signaling axis. This signaling axis can promote tumor cell proliferation and invasion [101]. Mechanistically, COL10A1 can function as a ligand for discoidin domain receptor 2 (DDR2), a collagen-activated receptor tyrosine kinase located at the cell-matrix interface [131, 132]. Binding to DDR2 leads to downstream activation of focal adhesion kinase (FAK), which is a key regulator of cell adhesion and signaling and is also associated with immune modulation and enhanced responses to combination immunotherapy strategies [133].

Integrin-mediated interactions further promote this process by coordinating cell-ECM communication and maintaining tissue integrity [134]. Supporting these findings, decellularization studies have identified COL10A1 as a tumor-associated ECM component that is largely absent from adjacent normal tissues [135]. In colorectal cancer, its expression correlates with CAF-related transcriptional programs, highlighting its role in mediating tumor-stroma interactions [99].

Overall, these observations suggest that COL10A1 functions not only as a structural ECM protein but also as a regulator of matrix remodeling, contributing to the formation of a microenvironment that supports tumor invasion and progression.

### *Tumor angiogenesis*

The ECM plays a central role in all stages of angiogenesis [136]. Its remodeling regulates vessel formation, pericyte recruitment, endothelial basement membrane assembly, and the availability of angiogenic factors [136]. In tumors, this process drives pathological angiogenesis, which supports tumor growth and

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metastasis [137, 138]. COL10A1 is specifically enriched in tumor vasculature and is largely absent in normal vessels [21]. Its association with angiogenesis is also observed in endochondral ossification, osteoarthritis-related neovascularization, and age-related macular degeneration [32, 139-141]. These findings suggest that COL10A1 functions as both a structural and signaling regulator of angiogenesis. In triple-negative breast cancer, COL10A1 promotes angiogenesis through paracrine mechanisms, as conditioned medium from COL10A1-overexpressing cells induces denser and more complex endothelial tubular networks with increased junctions and tube length [113].

### *Epithelial-mesenchymal transition*

Epithelial-mesenchymal transition (EMT) enables epithelial cells to acquire mesenchymal characteristics [142]. In cancer, EMT promotes invasion, metastasis, and therapy resistance [143, 144]. Among ECM components, COL10A1 not only induces EMT-associated phenotypic changes but also links ECM remodeling to tumor progression. In gastric cancer, the TGF- $\beta$ 1/SOX9/COL10A1 axis acts as a key inducer of EMT and is strongly associated with patient survival [114]. Mechanistically, TGF- $\beta$ 1 increases Smad2 phosphorylation and upregulates SOX9 and COL10A1 expression, while SOX9 directly activates COL10A1 transcription [114]. COL10A1 overexpression promotes EMT by upregulating mesenchymal markers (N-cadherin, vimentin, Snail/Slug) and downregulating epithelial markers (E-cadherin,  $\beta$ -catenin) [145]. In colorectal cancer, COL10A1+CAFs promote EMT through COL10A1 secretion [118]. Similarly, COL10A1 enhances EMT via activation of the Wnt/ $\beta$ -catenin pathway in triple-negative breast cancer [113]. Knockout studies further show that COL10A1 promotes breast cancer cell invasiveness and may facilitate penetration of the blood-brain barrier, contributing to intracranial metastasis [146]. Overall, COL10A1 promotes EMT and enhances tumor cell proliferation, migration, and invasion, suggesting that it may serve as a key molecular link between local invasion and distant metastasis, as well as a potential therapeutic target.

### **Signaling mechanisms of COL10A1 in cancer**

In cancer, the TGF- $\beta$ /Smad signaling pathway promotes COL10A1 expression. Elevated

COL10A1 further drives tumor progression through two major signaling axes: the COL10A1/DDR2/FAK axis, which promotes EMT and metastasis, and the COL10A1/ITGB1/PI3K/AKT axis, which enhances tumor cell proliferation and invasion (**Figure 3**).

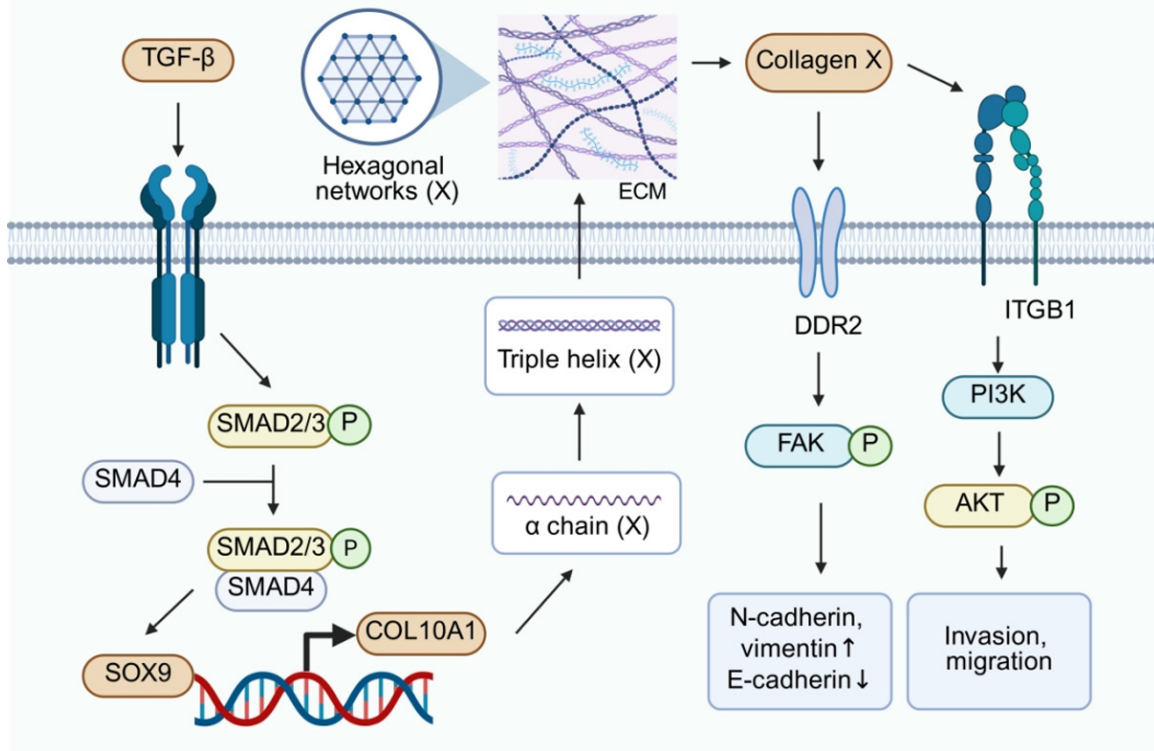
### *TGF- $\beta$ /Smad signaling pathway*

Transforming growth factor- $\beta$  (TGF- $\beta$ ) signaling is critical for embryonic development, tissue homeostasis, and tumor progression [147, 148]. In advanced cancer, this pathway often shifts from a tumor suppressor to a metastasis-promoting factor [149]. Its oncogenic effects involve EMT induction, abnormal ECM deposition, and CAF activation [147, 150]. Binding of TGF- $\beta$  to its cell surface receptors activates the canonical TGF- $\beta$ /Smad pathway [151], triggering a phosphorylation cascade that activates Smad2 and Smad3 [151]. These proteins form a complex with Smad4, translocate into the nucleus, and regulate downstream gene transcription [151, 152]. Within this network, COL10A1 has been identified as a key ECM component [135] and a downstream effector of TGF- $\beta$ 1 signaling [114]. Mechanistically, TGF- $\beta$ 1 enhances Smad2 phosphorylation, leading to increased expression of the transcription factor SOX9 [114]. SOX9 then directly binds to the COL10A1 promoter to activate its transcription [114]. The role of TGF- $\beta$  signaling in EMT regulation is well established [150, 153, 154]. In gastric cancer, TGF- $\beta$ 1 reduces epithelial marker E-cadherin while increasing the mesenchymal marker vimentin and activating transcription factors such as Snail and Slug, thereby enhancing migration and invasion [114]. Notably, COL10A1 knockdown attenuates TGF- $\beta$ 1-induced EMT [114]. In addition, COL10A1 knockdown in cervical cancer cells reduces TGF- $\beta$ 1 levels and decreases phosphorylation of Smad2 and Smad3 [155]. Overall, TGF- $\beta$ /Smad signaling promotes tumor progression partly through COL10A1, which is essential for EMT induction and maintenance of a tumor-permissive microenvironment.

### *COL10A1/DDR2/FAK signaling axis*

Discoidin domain receptors (DDR1 and DDR2) are a class of receptor tyrosine kinases activated by collagen binding in the extracellular matrix [156]. DDR2 preferentially binds specific

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**Figure 3.** Signaling mechanisms of COL10A1 in cancer. The TGF- $\beta$ /SMAD signaling pathway activates SOX9, which in turn promotes COL10A1 expression. Three identical type X  $\alpha$  chains form a triple helix that assembles into hexagonal networks within the extracellular matrix (ECM). COL10A1 binds to DDR2 to activate FAK and interacts with ITGB1 to stimulate the PI3K/AKT pathway. These signaling events induce epithelial-mesenchymal transition (EMT) and enhance tumor cell invasion and migration.

collagens, including type II and type X (COL10A1) [132]. Functional studies have identified focal adhesion kinase (FAK) as a key downstream effector of the COL10A1/DDR2 axis [101]. FAK is a non-receptor tyrosine kinase that regulates adhesion signaling, cell migration, and angiogenesis [133, 157]. In lung cancer cells, COL10A1 binds DDR2 and induces its phosphorylation (p-DDR2), which subsequently activates FAK phosphorylation (p-FAK) and downstream signaling [101].

The COL10A1/DDR2 axis regulates FAK through a dual mechanism, promoting both its phosphorylation and basal expression. COL10A1 expression positively correlates with FAK protein levels, as its upregulation increases FAK expression, whereas downregulation reduces FAK levels [101]. Activated FAK promotes EMT, characterized by decreased E-cadherin and increased N-cadherin and vimentin, thereby enhancing tumor cell migration and invasion [101]. Functional studies further indicate that

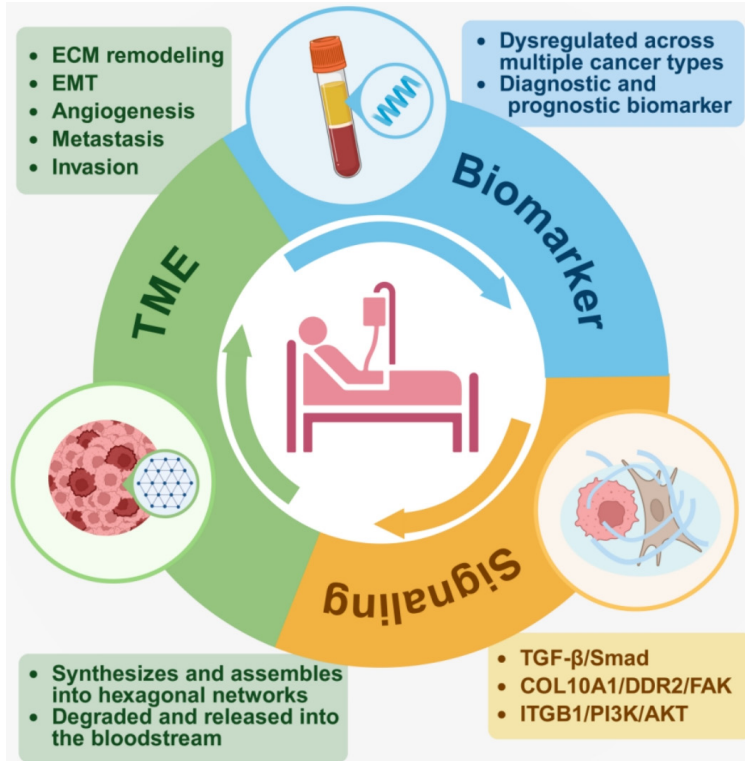
the migration-promoting effect of COL10A1 depends on FAK signaling, as FAK inhibition reverses this effect [101]. Thus, targeting the COL10A1/DDR2/FAK axis, particularly with DDR2 or FAK inhibitors, may represent a promising anticancer strategy.

### *ITGB1/PI3K/AKT signaling axis*

Integrins are heterodimeric transmembrane receptors composed of  $\alpha$  and  $\beta$  subunits that connect the extracellular matrix to the cytoskeleton and mediate both cell adhesion and signal transduction [158, 159]. Among them, integrin  $\beta$ 1 (ITGB1) plays an important role in sensing extracellular mechanical cues and has been implicated in processes such as angiogenesis, neuronal development, and tumor progression [160].

The PI3K/AKT pathway is a central regulator of malignant phenotypes, including cell survival, proliferation, and invasion, primarily through

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**Figure 4.** Functional and regulatory landscape of COL10A1 in cancer. COL10A1 promotes tumor progression by inducing ECM remodeling, EMT, and angiogenesis. COL10A1 is synthesized and assembled into hexagonal networks within the ECM, and its degradation products can be released into the circulation. COL10A1 is dysregulated in multiple cancers and functions as a dual tissue and serum biomarker for diagnosis and prognosis. In addition, COL10A1 drives tumor progression through the TGF- $\beta$ /Smad, COL10A1/DDR2/FAK, and ITGB1/PI3K/AKT signaling axes.

activation of AKT by phosphorylation [161]. Evidence suggests that COL10A1 can interact directly with ITGB1, leading to activation of its intracellular signaling domain and subsequent recruitment of PI3K [162, 163]. Activated PI3K generates PIP3, which facilitates the localization of AKT to the plasma membrane and promotes its phosphorylation, thereby initiating downstream signaling events [162].

Consistent with this mechanism, COL10A1 expression has been shown to correlate positively with AKT phosphorylation levels. Overexpression of COL10A1 enhances cancer cell proliferation, migration, and invasion, whereas these effects can be attenuated by PI3K/AKT inhibitors or by silencing ITGB1 [162]. Similar inhibitory effects are also observed in COL10A1-deficient cells, further supporting the functional link between COL10A1 and this pathway [162].

In addition, COL10A1 has been reported to interact with INHBA in prostate cancer, contributing to activation of PI3K/AKT signaling [164]. INHBA, a member of the TGF- $\beta$  superfamily, is known to promote tumor progression [165]. Its suppression inhibits TGF- $\beta$  signaling and reduces tumor cell migration and invasion [165, 166].

Overall, these findings suggest that activation of the PI3K/AKT pathway is a major mechanism underlying the tumor-promoting effects of COL10A1. Disrupting COL10A1-mediated signaling or targeting downstream components of this pathway may therefore represent a potential therapeutic approach.

### Conclusion and future perspectives

The extracellular matrix protein COL10A1 shows elevated expression in various solid tumors, and its expression is strongly associated with poor patient prognosis, highlighting its potential as both a histopathological and circulating biomarker for diagnosis and prognostic assessment (Figure 4) [21].

Currently, many findings regarding COL10A1 are derived from tumor transcriptomic analyses based on public databases. Therefore, the clinical translational potential of COL10A1 as a tissue or circulating biomarker still requires validation through large-scale prospective clinical studies. In addition, the mechanisms linking dynamic changes in circulating COL10A1 (or its fragments) with tumor burden, disease progression, and treatment response remain insufficiently understood. Further investigation into these relationships will be essential to facilitate the clinical translation of COL10A1 as a standardized liquid biopsy biomarker.

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#### Disclosure of conflict of interest

None.

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#### References

- [1] Huang J, Zhang L, Wan D, Zhou L, Zheng S, Lin S and Qiao Y. Extracellular matrix and its therapeutic potential for cancer treatment. *Signal Transduct Target Ther* 2021; 6: 153.
- [2] Hynes RO. The extracellular matrix: not just pretty fibrils. *Science* 2009; 326: 1216-1219.
- [3] Theocharis AD, Skandalis SS, Gialeli C and Karamanos NK. Extracellular matrix structure. *Adv Drug Deliv Rev* 2016; 97: 4-27.
- [4] Bonnans C, Chou J and Werb Z. Remodelling the extracellular matrix in development and disease. *Nat Rev Mol Cell Biol* 2014; 15: 786-801.
- [5] Carnazza M, Quaranto D, DeSouza N, Li XM, Tiwari RK, Di Martino JS and Geliebter J. The duality of collagens in metastases of solid tumors. *Int J Mol Sci* 2025; 26: 9745.
- [6] Winkler J, Abisoye-Ogunniyan A, Metcalf KJ and Werb Z. Concepts of extracellular matrix remodelling in tumour progression and metastasis. *Nat Commun* 2020; 11: 5120.
- [7] Ding DY, Jiang SY, Zu YX, Yang Y, Gan XJ, Yuan SX and Zhou WP. Collagen in hepatocellular carcinoma: a novel biomarker and therapeutic target. *Hepatol Commun* 2024; 8: e0489.
- [8] Slatter DA, Bihan DG and Farndale RW. The effect of purity upon the triple-helical stability of collagenous peptides. *Biomaterials* 2011; 32: 6621-6632.
- [9] Siadat SM and Ruberti JW. Mechanochemistry of collagen. *Acta Biomater* 2023; 163: 50-62.
- [10] Shoulders MD and Raines RT. Collagen structure and stability. *Annu Rev Biochem* 2009; 78: 929-958.
- [11] Ricard-Blum S. The collagen family. *Cold Spring Harb Perspect Biol* 2011; 3: a004978.
- [12] Fiala T, Barros EP, Heeb R, Riniker S and Wennemers H. Predicting collagen triple helix stability through additive effects of terminal residues and caps. *Angew Chem Int Ed Engl* 2023; 62: e202214728.
- [13] Kirkness MW, Lehmann K and Forde NR. Mechanics and structural stability of the collagen triple helix. *Curr Opin Chem Biol* 2019; 53: 98-105.
- [14] Leitinger B. Transmembrane collagen receptors. *Annu Rev Cell Dev Biol* 2011; 27: 265-290.
- [15] Kadler K. Extracellular matrix 1: Fibril-forming collagens. *Protein Profile* 1995; 2: 491-619.
- [16] Shaw LM and Olsen BR. FACIT collagens: diverse molecular bridges in extracellular matrices. *Trends Biochem Sci* 1991; 16: 191-194.
- [17] Heino J. The collagen family members as cell adhesion proteins. *Bioessays* 2007; 29: 1001-1010.
- [18] Gelse K, Pöschl E and Aigner T. Collagens-structure, function, and biosynthesis. *Adv Drug Deliv Rev* 2003; 55: 1531-1546.
- [19] Long F and Ornitz DM. Development of the endochondral skeleton. *Cold Spring Harb Perspect Biol* 2013; 5: a008334.
- [20] Fagerberg L, Hallström BM, Oksvold P, Kampf C, Djureinovic D, Odeberg J, Habuka M, Tahmasebpoor S, Danielsson A, Edlund K, Asplund A, Sjöstedt E, Lundberg E, Szilvarty CA, Skogs M, Takanen JO, Berling H, Tegel H, Mulder J, Nilsson P, Schwenk JM, Lindskog C, Danielsson F, Mardinoglu A, Sivertsson A, von Feilitzen K, Forsberg M, Zwahlen M, Olsson I, Navani S, Huss M, Nielsen J, Ponten F and Uhlén M. Analysis of the human tissue-specific expression by genome-wide integration of transcriptomics and antibody-based proteomics. *Mol Cell Proteomics* 2014; 13: 397-406.
- [21] Chapman KB, Prendes MJ, Sternberg H, Kidd JL, Funk WD, Wagner J and West MD. COL10A1 expression is elevated in diverse solid tumor types and is associated with tumor vasculature. *Future Oncol* 2012; 8: 1031-1040.
- [22] Yi Q, Zhu G, Zhu W, Wang J, Ouyang X, Yang K and Zhong J. Oncogenic mechanisms of COL10A1 in cancer and clinical challenges (Review). *Oncol Rep* 2024; 52: 162.
- [23] Suttmüller M, Buijij JA and de Heer E. Collagen types VIII and X, two non-fibrillar, short-chain collagens. Structure homologies, functions and involvement in pathology. *Histol Histopathol* 1997; 12: 557-566.
- [24] Chan D, Weng YM, Hocking AM, Golub S, McQuillan DJ and Bateman JF. Site-directed mutagenesis of human type X collagen. Expression of alpha1(X) NC1, NC2, and helical mutations in vitro and in transfected cells. *J Biol Chem* 1996; 271: 13566-13572.

## COL10A1: emerging biomarker and TME regulator in solid cancers

- [25] Barber RE and Kwan AP. Partial characterization of the C-terminal non-collagenous domain (NC1) of collagen type X. *Biochem J* 1996; 320: 479-485.
- [26] Bateman JF, Freddi S, McNeil R, Thompson E, Hermanns P, Savarirayan R and Lamandé SR. Identification of four novel COL10A1 missense mutations in schmid metaphyseal chondrodysplasia: further evidence that collagen X NC1 mutations impair trimer assembly. *Hum Mutat* 2004; 23: 396.
- [27] Hauta-Alus HH, Holmlund-Suila EM, Valkama SM, Enlund-Cerullo M, Rosendahl J, Coghlan RF, Andersson S and Mäkitie O. Collagen X Biomarker (CXM), linear growth, and bone development in a Vitamin D intervention study in infants. *J Bone Miner Res* 2022; 37: 1653-1664.
- [28] Coghlan RF, Olney RC, Boston BA, Coleman DT, Johnstone B and Horton WA. Norms for clinical use of CXM, a real-time marker of height velocity. *J Clin Endocrinol Metab* 2021; 106: e255-e264.
- [29] Anderson HC. Mechanism of mineral formation in bone. *Lab Invest* 1989; 60: 320-330.
- [30] Kirsch T and Pfäffle M. Selective binding of anchoring CII (annexin V) to type II and X collagen and to chondrocalcin (C-propeptide of type II collagen). Implications for anchoring function between matrix vesicles and matrix proteins. *FEBS Lett* 1992; 310: 143-147.
- [31] Chen Q, Linsenmayer C, Gu H, Schmid TM and Linsenmayer TF. Domains of type X collagen: alteration of cartilage matrix by fibril association and proteoglycan accumulation. *J Cell Biol* 1992; 117: 687-694.
- [32] Rolian C. Endochondral ossification and the evolution of limb proportions. *Wiley Interdiscip Rev Dev Biol* 2020; 9: e373.
- [33] Shen G. The role of type X collagen in facilitating and regulating endochondral ossification of articular cartilage. *Orthod Craniofac Res* 2005; 8: 11-17.
- [34] Anderson HC. Molecular biology of matrix vesicles. *Clin Orthop Relat Res* 1995; 266-280.
- [35] Kirsch T and Wuthier RE. Stimulation of calcification of growth plate cartilage matrix vesicles by binding to type II and X collagens. *J Biol Chem* 1994; 269: 11462-11469.
- [36] Kwan KM, Pang MK, Zhou S, Cowan SK, Kong RY, Pfordte T, Olsen BR, Sillence DO, Tam PP and Cheah KS. Abnormal compartmentalization of cartilage matrix components in mice lacking collagen X: implications for function. *J Cell Biol* 1997; 136: 459-471.
- [37] Rosati R, Horan GS, Pinero GJ, Garofalo S, Keene DR, Horton WA, Vuorio E, de Crombrughe B and Behringer RR. Normal long bone growth and development in type X collagen-null mice. *Nat Genet* 1994; 8: 129-135.
- [38] Jacenko O, LuValle PA and Olsen BR. Spondylo-metaphyseal dysplasia in mice carrying a dominant negative mutation in a matrix protein specific for cartilage-to-bone transition. *Nature* 1993; 365: 56-61.
- [39] Knuth CA, Andres Sastre E, Fahy NB, Witte-Bouma J, Ridwan Y, Strabbing EM, Koudstaal MJ, van de Peppel J, Wolvius EB, Narcisi R and Farrell E. Collagen type X is essential for successful mesenchymal stem cell-mediated cartilage formation and subsequent endochondral ossification. *Eur Cell Mater* 2019; 38: 106-122.
- [40] Warman ML, Abbott M, Apte SS, Hefferon T, McIntosh I, Cohn DH, Hecht JT, Olsen BR and Francomano CA. A type X collagen mutation causes Schmid metaphyseal chondrodysplasia. *Nat Genet* 1993; 5: 79-82.
- [41] Bateman JF, Wilson R, Freddi S, Lamandé SR and Savarirayan R. Mutations of COL10A1 in Schmid metaphyseal chondrodysplasia. *Hum Mutat* 2005; 25: 525-534.
- [42] Chan D, Weng YM, Graham HK, Sillence DO and Bateman JF. A nonsense mutation in the carboxyl-terminal domain of type X collagen causes haploinsufficiency in schmid metaphyseal chondrodysplasia. *J Clin Invest* 1998; 101: 1490-1499.
- [43] Wu H, Wang S, Li G, Yao Y, Wang N, Sun X, Fang L, Jiang X, Zhao J, Wang Y and Xu C. Characterization of a novel COL10A1 variant associated with Schmid-type metaphyseal chondrodysplasia and a literature review. *Mol Genet Genomic Med* 2021; 9: e1668.
- [44] Ho MS, Tsang KY, Lo RL, Susic M, Mäkitie O, Chan TW, Ng VC, Sillence DO, Boot-Handford RP, Gibson G, Cheung KM, Cole WG, Cheah KS and Chan D. COL10A1 nonsense and frameshift mutations have a gain-of-function effect on the growth plate in human and mouse metaphyseal chondrodysplasia type Schmid. *Hum Mol Genet* 2007; 16: 1201-1215.
- [45] Wang M and Kaufman RJ. Protein misfolding in the endoplasmic reticulum as a conduit to human disease. *Nature* 2016; 529: 326-335.
- [46] Wilson R, Freddi S, Chan D, Cheah KS and Bateman JF. Misfolding of collagen X chains harboring Schmid metaphyseal chondrodysplasia mutations results in aberrant disulfide bond formation, intracellular retention, and activation of the unfolded protein response. *J Biol Chem* 2005; 280: 15544-15552.
- [47] Wilson R, Freddi S and Bateman JF. Collagen X chains harboring Schmid metaphyseal chondrodysplasia NC1 domain mutations are selectively retained and degraded in stably trans-

## COL10A1: emerging biomarker and TME regulator in solid cancers

- ected cells. *J Biol Chem* 2002; 277: 12516-12524.
- [48] Tsang KY, Chan D, Cheslett D, Chan WC, So CL, Melhado IG, Chan TW, Kwan KM, Hunziker EB, Yamada Y, Bateman JF, Cheung KM and Cheah KS. Surviving endoplasmic reticulum stress is coupled to altered chondrocyte differentiation and function. *PLoS Biol* 2007; 5: e44.
- [49] Bialer M. Why are antiepileptic drugs used for nonepileptic conditions? *Epilepsia* 2012; 53 Suppl 7: 26-33.
- [50] Tan WH, Rücklin M, Larionova D, Ngoc TB, Joan van Heuven B, Marone F, Matsudaira P and Winkler C. A collagen10a1 mutation disrupts cell polarity in a medaka model for metaphyseal chondrodysplasia type Schmid. *iScience* 2024; 27: 109405.
- [51] Forouhan M, Sonntag S and Boot-Handford RP. Carbamazepine reduces disease severity in a mouse model of metaphyseal chondrodysplasia type Schmid caused by a premature stop codon (Y632X) in the Col10a1 gene. *Hum Mol Genet* 2018; 27: 3840-3853.
- [52] Mullan LA, Mularczyk EJ, Kung LH, Forouhan M, Wragg JM, Goodacre R, Bateman JF, Swanton E, Briggs MD and Boot-Handford RP. Increased intracellular proteolysis reduces disease severity in an ER stress-associated dwarfism. *J Clin Invest* 2017; 127: 3861-3865.
- [53] Saito T, Fukai A, Mabuchi A, Ikeda T, Yano F, Ohba S, Nishida N, Akune T, Yoshimura N, Nakagawa T, Nakamura K, Tokunaga K, Chung UI and Kawaguchi H. Transcriptional regulation of endochondral ossification by HIF-2 $\alpha$  during skeletal growth and osteoarthritis development. *Nat Med* 2010; 16: 678-686.
- [54] Li F, Lu Y, Ding M, Napierala D, Abbassi S, Chen Y, Duan X, Wang S, Lee B and Zheng Q. Runx2 contributes to murine Col10a1 gene regulation through direct interaction with its cis-enhancer. *J Bone Miner Res* 2011; 26: 2899-2910.
- [55] Gu J, Lu Y, Li F, Qiao L, Wang Q, Li N, Borgia JA, Deng Y, Lei G and Zheng Q. Identification and characterization of the novel Col10a1 regulatory mechanism during chondrocyte hypertrophic differentiation. *Cell Death Dis* 2014; 5: e1469.
- [56] Han T, Zhu T, Bian H, Chen J, Lu Y, Gu J, He TC, Qiao L and Zheng Q. Ddx5 participates in regulation of Col10a1 expression and chondrocyte hypertrophic differentiation in vitro. *Am J Transl Res* 2024; 16: 1454-1467.
- [57] Dy P, Wang W, Bhattaram P, Wang Q, Wang L, Ballock RT and Lefebvre V. Sox9 directs hypertrophic maturation and blocks osteoblast differentiation of growth plate chondrocytes. *Dev Cell* 2012; 22: 597-609.
- [58] Haseeb A, Kc R, Angelozzi M, de Charleroy C, Rux D, Tower RJ, Yao L, Pellegrino da Silva R, Pacifici M, Qin L and Lefebvre V. SOX9 keeps growth plates and articular cartilage healthy by inhibiting chondrocyte dedifferentiation/osteoblastic redifferentiation. *Proc Natl Acad Sci U S A* 2021; 118: e2019152118.
- [59] Melnik S, Gabler J, Dreher SI, Hecht N, Hofmann N, Großner T and Richter W. MiR-218 affects hypertrophic differentiation of human mesenchymal stromal cells during chondrogenesis via targeting RUNX2, MEF2C, and COL10A1. *Stem Cell Res Ther* 2020; 11: 532.
- [60] Etich J, Holzer T, Pitzler L, Bluhm B and Brachvogel B. MiR-26a modulates extracellular matrix homeostasis in cartilage. *Matrix Biol* 2015; 43: 27-34.
- [61] Coghlan RF, Oberdorf JA, Sienko S, Aiona MD, Boston BA, Connelly KJ, Bahney C, LaRouche J, Almubarak SM, Coleman DT, Girkontaite I, von der Mark K, Lunstrum GP and Horton WA. A degradation fragment of type X collagen is a real-time marker for bone growth velocity. *Sci Transl Med* 2017; 9: eaan4669.
- [62] Linsenmayer TF, Eavey RD and Schmid TM. Type X collagen: a hypertrophic cartilage-specific molecule. *Pathol Immunopathol Res* 1988; 7: 14-19.
- [63] Nicol LE, Coghlan RF, Cuthbertson D, Nagamani SCS, Lee B, Olney RC, Horton W and Orwoll E; Members of the Brittle Bone Disease Consortium. Alterations of a serum marker of collagen X in growing children with osteogenesis imperfecta. *Bone* 2021; 149: 115990.
- [64] Carroll RS, Olney RC, Duker AL, Coghlan RF, Schelhaas AJ, Mackenzie WG, Ditro CP, Brown CJ, O'Connell DA, Horton WA, Johnstone B, Espiner EA, Prickett TCR and Bober MB. C-type natriuretic peptide and collagen X marker are aberrant in skeletal dysplasias. *J Bone Miner Res* 2025; 40: 1052-1060.
- [65] Carroll RS, Olney RC, Duker AL, Coghlan RF, Mackenzie WG, Ditro CP, Brown CJ, O'Connell DA, Horton WA, Johnstone B, Espiner EA, Prickett TCR and Bober MB. Collagen X marker levels are decreased in individuals with achondroplasia. *Calcif Tissue Int* 2022; 111: 66-72.
- [66] Working ZM, Morris ER, Chang JC, Coghlan RF, Johnstone B, Miclau T 3rd, Horton WA and Bahney CS. A quantitative serum biomarker of circulating collagen X effectively correlates with endochondral fracture healing. *J Orthop Res* 2021; 39: 53-62.
- [67] Allen KD, Thoma LM and Golightly YM. Epidemiology of osteoarthritis. *Osteoarthritis Cartilage* 2022; 30: 184-195.
- [68] Hunter DJ and Bierma-Zeinstra S. Osteoarthritis. *Lancet* 2019; 393: 1745-1759.
- [69] Zhang Z, Hu Z, Zhao D, Huang H, Liang Y and Mao B. Arthroscopic surgery is not superior to conservative treatment in knee osteoarthritis:

## COL10A1: emerging biomarker and TME regulator in solid cancers

- a systematic review and meta-analysis of randomized controlled trials. *BMC Musculoskelet Disord* 2024; 25: 712.
- [70] Gratal P, Mediero A, Sánchez-Pernaute O, Prieto-Potin I, Lamuedra A, Herrero-Beaumont G and Largo R. Chondrocyte enlargement is a marker of osteoarthritis severity. *Osteoarthritis Cartilage* 2019; 27: 1229-1234.
- [71] Chawla S, Mainardi A, Majumder N, Dönges L, Kumar B, Occhetta P, Martin I, Egloff C, Ghosh S, Bandyopadhyay A and Barbero A. Chondrocyte hypertrophy in osteoarthritis: mechanistic studies and models for the identification of new therapeutic strategies. *Cells* 2022; 11: 4034.
- [72] Dreier R. Hypertrophic differentiation of chondrocytes in osteoarthritis: the developmental aspect of degenerative joint disorders. *Arthritis Res Ther* 2010; 12: 216.
- [73] Rim YA, Nam Y and Ju JH. The role of chondrocyte hypertrophy and senescence in osteoarthritis initiation and progression. *Int J Mol Sci* 2020; 21: 2358.
- [74] He Y, Manon-Jensen T, Arendt-Nielsen L, Petersen KK, Christiansen T, Samuels J, Abramson S, Karsdal MA, Attur M and Bay-Jensen AC. Potential diagnostic value of a type X collagen neo-epitope biomarker for knee osteoarthritis. *Osteoarthritis Cartilage* 2019; 27: 611-620.
- [75] He Y, Siebuhr AS, Brandt-Hansen NU, Wang J, Su D, Zheng Q, Simonsen O, Petersen KK, Arendt-Nielsen L, Eskehave T, Hoeck HC, Karsdal MA and Bay-Jensen AC. Type X collagen levels are elevated in serum from human osteoarthritis patients and associated with biomarkers of cartilage degradation and inflammation. *BMC Musculoskelet Disord* 2014; 15: 309.
- [76] Du Z, Zhao Y, Zhang K, Qin Q, Luo C, Wu J, Zhang H, Liu S, Xu Z, Zheng J, Fan S, Jiang X, Li X and Wang Y. Undercarboxylated OCN inhibits chondrocyte hypertrophy and osteoarthritis development through GPRC6A/HIF-1 $\alpha$  cascade. *Int J Biol Sci* 2025; 21: 4353-4373.
- [77] Huang S, Gao D, Li Z, He H, Yu X, You X, Wu D, Du Z, Zeng J, Shi X, Hu Q, Nie Y, Zhang Z, Luo Z, Wang D, Zhao Z, Li L, Wang G, Wang L, Zhou Z, Chen D and Yang F. Neuronal guidance factor Sema3A inhibits neurite ingrowth and prevents chondrocyte hypertrophy in the degeneration of knee cartilage in mice, monkeys and humans. *Bone Res* 2025; 13: 4.
- [78] Qian Y, Chu G, Zhang L, Wu Z, Wang Q, Guo JJ and Zhou F. M2 macrophage-derived exosomal miR-26b-5p regulates macrophage polarization and chondrocyte hypertrophy by targeting TLR3 and COL10A1 to alleviate osteoarthritis. *J Nanobiotechnology* 2024; 22: 72.
- [79] Chen J, Chen F, Wu X, Bian H, Chen C, Zhang X, Hei R, XiaotongYang, Yuan H, Wang Q, Lu Y, Qiao L and Zheng Q. DLX5 promotes Col10a1 expression and chondrocyte hypertrophy and is involved in osteoarthritis progression. *Genes Dis* 2023; 10: 2097-2108.
- [80] Lu Y, Ding M, Li N, Wang Q, Li J, Li X, Gu J, Im HJ, Lei G and Zheng Q. Col10a1-Runx2 transgenic mice with delayed chondrocyte maturation are less susceptible to developing osteoarthritis. *Am J Transl Res* 2014; 6: 736-745.
- [81] Zeng D, Umar M, Zhu Z, Pan H, Lu WW, Xiao G, Chen Y, Tong L and Chen D. Development of novel osteoarthritis therapy by targeting AMPK- $\beta$ -catenin-Runx2 signaling. *Genes Dis* 2025; 12: 101247.
- [82] Liu Z, Xie W, Li H, Liu X, Lu Y, Lu B, Deng Z and Li Y. Novel perspectives on leptin in osteoarthritis: focus on aging. *Genes Dis* 2024; 11: 101159.
- [83] Chen Z, Li M, Feng Y, Chen Y, Cai Z, Xu Y and Pang R. Identification and analysis of immune aging related biomarkers in cartilage and meniscus tissues of osteoarthritis. *Am J Clin Exp Immunol* 2025; 14: 204-222.
- [84] Crosby D, Bhatia S, Brindle KM, Coussens LM, Dive C, Emberton M, Esener S, Fitzgerald RC, Gambhir SS, Kuhn P, Rebbeck TR and Balasubramanian S. Early detection of cancer. *Science* 2022; 375: eaay9040.
- [85] Li W, Liu JB, Hou LK, Yu F, Zhang J, Wu W, Tang XM, Sun F, Lu HM, Deng J, Bai J, Li J, Wu CY, Lin QL, Lv ZW, Wang GR, Jiang GX, Ma YS and Fu D. Liquid biopsy in lung cancer: significance in diagnostics, prediction, and treatment monitoring. *Mol Cancer* 2022; 21: 25.
- [86] Vaidyanathan R, Soon RH, Zhang P, Jiang K and Lim CT. Cancer diagnosis: from tumor to liquid biopsy and beyond. *Lab Chip* 2018; 19: 11-34.
- [87] Lone SN, Nisar S, Masoodi T, Singh M, Rizwan A, Hashem S, El-Rifai W, Bedognetti D, Batra SK, Haris M, Bhat AA and Macha MA. Liquid biopsy: a step closer to transform diagnosis, prognosis and future of cancer treatments. *Mol Cancer* 2022; 21: 79.
- [88] Nagarajan A, Varadhan V, Manikandan MS, Kaliaperumal K, Palaniyandi T, Kaliamoorthy S, Baskar G, Rab SO, Balaramnavar VM and Kumarasamy S. Signature of collagen alpha-1(x) gene expression in human cancers and their therapeutic implications. *Pathol Res Pract* 2025; 266: 155811.
- [89] Necula L, Matei L, Dragu D, Pitica I, Neagu AI, Bleotu C, Dima S, Popescu I, Diaconu CC and Chivu-Economescu M. High plasma levels of COL10A1 are associated with advanced tumor stage in gastric cancer patients. *World J Gastroenterol* 2020; 26: 3024-3033.

## COL10A1: emerging biomarker and TME regulator in solid cancers

- [90] Thorlacius-Ussing J, Jensen C, Nissen NI, Cox TR, Kalluri R, Karsdal M and Willumsen N. The collagen landscape in cancer: profiling collagens in tumors and in circulation reveals novel markers of cancer-associated fibroblast subtypes. *J Pathol* 2024; 262: 22-36.
- [91] Andriani F, Landoni E, Mensah M, Facchinetti F, Miceli R, Tagliabue E, Giussani M, Callari M, De Cecco L, Colombo MP, Roz L, Pastorino U and Sozzi G. Diagnostic role of circulating extracellular matrix-related proteins in non-small cell lung cancer. *BMC Cancer* 2018; 18: 899.
- [92] Zhang M, Chen H, Wang M, Bai F and Wu K. Bioinformatics analysis of prognostic significance of COL10A1 in breast cancer. *Biosci Rep* 2020; 40: BSR20193286.
- [93] Zhou W, Li Y, Gu D, Xu J, Wang R, Wang H and Liu C. High expression COL10A1 promotes breast cancer progression and predicts poor prognosis. *Heliyon* 2022; 8: e11083.
- [94] Esposito M, Yerly L, Shukla P, Hermes V, Sella F, Balazs Z, Lattmann E, Tastanova A, Turko P, Lang R, Kolm I, Staeger R, Kuonen F, Krauthammer M, Hafner J, Levesque MP and Restivo G. COL10A1 expression distinguishes a subset of cancer-associated fibroblasts present in the stroma of high-risk basal cell carcinoma. *Br J Dermatol* 2024; 191: 775-790.
- [95] Chen S, Wei Y, Liu H, Gong Y, Zhou Y, Yang H and Tang L. Analysis of Collagen type X alpha 1 (COL10A1) expression and prognostic significance in gastric cancer based on bioinformatics. *Bioengineered* 2021; 12: 127-137.
- [96] Shen N, Zhu S, Zhang Z and Yong X. High expression of COL10A1 is an independent predictive poor prognostic biomarker and associated with immune infiltration in advanced gastric cancer microenvironment. *J Oncol* 2022; 2022: 1463316.
- [97] Lee JJ, Ng KY and Bakhtiar A. Extracellular matrix: unlocking new avenues in cancer treatment. *Biomark Res* 2025; 13: 78.
- [98] Huang H, Li T, Ye G, Zhao L, Zhang Z, Mo D, Wang Y, Zhang C, Deng H, Li G and Liu H. High expression of COL10A1 is associated with poor prognosis in colorectal cancer. *Onco Targets Ther* 2018; 11: 1571-1581.
- [99] Kahlert UD, Shi W, Strecker M, Scherpinski LA, Wartmann T, Dölling M, Perrakis A, Relja B, Mengoni M, Braun A and Croner RS. COL10A1 allows stratification of invasiveness of colon cancer and associates to extracellular matrix and immune cell enrichment in the tumor parenchyma. *Front Oncol* 2022; 12: 1007514.
- [100] Wang X, Bai Y, Zhang F, Li D, Chen K, Wu R, Tang Y, Wei X and Han P. Prognostic value of COL10A1 and its correlation with tumor-infiltrating immune cells in urothelial bladder cancer: a comprehensive study based on bioinformatics and clinical analysis validation. *Front Immunol* 2023; 14: 955949.
- [101] Liang Y, Xia W, Zhang T, Chen B, Wang H, Song X, Zhang Z, Xu L, Dong G and Jiang F. Upregulated collagen COL10A1 remodels the extracellular matrix and promotes malignant progression in lung adenocarcinoma. *Front Oncol* 2020; 10: 573534.
- [102] Wang C, Wang J, Chen S, Li K, Wan S and Yang L. COL10A1 as a prognostic biomarker in association with immune infiltration in prostate cancer. *Curr Cancer Drug Targets* 2024; 24: 340-353.
- [103] Liu Q, Zhao H, Guo Y, Zhang K, Shang F and Liu T. Bioinformatics-based analysis: noncoding RNA-mediated COL10A1 is associated with poor prognosis and immune cell infiltration in pancreatic cancer. *J Healthc Eng* 2022; 2022: 7904982.
- [104] Lapa RML, Barros-Filho MC, Marchi FA, Domingues MAC, de Carvalho GB, Drigo SA, Kowalski LP and Rogatto SR. Integrated miRNA and mRNA expression analysis uncovers drug targets in laryngeal squamous cell carcinoma patients. *Oral Oncol* 2019; 93: 76-84.
- [105] Xie C, Du LY, Guo F, Li X and Cheng B. Exosomes derived from microRNA-101-3p-overexpressing human bone marrow mesenchymal stem cells suppress oral cancer cell proliferation, invasion, and migration. *Mol Cell Biochem* 2019; 458: 11-26.
- [106] Solé X, Crous-Bou M, Cordero D, Olivares D, Guinó E, Sanz-Pamplona R, Rodríguez-Moranta F, Sanjuan X, de Oca J, Salazar R and Moreno V. Discovery and validation of new potential biomarkers for early detection of colon cancer. *PLoS One* 2014; 9: e106748.
- [107] Wang T, Bao X, Yang F, Pan S, Xu K and Ren T. Plasma COL10A1 level, a potential diagnostic and prognostic biomarker for pancreatic ductal adenocarcinoma. *Onco Targets Ther* 2024; 17: 949-959.
- [108] Giussani M, Landoni E, Merlino G, Turdo F, Veneroni S, Paolini B, Cappelletti V, Miceli R, Orlandi R, Triulzi T and Tagliabue E. Extracellular matrix proteins as diagnostic markers of breast carcinoma. *J Cell Physiol* 2018; 233: 6280-6290.
- [109] Shen W, Zhou H, Li L, Liu W, Lou Q, Tong CY, Gao J, Gao J and Shao P. Promising protein biomarkers for early gastric cancer: clinical performance of combined detection. *Clin Chem Lab Med* 2025; 63: 1864-1875.
- [110] Wu T and Dai Y. Tumor microenvironment and therapeutic response. *Cancer Lett* 2017; 387: 61-68.
- [111] Li C, Teixeira AF, Zhu HJ and Ten Dijke P. Cancer associated-fibroblast-derived exosomes in

## COL10A1: emerging biomarker and TME regulator in solid cancers

- cancer progression. *Mol Cancer* 2021; 20: 154.
- [112] Jiang Y, Zhang H, Wang J, Liu Y, Luo T and Hua H. Targeting extracellular matrix stiffness and mechanotransducers to improve cancer therapy. *J Hematol Oncol* 2022; 15: 34.
- [113] Peng J, Liu X, Mao Y, Lv M, Ma T, Liu J, Zhou Q, Han Y, Li X and Wang H. Upregulation of collagen type X alpha 1 promotes the progress of triple-negative breast cancer via Wnt/ $\beta$ -catenin signaling. *Mol Carcinog* 2024; 63: 1588-1598.
- [114] Li T, Huang H, Shi G, Zhao L, Li T, Zhang Z, Liu R, Hu Y, Liu H, Yu J and Li G. TGF- $\beta$ 1-SOX9 axis-inducible COL10A1 promotes invasion and metastasis in gastric cancer via epithelial-to-mesenchymal transition. *Cell Death Dis* 2018; 9: 849.
- [115] Raaijmakers KTPM, Adema GJ, Bussink J and Ansems M. Cancer-associated fibroblasts, tumor and radiotherapy: interactions in the tumor micro-environment. *J Exp Clin Cancer Res* 2024; 43: 323.
- [116] Masuda H. Cancer-associated fibroblasts in cancer drug resistance and cancer progression: a review. *Cell Death Discov* 2025; 11: 341.
- [117] Wang M, Feng R, Chen Z, Shi W, Li C, Liu H, Wu K, Li D and Li X. Identification of cancer-associated fibroblast subtype of triple-negative breast cancer. *J Oncol* 2022; 2022: 6452636.
- [118] Hu S, Ding M, Lou J, Qin J, Chen Y, Liu Z, Li Y, Nie J, Xu M, Sun H, Gu X, Xu T, Wang S, Wang S and Pan Y. COL10A1(+) fibroblasts promote colorectal cancer metastasis and M2 macrophage polarization with pan-cancer relevance. *J Exp Clin Cancer Res* 2025; 44: 243.
- [119] Geng X, Chen H, Zhao L, Hu J, Yang W, Li G, Cheng C, Zhao Z, Zhang T, Li L and Sun B. Cancer-Associated Fibroblast (CAF) heterogeneity and targeting therapy of CAFs in pancreatic cancer. *Front Cell Dev Biol* 2021; 9: 655152.
- [120] Lei X, Lei Y, Li JK, Du WX, Li RG, Yang J, Li J, Li F and Tan HB. Immune cells within the tumor microenvironment: biological functions and roles in cancer immunotherapy. *Cancer Lett* 2020; 470: 126-133.
- [121] Mao X, Xu J, Wang W, Liang C, Hua J, Liu J, Zhang B, Meng Q, Yu X and Shi S. Crosstalk between cancer-associated fibroblasts and immune cells in the tumor microenvironment: new findings and future perspectives. *Mol Cancer* 2021; 20: 131.
- [122] Meng Z, Zhang R, Wu X, Piao Z, Zhang M and Jin T. LncRNA HAGLROS promotes breast cancer evolution through miR-135b-3p/COL10A1 axis and exosome-mediated macrophage M2 polarization. *Cell Death Dis* 2024; 15: 633.
- [123] Xu S, Liu D, Qin Z, Liang Z, Xie H, Yi B, Wang K, Lin G, Liu R, Yang K, Xu Y and Zhang H. Experimental validation and pan-cancer analysis identified COL10A1 as a novel oncogene and potential therapeutic target in prostate cancer. *Aging (Albany NY)* 2023; 15: 15134-15160.
- [124] Wang W, Shen J, Song D, Fu K and Fu X. Identification of macrophage-related genes in bladder cancer patients using single-cell sequencing and construction of a prognostic model. *Am J Clin Exp Immunol* 2024; 13: 88-104.
- [125] Cai S, Sun Z, Yan Y, Li W and Wu Q. COL10A1 is a potential immunotherapy biomarker associated with immune infiltration and deficient mismatch repair in colon cancer. *Immunotherapy* 2023; 15: 1293-1308.
- [126] Johannet P, Rousseau B, Aghajanian C, Foote MB and Diaz LA Jr. Therapeutic targeting of mismatch repair-deficient cancers. *Nat Rev Clin Oncol* 2025; 22: 734-759.
- [127] Brodsky AS, Xiong J, Yang D, Schorl C, Fenton MA, Graves TA, Sikov WM, Resnick MB and Wang Y. Identification of stromal ColX $\alpha$ 1 and tumor-infiltrating lymphocytes as putative predictive markers of neoadjuvant therapy in estrogen receptor-positive/HER2-positive breast cancer. *BMC Cancer* 2016; 16: 274.
- [128] Mohan V, Das A and Sagi I. Emerging roles of ECM remodeling processes in cancer. *Semin Cancer Biol* 2020; 62: 192-200.
- [129] Najafi M, Farhood B and Mortezaee K. Extracellular matrix (ECM) stiffness and degradation as cancer drivers. *J Cell Biochem* 2019; 120: 2782-2790.
- [130] Yuzhalin AE, Lim SY, Kutikhin AG and Gordon-Weeks AN. Dynamic matrisome: ECM remodeling factors licensing cancer progression and metastasis. *Biochim Biophys Acta Rev Cancer* 2018; 1870: 207-228.
- [131] Trono P, Ottavi F and Rosano L. Novel insights into the role of Discoidin domain receptor 2 (DDR2) in cancer progression: a new avenue of therapeutic intervention. *Matrix Biol* 2024; 125: 31-39.
- [132] Leitinger B and Kwan AP. The discoidin domain receptor DDR2 is a receptor for type X collagen. *Matrix Biol* 2006; 25: 355-364.
- [133] Dawson JC, Serrels A, Stupack DG, Schlaepfer DD and Frame MC. Targeting FAK in anticancer combination therapies. *Nat Rev Cancer* 2021; 21: 313-324.
- [134] Xiong J, Yan L, Zou C, Wang K, Chen M, Xu B, Zhou Z and Zhang D. Integrins regulate stemness in solid tumor: an emerging therapeutic target. *J Hematol Oncol* 2021; 14: 177.
- [135] Moreira AM, Ferreira RM, Carneiro P, Figueiredo J, Osório H, Barbosa J, Preto J, Pinto-do-Ó P, Carneiro F and Seruca R. Proteomic Identification of a gastric tumor ECM signature associated with cancer progression. *Front Mol Biosci* 2022; 9: 818552.

## COL10A1: emerging biomarker and TME regulator in solid cancers

- [136] Senger DR and Davis GE. Angiogenesis. *Cold Spring Harb Perspect Biol* 2011; 3: a005090.
- [137] Hisano Y and Hla T. Bioactive lysolipids in cancer and angiogenesis. *Pharmacol Ther* 2019; 193: 91-98.
- [138] Ye ZW, Yu ZL, Chen G and Jia J. Extracellular vesicles in tumor angiogenesis and resistance to anti-angiogenic therapy. *Cancer Sci* 2023; 114: 2739-2749.
- [139] Mackie EJ, Ahmed YA, Tatarczuch L, Chen KS and Mirams M. Endochondral ossification: how cartilage is converted into bone in the developing skeleton. *Int J Biochem Cell Biol* 2008; 40: 46-62.
- [140] Liu Y, Xie HQ and Shen B. Type H vessels-a bridge connecting subchondral bone remodeling and articular cartilage degeneration in osteoarthritis development. *Rheumatology (Oxford)* 2023; 62: 1436-1444.
- [141] Lv D, Chen D, Wang Z, Cui Z, Ma JH, Ji S, Chen J and Tang S. COL10A1 is a novel factor in the development of choroidal neovascularization. *Microvasc Res* 2022; 139: 104239.
- [142] Pastushenko I and Blanpain C. EMT transition states during tumor progression and metastasis. *Trends Cell Biol* 2019; 29: 212-226.
- [143] Davis FM, Stewart TA, Thompson EW and Monteith GR. Targeting EMT in cancer: opportunities for pharmacological intervention. *Trends Pharmacol Sci* 2014; 35: 479-488.
- [144] Ramesh V, Brabletz T and Ceppi P. Targeting EMT in cancer with repurposed metabolic inhibitors. *Trends Cancer* 2020; 6: 942-950.
- [145] Necula LG, Dragu DL, Matei L, Pitica I, Dima SO, Bleotu C, Diaconu CC and Chivu-Economescu M. COL10A1 overexpression promotes gastric cancer aggressiveness through EMT and major oncogenic pathways. *Int J Mol Sci* 2025; 26: 11043.
- [146] Wang X, Ma S, Li S, Jia W and Zhang D. The regulatory effect of Col10A1 to the intracranial vascular invasion and cell proliferation in breast cancer via EMT pathway. *Sci Rep* 2025; 15: 11040.
- [147] Peng D, Fu M, Wang M, Wei Y and Wei X. Targeting TGF- $\beta$  signal transduction for fibrosis and cancer therapy. *Mol Cancer* 2022; 21: 104.
- [148] Xu X, Zheng L, Yuan Q, Zhen G, Crane JL, Zhou X and Cao X. Transforming growth factor- $\beta$  in stem cells and tissue homeostasis. *Bone Res* 2018; 6: 2.
- [149] Derynck R, Akhurst RJ and Balmain A. TGF-beta signaling in tumor suppression and cancer progression. *Nat Genet* 2001; 29: 117-129.
- [150] Xu J, Lamouille S and Derynck R. TGF-beta-induced epithelial to mesenchymal transition. *Cell Res* 2009; 19: 156-172.
- [151] Hata A and Chen YG. TGF- $\beta$  signaling from receptors to Smads. *Cold Spring Harb Perspect Biol* 2016; 8: a022061.
- [152] Shi Y and Massagué J. Mechanisms of TGF-beta signaling from cell membrane to the nucleus. *Cell* 2003; 113: 685-700.
- [153] Lee JH and Massagué J. TGF- $\beta$  in developmental and fibrogenic EMTs. *Semin Cancer Biol* 2022; 86: 136-145.
- [154] Zavadil J and Böttinger EP. TGF-beta and epithelial-to-mesenchymal transitions. *Oncogene* 2005; 24: 5764-5774.
- [155] Sun Y, Ling J and Liu L. Collagen type X alpha 1 promotes proliferation, invasion and epithelial-mesenchymal transition of cervical cancer through activation of TGF- $\beta$ /Smad signaling. *Physiol Int* 2022; 109: 204-214.
- [156] Leitinger B. Discoidin domain receptor functions in physiological and pathological conditions. *Int Rev Cell Mol Biol* 2014; 310: 39-87.
- [157] Zhao X and Guan JL. Focal adhesion kinase and its signaling pathways in cell migration and angiogenesis. *Adv Drug Deliv Rev* 2011; 63: 610-615.
- [158] Campbell ID and Humphries MJ. Integrin structure, activation, and interactions. *Cold Spring Harb Perspect Biol* 2011; 3: a004994.
- [159] Bachmann M, Kukkurainen S, Hytönen VP and Wehrle-Haller B. Cell adhesion by integrins. *Physiol Rev* 2019; 99: 1655-1699.
- [160] Su C, Mo J, Dong S, Liao Z, Zhang B and Zhu P. Integrin $\beta$ -1 in disorders and cancers: molecular mechanisms and therapeutic targets. *Cell Commun Signal* 2024; 22: 71.
- [161] He Y, Sun MM, Zhang GG, Yang J, Chen KS, Xu WW and Li B. Targeting PI3K/Akt signal transduction for cancer therapy. *Signal Transduct Target Ther* 2021; 6: 425.
- [162] Liu J, Chai XX, Qiu XR, Sun WJ, Tian YL, Guo WH, Yin DC and Zhang CY. Type X collagen knockdown inactivate ITGB1/PI3K/AKT to suppress chronic unpredictable mild stress-stimulated triple-negative breast cancer progression. *Int J Biol Macromol* 2024; 273: 133074.
- [163] Luckman SP, Rees E and Kwan AP. Partial characterization of cell-type X collagen interactions. *Biochem J* 2003; 372: 485-493.
- [164] Jiang K, Xu LZ, Cheng F and Ning JZ. COL10A1 facilitates prostate cancer progression by interacting with INHBA to activate the PI3K/AKT pathway. *J Cell Mol Med* 2024; 28: e70249.
- [165] Abdel Mouti M and Pauklin S. TGFB1/INHBA homodimer/Nodal-SMAD2/3 signaling network: a pivotal molecular target in PDAC treatment. *Mol Ther* 2021; 29: 920-936.
- [166] Chen ZL, Qin L, Peng XB, Hu Y and Liu B. INHBA gene silencing inhibits gastric cancer cell migration and invasion by impeding activation of

## COL10A1: emerging biomarker and TME regulator in solid cancers

- the TGF- $\beta$  signaling pathway. *J Cell Physiol* 2019; 234: 18065-18074.
- [167] Ikegawa S, Nakamura K, Nagano A, Haga N and Nakamura Y. Mutations in the N-terminal globular domain of the type X collagen gene (COL10A1) in patients with Schmid metaphyseal chondrodysplasia. *Hum Mutat* 1997; 9: 131-135.
- [168] Sawai H, Ida A, Nakata Y and Koyama K. Novel missense mutation resulting in the substitution of tyrosine by cysteine at codon 597 of the type X collagen gene associated with Schmid metaphyseal chondrodysplasia. *J Hum Genet* 1998; 43: 259-261.
- [169] Wallis GA, Rash B, Sweetman WA, Thomas JT, Super M, Evans G, Grant ME and Boot-Handford RP. Amino acid substitutions of conserved residues in the carboxyl-terminal domain of the alpha 1(X) chain of type X collagen occur in two unrelated families with metaphyseal chondrodysplasia type Schmid. *Am J Hum Genet* 1994; 54: 169-178.
- [170] McIntosh I, Abbott MH and Francomano CA. Concentration of mutations causing Schmid metaphyseal chondrodysplasia in the C-terminal noncollagenous domain of type X collagen. *Hum Mutat* 1995; 5: 121-125.
- [171] McIntosh I, Abbott MH, Warman ML, Olsen BR and Francomano CA. Additional mutations of type X collagen confirm COL10A1 as the Schmid metaphyseal chondrodysplasia locus. *Hum Mol Genet* 1994; 3: 303-307.
- [172] Mäkitie O, Susic M, Ward L, Barclay C, Glorieux FH and Cole WG. Schmid type of metaphyseal chondrodysplasia and COL10A1 mutations-findings in 10 patients. *Am J Med Genet A* 2005; 137A: 241-248.
- [173] Pateras J, Lodi M, Rana P and Ghosh P. Heterogeneous clustering of multiomics data for breast cancer subgroup classification and detection. *Int J Mol Sci* 2025; 26: 1707.
- [174] Yang W, Wu X and Zhou F. Collagen type X Alpha 1 (COL10A1) contributes to cell proliferation, migration, and invasion by targeting Prolyl 4-Hydroxylase Beta Polypeptide (P4HB) in breast cancer. *Med Sci Monit* 2021; 27: e928919.
- [175] Schultz S, Bartsch H, Sotlar K, Petat-Dutter K, Bonin M, Kahlert S, Harbeck N, Vogel U, Seeger H, Fehm T and Neubauer HJ. Progression-specific genes identified in microdissected formalin-fixed and paraffin-embedded tissue containing matched ductal carcinoma in situ and invasive ductal breast cancers. *BMC Med Genomics* 2018; 11: 80.
- [176] Famili-Youth EHH, Famili-Youth A, Yang D, Siddique A, Wu EY, Liu W, Resnick MB, Chen Q and Brodsky AS. Aberrant expression of collagen type X in solid tumor stroma is associated with EMT, immunosuppressive and pro-metastatic pathways, bone marrow stromal cell signatures, and poor survival prognosis. *BMC Cancer* 2025; 25: 247.
- [177] Vishnubalaji R, Shaath H, Elkord E and Alajez NM. Long non-coding RNA (lncRNA) transcriptional landscape in breast cancer identifies LINC01614 as non-favorable prognostic biomarker regulated by TGF $\beta$  and focal adhesion kinase (FAK) signaling. *Cell Death Discov* 2019; 5: 109.
- [178] Liu J, Qiu XR, Tian YL, Sun WJ, Wang YH, Liu H, Zhang G, Zhao WZ, Yin DC and Zhang CY. Urchin-like magnetic nanoparticles loaded with type X collagen siRNA and Stattic to treat triple negative breast cancer under rotating magnetic field like an "enchanted micro-scalpel". *Int J Biol Macromol* 2025; 300: 140318.
- [179] Liu Z, Sun L, Zhu W, Zhu J, Wu C, Peng X, Tian H, Huang C and Zhu Z. Disulfidptosis signature predicts immune microenvironment and prognosis of gastric cancer. *Biol Direct* 2024; 19: 65.
- [180] Aktas SH, Taskin-Tok T, Al-Khafaji K and Akin-Bali DF. A detailed understanding of the COL10A1 and SOX9 genes interaction based on potentially damaging mutations in gastric cancer using computational techniques. *J Biomol Struct Dyn* 2022; 40: 11533-11544.
- [181] You W, Ouyang J, Cai Z, Chen Y and Wu X. Comprehensive analyses of immune subtypes of stomach adenocarcinoma for mRNA vaccination. *Front Immunol* 2022; 13: 827506.
- [182] Liu Q, Zhang W, Wu Z, Liu H, Hu H, Shi H, Li S and Zhang X. Construction of a circular RNA-microRNA-messengerRNA regulatory network in stomach adenocarcinoma. *J Cell Biochem* 2020; 121: 1317-1331.
- [183] Cai Z, Wei Y, Chen S, Gong Y, Fu Y, Dai X, Zhou Y, Yang H, Tang L and Liu H. Screening and identification of key biomarkers in alimentary tract cancers: a bioinformatic analysis. *Cancer Biomark* 2020; 29: 221-233.
- [184] Qin Y, Miyake T, Muramoto K, Maekawa T, Nishina Y, Wang Y, Shimizu T and Tani M. Fibroblast activation protein- $\alpha$  expression in cancer-associated fibroblasts shows the poor survival of colorectal cancer via immune-mediated pathways: implications of FAP in cancer-associated fibroblasts link immune dysregulation to adverse survival in colorectal cancer. *Ann Surg Oncol* 2025; 32: 1941-1952.
- [185] Wu J, Ouyang P, Huang R, Cui Y, Yang Z, Xu W, Ma R, Xiang G, Zeng W, Wu W and Li J. MET- $\text{TL16}$  promotes stability of SYNPO2L mRNA and leading to cancer cell lung metastasis by secretion of COL10A1 and attract the cancer-associated fibroblasts. *Int J Biol Sci* 2024; 20: 4128-4145.

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- [186] Li Y, Wang X, Shi L, Xu J and Sun B. Predictions for high COL1A1 and COL10A1 expression resulting in a poor prognosis in esophageal squamous cell carcinoma by bioinformatics analyses. *Transl Cancer Res* 2020; 9: 85-94.
- [187] Song Y, Wang X, Wang F, Peng X, Li P, Liu S and Zhang D. Identification of four genes and biological characteristics of esophageal squamous cell carcinoma by integrated bioinformatics analysis. *Cancer Cell Int* 2021; 21: 123.
- [188] Purkayastha BPD, Chan ER, Ravillah D, Ravi L, Gupta R, Canto MI, Wang JS, Shaheen NJ, Willis JE, Chak A, Varadan V and Guda K. Genome-scale analysis identifies novel transcript-variants in esophageal adenocarcinoma. *Cell Mol Gastroenterol Hepatol* 2020; 10: 652-654, e617.
- [189] Jinato T, Khamjerm J, Manprasong S, Iadsee N, Tangkijvanich P, Nookaew I and Chuaypen N. Gut microbiota and host transcriptome interactions reveal diagnostic biomarkers in MASLD-associated hepatocellular carcinoma. *Gut Pathog* 2025; 17: 107.
- [190] Wang S, Yu L, Sun X and Zhang B. Establishment and verification of potential biomarkers for cholangiocarcinoma. *Exp Ther Med* 2022; 24: 546.
- [191] Ding X, Liu H, Xu Q, Ji T, Chen R, Liu Z and Dai J. Shared biomarkers and mechanisms in idiopathic pulmonary fibrosis and non-small cell lung cancer. *Int Immunopharmacol* 2024; 134: 112162.
- [192] Guo Q, Zheng M, Xu Y, Wang N and Zhao W. MiR-384 induces apoptosis and autophagy of non-small cell lung cancer cells through the negative regulation of Collagen  $\alpha$ -1(X) chain gene. *Biosci Rep* 2019; 39: BSR20181523.
- [193] Akçay S. Deciphering molecular overlaps between COPD and NSCLC subtypes (LUAD and LUSC): an integrative bioinformatics study. *Medicine (Baltimore)* 2025; 104: e43906.
- [194] Li Y, Li X, Deng M, Ye C, Peng Y and Lu Y. Cancer-associated fibroblasts hinder lung squamous cell carcinoma oxidative stress-induced apoptosis via METTL3 mediated m(6)A methylation of COL10A1. *Oxid Med Cell Longev* 2022; 2022: 4320809.
- [195] Guo W, Zheng X, Hua L, Zheng X, Zhang Y, Sun B, Tao Z and Gao J. Screening and bioinformatical analysis of differentially expressed genes in nasopharyngeal carcinoma. *J Cancer* 2021; 12: 1867-1883.
- [196] Zhao J, Wang R, Sun X, Huang K, Jin J, Lan L, Jian Y, Xu Z, Wu H, Wang S and Wang J. An integrative multi-omics analysis based on nomogram for predicting prostate cancer bone metastasis incidence. *Genet Res (Camb)* 2022; 2022: 8213723.
- [197] Cen S, Jiang D, Lv D, Xu R, Hou J, Yang Z, Wu P, Xiong X and Gao X. Comprehensive analysis of the biological functions of endoplasmic reticulum stress in prostate cancer. *Front Endocrinol (Lausanne)* 2023; 14: 1090277.
- [198] Wu SX, Huang J, Liu ZW, Chen HG, Guo P, Cai QQ, Zheng JJ, Qin HD, Zheng ZS, Chen X, Zhang RY, Chen SL and Lin TX. A genomic-clinicopathologic nomogram for the preoperative prediction of lymph node metastasis in bladder cancer. *EBioMedicine* 2018; 31: 54-65.
- [199] Xia F, Jiang B, Chen Y, Du X, Peng Y, Wang W, Wang Z and Li X. Prediction of novel target genes and pathways involved in tall cell variant papillary thyroid carcinoma. *Medicine (Baltimore)* 2018; 97: e13802.
- [200] Wang S, Zhong L, Li Y, Xiao D, Zhang R, Liao D, Lv D, Wang X, Wang J, Xie X, Chen J, Wu Y and Kang T. Up-regulation of PCOLCE by TWIST1 promotes metastasis in osteosarcoma. *Theranostics* 2019; 9: 4342-4353.
- [201] Li GB, Liu GY, Yang J and Li DW. Weighted gene correlation network analysis identifies the critical long non-coding RNAs participate in the progression of osteosarcoma. *Gen Physiol Biophys* 2021; 40: 173-182.
- [202] Ma R, Mandell J, Lu F, Heim T, Schoedel K, Duensing A, Watters RJ and Weiss KR. Do patient-derived spheroid culture models have relevance in chondrosarcoma research? *Clin Orthop Relat Res* 2021; 479: 477-490.
- [203] Sakimura R, Tanaka K, Yamamoto S, Matsunobu T, Li X, Hanada M, Okada T, Nakamura T, Li Y and Iwamoto Y. The effects of histone deacetylase inhibitors on the induction of differentiation in chondrosarcoma cells. *Clin Cancer Res* 2007; 13: 275-282.