

Original Article

Low expression of the long non-coding RNA NR_026827 in gastric cancer

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Abstract: Aim: The contribution of long non-coding RNAs (lncRNAs) to gastric cancer associated with *Helicobacter pylori* (*H. pylori*) infection remains largely unknown. Therefore, the present study aimed to investigate the expression of a lncRNA NR_026827 in gastric epithelial cells infected with *H. pylori*, and demonstrate its expression characteristic in gastric cancer. Materials and Methods: Gastric epithelial cell line cells, GES-1, were cultured and infected with *H. pylori*. A microarray was used to analyze the lncRNA profile of gastric epithelial cells. Eighty fresh gastric cancer tissues and the paired adjacent non-cancerous tissue samples were randomly selected from patients. The expression of the lncRNA NR_026827 was investigated using quantitative real-time polymerase chain reaction (qRT-PCR). Results: The expression of several lncRNAs was significantly altered in GES-1 cells following infection with *H. pylori*. Of these lncRNAs, NR_026827 was dramatically down-regulated in GES-1 cells infected with *H. pylori*. In addition, the expression of NR_026827 was decreased in gastric cancer tissues in comparison to the corresponding adjacent non-cancerous tissues. Moreover, the expression of NR_026827 did not change significantly in different gastric cancer stages. Conclusion: The lncRNA, NR_026827, is down-regulated in all stages of gastric cancer associated with *H. pylori* infection and could represent a potential biomarker for the diagnosis of gastric cancer.

Keywords: Long non-coding RNA, NR_026827, gastric cancer, *Helicobacter pylori*

Introduction

Globally, gastric cancer is the third leading cause of cancer death, and the fifth most common cancer, with an incidence of more than one million people experiencing it every year [1]. In China, gastric cancer is the most common type of cancer, and the second leading cause of cancer death [2]. It is well known that the chronic gastritis caused by persistent infection of the stomach with *Helicobacter pylori* (*H. pylori*) eventually causes gastric cancer [3]. *H. pylori* was discovered as a pathogen in 1983 and was certified as a definite carcinogen by the World Health Organization [4]. *H. pylori* infection represents a strong risk factor for gastric cancer [5].

Long non-coding RNAs (lncRNAs) are RNA species that are over 200 nucleotides in length and cannot be translated into protein [6].

Large-scale analyses of full-length cDNA sequences have detected large numbers of lncRNAs in humans [7]. These lncRNAs have been shown to play key roles in cell differentiation, proliferation, apoptosis, immune responses, tumorigenesis, and other biological processes [8-12].

In recent years, many lncRNAs have been reported to be associated with cancer. For example, the lncRNA GAPLINC regulates the invasion of gastric cancer cells by controlling the expression of CD44 [13]. Recently, Yang *et al.* described changes in the expression of various lncRNAs in *H. pylori* infected cells, and found that the expression of 23 lncRNAs was upregulated and the expression of 21 was downregulated in these cells [14]. However, the contribution of lncRNAs to gastric cancer associated with *H. pylori* infection still remains largely unknown. In this study, we investigated the lncRNA profile of gastric epithelial cells infected

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Table 1. The clinical characteristics of all patients

Clinicopathological features	Numbers
Age (years)	
< 60	28
≥ 60	52
Gender	
Male	45
Female	35
Differentiation	
Well	43
Moderate + Poor	37
Depth of invasion	
T1	8
T2	54
T3	13
T4	5
Lymph node metastasis	
N0	19
N1	22
N2	32
N3	7
Distant metastasis	
M0	68
M1	12
TNM stage	
I	23
II	32
III	13
IV	12

Table 2. Long non-coding RNAs up-regulated in the gastric epithelial cells infected by *H. pylori* compared to that of non-infection control by microarray-based profile assay

Long non-coding RNA	RNA length	Log2 Fold (infection/non-infection)
chr6:63131625-63144250+	12625	3.58
TCONS_00007049	1434	2.99
ENST00000581964	517	2.62
NR_036510	1458	2.01
TCONS_00003565	344	1.95
ENST00000538111	368	1.85
ENST00000571665	255	1.84
ENST00000452342	573	1.76
TCONS_00015979	810	1.75
ENST00000564300	600	1.71
ENST00000414795	506	1.69
NR_047576	2000	1.68

with *H. pylori* by microarray assay and found that the expression of several lncRNAs was significantly altered when compared with control cells. Of these lncRNAs, the expression of NR_026827 was first observed to be down-regulated 16-fold. Furthermore, we verified the low expression of NR_026827 in gastric epithelial cells infected with *H. pylori* by analyzing the expression of NR_026827 in gastric cancer tissues and the corresponding adjacent non-cancerous tissues using quantitative real-time polymerase chain reaction (qRT-PCR).

Materials and methods

Cell culture, infection, and microarray analysis

Cells from the gastric epithelial cell line GES-1 were maintained in RPMI 1640 supplemented with 10% fetal bovine serum at 37°C in a humidified air atmosphere containing 5% CO₂. To infect the GES-1 cells using *H. pylori* NCTC11637, cells were cultured at 5 × 10⁶ cells per flask. Approximately 5 × 10⁸ colony-forming units (CFU) of logarithmic-phase (OD₆₀₀ 0.5 to 0.6) anaerobically grown bacteria were pelleted, washed twice with phosphate buffered saline (PBS), resuspended in 1 ml of RPMI 1640, and then added to the cell monolayer at a multiplicity of infection of 100:1. After incubation for 20 min at 37°C, the infected cells were washed three times with pre-warmed PBS (pH 7.4), and the infected monolayers were lysed (0 h) from the tissue-culture dishes containing 100 µg/ml gentamicin to kill extracellular bacteria. The infected monolayers were then washed three times with pre-warmed PBS and further incubated for an additional 12 h in the presence of freshly supplemented tissue-culture medium containing 12 µg/ml gentamicin. The infected cells were then washed three times with pre-warmed PBS, the monolayers were lysed, and total RNA was extracted from the cells using TRIzol reagent (Invitrogen Life Technologies). Subsequently, total RNA was analyzed by microarray to investigate the lncRNA profile of GES-1 cells infected with *H. pylori*.

Patients and clinical samples

Eighty fresh gastric cancer tissues and their paired adjacent non-cancerous tissue samples were randomly selected from patients who had

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Table 3. Long non-coding RNAs down-regulated in the gastric epithelial cells infected by *H. pylori* compared to that of non-infection control by microarray-based profile assay

Long non-coding RNA	RNA length	Log2 Fold (infection/non-infection)
NR_026827	2322	-12.24
NR_028301	3278	-9.15
ENST00000523627	570	-8.63
ENST00000562166	1838	-6.77
AK022063	2196	-6.29
TCONS_00029621	276	-5.21
ENST00000453706	261	-4.82
TCONS_00005953	761	-4.62
ENST00000534653	529	-4.23
TCONS_00004713	206	-4.08
TCONS_00026381	324	-4.05
ENST00000553668	736	-4.01
ENST00000492461	415	-3.79
ENST00000444919	414	-3.37
uc010lfo.1	1470	-3.34
TCONS_00016207	728	-3.23
TCONS_00008376	784	-2.97
TCONS_00012531	222	-2.87
ENST00000528514	1361	-2.84
ENST00000542311	2041	-2.81
ENST00000455391	461	-2.69
TCONS_00026952	1692	-2.62
ENST00000554036	509	-2.59
ENST00000538025	534	-2.52
ENST00000512831	628	-2.45
ENST00000420187	549	-2.39
ENST00000422449	363	-2.36
ENST00000448671	574	-2.35
TCONS_00021051	459	-2.25
TCONS_00020407	1957	-2.19
ENST00000414135	976	-2.06
ENST00000503299	758	-1.97
uc002suu.2	3042	-1.96
ENST00000484222	2084	-1.95
ENST00000423186	1055	-1.94
ENST00000508986	331	-1.91
ENST00000532652	514	-1.85
NR_045027	1158	-1.84
uc002zis.1	570	-1.81
ENST00000531870	568	-1.69
ENST00000442305	514	-1.68

positive ¹³C-urea breathe tests and were undergoing surgery at the Second Affiliated Hospital

of Soochow University between January 1 and April 30, 2015. All experiments were performed according to the principles of the Declaration of Helsinki. The Ethics Committee of the Second Affiliated Hospital of Soochow University approved this study. The tissues were used to detect the expression of lncRNA NR_026827 using qRT-PCR. Fresh tissues were collected in the operating theatre and processed immediately, within 15 min. Each sample was frozen and stored at -80°C. The paired non-cancerous tissues were isolated from at least 2 cm away from the tumor border and were shown to lack tumor cells by histopathological evaluation. All patients in this study met the following inclusion criteria: the tissues were identified as gastric cancer tissue by pathological examination, and no anti-cancer treatments were administered before surgery. The clinical characteristics of all patients are listed in **Table 1**. The tumor staging was defined according to the 7th edition of the Tumor Node Metastasis (TNM) classification system published by the International Union Against Cancer. All patients provided written informed consent to participate in the study. The data do not contain any information that could identify the patients.

RNA extraction and cDNA synthesis

Total RNA was extracted from tissues and cells using TRIzol reagent according to the manufacturer's instructions. The quality of the total RNA was assessed using a NanoDrop[®]ND-1000 and denaturation agarose-gel electrophoresis. First-strand cDNA was synthesized from 1.5 µg of total RNA using specific primers for lncRNA NR_026827 and the MMLV reverse transcriptase (Epicentre).

qRT-PCR investigation

qRT-PCR analyses were performed with 2 × PCR master mixture (Arraystar) using a ViiA 7 Real-time PCR System (Applied Biosystems). Glyceraldehyde 3-phosphate dehydrogenase (GAPDH) was used as an internal control. The PCR primers for lncRNA NR_026827 and GAPDH were as follows: lncRNA NR_026827 sense, ACTGCCCATACGGACCTAC and reverse, TCCCAAGAGACAATGAAAAAG; GAPDH sense, GGGAACTGTGGCGTGAT and reverse, GAG-TGGGTGTCGCTGTTGA. The relative mRNA levels were calculated based on the Ct values and normalized to GAPDH levels. The results are

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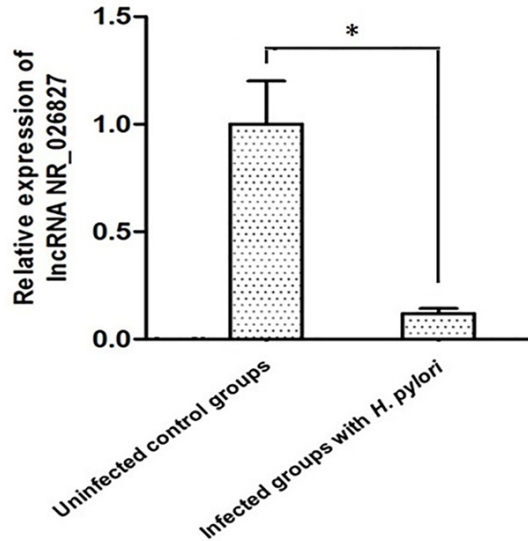


Figure 1. Relative expression of lncRNA NR_026827 in the gastric epithelial cells infected with *H. pylori* by qRT-PCR. *P < 0.01.

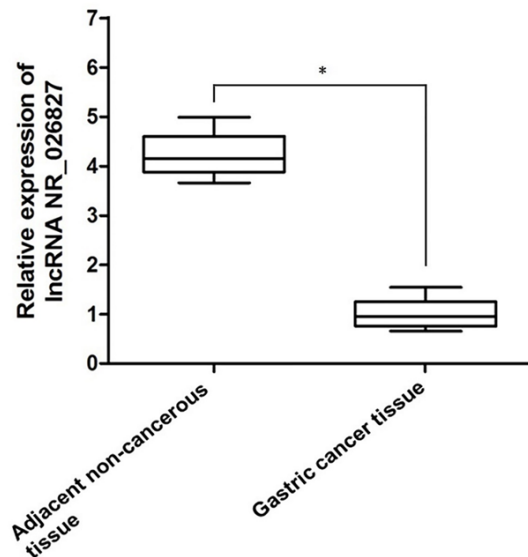


Figure 2. Relative expression of lncRNA NR_026827 in the gastric cancer tissues and the corresponding adjacent non-cancerous tissues by qRT-PCR. *P < 0.05.

expressed as means \pm standard deviation (SD) from three separate qRT-PCR tests performed on each of 80 gastric cancer specimens. The differences between the groups were analyzed using a two-tailed Student's t-test. Statistical analysis was performed using SPSS software and differences with *P*-values < 0.05 were considered significant.

Results

The lncRNA profile of GES-1 cells infected with H. pylori

The lncRNA profile of GES-1 cells infected by *H. pylori* differs to that of non-infection controls. The expression of several lncRNAs was significantly altered. lncRNAs that showed up-regulation and down-regulation are shown in **Tables 2** and **3**, respectively.

Low expression of lncRNA NR_026827 in GES-1 cells infected with H. pylori

To investigate the expression of lncRNA NR_026827 in the GES-1 cells infected with *H. pylori*, we compared the expression levels in the experimental group infected with *H. pylori* for 24 h with the levels in the non-infected control group using qRT-PCR. As shown in **Figure 1**, the expression of NR_026827 was significantly decreased in the GES-1 cells infected with *H. pylori* (*P* < 0.01).

Investigation of lncRNA NR_026827 expression in gastric cancer

We first examined the expression of lncRNA NR_026827 in 80 paired gastric cancer samples and their adjacent non-tumorous tissues using qRT-PCR. The expression of lncRNA NR_026827 was found to be significantly decreased in gastric cancer tissues compared with the corresponding adjacent non-tumorous tissues (*P* < 0.05; **Figure 2**).

Expression of lncRNA NR_026827 in different stages of gastric cancer

To study the expression characteristics of lncRNA NR_026827 in different stages of gastric cancer, we analyzed the data of 80 gastric cancer samples. Results showed that there were no obvious differences in the expression of lncRNA NR_026827 in gastric cancer at different stages (*P* > 0.05; **Figure 3**). This suggests that lncRNA NR_026827 is expressed stably in gastric cancer tissues at all stages.

Discussion

Gastric cancer is one of the most common malignancies worldwide, and ranks second in cancer-associated deaths because of its high

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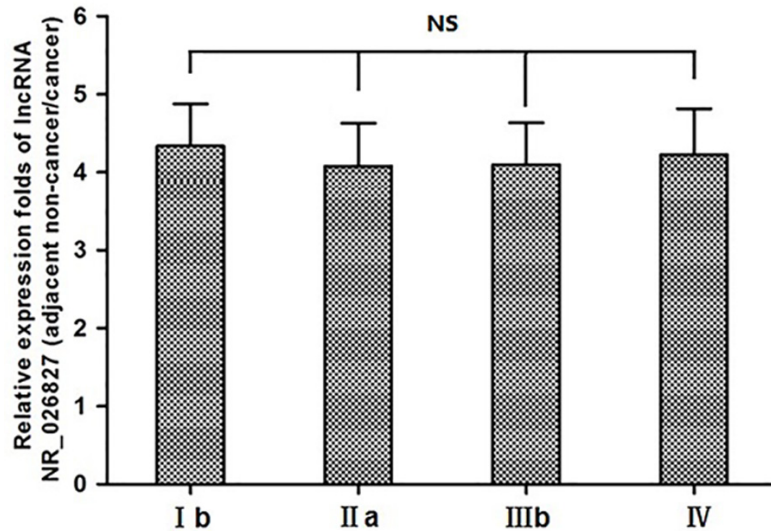


Figure 3. Relative expression folds of lncRNA NR_026827 (adjacent non-cancerous tissues/cancerous tissues). NS: $P > 0.05$.

lethality [1]. *H. pylori* is a gram-negative, spiral-shaped pathogenic bacterium that causes chronic gastritis, peptic ulcer disease, mucosa-associated lymphoid tissue lymphoma, and gastric cancer [15-17]. lncRNA, which is greater than 200 nucleotides in length, is a class of RNA species that does not have a protein coding function and is involved in different regulatory processes, such as the regulation of gene expression at the chromatin, transcriptional, or post-transcriptional levels [6]. In recent years, many studies have shown that lncRNAs are associated with cancers, and have been shown to be involved in both the occurrence and development of various cancers [18, 19]. However, the diagnostic significance of lncRNAs in gastric cancer is largely unknown [20]. Recently, Zhou *et al.* reported that the lncRNA LET was down-regulated in gastric cancer and was associated with poor prognosis [21]. It was also reported that the expression of the lncRNA AI364715 was down-regulated in gastric cancer tissues, and that its down-regulation was closely associated with tumor size and differentiation [22]. Therefore, new markers with high sensitivity and specificity for gastric cancer detection are urgently needed.

In this study, we found that the expression of a novel lncRNA, NR_026827, was down-regulated in gastric epithelial (GES-1) cells infected with *H. pylori*. Moreover, the expression of NR_026827 in gastric cancer tissues was also decreased in comparison to the corresponding

adjacent non-cancerous tissues. In addition, the expression of lncRNA NR_026827 did not differ in different stages of gastric cancer, suggesting that lncRNA NR_026827 may be involved in all stages of gastric cancer. We also analyzed the relationship between lncRNA NR_026827 expression and the age and sex of patients and our results show that there were no obvious relationships (data not shown). lncRNA NR_026827 is a long intergenic non-coding RNA that is 2322 nucleotides in length, and its gene is located on chromosome 10. The function of this novel

lncRNA was previously unclear. The results of this study may provide a novel perspective for better understanding the molecular basis of gastric cancer, and suggest that lncRNA NR_026827 might be a potential biomarker for the diagnosis of gastric cancer. However, the biological function of NR_026827 in the pathogenesis of gastric cancer should be elucidated in future studies.

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Disclosure of conflict of interest

None.

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