Original Article miR-4513 promotes breast cancer progression through targeting TRIM3

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Abstract: The biological function of microRNA-4513 (miR-4513) in human cancers is emerging. However, it remains unknown whether miR-4513 has a role in breast cancer (BC). In this study, we analyzed the expression of miR-4513 and tripartite motif containing 3 (TRIM3) in BC cell lines. The biological roles of miR-4513 and TRIM3 in BC were analyzed by cell counting kit-8 assay, colony formation assay, wound healing assay, and transwell invasion assay. The effects of miR-4513 or TRIM3 expression on the overall survival of BC patients were analyzed at Kaplan-Meier plotter website. We found miR-4513 or low TRIM3 expression was upregulated, whereas TRIM3 expression was downregulated in BC cell lines. Importantly, high miR-4513 or low TRIM3 expression was revealed as predictors for poorer overall survival of BC patients. Luciferase activity assay and western blot assay revealed TRIM3 was a direct target of miR-4513. Moreover, we showed miR-4513 was able to regulate BC cell proliferation, colony formation, cell migration, and cell invasion through regulating TRIM3. Taken together, our results suggested miR-4513 functions as an oncogene in the progression of BC and, therefore, miR-4513 may be validated as a potential therapeutic target in the future.

Keywords: miR-4513, TRIM3, breast cancer, oncogene, cell events

Introduction

Breast cancer (BC) remains a huge threat to women's health to date [1]. Worldwide, it was estimated there will be 2.1 million female BC patients been diagnosed at 2018 [1]. Even though significantly progresses on the treatment measures of BC have been achieved, however, the treatment of BC remains a challenge because of the lack of effective therapeutic targets [2, 3]. Therefore, it is essential to investigate the potential mechanisms underlying the progression of BC to identify novel diagnostic and therapeutic targets for BC.

microRNAs (miRNAs) are non-coding RNAs with 18-24 nucleotides in length [4]. miRNAs can bind to the 3'-untranslated region (3'-UTR) of targeted genes and hence resulted in message RNA degradation or protein translation inhibition [4]. Accumulating evidence has suggested that miRNAs function as crucial roles in human cancers [5]. Moreover, the importance of miR-NAs in regulating human cancer progression has been appreciated [6]. It was found miRNAs could regulate cancer cell proliferation, cell fate. And tumorigenesis [6]. Multiple miRNAs, including miR-27a and miR-181 have been reported to be upregulated in BC and thus play an oncogenic role, while the miRNAs including miR-196b-5p and miR-22 were found downregulated and function as tumor suppressive miR-NAs [7-10].

miR-4513, a newly identified miRNA, whose polymorphisms in the seed sequence are often associated with human diseases [11, 12]. It was found rs2168518: G>A, a genetic variant in the seed region of miR-4513, was found to have a dominant role on lipid and glucose homeostasis, blood pressure, and coronary artery disease [11]. Furthermore, the rs-2168518: G>A genetic variant was also found to be associated with age-related macular degeneration [12]. Very recently, miR-4513 was found to have a role in cancer as its expression was closely associated with the prognosis of lung adenocarcinoma after treatment with tyrosine kinase inhibitors targeting epidermal growth factor receptors [13]. However, little is known regarding the expression and molecular mechanism of miR-4513 in BC.

In this study, we investigated the expression patterns of miR-4513 in human BC cell lines, followed by functional analyses in BC cell lines. Our data revealed that miR-4513 was upregulated in BC cell lines. High expression of miR-4513 predicts poor overall survival of BC patients. Moreover, our results clearly suggested that miR-4513 promoted cell growth, colony formation, migration, and invasion in BC cell lines via down-regulating tripartite motif containing 3 (TRIM3) expression, further supporting miR-4513 as a potential therapeutic target in BC.

Materials and methods

Cell lines

Human BC cell lines (MCF-7, T-47D), and the breast epithelial cell line (MCF-10A) were purchased from the Type Culture Collection of the Chinese Academy of Sciences (Shanghai, China).

Cell culture and cell transfection

The above-mentioned cell lines were cultured in Dulbecco's modified Eagle's medium (DMEM, Invitrogen, Carlsbad, CA, USA) supplemented with 10% fetal bovine serum (FBS, Invitrogen) at a 37°C humidified incubator containing 5% of CO_2 .

For cell transfection, cells were seeded at the density of 6×10^4 cells/well and incubated for 24 h. Then, cells were transfected with miR-4513 inhibitor (5'-AUGGGCCUCCAGCCGUCAG-UCU-3'), inhibitor negative control (NC-inhibitor, 5'-GCUCCGGCUUGCGGUAUCACAC-3'), small-interfering RNA targeting TRIM3 (si-TRIM3, 5'-GCUCACUGUCACUACCAAATT-3'), or NC-siRNA (5'-UUCUCCGAACGUGUCACGUTT-3', all purchased from GenePharma, Shanghai, China) at the final concentration of 100 nmol/l using Lipofectamine 2000 (Invitrogen) according to the supplier's instruction. After transfection for 48 h, cells were collected for following analyses.

RNA isolation and real-time quantitative PCR (qRT-PCR)

Total RNA was isolated using TRIzol reagent (Invitrogen) according to the manufacturers'

protocols. The PrimeScript[™] RT-PCR kit (Takara, Dalian, Liaoning, China) was used to synthesized the first-strand complementary DNA (cDNA). SYBR Green Mater Mix Kit (Applied Biosystems, Foster City, CA, USA) was used for aRT-PCR at the Applied Biosystems 7300 system. U6 small nuclear RNA (U6 snRNA) was used as internal control and the relative miR-4513 expression levels were analyzed using the $2^{-\Delta\Delta Ct}$ method. The primers were as follows: miR-4513: forward, 5'-ACACTCCAGCTGGGA-GACTGACGGCTGGAG-3', reverse, 5'-CTCAACTG-GTGTCGTGGAGTCGGCAATTCAGTTGAGATGG-GC-3'; U6 snRNA: forward, 5'-CTCGCTTCG-GCAGCACA-3, reverse, 5'-ACGCTTCACGAATTTG-CGT-3'. Experiments were conducted in triplicates.

Protein isolation and western blot

Total protein was extracted from cell lines using RIPA lysis buffer (Beyotime, Haimen, Jiangsu, China) according to the manufacturer's instructions. 25 µg protein samples were isolated using 10% sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) and then transferred to polyvinylidene fluoride (PV-DF) membranes (Beyotime). After blocked with 5% fat-free milk, membranes were incubated with the primary antibodies (rabbit anti-TRIM3: ab111840, rabbit anti-GAPDH: ab181602; Abcam, Cambridge, MA, USA) for over-night at 4°C, followed by HRP-conjugated secondary antibody (ab6721, Abcam) for 3 h at room temperature. Western blot bands were visualized using BevoECL kit (Bevotime) and quantified with Image J 1.42 software (NIH, Bethesda, MD, USA). Experiments were conducted in triplicates.

Dual luciferase reporter assay

The potential miR-4513 binding site in the 3'-UTR of TRIM3 was predicted by TargetScan. TRIM3 3'-UTR sequence was amplified from genome and cloned into psiCHECK-2 luciferase vector (Promega, Madison, WI, USA) to generate TRIM3-wt. Site-direct mutagenesis kit (Takara) was used to produce TRIM3-mt. For dual luciferase reporter assay, cells were co-transfected with TRIM3-wt or TRIM3-mt and miR-4513 inhibitor or NC-inhibitor using Lipofectamine 2000 according to the manufacturer's protocols. After 48 h of transfection, Firefly and Renilla luciferase activities were detected using Dual Luciferase Reporter Assay System (Promega).

Cell counting kit-8 (CCK-8) assay

Cells were seeded at the density of 5×10^3 cells/well and allow to growth for 24 h. Then, CCK-8 solution (Beyotime) was added to the medium at indicated time (0, 24, 48 and 72 h) and further incubation for 2 h. Optical density at 450 nm was measured by a microplate reader (Bio-Rad, Hercules, CA, USA). Experiments were conducted in triplicates.

Colony formation assay

Cells were seeded at the density of 3×10^3 cells/well) and cultured for 14 days at the above-mentioned conditions. Subsequently, cells were fixed with methanol and stained with 0.1% crystal violet (Beyotime). Colony numbers were counted under a light microscope (Eclipse TS100; Nikon, Tokyo, Japan). Experiments were conducted in triplicates.

Wound healing assay

Cells were seeded at density of 6×10^4 cells/ well and incubated to 75% confluence. Wounds were created with 10 µl pipette tips at the cell surface. Then, cells were washed with PBS to remove debris and incubated at the abovementioned conditions. Cell movements in the scratch were observed and photographed at 0 and 48 h. The width of the scratch was detected with Image J 1.42 software (NIH). Experiments were conducted in triplicates.

Transwell invasion assay

Cell invasion ability was analyzed by transwell invasion assay. Cells were seeded into the upper chamber and supplemented with DMEM. The lower chamber was supplemented with DMEM supplemented with FBS. The membrane was pre-coated with Matrigel (Franklin Lakes, NJ, USA). The cells were incubated for 48 h and the invasion cells were fixed with 4% paraformaldehyde and stained with 0.1% crystal violet solution (Beyotime). Invasion numbers were counted under microscope. Experiments were conducted in triplicates.

Kaplan-Meier survival analysis

Kaplan-Meier plotter (www.kmplot.com) was used to assess the effect of miR-4513 or TRIM3 expression on the overall survival of BC. Cut-off value was auto-selected in the algorithm. Log-rank test was used to calculate the difference between low or high expression groups.

Statistical analysis

Data were presented as mean \pm standard deviation (SD) and analyzed using SPSS 21.0 (IBM, Armonk, NY, USA). The differences were assessed using a two-tailed Student's t-test or oneway ANOVA and Tukey post-hoc test for two or above groups, respectively. P<0.05 was considered to indicate a statistically significant.

Results

Upregulate expression of miR-4513 in BC

The expression of miR-4513 in BC cell lines was analyzed using qRT-PCR. As shown in **Figure 1A**, miR-4513 expression was significantly increased in BC cell lines (MCF-7 and T-47D) investigated compared with breast epithelial cell line (MCF-10A). Moreover, the analysis of overall survival data obtained from 1061 BC patients in the KM plotter website showed high miR-4513 predicts worse overall survival of BC patients (**Figure 1B**).

Downregulate expression of TRIM3 in BC

The expression of TRIM3 in BC cell lines was analyzed using western blot. As displayed in **Figure 2A**, TRIM3 expression was dramatically decreased in MCF-7 and T-47D cell lines compared to MCF-10A cell line. The overall survival analysis on 3951 BC patients showed that low TRIM3 was correlated with poor overall survival od BC patients (**Figure 2B**).

TRIM3 was a direct target of miR-4513

Considering the importance of miR-4513 and TRIM3, we are interested to investigate the relationship of them. The TargetScan prediction algorithm showed that TRIM3 contains a binding site for miR-4513 in its 3'-UTR (**Figure 3A**). Luciferase activity reporter assay showed that miR-4513 inhibitor transfection increased the luciferase activity in cells transfected with TRIM3-wt, while it did not have significance influence on the luciferase activity of cells transfected with TRIM3-mt (**Figure 3B**).

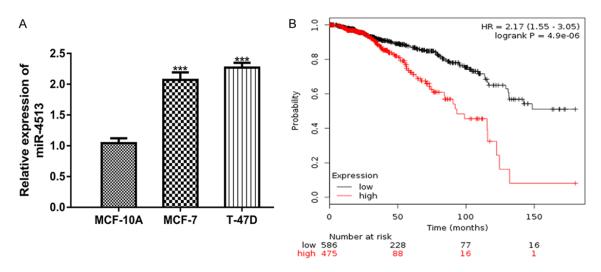
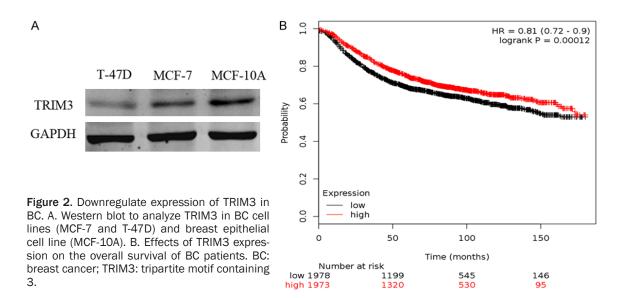


Figure 1. Overexpression of miR-4513 in BC. A. qRT-PCR to analyze miR-4513 in BC cell lines (MCF-7 and T-47D) and breast epithelial cell line (MCF-10A). B. Effects of miR-4513 expression on the overall survival of BC patients. (***P<0.001) qRT-PCR: real-time quantitative PCR; BC: breast cancer; miR-4513: microRNA-4513.



miR-4513 regulates BC cell proliferation, colony formation, cell migration, and cell invasion through targeting TRIM3

The above results have illustrated the importance of miR-4513 and TRIM3 in BC, we then interested to investigate the biological roles of miR-4513 and TRIM3. The transfection of miR-4513 inhibitor significantly decreased the expression of miR-4513 (**Figure 4A**). In the meantime, the transfection of si-TRIM3 significantly decreased the expression of TRIM3 (**Figure 4B**). Meanwhile, the stimulation effects of miR-4513 inhibitor on TRIM3 expression can be reversed by si-TRIM3 (**Figure 4B**). CCK-8 assay showed that cell proliferation was inhib-

ited by miR-4513 inhibitor but promoted by si-TRIM3 (Figure 4C). Colony formation assay confirmed the results of CCK-8 assay, which showed colony numbers were increased in si-TRIM3 group but decreased in miR-4513 group (Figure 4D). Wound-healing assay revealed that cell migration can be repressed by miR-4513 inhibitor but enhanced by si-TRIM3 (Figure 4E). Transwell invasion assay showed that cell invasion was inhibited by miR-4513 inhibitor but promoted by si-TRIM3 (Figure 4F). Furthermore, si-TRIM3 transfection could partially reversed the inhibitory effects of miR-4513 inhibitor on BC cell proliferation, colony formation, cell migration, and cell invasion in vitro (Figure 4C-F).

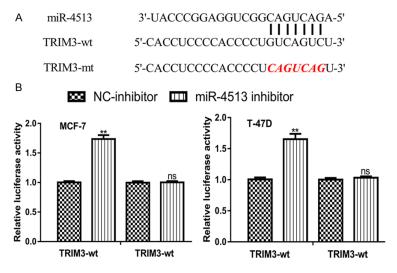


Figure 3. TRIM3 was a direct target of miR-4513. A. Binding site between miR-4513 and the 3'-UTR of TRIM3. B. Luciferase activity reporter assay to measure luciferase activity in cells transfected with TRIM3-wt or TRIM3-mt and miR-4513 inhibitor or NC-inhibitor. (ns not significant, **P<0.01) BC: breast cancer; TRIM3: tripartite motif containing 3; miR-4513: microR-NA-4513; UTR: untranslated region; wt: wild-type: mt: mutant; NC-inhibitor: negative control for miR-4513 inhibitor.

Discussion

TRIM3 located at chromosome 11p15.5, a region that has been reported to contain multiple cancer-related genes [14, 15]. Current studies have demonstrated that TRIM3 expression was reduced in several human cancers including liver cancer, gastric cancer, cervical cancer, and colorectal cancer [16-19]. TRIM3 was also revealed to regulate cell behaviors including cell proliferation, metastasis, cell migration, and cell invasion [16-19]. Upregulation of TRIM3 expression inhibited these cell malignancy behaviors, while the downregulation of TRIM3 promoted cell malignancy [16-19]. Importantly, TRIM3 was reported to function as tumor suppressor through regulating p21 (WAF1/CIP1) to prevent the accumulation of cyclin D1-cdk4 [20].

In this study, we found TRIM3 expression was significantly reduced in BC cell lines compared with the normal cell line. Then, we are interested to discovery the clinical significance of TRIM3 in BC and therefore we analyzed the overall survival data from 3951 BC patients. Notably, we found low TRIM3 expression was closely associated with poor overall survival of BC patients. These results illustrated that TRIM3 also functions as tumor suppressor in

BC, which is in consistent with the role of TRIM3 in other human cancers [16-19].

As a type of potential regulator, miRNAs serve a key role in the progression of BC [7-10]. The role of miR-4513 in human cancers is rarely investigated except a recent study showed the importance of miR-4513 in lung adenocarcinoma [13]. By measuring miR-4513 expression in BC cell lines, we found its expression was remarkedly enhanced in BC cell lines. Kaplan-Meier curve showed that high miR-4513 expression was correlated poorer overall survival of BC patients. These results showed miR-4513 might has an oncogenic role in the progression of BC. Then, functional

assays were conducted to investigate the role of miR-4513 in BC. It was found downregulation of miR-4513 inhibits BC cell proliferation, colony formation, cell migration, and cell invasion.

Then, software algorithms were performed to search for the possible connection between miR-4513 and TRIM3. Importantly, TargetScan analysis showed that miR-4513 can directly bind to the 3'-UTR of TRIM3. Luciferase activity reporter assay and western blot assay confirmed TRIM3 was a direct target of miR-4513. Recue experiments showed that downregulation of TRIM3 partially reversed the inhibitory effects of miR-4513 inhibitor on BC cell behaviors, which demonstrated that TRIM3 was a functional target of miR-4513 in BC. We also searched the targets of miR-4513 in miRTar-Base, a database contains the experimentally validated miRNA targets, and we found TRIM3 was not validated as target of miR-4513 to date. Therefore, the current study provided an experimental validated novel target of miR-4513, which will help us to understand the function of miR-4513 in human cancers. The limitation of this study was that we analyzed the effects of miR-4513 and TRIM3 on the overall survival of BC patients but we did not analyze the associations of miR-4513 and

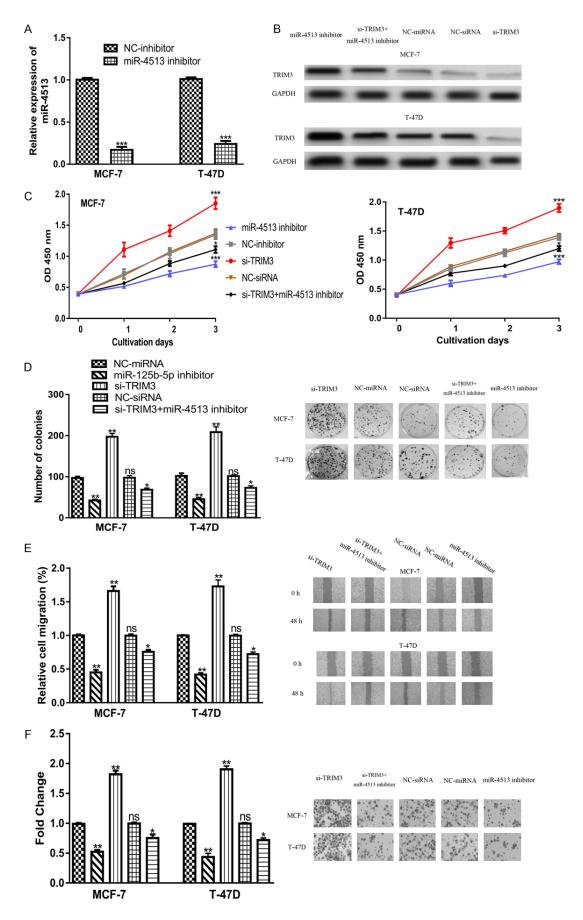


Figure 4. miR-4513 regulates BC cell proliferation, colony formation, and cell migration through targeting TRIM3. (A) qRT-PCR to analyze miR-4513 expression in cells after miR-4513 inhibitor or NC-inhibitor transfection. (B) TRIM3 expression, (C) Cell proliferation, (D) Colony formation, (E) Cell migration, and (F) Cell invasion in cells transfected with si-TRIM3, NC-siRNA, miR-4513 inhibitor, NC-inhibitor, or miR-4513 inhibitor and si-TRIM3 co-transfection. (ns not significant, *P<0.05, **P<0.01, ***P<0.001) BC: breast cancer; TRIM3: tripartite motif containing 3; miR-4513: microRNA-4513; qRT-PCR: real-time quantitative PCR; NC-inhibitor: negative control for miR-4513 inhibitor; NC-siRNA: negative control small interfering RNA.

TRIM3 expression with clinicopathological parameters. We will continue this work by recruiting BC patients to validate the value of miR-4513 and TRIM3 expression on the overall survival of BC patients in our local cohort. Moreover, the biological roles of miR-4513 and TRIM3 should be investigated in animal model in our following work.

Conclusion

In conclusion, the present study demonstrated that miR-4513 acted as an oncogenic role, whereas TRIM3 functions as a tumor suppressive role in BC cells. miR-4513 downregulation suppressed cell proliferation, colony formation, cell migration, and cell invasion through targeting TRIM3. Therefore, miR-4513 or TRIM3 may be potential novel therapeutic targets for BC.

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Disclosure of conflict of interest

None.

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References

[1] Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA and Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2018; 68: 394-424.

- Ha R, Chow D and Wynn R. Global trend in breast cancer imaging research 1992-2012: bibliometric study. Am J Roentgenol 2014; 202: 696-697.
- [3] Chaffer CL and Weinberg RA. A perspective on cancer cell metastasis. Science 2011; 331: 1559-1564.
- [4] Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. Cell 2004; 116: 281-297.
- [5] Reddy KB. MicroRNA (miRNA) in cancer. Cancer Cell Int 2015; 15: 38.
- [6] Hwang HW and Mendell JT. MicroRNAs in cell proliferation, cell death, and tumorigenesis. Br J Cancer 2006; 94: 776-780.
- [7] Jiang G, Shi W, Fang H and Zhang X. miR-27a promotes human breast cancer cell migration by inducing EMT in a FBXW7-dependent manner. Mol Med Rep 2018; 18: 5417-5426.
- [8] Tian Y, Fu X, Li Q, Wang Y, Fan D, Zhou Q, Kuang W and Shen L. MicroRNA-181 serves an oncogenic role in breast cancer via the inhibition of SPRY4. Mol Med Rep 2018; 18: 5603-5613.
- [9] Zhu X, Rao X, Yao W and Zou X. Downregulation of MiR-196b-5p impedes cell proliferation and metastasis in breast cancer through regulating COL1A1. Am J Transl Res 2018; 10: 3122-3132.
- [10] Song YK, Wang Y, Wen YY, Zhao P and Bian ZJ. MicroRNA-22 suppresses breast cancer cell growth and increases paclitaxel sensitivity by targeting NRAS. Technol Cancer Res Treat 2018; 17: 1533033818809997.
- [11] Ghanbari M, de Vries PS, de Looper H, Peters MJ, Schurmann C, Yaghootkar H, Dörr M, Frayling TM, Uitterlinden AG, Hofman A, van Meurs JB, Erkeland SJ, Franco OH and Dehghan A. A genetic variant in the seed region of miR-4513 shows pleiotropic effects on lipid and glucose homeostasis, blood pressure, and coronary artery disease. Hum Mutat 2014; 35: 1524-1531.
- [12] Ghanbari M, Erkeland SJ, Xu L, Colijn JM, Franco OH, Dehghan A, Klaver CCW and Meester-Smoor MA. Genetic variants in microRNAs and their binding sites within gene 3'UTRs associate with susceptibility to age-related macular degeneration. Hum Mutat 2017; 38: 827-838.
- [13] Zhang N, Li Y, Zheng Y, Zhang L, Pan Y, Yu J and Yang M. miR-608 and miR-4513 significantly

contribute to the prognosis of lung adenocarcinoma treated with EGFR-TKIs. Lab Invest 2018; 99: 568-576.

- [14] Koi M, Johnson LA, Kalikin LM, Little PF, Nakamura Y and Feinberg AP. Tumor cell growth arrest caused by subchromosomal transferable DNA fragments from chromosome 11. Science 1993; 260: 361-364.
- [15] El-Husseini AE, Fretier P and Vincent SR. Cloning and characterization of a gene (RFN22) encoding a novel brain expressed ring finger protein (BERP) that maps to human chromosome 11p15.5. Genomics 2000; 71: 363-367.
- [16] Huang XQ, Zhang XF, Xia JH, Chao J, Pan QZ, Zhao JJ, Zhou ZQ, Chen CL, Tang Y, Weng DS, Zhang JH and Xia JC. Tripartite motif-containing 3 (TRIM3) inhibits tumor growth and metastasis of liver cancer. Chin J Cancer 2017; 36: 77.

- [17] Fu H, Yang H, Zhang X, Wang B, Mao J, Li X, Wang M, Zhang B, Sun Z, Qian H and Xu W. Exosomal TRIM3 is a novel marker and therapy target for gastric cancer. J Exp Clin Cancer Res 2018; 37: 162.
- [18] Song Y, Guo Q, Gao S and Hua K. Tripartite motif-containing protein 3 plays a role of tumor inhibitor in cervical cancer. Biochem Biophys Res Commun 2018; 498: 686-692.
- [19] Piao MY, Cao HL, He NN, Xu MQ, Dong WX, Wang WQ, Wang BM and Zhou B. Potential role of TRIM3 as a novel tumour suppressor in colorectal cancer (CRC) development. Scand J Gastroenterol 2016; 51: 572-582.
- [20] Liu Y, Raheja R, Yeh N, Ciznadija D, Pedraza AM, Ozawa T, Hukkelhoven E, Erdjument-Bromage H, Tempst P, Gauthier NP, Brennan C, Holland EC and Koff A. TRIM3, a tumor suppressor linked to regulation of p21 (Waf1/ Cip1). Oncogene 2014; 33: 308-315.