

## Review Article

# Regulation of cancer stem cell activities by tumor-associated macrophages

Masahisa Jinushi, Muhammad Baghdadi, Shigeki Chiba, Hironori Yoshiyama

*Research Center for Infection-associated cancer, Institute for Genetic Medicine, Hokkaido University*

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**Abstract:** Recent studies revealed that tumor-associated macrophages play a decisive role in the regulation of tumor progression by manipulating tumor oncogenesis, angiogenesis and immune functions within tumor microenvironments. However, the role of cancer stem cells in the tumorigenic activities of tumor-associated macrophages during the course of transformation and treatment remains largely unknown. Recent studies have clarified the functional aspects of tumor-associated macrophages in the regulation of the tumorigenic activities and anticancer drug responsiveness of cancer stem cells through complex networks formed by distinct sets of cytokines, chemokines and growth factors. In this article we discuss recent advances and future perspectives regarding the molecular interplay between cancer stem cells and tumor-associated macrophages and provide future perspective about the therapeutic implication against treatment-resistant variants of cancer.

**Keywords:** Cancer stem cells, tumor associated macrophages, tumor microenvironments, MFG-E8, IL-6, TIM-3, M-CSF

### Introduction

Genetic and epigenetic alterations in heterogeneous tumor cell populations regulate tumor initiation, progression and therapeutic responses. On the other hands, emerging evidence unveiled that heterogeneous tumor microenvironments composed of tumor cells as well as normal cells including mesenchymal stem cells, fibroblasts, endothelial cells and immune cells, have a large impact on the behavior of tumorigenic cells during the course of tumor progression [1-3]. In addition, tumor cells may modify the biological properties of stromal cells, endothelial cells and immune cells in their unique microenvironments, thereby contributing to further tumor progression and the emergence of drug-resistant tumor phenotypes [4, 5]. Thus, the interaction between tumor cells and their surrounding normal cellular components may have a determining role in the regulation of tumor initiation, progression, and responsiveness to anticancer therapeutics.

Recent advances molecular immunology and the clinical success of drugs targeting immune-

regulatory circuits have emphasized the importance of tumor immune surveillance systems against nascent tumors [6-8]. On the other hand, tumor-infiltrating immune cells frequently promote tumor growth and incur invasive behavior through the coordinated activation of distinct inflammatory and angiogenic signals in the background of smoldering inflammation [9-11]. In particular, tumor-associated macrophages (TAM) have a critical role in modulating tumorigenic activities by activating oncogenic signals, angiogenesis, tissue/matrix remodeling and immune suppression [12-14].

In this review, we will describe recent advances in our understanding of the regulatory mechanisms whereby TAM impact cancer stem cell functions.

### The role of tumor microenvironments in the regulation of cancer stem cell activities

Mesenchymal stem cells (MSC) are one of the critical components in tumor microenvironments. MSC enter the circulation into bloodstream from the bone marrow [15] or reside in

normal stromal tissues [16, 17]. Emergent evidence has unveiled the critical role of MSC in positively regulating the tumorigenic activities of cancer stem cells in several murine tumor models [17]. Furthermore, immunohistochemical analysis has confirmed the proximity of MSC and cancer stem cells in biopsies obtained from cancer patients, raising the possibility that MSC have an impact on the clinical course of human malignancies by modulating the cancer stem cell functions [18].

In the tumor microenvironments, MSCs have the ability to differentiate into stromal fibroblasts, which also interact with and influence tumor cells through paracrine signals and various soluble factors [19, 20]. SDF-1 produced by breast carcinoma-associated fibroblasts (but not normal fibroblasts) accelerates the growth and metastatic potential of breast cancers, which express high levels of the SDF-1 receptor CXCR4 [21, 22]. Hepatocyte growth factor (HGF) provides a co-stimulatory signal to the Wnt pathway during colon carcinogenesis [23]. Since niche activities regulated by Wnt- $\beta$ -catenin cascades have a critical role in the survival and self-renewal of tissue and cancer stem cells, HGF released from stromal fibroblast may regulate cancer stem cell functions by stimulating Wnt- $\beta$ -catenin pathways in a paracrine fashion [24]. Moreover, additional factors produced by stromal fibroblasts, which include NOS, PDGF, Notch ligand and Hedgehog ligands, are potential candidates to regulate cancer stem cell activities [25, 26]. In addition, the stromal signals serve as a prognostic factor in patients with breast cancer, suggesting that stromal microenvironments composed of MSC and stromal fibroblasts greatly impact the biological behavior of tumorigenic cells in actual clinical settings [27].

Endothelial cells may impact biological behaviors of cancer stem cells in tumor microenvironment by direct interaction with tumor cells as well as by their role in blood vessel formation. Endothelial cells constitute an important component of normal hematopoietic and neuronal stem cell niches, but tumor vasculature is different from normal vasculature, raising the possibility that tumor microenvironments produce defined factors that modulate the genetic and epigenetic profiles of endothelial cells. Indeed, more than 1,000 genes are differentially expressed comparing tumor vs normal endothelial cells, including FGF receptors, MMPs and NF- $\kappa$ B

-regulated transcripts [28, 29]. In addition, cytokines produced by endothelial cells, which includes HGF, VEGF, PDGF and PIGF stimulate the self-renewal and survival of adjacent cancer stem cells [30, 31]. Interestingly, recent reports demonstrate that glioblastoma stem cells can trans-differentiate into endothelial cells to generate their own vasculature, thereby providing blood supply to adjacent tumor cells and further accelerating tumor progression [32, 33].

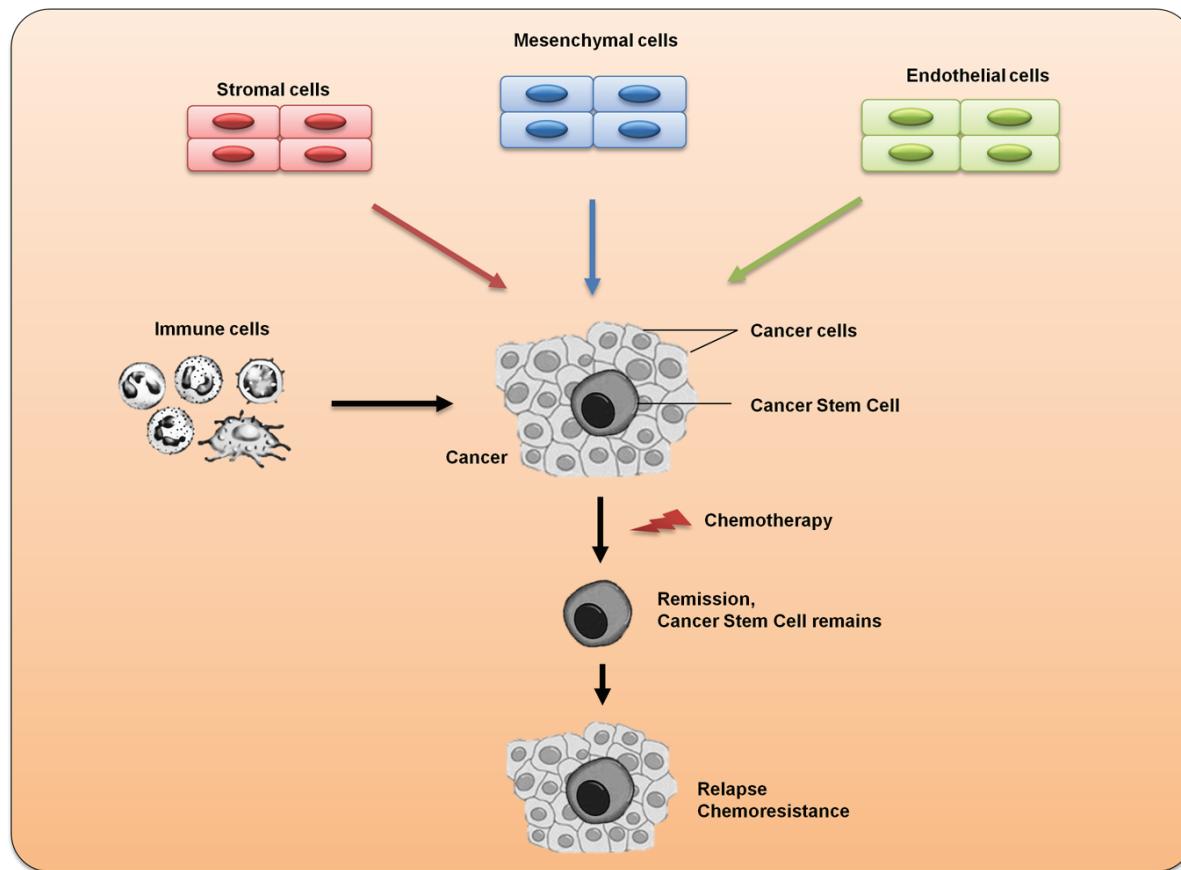
Together, the complex networks created by cancer stem cells and the surrounding normal cellular components contribute to the proliferation as well as the invasive activities of tumors in murine models as well as cancer patients (**Figure 1**).

#### The role of TAM in tumor progression and anti-cancer drug resistance

The immune system provides both inhibitory and stimulatory effectors in tumor initiation, promotion and metastasis, and the balance of these effects may be determined by different tumor microenvironments. Thus, the distinct modes of interplay between tumorigenic cells and host immunity may profoundly influence both the biological behaviors of tumors during the course of tumorigenicity and their responsiveness to anticancer modalities [6-11]. In particular, emerging evidences have unveiled the molecular mechanisms by which myeloid cells such as macrophages and myeloid-derived suppressor cells (MDSC), interact with tumor microenvironments to further accelerate tumor progression [12, 34].

Tumor-associated macrophages are characterized by distinct phenotypic polarization referred as "M1 and M2" subsets [14, 35]. The M1-polarized macrophages manifest high levels of proinflammatory cytokines, high production of reactive nitrogen and oxygen intermediates, and promote Th1 responses, which contributes to tumoricidal activity and antitumor immunity. On the other hands, M2 macrophages serve as the main players facilitating parasite containment, tissue remodeling and immune tolerance, which may be linked with tumor progression [36, 37]. The M1 polarization in macrophages is mainly regulated by distinct transcriptional networks consisting of IRF-1/5, Stat-1/4 and NF- $\kappa$ B, whereas M2 polarization is regulated through other transcription factors such as IRF-4, Stat-

## TAM regulation of CSC



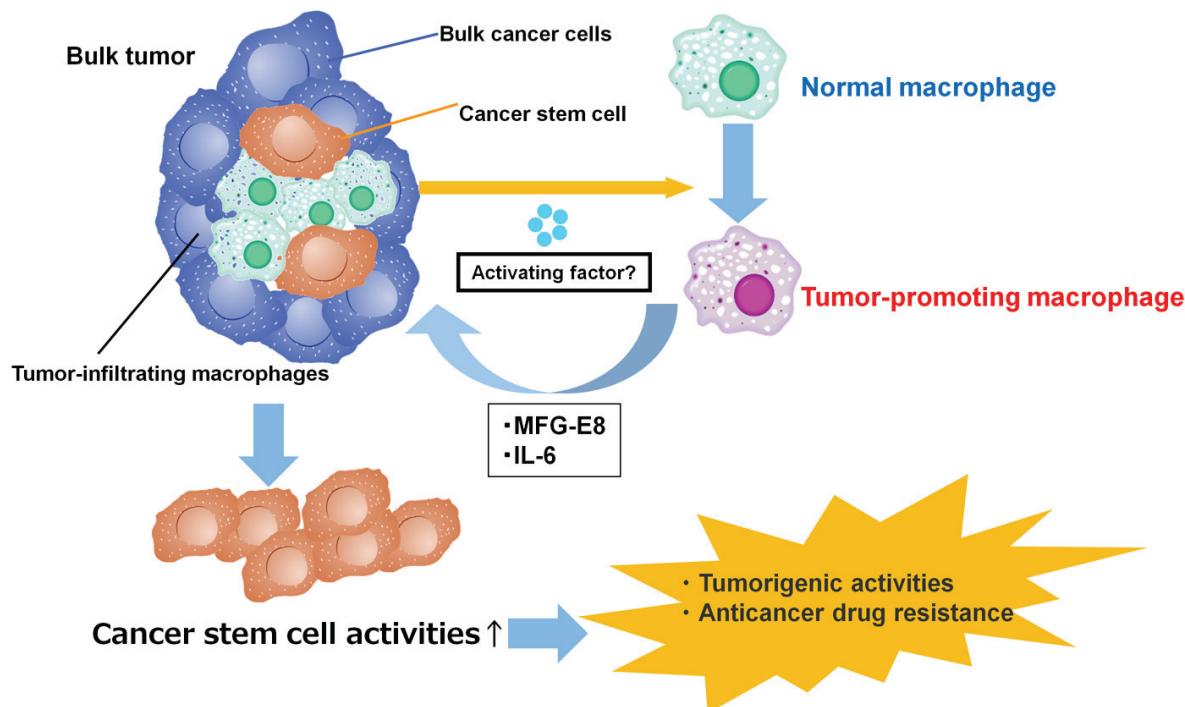
**Figure 1.** The complex networks created by cancer stem cells and their tumor microenvironments contribute to the tumorigenic and invasive activities of tumors.

3/6, PPAR- $\gamma$ , KLF-4 and histone demethylase such as Jmjd3 [38-42]. In several murine models of carcinogenesis, tumor progression is frequently associated with a phenotypic switch from M1 to M2 in TAM [43]. Furthermore, M1-polarized macrophages mediate elimination of senescent hepatocytes, which drive subsequent carcinogenesis [44]. It is therefore possible that classically activated M1 macrophages contribute to the tumor elimination and equilibrium phases during tumor progression via T cell-mediated mechanisms [6].

However, there is a growing appreciation that TAM are composed of several different populations that bear sufficient plasticity and flexibility to enable them to cause dynamic changes between the M1 and M2 phenotypes depending on the different tumor microenvironments [46, 47]. Furthermore, recent genetic profiling and phenotypic analyses have also revealed that

tumor cells, through distinct sets of signaling molecules, transcription factors, and epigenetic modifiers, manipulate tumor-infiltrating myeloid cells to differentiate them into peculiar subsets with tumor-promoting capacities [48, 49]. In this regard, tumor microenvironments characterized by smoldering inflammation and/or modulated by anticancer drug-mediated stress responses may serve as driving forces to alter the genetic and phenotypic profiles of tumor cells, thus promoting tumorigenic activities of myeloid cells in an autocrine and paracrine fashion.

Recent comprehensive analysis revealed that the numbers and activities of tumor-associated macrophages may influence the prognosis of patients with Hodgkin's lymphoma and breast cancer [49, 50]. Furthermore, treatment with M-CSF kinase inhibitors had significant antitumor activity against patients-derived primary tumors arising in immunodeficient mice when com-



**Figure 2.** TAM serves as the major components of niche microenvironments regulating cancer stem cell functions. MFG-E8 and IL-6 predominantly produced from TAM in a cancer stem cell-specific fashion trigger tumorigenesis and anticancer drug resistance in cancer stem cells through the coordinated activation of the Stat3 and Hedgehog pathways. The interplay between cancer stem cells and TAM accelerates tumor progression and drug resistance through autocrine positive-feedback mechanisms.

bined with conventional cytotoxic chemotherapy [50].

These findings suggest that different tumor microenvironments may have a distinct impact on the ability of TAM on tumor growth and therapeutic responses to chemotherapy. The identification of the various cellular and molecular pathways and their downstream factors that participate in the interaction between tumorigenic cells and tumor-infiltrating myeloid cells in various human cancers will translate our understanding of cancer-related inflammation to meaningful therapeutic advances.

#### The interplay between cancer stem cells and TAM in the regulation of tumorigenicity and anti-cancer drug responses

Recent studies have clarified the importance of TAM as major contributors in the regulation of both self-renewal and anticancer drug responses of cancer stem cells through distinct networks of cytokines, chemokines and growth

factors. In these processes, TAM interact with and promote the tumorigenicity of cancer stem cells via production of milk-fat globule-epidermal growth factor-VIII (MFG-E8) and IL-6 through coordinated activation of the STAT3 and sonic hedgehog pathways [51]. Interestingly, cancer stem cells are the major subset promoting the production of MFG-E8 and IL-6 from macrophages, implying that mediators specifically regulated by cancer stem cells render macrophages with the ability to facilitate the production of tumorigenic factors such as MFG-E8 and IL-6. In this sense, TAM might serve as a component of the “immunological niche”, by which cancer stem cell activities are maintained and amplified within tumor microenvironments (Figure 2).

Cancer stem cells have unique characteristics that manipulate complex signaling cascades which regulate oncogenesis, embryogenesis and self-renewal. In turn, these amplification loops lead to oncogenic addiction, stem cell maintenance, angiogenesis, and immune modu-

lation within tumor microenvironments [52, 53]. Several oncogenic pathways, including Wnt/β-catenin, Notch, TGF-β/FOXO cascades, support self-renewal capacity and anticancer drug resistance in cancer stem cells [54, 55]. In addition, recent studies have revealed the indispensable role of the IL-6-Stat3 signal cascade in stimulating cancer stem cell activities in coordination with NF-κB-dependent inflammatory signals derived from tumor cells and their microenvironment [56, 57]. Moreover, Hedgehog signals have been identified as sentinels linking oncogenic aberration with the developmental program of normal and cancer stem cells [58]. In this regards, our findings that MFG-E8 and IL-6 derived from TAM mediate self-renewal and anticancer drug resistance through activation of Stat3 and Hedgehog signals provide additional evidences that TAM play a critical role in activating distinct signals that are crucial to the maintenance of the stem cell properties of tumor cells.

Recent studies in human leukemia and lymphoma have suggested that tumor cells express the antigen CD47, which serves as a “don’t eat me” signal to tumor-associated macrophages by engaging their cognate receptor SIRT-1. Administration of a blocking antibody to CD47 induced macrophage phagocytosis of AML stem cells *in vitro* and in mouse models [59-61]. These findings provide the first evidence that macrophage phagocytosis serves as a critical mediator of tumor immunosurveillance against leukemia stem cells.

On the other hand, T cell immunoglobulin-mucin domain protein-3 (TIM-3), which is involved in apoptotic cell phagocytosis via recognition of phosphatidylserine, has been identified as a functional marker for dissecting acute leukemia stem cells from bulk tumor cells [62-64]. Furthermore, AML stem cells were eradicated by the administration of a TIM-3-depleting mAb [63]. Since TIM-3 expression is detectable in macrophages and dendritic cells upon stimulation with toll-like receptor ligands such as LPS, it is of great interest to examine whether TIM-3 is detected in tumor-associated myeloid cells and to determine the functional role of myeloid cell-derived TIM-3 and its phagocytic activity in the regulation of cancer stem cell functions.

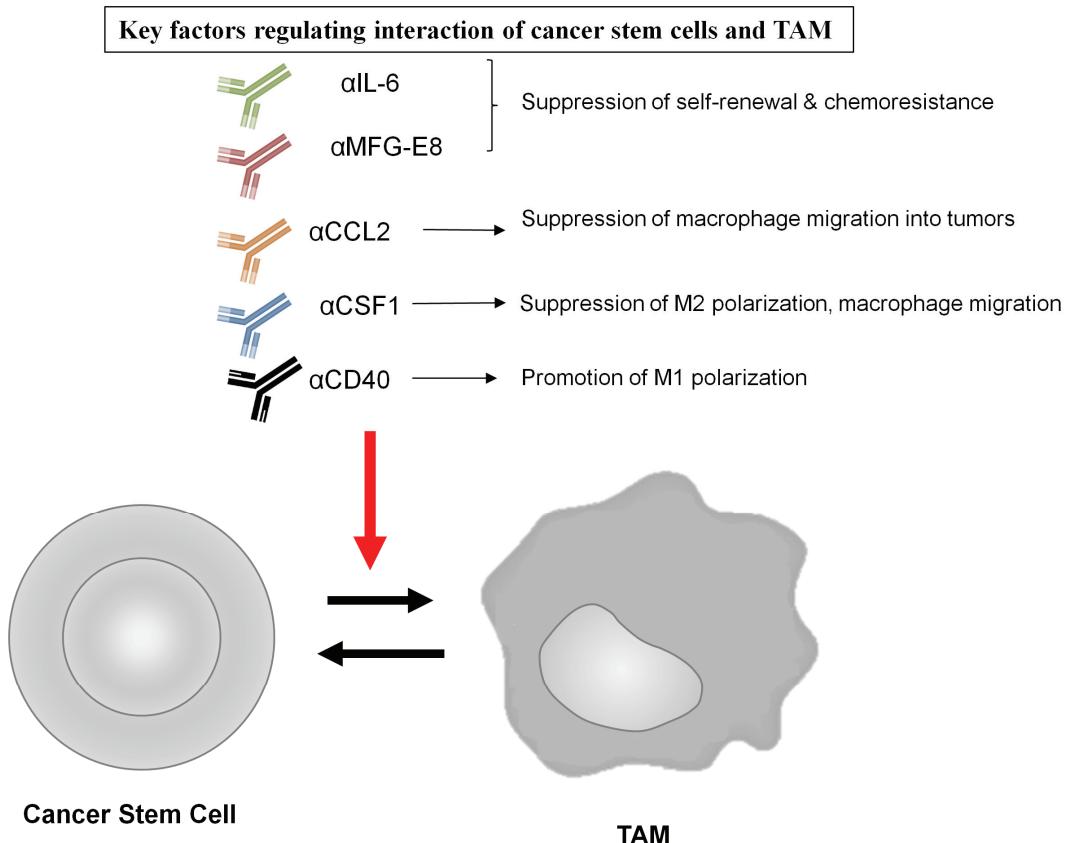
In addition, MFG-E8 not only serves as a positive modulator of cancer stem cell activities, but

also functions as an immunoregulatory factor within tumor microenvironments by promoting apoptotic cell phagocytosis and inducing Foxp3<sup>+</sup> infiltration into tumors [65, 66]. Moreover, TIM-4, expressed mainly on activated myeloid cells is also critically involved in the phagocytosis of apoptotic cells via the recognition of phosphatidylserine and the triggering of immune tolerance [67, 68]. The TAM receptor tyrosine kinase family composed of Axl, Mer-tyrosine kinase and Tyro-3, which serve as phagocytic receptors for apoptotic cells via recognition of Gas6, regulates innate immune responses and could be involved in the tumorigenic potentials of cancer cells [69, 70]. It is therefore likely that distinct sets of phagocytosis-associated molecules, such as CD47 / SIRT1, TIM-3, TIM-4, MFG-E8, Gas-6 etc. recognize distinct tumor subtypes including cancer stem cells, which arise from different backgrounds of oncogenic or epigenetic alterations and drug responsiveness. The identification and characterization of distinct sets of receptor / ligands on phagocytic macrophages may be an ideal strategy with which to investigate the interaction of cancer stem cells and TAM, and may lead to the exploration of new therapeutic targets against cancer stem cells.

#### **Therapeutic implication for targeting the interaction of cancer stem cells and TAM**

Comprehensive genetic approaches along with advances in the field of stem cell biology facilitated the identification of cancer stem cell-specific markers and multiple pathways potentially suitable for specifically targeting cancer stem cells [71, 72]. However, whether the target molecules identified from “pure” populations in cancer stem cells are actually effective against recurrent and multidrug-resistant variants of tumors remains largely uncharacterized. Thus, the development of drugs targeting the molecular networks between cancer stem cells and macrophages should provide useful tools with which to regulate cancer stem cell activities in coordination with those drugs targeting cancer stem cells and other factors derived from MSC, endothelial cells, fibroblasts and extracellular matrixes.

In addition, the targeting of TAM-derived downstream factors such as MFG-E8 and IL-6 may also be useful in repressing the emergence of chemoresistant tumors by controlling cancer



**Figure 3.** The strategy for targeting key mediators that influence the tumorigenic activities of TAM such as CCL2, M-CSF, CD40L and PPA has a potential to efficiently control cancer stem cell activities and might overcome the therapeutic limitations of conventional anticancer modalities.

stem cell activities [51, 73]. Moreover, therapeutic approach targeting TAM and their downstream effectors are now at the initial stages of clinical applications. Recruitment is a key determinant sustaining macrophage numbers at sites of inflammation and immunity. The CCL2/MCP-1 pathway has emerged as a new target for the prevention of myeloid cell recruitment to tumor microenvironments [74, 75]. CSF-1 receptor (c-fms) kinase inhibitors have been recently introduced into the clinical arena and exhibit antitumor and antiangiogenic activity in various murine and human tumor models [50, 76]. Furthermore, anti-CSF-1 antibodies augmented the antitumor effects of cytotoxic chemotherapy in a human breast cancer xenograft model [77].

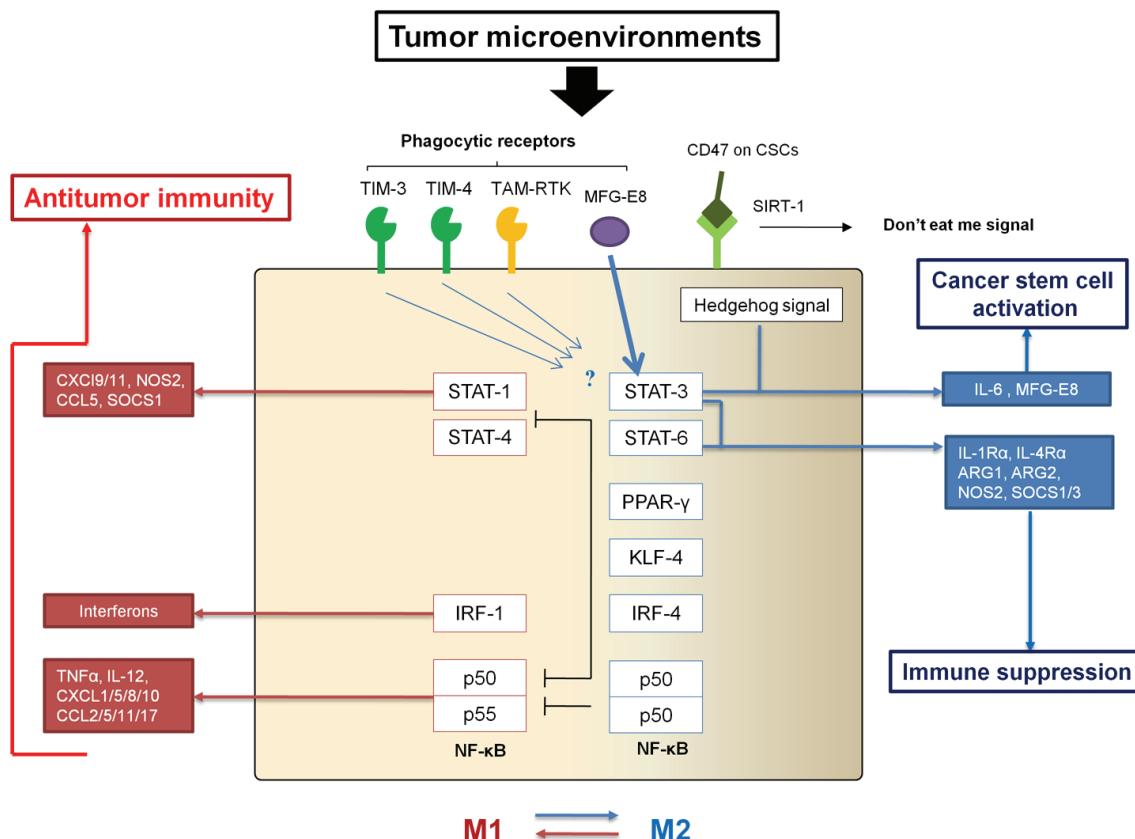
The manipulation of macrophage polarization may serve as another strategy for controlling tumorigenicity. Consistent with this concept,

CD40 agonist antibodies induce high expression of M1 markers in macrophages and augment antitumor responses to chemotherapy in pancreatic adenocarcinoma models [78]. Other therapeutic strategies that have been reported to affect macrophage polarization include PPAR- $\gamma$  agonist and TLR ligands (Poly (I: C) and CpG) [79-81].

Together, these findings validate the potential applicability of targeting key mediators that influence the tumorigenic activities of TAM, and suggest that this may negatively regulate cancer stem cell activities and overcome the therapeutic limitations of conventional anticancer modalities (**Figure 3**).

## Concluding remarks

We present the overview that TAM serve as a critical immunological niche in regulating cancer



**Figure 4.** Tumor microenvironments regulate distinct signal cascades that are critical for determining macrophage polarization and facilitating the expression of key molecules that control interactions with cancer stem cells.

stem cell functions (Figure 4). Accumulating evidence reveals that the quality of tumor microenvironments may determine the direction of interplay of tumors and immune cells throughout the different stages of carcinogenesis. In addition, tumor-infiltrating immune cells other than TAM, such as MDSC, dendritic cell, granulocytes, NKT cells, B cells and CD4<sup>+</sup> T cells, also serve as positive regulators of tumor progression and metastasis, raising the possibility that various sets of immune cells interact with cancer stem cells to modulate their biological activities [10]. In this regard, comprehensive analysis of molecular intersection between intrinsic and immune-mediated pathways enriched in tumor microenvironments should provide useful insights into the regulatory mechanisms of cancer stem cell activities as well as provide new therapeutic approaches for targeting the components of immunological niche in the future.

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**Address correspondence to:** Dr. Masahisa Jinushi, Research Center for Infection-associated cancer, Institute for Genetic Medicine, Hokkaido University Tel: +81-11-706-6073; Fax: +81-11-706-6071; E-mail: jinushi@igm.hokudai.ac.jp (M.J.)

#### References

- [1] Hanahan D and Weinberg RA. Hallmarks of cancer: the next generation. *Cell* 2011; 144: 646-674.
- [2] Grivennikov SI, Greten FR and Karin M. Immunity, inflammation, and cancer. *Cell* 2010; 140: 883-899.
- [3] Coussens LM and Werb Z. Inflammation and cancer. *Nature* 2002; 420: 860-867.
- [4] Gilbert LA and Hemann MT. DNA damage-

- mediated induction of a chemoresistant niche. *Cell* 2010; 143: 355-366.
- [5] Lord CJ and Ashworth A. The DNA damage response and cancer therapy. *Nature* 2012; 481: 287-294.
- [6] Schreiber RD, Old LJ and Smyth MJ. Cancer immunoediting: integrating immunity's roles in cancer suppression and promotion. *Science* 2011; 331: 1565-1570.
- [7] Mellman I, Coukos G and Dranoff G. Cancer immunotherapy comes of age. *Nature* 2011; 480: 480-489.
- [8] Vanneman M and Dranoff G. Combining immunotherapy and targeted therapies in cancer treatment. *Nat Rev Cancer* 2012; 12: 237-251.
- [9] Shiao SL, Ganeshan AP, Rugo HS and Coussens LM. Immune microenvironments in solid tumors: new targets for therapy. *Genes Dev* 2011; 25: 2559-2572.
- [10] Coussens LM and Pollard JW. Leukocytes in mammary development and cancer. *Cold Spring Harb Perspect Biol* 2011; 3: pii: a003285.
- [11] Zitvogel L, Apetoh L, Ghiringhelli F, Andre F, Tesniere A and Kroemer G. The anticancer immune response: indispensable for therapeutic success? *J Clin Invest* 2008; 118: 1991-2001.
- [12] Qian BZ and Pollard JW. Macrophage diversity enhances tumor progression and metastasis. *Cell* 2010; 141: 39-51.
- [13] Pollard JW. Tumour-educated macrophages promote tumour progression and metastasis. *Nat Rev Cancer* 2004; 4: 71-78.
- [14] Biswas SK and Mantovani A. Macrophage plasticity and interaction with lymphocyte subsets: cancer as a paradigm. *Nat Immunol* 2010; 11: 889-896.
- [15] Liu S, Ginestier C, Ou SJ, Clouthier SG, Patel SH, Monville F, Korkaya H, Heath A, Dutcher J, Kleer CG, Jung Y, Dontu G, Taichman R and Wicha MS. Breast cancer stem cells are regulated by mesenchymal stem cells through cytokine networks. *Cancer Res* 2011; 71: 614-624.
- [16] Korkaya H, Liu S and Wicha MS. Breast cancer stem cells, cytokine networks, and the tumor microenvironment. *J Clin Invest* 2011; 121: 3804-3809.
- [17] Karnoub AE, Dash AB, Vo AP, Sullivan A, Brooks MW, Bell GW, Richardson AL, Polyak K, Tubo R and Weinberg RA. Mesenchymal stem cells within tumour stroma promote breast cancer metastasis. *Nature* 2007; 449: 557-563.
- [18] Corre J, Labat E, Espagnolle N, Hebraud B, Avet-Loiseau H, Roussel M, Huynh A, Gadelorge M, Cordelier P, Klein B, Moreau P, Facon T, Fournie JJ, Attal M and Bourin P. Bioactivity and Prognostic Significance of Growth Differentiation Factor GDF15 Secreted by Bone Marrow Mesenchymal Stem Cells in Multiple Myeloma. *Cancer Res* 2012; 72: 1395-1406.
- [19] Pittenger MF, Mackay AM, Beck SC, Jaiswal RK, Douglas R, Mosca JD, Moorman MA, Simonetti DW, Craig S and Marshak DR. Multilineage potential of adult human mesenchymal stem cells. *Science* 1999; 284: 143-147.
- [20] Erez N, Truitt M, Olson P, Arron ST and Hanahan D. Cancer-Associated Fibroblasts Are Activated in Incipient Neoplasia to Orchestrate Tumor-Promoting Inflammation in an NF-kappaB-Dependent Manner. *Cancer Cell* 2010; 17: 135-147.
- [21] Orimo A, Gupta PB, Sgroi DC, Arenzana-Seisdedos F, Delaunay T, Naeem R, Carey VJ, Richardson AL and Weinberg RA. Stromal fibroblasts present in invasive human breast carcinomas promote tumor growth and angiogenesis through elevated SDF-1/CXCL12 secretion. *Cell* 2005; 121: 335-348.
- [22] Kojima Y, Acar A, Eaton EN, Mellody KT, Scheel C, Ben-Porath I, Onder TT, Wang ZC, Richardson AL, Weinberg RA and Orimo A. Autocrine TGF-beta and stromal cell-derived factor-1 (SDF-1) signaling drives the evolution of tumor-promoting mammary stromal myofibroblasts. *Proc Natl Acad Sci USA* 2010; 107: 20009-20014.
- [23] Vermeulen L, De SEMF, van der HM, Cameron K, de Jong JH, Borovski T, Tuynman JB, Todaro M, Merz C, Rodermond H, Sprick MR, Kemper K, Richel DJ, Stassi G and Medema JP. Wnt activity defines colon cancer stem cells and is regulated by the microenvironment. *Nat Cell Biol* 2010; 12: 468-476.
- [24] Watt FM and Collins CA. Role of beta-catenin in epidermal stem cell expansion, lineage selection, and cancer. *Cold Spring Harb Symp Quant Biol* 2008; 73: 503-512.
- [25] Hoey T, Yen WC, Axelrod F, Basi J, Donigian L, Dylla S, Fitch-Bruhns M, Lazetic S, Park IK, Sato A, Satyal S, Wang X, Clarke MF, Lewicki J and Gurney A. DLL4 blockade inhibits tumor growth and reduces tumor-initiating cell frequency. *Cell Stem Cell* 2009; 5: 168-177.
- [26] Anderberg C, Li H, Fredriksson L, Andrae J, Betsholtz C, Li X, Eriksson U and Pietras K. Paracrine signaling by platelet-derived growth factor-CC promotes tumor growth by recruitment of cancer-associated fibroblasts. *Cancer Res* 2009; 69: 369-378.
- [27] Finak G, Bertos N, Pepin F, Sadekova S, Souleimanova M, Zhao H, Chen H, Omeroglu G, Meterissian S, Omeroglu A, Hallett M and Park M. Stromal gene expression predicts clinical outcome in breast cancer. *Nat Med* 2008; 14: 518-527.
- [28] Bhati R, Patterson C, Livasy CA, Fan C, Ketelsen D, Hu Z, Reynolds E, Tanner C, Moore DT, Gabrielli F, Perou CM and Klauber-DeMore N. Molecular characterization of human breast tumor vascular cells. *Am J Pathol* 2008; 172: 1381-1390.
- [29] Ferrara N. VEGF and the quest for tumour angiogenesis factors. *Nat Rev Cancer* 2002; 2: 795-803.

- [30] Hamerlik P, Lathia JD, Rasmussen R, Wu Q, Bartkova J, Lee M, Moudry P, Bartek J Jr, Fischer W, Lukas J, Rich JN and Bartek J. Autocrine VEGF-VEGFR2-Neuropilin-1 signaling promotes glioma stem-like cell viability and tumor growth. *J Exp Med* 2012; 209: 507-520.
- [31] Okamoto R, Ueno M, Yamada Y, Takahashi N, Sano H, Suda T and Takakura N. Hematopoietic cells regulate the angiogenic switch during tumorigenesis. *Blood* 2005; 105: 2757-2763.
- [32] Wang R, Chadalavada K, Wilshire J, Kowalik U, Hovinga KE, Geber A, Fligelman B, Leversha M, Brennan C and Tabar V. Glioblastoma stem-like cells give rise to tumour endothelium. *Nature* 2010; 468: 829-833.
- [33] Ricci-Vitiani L, Pallini R, Biffoni M, Todaro M, Invernici G, Cencio T, Maira G, Parati EA, Stassi G, Larocca LM and De MR. Tumour vascularization via endothelial differentiation of glioblastoma stem-like cells. *Nature* 2010; 468: 824-828.
- [34] Gabrilovich DI and Nagaraj S. Myeloid-derived suppressor cells as regulators of the immune system. *Nat Rev Immunol* 2009; 9: 162-174.
- [35] Mosser DM and Edwards JP. Exploring the full spectrum of macrophage activation. *Nat Rev Immunol* 2008; 8: 958-969.
- [36] Mantovani A, Sozzani S, Locati M, Allavena P and Sica A. Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes. *Trends Immunol* 2002; 23: 549-555.
- [37] Gordon S and Martinez FO. Alternative activation of macrophages: mechanism and functions. *Immunity* 2010; 32: 593-604.
- [38] Krausgruber T, Blazek K, Smallie T, Alzabin S, Lockstone H, Sahgal N, Hussell T, Feldmann M and Udalova IA. IRF5 promotes inflammatory macrophage polarization and TH1-TH17 responses. *Nat Immunol* 2011; 12: 231-238.
- [39] Hagemann T, Lawrence T, McNeish I, Charles KA, Kulbe H, Thompson RG, Robinson SC and Balkwill FR. "Re-educating" tumor-associated macrophages by targeting NF-kappaB. *J Exp Med* 2008; 205: 1261-1268.
- [40] Szanto A, Balint BL, Nagy ZS, Barta E, Dezso B, Pap A, Szeles L, Poliska S, Oros M, Evans RM, Barak Y, Schwabe J and Nagy L. STAT6 transcription factor is a facilitator of the nuclear receptor PPARgamma-regulated gene expression in macrophages and dendritic cells. *Immunity* 2010; 33: 699-712.
- [41] Liao X, Sharma N, Kapadia F, Zhou G, Lu Y, Hong H, Paruchuri K, Mahadeleshwar GH, Dalmas E, Venteclef N, Flask CA, Kim J, Doreian BW, Lu KQ, Kaestner KH, Hamik A, Clement K and Jain MK. Kruppel-like factor 4 regulates macrophage polarization. *J Clin Invest* 2011; 121: 2736-2749.
- [42] Satoh T, Takeuchi O, Vandenberg A, Yasuda K, Tanaka Y, Kumagai Y, Miyake T, Matsushita K, Okazaki T, Saitoh T, Honma K, Matsuyama T, Yui K, Tsujimura T, Standley DM, Nakanishi K, Nakai K and Akira S. The JmjD3-Irf4 axis regulates M2 macrophage polarization and host responses against helminth infection. *Nat Immunol* 2010; 11: 936-944.
- [43] Zaynagetdinov R, Sherrill TP, Polosukhin IV, Han W, Ausborn JA, McLoed AG, McMahon FB, Gleaves LA, Degryse AL, Stathopoulos GT, Yull FE and Blackwell TS. A critical role for macrophages in promotion of urethane-induced lung carcinogenesis. *J Immunol* 2011; 187: 5703-5711.
- [44] Kang TW, Yevsa T, Woller N, Hoenicke L, Wuestefeld T, Dauch D, Hohmeyer A, Gereke M, Rudalska R, Potapova A, Iken M, Vucur M, Weiss S, Heikenwalder M, Khan S, Gil J, Bruder D, Manns M, Schirmacher P, Tacke F, Ott M, Luedde T, Longerich T, Kubicka S and Zender L. Senescence surveillance of pre-malignant hepatocytes limits liver cancer development. *Nature* 2011; 479: 547-551.
- [45] Saccani A, Schioppa T, Porta C, Biswas SK, Nebuloni M, Vago L, Bottazzi B, Colombo MP, Mantovani A and Sica A. p50 nuclear factor-kappaB overexpression in tumor-associated macrophages inhibits M1 inflammatory responses and antitumor resistance. *Cancer Res* 2006; 66: 11432-11440.
- [46] Guiducci C, Vicari AP, Sangaletti S, Trinchieri G and Colombo MP. Redirecting in vivo elicited tumor infiltrating macrophages and dendritic cells towards tumor rejection. *Cancer Res* 2005; 65: 3437-3446.
- [47] Ojalvo LS, King W, Cox D and Pollard JW. High-density gene expression analysis of tumor-associated macrophages from mouse mammary tumors. *Am J Pathol* 2009; 174: 1048-1064.
- [48] Movahedi K, Laoui D, Gysemans C, Baeten M, Stange G, Van BJ, Mack M, Pipeleers D, In't VP, De BP and Van GJA. Different tumor microenvironments contain functionally distinct subsets of macrophages derived from Ly6C(high) monocytes. *Cancer Res* 2010; 70: 5728-5739.
- [49] Steidl C, Lee T, Shah SP, Farinha P, Han G, Nayar T, Delaney A, Jones SJ, Iqbal J, Weisenburger DD, Bast MA, Rosenwald A, Muller-Hermelink HK, Rimsza LM, Campo E, Delabie J, Braziel RM, Cook JR, Tubbs RR, Jaffe ES, Lenz G, Connors JM, Staudt LM, Chan WC and Gascoyne RD. Tumor-associated macrophages and survival in classic Hodgkin's lymphoma. *N Engl J Med* 2010; 362: 875-885.
- [50] Denardo DG, Brennan DJ, Rexhepaj E, Ruffell B, Shiao SL, Madden SF, Gallagher WM, Wadhwani N, Keil SD, Junaid SA, Rugo HS, Hwang ES, Jirstrom K, West BL and Coussens LM. Leukocyte Complexity Predicts Breast Cancer Survival and Functionally Regulates Response to Chemotherapy. *Cancer Discov* 2011; 1: 54-67.
- [51] Jinushi M, Chiba S, Yoshiyama H, Masutomi K,

- Kinoshita I, Dosaka-Akita H, Yagita H, Takaoka A and Tahara H. Tumor-associated macrophages regulate tumorigenicity and anticancer drug responses of cancer stem/initiating cells. *Proc Natl Acad Sci USA* 2011; 108: 12425-12430.
- [52] Scheel C, Eaton EN, Li SH, Chaffer CL, Reinhardt F, Kah KJ, Bell G, Guo W, Rubin J, Richardson AL and Weinberg RA. Paracrine and autocrine signals induce and maintain mesenchymal and stem cell states in the breast. *Cell* 2011; 145: 926-940.
- [53] Naka K, Hoshii T, Muraguchi T, Tadokoro Y, Ooshio T, Kondo Y, Nakao S, Motoyama N and Hirao A. TGF-beta-FOXO signalling maintains leukaemia-initiating cells in chronic myeloid leukaemia. *Nature* 2010; 463: 676-680.
- [54] Sykes SM, Lane SW, Bullinger L, Kalaitzidis D, Yusuf R, Saez B, Ferraro F, Mercier F, Singh H, Brumme KM, Acharya SS, Scholl C, Tothova Z, Attar EC, Frohling S, DePinho RA, Armstrong SA, Gilliland DG and Scadden DT. AKT/FOXO signaling enforces reversible differentiation blockade in myeloid leukemias. *Cell* 2011; 146: 697-708.
- [55] Zhou J, Wulfkuhle J, Zhang H, Gu P, Yang Y, Deng J, Margolick JB, Liotta LA, Petricoin E3 and Zhang Y. Activation of the PTEN/mTOR/STAT3 pathway in breast cancer stem-like cells is required for viability and maintenance. *Proc Natl Acad Sci USA* 2007; 104: 16158-16163.
- [56] Marotta LL, Almendro V, Marusyk A, Shipitsin M, Schemme J, Walker SR, Bloushtain-Qimron N, Kim JJ, Choudhury SA, Maruyama R, Wu Z, Gonen M, Mulvey LA, Bessarabova MO, Huh SJ, Silver SJ, Kim SY, Park SY, Lee HE, Anderson KS, Richardson AL, Nikolskaya T, Nikolsky Y, Liu XS, Root DE, Hahn WC, Frank DA and Polyak K. The JAK2/STAT3 signaling pathway is required for growth of CD44CD24 stem cell-like breast cancer cells in human tumors. *J Clin Invest* 2011; 121: 2723-2735.
- [57] Iliopoulos D, Hirsch HA, Wang G, Struhl K. Inducible formation of breast cancer stem cells and their dynamic equilibrium with non-stem cancer cells via IL6 secretion. *Proc Natl Acad Sci USA* 2011; 108: 1397-1402.
- [58] Zhao C, Chen A, Jamieson CH, Fereshteh M, Abrahamsson A, Blum J, Kwon HY, Kim J, Chute JP, Rizzieri D, Munchhof M, VanAarsdale T, Beachy PA and Reya T. Hedgehog signalling is essential for maintenance of cancer stem cells in myeloid leukaemia. *Nature* 2009; 458: 776-779.
- [59] Jaiswal S, Jamieson CH, Pang WW, Park CY, Chao MP, Majeti R, Traver D, van Rooijen N and Weissman IL. CD47 is upregulated on circulating hematopoietic stem cells and leukemia cells to avoid phagocytosis. *Cell* 2009; 138: 271-285.
- [60] Majeti R, Chao MP, Alizadeh AA, Pang WW, Jaiswal S, Gibbs KDJ, van Rooijen N and Weissman IL. CD47 is an adverse prognostic factor and therapeutic antibody target on human acute myeloid leukemia stem cells. *Cell* 2009; 138: 286-299.
- [61] Chao MP, Alizadeh AA, Tang C, Myklebust JH, Varghese B, Gill S, Jan M, Cha AC, Chan CK, Tan BT, Park CY, Zhao F, Kohrt HE, Malumbres R, Briones J, Gascoyne RD, Lossos IS, Levy R, Weissman IL and Majeti R. Anti-CD47 antibody synergizes with rituximab to promote phagocytosis and eradicate non-Hodgkin lymphoma. *Cell* 2010; 142: 699-713.
- [62] Nakayama M, Akiba H, Takeda K, Kojima Y, Hashiguchi M, Azuma M, Yagita H and Okumura K. Tim-3 mediates phagocytosis of apoptotic cells and cross-presentation. *Blood* 2009; 113: 3821-3830.
- [63] Kikushige Y, Shima T, Takayanagi S, Urata S, Miyamoto T, Iwasaki H, Takenaka K, Teshima T, Tanaka T, Inagaki Y and Akashi K. TIM-3 is a promising target to selectively kill acute myeloid leukemia stem cells. *Cell Stem Cell* 2010; 7: 708-717.
- [64] Jan M, Chao MP, Cha AC, Alizadeh AA, Gentles AJ, Weissman IL and Majeti R. Prospective separation of normal and leukemic stem cells based on differential expression of TIM3, a human acute myeloid leukemia stem cell marker. *Proc Natl Acad Sci USA* 2011; 108: 5009-5014.
- [65] Hanayama R, Tanaka M, Miyasaka K, Aozasa K, Koike M, Uchiyama Y and Nagata S. Autoimmune disease and impaired uptake of apoptotic cells in MFG-E8-deficient mice. *Science* 2004; 304: 1147-1150.
- [66] Jinushi M, Nakazaki Y, Dougan M, Carrasco DR, Mihm M and Dranoff G. MFG-E8-mediated uptake of apoptotic cells by APCs links the pro- and antiinflammatory activities of GM-CSF. *J Clin Invest* 2007; 117: 1902-1913.
- [67] Kobayashi N, Karisola P, Pena-Cruz V, Dorfman DM, Jinushi M, Umetsu SE, Butte MJ, Nagumo H, Chernova I, Zhu B, Sharpe AH, Ito S, Dranoff G, Kaplan GG, Casasnovas JM, Umetsu DT, Dekruyff RH and Freeman GJ. TIM-1 and TIM-4 glycoproteins bind phosphatidylserine and mediate uptake of apoptotic cells. *Immunity* 2007; 27: 927-940.
- [68] Miyanishi M, Tada K, Koike M, Uchiyama Y, Kitamura T and Nagata S. Identification of Tim4 as a phosphatidylserine receptor. *Nature* 2007; 450: 435-439.
- [69] Linger RM, Keating AK, Earp HS and Graham DK. TAM receptor tyrosine kinases: biologic functions, signaling, and potential therapeutic targeting in human cancer. *Adv Cancer Res* 2008; 100: 35-83.
- [70] Rothlin CV, Ghosh S, Zuniga EI, Oldstone MBA, Lemke G. TAM receptors are pleiotrophic inhibitors of the innate immune response. *Cell* 2007; 131: 1124-1136.
- [71] Gupta PB, Onder TT, Jiang G, Tao K, Kuper-

- wasser C, Weinberg RA and Lander ES. Identification of selective inhibitors of cancer stem cells by high-throughput screening. *Cell* 2009; 138: 645-659.
- [72] Liu S and Wicha MS. Targeting breast cancer stem cells. *J Clin Oncol* 2010; 28: 4006-4012.
- [73] Kim S, Takahashi H, Lin WW, Descargues P, Grivennikov S, Kim Y, Luo JL and Karin M. Carcinoma-produced factors activate myeloid cells through TLR2 to stimulate metastasis. *Nature* 2009; 457: 102-106.
- [74] Qian BZ, Li J, Zhang H, Kitamura T, Zhang J, Campion LR, Kaiser EA, Snyder LA and Pollard JW. CCL2 recruits inflammatory monocytes to facilitate breast-tumour metastasis. *Nature* 2011; 475: 222-225.
- [75] Loberg RD, Ying C, Craig M, Day LL, Sargent E, Neeley C, Wojno K, Snyder LA, Yan L, Pienta KJ. Targeting CCL2 with systemic delivery of neutralizing antibodies induces prostate cancer tumor regression *in vivo*. *Cancer Res* 2007; 67: 9417-9424.
- [76] Kubota Y, Takubo K, Shimizu T, Ohno H, Kishi K, Shibuya M, Saya H and Suda T. M-CSF inhibition selectively targets pathological angiogenesis and lymphangiogenesis. *J Exp Med* 2009; 206: 1089-1102.
- [77] Paulus P, Stanley ER, Schafer R, Abraham D and Aharinejad S. Colony-stimulating factor-1 antibody reverses chemoresistance in human MCF-7 breast cancer xenografts. *Cancer Res* 2006; 66: 4349-4356.
- [78] Beatty GL, Chiorean EG, Fishman MP, Saboury B, Teitelbaum UR, Sun W, Huhn RD, Song W, Li D, Sharp LL, Torigian DA, O'Dwyer PJ and Von derheide RH. CD40 agonists alter tumor stroma and show efficacy against pancreatic carcinoma in mice and humans. *Science* 2011; 331: 1612-1616.
- [79] Charo IF. Macrophage polarization and insulin resistance: PPARgamma in control. *Cell Metab* 2007; 6: 96-98.
- [80] Shirota Y, Shirota H and Klinman DM. Intratumoral injection of CpG oligonucleotides induces the differentiation and reduces the immunosuppressive activity of myeloid-derived suppressor cells. *J Immunol* 2012; 188: 1592-1599.
- [81] Shime H, Matsumoto M, Oshiumi H, Tanaka S, Nakane A, Iwakura Y, Tahara H, Inoue N and Seya T. Toll-like receptor 3 signaling converts tumor-supporting myeloid cells to tumoricidal effectors. *Proc Natl Acad Sci USA* 2012; 109: 2066-2071.