# Original Article Oxymatrine enhanced anti-tumor effects of Bevacizumab against triple-negative breast cancer via abating Wnt/β-Catenin signaling pathway

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Abstract: Bevacizumab, a monoclonal antibody targeting vascular endothelial growth factor A (VEGF-A), was used in combination with traditional chemotherapy as the first line treatment for metastatic colorectal cancer (mCRC), nonsmall cell lung cancer (NSCLC) and advanced ovarian cancer. However, it shows limited efficacy for human triplenegative breast cancer (TNBC). Bevacizumab shows potent anti-angiogenesis activity, meanwhile, it also increases invasive and metastatic properties of TNBC cells by activiting Wnt/β-Catenin pathway. To overcome this problem, and fully utilize its potency against cancer, further synergistic strategy is recommended to be developed, especially the concurrent use with those Wnt-targeting agents. Here, by screening a small library of traditional Chinese medicine, we identified a Chinese herb derived Oxymatrine, which could target Wnt/B-Catenin signaling and compromise the oncogenic effects of Bevacizumab. Bevacizumab was validated to induce epithelial-mesenchymal cell transformation (EMT) and cancer stem-like properties of TNBC cells in hypoxia/nutritional stress environment. On the contrary, Oxymatrine reversed the EMT phenotype and depleted the subpopulation of TNBC stem cells induced by Bevacizumab. Oxymatrine enhanced the anti-tumor effects of Bevacizumab in vivo, and holded the potential of reducing the risk of relapse and metastasis by impairing the self-renewal ability of TNBC stem cells. The underlying mechanism was elucidated: Bevacizumab stimulated Wnt/B-Catenin signaling pathway, and Oxymatrine could compromise this effect. On this foundation, factoring into the satisfactory anti-angiogenic activity and low toxicity, Oxymatrine is a good candidate for the synergistic therapy together with Bevacizumab for the treatment of TNBC.

Keywords: Oxymatrine, bevacizumab, TNBC, EMT, Wnt/β-Catenin pathway

#### Introduction

TNBC, characterized by the lack of expression of estrogen receptor (ER), progesterone receptor (PR), and human epidermal growth factor receptor 2 (HER2), is an aggressive subtype of breast cancer. Breast cancer incidence rate ranks the first in overall cancer in women, among them, TNBC accounts for around 10-20% [1, 2]. Currently TNBC still represents the most therapeutically intractable subtype as the negative expressions of ER, PR, and HER2, and cytotoxic chemotherapy remains the standard of care for patients with metastatic TNBC. However, TNBC is clinically characterized as more aggressive and less responsive to standard treatment with a poorer overall patient prognosis [3-5].

Bevacizumab is a recombinant humanized monoclonal antibody that blocks angiogenesis by inhibiting VEGF-A approved by FDA in 2004. To date, FDA has approved its application for the treatment of mCRC, NSCLC and advanced ovarian cancer combined with the chemotherapy drugs. Meanwhile, it has also been tested by combination with the cytotoxic chemotherapy in patients with TNBC (clinical trial number NCT-00528567). The result from the clinical trial showed that this combination regime increases objective response rate (ORR) and progression-free survival (PFS) vs. chemotherapy alone, however the benefit of overall survival (OS) is not enough to support its further application for this indication [6-8].

Bevacizumab, as well as some other anti-angiogenic agents, has been recently shown to be able to increase the invasive and metastatic capacity and induce EMT in breast cancer cells [9]. Bevacizumab activates Wnt/β-Catenin signaling pathway excessively, further facilitates the growth, migration and invasion of TNBC, which normally associates with the unfavorable outcome of patients [10-13]. In addition, the important angiogenesis inducing factor VEGF, a target gene of  $\beta$ -Catenin, is up-regulated by the activated Wnt/B-Catenin signaling, possibly leading to the incomplete blocking of tumor angiogenesis [14, 15]. So it seems that the therapeutic effects of Bevacizumab against TNBC may be partially counteracted by the induced activation of Wnt/β-Catenin signaling, indicating that, to improve TNBC patient outcome, anti-angiogenic agents need to be combined with Wnt/β-Catenin signaling-targeting drugs.

TCM has a long history with various pharmacological activities and low toxicity. Thereby, those Chinese herbs hold great potential for the therapy development against cancer, and several agents like 20(S)-ginsenoside Rg3, Lupeol and Sophoridine has entered into market [16, 17]. We have carried out some work about the development of Chinese herb-derived triptolide [18, 19], and established a small Chinese herb library focusing on cancer therapy. Oxymatrine  $(C_{15}H_{24}N_{2}O_{2})$ , one compound in our library, is an alkaloid compound extracted from the roots of Sophora flavescens Ait. It attracts our interest as it also shows multiple biological activities like triptlide, but it is well tolerable. Oxymatrine is clinically demonstrated to display anti-inflammatory, anti-allergic, anti-virus, antifibrotic, and cardiovascular protective effects [20, 21]. Recently, extensive experimental studies on Oxymatrine have demonstrated its potential applications to the treatment of various cancer [22-24], and showed that Oxymatrine can effectively repress Wnt/β-Catenin pathway in some tumors [25, 26]. The potential anti-tumor activities of Oxymatrine against TNBC were verified in our preliminary study (Supplementary Figure <u>1</u>).

Herein, the double-edged role of Bevacizumab for TNBC therapy deserves attention, and may be overcame by synergistic therapy. Chinese herbs are promising candidates for such design. We firstly verified the EMT and CSC phenotype caused by Bevacizumab with some routine assays like wound healing, transwell invasion, and ALDEFLUOR test. Then Oxymatrine was selected from our established Chinese herb library for further evaluation as it demonstrated good synergistic effects with Bevacizumab. In the end, the *in vivo* effects were also evaluated, and Wnt/ $\beta$ -Catenin signaling pathway were well dissected as the possible mechanism for such synergistic effects.

#### Materials and methods

#### Cell culture and reagents

Oxymatrine (purity  $\geq$  98%, Macklin Biochemical Co., Ltd., Shanghai, China) and Bevacizumab (Roche, Paris, France) was diluted in PBS. Docetaxel (Hengrui Medicine, Jiangsu, China) was suspended in the injection solvent and then diluted in PBS. FH535 ( $\geq$  98%, absin, China) was suspended in DMSO. The human embryonic kidney cell line HEK293 (Cell bank of the Chinese Academy of Sciences, Shanghai, China) was cultured in DMEM high glucose medium (Gibco, New York, USA), supplemented with 10% (v/v) FBS (Gibco, Auckland, NZ). The human breast cancer cell lines MDA-MB-231/ MDA-MB-468 (American Type Culture Collection, New York, USA) were maintained in L-15 medium (Gibco, New York, USA) supplemented with 10% (v/v) FBS. HUVECs (ScienCell, San Diego, CA) were maintained in endothelial culture medium (ECM, ScienCell) supplemented with 5% (v/v) FBS and 1% (v/v) endothelial cell growth supplement (ECGS, ScienCell). All cells were maintained at 37°C in a humidified atmosphere with 5% CO<sub>2</sub>.

#### Transwell invasion assay

 $1 \times 10^4$  of MDA-MB-231/MDA-MB-468 cells were suspended in serum-free medium with the addition of Oxymatrine (4 mM)/Bevacizumab (200 nM)/Oxymatrine (4 mM) + Bevacizumab (200 nM) and then plated into the upper wells of 24-well transwell chambers (Millipore, Billerica, USA) coated with 20 µL Matrigel (Corning, Bedford, USA). The lower chambers were filled with L-15 medium containing 10% (v/v) FBS. Similarly,  $1 \times 10^4$  HUVECs and different concentrations of Oxymatrine (0, 1, 2 and 4 mM) were suspended in serum-free medium and added to the top 24-well transwell chambers. The bottom chambers were filled with 600 µL ECM containing 5% (v/v) FBS and 1% (v/v) ECGS. The next experimental procedure was performed as previously described [27].

#### Wound healing assay

 $3 \times 10^4$  MDA-MB-231/MDA-MB-468 cells were placed into the each well of Culture-Insert (lbidi, Martinsried, Germany) in 24-well plate. After adherence, the Culture-Insert was removed and cells were rinsed with PBS. Oxymatrine/ Bevacizumab/Oxymatrine + Bevacizumab dissolved in serum-free medium was added into the wells. After 24 h-incubation, the drug-dissolved medium was replaced by the serum-free medium. Images were taken with OLYM-PUS inversion fluorescence microscope at 0, 24, 48 h after the addition of the treatments. The wound migrated distances were measured with Image-Pro-Plus program and calculated as follows: Ln = (L<sub>0</sub>-L<sub>time</sub>)/2.

#### ALDEFLUOR assay

ALDEFLUOR™ kit (STEMCELL Technologies, Canada) was used to analyze the level of the enzyme aldehyde dehydrogenase (ALDH) in cells. 5 × 10<sup>5</sup> MDA-MB-231/MDA-MB-468 cells were seeded in 6-well plates. When the confluence reached 70%-80%, cells were incubated with Oxymatrine/Bevacizumab/Oxymatrine + Bevacizumab for 24 h. Whereafter, cells were trypsinized, resuspended in ALDEFLUOR<sup>™</sup> assay buffer and incubated with the activated ALDEFLUOR™ fluorescent reagent for 30 min at 37°C. For ALDH1 inhibitor control, diethylaminobenzaldehyde (DEAB) was added along with the fluorescent reagent. Cells were then stained with 7-AAD (BD Biosciences, San Diego, USA), green fluorescence positive cells in live cells (7-AAD) were analyzed using a BD FA-CSMelody flow cytometer by comparing the fluorescence intensity of DEAB-treated samples.

#### IF analysis

 $5 \times 10^5$  MDA-MB-231/MDA-MB-468 cells were seeded on cover slips in 6-well plates and allowed to adhere to the glass overnight to reach 70% confluence. Then, the cells were treated with Oxymatrine/Bevacizumab/Oxymatrine + Bevacizumab/FH535 for 24 h. Subsquently, the subcellar location of  $\beta$ -Catenin was detected as previously described [28]. Finally, images were taken by an Leica SP2 Confocal System at 400-times magnification.

#### Immunoblotting assay

5 × 10<sup>5</sup> MDA-MB-231/MDA-MB-468 cells were seeded into the wells of 6-well plates, allowed to adhere overnight to reach 70%-80% confluence, treated with Oxymatrine/Bevacizumab/ Oxymatrine + Bevacizumab/FH535 (20 µM, a small molecule inhibitor of Wnt/β-Catenin pathway) for 24 h, then the whole cell protein was extracted from cells using RIPA buffer (Beyotime, Shanghai, China) and the nuclear extracts were prepared with a NE-PER Nuclear and Cytoplasmic Extraction Kit (Pierce Biotechnology, Rockford, USA). The next experimental procedure was performed as previously described [28]. The primary antibodies used included anti-β-actin, anti-Histone H3, anti-β-Catenin, anti-c-Myc, anti-Snail, anti-Slug, anti-Vimentin, anti-TCF8/ZEB1, anti-CD44, anti-E-cadherin, anti-N-cadherin (CST, MA, USA), anti-Cyclin D1, anti-VEGFA (Abcam, Cambridge, UK).

#### Cell xenografts in nude mice assay

5 to 6-week-old female BALB/c-nude mice were purchased from Shanghai Lab. Animal Research Center, China, maintained in a pathogen-free environment (23 ± 2°C, 55 ± 5% humidity) on a 12 h light/12 h dark cycle with food and water supplied adlibitum throughout the experimental period. All animal experiments were performed in accordance with the National Institutes of Health guide for the care and use of Laboratory animals and protocols approved by the Animal Ethics Committee of Shanghai University of Medicine and Health Sciences. Each mouse was subcutaneously inoculated with injections of 1 × 107 MDA-MB-231/MDA-MB-468 cells into its left axilla. When the tumors reached 50 mm<sup>3</sup>, mice were randomized into 8 groups (n = 5 for each treatment group), and the administration began: (1) PBS control; (2) 5 mg/kg Oxymatrine (intraperitoneal injection, once every other day); (3) 25 mg/kg Oxymatrine; (4) 25 mg/kg FH535 (intraperitoneal injection, once every other day); (5) 5 mg/kg Bevacizumab (intravenous injection, every 3 days); (6) 5 mg/kg Bevacizumab + 25 mg/kg Oxymatrine; (7) 5 mg/kg Bevacizumab + 25 mg/kg FH535; (8) 5 mg/kg Bevacizumab + 10 mg/kg Docetaxel (intravenous injection, every 3 days). Tumor development was measured periodically and the tumor volume was determined using the formula V = (length × width<sup>2</sup>)/2. The administration end at day 31 and the tumors of each group were isolated for further study.

### IHC and IF analysis

Paraffin sections were cut into 5 µm sections and fixed in 4% paraformaldehyde. For IHC staining, the sections were incubated with primary antibodies anti-Ki-67 (CST, MA, USA), anti-VEGFA antibody (Abcam, Cambridge, UK), and horseradish peroxidase (HRP)-labeled secondary antibody and analyzed with the Vectastain ABC Kit (Dako, Copenhagen, Denmark). A subset of MDA-MB-231/MDA-MB-468 tumor tissues were subjected to Western blot to examine the VEGFA protein levels and show changes in VEGFA expression. For IF staining, the sections were incubated with primary antibodies anti-β-Catenin, anti-CD31 (CST, MA, USA) and Alexafluor-488/594 labeled antibody (Molecular Probes, Eugene, USA). The slides were sealed with Prolong Gold Antifade (Invitrogen, Carlsbad, USA), which contained DAPI. Subsquently, images were taken by an Leica SP2 Confocal System at 400-times magnification.

#### Sphere formation assay

MDA-MB-231/MDA-MB-468 tumors tissues were minced into small pieces and processed with collagenase for 2 h at 37°C. After centrifugation, the cell pellets were trypsinized and passed through an 80-µm filter to produce single-cell suspension. Then  $5 \times 10^3$  dissociated tumor cells were seeded on ultralow attachment 6-well plates in serum-free medium DM-EM/F12 (Gibco, Grand Island, USA) supplemented with B27 (Gibco, Grand Island, USA), 20 ng/mL human recombinant basic fibroblast growth factor (FGF)-basic, and 20 ng/mL epidermal growth factor (EGF, Sino Biological Inc., Beijing, China). The medium was changed every other day until the tumor sphere formation was observed in 2 weeks, and the mammospheres with diameter > 60  $\mu$ m were counted under an OLYMPUS inversion fluorescence microscope [29].

#### Cell proliferation assay

HUVECs/HEK293 cells (negative control) were seeded at a density of 2000 cells/well in 96-well plates. After cultured overnight, fresh medium containing different concentrations of Oxymatrine were replenished for 48 h. Subsequently, cell viability was quantified by MTT assay and the proliferation rate was expressed as percentages of the vehicle control (Oxymatrine untreated group).

### Tube formation assay

 $2 \times 10^4$  HUVECs were seeded into the 96-well plate coated with 50 µL Matrigel and added with 100 µL ECM supplemented with 2% (v/v) FBS, 1% (v/v) ECGS and various concentrations of Oxymatrine (0, 1, 4 and 16 mM). The next experimental procedure was performed as previously described [27].

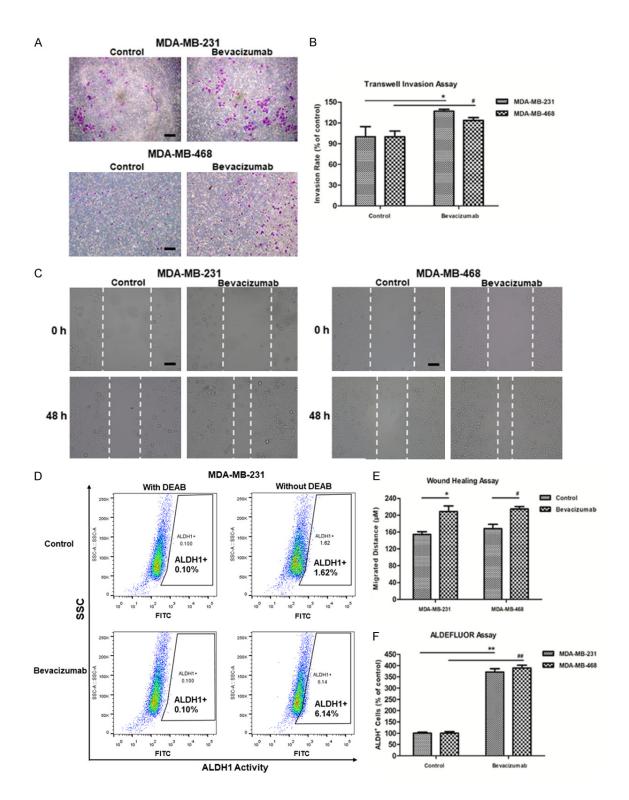
### Statistical analysis

All data were presented as the mean  $\pm$  standard deviation (SD). Differences between multiple groups were analyzed by the student's *t* test and *p* values of 0.05 or less were considered statistically significant. All figures were produced with the GraphPad Prism 5 software program.

#### Results

Bevacizumab induced transwell, migration of TNBC cells and increased subpopulation of putative TNBC stem-like cells under serum starvation condition

Wound healing and transwell invasion assays were performed for the evaluation of cell migration. Following Bevacizumab treatments under serum starvation, increased cell invasion (Figure 1A, 1B) and migration (Figure 1C, 1E) were detected. As increased invasion and migration are outcomes of the EMT process [30, 31], which indicated Bevacizumab could induce EMT of TNBC cells in serum-deprived environment. Moreover, researchers have shown that EMT could promote stem cell properties and further generate cells with the features of CSC [32, 33], conversely, CSC could accelerate tumor invasion and metastasis. In consideration of the interplay of EMT and CSC, we uti-



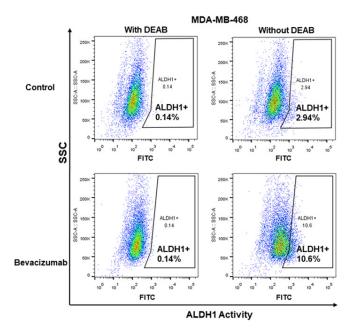


Figure 1. Bevacizumab promoted the invasion, migration of TNBC cells, and increased percent of ALDH1<sup>+</sup> TNBC cells. (A) Microscopic views from transwell assay to estimate MDA-MB-231/MDA-MB-468 cells invasion following 24 h treatment of Bevacizumab (200 nM), bar = 100  $\mu$ m. (B) Quantitative analysis of (A) by Image J. (C)Photomicrographs of cell migration from wound healing assay in MDA-MB-231/MDA-MB-468 cells treated with Bevacizumab, bar =  $100 \mu m. (D)$ MDA-MB-231/MDA-MB-468 cells were treated with Bevacizumab for 24 h, then ALDH1<sup>+</sup> cells were determined by Alderfluor assay and viable cells (7-AAD negative) were used for analysis. DEAB-treated cells were used as negative control. (E) Quantitative analysis of (C) by Image J. (F) Quantitative analysis of (D). Data were presented as the mean  $\pm$  SD, n = 3,  ${}^*P < 0.05$ ,  ${}^{**}P < 0.01$  (MDA-MB-231 cells); \*P < 0.05, \*\*P < 0.01 (MDA-MB-468 cells).

lized ALDEFLUOR assay to detect the subpopulation of TNBC stem-like cells [34]. Previously, intracellular ALDH activity was identified to be a functional marker for CSCs derived from different cancer types [35-37]. As shown in **Figure 1D**, **1F**, in comparison with the control cells, the Bevacizumab-treated TNBC cells possessed higher ALDH1<sup>+</sup> population notably (MDA-MB-231: \*\*P < 0.01, MDA-MB-468: ##P < 0.01), which indicated that Bevacizumab increased the subpopulation of TNBC stem-like cells remarkably under serum starvation condition.

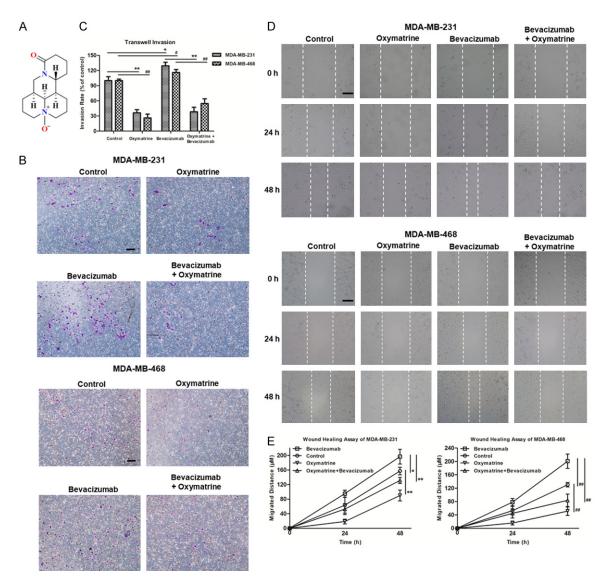
Oxymatrine suppressed Bevacizumab-induced transwell, migration of TNBC cells and depleted subpopulation of putative TNBC stem-like cells induced by Bevacizumab

Figure 2 demonstrated the potential of Oxymatrine to reduce the invasiveness and metastasis of TNBC cells (Oxymatrine group vs. control group). Further, the results showed that Oxymatrine indeed attenuated Bevacizumabinduced invasion and migration of MDA-MB-231 cells (\*\*P < 0.01) and MDA-MB-468 cells (##P < 0.01), which indicated Oxymatrine strongly suppressed Bevacizumab-induced EMT of TNBC cells. We further determined the effects of Oxymatrine on the proportion of ALDH1<sup>+</sup> cells in TNBC. Figure 3A, 3B revealed that, Oxymatrine reduced ALDH1<sup>+</sup> population cells of TNBC significantly (Oxymatrine group vs. control group, MDA-MB-231: \*\*P < 0.01, MDA-MB-468:

<sup>##</sup>*P* < 0.01), and Oxymatrine reversed the aberrantly increased proportion of ALDH1<sup>+</sup> cells induced by Bevacizumab after treatment together with Bevacizumab (Bevacizumab + Oxymatrine group vs. Bevacizumab group, MDA-MB-231: <sup>\*\*</sup>*P* < 0.01, MDA-MB-468: <sup>##</sup>*P* < 0.01). The specificity of ALDEFLUOR assay was determined by treating the cells with DEAB, a specific inhibitor of ALDH. Above all, Oxymatrine not only reversed Bevacizumab induced EMT of TNBC cells, but also suppressed Bevacizumab-increased TNBC stem-like cells population.

Oxymatrine inhibited EMT and Wnt/β-Catenin pathway activated by Bevacizumab in TNBC cells

Firstly, the expression level of various classical EMT markers in TNBC cells were detected by Western blot. Following treatment of Bevacizumab in serum-deprived environment, the expression level of N-cadherin and Vimentin were increased; and the expression of epithelialassociated gene E-cadherin decreased compared with control. The expression of EMT related transcription factors including ZEB, Snail, and Slug were also increased (Figure 4A, 4D). Meanwhile, Oxymatrine and FH535 exhibited an exactly opposite effect on EMT of TNBC ce-Ils compared to Bevacizumab (Figure 4A, 4D). Further, the analysis of the data from Bevacizumab + Oxymatrine group and Bevacizumab group indicated that, Oxymatrine reversed the

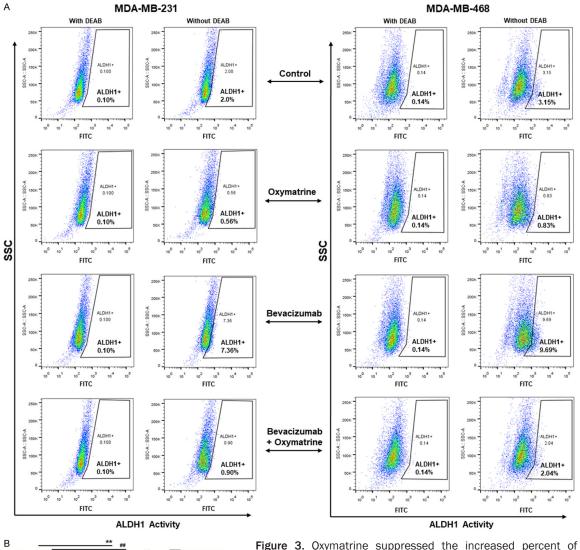


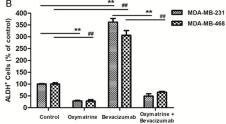
**Figure 2.** Oxymatrine inhibited Bevacizumab-induced transwell and migration of TNBC cells. (A) Chemical structure of Oxymatrine. (B) Microscopic views from transwell assay to estimate MDA-MB-231/MDA-MB-468 cells invasion following 24 h treatment of Oxymatrine (4 mM)/Bevacizumab (200 nM)/Oxymatrine (4 mM) + Bevacizumab (200 nM), bar = 100  $\mu$ m. (C) Quantitative analysis of (B) by Image J. (D) Photomicrographs of cell migration from wound healing assay in MDA-MB-231/MDA-MB-468 cells treated with Oxymatrine/Bevacizumab/Oxymatrine + Bevacizumab, bar = 200  $\mu$ m. (E) Quantitative analysis of (D) by Image J. The quantitative analyses of transwell invasion assay and wound healing assay were based on the mean of the 5 regions of each group. Data were presented as the mean ± SD, n = 5, \*P < 0.05, \*\*P < 0.01 (MDA-MB-231 cells); #P < 0.05, #P < 0.01 (MDA-MB-468 cells).

Bevacizumab-induced increase in the expression of N-cadherin, Vimentin, ZEB, Snail and Slug, and increased E-cadherin expression in TNBC cells. Our findings strongly suggested that Oxymatrine effectively repressed Bevacizumab-induced EMT in TNBC cells.

Since  $Wnt/\beta$ -Catenin signaling played a crucial role in EMT, we further demonstrated that whether Bevacizumab-induced EMT involved

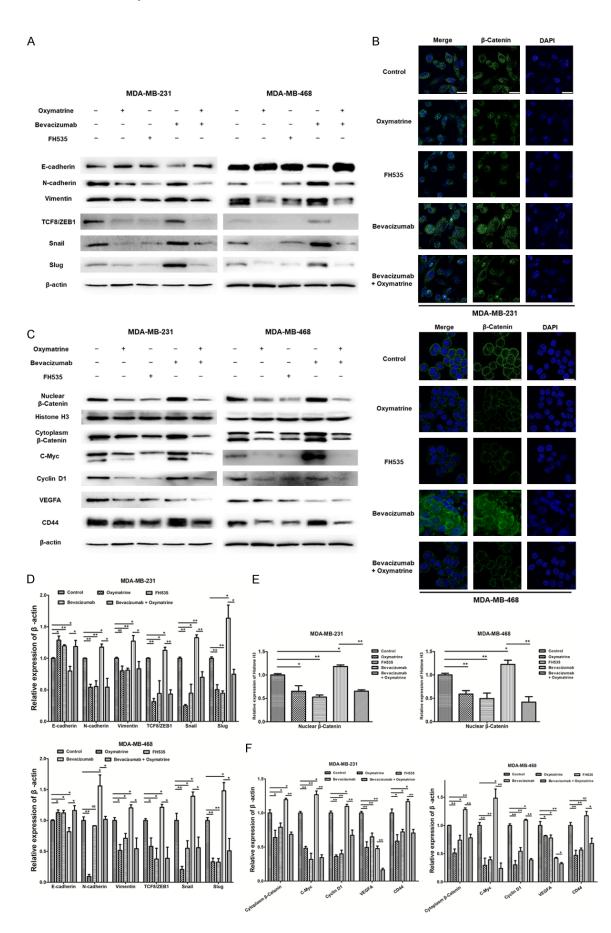
the activation of Wnt/ $\beta$ -Catenin signaling [10, 38]. IF staining of  $\beta$ -Catenin was performed to determine the localization of  $\beta$ -Catenin, and thereby the effect of Bevacizumab on nuclear  $\beta$ -Catenin accumulation was verified [39]. Figure 4B showed that, compared to the control group, Bevacizumab promoted the translocation of  $\beta$ -Catenin from the cytoplasm to the nucleus in TNBC cells. Figure 4C, 4E further confirmed the increased  $\beta$ -Catenin level in nu-





**Figure 3.** Oxymatrine suppressed the increased percent of ALDH1<sup>+</sup> TNBC cells induced by Bevacizumab. (A) MDA-MB-231/MDA-MB-468 cells were treated with Oxymatrine/Bevacizumab/Oxymatrine + Bevacizumab for 24 h, then ALDH1<sup>+</sup> cells were determined by Alderfluor assay and viable cells (7-AAD negative) were used for analysis. DEAB-treated cells were used as negative control. (B) Quantitative analysis of (A). Data were presented as the mean  $\pm$  SD, n = 3, \*\**P* < 0.01 (MDA-MB-231 cells); ##*P* < 0.01 (MDA-MB-468 cells).

cleus (Bevacizumab group vs. control group). Intriguingly, both cytoplasmatic and nuclear  $\beta$ -Catenin expression level of Bevacizumab-treated TNBC cells were improved compared to the control. Bevacizumab also induced the expression of c-Myc, Cyclin D1 and CD44, downstream components of the Wnt/ $\beta$ -Catenin pathway (Figure 4C, 4F). To sum up, the nuclear accumulation of  $\beta$ -Catenin, along with the increased expression level of total  $\beta$ -Catenin protein and downstream components strongly indicated Bevacizumab activated Wnt/ $\beta$ -Catenin pathway remarkably. On the contrary, Oxymatrine exhibited the potency of disrupting Wnt/ $\beta$ -Catenin pathway (vs. control group), which was similar to FH535 (**Figure 4B, 4C**). After making a further comparative study of Bevacizumab + Oxymatrine group and Bevacizumab group, we found  $\beta$ -Catenin staining was attenuated in the nuclei and remained in the cytoplasm, and total  $\beta$ -Catenin protein level declined notably in Bevacizumab + Oxymatrine group (**Figure 4B**,



**Figure 4.** Oxymatrine inhibited EMT and Wnt/β-Catenin signaling induced by Bevacizumab. (A) Oxymatrine inhibited Bevacizumab-induced EMT in TNBC cells. MDA-MB-231/MDA-MB-468 cells were incubated with Oxymatrine/ FH535 (20 µM)/Bevacizumab/Oxymatrine + Bevacizumab for 24 h. Then the results of Western blot analysis for EMT marker proteins were shown. (B) Oxymatrine repressed the accumulation of β-Catenin in the nucleus induced by Bevacizumab. IF stainings of β-Catenin (green) were shown, and nuclei were counterstained with DAPI (blue), bar = 20 µm. (C) Oxymatrine suppressed the activation of Wnt/β-Catenin pathway induced by Bevacizumab. The results of Western blot analysis for the expression of β-Catenin and downstream oncoproteins were shown. Histone H3 was used as loading control for nulear proteins, and β-actin was for cytoplasmic proteins (A, C). (D) Relative expression levels of the proteins indicated in (A). (E) Relative expression levels of nuclear β-Catenin indicated in (C). (F) Relative expression levels of the proteins indicated in (C) except nuclear β-Catenin. Data were presented as the mean ± SD, n = 3, \*P < 0.05, \*\*P < 0.01, NS: no significance.

**4C**). Furthermore, the expression of Wnt/ $\beta$ -Catenin downstream oncoproteins (c-Myc, Cyclin D1, CD44) was repressed in Bevacizumab + Oxymatrine group (vs. Bevacizumab group, Figure 4C, 4F). It was noteworthy that, VEGF, an important target gene of Wnt/β-Catenin pathway and also a direct target of Bevacizumab, was down-regulated by Bevacizumab and Oxymatrine synergistically (Figure 4C, 4F). In the immunoblotting assay, Histone H3 was used as an internal control for nuclear protein and β-actin was used as the control for cytoplasm protein. The comprehensive analysis of IF and Western blot indicated that Oxymatrine markedly suppressed Wnt/β-Catenin pathway activated by Bevacizumab, and thus inhibited EMT in TNBC cells.

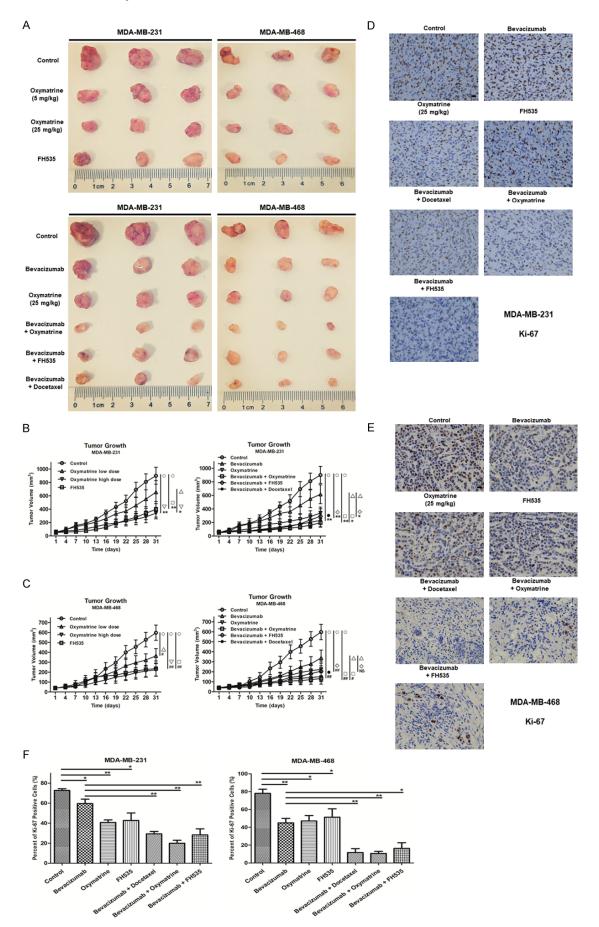
# Oxymatrine enhanced the anti-tumor activity of Bevacizumab against TNBC in vivo

Since Oxymatrine was demonstrated to suppress Wnt/ $\beta$ -Catenin pathway, inhibit EMT and deplete subpopulation of CSC effectively in vitro, TNBC cell xenografts in nude mice assay was executed to investigate the anti-tumor capacity of Oxymatrine in vivo. The tumor photographs (Figure 5A, 3 tumors of each group were shown, the other 2 not shown were treated for further study in the fresh condition) and tumor growth curves (Figure 5B, 5C) of each group showed that, Oxymatrine inhibited the tumor growth in MDA-MB-231/MDA-MB-468 xenografts. Notably, the high dose (25 mg/kg) of Oxymatrine achieved 62.16 ± 5.82% MDA-MB-231 tumor growth inhibition (Figure 5B) and 62.54 ± 4.29% MDA-MB-468 tumor growth inhibition (Figure 5C) compared to the PBS control. After making a further comparative study of the data from Bevacizumab group and Bevacizumab + Oxymatrine group, we found Bevacizumab + Oxymatrine significantly reduced tumor growth when compared either to PBS control (MDA-MB-231: \*\*P < 0.01, MDA- MB-468:  $^{\#P}$  < 0.01) or Bevacizumab alone treatment group (MDA-MB-231: \*P < 0.05, MDA-MB-468: #P < 0.05). Figure 5B, 5C verified that FH535 showed a similar effect with Oxymatrine. What differentiated FH535 from Oxymatrine was that, the anti-tumor synergistic effect of FH535 was weaker than Oxymatrine when combined with Bevacizumab; as showed in Figure 5C, there was no significant difference between the MDA-MB-468 tumor volume of Bevacizumab group and Bevacizumab + FH535 group at the endpoint of administration. Moreover, let it be noted that Bevacizumab + Oxymatrine exhibited slightly better anti-proliferative activity than Bevacizumab + Docetaxel, which was one of the relatively effective clinical therapies of TNBC.

Transplanted tumor tissues were harvested for IHC at the end of the treatment period. IHC demonstrated that there was a significant decrease in the numbers and intensity of cell proliferation marker Ki-67 in Bevacizumab + Oxymatrine group compared to PBS control or Bevacizumab group (**Figure 5D-F**), which was consistent with the results of the inhibition of TNBC tumor growth. In conclusion, Oxymatrine could enhance the anti-tumor effects of Bevacizumab against TNBC effectively.

Oxymatrine abated  $Wnt/\beta$ -Catenin pathway of TNBC activated by Bevacizumab in vivo and weakened the self-renew capacity of TNBC cells enhanced by Bevacizumab

We have verified that Oxymatrine indeed abated Wnt/ $\beta$ -Catenin pathway in TNBC cells under conditions with and without Bevacizumab treatment *in vitro*. To extend these observations to the mouse models, we examined the expression and localization of  $\beta$ -Catenin in MDA-MB-231/MDA-MB-468 tumors by IF. As shown in **Figure 6A**, **6B**,  $\beta$ -Catenin was primarily detected in the cytoplasm of tumor cells from PBS



**Figure 5.** Oxymatrine combined with Bevacizumab significantly inhibited tumor growth in MDA-MB-231/MDA-MB-468 xenografts. A. Representative images of isolated tumors from MDA-MB-231 (the left 3 rows)/MDA-MB-468 (the right 3 rows)-tumor bearing nude mice. B, C. MDA-MB-231/MDA-MB-468 tumor growth curves of each group under different treatments. Data were given as the mean  $\pm$  SD (n = 5), \**P* < 0.05, \*\**P* < 0.01 (MDA-MB-231 tumor); \**P* < 0.05, #\**P* < 0.01, NS: no significance (MDA-MB-468 tumor). D, E. IHC staining of Ki-67 on paraffin sections of MDA-MB-231/MDA-MB-468 xenografted tumor. Ki-67<sup>+</sup> cells were identified with an anti-Ki-67 antibody (brown staining), bar = 20 µm. F. Ki-67 positive cells of tumor tissues were counted using Image-Pro-Plus program, and the percent of Ki-67 positive cells of each group was shown. Data were given as the mean  $\pm$  SD (n = 3), \**P* < 0.05, \*\**P* < 0.01.

control group. In contrast, cells within Bevacizumab-treated tumors displayed distinct nuclear localization of  $\beta$ -Catenin (typically as the white arrow indicated). The activated Wnt/ $\beta$ -Catenin pathway was probably induced by Bevacizumab *via* generating intratumoral hypoxia in human TNBC xenografts [11, 30]. Compared to Bevacizumab group, we found Oxymatrine inhibited the accumulation of nuclear  $\beta$ -Catenin and reduced total expression level of  $\beta$ -Catenin, which were in agreement with the data *in vitro*. Hence, we came to conclusion that Oxymatrine could abate Wnt/ $\beta$ -Catenin pathway of TNBC activated by Bevacizumab *in vivo*.

Multiple researches have verified that Wnt/β-Catenin pathway played a key role in the behavior of CSCs, especially the self-renew capacity of cancer cells. Then, mammosphere formation assay was executed to evaluate the self-renew capacity of TNBC cells. Figure 6C, 6D showed an increase in mammosphere formation in Bevacizumab treatment group compared to the control, which indicated Bevacizumab could enhance the self-renew capacity of TNBC cells. It was also observed in Figure 6C, 6D that, Oxymatrine attenuated mammosphere formation capacity of TNBC cells under conditions with and without Bevacizumab treatment. Moreover, compared to Bevacizumab + Docetaxel group, TNBC cells of Bevacizumab + Oxymatrine group exhibited significantly weaker mammosphere formation capacity (MDA-MB-231: \*P < 0.05, MDA-MB-468: \*P < 0.05), which highlighted the advantage of Oxymatrine when combined with Bevacizumab. Above all, Oxymatrine suppressed Bevacizumab-enhanced self-renew capacity of TNBC cells, which predicted Oxymatrine could prevent TNBC recurrence and metastasis after bevacizumab treatment.

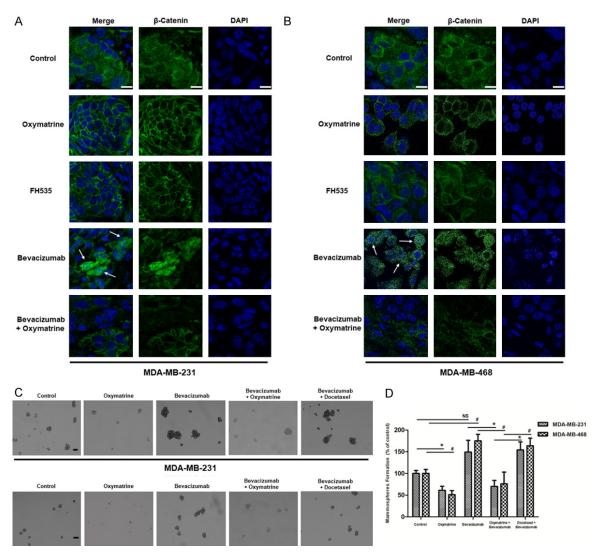
# Oxymatrine exhibited potential anti-angiogenic activity in vivo and in vitro

Tumor angiogenesis is one of the important components of tumor microenvironment, play-

ing an oncogenic role in tumorigenesis, also is the target of the anti-angiogenic agent. The CD31 staining (**Figure 7A**, **7B**) was executed to evaluate tumor angiogenesis of each treatment group. We were delighted to find that Oxymatrine treatment reduced the blood vessel density effectively (Oxymatrine group vs. PBS control). The remarkably reduced intensity of CD31 staining in Bevacizumab + Oxymatrine group indicated that the density of tumor neovascularization was significantly lower in Bevacizumab + Oxymatrine group compared to Bevacizumab group or Oxymatrine group, further illustrating the synergistic anti-angiogenesis synergistic effect of Bevacizumab and Oxymatrine.

VEGF, a key pro-angiogenic factor, was then detected by IHC and Western blot. As an important target gene of Wnt/ $\beta$ -Catenin pathway, VE-GF was supposed to be down regulated by Oxymatrine. **Figure 8A-D** corroborated the speculation that the expression level of VEGFA declined after Oxymatrine treatment. Compared to other treatment groups, Bevacizumab + Oxymatrine group exhibited the lowest expression of VEGFA, which was consistent with the results of CD31 staining.

To further verify the anti-angiogenic potential of Oxymatrine, we studied its effects on the proliferation, transwell invasion, and tube formation of HUVECs. Figure 8E showed that, Oxymatrine effectively inhibited the growth of HU-VECs in a dose-dependent manner in vitro. After 48 h effect of Oxymatrine, the  $IC_{50}$  value was 0.51 mM. However, no inhibition on HE-K293 was seen. As shown in Figure 8F, 8G, the invasive cells to the lower chamber were significantly reduced with increasing concentration of Oxymatrine, and the inhibitory rate of 4 mM Oxymatrine on the invasion of HUVECs reached 76.2 ± 3.43%. The maturation of endothelial cells into a capillary tube, a critical early step of angiogenesis, was gradually abrogated with increasing concentrations of Oxymatrine, and the inhibitory rate of 4 mM Oxymatrine on the tube formation of HUVECs reached 92.8 ±



**Figure 6.** Oxymatrine repressed Bevacizumab-induced accumulation of β-Catenin in nucleus of MDA-MB-231/MDA-MB-468-tumor xenografts, and reduced mammosphere formation of tumor cells. A, B. IF staining of β-Catenin (green) on paraffin sections of MDA-MB-231/MDA-MB-468 xenografted tumor. Nuclei were counterstained with DAPI (blue), bar = 10 µm. C.  $5 \times 10^3$  single cells isolated from MDA-MB-231/MDA-MB-468 xenografts were cultured in low attachment 6 well culture plates for 2 weeks to form the mammospheres, bar = 100 µm. D. Quantitation of the mammosphere-forming efficiency. The numbers of mammospheres (> 60 µm) were recorded after 2 weeks of culture. Data were presented as the mean ± SD, n = 3, \**P* < 0.05, NS: no significance (MDA-MB-231 cells); \**P* < 0.05 (MDA-MB-468 cells).

1.89% (Figure 8H, 8I). Oxymatrine did not show any cytotoxicity in the transwell and tube formation assays (data not shown). In conclusion, Oxymatrine exhibited superior anti-angiogenic activities *in vivo* and *in vitro*, and enhanced the anti-angiogenic effect when combined with Bevacizumab.

#### Discussion

Anti-angiogenic therapy has been demonstrated to hold significant potential for the treatment of various cancers, including TNBC. However, the efficacy of such therapy, especially in TNBC patients, has been called into question, as recent clinical trials reveal only limited effectiveness of anti-angiogenic agents in prolonging patient survival [40]. Conley SJ et al [11], found that anti-angiogenic agents increased the population of CSCs by generating intratumoral hypoxia in human TNBC xenografts, further demonstrated hypoxia-driven CSC stimulation limited the effectiveness of antiangiogenic agents. Huang W et al [30], showed that

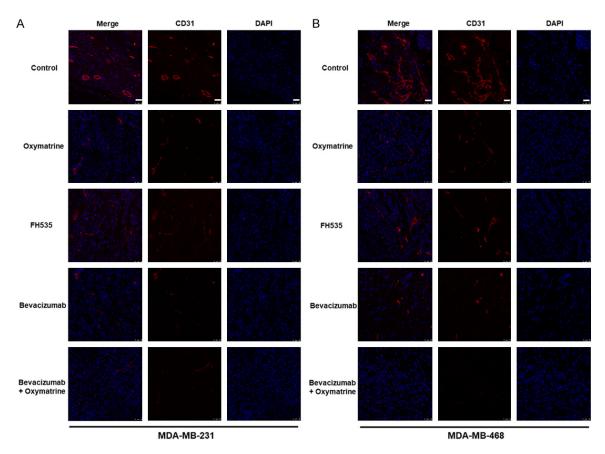
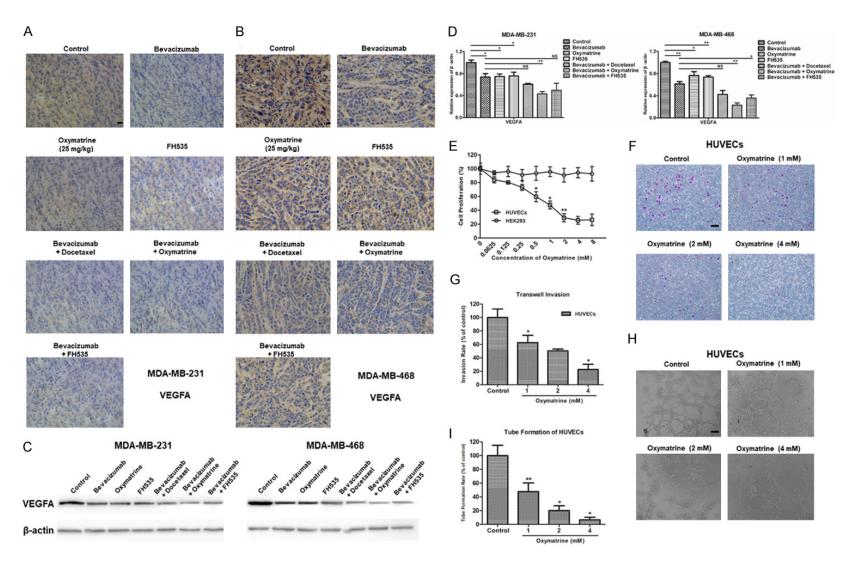


Figure 7. Oxymatrine combined with Bevacizumab reduced markers of angiogenesis in MDA-MB-231/MDA-MB-468-tumor xenografts. A, B. IF staining of CD31 on paraffin sections of MDA-MB-231/MDA-MB-468 implanted tumor. The CD31<sup>+</sup> blood vessels were identified with an anti-CD31 antibody (red), and nuclei were counterstained with DAPI (blue), bar =  $25 \mu m$ .

Bevacizumab induced EMT and promoted tumor invasion in malignancies, which compromised its efficacy severely. Sun H et al [40], indicated that anti-angiogenic treatment induced the formation of vasculogenic mimicry (VM) channels via causing acute hypoxic stress in TNBC; this effect might influence eventual tumor progression, invasion and angiogenesis rebound. Furthermore, there has been increasing evidence for that, the Wnt/β-Catenin pathway involved in EMT/CSC was activated in Sunitinib/Bevacizumab-treated TNBC cells under hypoxia/serum deprivation condition in vitro and in Sunitinib/Bevacizumab-treated mouse xenografts [41, 42]. These studies suggest that to improve patient outcome, anti-angiogenic agents need to be combined with Wnt/β-Catenin pathway/CSC-targeting drugs. Additional approaches for targeting the hypoxic tumor microenvironment are being investigated in pre-clinical and clinical studies, which also have potential for producing synthetic lethality when combined with anti-angiogenic therapy [43, 44].

Through a series of experiments, Marjon PL et al [45], found that TNBC cell line was sensitive to serum deprivation rather than hypoxia. There's more evidence that hypoxia was just one of the triggering factors for the production of tumor specific growth factors (TSGFs) [46, 47]. Thus, in this study, we utilized cell serum starvation model to confirm Bevacizumab treatment induced TNBC cells transwell, migration in vitro. Figure 1A, 1C revealed that, compared to serum starvation control group, invasion and migration of TNBC cells were enhanced after Bevacizumab treatment, which could be due to the expression of invasive factors and transfer factors stimulated by the ablation of VEGF pathway [48]. Figures 1D and 4 indicated that, in serum-deprived environment, Bevacizumab increased the subpopulation of TNBC stem-like cells and promoted EMT of TNBC via activating



**Figure 8.** Oxymatrine exhibited potential anti-angiogenic activity in *vivo* and in *vitro*. (A, B) IHC staining of VEGFA (brown staining) on paraffin sections of MDA-MB-231/MDA-MB-468 xenografted tumor, bar = 20  $\mu$ m. (C) A subset of MDA-MB-231/MDA-MB-468 tumor tissues were subjected to Western blot to examine the VEGFA protein levels.  $\beta$ -actin was used as loading control for proteins. (D) Relative expression levels of VEGFA protein after gray scanning. Data were presented as the mean  $\pm$  SD, n = 3, \*P < 0.05, \*\*P < 0.01, NS: no significance. (E) The viability of HUVECs was assessed by MTT assay at 48 h after treatment with different concentrations of Oxymatrine (ranging from 0 to 8 mg/mL), data were presented as the mean  $\pm$  SD, n = 3, \*P < 0.05, \*\*P < 0.01, vs. the previous group. (F) Microscopic views from transwell assay to estimate HUVECs invasion following 24 h treatment of different concentrations of Oxymatrine (0, 1, 2, and 4 mM), bar = 100  $\mu$ m. (G) Quantitative analysis of (F) by Image J. (H) Photomicrographs of HUVECs tube formation treated by different concentrations of Oxymatrine for 8 h, bar = 100  $\mu$ m. (I) Quantitative analysis of (H) by Image J. The quantitative analyses of transwell invasion and HUVECs tube formation (a complete polygon was considered as a tube) were based on the mean of the 5 regions of each group. Data were presented as the mean  $\pm$  SD, n = 5, \*P < 0.05, \*\*P < 0.01, vs. the previous group.

Wht/ $\beta$ -Catenin pathway, which probably accounted for the invasion and migration of TNBC cells. Similarly, Wht/ $\beta$ -Catenin pathway was also activated in Bevacizumab-treated mouse xenografts (**Figure 6A, 6B**), mainly due to the generation of tumor hypoxia [49]. As a key regulator of breast CSC self-renewal [38], activated Wht/ $\beta$ -Catenin pathway further enhanced mammosphere formation capacity of TNBC cells (**Figure 6C**).

This study showed that, Oxymatrine reversed the Bevacizumab-activated Wnt/β-Catenin pathway and Bevacizumab-induced EMT in TNBC effectively; in addition, Oxymatrine exhibited potent anti-angiogenic activities, which implied Oxymatrine was a potential candidate for the drug combination with anti-angiogenic agents in the treatment of TNBC. Figures 2, 3 suggested Oxymatrine inhibited the invasion, migration of TNBC cells, and decreased the subpopulation of TNBC stem-like cells through repressing EMT and Wnt/ $\beta$ -Catenin pathway stimulated by Bevacizumab. Furthermore, in vivo (Figure 6A, **6B**), Wnt/β-Catenin pathway was disrupted remarkably in the Bevacizumab + Oxymatrine group compared to the Bevacizumab group, which was consistent with cell studies. Moreover, Figures 7 and 8A-D clearly demonstrated Oxymatrine strengthened the anti-angiogenic potency of Bevacizumab in a synergistic mechanism. In view of the above, Bevacizumab + Oxymatrine treatment achieved a better curative effect, which embodied in tumor growth curves (Figure 5B, 5C) and tumor proliferation (Figure 5D, 5E). The enhanced TNBC CSC self-renewal capacity portended tumor recurrence and metastasis, going straight to the heart of Bevacizumab's problem, which was also an key problem during the clinical application of Docetaxel. Notably, Figure 6C verified that Oxymatrine suppressed Bevacizumab-enhanced selfrenew capacity of TNBC cells, indicating the combination of Oxymatrine and Bevacizumab possibly reduced the risk of tumor recurrence and metastasis caused by Bevacizumab treatment alone.

Since anti-angiogenic agents increased TNBC CSCs via the generation of tumor hypoxia, which was demonstrated to be mediated by hypoxia-inducible factor  $1\alpha$  (HIF- $1\alpha$ ), an important upstream factor of the  $\beta$ -Catenin response [38, 50, 51]; Oxymatrine was hypothesized to stimulate TNBC CSC behavior as a direct effect of

the hypoxia induced by the angiogenic inhibition. While, mammosphere formation assay (Figure 6C) exhibited a contrary result that, Oxymatrine negatively regulated TNBC stem cell self-renewal. Figure 6A, 6B indicated the effects of Oxymatrine was at least partly regulated by blocking  $\beta$ -Catenin signaling; but the effects of Oxymatrine on the critical factor HIF-1 $\alpha$  needed to be further investigated, on account of that HIF-1 $\alpha$  was reported to enhance  $\beta$ -Catenin signaling in hypoxic embryonic stem cells and be involved in EMT.

The findings presented here provided additional evidence suggesting that Oxymatrine was sufficient to inhibit Bevacizumab-induced cancer stem-like properties and EMT with a suppressive effect on Wnt/ $\beta$ -Catenin signaling. The combination of Oxymatrine and Bevacizumab achieved better anti-tumor and anti-angiogenic effects, further reduced the risk of TNBC recurrence and metastasis induced by Bevacizumab treatment alone. In conclusion, the concurrent use of Oxymatrine and Bevacizumab provides an effective candidate for the clinical treatment of TNBC, and lays foundation for the synergistic strategy of combining anti-angiogenic agents with Wnt/CSC-targeting agents.

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# Disclosure of conflict of interest

None.

# Abbreviations

ALDH, aldehyde dehydrogenase; CSC, cancer stem cell; DEAB, diethylaminobenzaldehyde; ECGS, endothelial cell growth supplement; ECM, endothelial culture medium; EGF, epidermal growth factor; EMT, epithelial-mesenchymal cell transformation; FGF, fibroblast growth factor; HIF-1 $\alpha$ , hypoxia-inducible factor 1 $\alpha$ ; HUVECs, human umbilical vascular endothelial cells; IF, immunofluorescence; IHC, immunohistochemistry; mCRC, metastatic colorectal cancer; NSCLS, non-small cell lung cancer; ORR, objective response rate; OS, overall survival; PFS, progression-free survival; TCM, traditional Chinese medicine; TNBC, triple-negative breast cancer; TSGF, tumor specific growth factor; VEGF-A, vascular endothelial growth factor A.

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#### References

- [1] Siegel RL, Miller KD and Jemal A. Cancer statistics, 2019. CA Cancer J Clin 2019; 69: 7-34.
- [2] Sikov WM, Berry DA, Perou CM, Singh B, Cirrincione CT, Tolaney SM, Kuzma CS, Pluard TJ, Somlo G, Port ER, Golshan M, Bellon JR, Collyar D, Hahn OM, Carey LA, Hudis CA and Winer EP. Impact of the addition of carboplatin and/or bevacizumab to neoadjuvant once-perweek paclitaxel followed by dose-dense doxorubicin and cyclophosphamide on pathologic complete response rates in stage II to III triple-negative breast cancer: CALGB 40603 (Alliance). J Clin Oncol 2015; 33: 13-21.
- [3] Bardia A, Mayer IA, Vahdat LT, Tolaney SM, Isakoff SJ, Diamond JR, O'Shaughnessy J, Moroose RL, Santin AD, Abramson VG, Shah NC, Rugo HS, Goldenberg DM, Sweidan AM, Iannone R, Washkowitz S, Sharkey RM, Wegener WA and Kalinsky K. Sacituzumab govitecan-hziy in refractory metastatic triple-negative breast cancer. N Engl J Med 2019; 380: 741-751.
- [4] Carey L, Winer E, Viale G, Cameron D and Gianni L. Triple-negative breast cancer: disease entity or title of convenience? Nat Rev Clin Oncol 2010; 7: 683-692.
- [5] Cameron D, Brown J, Dent R, Jackisch C, Mackey J, Pivot X, Steger GG, Suter TM, Toi M, Parmar M, Laeufle R, Im YH, Romieu G, Harvey V, Lipatov O, Pienkowski T, Cottu P, Chan A, Im SA, Hall PS, Bubuteishvili-Pacaud L, Henschel V, Deurloo RJ, Pallaud C and Bell R. Adjuvant bevacizumab-containing therapy in triple-negative breast cancer (BEATRICE): primary results of a randomised, phase 3 trial. Lancet Oncol 2013; 14: 933-942.

- [6] Dieras V, Campone M, Yardley DA, Romieu G, Valero V, Isakoff SJ, Koeppen H, Wilson TR, Xiao Y, Shames DS, Mocci S, Chen M and Schmid P. Randomized, phase II, placebo-controlled trial of onartuzumab and/or bevacizumab in combination with weekly paclitaxel in patients with metastatic triple-negative breast cancer. Ann Oncol 2015; 26: 1904-1910.
- [7] Miles DW, Dieras V, Cortes J, Duenne AA, Yi J and O'Shaughnessy J. First-line bevacizumab in combination with chemotherapy for HER2negative metastatic breast cancer: pooled and subgroup analyses of data from 2447 patients. Ann Oncol 2013; 24: 2773-2780.
- [8] Miller K, Wang M, Gralow J, Dickler M, Cobleigh M, Perez EA, Shenkier T, Cella D and Davidson NE. Paclitaxel plus bevacizumab versus paclitaxel alone for metastatic breast cancer. N Engl J Med 2007; 357: 2666-2676.
- [9] Hurwitz H, Fehrenbacher L, Novotny W, Cartwright T, Hainsworth J, Heim W, Berlin J, Baron A, Griffing S, Holmgren E, Ferrara N, Fyfe G, Rogers B, Ross R and Kabbinavar F. Bevacizumab plus irinotecan, fluorouracil, and leucovorin for metastatic colorectal cancer. N Engl J Med 2004; 350: 2335-2342.
- [10] Gurney A, Axelrod F, Bond CJ, Cain J, Chartier C, Donigan L, Fischer M, Chaudhari A, Ji M, Kapoun AM, Lam A, Lazetic S, Ma S, Mitra S, Park IK, Pickell K, Sato A, Satyal S, Stroud M, Tran H, Yen WC, Lewicki J and Hoey T. Wnt pathway inhibition via the targeting of Frizzled receptors results in decreased growth and tumorigenicity of human tumors. Proc Natl Acad Sci U S A 2012; 109: 11717-11722.
- [11] Conley SJ, Gheordunescu E, Kakarala P, Newman B, Korkaya H, Heath AN, Clouthier SG and Wicha MS. Antiangiogenic agents increase breast cancer stem cells via the generation of tumor hypoxia. Proc Natl Acad Sci U S A 2012; 109: 2784-2789.
- [12] Linke F, Harenberg M, Nietert MM, Zaunig S, von Bonin F, Arlt A, Szczepanowski M, Weich HA, Lutz S, Dullin C, Janovska P, Krafcikova M, Trantirek L, Ovesna P, Klapper W, Beissbarth T, Alves F, Bryja V, Trumper L, Wilting J and Kube D. Microenvironmental interactions between endothelial and lymphoma cells: a role for the canonical WNT pathway in Hodgkin lymphoma. Leukemia 2017; 31: 361-372.
- [13] Zhang L, Wang H, Li C, Zhao Y, Wu L, Du X and Han Z. VEGF-A/neuropilin 1 pathway confers cancer stemness via activating wnt/beta-Catenin axis in breast cancer cells. Cell Physiol Biochem 2017; 44: 1251-1262.
- [14] Zhang L, Xiong W, Xiong Y, Liu H and Liu Y. 17 beta-Estradiol promotes vascular endothelial growth factor expression via the Wnt/betacatenin pathway during the pathogenesis of

endometriosis. Mol Hum Reprod 2016; 22: 526-535.

- [15] Paez-Ribes M, Allen E, Hudock J, Takeda T, Okuyama H, Vinals F, Inoue M, Bergers G, Hanahan D and Casanovas O. Antiangiogenic therapy elicits malignant progression of tumors to increased local invasion and distant metastasis. Cancer Cell 2009; 15: 220-231.
- [16] Saleem M, Maddodi N, Abu Zaid M, Khan N, bin Hafeez B, Asim M, Suh Y, Yun JM, Setaluri V and Mukhtar H. Lupeol inhibits growth of highly aggressive human metastatic melanoma cells in vitro and in vivo by inducing apoptosis. Clin Cancer Res 2008; 14: 2119-2127.
- [17] Wang XJ, Zhou RJ, Zhang N and Jing Z. 20(S)ginsenoside Rg3 sensitizes human non-small cell lung cancer cells to icotinib through inhibition of autophagy. Eur J Pharmacol 2019; 850: 141-149.
- [18] Zhou ZL, Yang YX, Ding J, Li YC and Miao ZH. Triptolide: structural modifications, structureactivity relationships, bioactivities, clinical development and mechanisms. Nat Prod Rep 2012; 29: 457-475.
- [19] Li Z, Zhou ZL, Miao ZH, Lin LP, Feng HJ, Tong LJ, Ding J and Li YC. Design and synthesis of novel C14-hydroxyl substituted triptolide derivatives as potential selective antitumor agents. J Med Chem 2009; 52: 5115-5123.
- [20] Liu Y, Xu Y, Ji W, Li X, Sun B, Gao Q and Su C. Anti-tumor activities of matrine and oxymatrine: literature review. Tumour Biol 2014; 35: 5111-5119.
- [21] Lu ML, Xiang XH and Xia SH. Potential signaling pathways involved in the clinical application of oxymatrine. Phytother Res 2016; 30: 1104-1112.
- [22] Wu J, Cai Y, Li M, Zhang Y, Li H and Tan Z. Oxymatrine promotes s-phase arrest and inhibits cell proliferation of human breast cancer cells in vitro through mitochondria-mediated apoptosis. Biol Pharm Bull 2017; 40: 1232-1239.
- [23] Li W, Yu X, Tan S, Liu W, Zhou L and Liu H. Oxymatrine inhibits non-small cell lung cancer via suppression of EGFR signaling pathway. Cancer Med 2018; 7: 208-218.
- [24] Lin B, Li D and Zhang L. Oxymatrine mediates Bax and Bcl-2 expression in human breast cancer MCF-7 cells. Pharmazie 2016; 71: 154-157.
- [25] Zhang Y, Piao B, Zhang Y, Hua B, Hou W, Xu W, Qi X, Zhu X, Pei Y and Lin H. Oxymatrine diminishes the side population and inhibits the expression of beta-catenin in MCF-7 breast cancer cells. Med Oncol 2011; 28 Suppl 1: S99-107.
- [26] Song MQ, Zhu JS, Chen JL, Wang L, Da W, Zhu L and Zhang WP. Synergistic effect of oxyma-

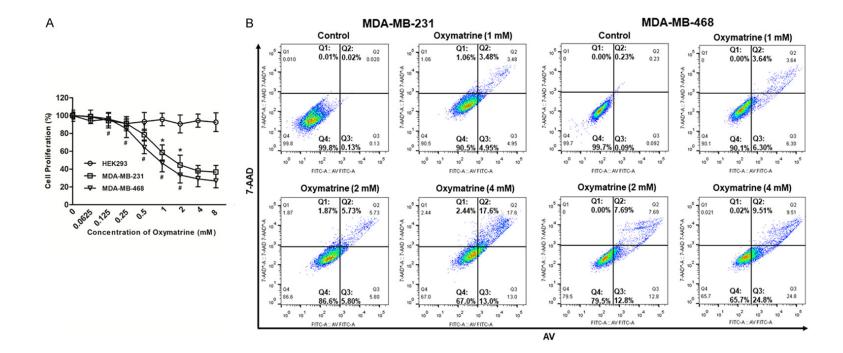
trine and angiogenesis inhibitor NM-3 on modulating apoptosis in human gastric cancer cells. World J Gastroenterol 2007; 13: 1788-1793.

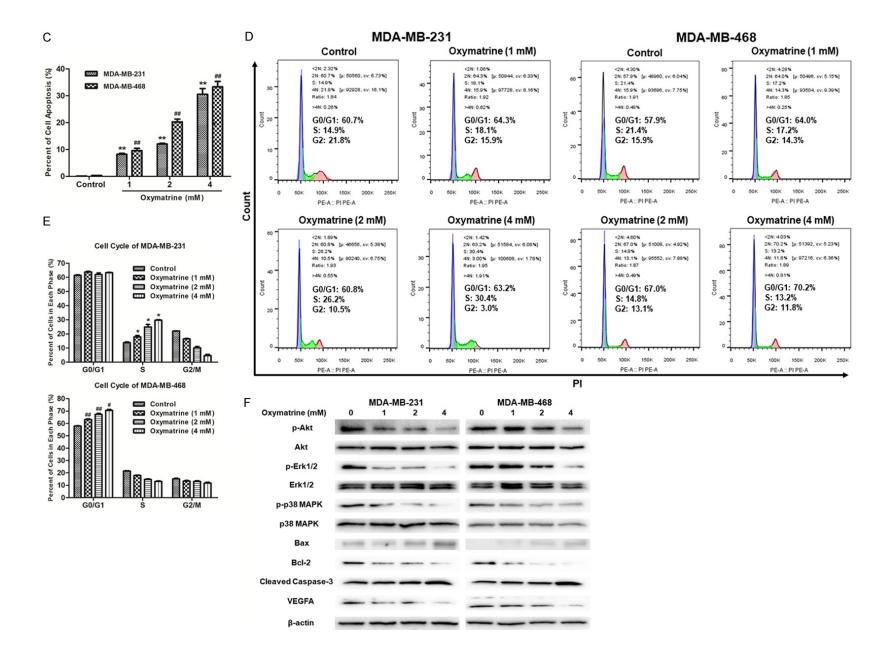
- [27] Xie W, Li D, Zhang J, Li Z, Acheampong DO, He Y, Wang Y, Chen Z and Wang M. Generation and characterization of a novel human IgG1 antibody against vascular endothelial growth factor receptor 2. Cancer Immunol Immunother 2014; 63: 877-888.
- [28] Xie W, Zhang Y, He Y, Zhang K, Wan G, Huang Y, Zhou Z, Huang G and Wang J. A novel recombinant human Frizzled-7 protein exhibits antitumor activity against triple negative breast cancer via abating Wnt/beta-catenin pathway. Int J Biochem Cell Biol 2018; 103: 45-55.
- [29] Xu Z, Wang Z, Jia X, Wang L, Chen Z, Wang S, Wang M, Zhang J and Wu M. MMGZ01, an anti-DLL4 monoclonal antibody, promotes nonfunctional vessels and inhibits breast tumor growth. Cancer Lett 2016; 372: 118-127.
- [30] Huang W, Zhang C, Cui M, Niu J and Ding W. Inhibition of bevacizumab-induced epithelialmesenchymal transition by BATF2 overexpression involves the suppression of wnt/betacatenin signaling in glioblastoma cells. Anticancer Res 2017; 37: 4285-4294.
- [31] Gupta GP and Massague J. Cancer metastasis: building a framework. Cell 2006; 127: 679-695.
- [32] Mani SA, Guo W, Liao MJ, Eaton EN, Ayyanan A, Zhou AY, Brooks M, Reinhard F, Zhang CC, Shipitsin M, Campbell LL, Polyak K, Brisken C, Yang J and Weinberg RA. The epithelial-mesenchymal transition generates cells with properties of stem cells. Cell 2008; 133: 704-715.
- [33] Lo JF, Yu CC, Chiou SH, Huang CY, Jan CI, Lin SC, Liu CJ, Hu WY and Yu YH. The epithelialmesenchymal transition mediator S100A4 maintains cancer-initiating cells in head and neck cancers. Cancer Res 2011; 71: 1912-1923.
- [34] Qiu M, Peng Q, Jiang I, Carroll C, Han G, Rymer I, Lippincott J, Zachwieja J, Gajiwala K, Kraynov E, Thibault S, Stone D, Gao Y, Sofia S, Gallo J, Li G, Yang J, Li K and Wei P. Specific inhibition of Notch1 signaling enhances the antitumor efficacy of chemotherapy in triple negative breast cancer through reduction of cancer stem cells. Cancer Lett 2013; 328: 261-270.
- [35] Bhola NE, Balko JM, Dugger TC, Kuba MG, Sanchez V, Sanders M, Stanford J, Cook RS and Arteaga CL. TGF-beta inhibition enhances chemotherapy action against triple-negative breast cancer. J Clin Invest 2013; 123: 1348-1358.
- [36] Liu P, Kumar IS, Brown S, Kannappan V, Tawari PE, Tang JZ, Jiang W, Armesilla AL, Darling JL and Wang W. Disulfiram targets cancer stemlike cells and reverses resistance and cross-

resistance in acquired paclitaxel-resistant triple-negative breast cancer cells. Br J Cancer 2013; 109: 1876-1885.

- [37] Chang WW, Hu FW, Yu CC, Wang HH, Feng HP, Lan C, Tsai LL and Chang YC. Quercetin in elimination of tumor initiating stem-like and mesenchymal transformation property in head and neck cancer. Head Neck 2013; 35: 413-419.
- [38] Korkaya H, Paulson A, Charafe-Jauffret E, Ginestier C, Brown M, Dutcher J, Clouthier SG and Wicha MS. Regulation of mammary stem/ progenitor cells by PTEN/Akt/beta-catenin signaling. PLoS Biol 2009; 7: e1000121.
- [39] Yang L, Wu X, Wang Y, Zhang K, Wu J, Yuan YC, Deng X, Chen L, Kim CC, Lau S, Somlo G and Yen Y. FZD7 has a critical role in cell proliferation in triple negative breast cancer. Oncogene 2011; 30: 4437-4446.
- [40] Sun H, Zhang D, Yao Z, Lin X, Liu J, Gu Q, Dong X, Liu F, Wang Y, Yao N, Cheng S, Li L and Sun S. Anti-angiogenic treatment promotes triplenegative breast cancer invasion via vasculogenic mimicry. Cancer Biol Ther 2017; 18: 205-213.
- [41] Ebos JM, Lee CR, Cruz-Munoz W, Bjarnason GA, Christensen JG and Kerbel RS. Accelerated metastasis after short-term treatment with a potent inhibitor of tumor angiogenesis. Cancer Cell 2009; 15: 232-239.
- [42] Schomber T, Zumsteg A, Strittmatter K, Crnic I, Antoniadis H, Littlewood-Evans A, Wood J and Christofori G. Differential effects of the vascular endothelial growth factor receptor inhibitor PTK787/ZK222584 on tumor angiogenesis and tumor lymphangiogenesis. Mol Cancer Ther 2009; 8: 55-63.
- [43] Vasudev NS, Goh V, Juttla JK, Thompson VL, Larkin JM, Gore M, Nathan PD and Reynolds AR. Changes in tumour vessel density upon treatment with anti-angiogenic agents: relationship with response and resistance to therapy. Br J Cancer 2013; 109: 1230-1242.
- [44] McIntyre A and Harris AL. Metabolic and hypoxic adaptation to anti-angiogenic therapy: a target for induced essentiality. EMBO Mol Med 2015; 7: 368-379.

- [45] Marjon PL, Bobrovnikova-Marjon EV and Abcouwer SF. Expression of the pro-angiogenic factors vascular endothelial growth factor and interleukin-8/CXCL8 by human breast carcinomas is responsive to nutrient deprivation and endoplasmic reticulum stress. Mol Cancer 2004; 3: 4.
- [46] Mazzone M, Dettori D, de Oliveira RL, Loges S, Schmidt T, Jonckx B, Tian YM, Lanahan AA, Pollard P, de Almodovar CR, De Smet F, Vinckier S, Aragones J, Debackere K, Luttun A, Wyns S, Jordan B, Pisacane A, Gallez B, Lampugnani MG, Dejana E, Simons M, Ratcliffe P, Maxwell P and Carmeliet P. Heterozygous deficiency of PHD2 restores tumor oxygenation and inhibits metastasis via endothelial normalization. Cell 2009; 136: 839-851.
- [47] Kallergi G, Markomanolaki H, Giannoukaraki V, Papadaki MA, Strati A, Lianidou ES, Georgoulias V, Mavroudis D and Agelaki S. Hypoxia-inducible factor-1alpha and vascular endothelial growth factor expression in circulating tumor cells of breast cancer patients. Breast Cancer Res 2009; 11: R84.
- [48] Li A, Dubey S, Varney ML, Dave BJ and Singh RK. IL-8 directly enhanced endothelial cell survival, proliferation, and matrix metalloproteinases production and regulated angiogenesis. J Immunol 2003; 170: 3369-3376.
- [49] Liu L, Zhu XD, Wang WQ, Shen Y, Qin Y, Ren ZG, Sun HC and Tang ZY. Activation of beta-catenin by hypoxia in hepatocellular carcinoma contributes to enhanced metastatic potential and poor prognosis. Clin Cancer Res 2010; 16: 2740-2750.
- [50] Mazumdar J, O'Brien WT, Johnson RS, LaManna JC, Chavez JC, Klein PS and Simon MC. 02 regulates stem cells through Wnt/beta-catenin signalling. Nat Cell Biol 2010; 12: 1007-1013.
- [51] Mendez O, Zavadil J, Esencay M, Lukyanov Y, Santovasi D, Wang SC, Newcomb EW and Zagzag D. Knock down of HIF-1alpha in glioma cells reduces migration in vitro and invasion in vivo and impairs their ability to form tumor spheres. Mol Cancer 2010; 9: 133.





Supplementary Figure 1. Oxymatrine inhibited the proliferation and induced the apoptosis of TNBC cells. A. The viability of MDA-MB-231/MDA-MB-468 cells was assessed by MTT assay at 48 h after treatment with different concentrations of Oxymatrine. HEK293 cell was set as a negative control. Oxymatrine effectively inhibited the growth of MDA-MB-231/MDA-MB-468 cells in a dose-dependent manner, and the IC<sub>E0</sub> value was 0.57 mM/0.68 mM respectively. B. Representative plots showing the apoptosis patterns of Oxymatrine treated MDA-MB-231/MDA-MB-468 cells, the percentage of cells in each quadrant was indicated. C. Quantitative analysis of apoptosis assay. D. Representative plots of the cell cycle phases following different concentrations of Oxymatrine treated MDA-MB-231/MDA-MB-468 cells, the percentage of cells in each phase was indicated. E. Statistical analysis of cell numbers at different cell cycle phases. Data were presented as the mean  $\pm$  SD, n = 3, \*P < 0.05, \*\*P < 0.01 (MDA-MB-231 cells), vs. the previous group; \*P < 0.05, \*\*P < 0.01 (MDA-MB-468 cells), vs. the previous group. Oxymatrine promoted S-phase arrest in MDA-MB-231 cells. Unlike MDA-MB-231 cells, Oxymatrine promoted GO/G1-phase arrest in MDA-MB-468 cells. The Oxymatrine-induced S-phase or G0/G1-phase arrest probably accounted for the suppression of cell proliferation and induction of cell apoptosis. F. Preliminary study on the anti-tumor mechanism of Oxymatrine via Western blot. MDA-MB-231/MDA-MB-468 cells were incubated with different concentrations of oxymatrine (0, 1, 2 and 4 mg/mL) for 48 h. The protein expression levels of p-Akt/Akt, p-Erk/Erk, p-p38/p38, VEGFA and the apoptosis-related protein expression levels of Cleaved Caspase-3, Bax, Bcl-2 were evaluated by Western blot. Equal loading of protein was confirmed by stripping the immunoblot and reprobing it for β-actin. Oxymatrine significantly interfered with the phosphorylation of Akt, Erk and p38 MAPK in a dose-dependent manner, strongly suggesting that Oxymatrine inhibited proliferation and migration of TNBC cells by suppressing the activities of Akt, Erk and p38 MAPK protein kinase. Subsquently, examination of apoptosis-associated proteins showed that Oxymatrine treatment greatly reduced the expression of anti-apoptotic protein Bcl-2, also increased the expression of pro-apoptotic protein Bax. In addition, the immunoblot of cleaved Caspase-3 further verified that, Oxymatrine could induce the apoptosis of TNBC cells. It was noteworthy that, consistent with the anti-angiogenetic potency in vitro, Oxymatrine also significantly decreased the level of cytoplasmatic pro-angiogenic VEGF, which needed to be further confirmed in tumor tissue samples.