

Original Article

FOXP3⁺ Tregs exhibit different infiltrating status and predict a distinct prognosis in primary lesions and hepatic metastases in stage III&IV advanced gastric cancer

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Abstract: Advanced gastric cancer (AGC) patients with hepatic metastasis have a somber prognosis. Furthermore, understanding the molecular mechanisms and immune cells infiltrating status in the hepatic metastases event in gastric cancer become quite imperative and pressing. In this study, CD3⁺ T lymphocytes, CD8⁺ T lymphocytes and PD-L1 were favorable prognostic indicators. The positive expression of PD-L1 indicates better prognosis, and FOXP3^{high}PD-L1^{neg} could be regarded as a poor prognostic factor in the multivariate analysis in primary lesions. The infiltration of FOXP3⁺ Treg is significantly higher in primary tumor lesions than paired hepatic metastatic lesions (P<0.0001). In AGC patients with hepatic metastasis, low infiltration of FOXP3⁺ Tregs both on primary lesions and metastatic lesions indicate better prognosis. Besides, compared with this in hepatic metastases, the proportion of PD-1⁺CD8⁺ T lymphocytes in CD8⁺ T lymphocytes was elevated in the primary lesions. Moreover, compared with Tregs which were infiltrated in primary lesions, they exhibit higher immunosuppressive effects on hepatic metastases despite the decrease in number. Thus, FOXP3⁺ Tregs exhibit different infiltrating status and predict a distinct prognosis in primary lesions and hepatic metastases, implying the immunological heterogeneity of primary and metastatic lesions in AGC. These conclusions would provide further theoretical basis and a potential target for immunotherapy of AGC.

Keywords: Gastric carcinoma, FOXP3⁺ regulatory T cell, PD-L1, hepatic metastasis, prognosis

Introduction

Gastric cancer (GC), still ranking the fifth most frequently diagnosed cancer and the third leading cause of cancer death, is a high-mortality disease with limited effective therapeutic strategies [1, 2]. GC is most often diagnosed at an advanced stage, and patients with advanced disease have a somber prognosis [3].

At the time of diagnosis, 35% of gastric cancer patients have evidence of distant metastases, 31% with peritoneal disease, 14% with hepatic metastases, and 16% with lung metastases. Furthermore, distant metastasis have commonly been considered invariably fatal situations of gastric cancer [4]. Actually, clinical approach to gastric cancer patients with hepatic

metastasis is still debated, only a few patients are candidates for hepatic resection because these are often multiple, scattered, bilobar metastases, and recurrence usually occurs with a combination of various patterns, such as peritoneal dissemination, lymph node metastases, and distant metastases [5, 6]. However, the researches about the mechanisms of remodelling the immune microenvironment of gastric cancer hepatic metastases were quite rare [7]. Thus, understanding the molecular mechanisms and immune cells infiltrating status which may drive the hepatic metastases event in gastric cancer become quite imperative and pressing.

In recent years, immunotherapy with immune checkpoint inhibitors has revolutionised the

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oncology landscape by targeting the host immune system in advanced cancer and exhibited quite promising perspective [8-10]. Early-phase clinical trials demonstrating the potential applications of Programmed death-1 (PD-1) therapy in gastric cancer were performed in patients with positive immunohistochemical expression of Programmed death-ligand 1 (PD-L1), a ligand for PD-1 [11], however, the therapeutic effect is still not ideal [12]. As for the expression of PD-L1 in tumor cells be an independent factor which could indicate either better or worse prognosis in gastric cancer had been widely reported [13]. Thus, a study of tumor PD-L1 expression in hepatic metastases from gastric cancer is needed.

Tumor infiltrative forkhead box P3-positive (FOXP3⁺) regulatory T cells (Tregs) can suppress anticancer immunity, thereby hindering protective immunosurveillance of neoplasia and hampering effective antitumor immune responses in tumor-bearing hosts, thus promoting tumor development and progression in human cancers including gastric cancer [14-17]. FOXP3 is a transcription factor that is specifically expressed by natural Tregs. Its expression and stability are crucial for Tregs to regulate effector T cells' function [18, 19], in addition, CD4⁺CD25⁺CD127^{low/-} T cell population had typical characteristics of Treg cells and FOXP3 expression was significantly higher than other groups and correlated positively with the classic regulatory T cells [20]. However, predicting the prognosis of patients with the infiltration of Tregs in the microenvironment of GC is still controversial [21, 22]. Moreover, the studies about the infiltrating status between primary tumors and hepatic metastases, and the correlation between the infiltration of Tregs in hepatic metastases and the GC patients' prognosis are still quite rare.

In this study, a tissue microarray (TMA) including 266 advanced gastric cancer (AGC) primary lesion specimens (including 68 paired hepatic metastases lesions) was used to investigate the expression of PD-L1, quantified tumor infiltrating CD3⁺, CD8⁺ T lymphocytes, FOXP3⁺ Tregs density to determine their relationships with clinicopathological features and patients' prognosis in advanced gastric cancer patients (Figure 1A-D). The immune microenvironment of hepatic metastasis was also assessed and

compared with that of the primary tumor from the same case.

Materials and methods

Patients and samples

To evaluate the immune indices in the advanced gastric cancer samples, we retrospectively assessed 266 advanced stage III-IV gastric cancer samples (using TMA, including 68 cases with hepatic metastasis) from patients who underwent primary or metastatic tumor resection between December 2010 to June 2016 at Department of Gastrointestinal Surgery, Renji Hospital, School of Medicine, Shanghai Jiao Tong University. All the samples were definitely diagnosed as gastric cancer by Department of Pathology.

In this study, we excluded the following types of patients: (1). Patients without complete clinical information, follow-up data, etc. (2). Patients with non-neoplastic resection such as palliative gastrointestinal bypass surgery and non-adenocarcinoma patients. (3). Patients receiving other neoadjuvant treatment or previous radiotherapy. (4). Patients with perioperative death from various surgical complications. (5). Broken tissue samples were unavailable for TMA. Overall Survival time was defined as the interval between the gastrectomy and patient death or survival, and the final follow-up date was January 8, 2018, for all cases examined [23, 24]. All patients received the standard treatments such as D2 radical resection for primary lesions and hepatic metastases resection for the hepatic metastases lesions. Tumor TNM stage was assigned based on pathological tumor, node, and metastasis staging by the American Joint Committee on Cancer (AJCC 8th edition) staging system. For each case, the diagnosis was confirmed by a senior pathologist.

Every patient's tumor formalin-fixed, paraffin-embedded (FFPE) tissues on the TMA was consecutive, and the TMA was constructed using a tissue arrayer with 5 μm thickness. According to the manufacturer's instructions, the immunohistochemical staining was performed with the manual of Dako REAL EnVision Detection System (K5007, Dako). The following primary antibodies were used: Anti-CD3 (1:100, ab-16669, Abcam); Anti-CD8 (1:100, ab4055,

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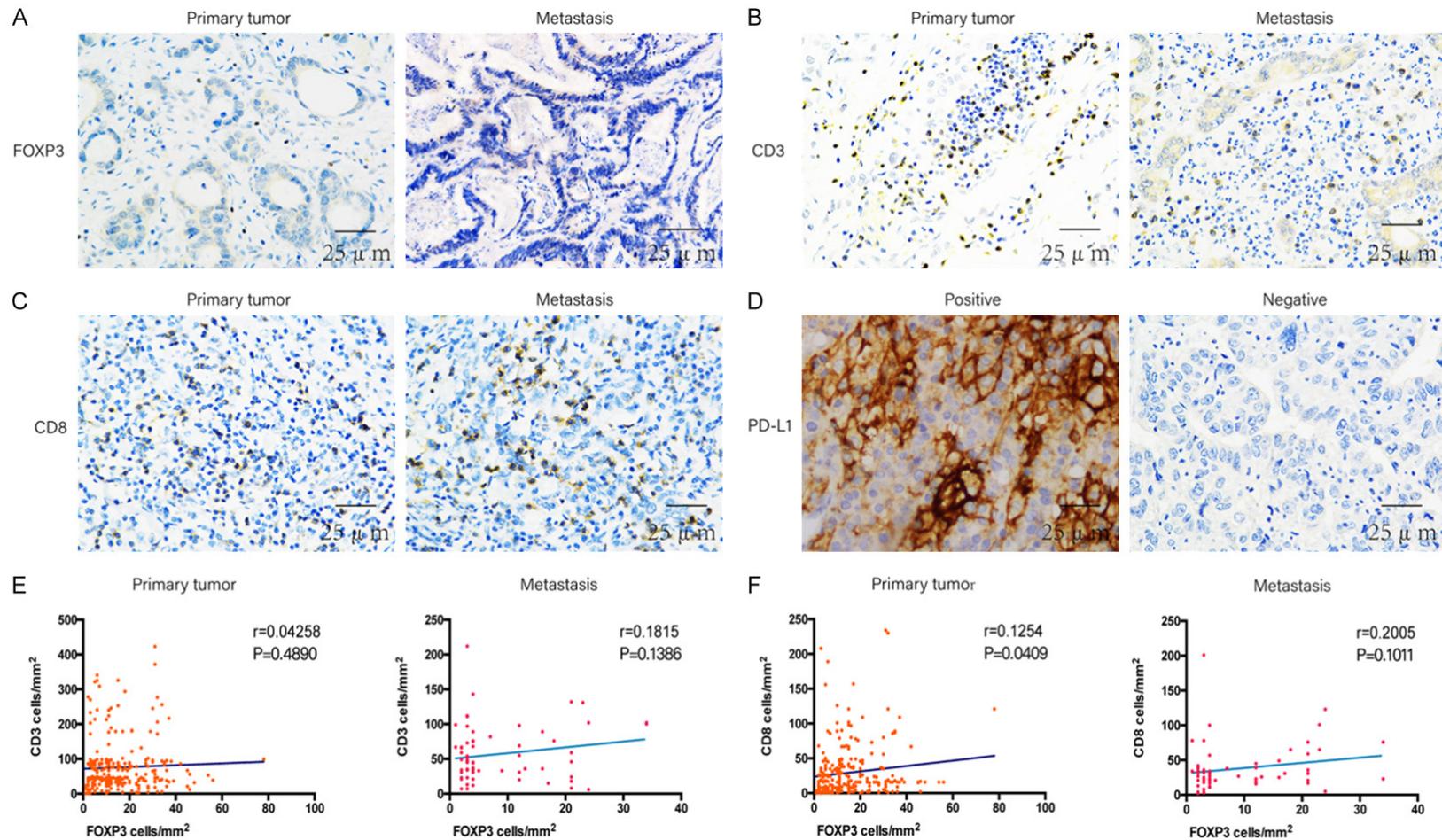


Figure 1. FOXP3⁺ Tregs, CD3⁺ T lymphocytes, CD8⁺ T lymphocytes and PD-L1 expression in primary lesions or hepatic metastases by immunohistochemistry (×400). The representative images of FOXP3⁺ Tregs (A); CD3⁺ T lymphocytes (B); CD8⁺ T lymphocytes (C); the representative positive or negative expression of PD-L1 in tumor tissues (D); the correlations between FOXP3⁺ Tregs and CD3⁺ T lymphocytes in primary lesions or hepatic metastases (E); the correlations between FOXP3⁺ Tregs and CD8⁺ T lymphocytes in primary lesions or hepatic metastases (F).

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Abcam); Anti-FOXP3 (1:100, ab20034, Abcam); Anti-PD-L1 (1:100, 22C3, Dako). 6 pairs of TNM stage IV Fresh AGC samples with hepatic metastasis were collected from treatment-naive adults undergoing surgery for AGC between October 2017 to September 2019 after informed consent and approval from the Shanghai Jiaotong University School of Medicine, Renji Hospital Ethics Committee. This retrospective study was performed according to the criteria of the Ethical Committee of the Shanghai Jiao Tong University School of Medicine. Renji Hospital consent from the patient was obtained.

Quantitative immunohistochemical analysis

In brief, each primary antibody-probed section was incubated with 0.3% hydrogen peroxide for 30 minutes, and then blocked with 10% BSA (Sangon, Shanghai, China) at 4°C overnight, and followed by the HRP second antibody (Thermo Scientific, US) at room temperature for 1 h. After that, positive staining was visualized with DAB substrate liquid (Gene Tech, Shanghai) and counterstained with hematoxylin. Digital images of the sections were obtained at 40× and 200× magnifications by ZEISS Axio Vert. A1 microscope system. 5 visual areas showing highest infiltrating densities at 40× magnification were chosen firstly and then counted the cell numbers at 200× magnification. After counting the numbers, cell density were calculated in mm² for further statistics. In a two-category immunoscore analysis, patients with cell numbers greater than the median were defined as 'high' and those were smaller cell numbers were defined as 'low' (The median count of hepatic metastatic lesions was comparable to that of the primary lesion). And PD-L1 positivity was defined as staining in 1% or more of tumor cells [25, 26]. PD-L1 expression on tumor cells, instead of stroma, was immunohistochemically analyzed by an experienced pathologist [27].

Flow cytometry

For analysis of surface markers, cells were stained in PBS containing 2% fetal bovine serum (FBS) with antibodies as indicated. Antibodies staining was performed according to the manufacturer's instructions (eBioscience). Cells were stained with fixable viability dye eFluor™ 780 and antibodies as indicated. All samples were processed on a LSRFortessa™ X-20 flow cytometer (BD Biosciences) and

data were analyzed by FlowJo software (Tree-Star). The following flow cytometry antibodies were purchased from Biolegend: CD3 (OKT3), CD8 (RPA-T8), PD-1 (EH12.2H7), CD127 (A01-9D5); FOXP3 (PCH101), CD25 (BC96), CTLA-4 (14D3) were from eBiosciences.

In vitro suppression assay

All tissue samples were cut into small pieces and incubated in 300 U/mL type IV collagenase (SIGMA) for 40 min at 37°C. After passing through a 300 mesh filter, cells were washed twice with PBS. Tumor infiltrating lymphocyte was isolated by density gradient centrifugation with Ficoll-Paque (GE Healthcare). Tregs isolated from gastric cancer tumor tissues by FACS on a BD FACS ARIA II sorter (BD Biosciences). Responder T cells were sorted from human peripheral blood and labelled with CellTrace Violet following the protocol from CellTrace Violet Cell Proliferation Kit (Invitrogen). Labeled responder T cells were then either cultured alone or mixed with different ratios of Tregs from primary tumor or hepatic metastasis (1:0; 2:1; 4:1; 8:1). Cell mixtures were stimulated under anti-CD3/anti-CD28 antibody for 48 h. Proliferative cells were further detected by flow cytometry.

Statistical analysis

SPSS 23.0 and GraphPad Prism 6.0 were used for statistics in this study. Box and whiskers plot diagrams represent the median, the interquartile range, minimum (Min) and maximum (Max) of positive cell counting numbers per mm² (Including CD3, CD8, FOXP3). Chi-square tests were performed to compare CD3, CD8, FOXP3 or PD-L1 expression with clinical features. Overall survival analysis was performed using the Kaplan-Meier method and the long-rank test (Time unit: month). The hazard ratio (HR) of mortality was assessed by the Cox regression model. Two-sided, *P*-values <0.05 were considered to be significant.

Results

Clinicopathologic characteristics

A retrospective study of 266 AGC patients, including 193 TNM stage III cases, and 73 TNM stage IV cases, was conducted (68 cases with paired primary lesions and hepatic metastasis).

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ses). The median age of the AGC patients was 63 (33-89) years, and the median OS time was 32 (0-99) months. A total of 201 (75.56%) patients died during the follow-up period. The detailed clinicopathological characteristics of the patients are presented in **Table 1**.

Correlation of CD3⁺ lymphocytes, CD8⁺ lymphocytes and PD-L1 with AGC patients' clinicopathological parameters

In this study, CD3⁺ T lymphocytes could be favorable prognostic factor (HR: 0.694, 95% CI: 0.524-0.918, P=0.010), high CD3 expression indicates better prognosis (5-year OS: 17.5 ± 0.034 vs 32.5 ± 0.042, P=0.010), at the same time, CD8⁺ T lymphocytes could also be favorable prognostic factor (HR: 0.645, 95% CI: 0.487-0.854, P=0.002), high CD8 expression indicates benign prognosis (5-year OS: 15.3 ± 0.032 vs 35.0 ± 0.043, P=0.002). Besides, 53 out of 266 cases (19.9%) showed positive PD-L1 expression in primary tumor cells, whereas 213 patients out of 266 cases (80.1%) showed negative PD-L1 expression. Nevertheless, 6 out of 68 cases (8.8%) showed positive PD-L1 expression in hepatic metastatic tumor cells, whereas 62 patients out of 68 cases (91.2%) showed negative PD-L1 expression (**Figure 2B**). The positive expression of PD-L1 in primary tumor indicates better prognosis (HR: 0.623, 95% CI: 0.430-0.901, P=0.012; 5-year OS: 22.0 ± 0.029 vs 36.1 ± 0.069, P=0.012). However, in hepatic metastases, all of these indicators has no statistically significant indication for AGC patients' prognosis (**Tables 1, 2; Figure 3B-D**).

PD-L1 expression and FOXP3⁺ Treg infiltration are associated with AGC patients' overall survival

In this research, high FOXP3 expression indicates poor prognosis in primary tumors, while the correlation between FOXP3⁺ Tregs and patients' prognosis features did not show significant differences in hepatic metastases (**Figure 3A**). The ratio of Tregs to CD3⁺ T cells or CD8⁺ T cells was dichotomous, in primary tumors, both high ratio of FOXP3/CD3 and FOXP3/CD8 exhibit worse prognosis (5-year OS: 29.5 ± 0.041 vs 20.2 ± 0.036, P=0.013; 30.4 ± 0.042 vs 19.3 ± 0.035, P=0.026, respectively). Then, when FOXP3 was combined with PD-L1, FOXP3^{high}PD-L1^{neg} could be regard-

ed as a poor prognostic factor in the multivariate analysis (HR: 1.514, 95% CI: 1.119-2.049, P=0.007; 5-year OS: 30.6 ± 0.040 vs 18.3 ± 0.036, P=0.007) (**Table 2; Figure 4B, 4C**).

Different immune cells infiltrating status between primary tumor lesions and hepatic metastatic lesions indicates diverse prognosis

68 pairs of primary tumor lesions and hepatic metastatic lesions were analysed about the infiltrating status of FOXP3⁺ Tregs, CD3⁺ T cells, CD8⁺ T cells. The infiltration of FOXP3⁺ Treg is significantly higher in primary tumor lesions than paired hepatic metastatic lesions, (P< 0.0001= nevertheless, the infiltration of CD3⁺, CD8⁺ lymphocytes did not show significant difference between primary lesions and metastatic lesions) (**Figure 2A**). This phenomenon is also confirmed in 6 pairs of fresh advanced gastric cancer samples by Flow Cytometry, the proportion of CD4⁺CD25⁺CD127^{low} Tregs is significantly upregulated in primary tumors than metastases. However, the proportion of CD3⁺ and CD8⁺ T lymphocytes didn't show difference (**Figure 5A-C**). Also, the patients without metastases have a significantly lower Treg infiltration than the ones with metastases in primary lesions, adversely, the patients without metastases have a significantly higher CD3⁺ T cells infiltration than the ones with metastases in primary lesions (**Figure 2C**). In addition, CD8⁺ T lymphocytes and FOXP3⁺ Tregs show a slight positive correlation in primary tumors without hepatic metastasis, however, CD3⁺ T lymphocytes and FOXP3⁺ Treg did not show any correlation (**Figure 1E, 1F**). Moreover, in AGC patients with hepatic metastases, low infiltration of FOXP3⁺ Tregs both on primary lesions and metastatic lesions show better prognosis. It would also be an independent favorable prognostic factor (HR: 0.244, 95% CI: 0.075-0.792, P=0.019) (**Figure 4D**).

Tregs exhibit higher immunosuppressive effects on hepatic metastases than primary tumors despite the decrease in number

To further investigate the differences of Treg's functional markers and the expression of PD-1 on CD8⁺ T lymphocytes between the primary tumors and the hepatic metastases, we examined them using fresh AGC samples by Flow Cytometry. Compared with this in the hepatic metastases, the proportion of PD-1⁺CD8⁺ T lym-

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Table 1. Clinicopathologic characteristics of the total 266 gastric cancer patients

	n	T cells									Tumor cells		
		FOXP3			CD3			CD8			PD-L1		
		Low (%)	High (%)	p-value	Low (%)	High (%)	p-value	Low (%)	High (%)	p-value	Neg (%)	Pos (%)	p-value
Gender	266			0.568			0.435			0.322			0.066
Female	78	41	59		55.1	44.9		47.4	52.6		87.2	12.8	
Male	188	44.7	55.3		49.5	50.5		53.7	46.3		77.1	22.9	
Age	266			0.089			0.322			0.671			0.279
≤60	82	52.4	47.6		46.3	53.7		50	50		84.1	15.9	
>60	184	39.7	60.3		53.2	46.8		52.7	47.3		78.3	21.7	
T stage	266			0.02			0.013			0.001			0.356
2	5	40	60		60	40		40	60		80	20	
3	69	56.5	43.5		40.1	59.9		33.3	66.7		73.9	26.1	
4	192	39.1	60.9		54.7	45.3		58.9	41.1		82.3	17.7	
N stage	266			<0.0001			0.777			0.395			0.228
N0	30	16.7	83.3		53.3	46.7		56.7	43.3		73.3	26.7	
N+	236	47	53		50.8	49.2		51.3	48.7		80.5	19.5	
M stage	266			<0.0001			0.023			<0.0001			0.202
M0	193	57	43		46.6	53.4		43.5	56.5		78.2	21.8	
M+	73	9.6	90.4		63	37		73.9	26.1		84.9	15.1	
pTNM	266			<0.0001			0.023			<0.0001			0.202
3	193	57	43		46.6	53.4		43.5	56.5		78.2	21.8	
4	73	9.6	90.4		63	37		73.9	26.1		84.9	15.1	

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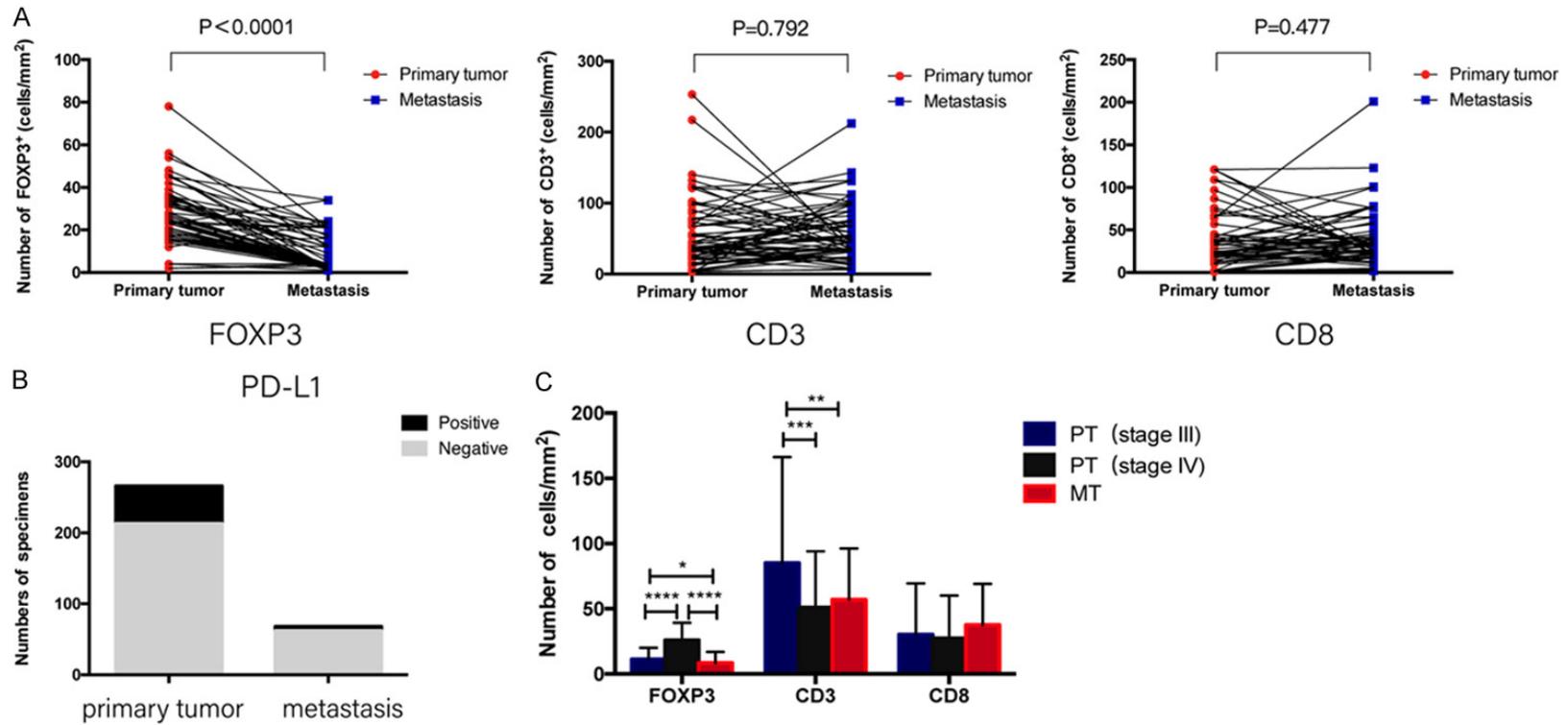


Figure 2. Cell counts comparison between primary tumor and paired hepatic metastasis. A: Cell counts comparison between primary tumor and paired hepatic metastasis about FOXP3⁺ Tregs, CD3⁺ T lymphocytes and CD8⁺ T lymphocytes; B: The proportion of PD-L1 positive patients between primary tumor and paired hepatic metastasis; C: Cell counts comparison between primary tumor (with or without hepatic metastasis) and hepatic metastasis about FOXP3⁺ Tregs, CD3⁺ T lymphocytes, CD8⁺ T lymphocytes (PT: primary tumor; stage III: without hepatic metastasis; stage IV: with hepatic metastasis).

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Table 2. Univariate and multivariate analysis for prognostic factors in 266 GC patients

Risk factors	Univariate analysis		Multivariate analysis	
	HR (95% CI)	P	HR (95% CI)	P
Age: female	1.004 (0.990-1.019)	0.582		
Gender	0.837 (0.620-1.130)	0.245		
T stage	1.150 (0.864-1.530)	0.339		
N stage	0.854 (0.565-1.291)	0.454		
M stage	2.186 (1.617-2.955)	<i><0.0001</i>		
pTNM	2.186 (1.617-2.955)	<i><0.0001</i>	1.675 (1.192-2.354)	0.003
FOXP3 ⁺ Tregs density: high	1.519 (1.142-2.021)	0.004		
CD3 ⁺ T cells density: high	0.694 (0.524-0.918)	0.010	0.817 (0.598-1.116)	0.193
CD8 ⁺ T cells density: high	0.645 (0.487-0.854)	0.002	0.745 (0.543-1.023)	0.069
Tumor PD-L1 expression: positive	0.623 (0.430-0.901)	0.012		
FOXP3 ⁺ Tregs density/CD3 ⁺ T cells density: high	1.422 (1.077-1.878)	0.013		
FOXP3 ⁺ Tregs density/CD8 ⁺ T cells density: high	1.370 (1.038-1.809)	0.026		
FOXP3 ⁺ Tregs density: high & PD-L1: negtive	1.655 (1.253-2.185)	0.0001	1.514 (1.119-2.049)	0.007

Tumor PD-L1 expression, CD3⁺ cell density, CD8⁺ cell density, FOXP3⁺ Treg density are evaluated in primary tumors, and variables showing *P* values less than 0.05 are presented in italic.

phocytes in the CD8⁺ T lymphocytes was elevated in the primary lesions (**Figure 5D**), we next sought to determine the functionality of tumor-infiltrated Tregs by investigating Treg signature markers and found that Treg's functional marker CTLA-4 was highly expressed by Treg infiltrated in hepatic metastases compared with Tregs infiltrated in the primary lesions, rather than CD25 and FOXP3 (**Figure 6A, 6B**). Moreover, we examined the activity of Tregs using an in vitro suppression assay, and found that Tregs from hepatic metastases did exhibit higher immunosuppressive effects than which from primary tumors, revealing that Tregs show higher immunosuppressive effects on hepatic metastases than primary tumors despite the decrease in number (**Figure 6C, 6D**). These data implied that the different immune status between the primary lesions and the hepatic metastases might be attributed to the diverse responses to the immunotherapy.

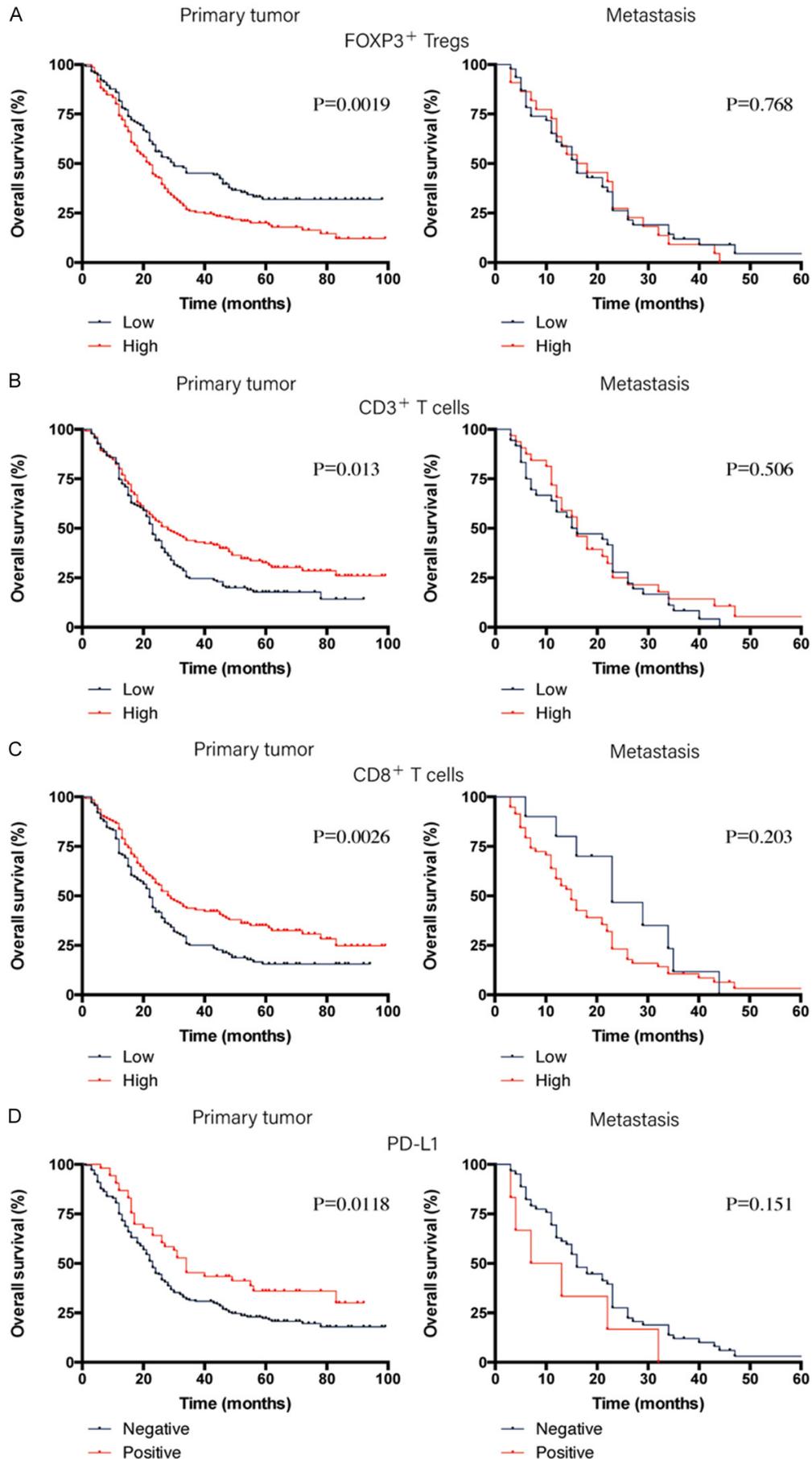
Discussion

Immune cells infiltrated in tumor microenvironment have quite crucial influence on the occurrence and development of tumor [28, 29]. In this study, we conducted immunohistochemical evaluation of some certain immune indices from 266 gastric cancer samples, and mainly focused on the different infiltrating status of some immune cells predicting a distinct prognosis in primary lesions and hepatic metastases.

In this study, CD3⁺ T lymphocytes, CD8⁺ T lymphocytes also generally be favorable prognostic indicators, and the positive expression of PD-L1 indicates better prognosis, which is consistent with the trend in some previous reports [30, 31]. Besides, accumulating studies have demonstrated that a large number of Tregs infiltrate into various types of tumors in humans. And high frequency of tumor-infiltrating FOXP3⁺ Tregs was often significantly negatively correlated with patients' survival [32, 33]. Our results also exhibit that high infiltrating of the FOXP3⁺ Tregs indicate poor prognosis in stage III-IV AGC patients. A relative study reported that high densities of PD-L1 in patients with high CD8/FOXP3 and low CD8/PD-L1 ratios correlated with increased survival in GC [34]. In our study, it showed that the combination of FOXP3 and CD3 or CD8 can predict the prognosis in the primary lesions of AGC patients. Combining two indicators to predict prognosis would be a good idea, suggesting that high expression of FOXP3 in AGC indicates poor prognosis, while higher infiltrating of CD3⁺ T cells and CD8⁺ T cells suggests a favorable prognosis. In addition, the combination of FOXP3⁺ Tregs infiltration and the expression of PD-L1 in tumor cells can be considered as an independent predictor of multivariate analysis.

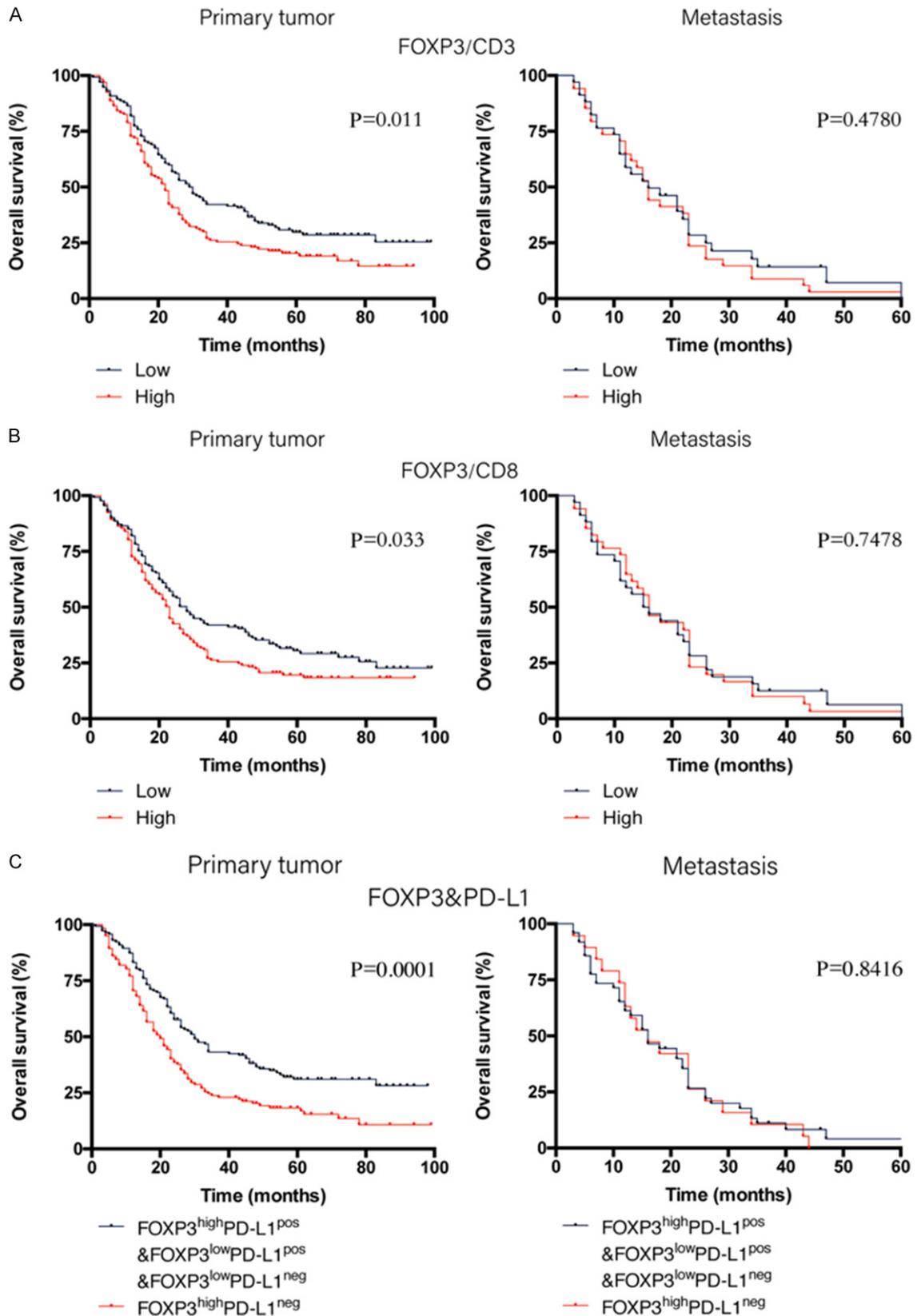
In order to elucidate the difference of the expression of immune indices between primary lesions and hepatic metastatic lesions, we used the paired primary focus and metastatic

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Figure 3. Correlation of immune indicators expressed on primary lesions or hepatic metastases with AGC patients' overall survival. Correlation of FOXP3⁺ Tregs (A); CD3⁺ T lymphocytes (B); CD8⁺ T lymphocytes (C) and PD-L1 (D) expressed on primary lesions or hepatic metastases with AGC patients' overall survival.



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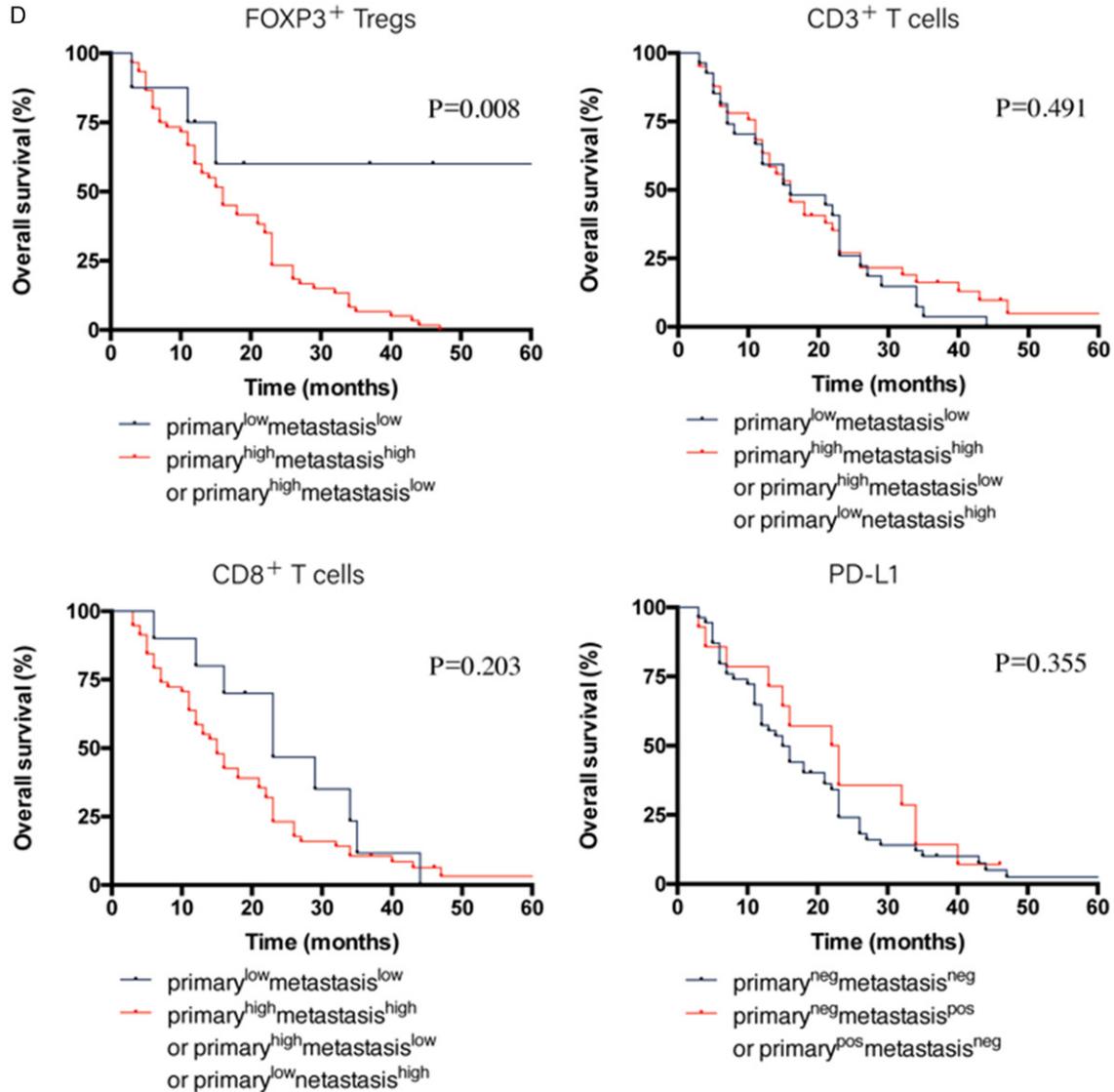


Figure 4. Combination of some indicators to predict AGC patients' prognosis. Correlation of FOXP3⁺ Tregs/CD3⁺ T lymphocytes ratio (A); FOXP3⁺ Tregs/CD8⁺ T lymphocytes ratio (B); FOXP3⁺ Tregs & PD-L1 (C) on primary lesions or hepatic metastases with AGC patients' overall survival; Correlation of 4 indicators (comparison between primary lesions and hepatic metastases) with AGC patients' overall survival (D).

lesions in AGC patients with hepatic metastasis. Compared with the primary lesion, although the infiltration of Tregs is significantly higher than the metastatic lesions, it exhibited that the expression of the Treg functional marker CTLA-4 in Tregs in metastatic lesions was elevated than matched primary lesions in fresh clinical AGC samples, which suggest a more robust inhibition. Moreover, Tregs also show higher immunosuppressive effects on hepatic metastases than primary tumors by in vitro suppression assay. At present, there are a number of studies dedicated to discovering the heterogeneity of tumor-infiltrating Tregs, the sub-

sets with strong inhibiting capacity [35, 36], and the Tregs infiltrated in hepatic metastases just exhibit a strong inhibitory function.

At the same time, the expression of PD-1 in the infiltrating CD8⁺ T lymphocytes in the hepatic metastases was lower than that in the matched primary lesions. Moreover, the expression of PD-L1 in hepatic metastases was also lower than that of primary lesions. Up to now, the predictive role of PD-L1 expression is still controversial in the clinic, the tumor stratification based on the presence of T lymphocytes and PD-L1 might be a promising predictive tool

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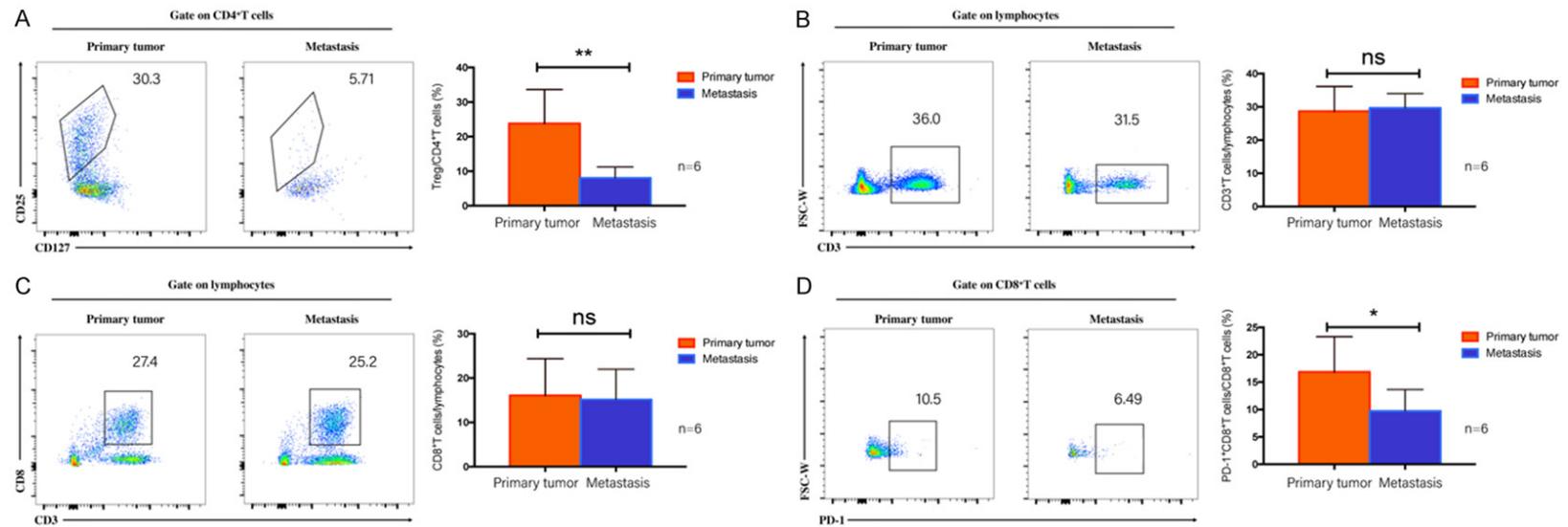


Figure 5. FOXP3⁺ Tregs, CD3⁺ T lymphocytes, CD8⁺ T lymphocytes and PD-L1 expression in primary lesions or hepatic metastases of AGC by FACS. Proportions of CD4⁺CD25⁺CD127^{low} Tregs (A); CD3⁺ T lymphocytes (B); CD8⁺ T lymphocytes (C) between primary lesions and paired hepatic metastases. Proportions of PD-1⁺CD8⁺ T lymphocytes between primary lesions and paired hepatic metastases (D). *, P<0.05; **, P<0.01; ns: no significance.

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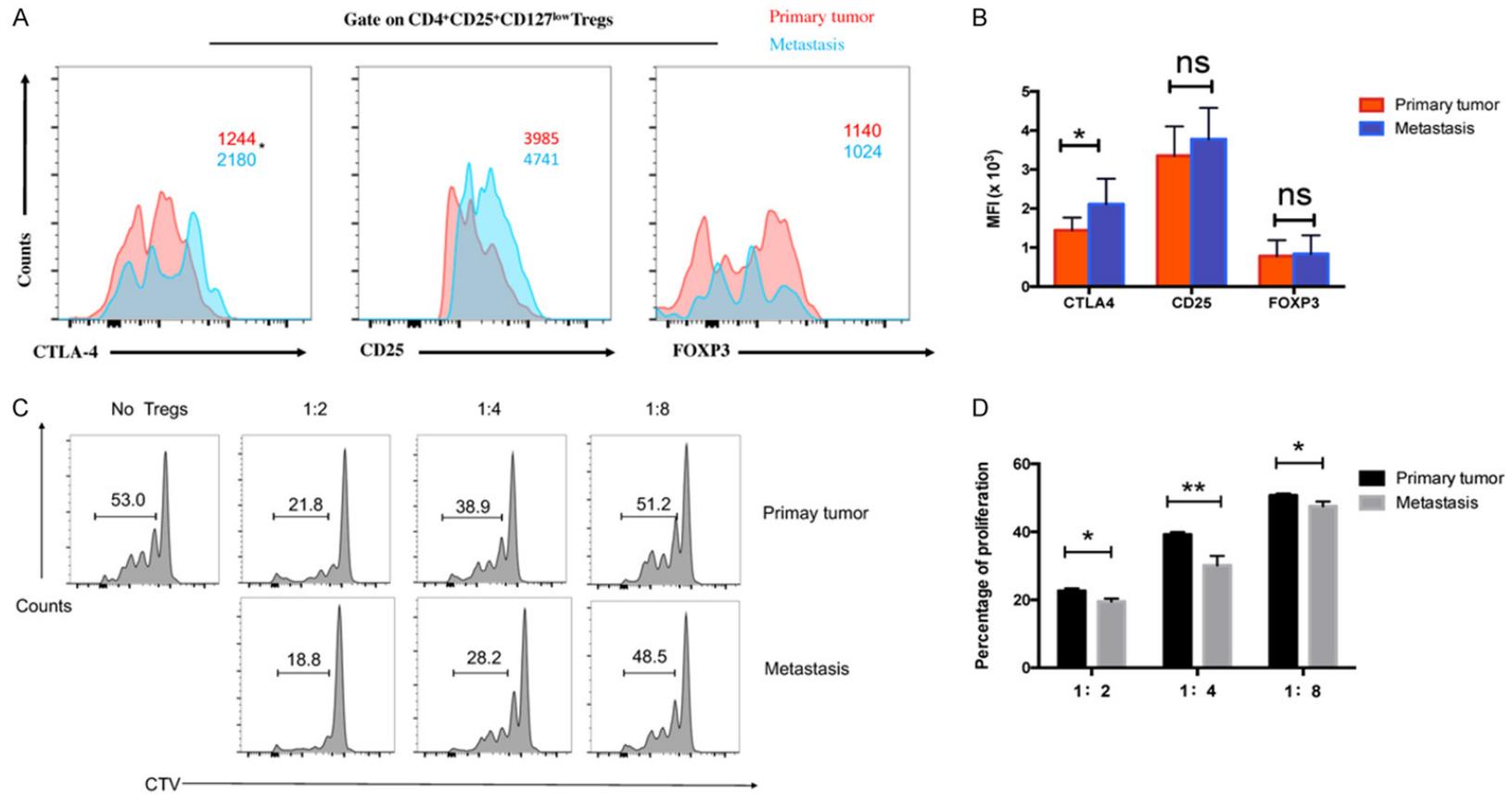


Figure 6. Tregs exhibit higher immunosuppressive effects on hepatic metastases. MFI of various Treg functional marker expression by Treg cells within primary lesions and hepatic metastases as indicated (A) (n=6). *, P<0.05; quantitative analysis of the MFI of Treg functional markers between primary lesions and paired hepatic metastases; (B) *, P<0.05. In vitro suppression assay was performed in Tregs from primary tumor or hepatic metastasis. Labeled responder T cells were either cultured alone or mixed with different ratios of Tregs from primary tumor or hepatic metastasis (1:0; 2:1; 4:1:8:1) (C); percentage of proliferated responder T cells was assessed (D). All data represent mean ± S.D. *, P<0.05; **, P<0.01.

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to define optimal therapy for patients with advanced cancer. In this research, the above immune indicators suggest the immunological heterogeneity of primary and metastatic lesions, which may indicate that anti-PD-L1/PD-1 treatment is more difficult to achieve the desired effect on hepatic metastases, while targeting the certain subset of Tregs, which exhibit strong inhibition in the tumor immune microenvironment, may become a promising therapeutic strategy. However, the number of samples of matched primary and metastatic lesions is still quite rare, more samples are needed to enhance persuasiveness. Due to the limitations of single-center retrospective studies, more multicenter studies should be conducted to validate all of these results, moreover, the specific pathways or related mechanisms may require further in-depth exploration in the future.

Conclusions

In this study, large scale advanced gastric cancer samples (including AGC with hepatic metastasis) were used to elucidate both CD3⁺, CD8⁺ T lymphocytes and PD-L1 in tumor cells with positive prognostic effects, the FOXP3⁺ Tregs, with poor prognostic implications. At the same time, the differences of immune molecules between primary and metastatic lesions were also been compared. In addition, the immunological heterogeneity of primary and metastatic lesions was also exhibited. It would provide further theoretical basis and potential target for immunotherapy of advanced gastric cancer.

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Disclosure of conflict of interest

None.

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References

- [1] Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA and Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2018; 68: 394-424.
- [2] Van Cutsem E, Sagaert X, Topal B, Haustermans K and Prenen H. Gastric cancer. *Lancet* 2016; 388: 2654-2664.
- [3] Digkila A and Wagner AD. Advanced gastric cancer: current treatment landscape and future perspectives. *World J Gastroenterol* 2016; 22: 2403-2414.
- [4] Kerkar SP, Kemp CD, Duffy A, Kammula US, Schrupp DS, Kwong KF, Quezado M, Goldspiel BR, Venkatesan A, Berger A, Walker M, Toomey MA, Steinberg SM, Giaccone G, Rosenberg SA and Avital I. The GYMSSA trial: a prospective randomized trial comparing gastrectomy, metastasectomy plus systemic therapy versus systemic therapy alone. *Trials* 2009; 10: 121.
- [5] Okano K, Maeba T, Ishimura K, Karasawa Y, Goda F, Wakabayashi H, Usuki H and Maeta H. Hepatic resection for metastatic tumors from gastric cancer. *Ann Surg* 2002; 235: 86-91.
- [6] Tiberio GA, Baiocchi GL, Morgagni P, Marrelli D, Marchet A, Cipollari C, Graziosi L, Ministrini S, Vittimberga G, Donini A, Nitti D, Roviello F, Coniglio A and de Manzoni G. Gastric cancer and synchronous hepatic metastases: is it possible to recognize candidates to R0 resection? *Ann Surg Oncol* 2015; 22: 589-596.
- [7] Zhang H, Deng T, Liu R, Bai M, Zhou L, Wang X, Li S, Wang X, Yang H, Li J, Ning T, Huang D, Li H, Zhang L, Ying G and Ba Y. Exosome-delivered EGFR regulates liver microenvironment to promote gastric cancer liver metastasis. *Nat Commun* 2017; 8: 15016.
- [8] Pardoll DM. The blockade of immune checkpoints in cancer immunotherapy. *Nat Rev Cancer* 2012; 12: 252-264.
- [9] Bockorny B and Pectasides E. The emerging role of immunotherapy in gastric and esophageal adenocarcinoma. *Future Oncol* 2016; 12: 1833-1846.
- [10] Bonotto M, Garattini SK, Basile D, Ongaro E, Fanotto V, Cattaneo M, Cortiula F, Iacono D,

Treg exhibit different infiltrating status in hepatic metastases in AGC

- Cardellino GG, Pella N, Fasola G, Antonuzzo L, Silvestris N and Aprile G. Immunotherapy for gastric cancers: emerging role and future perspectives. *Expert Rev Clin Pharmacol* 2017; 10: 609-619.
- [11] Muro K, Chung HC, Shankaran V, Geva R, Catenacci D, Gupta S, Eder JP, Golan T, Le DT, Burtness B, McRee AJ, Lin CC, Pathiraja K, Lunceford J, Emancipator K, Juco J, Koshiji M and Bang YJ. Pembrolizumab for patients with PD-L1-positive advanced gastric cancer (KEYNOTE-012): a multicentre, open-label, phase 1b trial. *Lancet Oncol* 2016; 17: 717-726.
- [12] Shitara K, Özgüroğlu M, Bang YJ, Di Bartolomeo M, Mandalà M, Ryu MH, Fornaro L, Olesiński T, Caglevic C, Chung HC, Muro K, Goekkurt E, Mansoor W, McDermott RS, Shacham-Shmueli E, Chen X, Mayo C, Kang SP, Ohtsu A and Fuchs CS; KEYNOTE-061 investigators. Pembrolizumab versus paclitaxel for previously treated, advanced gastric cancer or gastro-oesophageal junction cancer (KEYNOTE-061): a randomized, open-label, controlled, phase 3 trial. *Lancet* 2018; 392: 123-133.
- [13] Gu L, Chen M, Guo D, Zhu H, Zhang W, Pan J, Zhong X, Li X, Qian H and Wang X. PD-L1 and gastric cancer prognosis: a systematic review and meta-analysis. *PLoS One* 2017; 12: e0182692.
- [14] Sakaguchi S, Yamaguchi T, Nomura T and Ono M. Regulatory T cells and immune tolerance. *Cell* 2008; 133: 775-787.
- [15] Li Z, Li D, Tsun A and Li B. FOXP3+ regulatory T cells and their functional regulation. *Cell Mol Immunol* 2015; 12: 558-565.
- [16] Togashi Y, Shitara K and Nishikawa H. Regulatory T cells in cancer immunosuppression - implications for anticancer therapy. *Nat Rev Clin Oncol* 2019; 16: 356-371.
- [17] Zhu F, Yi G, Liu X, Zhu F, Zhao A, Wang A, Zhu R, Chen Z, Zhao B, Fang S, Yu X, Lin R, Liang R, Li D, Zhao W, Zhang Z, Guo W, Zhang S, Ge S, Fan X, Zhao G and Li B. Ring finger protein 31-mediated atypical ubiquitination stabilizes forkhead box P3 and thereby stimulates regulatory T-cell function. *J Biol Chem* 2018; 293: 20099-20111.
- [18] Hori S, Nomura T and Sakaguchi S. Control of regulatory T cell development by the transcription factor Foxp3. *Science* 2003; 299: 1057-1061.
- [19] Fontenot JD, Gavin MA and Rudensky AY. Foxp3 programs the development and function of CD4+CD25+ regulatory T cells. *Nat Immunol* 2003; 4: 330-336.
- [20] Yu N, Li X, Song W, Li D, Yu D, Zeng X, Li M, Leng X and Li X. CD4(+)CD25(+)CD127 (low/-) T cells: a more specific Treg population in human peripheral blood. *Inflammation* 2012; 35: 1773-1780.
- [21] Perrone G, Ruffini PA, Catalano V, Spino C, Santini D, Muretto P, Spoto C, Zingaretti C, Sisti V, Alessandrini P, Giordani P, Cicetti A, D'Emidio S, Morini S, Ruzzo A, Magnani M, Tonini G, Rabitti C and Graziano F. Intratumoural FOXP3-positive regulatory T cells are associated with adverse prognosis in radically resected gastric cancer. *Eur J Cancer* 2008; 44: 1875-1882.
- [22] Haas M, Dimmler A, Hohenberger W, Grabenbauer GG, Niedobitek G and Distel LV. Stromal regulatory T-cells are associated with a favourable prognosis in gastric cancer of the cardia. *BMC Gastroenterol* 2009; 9: 65.
- [23] McShane LM, Altman DG, Sauerbrei W, Taube SE, Gion M and Clark GM. Reporting recommendations for tumour MARKer prognostic studies (REMARK). *Br J Cancer* 2005; 93: 387-391.
- [24] Wang Y, Zhu C, Song W, Li J, Zhao G and Cao H. PD-L1 expression and CD8+ T cell infiltration predict a favorable prognosis in advanced gastric cancer. *J Immunol Res* 2018; 4180517.
- [25] Kang YK, Boku N, Satoh T, Ryu MH, Chao Y, Kato K, Chung HC, Chen JS, Muro K, Kang WK, Yeh KH, Yoshikawa T, Oh SC, Bai LY, Tamura T, Lee KW, Hamamoto Y, Kim JG, Chin K, Oh DY, Minashi K, Cho JY, Tsuda M and Chen LT. Nivolumab in patients with advanced gastric or gastro-oesophageal junction cancer refractory to, or intolerant of, at least two previous chemotherapy regimens (ONO-4538-12, ATTRACTION-2): a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet* 2017; 390: 2461-2471.
- [26] Nomi T, Sho M, Akahori T, Hamada K, Kubo A, Kanehiro H, Nakamura S, Enomoto K, Yagita H, Azuma M and Nakajima Y. Clinical significance and therapeutic potential of the programmed death-1 ligand/programmed death-1 pathway in human pancreatic cancer. *Clin Cancer Res* 2007; 13: 2151-2157.
- [27] Liu X, Xu D, Huang C, Guo Y, Wang S, Zhu C, Xu J, Zhang Z, Shen Y, Zhao W and Zhao G. Regulatory T cells and M2 macrophages present diverse prognostic value in gastric cancer patients with different clinicopathologic characteristics and chemotherapy strategies. *J Transl Med* 2019; 17: 192.
- [28] Tanaka A and Sakaguchi S. Regulatory T cells in cancer immunotherapy. *Cell Res* 2017; 27: 109-118.
- [29] Joyce JA and Fearon DT. T cell exclusion, immune privilege, and the tumor microenvironment. *Science* 2015; 348: 74-80.
- [30] Lee HE, Chae SW, Lee YJ, Kim MA, Lee HS, Lee BL and Kim WH. Prognostic implications of type and density of tumour-infiltrating lympho-

Treg exhibit different infiltrating status in hepatic metastases in AGC

- cytes in gastric cancer. *Br J Cancer* 2008; 99: 1704-1711.
- [31] Xing X, Guo J, Ding G, Li B, Dong B, Feng Q, Li S, Zhang J, Ying X, Cheng X, Guo T, Du H, Hu Y, Zhou T, Wang X, Li L, Li Q, Xie M, Li L, Gao X, Shan F, Li Z, Jia S, Wen X, Wang J and Ji J. Analysis of PD1, PDL1, PDL2 expression and T cells infiltration in 1014 gastric cancer patients. *Oncoimmunology* 2017; 7: e1356144.
- [32] Ward-Hartstonge KA and Kemp RA. Regulatory T-cell heterogeneity and the cancer immune response. *Clin Transl Immunology* 2017; 6: e154.
- [33] Wing JB, Tanaka A and Sakaguchi S. Human FOXP3+ regulatory T cell heterogeneity and function in autoimmunity and cancer. *Immunity* 2019; 50: 302-316.
- [34] Ying L, Yan F, Meng Q, Yu L, Yuan X, Gantier MP, Williams BRG, Chan DW, Shi L, Tu Y, Ni P, Wang X, Chen W, Zang X, Xu D and Hu Y. PD-L1 expression is a prognostic factor in subgroups of gastric cancer patients stratified according to their levels of CD8 and FOXP3 immune markers. *Oncoimmunology* 2018; 7: e1433520.
- [35] Mao FY, Kong H, Zhao YL, Peng LS, Chen W, Zhang JY, Cheng P, Wang TT, Lv YP, Teng YS, Fu XL, Liu YG, Wu XL, Hao CJ, You N, Luo P, Yu PW, Zou QM, Guo G and Zhuang Y. Increased tumor-infiltrating CD45RA-CCR7- regulatory T Cell subset with immunosuppressive properties foster gastric cancer progress. *Cell Death Dis* 2017; 8: e3002
- [36] Nagase H, Takeoka T, Urakawa S, Morimoto-Okazawa A, Kawashima A, Iwahori K and Takiguchi S. ICOS+ Foxp3+ TILs in gastric cancer are prognostic markers and effector regulatory T cells associated with *Helicobacter pylori*. *Int J Cancer* 2017; 140: 686-695