Original Article Analysis of difference of association between polymorphisms in the XRCC5, RPA3 and RTEL1 genes and glioma, astrocytoma and glioblastoma

Tianbo Jin^{1,2}, Yuan Wang³, Gang Li⁴, Shuli Du², Hua Yang², Tingting Geng², Peng Hou⁵, Yongkuan Gong¹

¹Key Laboratory of Synthetic and Natural Functional Molecule Chemistry of Ministry of Education, College of Chemistry and Materials Science, Northwest University, Xi'an 710069, China; ²National Engineering Research Center for Miniaturized Detection Systems, School of Life Sciences, Northwest University, Xi'an 710069, China; ³Department of Trauma, The Second Affiliated Hospital, Inner Mongolia Medical University, Hohhot 010030, China; ⁴Department of Neurosurgery, Tangdu Hospital, The Fourth Military Medical University, Xi'an 710038, China; ⁵Department of Endocrinology, The First Affiliated Hospital of Xi'an Jiaotong University School of Medicine, Xi'an 710061, China

Received May 29, 2015; Accepted June 10, 2015; Epub June 15, 2015; Published July 1, 2015

Abstract: Background: Gliomas are the most common aggressive brain tumors and have many complex pathological types. Previous reports have discovered that genetic mutations are associated with the risk of glioma. However, it is unclear whether uniform genetic mutations exist difference between glioma and its two pathological types in the Han Chinese population. Materials and methods: We evaluated 20 SNPs of 703 glioma cases (338 astrocytoma cases, 122 glioblastoma cases) and 635 controls in a Han Chinese population using χ^2 test and genetic model analysis. Results: In three case-control studies, we found rs9288516 in *XRCC5* gene showed a decreased risk of glioma (OR, 0.85; 95% CI, 0.73-0.99; *P* = 0.042) and glioblastoma (OR, 0.70; 95% CI, 0.52-0.92; *P* = 0.001) in the allele model. We identified rs414805 in *RPA3* gene showed an increased risk of glioblastoma in allele model (OR, 1.38; 95% CI, 1.00-1.89; *P* = 0.047) and dominant model (OR, 1.57; 95% CI, 1.05-2.35; *P* = 0.027), analysis respectively. Meanwhile, rs2297440 in *RTEL1* gene showed an increased risk of glioma (OR, 1.30; 95% CI, 1.10-1.54; *P* = 0.002) and astrocytoma (OR, 1.26; 95% CI, 1.02-1.54; *P* = 0.029) in the allele model. In addition, we also observed a haplotype of "GCT" in the *RTEL1* gene with an increased risk of astrocytoma (*P* = 0.005). Conclusions: Polymorphisms in the *XRCC5, RPA3* and *RTEL1* genes, combinating with previous reaserches, are associated with glioma developing. However, those genes mutations may play different roles in the glioma, astrocytoma and glioblastoma, respectively.

Keywords: XRCC5, RPA3, RTEL1, glioma, astrocytoma, glioblastoma, case-control study

Introduction

Glial cell is regard as the origin of glioma [1]. Complex subtypes of glioma exist including World Health Organization classification astrocytoma grades I, II (astrocytoma), III (anaplastic astrocytoma), and IV (glioblastoma), oligodendrogliomas, ependy momas, and mixed gliomas [2]. Glioma is the most common and aggressive type of brain tumor and its morbidity rate is approximate six per 100,000 each year [3, 4].

Affecting glioma risk has been only found a few factors, such as family history, genetic syn-

dromes and exposure to ionizing radiation [5, 6]. Although the etiology of gliomas have not been illuminated clearly so far, increasing data indicated that genetic variants have momentous effect on the kind of tumor [7]. Some genetic mutations in glioma have been known for years, such as *EGFR*, *CCDC26*, *TREH*, *XRCC1* and *GSTP1* gene [8-10]. It is previously reported that *TEL1*, *TERT* gene were associated with susceptibility to astrocytoma [11]. Recent studies demonstrated that mutations in *FLT3*, *EGFR*, *NEIL3* and *ALOX5* genes were associated with glioblastoma survival [12]. However, we found genes are alone with a separate analysis of glioma, glioblastoma, astrocytoma.

Pathology	Parameters	Cases	Healthy controls	P value
Gliomas	Male/female	385 (54.8%)/318 (45.2%)	311 (49.0%)/324 (51.0%)	0.34
	Age (years)	42.2 ± 17.4	38.0 ± 16.5	
	Total (N)	703	635	
Astrocytoma	Male/female	191 (56.5%)/147 (43.5%)	311 (49.0%)/324 (51.0%)	0.25
	Age (years)	42.5 ± 16.7	38.0 ± 16.5	
	Total (N)	338	635	
Glioblastoma	Male/female	70 (57.4%)/52 (42.6%)	311 (49.0%)/324 (51.1%)	0.89
	Age (years)	46.9 ± 15.1	38.0 ± 16.6	
	Total (N)	122	636	

 Table 1. Characteristics of patients with cases and controls

P value is based on the age and sex versus healthy controls in the study.

In our study, we selected 20 SNPs in fifteen genes which have previously been reported to be associated with glioma, astrocytoma or glioblastoma onset in European. We conducted three case-controls from 703 cases and 635 controls: 1) glioma case-control (703 cases and 635 controls); 2) astrocytoma case-control (338 cases and 635 controls); 3) glioblastoma case-control (122 cases and 635 controls). A denotative association analysis was performed in a Han Chinese population by three case-control studies. The aim of the study was to investigate the different influence between mutations of *XRCC5, RPA3* and *RTEL1* genes and glioma, astrocytoma and glioblastoma, respectively.

Materials and methods

Ethics statement

The protocol in this study was cautiously affirmed to the principles of the Declaration of Helsiinki and was ratified by the Ethical Committee of Tangdu Hospital. The paticipants all had signed informed consents.

Study population

A total of 703 patients with glioma, includes astrocytoma 338 patients and glioblastoma 122 patients, between December 2010 and November 2014 were recruited from the department of Neurosurgery at Tangdu Hospital, all of the study participants are Han Chinese living in the area of Xi'an, China. Confirmed cases who were newly diagnosed and histologically ensured. All glioma cases never undergone radiotherapy, chemotherapy and cancer. According to WHO classifications [2], all the pathologies of glioma tissues were reevaluated. The clinical pathology and characteristics of all the patients were indicated in Table 1.

According to the recruitment and exclusion standards, the controls were 635 healthy individuals who be selected from June 2011 to October 2014 from the medical examination center, Tangdu Hospital. The controls were all Han Chinese living in Xi'an city and around area. Meanwhile, we also excluded subjucts with chronic diseases of kidney, heart, liver and brain by detailed exclusion criteria. The factors, environmental and therapeutic, may contribute to mutate, and we maximized the data in the study to be more persuative. Finally, we selected 635 unrelated healthy controls in this study.

Genotyping

Combinating genome-wide association studies (GWAS), a powerful research strategy, used to identify susceptibility genes which previously reported to be associated with glioma, astrocytoma and glioblastoma risk [8-12], we genotyped twenty tSNPs with minor allele frequency (MAF) > 5% in HapMap Asian population in fifteen genes. Genomic DNA was stored at -20°C and was extracted from whole blood by the phenol-chloroform extraction method. Using an extraction kit (GoldMag, China), we ioslated DNA from the samples. DNA concentration was measured by spectrometry (DU530 UV/VIS spectrophotometer, Beckman Instruments, Fullerton, CA, USA). We designed Multiplexed SNP Mass EXTEND assay by Sequenom MassARRAY Assay Design 4.0 Software [13]. Using the Sequenom Mass ARRAY RS1000, genotyped SNP was recommended by the manufacturer with a standard protocol [14]. Data

		Position	Gene	HWE P value	Alleles A/B	MAF control	MAF case		
SNP ID	Chromosome						Glioma	Astrocytoma	Glioblastoma
rs12022378	1	114448389	DCLRE1B	0.546	C/T	0.358	0.391	0.370	0.402
rs1800871	1	206946634	IL10	0.111	C/T	0.342	0.349	0.356	0.316
rs3770502	2	217045059	XRCC5	0.754	A/G	0.149	0.164	0.169	0.172
rs9288516	2	217053264	XRCC5	0.633	A/T	0.479	0.440	0.438	0.389
rs12645561	4	178260872	NEIL3	0.111	T/C	0.275	0.268	0.254	0.287
rs2853676	5	1288547	TERT	0.03	A/G	0.165	0.223	0.232	0.209
rs2243248	5	132008644	IL4	1	G/T	0.061	0.056	0.047	0.057
rs2070874	5	132009710	IL4	0.47	C/T	0.208	0.220	0.217	0.209
rs1801270	6	36651971	CDKN1A	0.686	A/C	0.427	0.458	0.436	0.471
rs4140805	7	7727101	RPA3	0.622	G/T	0.202	0.195	0.189	0.258
rs6947203	7	7737048	RPA3	0.719	T/C	0.126	0.120	0.126	0.164
rs7003908	8	48770702	PRKDC	0.098	C/A	0.236	0.213	0.223	0.230
rs12917	10	131506283	MGMT	0.208	T/C	0.106	0.087	0.090	0.102
rs1695	11	67352689	GSTP1	0.349	G/A	0.217	0.204	0.222	0.189
rs1042522	17	7579472	TP53	0.333	C/G	0.431	0.435	0.428	0.467
rs2952155	17	37861718	ERBB2	0.749	C/T	0.460	0.476	0.476	0.492
rs2992	19	4443046	UBXN6	0.376	A/G	0.438	0.435	0.442	0.430
rs6010620	20	62309839	RTEL1	0.189	G/A	0.269	0.321	0.314	0.320
rs2297440	20	62312299	RTEL1	0.264	C/T	0.265	0.320	0.312	0.316
rs4809324	20	62318220	RTEL1	0.699	C/T	0.115	0.116	0.111	0.090

Table 2. Basic information of candidate SNPs in this study

sSNP, single-nucleotide polymorphism, A/B stands for minor/major alleles on the control sample frequencies. HWE, Hardy-Weinberg equilibrium, The SNPs are excluded at 5% HWE P level.

analyses and management were conducted by Sequenom Typer 4.0 Software [15].

Statistical analysis

The data analysis was used by SPSS 16.0 statistical package (SPSS, Chicago, IL) and microsoft Excel. We excluded the *P* value which $P \ge$ 0.05 was considered the deviation value of statistical significance. Each SNP of the genotype frequencies in control subjects were checked by using Hardy-Weinberg equilibrium (HWE). The genotype frequencies of cases and controls were calculated by using χ^2 test [16, 17]. Odds ratios (ORs) and 95% confidence intervals (CIs) were tested by using unconditional logistic regression analysis with adjustment for age and gender [18]. The three genetic models (allele, dominant and Log-additive) were applied by PLINK software (http://pngu.mgh.harvard. edu/purcell/plink/) to assess the association of SNP with the risk of glioma, astrocytoma and glioblastoma. Finally, we analysed haplotype construction, and genetic association at polymorphism loci by the SHEsis software platform (www.nhgg.org/analysis/) [19].

Results

In this study, 703 glioma patients contained 385 males and 318 females which those cases diagnosed mean age was 42.2 ± 17.4 years. The 703 cases included 338 cases of astrocytoma, 122 cases of glioblastoma, 35 cases of ependymoma, 24 cases of oligodendrogliomas, and 98 cases of other glioma types. The 635 healthy controls included 311 males and 324 females which those controls diagnosed mean age was 38.0 ± 16.5 years. We found no differences between gender and age distribution by p value. The Characteristics of patients with cases and controls are shown in Table 1. 20 SNPs in the fifteen genes were analyzed in this study. Chromosomal position, gene, Allele, HWE test results and MAF of cases and controls of all the SNPs were appeared in Table 2. The minor allele of each SNP, a risk factor, was compared to the wild-type allele. A total of 20 SNPs were conducted in patients and controls, and rs2853676 were cut off at 3% HWE P level.

Further model association analyses used logistic tests were presented in **Table 3**. rs9288516 was observed to be associated with the

Madal	Construct	Glioma		Astrocytoma		Glioblastoma	
wouer	Genetype	OR (95% CI)	P value	OR (95% CI)	P value	OR (95% CI)	P Value
Allele model	A/T	0.85 (0.73-0.99)	0.042*	0.85 (0.70-1.02)	0.08	0.70 (0.52-0.92)	0.001*
Dominant model	A/T-A/A	0.85 (0.67-1.08)	0.2	0.90 (0.67-1.21)	0.48	0.70 (0.46-1.06)	0.096
Log-additive model		0.87 (0.74-1.01)	0.07	0.86 (0.71-1.04)	0.13	0.72 (0.55-0.96)	0.026*
Allele model	G/T	0.96 (0.79-1.16)	0.657	0.92 (0.73-1.17)	0.507	1.38 (1.00-1.89)	0.047*
Dominant model	G/T-G/G	0.94 (0.75-1.18)	0.62	0.92 (0.69-1.21)	0.54	1.57 (1.05-2.35)	0.027*
Log-additive model		0.94 (0.78-1.14)	0.56	0.92 (0.73-1.17)	0.52	1.36 (0.98-1.88)	0.07
Allele model	C/T	1.30 (1.10-1.54)	0.002*	1.26 (1.02-1.54)	0.029*	1.28 (0.95-1.72)	0.107
Dominant model	T/C-C/C	1.25 (1.01-1.56)	0.042*	1.28 (0.98-1.67)	0.073	1.12 (0.75-1.67)	0.57
Log-additive model		1.31 (1.10-1.55)	0.002*	1.28 (1.04-1.59)	0.022*	1.29 (0.95-1.77)	0.11
	Dominant model Log-additive model Allele model Dominant model Log-additive model Allele model Dominant model	Allele modelA/TDominant modelA/T-A/ALog-additive modelAllele modelG/TDominant modelG/T-G/GLog-additive modelAllele modelC/TDominant modelT/C-C/C	Model Genetype OR (95% Cl) Allele model A/T 0.85 (0.73-0.99) Dominant model A/T-A/A 0.85 (0.67-1.08) Log-additive model 0.87 (0.74-1.01) Allele model G/T 0.96 (0.79-1.16) Dominant model G/T-G/G 0.94 (0.75-1.18) Log-additive model 0.94 (0.78-1.14) Allele model C/T 1.30 (1.10-1.54) Dominant model T/C-C/C 1.25 (1.01-1.56)	Model Genetype OR (95% Cl) P value Allele model A/T 0.85 (0.73-0.99) 0.042* Dominant model A/T-A/A 0.85 (0.67-1.08) 0.2 Log-additive model 0.87 (0.74-1.01) 0.07 Allele model G/T 0.96 (0.79-1.16) 0.657 Dominant model G/T-G/G 0.94 (0.75-1.18) 0.62 Log-additive model 0.94 (0.78-1.14) 0.56 Allele model C/T 1.30 (1.10-1.54) 0.002* Dominant model T/C-C/C 1.25 (1.01-1.56) 0.042*	Model Genetype OR (95% Cl) P value OR (95% Cl) Allele model A/T 0.85 (0.73-0.99) 0.042* 0.85 (0.70-1.02) Dominant model A/T-A/A 0.85 (0.67-1.08) 0.2 0.90 (0.67-1.21) Log-additive model 0.87 (0.74-1.01) 0.07 0.86 (0.71-1.04) Allele model G/T 0.96 (0.79-1.16) 0.657 0.92 (0.73-1.17) Dominant model G/T-G/G 0.94 (0.78-1.14) 0.62 0.92 (0.69-1.21) Log-additive model 0.94 (0.78-1.14) 0.56 0.92 (0.73-1.17) Allele model C/T 1.30 (1.10-1.54) 0.002* 1.26 (1.02-1.54) Dominant model T/C-C/C 1.25 (1.01-1.56) 0.042* 1.28 (0.98-1.67)	Model Genetype OR (95% Cl) P value OR (95% Cl) P value Allele model A/T 0.85 (0.73-0.99) 0.042* 0.85 (0.70-1.02) 0.08 Dominant model A/T-A/A 0.85 (0.67-1.08) 0.2 0.90 (0.67-1.21) 0.48 Log-additive model 0.87 (0.74-1.01) 0.07 0.86 (0.71-1.04) 0.13 Allele model G/T 0.96 (0.79-1.16) 0.657 0.92 (0.73-1.17) 0.507 Dominant model G/T-G/G 0.94 (0.78-1.14) 0.62 0.92 (0.73-1.17) 0.524 Log-additive model 0.94 (0.78-1.14) 0.56 0.92 (0.73-1.17) 0.524 Log-additive model C/T 1.30 (1.10-1.54) 0.002* 1.26 (1.02-1.54) 0.029* Allele model C/T 1.25 (1.01-1.56) 0.042* 1.28 (0.98-1.67) 0.073	Model Genetype OR (95% Cl) P value OR (95% Cl) OR (95% Cl) OR (95% Cl) De value OR (0.52-0.92) O.04 O.05 O.050 (0.67-1.01) O.48 O.70 (0.46-1.06) O.48 O.70 (0.46-1.06) O.72 (0.55-0.96) O.

 Table 3. Association of tSNPs with glioma, astrocytoma and glioblastoma risk based on logistic tests (adjusted for sex + age)

OR, odds ratio, 95% CI, 95% confidence interval. *P < 0.05, statistical significance.

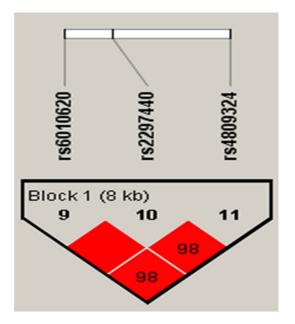


Figure 1. Haplotype-Block Map for *RTEL1* based on SNPs rs6010620, rs2297440 and rs4809324 which were included in Block 1.

decreased glioblastoma risk by both allel model analyses (OR, 0.70; 95% Cl, 0.52-0.92; P =0.001) and log-additive model analyses (OR, 0.72; 95% Cl, 0.55-0.96; P = 0.026). We also found rs9288516 decreased glioma risk by allele model analyses (OR, 0.85; 95% Cl, 0.73-0.99; P = 0.042). Otherwise, individual with the genetype "G/T-G/G" of rs4140805 only increaced glioblastoma risk in an allele model (OR, 1.38; 95% Cl, 1.00-1.89; P = 0.047) and in a dominant model (OR, 1.57; 95% Cl, 1.05-2.35; P = 0.027). Meanwhile, we discovered the genetype "C/T-C/C" of rs2297440 as the risk for glioma in the allele model (OR, 1.30; 95% Cl, 1.10-1.54; P = 0.002), in the dominant model (OR, 1.25; 95% Cl, 1.01-1.56; P = 0.042) and in the log-additive model (OR, 1.31; 95% Cl, 1.10-1.55; P = 0.0018). We also discovered rs2297440 as the risk for astrocytoma in the allele model (OR, 1.26; 95% Cl, 1.02-1.54; P = 0.029) and in the log-additive model (OR, 1.28; 95% Cl, 1.04-1.59; P = 0.022), analysis respectively.

Only one block was detected in *RTEL1* gene by haplotype analysis (**Figure 1**). Global result for the block was: total case = 338, total control = 635, global haplotype association pvalue: 0.038. The results of the association between the *RTEL1* gene haplotype "GCT" and the risk of astrocytoma (OR, 1.45; 95% Cl, 1.12-1.86; Pearson's P = 0.0046) are presented in **Table 4**.

Disscusion

In three case-control studies, we observed rs9288516 in the *XRCC5* gene was associated with a decreased risk of glioma and glioblastoma, rs4140805 in the *RPA3* gene was only associated with an increased risk of glioblastoma and rs2297440 in the *RTEL1* gene was associated with an increased risk of glioma and astrocytoma. In addition, we showed that haplotype "GCT" was associated with the risk of astrocytoma at a 5% level by haplotype association analysis.

The *XRCC5* gene, which is located in 2q35, a risk SNP has been found for hepatocellular carcinoma [20]. XRCC5 gene mutation of a single patient can not be discovered, because XRCC5 gene is a vital evolved gene for human life, indi-

Table 4. RTEL1 haplotype frequencies and the association with astrocytoma risk (n = 973, adjusted for sex + age)

Haplotype	Freq (case + control)	Chi2	OR (95% CI)	P value
GCT	0.1691	8.122	1.45 (1.12-1.86)	0.005
GCC	0.1125	0.074	1.05 (0.77-1.44)	0.76

Loci chosen for hap-analysis: rs6010620, rs2297440 and

rs4809324 in <code>RTEL1</code> gene. OR, odd ratio; Cl, confidence interval.

cating that genetic variations in non-coding regions may be the underlying basis of differing levels of gene transcription and translation [21]. It was also reported in the chronic obstructive pulmonary disease, breast cancer and digestive system cancer as a risk factor [22-24]. In our study, we found genotype "A/T" of rs9288516 as a protective factor was just associated with a decreased risk of glioma and glioblastoma in Chinese patients. Comparing with previous studies, we found that XRCC5 gene is not a risk factor, can be used as a protective factor in glioma and glioblastoma. Our finding suggested that this gene may play a different role in complex diseases. In further studies, we should realize XRCC5 gene different disease mechanisms in glioma, astrocytoma and glioblastoma.

The RPA3 gene, which is located in 7p21.3, a risk SNP (genetype "G/T-G/G" of rs4140805) has been found in our study. Meanwhile, we only found rs4140805 of RPA3 gene with an increased risk of developing glioblastoma. However, glioma and astrocytoma have no significance. It is a single stranded DNA-binding protein that functions in many aspects of DNA metabolism and has a central role in DNA replication, playing an essential function in both initiation and elongation [25]. It was reported that, as a protective risk, RPA3 gene showed decreased risk of glioma [26]. However, we have not yet found about relationship between RPA3 gene and risk of glioblastoma and astrocytoma in previous studies. According to World Health Organization classification astrocytoma grades IV (glioblastoma) [2], whether rs4140805 of the RPA3 gene only for high malignant degree of glioblastoma is in danger. Combinating with previous reseaches, we discovered uniform SNP of gene in glioma and glioblastoma showed the opposite effect. Therefore, it is necessary to study biological functions of the RPA3 gene in further reseach.

The RTEL1 gene locates in 20q13.3, including 40 exons. It was reported that RTEL1 kept genomic stability in suppressing homologous recombination [27]. A recent review point out that RTEL1 was an significant helicase for telomere maintenance and the regulation of homologous recombination [28]. In this study, we found rs2297440 of RTEL1 gene with an increased risk of developing glioma and astrocytoma. The result is consistent with the previous research [11, 29]. However, we found no significant between RTEL1 gene and glioblastoma, this may be related to our sample is less. Previous study suggested polymorphism in the RTEL1 gene was associated with glioblastoma survival [30]. Therefore, relationship between RTEL1 gene and glioblastoma is of great interest and warrant further investigation.

This study was the first associated study between polymorphisms of XRCC5, RPA3 and RTEL1 genes and glioma, astrocytoma and glioblastoma risk respectively in a Chinese population. However, some limitations must be mentioned in our study. Firstly, we collected all the samples from the same hospital for avoiding two or more definite selection bias, and they maintained adiaphorous, did not exist difference in genotype frequencies. Together, all the samples were selected from Han Chinese population who lived in Xi'an city or aroud area. Substantial population of confounding factors, which may cause type-I error for association study. Secondly, the sample size (703 cases and 635 controls) is not large enough in our study for association studies, especially, included 338 astrocytoma cases and 122 glioblastoma cases, large sample size will be convincing. Thirdly, it was ensure that analyzing data was convince, and we select SNPs with MAF > 5% in HapMap Asian population. However, this method will ignore some significant SNPs in other studies.

In conclusion, our study may provides new evidence that *XRCC5, RPA3* and *RTEL1* genes mutations play different roles in the glioma, astrocytoma and glioblastoma in Chinese Han population.

Acknowledgements

This work is supported by China Postdoctoral Science Foundation funded projects (No. 2012-

M521798 and No. 2013T60886). We are also grateful to thank the clinicians and other hospital staff who contributed to the blood sample and data collection for this study.

Disclosure of conflict of interest

None.

Address correspondence to: Dr. Yongkuan Gong, Key Laboratory of Synthetic and Natural Functional Molecule Chemistry of Ministry of Education, College of Chemistry and Materials Science. Northwest University, Xi'an 710069, China. Tel: +86 29 88302109; Fax: +86 29 88302604; E-mail: yongkuangong@ 126.com

References

- Schwartzbaum JA, Fisher JL, Aldape KD, Wrensch M. Epidemiology and molecular pathology of glioma. Nat Clin Pract Neurol 2006; 2: 494-503; quiz 1 p following 16.
- [2] Radner H, Blumcke I, Reifenberger G, Wiestler OD. [The new WHO classification of tumors of the nervous system 2000. Pathology and genetics]. Der Pathologe 2002; 23: 260-83.
- [3] Wen PY, Kesari S. Malignant gliomas in adults. N Engl J Med 2008; 359: 492-507.
- [4] Okada H, Scheurer ME, Sarkar SN, Bondy ML. Integration of epidemiology, immunobiology, and translational research for brain tumors. Ann N Y Acad Sci 2013; 1284: 17-23.
- [5] Bondy ML, Scheurer ME, Malmer B, Barnholtz-Sloan JS, Davis FG, Il'yasova D, Kruchko C, McCarthy BJ, Rajaraman P, Schwartzbaum JA, Sadetzki S, Schlehofer B, Tihan T, Wiemels JL, Wrensch M, Buffler PA; Brain Tumor Epidemiology Consortium. Brain tumor epidemiology: consensus from the Brain Tumor Epidemiology Consortium. Cancer 2008; 113: 1953-68.
- [6] Ostrom QT, Barnholtz-Sloan JS. Current state of our knowledge on brain tumor epidemiology. Curr Neurol Neurosci Rep 2011; 11: 329-35.
- [7] Liu Y, Shete S, Hosking F, Robertson L, Houlston R, Bondy M. Genetic advances in glioma: susceptibility genes and networks. Curr Opin Genet Develop 2010; 20: 239-44.
- [8] Wang X, Zhang H, Wang D, Li X. Association of Genetic Polymorphisms of EGFR with Glioma in a Chinese Population. Genet Test Mol Biomarkers 2015; 19: 59-62.
- [9] Li S, Jin T, Zhang J, Lou H, Yang B, Li Y, Chen C, Zhang Y. Polymorphisms of TREH, IL4R and CCDC26 genes associated with risk of glioma. Cancer Epidemiol 2012; 36: 283-7.
- [10] Li G, Jin TB, Wei XB, He SM, Liang HJ, Yang HX, Cui Y, Chen C, Cai LB, Gao GD. Selected poly-

morphisms of GSTP1 and TERT were associated with glioma risk in Han Chinese. Cancer Epidemiol 2012; 36: 525-7.

- [11] Jin TB, Zhang JY, Li G, Du SL, Geng TT, Gao J, Liu QP, Gao GD, Kang LL, Chen C, Li SQ. RTEL1 and TERT polymorphisms are associated with astrocytoma risk in the Chinese Han population. Tumour Biol 2013; 34: 3659-66.
- [12] Jin TB, Li XL, Yang H, Jiri M, Shi XG, Yuan DY, Kang LL, Li SQ. Association of polymorphisms in FLT3, EGFR, ALOX5, and NEIL3 with glioblastoma in the Han Chinese population. Med Oncol 2013; 30: 718.
- [13] Trembizki E, Smith H, Lahra MM, Chen M, Donovan B, Fairley CK, Guy R, Kaldor J, Regan D, Ward J, Nissen MD, Sloots TP, Whiley DM. High-throughput informative single nucleotide polymorphism-based typing of Neisseria gonorrhoeae using the Sequenom MassARRAY iPLEX platform. J Antimicrob Chemother 2014; 69: 1526-32.
- [14] Gabriel S, Ziaugra L, Tabbaa D. SNP genotyping using the Sequenom MassARRAY iPLEX platform. Current protocols in human genetics/editorial board, Jonathan L Haines [et al] 2009; 2: 2-12.
- [15] Thomas RK, Baker AC, Debiasi RM, Winckler W, Laframboise T, Lin WM, Wang M, Feng W, Zander T, MacConaill L, Lee JC, Nicoletti R, Hatton C, Goyette M, Girard L, Majmudar K, Ziaugra L, Wong KK, Gabriel S, Beroukhim R, Peyton M, Barretina J, Dutt A, Emery C, Greulich H, Shah K, Sasaki H, Gazdar A, Minna J, Armstrong SA, Mellinghoff IK, Hodi FS, Dranoff G. Mischel PS, Cloughesy TF, Nelson SF, Liau LM, Mertz K, Rubin MA, Moch H, Loda M, Catalona W. Fletcher J. Signoretti S. Kaye F. Anderson KC, Demetri GD, Dummer R, Wagner S, Herlyn M, Sellers WR, Meyerson M, Garraway LA. High-throughput oncogene mutation profiling in human cancer. Nat Genet 2007; 39: 347-51.
- [16] Kochl S, Niederstatter H, Parson W. DNA extraction and quantitation of forensic samples using the phenol-chloroform method and realtime PCR. Methods Mol Biol 2005; 297: 13-30.
- [17] Adamec C. [Example of the Use of the Nonparametric Test. Test X2 for Comparison of 2 Independent Examples]. Cesk Zdrav 1964; 12: 613-9.
- [18] Bland JM, Altman DG. Statistics notes. The odds ratio. BMJ 2000; 320: 1468.
- [19] Shi YY, He L. SHEsis, a powerful software platform for analyses of linkage disequilibrium, haplotype construction, and genetic association at polymorphism loci. Cell Res 2005; 15: 97-8.
- [20] Li R, Yang Y, An Y, Zhou Y, Liu Y, Yu Q, Lu D, Wang H, Jin L, Zhou W, Qian J, Shugart YY. Genetic polymorphisms in DNA double-strand

break repair genes XRCC5, XRCC6 and susceptibility to hepatocellular carcinoma. Carcinogenesis 2011; 32: 530-6.

- [21] Wang Y, Ghosh G, Hendrickson EA. Ku86 represses lethal telomere deletion events in human somatic cells. Proc Natl Acad Sci U S A 2009; 106: 12430-5.
- [22] Wang B, Yang J, Xiao J, Liang B, Zhou HX, Su Z, Xu S, Chen H, Ma C, Deng J, Li D, Zhou H, Ou X, Feng Y. Association of XRCC5 polymorphisms with COPD and COPD-related phenotypes in the Han Chinese population: a case-control cohort study. Genet Mol Res 2014; 13: 7070-8.
- [23] Yang MD, Tsai CW, Chang WS, Tsou YA, Wu CN, Bau DT. Predictive role of XRCC5/XRCC6 genotypes in digestive system cancers. World J Gastrointest Oncol 2011; 3: 175-81.
- [24] Rajaei M, Saadat I, Omidvari S, Saadat M. Association between polymorphisms at promoters of XRCC5 and XRCC6 genes and risk of breast cancer. Med Oncol 2014; 31: 885.
- [25] Fanning E, Klimovich V, Nager AR. A dynamic model for replication protein A (RPA) function in DNA processing pathways. Nucleic Acids Res 2006; 34: 4126-37.
- [26] Jin T, Zhang J, Li G, Li S, Yang B, Chen C, Cai L. TP53 and RPA3 gene variations were associated with risk of glioma in a Chinese Han population. Cancer Biother Radiopharm 2013; 28: 248-53.

- [27] Uringa EJ, Lisaingo K, Pickett HA, Brind'Amour J, Rohde JH, Zelensky A, Essers J, Lansdorp PM. RTEL1 contributes to DNA replication and repair and telomere maintenance. Mol Biol Cell 2012; 23: 2782-92.
- [28] Uringa EJ, Youds JL, Lisaingo K, Lansdorp PM, Boulton SJ. RTEL1: an essential helicase for telomere maintenance and the regulation of homologous recombination. Nucleic Acids Res 2011; 39: 1647-55.
- [29] Li G, Jin T, Liang H, Zhang Z, He S, Tu Y, Yang H, Geng T, Cui G, Chen C, Gao G. RTEL1 tagging SNPs and haplotypes were associated with glioma development. Diagn Pathol 2013; 8: 83.
- [30] Liu Y, Shete S, Etzel CJ, Scheurer M, Alexiou G, Armstrong G, Tsavachidis S, Liang FW, Gilbert M, Aldape K, Armstrong T, Houlston R, Hosking F, Robertson L, Xiao Y, Wiencke J, Wrensch M, Andersson U, Melin BS, Bondy M. Polymorphisms of LIG4, BTBD2, HMGA2, and RTEL1 genes involved in the double-strand break repair pathway predict glioblastoma survival. J Clin Oncol 2010; 28: 2467-74.