

Review Article

Genetic and chromosomal dysregulations of progenitor cells in the etiology and pathogenesis of pediatric cancers

Kirtana Reddy, Balaji Ramachandran

Department of Molecular Oncology, Cancer Institute (W.I.A), No. 38, Sardar Patel Road, Adyar, Chennai 600036, India

Received July 2, 2026; Accepted August 25, 2026; Epub August 25, 2026; Published August 30, 2026

Abstract: Pediatric cancers (PCs) differ significantly from adult cancers in terms of etiology and response to treatment. Although environmental factors have been implicated in the development of some PCs, accumulating evidence indicates that these cancers may arise from genetic dysregulation, which introduces errors within the precursor cells and the progenitor cell machinery during the early developmental stages. Such dysregulations are often explained by a “multi-hit model”, in which progenitor cells acquire an initial prenatal mutation that predisposes them to subsequent postnatal hits, eventually driving malignant transformation. This review aims to consolidate emerging evidence on the role of these dysregulated progenitor cells (dPCs) as tumor cells of origin in PCs and to examine how their identity and developmental timing shape tumor phenotype, latency, and clinical behavior. Evidence drawn from monozygotic twin studies, germline predisposition syndromes, and genetically engineered mouse models were discussed for selected PC categories. This review also analyzes the current targeted therapeutic approaches. Such evidences indicate a firm and recurring role for dPCs across diverse PC subtypes, though a complete functional understanding of their dysregulation and the identification of reliable therapeutic targets remain in their early stages. Unlike adult cancers, therapeutic strategies for PCs should account for developmentally timed vulnerabilities intrinsic to their cells of origin, positioning dPCs as a potential therapeutic target.

Keywords: Multi-hit model, germline predisposition, targeted therapy, tumor cell of origin, tumor microenvironment, leukemia, neuroblastoma, Ewing sarcoma, epigenetic dysregulation and dysregulated progenitor cells

Introduction

In children, cancer is the second leading cause of death after accidents [1]. Though prenatal and early life exposures to environmental risk factors such as maternal and paternal smoking, alcohol consumption, and radiation exposure during pregnancy may impact the development of the fetus, they do not form a sole significant attribute for the incidence of various Pediatric cancers (PCs) [2, 3]. Also, PCs are much less complex than adult cancers in terms of mutation burden [3]. They often exhibit specific genetic alterations that are highly oncogenic, such as chromosomal rearrangements or gene fusions, endogenous developmental mutations, and predisposing germline mutations, as in Li-Fraumeni syndrome and Down syndrome, which increase the risk of PCs [4]. These alterations can activate oncogenes or inactivate tumor suppressor genes, profoundly

affecting cell behavior even with fewer overall mutations [5]. The significant impact of relatively few mutations reflects the fact that stem and progenitor cells rely on tightly coordinated gene expression programs and stable chromosomal architecture to self-renew and differentiate appropriately. When these regulatory circuits are disrupted, whether through structural chromosomal abnormalities such as translocations, aneuploidy, and chromothripsis, or through dysregulated expression of key transcription factors and epigenetic regulators, stem and progenitor cells can become locked in an aberrant, proliferative state rather than progressing through normal differentiation [6, 7]. This convergence of chromosomal instability and abnormal gene regulation within the stem cell compartment is increasingly recognized as a defining feature underlying childhood cancers, distinguishing their pathogenesis from the more gradually acquired mutation-

al landscape typical of adult malignancies [8]. Historically, progress in treating PCs has largely relied on the empirical application of multimodal therapy; however, a deeper understanding of the biology and origin of PCs is increasingly recognized as essential to further improving outcomes. Hence, it is necessary to examine not only the range of molecular abnormalities associated with distinct cancer subtypes but also the role of dysregulated progenitor cells (dPCs) during early developmental stages and their transition into tumor cells of origin with unique biological properties, thereby elucidating the process of oncogenesis. This review presents current information, summarizing emerging evidence suggesting that genetically dysregulated progenitor cells (dPCs) serve as tumor cells of origin in PCs.

Significance of dPCs

During embryonic development, cells undergo a highly orchestrated process of rapid division and differentiation, resulting in the formation of specialized tissues and organs. This process is governed by the activation and regulation of various genetic pathways, which can be disrupted by genetic mutations in progenitor cells, thereby generating dPCs [9]. These dysregulated cells are highly susceptible to further genetic mutations due to their active proliferation and genetic changes [10]. For example, in most pediatric malignancies, the origin of tumor cells can be traced to specific progenitor populations; in some cases, however, as in Ewing's sarcoma, the tumor cell of origin remains the subject of ongoing research and is associated with ambiguous stem or progenitor cells [11]. Genetic dysregulation in progenitor or precursor cells that leads to various PCs is summarized in (Table 1). It can be presumed that mutations such as chromosome rearrangements, chromothripsis, and other endogenous genomic alterations can occur in these susceptible dPCs with impaired DNA repair mechanisms during development [12]. The mechanistic basis of such alterations is increasingly attributed to errors occurring during mitosis in rapidly dividing progenitor cells. Mis-segregated chromosomes can become sequestered into micronuclei, where they undergo asynchronous replication and premature chromosome condensation, ultimately leading to fragmentation, or pulverization, of the enclosed chromo-

some. These fragments are subsequently re-incorporated into the daughter nucleus and rejoined, often through error-prone non-homologous end joining, generating the clustered rearrangements characteristic of chromothripsis [13, 14]. Similar catastrophic rearrangements can also arise from dicentric chromosome bridges formed during anaphase, as well as from telomere erosion and replication stress at fragile genomic sites [13]. Progenitor cells are particularly susceptible to these events when canonical DNA damage response and repair pathways are compromised: loss of TP53 function, in particular, permits cells harboring pulverized chromosomes to bypass apoptosis and cell-cycle checkpoints, allowing chromothriptic rearrangements to persist and propagate [14, 15]. Constitutional DNA repair disorders such as ataxia telangiectasia, Nijmegen breakage syndrome, and Bloom syndrome, which impair double-strand break sensing and homologous recombination, similarly predispose progenitor cells to chromosome instability and complex rearrangements [14]. In pediatric malignancies such as medulloblastoma and osteosarcoma, these processes are especially prominent, with chromothripsis detected in a substantial proportion of tumor genomes and frequently co-occurring with germline or somatic TP53 alterations [15, 16]. Collectively, these findings suggest that the combination of high proliferative activity, developmentally permissive chromatin states, and compromised genome surveillance renders progenitor cells uniquely vulnerable to the endogenous genomic catastrophes that form the basis of pediatric oncogenesis.

Beyond chromosomal instability, genetic regulatory abnormalities in progenitor cells commonly arise from the hijacking of normal transcriptional and epigenetic control mechanisms. Chromosomal translocations that generate fusion oncoproteins, such as EWSR1-FLI1 in Ewing sarcoma or PAX3-FOXO1 in alveolar rhabdomyosarcoma, do not merely create novel proteins but actively rewire the progenitor cell's transcriptional network, binding to and activating lineage-inappropriate enhancers and repressing normal differentiation programs [17, 18]. In parallel, aberrant epigenetic reprogramming, including altered DNA methylation at tumor suppressor loci and dysregulated histone modifications, can lock progenitor cells

Progenitor cell dysregulation in pediatric cancers

Table 1. List of Genetic dysregulations of progenitor cells in various pediatric cancers (PCs)

Cancer	Mutations or Dysregulation	Cell of Origin	Onset of Disease	References
AML	<i>KMT2A</i> (MLL), <i>CBFA2T3</i> , <i>MNX1</i> , t(8;21) translocations (<i>RUNX1-RUNXT1</i>), <i>CBFB</i> , and <i>RARA</i>	HSPCs	Infancy	[86]
ALL	<i>KMT2A</i> (MLL), <i>CBFA2T3</i> , <i>MNX1</i> , <i>ETV6-RUNX1</i> (TEL-AML1), and <i>TCF3-PBX1</i>	HSPCs	Post Infancy	
NHL	t(8;14)(q24; lq32), t(1;1 7)(p36;q2 1), t(1;14)(p36;q22) and disorder of immune dysfunction may also predispose the child to NHL	GC-B cells of lymph nodes) and DZ centroblast	Early childhood to adolescents	[87]
Neuroblastoma	<i>PHOX2B</i> , <i>ALK</i> , <i>KIF1Bb</i> and predisposing syndromes Costello syndrome, Noonan syndrome, and neurofibromatosis type 1	Neural crest progenitor cells/sympathoadrenal lineage of neural crest cells and Schwann cell precursors that were recently identified as the source of adrenal chromaffin cells.	Infant to toddler	[88]
Retinoblastoma	<i>RB1</i> gene mutation	retinal progenitor cells and retinal transition cells/maturing cone precursor	Young children	[63, 64]
Ewing Sarcoma	EWS-FLI1 (t(11;22)(q24;q12)); CNV (gain of chromosome 1q, 8, 12 and loss of 9p21 and 16q) and gene mutations such as in <i>STAG2</i> , <i>TP53</i> and <i>Rb1</i> genes	MSCs/NCCs/OCPCs	Children and young adults	[89, 90]
Osteosarcoma	recurrent deletions <i>TP53</i> , <i>RB1</i> , <i>CDKN2A/B</i> genes, or recurrent amplifications, <i>COPS3</i> , <i>CCNE1</i> , <i>MDM2/CDK4</i> , <i>MYC</i> genes, and 6p12.3 amplifications	MSCs/MSD-derived osteogenic cell types	Children and adolescents	[91]
Glioblastoma	H3 gene mutations, alterations in MAP kinase pathway, mutations in <i>TP53</i> or <i>PPM1D</i> , homozygous deletion of <i>CDKN2A</i> or <i>CDKN2B</i> ; <i>NF1</i> ; <i>TST1</i> or <i>CMMRD</i> ; Nevroid BCC Syndrome/(GS)	NSC/OCPCs	Younger children (midline Glioma) and Adolescents and young adults (high grade gliomas)	[36, 92]
-Rhabdomyosarcoma	Fusion-Negative: <i>TP53</i> , <i>RAS</i> Pathway mutations, alterations in the <i>PI3K-AKT-mTOR</i> pathway, and alterations in the <i>MYCN</i> oncogene Fusion-Positive: <i>Pax3:Foxo</i> t(1;13) or t(2;13)	MSCs/EPCs	Children and young adults	[93]

Abbreviations: AML: Acute Myeloid Leukemia; HSPCs: Hematopoietic stem or progenitor cells; ALL: Acute Lymphoblastic Leukemia; NHL: Non-Hodgkins lymphomas; GC: Germinal Center; DZ: Dark Zone; CNV: Copy Number Variation; MSCs: Mesenchymal Stem Cells; NCCs: Neural Crest Cells; OCPCs: Osteochondrogenic Progenitor Cells; *NF1*: Neurofibromatosis Type 1; *TST1*: Turcot Syndrome Type 1; *CMMRD*: Constitutional Mismatch Repair Deficiency; BCC: Basal Cell Carcinoma; GS: Gorlin Syndrome; NSCs: Neural Stem Cell; OCPCs: Oligodendrocyte precursor cells; EPCs: Endothelial progenitor cells.

into a self-renewing, undifferentiated state by silencing the genetic programs that would normally drive maturation [19]. Because progenitor cells rely on finely tuned, developmentally timed activation and repression of transcription factor networks to progress through differentiation, even a single regulatory lesion introduced at a permissive developmental window can propagate across subsequent cell divisions, compounding into the broader dysregulated phenotype that characterizes dPCs. These aberrations can persist throughout the developmental stage of the fetus and can either present with cancer at an early age or in adolescence [20]. Identification of such distinctive age-related patterns in the onset of cancer implies a specific developmental phase during which certain tissues or cells exhibit a heightened susceptibility to molecular and cellular changes leading to the disease [21]. Hence, the complex and heterogeneous nature of cancer biology contributes to ongoing challenges, including genetic variability within tumors, risk of secondary malignancies, aggressive recurrence, and resistance to therapy, which remain difficult to fully overcome [22, 23]. As one avenue for identifying novel cancer therapeutic targets, it makes a compelling case for understanding the origin of tumor-initiating dPCs and their roles in PC initiation, tumorigenesis, and metastasis.

Fundamental basis of precursor cell dysregulations

Studies conducted in mouse models report that PCs exhibit a “multi-hit model” of disease development, which suggests that cancer is caused by the accumulation of multiple genetic mutations or “hits” over time [24]. This model proposes that children born with a small number of mutations in progenitor cells during development are predisposed to additional mutations that accumulate and transform these cells into dPCs as they grow and mature, ultimately leading to cancer. These dPCs are susceptible to malignant transformation and sustain disease progression (**Figure 1**).

Leukemia

A study on the whole genomes of leukemic cells from two sets of monozygotic twins with acute lymphoblastic leukemia (ALL), which is

the most prevalent subtype of leukemia, accounting for approximately 80% of cases, suggests that childhood ALL can develop in the absence of genetic instability and with only a small number of driver mutations [25]. It was revealed that shared, prenatal coding regions with single-nucleotide variants were limited to the putative initiating lesions. In contrast, all other nonsynonymous single-nucleotide variants were distinct between tumors and therefore secondary and postnatal [26]. Furthermore, noncoding mutational changes were almost entirely discordant in twin pairs, likely representing passenger mutations acquired during leukemic cell proliferation. Similarly, another study on monozygotic twins with concordant hyperdiploid B-ALL but different RAS mutations provides further evidence that hyperdiploidy occurs prenatally, with RAS mutations developing postnatally. This supports the multi-hit model, in which environmental exposures did not appear to be a clear contributing factor [27]. Certain germline mutations or syndromes were also known to predispose the child to ALL. These genetic conditions often involve the same genes that are targeted by somatic mutations in ALL blasts, such as *PAX5*, *IKZF1*, *ETV6*, and *PTPN11* [28]. Some of these conditions, including those caused by germline alterations in *PAX5*, *IKZF1*, and *ETV6*, primarily predispose individuals to develop ALL. Examination of the genetic status of leukemic blasts provides important clues to the underlying ALL predisposing condition, such as a low hypodiploid karyotype in blasts from individuals with LFS, or a hypermutator phenotype in blasts from individuals with Constitutional Mismatch Repair Deficiency (CMMRD) [29]. Full-blown leukemia in Down syndrome was dependent on additional genetic mutations, specifically deletion of cohesion genes, including *STAG2* (*STAG2ko*). Such mutations occurred in progenitor cells during the fetal and early postnatal stages; however, adult-derived bone marrow hematopoietic stem and progenitor cells were unable to undergo the same leukemic transformation [30]. This suggests that it is crucial to identify and understand the cell of origin (COO) of PCs and the mechanism of their dysregulation to develop an effective therapy. For instance, a study on AML that transplanted bone marrow cells transduced with a retrovirus expressing NUP98-HOXA9 revealed that the age of the COO determines not only the latency

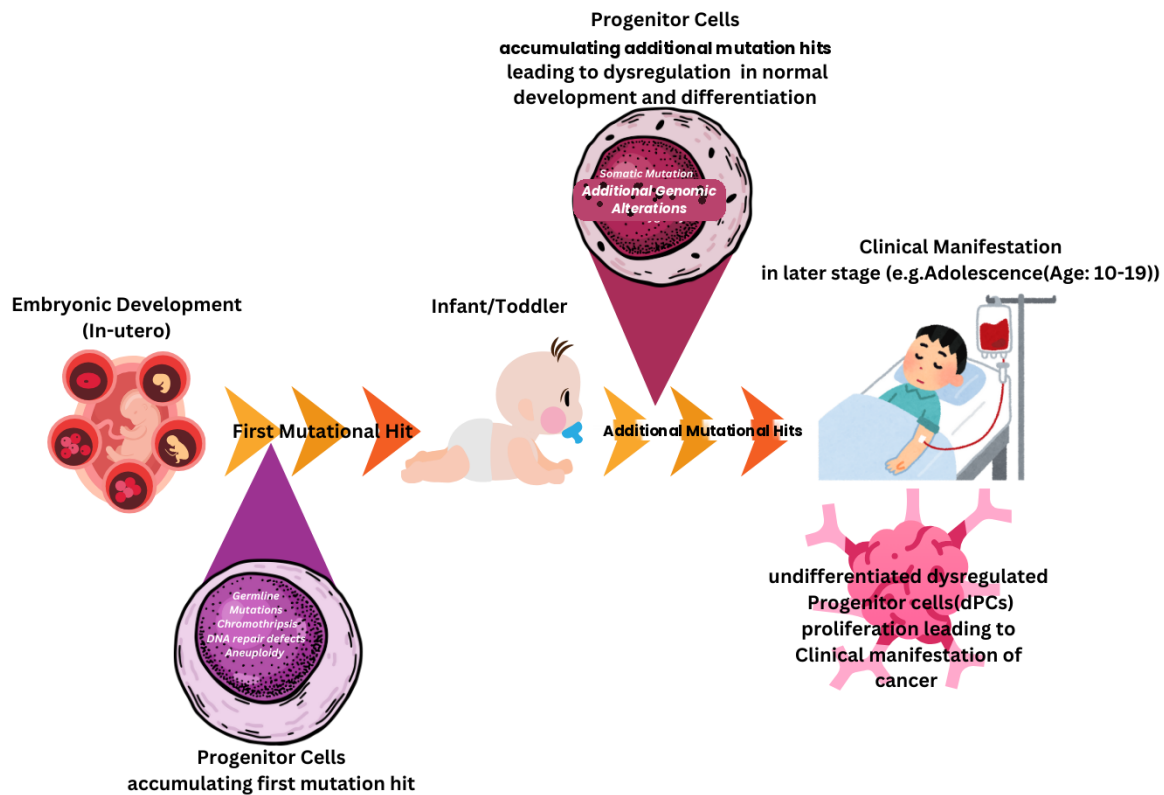


Figure 1. Schematic illustration of multi-hit model of progenitor cell dysregulation. Majority of pediatric cancers (PCs) exhibit the multi-hit model of cancer manifestation, suggesting that the multiple genetic mutation hits are accumulated over time across various developmental phases initiating from *in utero* till adolescence. Mutations acquired in early developmental phases such as during embryonic development (*in utero*) predispose them to additional mutations that transform the progenitor cells to a dysregulated state, ultimately leading to the development of various PCs.

period for disease development but also the lineage phenotype and changes in the bone marrow (BM) niche, highlighting the importance of COO [31]. But in the current scenario, there is still ambiguity in defining or pinpointing a COO for any PCs, as the embryonic developmental phases are layered. However, it is believed that the COO may be hematopoietic stem cells (HSCs) or progenitor cells that undergo prenatal genetic aberrations. Studies have suggested that HSC-independent progenitor cells can also be subject to genetic hits occurring prenatally due to their proliferation rate in a limited time frame [32], leading to a pre-leukemic state which then requires multiple genetic hits before an overt manifestation of the disease [33]. For instance, erythro-myeloid progenitors (EMPs), which sustain fetal myelopoiesis and persist in adults, can harbor mutations such as *PTPN11*, which lead to a distinctive case in children around 2 years of age known as juvenile myelomonocytic leukemia (JMML), a type of leukemia

with high relapse rates after HSC transplantation and plausible in *utero* initiation [34]. Next, lymphoid-primed multipotent progenitors are another type of HSC-independent progenitor that has emerged as a potential cell of origin in B-cell acute lymphoblastic leukemia (B-ALL) with *MLL-AF4* translocation [35].

Pediatric brain tumors

Pediatric brain tumors encompass a heterogeneous group of cancers with diverse developmental origins and genetic mutations. Midline gliomas primarily affect younger children and are associated with specific mutations in the H3 genes. In contrast, high-grade gliomas within the cerebral lobes occur in adolescents and young adults and exhibit different mutations [36]. Medulloblastomas, a type of embryonal tumor, are believed to arise from immature neuronal precursors in the external granular cell layer of the cerebellum, contributing to their

localization primarily in the pediatric cerebellum [37]. Certain rare pediatric CNS tumor types are associated with germline mutations in predisposition genes; *DICER1* [38] and *RB1* [39] mutations can predispose patients to pineoblastoma. Dysplastic cerebellar gangliocytoma is pathognomonic for Cowden syndrome, caused by germline mutations in the *PTEN* gene [37]. Pediatric high-grade gliomas (pHGGs) exhibit distinct molecular alterations, including mutations in the genes *H3F3A* and *HIST1H3B*, leading to specific amino acid substitutions in histone proteins. These mutations exhibit a characteristic distribution pattern, suggesting distinct cell-of-origin for pHGG subgroups [40]. High-grade gliomas (HGGs) in children and adolescents exhibit distinct patterns of occurrence by location and genetic mutations [41]. Recent evidence from genetic mouse models indicates that introducing classic genomic alterations associated with midline gliomas during prenatal brain development does not result in immediate tumor formation but instead gives rise to neoplasms only in the postnatal period. This highlights that glioma initiation is constrained to a specific developmental window, emphasizing the importance of neurodevelopmental timing in tumor latency and onset [42]. Additionally, while these mutations *in vitro* can transform neural stem cells, the resulting tumors do not resemble the typical histological appearance of diffuse midline gliomas when transplanted into the mouse brain. Similar mechanisms have been observed in adult glioblastoma (GBM), where mutations in neural stem cells give rise to tumors only when those stem cells differentiate into oligodendrocyte precursor cells (OPCs) [43], which are neural precursor cells involved in myelination. It is believed that GBMs can originate from both neural stem cells and OPCs, but the specific molecular subtype is determined by the COO [44]. A study explored the cellular origins of CNS tumors by analyzing *INSM1* expression at the transcriptional and translational levels using publicly available microarray data. The findings showed higher *INSM1* transcript levels in medulloblastoma than in extracerebellar embryonal tumors. Notably, most medulloblastomas arise from immature neuronal precursors within the external granular layer of the developing cerebellum, which explains their predominant localization in the pediatric cerebellum [37].

Pediatric embryonal tumors

Neuroblastoma, a pediatric embryonal tumor, is an aggressive childhood cancer arising from the developing peripheral nervous system, specifically the neural crest [45]. While the exact cause is unknown, genetic predisposition and dysregulation of neural crest development are believed to play a role. Neuroblastoma was presumed to originate from sympathoadrenal precursor cells, a specific lineage of neural crest precursor cells that fail to mature into functional nerve cells, and mutations in *MYCN*, *PHOX2B*, *ALK*, and *RAS* pathway genes in these cells have been demonstrated to increase the risk [45]. Thus, understanding the precise origin and the factors that influence the development of neuroblastoma is crucial for developing targeted therapies and improving prognosis to overcome treatment resistance.

Another type of embryonic tumor prevalent in children is retinoblastoma, which arises from the abnormal development of retinal cells, most often in young children. Earlier studies suggested that retinoblastoma develops due to mutations in the *Rb1* gene. However, studies using an inherited mouse model with *Rb* loss challenge this view by findings suggesting that the COO for retinoblastoma may be a specific retinal cell type that is naturally resistant to cell death triggered by *Rb* loss, likely a susceptible progenitor cell [46]. Tumor formation appears to involve these cells escaping a normal growth arrest process that occurs during retinal development, rather than acquiring resistance to cell death [47].

Lymphoma

Pediatric lymphomas are influenced by the age-specific pathway of cell transformation and the cells in which the mutational hits are acquired. Burkitt lymphoma (BL), for example, may develop from a centroblast [48], which, in turn, is a proliferative, immature cell at risk of transformation by a GECC (great effect chromosomal change) carrying Ig/*MYC* translocations. However, the rarity of centroblast transformation by a GECC is attributed to the death of many cells with severe DNA injuries caused by EBV infections. Anaplastic large cell lymphoma (*ALK+* ALCL) is another pediatric lymphoma that primarily affects children and adolescents. It is thought to result from rapid cell transfor-

mation of a differentiating T-cell, possibly a “T-immunoblast” precursor [49]. These T-immunoblasts, which exhibit an anaplastic morphology, have a risk of transformation by GECCs such as t(2;5). T-cell lymphoblastic lymphoma (T-LBL), a rare lymphoma occurring primarily in older children and young adults, was believed to arise from the rapid transformation of a T-lymphoblast, possibly originating in the thymus, as this stage is naturally high-risk because DNA is actively being modified. Thymic involution during puberty may contribute to the risk of T-lymphoblast injury, leading to T-LBL. GECCs, such as certain TCR (T-Cell Receptor) gene-related translocations, were speculated to trigger oncogenic transformation in T-LBL [50].

Cancer of connective tissues

Rhabdomyosarcoma (RMS): The precise COO for RMS remains under investigation, with various candidates being explored. Mesenchymal Stem Cells (MSCs) are considered potential candidates, particularly due to their ability to differentiate into various cell types, including muscle cells [51]. While a few reports suggest a link, conclusive evidence that MSCs are the sole source remains lacking [52]. A study suggested that genetic modifications in muscle progenitor cells, particularly those expressing Myf6 or MyoD, can lead to RMS development, especially embryonal RMS (eRMS) [53]. Fibroadipogenic Progenitor (FAPs) cells were found to contribute to muscle repair and can switch between muscle and fat lineages. FAPs can be considered potential candidates for studying the origin of RMS, especially given their role in dystrophic muscle degeneration [54]. Pericytes and PW1+Pax7-Interstitial Cells are other cell types residing within muscle tissue with progenitor potential. Their role in RMS development is underexplored but cannot be ruled out [55]. RMS development likely involves a combination of genetic and environmental factors: Specific mutations, like *PAX3-FOXO1* gene fusions, are almost always observed in alveolar RMS (aRMS) and are considered a hallmark of this subtype [52]. Mutations in genes like p53, dystrophin, and telomerase may also play a role [56]. The Sonic hedgehog (Shh)-Smoothened (Smo)-Patched (Ptch) signaling pathway appears to be involved in eRMS development, potentially by activating early myogenic genes

in adipogenic cells [57]. RMS development may be specific to certain cell populations or tissues due to the permissive environment of these tissues. For example, Shh signaling induced tumors only in brown adipose tissue, not in white adipose tissue, highlighting the importance of cellular context [57]. A study on FN-RMS (Fusion Negative-Rhabdomyosarcoma) traced the development of cancer and found that it originates from endothelial progenitor cells, but not from the skeletal muscle cells, within the head and neck region. Most importantly, it was found that the hedgehog signaling pathway, aberrantly activated in FN-RMS, causes these progenitor cells to adopt a skeletal muscle-like fate. This finding suggests that FN-RMS may have multiple cellular origins and that understanding these origins is crucial for developing targeted therapies [58].

Ewing sarcoma (ES): ES presents a significant challenge in understanding its cellular origins [59]. Despite the absence of distinct genetic subtypes, suggesting a single source cell, the precise lineage from which this malignancy arises remains elusive [60]. This obscurity is complicated by the tumor’s undifferentiated phenotype and its unique gene expression profile [60, 61]. One hypothesis posits that neural crest stem cells, when exposed to EWSR1-FLI1, exhibit a gene expression pattern strikingly similar to that of ES [62, 63]. MSCs also emerge as a candidate source [64]. These cells are susceptible to EWSR1-FLI1, and conversely, knockdown of this protein shifts the gene expression profile of ES cells closer to that of MSCs. Osteochondrogenic progenitors that give rise to bone and cartilage are also under investigation for their roles in YAP/TAZ signaling in bone development and sarcoma pathogenesis [65].

Emerging therapeutic targets of dPCs

Understanding dysregulation in progenitor cells and its contribution to cancer pathogenesis paves the way for predicting prognosis and identifying new therapeutic targets that can overcome the limitations of treating pediatric patients (**Figure 2**). Several important gene dysregulations that highlight their normal and tumorigenic roles in PCs are listed in (**Table 2**). The unique dysregulated expression of these genes during tumorigenesis underscores their

Progenitor cell dysregulation in pediatric cancers

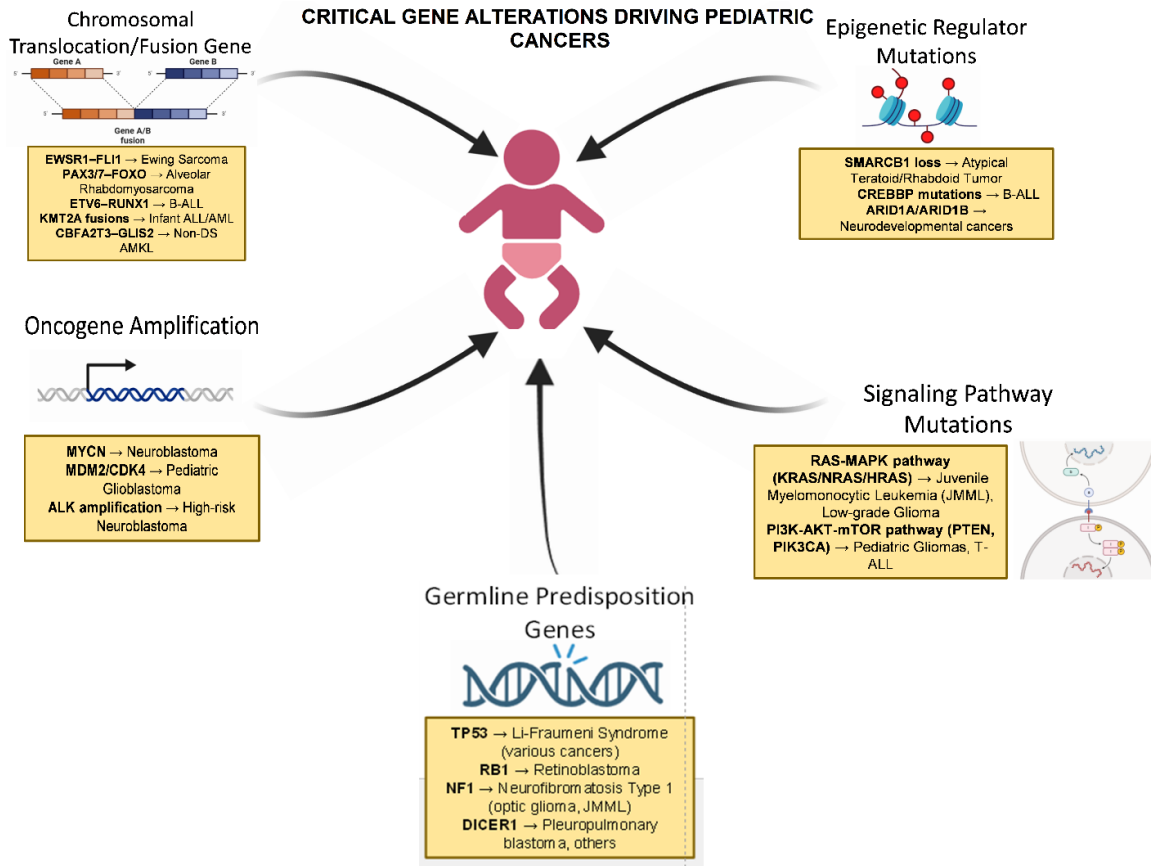


Figure 2. Schematic overview of key genomic perturbations driving pediatric cancers (PCs). Five main genomic alterations linked to pediatric tumors: Chromosomal translocations/fusions, Oncogene amplifications, Germline predisposition genes, Signaling pathway mutations, and Epigenetic regulator mutations.

importance as therapeutic targets. Fusion proteins resulting from chromosomal translocations and other genomic rearrangements are non-physiological and drive PC progression through continuous activation of growth and survival pathways, altered transcriptional regulation, and evasion of apoptosis. Current therapeutic strategies targeting these fusion genes include small-molecule inhibitors like imatinib for Philadelphia chromosome-positive ALL [66] and BCR-ABL-positive chronic myeloid leukemia in pediatric patients [67], and monoclonal antibodies such as blinatumomab for *ETV6-RUNX1* in ALL [68]. Current research on fusion oncoproteins in childhood sarcomas such as ES identifies several promising therapeutic strategies, including RNA interference [69] and immunotherapies targeting breakpoint regions [70]. Next, aberrant DNA methylation contributes significantly to tumorigenesis, as seen in ALL, where silencing of genes such as p15INK4B and p16INK4A has been observed

[71], and in medulloblastoma, where methylation affects key *WNT* pathway genes [72]. Histone modifications, such as H3K27M mutations in DIPG [73] and HDAC overexpression in neuroblastoma [74], further contribute to oncogenesis. Non-coding RNAs, like miR-206 in RMS and *EWSAT1* in ES, also influence tumor behavior [75, 76]. The intricate interplay between the tumor and its tumor microenvironment (TME), comprising extracellular matrix, signaling molecules, and immune cells, determines the fate of tumor progression. For instance, studies on the tumor-progression mouse glioma model showed that macrophages transition from an immune-active to an immune-suppressive state, contributing to therapy resistance. That myeloid-derived suppressor cells (MDSCs) further strengthen this suppression [77]. Elevated levels of immune checkpoint molecules such as CD200, PD-L1, CTLA-4, and B7-H3 on tumor cells suppress anti-tumor immunity by inhibiting T-cell activa-

Progenitor cell dysregulation in pediatric cancers

Table 2. Summary of genes indicating both their developmental role and their tumorigenic functions during their state of dysregulation/translocation or chimeric fusion in various pediatric cancers (PCs)

Cancer/Syndromes	Dysregulated genes	Normal Developmental Role	Tumorigenic Mechanism	References
Ewing Sarcoma	EWSR1-FLI1	<i>EWSR1</i> : Involved in RNA processing and transcription regulation. <i>FLI1</i> : Transcription factor regulating hematopoiesis and endothelial cell development.	EWS-FLI1 fusion acts as an aberrant transcription factor, binds to GGAA microsatellites and dysregulate gene expression. It blocks differentiation and promotes proliferation in mesenchymal progenitor cells.	[94]
Alveolar Rhabdomyosarcoma	PAX3/7-FOXO1	<i>PAX3</i> : Key in early muscle and neural crest development, promotes progenitor migration and commitment. <i>PAX7</i> : Maintains satellite (muscle stem) cells and supports muscle regeneration. <i>FOXO1</i> : Apoptosis regulator, cell proliferation, and stress response	Blocks myogenic differentiation, induces uncontrolled proliferation and survival signals, highly aggressive subtype.	[95, 96]
Neuroblastoma, Medulloblastoma	<i>MYCN</i>	Transcription factor regulating embryonic development, especially neural crest cell proliferation and differentiation.	Uncontrolled cell proliferation, impaired differentiation, It suppresses ER α and (NGF) signaling, maintaining an undifferentiated state and poor prognosis.	[97]
Neuroblastoma	<i>ALK</i>	Receptor tyrosine kinase involved in neuronal development and synaptic signaling.	<i>ALK</i> amplification or mutation leads to constitutive activation of MAPK/PI3K pathways, leading to proliferation.	[98]
Infant gliomas	<i>CDK4/6</i>	Kinases that promote G1-S phase progression in the cell cycle by phosphorylating RB protein.	Amplification overrides cell cycle checkpoints, leading to hyperproliferation.	[99]
Atypical teratoid/rhabdoid tumors	<i>SMARCB1 (INI1)</i>	Core component of the SWI/SNF chromatin remodeling complex; regulates gene accessibility and transcription.	Loss of function leads to widespread dysregulation of transcriptional programs and uncontrolled proliferation.	[100]
Rhabdoid tumors, Medulloblastoma	<i>EZH2</i>	Catalytic subunit of PRC2 complex; adds repressive H3K27me3 marks to silence gene expression.	Gain-of-function mutations lead to excessive gene silencing, particularly of tumor suppressor genes.	[101]
Leukemias	<i>CREBBP</i>	Histone acetyltransferase; promotes open chromatin and transcriptional activation.	Loss of function results in reduced acetylation, leading to impaired tumor suppressor gene expression.	[102]
Neuroblastoma	<i>ALK</i>	Receptor tyrosine kinase involved in neural crest development and survival.	Activating mutations cause constitutive signaling via MAPK and PI3K pathways, promoting growth.	[103]
Low-grade glioma, Juvenile JMML, Embryonal rhabdomyosarcoma	<i>RAS (HRAS, KRAS, NRAS)</i>	Regulates cell proliferation and differentiation via MAPK pathway.	Activating mutations lock RAS in GTP-bound active state, driving uncontrolled cell division.	[104, 105]
Pediatric ALL	<i>JAK2</i>	Key mediator in cytokine receptor signaling for blood cell development.	Mutations result in cytokine-independent activation of JAK/STAT pathway, driving leukemogenesis.	[106]
Li-Fraumeni Syndrome (various sarcomas, brain tumors, leukemias, adrenocortical carcinoma)	<i>TP53</i>	Tumor suppressor; guardian of the genome. Regulates cell cycle arrest, DNA repair, apoptosis.	Germline mutation leads to failure in responding to DNA damage, allowing accumulation of mutations.	[107]
Hereditary Retinoblastoma	<i>RB1</i>	Regulates cell cycle by inhibiting E2F transcription factors.	Loss of both alleles removes cell cycle checkpoint, leading to uncontrolled retinal cell proliferation.	[108]
<i>DICER1</i> syndrome (pleuropulmonary blastoma, ovarian Sertoli-Leydig tumors, others)	<i>DICER1</i>	MicroRNA processing; regulates gene expression post-transcriptionally.	Mutation leads to dysregulation of miRNA networks, disrupting normal development and allowing oncogenesis.	[109]
Fanconi Anemia subtype D1, also linked to pediatric leukemia	<i>BRCA2</i>	Homologous recombination repair of DNA double-strand breaks.	Germline defects cause genomic instability, increasing susceptibility to leukemias and solid tumors.	[110]

Abbreviations: ER α : Estrogen Receptor Alpha; NGF: Nerve Growth Factor; JMML: Juvenile myelomonocytic leukemia; ALL: Acute Lymphoblastic Leukemia.

tion, leading to immune evasion and treatment resistance [78-80]. A dysregulated immune response reflects the state of the TME and can serve as a prognostic biomarker [81]. In recent times, immunotherapy approaches, along with traditional approaches such as surgery, radiation, and chemotherapy, have shown significant success, including CAR T-cell therapy, monoclonal antibody (mAb) therapy, and cytokine-based treatments, which are under active investigation [82]. Approximately 10% of PCs are linked to germline mutation variants, a significantly higher prevalence than in adult cancers [83, 84]. In PCs, as the somatic mutation burden is low compared to adult cancers, germline variants confer inherited risks of aiding tumor progression and present with cancers that are more aggressive and resistant to conventional therapies [39]. These germline mutations often affect key tumor suppressor genes and oncogenes, contributing to early-onset malignancies. Hence, identifying these germline variants is crucial for risk stratification, early diagnosis, and personalized therapeutic approaches. Genetic testing and counseling are recommended for pediatric patients' families to identify inherited predispositions, enabling targeted surveillance and early intervention [85]. Overall, integrating insights into germline variants enhances personalized pediatric oncology. Collectively, these findings underscore that, unlike adult cancers where targeted therapy chiefly addresses mutations accumulated over a lifetime, therapeutic strategies for PCs must additionally account for developmentally timed vulnerabilities intrinsic to dPCs.

Discussion

PCs, despite exhibiting fewer genetic mutations than adult cancers [3], exhibit intricate evolutionary trajectories that contribute significantly to their clinical complexity. During fetal development, a series of progressive changes occur in progenitor cells at the genetic, epigenetic, and microenvironmental levels, pushing them toward a dysregulated state and driving the disease from its origin to its full clinical presentation. Further, heterogeneities arise from a multitude of factors, including distinct mutational landscapes across different tumor types, cellular origins from embryonic and developmental precursors, and variations in TME, which collectively complicate diagnosis and treatment. While heterogeneity poses challenges, it

simultaneously presents unique opportunities for therapeutic and preventive interventions. Targeted therapies tailored to specific genetic mutations or molecular pathways can exploit this heterogeneity to achieve more precise and effective treatment outcomes. It is imperative to understand the role of dPCs and the mode of dysregulation at the functional level to specifically identify appropriate therapeutic targets to combat PCs. However, full understanding of the heterogeneity and complexity of PCs, identification of foolproof drug targets, and effective therapeutics are still evolving and remain at a nascent stage; more research is warranted in this area. In our view, a major limitation constraining this field is the continued reliance on retrospective genomic and transcriptomic correlations to infer the cell of origin, rather than direct, real-time lineage tracing of the dysregulation event as it occurs. Much of the current evidence, while compelling, is circumstantial: it associates a mutational signature or gene expression profile with a candidate progenitor population without directly capturing the transformation event within that cell as it happens *in vivo*. We believe that dual-reporter and inducible lineage-tracing models, capable of marking a specific progenitor population at a defined developmental window and tracking its fate through to overt malignancy, represent a more direct approach to resolving COO ambiguity than correlative profiling alone, particularly for tumors such as Ewing sarcoma and rhabdomyosarcoma, where multiple candidate cells of origin remain under debate. We further contend that the field would benefit from greater emphasis on the developmental timing of the initiating lesion, not merely its identity, since the same mutation can yield markedly different phenotypic and clinical outcomes depending on when in the progenitor cell's differentiation trajectory it occurs. Addressing this temporal dimension, in our opinion, is likely to be as important for identifying viable therapeutic windows as identifying the responsible gene or fusion event itself.

Conclusion

In this review, we have highlighted how dysregulations in progenitor cell populations contribute to the initiation of PCs. To our knowledge, we have comprehensively summarized the existing literature, which strongly indicates that dPCs play a firm role in PCs, and that so-

lutions to target or circumvent them are only emerging. At the same time, their complete functional understanding appears quite challenging. Genetic and chromosomal dysregulations, arising from both structural genomic instability and aberrant transcriptional or epigenetic regulation, converge to lock progenitor cells into an oncogenic trajectory during a developmentally restricted window, distinguishing the pathogenesis of PCs from the more gradually acquired mutational landscape of adult malignancies. Continued application of lineage-tracing and reporter-based models will be essential to translate this understanding into viable, developmentally informed therapeutic strategies. These dPCs could serve as a potential “Achilles heel”, offering promising avenues for targeted therapeutic intervention in the future.

Acknowledgements

The authors wish to acknowledge the support from the Science and Engineering Research Board to Balaji Ramachandran (CRG/2019/000546). During the preparation of this manuscript, the authors used ChatGPT (GPT-5.6) and Grammarly for language editing and clarity. The authors reviewed and edited the content as needed and take full responsibility for the content of this publication.

Disclosure of conflict of interest

None.

Address correspondence to: Dr. Balaji Ramachandran, Department of Molecular Oncology, Cancer Institute (W.I.A), No. 38, Sardar Patel Road, Adyar, Chennai 600036, India. Tel: +91-4422209150; E-mail: r.balaji@cancerinstitutewia.org

References

- [1] Siegel RL, Miller KD, Fuchs HE and Jemal A. Cancer statistics, 2022. *CA Cancer J Clin* 2022; 72: 7-33.
- [2] Wilson DA, Sullivan RM, Smiley JF, Saito M and Rainei C. Developmental alcohol exposure is exhausting: sleep and the enduring consequences of alcohol exposure during development. *Neurosci Biobehav Rev* 2024; 158: 105567.
- [3] Ricci AM, Emeny RT, Bagley PJ, Blunt HB, Butow ME, Morgan A, Alford-Teaster JA, Titus L, Walston RR 3rd and Rees JR. Causes of childhood cancer: a review of the recent literature: part I-childhood factors. *Cancers (Basel)* 2024; 16: 1297.
- [4] Yuan S, Almagro J and Fuchs E. Beyond genetics: driving cancer with the tumour microenvironment behind the wheel. *Nat Rev Cancer* 2024; 24: 274-286.
- [5] Kulyova SA, Ryazankina AA and Vasileva MM. Hereditary diseases and tumors in children. *Support Ther Oncol* 2025; 2: 21-29.
- [6] Zhao Y, Carter R, Natarajan S, Varn FS, Compton DA, Gawad C, Cheng C and Godek KM. Single-cell RNA sequencing reveals the impact of chromosomal instability on glioblastoma cancer stem cells. *BMC Med Genomics* 2019; 12: 79.
- [7] Feinberg AP, Ohlsson R and Henikoff S. The epigenetic progenitor origin of human cancer. *Nat Rev Genet* 2006; 7: 21-33.
- [8] Vrobelova K, Jakl L, Skorvaga M, Kosik P, Durdik M, Markova E, Jakubikova J, Holop M, Kubes M, Cermak M, Puskacova J, Kolenova A and Belyaev I. Genetic instability in HSPC subpopulations of umbilical cord blood from patients with childhood acute lymphoblastic leukemia. *Sci Rep* 2025; 15: 8953.
- [9] Derry WB. Role of developmental pathways in disease. *FEBS J* 2023; 290: 3296-3299.
- [10] Balachandran S and Narendran A. The developmental origins of cancer: a review of the genes expressed in embryonic cells with implications for tumorigenesis. *Genes (Basel)* 2023; 14: 604.
- [11] Hernández-Muñoz I, Cuervas I, Prada E, Pulecio J, Gimeno R, Andrades E, Gómez-González S, Berenguer-Molins P, Acedo-Terrades A, Perra-Bel J, Bódalo-Torruella M, Nonell L, Pérez E, Grases D, Mata C, Yélamos J, Richaud-Patin Y, Vidal E, Cuartero Y, Le Dily F, Suñol M, Manzanares A, Raya A and Mora J. EWS::FLI1 expression in human embryonic mesenchymal stem cells leads to transcriptional reprogramming, defective DNA damage repair and Ewing sarcoma. *Nat Commun* 2025; 16: 9427.
- [12] Trakala M, Aggarwal M, Sniffen C, Zasadil L, Carroll A, Ma D, Su XA, Wangsa D, Meyer A, Sieben CJ, Zhong J, Hsu PH, Paradis G, Ried T, Holland A, Van Deursen J and Amon A. Clonal selection of stable aneuploidies in progenitor cells drives high-prevalence tumorigenesis. *Genes Dev* 2021; 35: 1079-1092.
- [13] Ly P and Cleveland DW. Rebuilding chromosomes after catastrophe: emerging mechanisms of chromothripsis. *Trends Cell Biol* 2017; 27: 917-930.
- [14] Nazaryan-Petersen L, Bjerregaard VA, Nielsen FC, Tommerup N and Tümer Z. Chromothripsis and DNA repair disorders. *J Clin Med* 2020; 9: 613.

- [15] Rausch T, Jones DT, Zapatka M, Stütz AM, Zichner T, Weischenfeldt J, Jäger N, Remke M, Shih D, Northcott PA, Pfaff E, Tica J, Wang Q, Massimi L, Witt H, Bender S, Pleier S, Cin H, Hawkins C, Beck C, von Deimling A, Hans V, Brors B, Eils R, Scheurlen W, Blake J, Benes V, Kulozik AE, Witt O, Martin D, Zhang C, Porat R, Merino DM, Wasserman J, Jabado N, Fontebasso A, Bullinger L, Rücker FG, Döhner K, Döhner H, Koster J, Molenaar JJ, Versteeg R, Kool M, Tabori U, Malkin D, Korshunov A, Taylor MD, Lichter P, Pfister SM and Korbel JO. Genome sequencing of pediatric medulloblastoma links catastrophic DNA rearrangements with TP53 mutations. *Cell* 2012; 148: 59-71.
- [16] Li G, Wu N, Ghabrial J, Stinnett V, Klausner M, Morsberger L, Long P, Baraban E, Gross JM and Zou YS. Chromoanagenesis in osteosarcoma. *Biomolecules* 2025; 15: 833.
- [17] Kucinski J, Tallan A, Taslim C, Vontell AM, Silvius KM, Wang M, Cannon MV, Stanton BZ and Kendall GC. Rhabdomyosarcoma fusion oncoprotein initially pioneers a neural signature in vivo. *Cell Rep* 2025; 44: 115923.
- [18] Gryder BE, Wachtel M, Chang K, El Demerdash O, Aboreden NG, Mohammed W, Ewert W, Pomella S, Rota R, Wei JS, Song Y, Stanton BZ, Schäfer B, Vakoc CR and Khan J. Miswired enhancer logic drives a cancer of the muscle lineage. *iScience* 2020; 23: 101103.
- [19] Liapodimitri A, Tetens AR, Craig-Schwartz J, Lunsford K, Skalitzy KO and Koldobskiy MA. Progress toward epigenetic targeted therapies for childhood cancer. *Cancers (Basel)* 2024; 16: 4149.
- [20] Oliver TRW and Behjati S. Developmental dysregulation of childhood cancer. *Cold Spring Harb Perspect Med* 2024; 14: a041580.
- [21] Zhang S, Xiao X, Yi Y, Wang X, Zhu L, Shen Y, Lin D and Wu C. Tumor initiation and early tumorigenesis: molecular mechanisms and interventional targets. *Signal Transduct Target Ther* 2024; 9: 149.
- [22] Hudson MM, Bhatia S, Casillas J and Landier W; Section on Hematology/Oncology, Children's Oncology Group, American Society of Pediatric Hematology/Oncology. Long-term follow-up care for childhood, adolescent, and young adult cancer survivors. *Pediatrics* 2021; 148: e2021053127.
- [23] Sun X, Zhang Y, Li H, Zhou Y, Shi S, Chen Z, He X, Zhang H, Li F, Yin J, Mou M, Wang Y, Qiu Y and Zhu F. DRESIS: the first comprehensive landscape of drug resistance information. *Nucleic Acids Res* 2023; 51: D1263-D1275.
- [24] Isidro-Hernández M, Alemán-Arteaga S, Casado-García A, Ruiz-Corzo B, Riesco S, Prieto-Matos P, Martínez-Cano J, Sánchez L, Cobaleda C, Sánchez-García I and Vicente-Dueñas C. Childhood B-cell preleukemia mouse modeling. *Int J Mol Sci* 2022; 23: 7562.
- [25] Filipek-Gorzał J, Kwiecińska P, Szade A and Szade K. The dark side of stemness - the role of hematopoietic stem cells in development of blood malignancies. *Front Oncol* 2024; 14: 1308709.
- [26] Inaba H and Mullighan CG. Pediatric acute lymphoblastic leukemia. *Haematologica* 2020; 105: 2524-2539.
- [27] Davidow K, Mumanachit S and Mangum DS. The two-hit hypothesis in practice: monozygotic twins with simultaneous hyperdiploid acute lymphoblastic leukemia. *Pediatr Blood Cancer* 2022; 69: e29885.
- [28] Wagener R, Elitzur S, Brozou T and Borkhardt A. Functional damaging germline variants in ETV6, IKZF1, PAX5 and RUNX1 predisposing to B-cell precursor acute lymphoblastic leukemia. *Eur J Med Genet* 2023; 66: 104725.
- [29] Bloom M, Maciaszek JL, Clark ME, Pui CH and Nichols KE. Recent advances in genetic predisposition to pediatric acute lymphoblastic leukemia. *Expert Rev Hematol* 2020; 13: 55-70.
- [30] Wagenblast E, Araújo J, Gan OI, Cutting SK, Murison A, Krivdova G, Azkanaz M, McLeod JL, Smith SA, Gratton BA, Marhon SA, Gabra M, Medeiros JF, Manteghi S, Chen J, Chan-Seng-Yue M, Garcia-Prat L, Salmena L, De Carvalho DD, Abelson S, Abdelhaleem M, Chong K, Roifman M, Shannon P, Wang JCY, Hitzler JK, Chitayat D, Dick JE and Lechman ER. Mapping the cellular origin and early evolution of leukemia in Down syndrome. *Science* 2021; 373: eabf6202.
- [31] Yurkovich JT, Tian Q, Price ND and Hood L. A systems approach to clinical oncology uses deep phenotyping to deliver personalized care. *Nat Rev Clin Oncol* 2020; 17: 183-194.
- [32] Barone C, Orsenigo R, Cazzola A, D'Errico E, Patelli A, Quattrini G, Vergani B, Bombelli S, De Marco S, D'Orlando C, Bianchi C, Leone BE, Meneveri R, Biondi A, Cazzaniga G, Rabbitts TH, Brunelli S and Azzoni E. Hematopoietic stem cell (HSC)-independent progenitors are susceptible to MLL-Af9-induced leukemic transformation. *Cancers (Basel)* 2023; 15: 3624.
- [33] Udroui I and Sgura A. Hematopoietic ontogeny and its relevance for pediatric leukemias. *Med Hypotheses* 2016; 88: 70-73.
- [34] Tarnawsky SP, Yoshimoto M, Deng L, Chan RJ and Yoder MC. Yolk sac erythromyeloid progenitors expressing gain of function PTPN11 have functional features of JMML but are not sufficient to cause disease in mice. *Dev Dyn* 2017; 246: 1001-1014.
- [35] Malouf C and Ottersbach K. The fetal liver lymphoid-primed multipotent progenitor provides the prerequisites for the initiation of

- t(4;11) *MLL-AF₄* infant leukemia. *Haematologica* 2018; 103: e571-e575.
- [36] Filbin M and Monje M. Developmental origins and emerging therapeutic opportunities for childhood cancer. *Nat Med* 2019; 25: 367-376.
- [37] Ames HM, Rooper LM, Lattera JJ, Eberhart CG and Rodriguez FJ. INSM1 expression is frequent in primary central nervous system neoplasms but not in the adult brain parenchyma. *J Neuropathol Exp Neurol* 2018; 77: 374-382.
- [38] de Kock L, Sabbaghian N, Druker H, Weber E, Hamel N, Miller S, Choong CS, Gottardo NG, Kees UR, Rednam SP, van Hest LP, Jongmans MC, Jhangiani S, Lupski JR, Zacharin M, Bouron-Dal Soglio D, Huang A, Priest JR, Perry A, Mueller S, Albrecht S, Malkin D, Grundy RG and Foulkes WD. Germ-line and somatic DICER1 mutations in pineoblastoma. *Acta Neuropathol* 2014; 128: 583-595.
- [39] Zhang J, Walsh MF, Wu G, Edmonson MN, Gruber TA, Easton J, Hedges D, Ma X, Zhou X, Yergeau DA, Wilkinson MR, Vadodaria B, Chen X, McGee RB, Hines-Dowell S, Nuccio R, Quinn E, Shurtleff SA, Rusch M, Patel A, Becksfort JB, Wang S, Weaver MS, Ding L, Mardis ER, Wilson RK, Gajjar A, Ellison DW, Pappo AS, Pui CH, Nichols KE and Downing JR. Germline mutations in predisposition genes in pediatric cancer. *N Engl J Med* 2015; 373: 2336-2346.
- [40] Guerreiro Stucklin AS, Ramaswamy V, Daniels C and Taylor MD. Review of molecular classification and treatment implications of pediatric brain tumors. *Curr Opin Pediatr* 2018; 30: 3-9.
- [41] Kazarian E, Marks A, Cui J, Darbinyan A, Tong E, Mueller S, Cha S and Aboian MS. Topographic correlates of driver mutations and endogenous gene expression in pediatric diffuse midline gliomas and hemispheric high-grade gliomas. *Sci Rep* 2021; 11: 14377.
- [42] Pathania M, De Jay N, Maestro N, Harutyunyan AS, Nitarska J, Pahlavan P, Henderson S, Mikael LG, Richard-Londt A, Zhang Y, Costa JR, Hébert S, Khazaei S, Ibrahim NS, Herrero J, Riccio A, Albrecht S, Ketteler R, Brandner S, Kleinman CL, Jabado N and Salomoni P. H3.3K27M cooperates with Trp53 loss and PDGFRA gain in mouse embryonic neural progenitor cells to induce invasive high-grade gliomas. *Cancer Cell* 2017; 32: 684-700, e9.
- [43] Liu C, Sage JC, Miller MR, Verhaak RG, Hippenmeyer S, Vogel H, Foreman O, Bronson RT, Nishiyama A, Luo L and Zong H. Mosaic analysis with double markers reveals tumor cell of origin in glioma. *Cell* 2011; 146: 209-221.
- [44] Alcantara Llaguno SR, Wang Z, Sun D, Chen J, Xu J, Kim E, Hatanpaa KJ, Raisanen JM, Burns DK, Johnson JE and Parada LF. Adult lineage-restricted CNS progenitors specify distinct glioblastoma subtypes. *Cancer Cell* 2015; 28: 429-440.
- [45] Johnsen JI, Dyberg C and Wickström M. Neuroblastoma-a neural crest derived embryonal malignancy. *Front Mol Neurosci* 2019; 12: 9.
- [46] Singh HP, Wang S, Stachelek K, Lee S, Reid MW, Thornton ME, Craft CM, Grubbs BH and Cobrinik D. Developmental stage-specific proliferation and retinoblastoma genesis in RB-deficient human but not mouse cone precursors. *Proc Natl Acad Sci U S A* 2018; 115: E9391-E9400.
- [47] Rozanska A, Cerna-Chavez R, Queen R, Collin J, Zerti D, Dorgau B, Beh CS, Davey T, Coxhead J, Hussain R, Al-Aama J, Steel DH, Benvenisty N, Armstrong L, Parulekar M and Lako M. pRB-depleted pluripotent stem cell retinal organoids recapitulate cell state transitions of retinoblastoma development and suggest an important role for pRB in retinal cell differentiation. *Stem Cells Transl Med* 2022; 11: 415-433.
- [48] Nguyen L, Papenhausen P and Shao H. The role of c-MYC in B-cell lymphomas: diagnostic and molecular aspects. *Genes (Basel)* 2017; 8: 116.
- [49] Wang-Michelitsch J and Michelitsch TM. Pediatric lymphoma may develop by “one-step” cell transformation of a lymphoid cell. *arXiv preprint arXiv:1804.11078* 2018.
- [50] L'Abbate A, Macchia G, D'Addabbo P, Lonoce A, Tolomeo D, Trombetta D, Kok K, Bartenhagen C, Whelan CW, Palumbo O, Severgnini M, Cifola I, Dugas M, Carella M, De Bellis G, Rocchi M, Carbone L and Storlazzi CT. Genomic organization and evolution of double minutes/homogeneously staining regions with MYC amplification in human cancer. *Nucleic Acids Res* 2014; 42: 9131-45.
- [51] Martin-Giacalone BA, Weinstein PA, Plon SE and Lupo PJ. Pediatric rhabdomyosarcoma: epidemiology and genetic susceptibility. *J Clin Med* 2021; 10: 2028.
- [52] Charytonowicz E, Cordon-Cardo C, Matushansky I and Ziman M. Alveolar rhabdomyosarcoma: is the cell of origin a mesenchymal stem cell? *Cancer Lett* 2009; 279: 126-136.
- [53] Blum JM, Añó L, Li Z, Van Mater D, Bennett BD, Sachdeva M, Lagutina I, Zhang M, Mito JK, Dodd LG, Cardona DM, Dodd RD, Williams N, Ma Y, Lepper C, Linardic CM, Mukherjee S, Grosveld GC, Fan CM and Kirsch DG. Distinct and overlapping sarcoma subtypes initiated from muscle stem and progenitor cells. *Cell Rep* 2013; 5: 933-940.
- [54] Penton CM, Thomas-Ahner JM, Johnson EK, McAllister C and Montanaro F. Muscle side

- population cells from dystrophic or injured muscle adopt a fibro-adipogenic fate. *PLoS One* 2013; 8: e54553.
- [55] Judson RN, Zhang RH and Rossi FM. Tissue-resident mesenchymal stem/progenitor cells in skeletal muscle: collaborators or saboteurs? *FEBS J* 2013; 280: 4100-4108.
- [56] Boscolo Sesillo F, Fox D and Sacco A. Muscle stem cells give rise to rhabdomyosarcomas in a severe mouse model of Duchenne muscular dystrophy. *Cell Rep* 2019; 26: 689-701, e6.
- [57] Hatley ME, Tang W, Garcia MR, Finkelstein D, Millay DP, Liu N, Graff J, Galindo RL and Olson EN. A mouse model of rhabdomyosarcoma originating from the adipocyte lineage. *Cancer Cell* 2012; 22: 536-546.
- [58] Drummond CJ, Hanna JA, Garcia MR, Devine DJ, Heyrana AJ, Finkelstein D, Rehg JE and Hatley ME. Hedgehog pathway drives fusion-negative rhabdomyosarcoma initiated from non-myogenic endothelial progenitors. *Cancer Cell* 2018; 33: 108-124.
- [59] Sand LG, Suzhai K and Hogendoorn PC. Sequencing overview of Ewing sarcoma: a journey across genomic, epigenomic and transcriptomic landscapes. *Int J Mol Sci* 2015; 16: 16176-16215.
- [60] Jedlicka P. Ewing sarcoma, an enigmatic malignancy of likely progenitor cell origin, driven by transcription factor oncogenic fusions. *Int J Clin Exp Pathol* 2010; 3: 338-347.
- [61] Riggì N and Stamenkovic I. The biology of Ewing sarcoma. *Cancer Lett* 2007; 254: 1-10.
- [62] von Levetzow C, Jiang X, Gwyne Y, von Levetzow G, Hung L, Cooper A, Hsu JH and Lawlor ER. Modeling initiation of Ewing sarcoma in human neural crest cells. *PLoS One* 2011; 6: e19305.
- [63] Castillero-Trejo Y, Eliazer S, Xiang L, Richardson JA and Ilaria RL Jr. Expression of the EWS/FLI-1 oncogene in murine primary bone-derived cells results in EWS/FLI-1-dependent, Ewing sarcoma-like tumors. *Cancer Res* 2005; 65: 8698-8705.
- [64] Sole A, Grossetête S, Heintzé M, Babin L, Zaïdi S, Revy P, Renouf B, De Cian A, Giovannangeli C, Pierre-Eugène C, Janoueix-Lerosey I, Couronné L, Kaltenbach S, Tomishima M, Jasin M, Grünwald TGP, Delattre O, Surdez D and Brunet E. Unraveling ewing sarcoma tumorigenesis originating from patient-derived mesenchymal stem cells. *Cancer Res* 2021; 81: 4994-5006.
- [65] Kovar H, Bierbaumer L and Radic-Sarikas B. The YAP/TAZ pathway in osteogenesis and bone sarcoma pathogenesis. *Cells* 2020; 9: 972.
- [66] Schultz KR, Carroll A, Heerema NA, Bowman WP, Aledo A, Slayton WB, Sather H, Devidas M, Zheng HW, Davies SM, Gaynon PS, Trigg M, Rutledge R, Jorstad D, Winick N, Borowitz MJ, Hunger SP, Carroll WL and Camitta B; Children's Oncology Group. Long-term follow-up of imatinib in pediatric Philadelphia chromosome-positive acute lymphoblastic leukemia: children's Oncology Group study AALL0031. *Leukemia* 2014; 28: 1467-1471.
- [67] Shima H, Tokuyama M, Tanizawa A, Tono C, Hamamoto K, Muramatsu H, Watanabe A, Hotta N, Ito M, Kurosawa H, Kato K, Tsurusawa M, Horibe K and Shimada H. Distinct impact of imatinib on growth at prepubertal and pubertal ages of children with chronic myeloid leukemia. *J Pediatr* 2011; 159: 676-681.
- [68] Wu Y, Li Y, Fan J, Qi P, Lin W, Yang J, Liu H, Wang X, Zheng H, Wang T and Zhang R. Blinatumomab for treating pediatric B-lineage acute lymphoblastic leukemia: a retrospective real-world study. *Front Pediatr* 2022; 10: 1034373.
- [69] Wilky BA, Kim C, McCarty G, Montgomery EA, Kammers K, DeVine LR, Cole RN, Raman V and Loeb DM. RNA helicase DDX3: a novel therapeutic target in Ewing sarcoma. *Oncogene* 2016; 35: 2574-2583.
- [70] Reuter D, Staeger MS, Kühnöl CD and Föll J. Immunostimulation by OX40 ligand transgenic Ewing sarcoma cells. *Front Oncol* 2015; 5: 242.
- [71] Okuda T, Shurtleff SA, Valentine MB, Raimondi SC, Head DR, Behm F, Curcio-Brint AM, Liu Q, Pui CH and Sherr CJ. Frequent deletion of p16INK4a/MTS1 and p15INK4b/MTS2 in pediatric acute lymphoblastic leukemia. *Blood* 1995; 85: 2321-2330.
- [72] Park AK, Lee SJ, Phi JH, Wang KC, Kim DG, Cho BK, Haberler C, Fattet S, Dufour C, Puget S, Sainte-Rose C, Bourdeaut F, Grill J, Delattre O, Kim SK and Park WY. Prognostic classification of pediatric medulloblastoma based on chromosome 17p loss, expression of MYCC and MYCN, and Wnt pathway activation. *Neuro Oncol* 2012; 14: 203-214.
- [73] Chung C, Sweha SR, Pratt D, Tamrazi B, Panwalkar P, Banda A, Bayliss J, Hawes D, Yang F, Lee HJ, Shan M, Cieslik M, Qin T, Werner CK, Wahl DR, Lyssiotis CA, Bian Z, Shotwell JB, Yadav VN, Koschmann C, Chinnaiyan AM, Blüml S, Judkins AR and Venneti S. Integrated metabolic and epigenomic reprogramming by H3K27M mutations in diffuse intrinsic pontine gliomas. *Cancer Cell* 2020; 38: 334-349, e9.
- [74] Condorelli F, Gnemmi I, Vallario A, Genazzani AA and Canonico PL. Inhibitors of histone deacetylase (HDAC) restore the p53 pathway in neuroblastoma cells. *Br J Pharmacol* 2008; 153: 657-668.
- [75] Ciesla M, Marona P, Kozakowska M, Jez M, Seczynska M, Loboda A, Bukowska-Strakova K, Szade A, Walawender M, Kusior M,

- Stepniwski J, Szade K, Krist B, Yagensky O, Urbanik A, Kazanowska B, Dulak J and Jozkowicz A. Heme oxygenase-1 controls an HDAC4-miR-206 pathway of oxidative stress in rhabdomyosarcoma. *Cancer Res* 2016; 76: 5707-5718.
- [76] Marques Howarth M, Simpson D, Ngok SP, Nieves B, Chen R, Siprashvili Z, Vaka D, Breese MR, Crompton BD, Alexe G, Hawkins DS, Jacobson D, Brunner AL, West R, Mora J, Stegmaier K, Khavari P and Sweet-Cordero EA. Long noncoding RNA EWSAT1-mediated gene repression facilitates Ewing sarcoma oncogenesis. *J Clin Invest* 2014; 124: 5275-5290.
- [77] Kumar V, Patel S, Tcyganov E and Gabrilovich DI. The nature of myeloid-derived suppressor cells in the tumor microenvironment. *Trends Immunol* 2016; 37: 208-220.
- [78] Xin C, Zhu J, Gu S, Yin M, Ma J, Pan C, Tang J, Zhang P, Liu Y, Bai XF, Mo X, Xu M and Zhu H. CD200 is overexpressed in neuroblastoma and regulates tumor immune microenvironment. *Cancer Immunol Immunother* 2020; 69: 2333-2343.
- [79] Ligon JA, Choi W, Cojocaru G, Fu W, Hsiue EH, Oke TF, Siegel N, Fong MH, Ladle B, Pratilas CA, Morris CD, Levin A, Rhee DS, Meyer CF, Tam AJ, Blosser R, Thompson ED, Suru A, McConkey D, Housseau F, Anders R, Pardoll DM and Llosa N. Pathways of immune exclusion in metastatic osteosarcoma are associated with inferior patient outcomes. *J Immunother Cancer* 2021; 9: e001772.
- [80] Do P, Beckwith KA, Cheney C, Tran M, Beaver L, Griffin BG, Mo X, Liu Y, Lapalombella R, Hertlein E, Muthusamy N and Byrd JC. Leukemic B cell CTLA-4 suppresses costimulation of T cells. *J Immunol* 2019; 202: 2806-2816.
- [81] Dakal TC, George N, Xu C, Suravajhala P and Kumar A. Predictive and prognostic relevance of tumor-infiltrating immune cells: tailoring personalized treatments against different cancer types. *Cancers (Basel)* 2024; 16: 1626.
- [82] Singh M, Morris VK, Bandey IN, Hong DS and Kopetz S. Advancements in combining targeted therapy and immunotherapy for colorectal cancer. *Trends Cancer* 2024; 10: 598-609.
- [83] Kratz CP, Jongmans MC, Cavé H, Wimmer K, Behjati S, Guerrini-Rousseau L, Milde T, Pajtlér KW, Golmard L, Gauthier-Villars M, Jewell R, Duncan C, Maher ER, Brugieres L, Pritchard-Jones K and Bourdeaut F. Predisposition to cancer in children and adolescents. *Lancet Child Adolesc Health* 2021; 5: 142-154.
- [84] Kamihara J, Bourdeaut F, Foulkes WD, Molenaar JJ, Mossé YP, Nakagawara A, Parareda A, Scollon SR, Schneider KW, Skalet AH, States LJ, Walsh MF, Diller LR and Brodeur GM. Retinoblastoma and neuroblastoma predisposition and surveillance. *Clin Cancer Res* 2017; 23: e98-e106.
- [85] Zelly K, Schiend J, Gallinger B, Kohlmann WK, McGee RB, Scollon SR and Schneider KW. Update on genetic counselor practice and recommendations for pediatric cancer predisposition evaluation and surveillance. *Clin Cancer Res* 2024; 30: 3983-3989.
- [86] Cazzola A, Cazzaniga G, Biondi A, Meneveri R, Brunelli S and Azzoni E. Prenatal origin of pediatric leukemia: lessons from hematopoietic development. *Front Cell Dev Biol* 2021; 8: 618164.
- [87] Li W. Pathogenesis and pathology of pediatric lymphoma. Exon Publications 2021: 1-26.
- [88] Ponzoni M, Bachetti T, Corrias MV, Brignole C, Pastorino F, Calarco E, Bensa V, Giusto E, Ceccherini I and Perri P. Recent advances in the developmental origin of neuroblastoma: an overview. *J Exp Clin Cancer Res* 2022; 41: 92.
- [89] Grünwald TGP, Cidre-Aranaz F, Surdez D, Tomazou EM, de Álava E, Kovar H, Sorensen PH, Delattre O and Dirksen U. Ewing sarcoma. *Nat Rev Dis Primers* 2018; 4: 5.
- [90] Pfaltzgraff ER, Apfelbaum A, Kassa AP, Song JY, Jiang W, Suhan TK, Wellik DM and Lawlor ER. Anatomic origin of osteochondrogenic progenitors impacts sensitivity to EWS-FLI1-induced transformation. *Cancers (Basel)* 2019; 11: 313.
- [91] Gaspar N, Marques da Costa ME, Fromigue O, Droit R, Berlanga P and Marchais A. Recent advances in understanding osteosarcoma and emerging therapies. *Fac Rev* 2020; 9: 18.
- [92] Muskens IS, Zhang C, de Smith AJ, Biegel JA, Walsh KM and Wiemels JL. Germline genetic landscape of pediatric central nervous system tumors. *Neuro Oncol* 2019; 21: 1376-1388.
- [93] Abraham J, Nuñez-Álvarez Y, Hettmer S, Carrió E, Chen HI, Nishijo K, Huang ET, Prajapati SI, Walker RL, Davis S, Rebeles J, Wiebush H, McCleish AT, Hampton ST, Bjornson CR, Brack AS, Wagers AJ, Rando TA, Capecchi MR, Marini FC, Ehler BR, Zarzabal LA, Goros MW, Michalek JE, Meltzer PS, Langenau DM, LeGallo RD, Mansoor A, Chen Y, Suelves M, Rubin BP and Keller C. Lineage of origin in rhabdomyosarcoma informs pharmacological response. *Genes Dev* 2014; 28: 1578-91.
- [94] Riggí N, Knoechel B, Gillespie SM, Rheinbay E, Boulay G, Suvà ML, Rossetti NE, Boonseng WE, Oksuz O, Cook EB, Formey A, Patel A, Gymer M, Thapar V, Deshpande V, Ting DT, Hornicek FJ, Nielsen GP, Stamenkovic I, Aryee MJ, Bernstein BE and Rivera MN. EWS-FLI1 utilizes divergent chromatin remodeling mechanisms to directly activate or repress enhancer elements in Ewing sarcoma. *Cancer Cell* 2014; 26: 668-681.

- [95] Buckingham M and Relaix F. PAX3 and PAX7 as upstream regulators of myogenesis. *Semin Cell Dev Biol* 2015; 44: 115-125.
- [96] Schmitt-Ney M and Camussi G. The PAX3-FOXO1 fusion protein present in rhabdomyosarcoma interferes with normal FOXO activity and the TGF- β pathway. *PLoS One* 2015; 10: e0121474.
- [97] Dzieran J, Rodriguez Garcia A, Westermarck UK, Henley AB, Eyre Sánchez E, Träger C, Johansson HJ, Lehtiö J and Arsenian-Henriksson M. *MYCN*-amplified neuroblastoma maintains an aggressive and undifferentiated phenotype by deregulation of estrogen and NGF signaling. *Proc Natl Acad Sci U S A* 2018; 115: E1229-E1238.
- [98] Hurley SP, Clary DO, Copie V and Lefcort F. Anaplastic lymphoma kinase is dynamically expressed on subsets of motor neurons and in the peripheral nervous system. *J Comp Neurol* 2006; 495: 202-212.
- [99] Fassl A, Geng Y and Sicinski P. CDK4 and CDK6 kinases: from basic science to cancer therapy. *Science* 2022; 375: eabc1495.
- [100] Kim KH and Roberts CW. Mechanisms by which SMARCB1 loss drives rhabdoid tumor growth. *Cancer Genet* 2014; 207: 365-372.
- [101] Alimova I, Venkataraman S, Harris P, Marquez VE, Northcott PA, Dubuc A, Taylor MD, Foreman NK and Vibhakkar R. Targeting the enhancer of zeste homologue 2 in medulloblastoma. *Int J Cancer* 2012; 131: 1800-9.
- [102] Mullighan CG, Zhang J, Kasper LH, Lerach S, Payne-Turner D, Phillips LA, Heatley SL, Holmfeldt L, Collins-Underwood JR, Ma J, Buetow KH, Pui CH, Baker SD, Brindle PK and Downing JR. CREBBP mutations in relapsed acute lymphoblastic leukaemia. *Nature* 2011; 471: 235-9.
- [103] Wulf AM, Moreno MM, Paka C, Rampasekova A and Liu KJ. Defining pathological activities of ALK in neuroblastoma, a neural crest-derived cancer. *Int J Mol Sci* 2021; 22: 11718.
- [104] Flotho C, Kratz C and Niemeyer CM. Targeting RAS signaling pathways in juvenile myelomonocytic leukemia. *Curr Drug Targets* 2007; 8: 715-725.
- [105] Niemeyer CM. RAS diseases in children. *Haematologica* 2014; 99: 1653-1662.
- [106] Vainchenker W and Constantinescu SN. JAK/STAT signaling in hematological malignancies. *Oncogene* 2013; 32: 2601-2613.
- [107] Merino D and Malkin D. p53 and hereditary cancer. In: *Mutant p53 and MDM2 in Cancer*. Subcell Biochem 2014; 85: 1-16.
- [108] Mendoza PR and Grossniklaus HE. The biology of retinoblastoma. *Prog Mol Biol Transl Sci* 2015; 134: 503-516.
- [109] Wang Y, Chen J, Yang W, Mo F, Senz J, Yap D, Anglesio MS, Gilks B, Morin GB and Huntsman DG. The oncogenic roles of DICER1 RNase IIIb domain mutations in ovarian Sertoli-Leydig cell tumors. *Neoplasia* 2015; 17: 650-660.
- [110] Dodgshun AJ, Sexton-Oates A, Saffery R and Sullivan MJ. Biallelic FANCD1/BRCA2 mutations predisposing to glioblastoma multiforme with multiple oncogenic amplifications. *Cancer Genet* 2016; 209: 53-56.