Original Article Prognostic value of the PDLIM family in acute myeloid leukemia

Longzhen Cui^{1,2,3,4*}, Zhiheng Cheng^{5*}, Kai Hu^{6*}, Yifan Pang⁷, Yan Liu³, Tingting Qian^{1,2}, Liang Quan^{1,2}, Yifeng Dai⁸, Ying Pang¹, Xu Ye¹, Jinlong Shi⁹, Lin Fu^{1,2,4}

¹Department of Hematology, ²Translational Medicine Center, The Second Affiliated Hospital of Guangzhou Medical University, Guangzhou 510260, Guangdong, China; ³Translational Medicine Center, ⁴Department of Hematology, Huaihe Hospital of Henan University, Kaifeng 475000, Henan, China; ⁵Department of Pathology and Medical Biology, University Medical Center Groningen, University of Groningen, Groningen, Netherlands; ⁶Department of Hematology and Lymphoma Research Center, Peking University, Third Hospital, Beijing 100191, China; ⁷Department of Medicine, William Beaumont Hospital, Royal Oak, MI 48073, USA; ⁸Immunoendocrinology, Division of Medical Biology, Department of Pathology and Medical Biology, University Medical Center Groningen, University of Groningen, Groningen, Netherlands; ⁹Department of Medical Big Data, Chinese PLA General Hospital, Beijing 100853, China. *Equal contributors.

Received April 22, 2019; Accepted June 26, 2019; Epub September 15, 2019; Published September 30, 2019

Abstract: Acute myeloid leukemia (AML) is a genetically complex, highly aggressive hematological malignancy. Prognosis is usually with grim. PDZ and LIM domain proteins (PDLIM) are involved in the regulation of a variety of biological processes, including cytoskeletal organization, cell differentiation, organ development, neural signaling or tumorigenesis. The clinical and prognostic value of the PDLIM family in AML is unclear. To understand the role of PDLIM expression in AML, The Cancer Genome Atlas (TCGA) database was screened and 155 de novo AML patients with complete clinical information and the expression data of the PDLIM family were included in the study. The clinical and molecular characteristics associated with the expression of different members of the PDLIM family were summarized using various statistical methods. In 84 patients who only received chemotherapy, univariate analysis indicated that high expression of PDLIM2 or PDLIM7 was associated with shorter EFS and OS (both P<0.05 for PDLIM2, and both P<0.01 for PDLIM7). Multivariate analysis suggested that high expression of PDLIM7 was an independent risk factor for EFS and OS (both P<0.05). In the other 71 patients who underwent allogeneic hematopoietic stem cell transplantation (allo-HSCT), survival was unaffected by PDLIM expressions. In summary, high expression of *PDLIM2* and *PDLIM7*, especially the latter, could serve as adverse prognostic factors for AML, but their prognostic effects could be reversed by allo-HSCT.

Keywords: Acute myeloid leukemia, PDLIM, prognosis, next generation sequencing, mutational spectrum

Introduction

Acute myeloid leukemia (AML) is caused by dysregulated clonal expansion of mutant hematopoietic progenitor cells. It is a very heterogeneous disease with various clinical and laboratory manifestations. Despite advances in AML research and treatment, it is still a deadly disease, with a survival rate of 35% to 40% in patients younger than 60 years old, and only 5% to 15% in patients older than 60 [1]. In the past decade, lots of work have been done to improve AML prognostication and individualized targeted therapy. Our team and many other

groups have identified that FLT3-ITD and DNMT3A mutations, or high expressions of DOK4/5, PDK2/3, FHL2, and iASPP, are associated with adverse prognosis, whereas mutations of NPM1 and CEBPA, or high expression of DOK7 are indicative of favorable prognosis [2-4]. Many studies are underway to find new epigenetic or genetic factors that participate in leukemogenesis, affect prognosis or can be potential therapeutic targets.

The PDZ and LIM domains (PDLIM) are interacting structural modules shared by various proteins [5]. There are five genes encoding five dif-

ferent PDLIM isoforms, PDLIM1, 2, 4, 5, and 7. The PDZ domain interacts with certain peptide domains on various proteins, to exert different functions, particularly those related to cell polarity, intercellular junctions, recognition of immune cells, and control of proliferation and cellular migration [6, 7]. PDLIM4 plays crucial roles in many fundamental biological processes and reduced activities have been observed in some pathological processes including oncogenesis [8]. PDLIM5 and PDLIM1 are up-regulated in papillary thyroid carcinoma and PDLIM5 can promote this malignancy via activating the Ras-ERK pathway [9]. PDLIM7 is an important stabilizer of MDM2. After binding to the latter, it prevents the autoubiquitination of MDM2, which enables MDM2 to trans-ubiquitinate p53. MDM2 stabilization has been proposed to be one of the mechanisms of resistance to CDK4/6 inhibitors [10-12]. One study showed that AML patients were more likely to have low PDLIM4 expression than healthy controls; interestingly, among the AML patients, those with lower PDLIM4 expression had relatively longer overall survival than normal expressors [13]. Research is still limited on the prognostic impact of the expression of the other PDLIM members on AML, which we aimed to help elucidate with this study.

Methods

Patients

The Cancer Genome Atlas (TCGA) database was screened for de novo AML patients with complete clinical and PDLIM expression data. A total of 155 patients who met the criteria were included in the study, among which 84 were treated only with chemotherapy, and 71 later received allogeneic hematopoietic stem cell transplantation (allo-HSCT). Clinical features at diagnosis were described, including age, peripheral blood (PB) white blood cell (WBC) counts, blast percentages in the PB and the bone marrow (BM), French-American-British (FAB) subtypes, cytogenetic risk, and the frequencies of known recurrent genetic mutations. Event-free survival (EFS) and overall survival (OS) were the primary endpoints of this study. EFS was defined as the time from diagnosis to withdrawal of the study due to lack of complete remission, relapse, or death, or was censored at the last follow-up. OS was defined

as the time from diagnosis to death from any cause, or was censored at the last follow-up. All patients provided informed consent. The study protocol of TCGA database was approved by the University of Washington Human Research Committee.

Statistical analysis

Patients' clinical and molecular characteristics were outlined by descriptive statistical methods. Numerical data was described with median and/or range; intergroup comparison was done by the Mann-Whitney *U*-test. For categorical data, we used the chi-square test to perform intergroup comparisons. Survival of each group or subgroup was estimated using the Kaplan-Meier method and compared using the log-rank test. The multivariate Cox proportional hazard models of EFS and OS were constructed using a limited backward elimination process. The statistical significance level (P) was less than (≤) 0.05 for a two-tailed test. All statistical analyses were performed using the SPSS software 20.0 and the GraphPad Prism software 7.0.

Results

Prognostic significance of PDLIM family expression in AML

Both chemotherapy-only and allo-HSCT groups were divided into subgroups based on the median expression levels of each of the five PDLIM members, respectively. The high expression of PDLIM2 or PDLIM7 was noted to be associated with inferior EFS and OS in the chemotherapy-only group (all P<0.05, **Table 1** and **Figure 1**). In the allo-HSCT group, survival was independent of the expression levels of any PDLIM member.

To assess the prognostic significance of PDLIM2, PDLIM7, and other clinical and molecular factors in the chemotherapy-only group, we chose the expression levels of PDLIM2 and PDLIM7 (high vs. low), WBC count (\geq 15 vs. <15×10 9 /L), BM blasts (\geq 70 vs. <70%), FLT3-ITD (positive vs. negative), and other common genetic mutations (NPM1, RUNX1 and NRAS/ KRAS; mutated vs. wild) to construct Cox proportional hazard models for multivariate analysis (**Table 2**). Based on the results, there were two independent risk factors for both EFS and

Table 1. Comparison of EFS and OS between different expression levels of *PDLIM* members

Variables	E	FS	OS		
Variables	χ^2	P-value	χ^2	P-value	
Chemotherapy-only group					
PDLIM1 (high vs. low)	0.512	0.474	0.643	0.423	
PDLIM2 (high vs. low)	4.796	0.029	6.134	0.013	
PDLIM4 (high vs. low)	2.864	0.091	2.098	0.147	
PDLIM5 (high vs. low)	0.707	0.401	0.955	0.328	
PDLIM7 (high vs. low)	6.830	0.009	7.759	0.005	
Allo-HSCT group					
PDLIM1 (high vs. low)	0.020	0.888	1.157	0.282	
PDLIM2 (high vs. low)	2.236	0.135	0.644	0.422	
PDLIM4 (high vs. low)	0.008	0.928	0.298	0.585	
PDLIM5 (high vs. low)	0.236	0.627	1.077	0.299	
PDLIM7 (high vs. low)	1.505	0.220	1.126	0.289	

Abbreviations: EFS, event-free survival; OS, overall survival; Allo-HSCT, allogeneichematopoietic stem cell transplantation.

OS, which were high PDLIM7 expression and BM blasts \geq 70% (all P<0.05). *RUNX1* mutation was an independent risk factor only for OS (P=0.042).

Comparison of the other clinical and molecular characteristics of the patients with different PDLIM2 and PDLIM7 expression levels

The comparison of clinical and molecular characteristics between high and low PDLIM2 and PDLIM7 expression subgroups in the chemotherapy-only group were shown in Table 3. Firstly, the PDLIM2high subgroup were older (P= 0.029), had fewer patients with FAB-M2 (P= 0.006), more patients with complex karyotype (P=0.004), less frequent RUNX1-RUNX1T1 (P= 0.011) but more NRAS/KRAS mutations (P= 0.004) than the PDLIM2^{low} group. No significant differences were found in gender distribution, WBC count, BM blasts, PB blasts, cytogenetic risk group distribution, and the frequencies of other recurrent genetic mutations (FLT3, NPM1, DNMT3A, IDH1/IDH2, RUNX1, TET2 and TP53) between the two subgroups. Meanwhile, comparing with the PDLIM7^{low} subgroup, PDLIM7^{high} patients had higher WBC count (P=0.045), fewer good-risk (P=0.016), and more frequent DNMT3A mutation (P=0.028). No significant differences were found in age, gender distribution, BM blasts, PB blasts, FAB subtypes, karyotype, and the frequencies of other recurrent genetic mutations (FLT3, NPM1, IDH1/IDH2, RUNX1, NRAS/KRAS, TET2 and TP53) between the two subgroups.

Discussion

PDLIMs are important peptide modules that controls and mediates various cellular and intercellular activities. We postulated that some of the PDLIMs could play roles in AML leukemogenesis and have prognostic meanings. In this registration-based study, we were able to find that that high expression of PDLIM2 and PDLIM7 were poor prognostic factors for AML, but their effects on survival were not observed in those who underwent allo-HSCT, indicating that allo-HSCT might prevail over the deleterious influence of PDLIM2 and PDLIM7 on AML.

PDLIM2 regulates the stability of a variety of transcription factors in the hematopoietic and epithelial cells [14]. Previous work suggested that its role in oncogene-

sis might be complex, with distinctive behavior in different cancer types. In some, its expression is epigenetically suppressed, such as adult T-cell lymphoma [15-17], colorectal carcinoma [18, 19], and breast cancer [20, 21]. In these tumors, in vitro and in vivo studies have observed inhibition of tumorigenicity and increasing tumor cell death after inducing PDLIM2 expression. On the other hand, PDLIM2 may have oncogenic role in other malignancies, such as prostate cancer. It is highly expressed in cell lines derived from metastatic prostate cancer and its expression is associated with tumor progression and metastasis [22]. Another study showed that PDLIM2 was capable of activating the COP9 signaling pathway, and its high expression could promote tumor growth [23]. The contradicting functions of PDLIM2 could be explained by the evolution of tumor cells when they were grown in vitro; it also highlights the complexity of oncogenesis. In this study, high PDLIM2 expression was more likely to coexist with complex karyotype and NRAS/ KRAS mutations and was a poor prognostic factor for AML. Whether and how does PDLIM2 participate in the formation and thriving of AML, and if interactions exist between PDLIM2 and NRAS/KRAS, remain to be answered by future investigations.

Studies on PDLIM7 have more uniform conclusions than those on PDLIM2, that PDLIM7 often acts as a pro-oncogenic or oncogenic factor in cancers. A gene expression analysis of skin

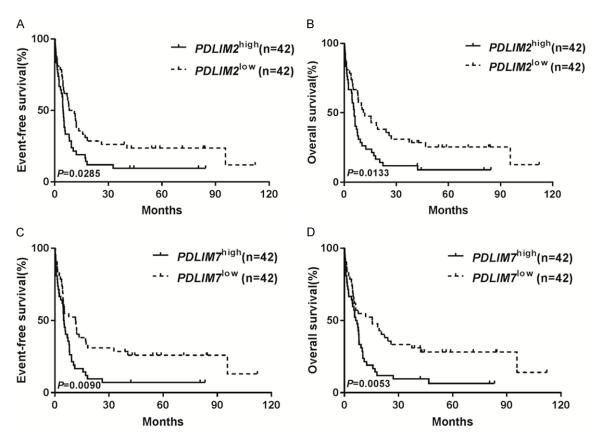


Figure 1. Kaplan-Meier curves of event-free survival (EFS) and overall survival (OS) in different expression levels of *PDLIM2* or *PDLIM7*. A, B. High *PDLIM2* expressers had shorter EFS and OS than the low expressers; C. D. High *PDLIM7* expressers had shorter EFS and OS than the low expressers.

Table 2. Multivariate analysis of EFS and OS

Variables	EFS		OS		
variables	HR (95% CI)	P-value	HR (95% CI)	P-value	
PDLIM2 (high vs. Low)	1.605 (0.917-2.809)	0.098	1.688 (0.965-2.952)	0.066	
PDLIM7 (high vs. Low)	1.878 (1.062-3.320)	0.030	1.995 (1.139-3.495)	0.016	
WBC (≥15 vs. <15×10 ⁹ /L)	0.767 (0.441-1.333)	0.346	0.819 (0.476-1.407)	0.470	
BM blasts (≥70 vs. <70%)	2.073 (1.189-3.613)	0.010	2.014 (1.155-3.514)	0.014	
FLT3-ITD (positive vs. negative)	0.960 (0.488-1.886)	0.905	1.042 (0.523-2.076)	0.906	
NPM1 (mutated vs. wild)	0.943 (0.495-1.797)	0.859	0.842 (0.437-1.621)	0.606	
RUNX1 (mutated vs. wild)	1.977 (0.877-4.457)	0.100	2.338 (1.032-5.296)	0.042	
N/KRAS (mutated vs. wild)	0.719 (0.332-1.561)	0.405	0.774 (0.357-1.678)	0.516	

Abbreviations: EFS, Event-free survival; OS, Overall survival; HR, hazard ratio; CI, confidence interval; WBC, white blood cell; BM, bone marrow.

tumors has shown that PDLIM7 expression is higher in metastatic compared with nonmetastatic tumors [24]. PDLIM7 is a suppressor of p53, decreasing the latter's proapoptotic activity and triggering mitosis [11]. In breast cancer, high expression level of PDLIM7 has been associated with low survival rate [25]. One

explanation involves Afadin, a protein that interacts with Claudin-2 via the PDLIM7 domain, and such interaction promotes breast cancer cell growth and metastasis [26]. Another proposed mechanism of PDLIM7's oncogenic property is that as one of the major *SRF/IGF2BP1*-enhanced genes, it has conserved

Prognostic role of PDLIM family in AML

Table 3. Comparison of clinical and molecular characteristics in different groups

Characteristics	PDLIM2		P	PDLIM7		Р
	High (n=42)	Low (n=42)	0.000*	High (n=42)	Low (n=42)	0.040*
Age/years, median (range)	68 (35-88)	63.5 (22-82)	0.029*	66.5 (35-81)	66.5 (22-88)	0.946*
Age group/n (%)	40 (00 0)	47 (40 5)	0.102§	40 (00 0)	45 (05.7)	0.483§
<60 years	10 (23.8)	17 (40.5)		12 (28.6)	15 (35.7)	
≥60 years	32 (76.2)	25 (59.5)	0.4005	30 (71.4)	27 (64.3)	0.5408
Gender/n (%)	00 (04 0)	40 (45 0)	0.126§	04 (57.4)	04 (50.0)	0.512§
Male	26 (61.9)	19 (45.2)		24 (57.1)	21 (50.0)	
Female	16 (38.1)	18 (54.8)		18 (42.9)	21 (50.0)	
WBC/×10 ⁹ /L, median (range)		13.7 (1-297.4)		38 (1.5-171.9)		
BM blasts/%, median (range)	73.5 (30-98)	71.5 (32-99)	0.655*	75 (30-98)	71.5 (32-99)	0.823*
PB blasts/%, median (range)	20 (0-97)	47 (0-98)	0.058*	22.5 (0-91)	32 (0-98)	0.613*
FAB subtypes/n (%)						
MO	5 (11.9)	2 (4.8)	0.236§	4 (9.5)	3 (7.1)	0.693§
M1	10 (23.8)	10 (23.8)	1.000§	7 (16.7)	13 (31.0)	0.124§
M2	5 (11.9)	16 (38.1)	0.006§	11 (26.2)	10 (23.8)	0.801§
M4	12 (28.6)	8 (19.0)	0.306§	9 (21.4)	11 (26.2)	0.608§
M5	8 (19.0)	4 (9.5)	0.212§	8 (19.0)	4 (9.5)	0.212§
M6	1 (2.4)	0 (0.0)	0.314§	0 (0.0)	1 (2.4)	0.314§
M7	1 (2.4)	2 (4.8)	0.557§	3 (7.1)	0 (0.0)	0.078§
Karyotype/n (%)						
Normal	21 (50.0)	19 (45.2)	0.662§	22 (52.4)	18 (42.9)	0.382§
Complex	10 (23.8)	1 (2.4)	0.004§	8 (19.0)	3 (7.1)	0.106§
inv(16)/CBFβ-MYH11	3 (7.1)	3 (7.1)	1.000§	1 (2.4)	5 (11.9)	0.090§
t(8;21)/RUNX1-RUNX1T1	0 (0.0)	6 (16.3)	0.011§	1 (2.4)	5 (11.9)	0.090§
11q23/MLL	1 (2.4)	2 (4.8)	0.557§	1 (2.4)	2 (4.8)	0.557§
-7/7q-	2 (4.8)	3 (7.1)	0.645§	3 (7.1)	2 (4.8)	0.645§
t(9;22)/BCR-ABL1	0 (0.0)	1 (2.4)	0.314§	1 (2.4)	0 (0.0)	0.314§
Others	5 (11.9)	7 (16.7)	0.533§	5 (11.9)	7 (16.7)	0.533§
Risk/n (%)						
Good	3 (7.3)	9 (22.0)	0.061§	2 (5.0)	10 (23.8)	0.016§
Intermediate	23 (56.1)	23 (56.1)	1.000§	23 (56.1)	23 (56.1)	0.803§
Poor	15 (36.6)	9 (22.0)	0.145§	15 (37.5)	9 (21.4)	0.110§
FLT3/n (%)			0.137§			0.567§
FLT3-ITD	4 (9.5)	11 (26.2)		7 (16.7)	8 (19.0)	
FLT3-TKD	5 (11.9)	4 (9.5)		6 (14.3)	3 (7.1)	
Wild type	33 (78.6)	27 (64.3)		29 (69.0)	31 (73.8)	
<i>NPM1</i> /n (%)	, ,	, ,	0.815§	, ,	, ,	0.102§
Mutation	14 (33.3)	13 (31.0)		17 (40.5)	10 (23.8)	
Wildtype	28 (66.7)	29 (69.0)		25 (59.5)	32 (76.2)	
DNMT3A/n (%)	- (/	- (,	0.807§	- (/	- (- ,	0.028§
Mutation	12 (28.6)	11 (26.2)		16 (38.1)	7 (16.7)	
Wildtype	30 (71.4)	31 (73.8)		26 (61.9)	35 (83.3)	
IDH1/IDH2/n (%)	(,	0= (1010)	0.266§	_= (====)	()	0.266§
Mutation	6 (14.3)	10 (23.8)		6 (14.3)	10 (28.8)	
Wildtype	36 (85.7)	32 (76.2)		36 (85.7)	32 (76.2)	
RUNX1/n (%)	33 (33.17)	02 (10.2)	1.000§	00 (00.17	02 (10.2)	0.137§
Mutation	4 (9.5)	4 (9.5)	1.000	2 (4.8)	6 (14.3)	5.101
Matadon	- (3.3)	- (3.3)		2 (4.0)	0 (14.0)	

Prognostic role of PDLIM family in AML

Wildtype	38 (90.5)	38 (90.5)		40 (95.2)	36 (95.7)	
NRAS/KRAS/n (%)			0.004§			0.746§
Mutation	10 (23.8)	1 (2.4)		6 (14.3)	5 (11.9)	
Wildtype	32 (76.2)	41 (97.6)		36 (85.7)	37 (88.1)	
TET2/n (%)			0.365§			0.763§
Mutation	5 (11.9)	8 (19.0)		6 (14.3)	7 (16.7)	
Wildtype	37 (88.1)	34 (81.0)		36 (85.7)	35 (83.3)	
TP53/n (%)			0.061§			0.061§
Mutation	9 (21.4)	3 (7.1)		9 (21.4)	3 (7.1)	
Wildtype	33 (78.6)	39 (92.9)		33 (78.6)	39 (92.9)	

Abbreviations: WBC, white blood cell; BM, bone marrow; PB, peripheral blood; FAB, French American British. '*' denotes Mann-Whitney U test; ' \S ' denotes chi-square test.

upregulation with SRF and IGF2BP1, and together they promote tumor cell growth and invasion [27]. We found that high PDLIM7 expression was more likely to coexist with high WBC count and DNMT3A mutations and was also associated with inferior prognosis of AML. In contrast to PDLIM2, PDLIM7 had an independent prognostic effect, and could be a better marker than the former.

In multivariate analysis, BM blasts ≥70% was an independent risk factor for EFS and OS. This was consistent with former finding that abnormal proliferation of BM blasts could exert significant negative effect on AML survival [28]. RUNX1 mutation was also an independent risk factor for EFS and OS in AML, in line with former findings that somatic mutation of RUNX1 was an indicator of low OS in patients with myelodysplastic syndrome, and that RUNX1 mutation would predict poor outcomes in AML [29].

Conclusions

In conclusion, our study indicated that high expression of PDLIM2 and PDLIM7 were poor prognostic factors for AML, which could be overcome by allo-HSCT. The study was limited by its registration-based, retrospective nature and a small sample size. Therefore, larger subsequent clinical studies and laboratory investigations are needed to verify our findings and decipher the role of the PDLIM family in tumorigenesis.

Acknowledgements

This work was supported by grants from the National Natural Science Foundation of China (81500118, 61501519), the China Postdo-

ctoral Science Foundation funded project (2016M600443), and Jiangsu Province Postdoctoral Science Foundation funded project (1701184B). These funding bodies were not involved in the design of the study, the collection, analysis, and interpretation of data or the writing of the manuscript.

Disclosure of conflict of interest

None.

Address correspondence to: Dr. Lin Fu, Department of Hematology, The Second Affiliated Hospital of Guangzhou Medical University, Guangzhou 510260, Guangdong, China. Tel: +86-020-34152860; Fax: +86-020-34152860; E-mail: fulin022@126.com

References

- [1] Döhner H, Weisdorf DJ, Bloomfield CD. Acute myeloid leukemia. N Engl J Med 2015; 373: 1136-52.
- [2] Cheng Z, Dai Y, Pang Y, Jiao Y, Zhao H, Zhang Z, Qin T, Hu N, Zhang Y, Ke X, Chen Y, Wu D, Shi J, Fu L. Enhanced expressions of FHL2 and iASPP predict poor prognosis in acute myeloid leukemia. Cancer Gene Ther 2019; 26: 17-25.
- [3] Zhang L, Li R, Hu K, Dai Y, Pang Y, Jiao Y, Liu Y, Cui L, Shi J, Cheng Z, Fu L. Prognostic role of DOK family adapters in acute myeloid leukemia. Cancer Gene Ther 2018; [Epub ahead of print].
- [4] Cui L, Cheng Z, Liu Y, Dai Y, Pang Y, Jiao Y, Ke X, Cui W, Zhang Q, Shi J, Fu L. Overexpression of PDK2 and PDK3 reflects poor prognosis in acute myeloid leukemia. Cancer Gene Ther 2018; [Epub ahead of print].
- [5] Ott EB, Sakalis PA, Marques IJ, Bagowski CP. Characterization of the enigma family in zebrafish. Dev Dyn 2007; 236: 3144-54.

- [6] Simpson N, Shaw R, Crepin VF, Mundy R, FitzGerald AJ, Cummings N, Straatman-Iwanowska A, Connerton I, Knutton S, Frankel G. The enteropathogenic escherichia coli type III secretion system effector Map binds EBP50/NHERF1: implication for cell signalling and diarrhoea. Mol Microbiol 2006; 60: 349-63.
- [7] Alto NM, Shao F, Lazar CS, Brost RL, Chua G, Mattoo S, McMahon SA, Ghosh P, Hughes TR, Boone C, Dixon JE. Identification of a bacterial type III effector family with G protein mimicry functions. Cell 2006; 124: 133-45.
- [8] Hunter CS, Rhodes SJ. LIM-homeodomain genes in mammalian development and human disease. Mol Biol Rep 2005; 32: 67-77.
- [9] Wei X, Zhang Y, Yu S, Li S, Jiang W, Zhu Y, Xu Y, Yang C, Tian G, Mi J, Bergquist J, Zhao M, Song F. PDLIM5 identified by label-free quantitative proteomics as a potential novel biomarker of papillary thyroid carcinoma. Biochem Biophys Res Commun 2018; 499: 338-44.
- [10] Klein ME, Dickson MA, Antonescu C, Qin LX, Dooley SJ, Barlas A, Manova K, Schwartz GK, Crago AM, Singer S, Koff A, Tap WD. PDLIM7 and CDH18 regulate the turnover of MDM2 during CDK4/6 inhibitor therapy-induced senescence. Oncogene 2018; 37: 5066-78.
- [11] Jung CR, Lim JH, Choi Y, Kim DG, Kang KJ, Noh SM, Im DS. Enigma negatively regulates p53 through MDM2 and promotes tumor cell survival in mice. J Clin Invest 2010; 120: 4493-506.
- [12] Riley MF, Lozano G. The many faces of MDM2 binding partners. Genes Cancer 2012; 3: 226-39.
- [13] Li Y, Qian J, Lin J, Qian W, Yang J, Chai HY, Wang CZ, Deng ZQ, Yao DM, Chen Q, Ma JC. Reduced expression of PDLIM4 gene correlates with good prognosis in acute myeloid leukemia. Zhongguo Shi Yan Xue Ye Xue Za Zhi 2013; 21: 1111-5.
- [14] Deevi RK, Cox OT, O'Connor R. Essential function for PDLIM2 in cell polarization in three-dimensional cultures by feedback regulation of the β 1-integrin-RhoA signaling axis. Neoplasia 2014; 16: 422-31.
- [15] Zhao T, Yasunaga J, Satou Y, Nakao M, Takahashi M, Fujii M, Matsuoka M. Human T-cell leukemia virus type 1 bZIP factor selectively suppresses the classical pathway of NFkappaB. Blood 2009; 113: 2755-64.
- [16] Yan P, Qu Z, Ishikawa C, Mori N, Xiao G. Human T-cell leukemia virus type I-mediated repression of PDZ-LIM domain-containing protein 2 involves DNA methylation but independent of the viral oncoprotein tax. Neoplasia 2009; 11: 1036-41.

- [17] Fu J, Yan P, Li S, Qu Z, Xiao G. Molecular determinants of PDLIM2 in suppressing HTLV-I Tax-mediated tumorigenesis. Oncogene 2010; 29: 6499-507.
- [18] Qu Z, Yan P, Fu J, Jiang J, Grusby MJ, Smithgall TE, Xiao G. DNA methylation-dependent repression of PDZ-LIM domain-containing protein 2 in colon cancer and its role as a potential therapeutic target. Cancer Res 2010; 70: 1766-72.
- [19] Liu S, Sun X, Wang M, Hou Y, Zhan Y, Jiang Y, Liu Z, Cao X, Chen P, Liu Z, Chen X, Tao Y, Xu C, Mao J, Cheng C, Li C, Hu Y, Wang L, Chin YE, Shi Y, Siebenlist U, Zhang X. A microRNA 221-and 222-mediated feedback loop maintains constitutive activation of NFkB and STAT3 in colorectal cancer cells. Gastroenterology 2014; 147: 847-59.
- [20] Qu Z, Fu J, Yan P, Hu J, Cheng SY, Xiao G. Epigenetic repression of PDZ-LIM domain-containing protein 2: implications for the biology and treatment of breast cancer. J Biol Chem 2010; 285: 11786-92.
- [21] Vanoirbeek E, Eelen G, Verlinden L, Carmeliet G, Mathieu C, Bouillon R, O'Connor R, Xiao G, Verstuyf A. PDLIM2 expression is driven by vitamin D and is involved in the pro-adhesion, and anti-migration and -invasion activity of vitamin D. Oncogene 2014; 33: 1904-11.
- [22] Bowe RA, Cox OT, Ayllón V, Tresse E, Healy NC, Edmunds SJ, Huigsloot M, O'Connor R. PDLIM2 regulates transcription factor activity in epithelial-to-mesenchymal transition via the COP9 signalosome. Mol Biol Cell 2014; 25: 184-95.
- [23] Adler AS, Littlepage LE, Lin M, Kawahara TL, Wong DJ, Werb Z, Chang HY. CSN5 isopeptidase activity links COP9 signalosome activation to breast cancer progression. Cancer Res 2008; 68: 506-15.
- [24] McCreery MQ, Halliwill KD, Chin D, Delrosario R, Hirst G, Vuong P, Jen KY, Hewinson J, Adams DJ, Balmain A. Evolution of metastasis revealed by mutational landscapes of chemically induced skin cancers. Nat Med 2015; 21: 1514-20.
- [25] Kales SC, Nau MM, Merchant AS, Lipkowitz S. Enigma prevents Cbl-c-mediated ubiquitination and degradation of RETMEN2A. PLoS One 2014; 9: e87116.
- [26] Tabariès S, McNulty A, Ouellet V, Annis MG, Dessureault M, Vinette M, Hachem Y, Lavoie B, Omeroglu A, Simon HG, Walsh LA, Kimbung S, Hedenfalk I, Siegel PM. Afadin cooperates with Claudin-2 to promote breast cancer metastasis. Genes Dev 2019; 33: 180-93.
- [27] Müller S, Glaß M, Singh AK, Haase J, Bley N, Fuchs T, Lederer M, Dahl A, Huang H, Chen J,

Prognostic role of PDLIM family in AML

- Posern G, Hüttelmaier S. IGF2BP1 promotes SRF-dependent transcription in cancer in a m6A- and miRNA-dependent manner. Nucleic Acids Res 2019; 47: 375-90.
- [28] Zaidi SK, Dowdy CR, van Wijnen AJ, Lian JB, Raza A, Stein JL, Croce CM, Stein GS. Altered Runx1 subnuclear targeting enhances myeloid cell proliferation and blocks differentiation by activating a miR-24/MKP-7/MAPK network. Cancer Res 2009; 69: 8249-55.
- [29] Maiques-Diaz A, Chou FS, Wunderlich M, Gómez-López G, Jacinto FV, Rodriguez-Perales S, Larrayoz MJ, Calasanz MJ, Mulloy JC, Cigudosa JC, Alvarez S. Chromatin modifications induced by the AML1-ETO fusion protein reversibly silence its genomic targets through AML1 and Sp1 binding motifs. Leukemia 2012; 26: 1329-37.