Original Article Correlations between stemness indices for hepatocellular carcinoma, clinical characteristics, and prognosis

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Abstract: Recent studies have shown that cancer stem cells (CSCs) are involved in the occurrence and development of hepatocellular carcinoma (HCC). However, potential mechanisms for this have not yet been elucidated. We constructed a model based on the Progenitor Cell Biology Consortium database to generate stemness indices. We then utilized RNA-seq data and clinical information from the Cancer Genome Atlas (CGA) and International Cancer Genome Consortium (ICGC) for model predictions and verification. An mRNA gene expression-based stemness index (mRNAsi) and a DNA methylation-based stemness index (mDNAsi) were both calculated through one-class logistic regression. By applying univariate Cox regression analysis, we found that the mRNAsi and the mDNAsi correlated significantly with overall survival. Functional prediction analyses were used to characterize implicated genes and their degree of involvement as network hubs through protein-protein interaction analysis, and Spearman's rank correlation coefficient test was used to assess the relationship between hub genes and indices for stemness. The mRNAsi values for CGA and ICGC carcinoma samples correlated significantly with negative clinical characteristics and overall survival, whereas gene and protein-protein interaction analyses revealed that SNAP25, KPT19, GABBR1, and EPCAM were negatively associated with clinical mDNAsi scores. Collectively, the data suggest that our new stemness model based on related genes may predict patient prognoses.

Keywords: Hepatocellular carcinoma, cancer stem cell, mRNAsi, mDNAsi, prognosis

Introduction

Hepatocellular carcinoma (HCC) is the sixth most common cancer, and the third most frequent cause of cancer death [1, 2]. Although surgical resection is the most promising treatment during early stages, the 5-year survival rate for HCC is less than 20%, mainly due to limited treatment efficacy for advanced HCC [3-6]. Due to the absence of clinical symptoms during early stages, most patients with HCC are diagnosed at advanced stages, with poor prognoses [4, 7]. As the pathogenesis of HCC is unclear, it is essential to explore the mechanisms of HCC development and progression so that early diagnosis and targeted treatments can be achieved.

The theory of cancer stem cells (CSCs) indicated to researchers that malignant solid tumors

contained a highly heterogeneous population of CSCs [8, 9]. CSCs represent the small number of cancer cells that can self-replicate as stem cells and support both tumor growth and tumor characteristics, just as normal stem cells can renew and maintain organs and tissues [10, 11]. Growing evidence suggests that CSCs play crucial roles in tumorigenesis, metastasis, recurrence, and resistance to both radiation and chemotherapy [12-15]. However, how CSCs maintain malignant tumors, including potential molecular mechanisms and signaling pathways involved, is not clear.

Recent studies have shown the potential for CSCs to be targets of cancer therapy [16, 17]. Yang et al. [18] found that HGF/c-Met promoted the enrichment of renal CSCs, which could be a mechanism for metastasis and recurrence to leverage as a new target for treating renal cell

carcinoma. Additional evidence has suggested that clarifying the source of CSCs may offer novel methods for individualized cancer treatment [19], but for HCC, little is known about the source, expression patterns, and molecular mechanisms involved.

In this study, we constructed a model based on stem-cell sequencing data from the Progenitor Cell Biology Consortium (PCBC) to predict stem cell-associated molecular-feature (*stemness*) indices for HCC samples. We then utilized a Cancer Genome Atlas (CGA) cohort to make predictions and validated our findings through an independent dataset, analyzing relationships between stemness indices, clinical characteristics, and overall patient survival. Furthermore, we evaluated genes based on stemness and performed protein-protein interaction (PPI) analyses to identify potential hub genes in signaling networks, which could provide CSCspecific targets for the treatment of HCC.

Methods

Data processing

The following databases were used: the Cancer Genome Atlas (CGA, https://cancergenome. nih.gov/) database, International Cancer Genome Consortium (ICGC, https://icgc.org/) database, Progenitor Cell Biology Consortium (PCBC, https://progenitorcells.org/frontpage) database, and the STRING (https://string-db. org/) database. Expression data (syn2701943) for pluripotent stem-cell samples, including embryonic stem cells (ESCs) and induced pluripotent stem cells (iPSCs), were downloaded from the PCBC using the R package synapse (v0.6.61). RNA-Seq data and clinical follow-up information were downloaded concurrently for 424 samples in the CGA, and 203 samples in the ICGC database, the latter used as a validation set. Clinical follow-up information included gender, age, pathology grading, Tumor Node Metastasis (TNM) staging, hepatitis B virus (HBV) and hepatitis C virus (HCV) infection status, immune-cell score, and iCluster typing. Data were processed as previously described [20-22].

Calculation of a stemness index based on the one-class logistic regression (OCLR) method

To predict and calculate stemness indices, ESC and iPSC expression data from the PCBC data-

base were evaluated using the OCLR method, and this method was performed as previously described [23]. There were 78 total ESC and iPSC samples with 12,998 mRNA expression profiles of genes per sample. For DNA methylation-based stemness index (mDNAsi) calculations, 99 samples with 219 stem-cell probe signatures in the PCBC database were divided into two groups, of which 44 samples were stem-cell samples, and 55 samples were nonstem-cell samples. Expression profiles and methylation data were centralized by the average value for each sample. Both the mRNA gene expression-based stemness index (mRNAsi), and the mDNAsi weight vector for each gene were then calculated by the OCLR method using the R package gel net (v1.2.1).

For the expression profiles and methylation data from the CGA and ICGC HCC samples, we calculated Spearman's rank correlation coefficient for each sample gene with the model's gene weight vector. We then performed a linear conversion to the Spearman's rank correlation coefficient obtained from the sample. Conversion methods were used to identify a Spearman's rank correlation coefficient for each sample, minus a minimum, which was then divided by the maximum value. The values obtained became the mRNAsi and mDNAsi for each sample, with a distribution range of stemness indices between zero and one.

Analysis and screening of differentially expressed genes

We used the R package DESeq2 (v1.24.0) to perform a differential analysis of the RNA-seq data from mRNAsi-high/mDNAsi-high and mR-NAsi-low/mDNAsi-low samples and then determined differentially expressed genes (DEGs) between the these two samples [23, 24]. These DEGs were then filtered according to a threshold false discovery rate (FDR) < 0.05 and |log-2FC| > 1 [22].

Enrichment analysis of gene function and pathways

A Kyoto Encyclopedia of Genes and Genomes (KEGG) (https://www.kegg.jp/ or https://www. genome.jp/kegg/) pathway-enrichment analysis was used to determine networks for metabolic, signaling, and other molecular interactions [25]. Gene Ontology (GO) functional enrichment analysis is an international standard classification system for gene function, including Biological Process (BP), Cellular Components (CC), and Molecular Function (MF) [26]. Through these categories, gene functions were defined and described accordingly [27]. To examine possible functions related to the stemness indices of DEGs, we performed KE-GG-pathway analyses and GO functional enrichment analyses with mRNAsi and mDNAsi through the R package cluster Profiler (v3.14.0). The significance level was defined as FDR < 0.05.

PPI network analysis

The STRING database was used to analyze the PPI network of DEGs, and the degree algorithm in the cytoHubba plugin of Cityscape software was applied to identify hub genes. PPI network analysis is commonly used to elucidate molecular bases of diseases, and provides detailed knowledge about protein interactions which can be used to determine underlying factors for prevention, diagnosis, and treatment of disease [28]. This PPI analysis also provides information about the genes and proteins shared in a disease and describes their interactions.

Statistics

A t-test was used for continuous variables. Statistical analyses were performed using IBM SPSS 24.0 software. All statistical tests were two-sided, and the significance level was defined as P < 0.05. Spearman's rank correlation coefficients were used to analyze correlations between two variables. Kaplan-Meier survival curves were generated, after stratification of the data, and compared using a log-rank test.

Results

Relationships between indices for mRNA expression- and DNA methylation-based stemness and clinical characteristics

We first investigated relationships between stem-cell index distributions and clinical characteristics by ranking HCC samples according to their mRNAsi and mDNAsi values, and then tested whether any clinical features correlated with low- or high-stemness values. Clinical features for tumor samples included gender, age, pathology staging, HBV or HCV infection status, immune-cell score, and iCluster typing. mRNAsi and mDNAsi exponential distributions for the CGA HCC samples are shown in **Figure 1A**, **1B**, respectively. The mRNAsi exponential distributions for the ICGC HCC samples are shown in **Figure 1C**.

We then performed linear-regression testing for the above characteristics. As shown in **Table 1**, the mRNAsi values for the CGA HCC samples correlated significantly with pathology grading, HBV infection, immune-cell score, and iCluster typing. mDNAsi values were significantly associated with immune-cell score and iCluster typing. Analysis of the ICGC HCC validation cohort dataset indicated that mRNAsi values were significantly associated with TNM stage (**Table 2**).

For the CGA HCC samples, mRNAsi scores were significantly higher in HBV-infected samples compared to uninfected samples (Figure 2A), and in iCluster3 samples compared to iCluster1 and iCluster2 samples (Figure 2B, 2C). In addition, samples from HCC patients with late pathology grading showed higher mRNAsi scoring (Figure 2D, 2E). iCluster typing also significantly correlated with mDNAsi values. mDNAsi scores were lowest in iCluster1 samples and highest in iCluster3 samples, while HBV infection and pathology grading had no obvious correlations with mDNAsi scores (Figure S1A-E). In the ICGC HCC samples, TNM stage was significantly associated with mRNAsi values in an increasing manner for the first three stages, but no relationship between HBV infection and mRNAsi values was found (Figure 2F, 2G). Immune scores were negatively correlated with both mRNAsi and mDNAsi scores in the CGA HCC samples (Figures 2H, S1F). Furthermore, there was a positive correlation between the mRNAsi and mDNAsi scores in the CGA HCC samples (R = 0.30, Figure 2I). Taken together, higher stemness-index values in HCC samples correlated with worse clinical characteristics, demonstrating that stemness indices may characterize tumor progression.

Correlations between HCC stemness indices and overall survival

To better understand the relationship between stemness-index values and prognosis, we first compared overall survival with clinical features such as gender, age, pathology grading, iCluster typing, immune-cell scores, and HBV or HCV



Figure 1. Exponential mRNAsi and mDNAsi distributions based on clinical characteristics of HCC samples. A. The mRNAsi distribution based on the clinical characteristics of the CGA HCC samples. B. The mDNAsi distribution based on the clinical characteristics of the CGA HCC samples. C. The mRNAsi distribution based on the clinical characteristics of the ICGC HCC samples. HCC: hepatocellular carcinoma; mRNAsi: mRNA gene expression-based stemness index; mDNAsi: DNA methylation-based stemness index; CGA: Cancer Genome Atlas; ICGC: International Cancer Genome Consortium.

	Percent/Average	p (mRNAsi)	p (mDNAsi)
Sex	Male: 65.6%	0.80	0.12
	Female: 34.4%		
Age	60.13	0.47	0.23
	G1		
Grade	G2	3.71E-05	0.67
	G3		
HBV	23.00%	3.04E-04	0.47
HCV	18.6%	0.76	0.15
Immune score	166.12	1.2E-03	0.018
	iCluster1		
iCluster	iCluster2	1.29E-07	5.64E-31
	iCluster3		

Table 1. Correlations between CGA HCC samplemRNAsi and mDNAsi values and clinical features andmolecular typing

Table 2. Correlations between ICGC HCC samplemRNAsi values and clinical features and molecular typing

	Percent/Average	p (mRNAsi)	
Sex	Male: 75.4%	0.12	
	Female: 24.6%		
Age	67	0.2	
	I: 16.3%		
TNM stage	II: 47.3%	9.09E-03	
	III: 29.1%		
	IV: 7.4%		
HBV	23.00%	0.81	
HCV	18.6%	0.64	
Immune score	166.12	0.97	

infection status, among the HCC samples. The results demonstrated that gender, age, HCV infection, pathology grade, and iCluster typing were not significantly correlated with overall survival (Figure 3A-E). Significant survival differences were identified in these samples based on HBV infection and immune-cell scoring. The high immune-score group demonstrated higher survival compared to the low immunescore group (Figure 3F), and patient survival for HBV-infected samples was lower than for noninfected samples (Figure 3G). Importantly, overall survival for the mRNAsi-low group was higher compared to the mRNAsi-high group (Figure 3H), and survival for the mDNAsi-high group was higher compared to the mDNAsi-low group (Figure S2A). Samples grouped by mRNAsi and mDNAsi values were then assessed

according to iCluster typing (**Table 3**). Only iCluster2 samples grouped according to mRNAsi values showed a significant survival difference (**Figures 3I-K** and <u>S2B-D</u>). However, this evaluation was complicated by the small sample sizes and short survival times of the three groups.

We next used the above analysis on the ICGC HCC dataset, and validated that there were no correlations between survival and gender, age, immune-cell scoring, and HBV infection (Figure 4A-D). Overall survival was lower in the presence of HCV infection (Figure 4E) and was lower with increasing TNM stage (Figure 4F). For the stemness indices, overall survival of the mRNAsi-low group was longer than for the mRNAsi-high group (Figure 4G). Collectively, these data

suggest that higher mRNAsi values correlate with poorer survival, which may offer new criteria for determining prognoses in patients with HCC.

Screening for stem cell-related genes in HCC samples

Based on the above results, we further analyzed differentially expressed genes (DEGs) in the CGA HCC cohort. After threshold-filtering, 136 DEGs were identified, with 110 genes downregulated and 26 genes upregulated in the mRNAsi-high group compared to the mRNAsi-low group (Figure 5A). In the mDNAsi set, 569 DEGs were identified, with 546 genes downregulated and 23 genes upregulated in the mDNAsi-high group compared to mDNAsilow group (Figure 5B). Common DEGs between the mRNAsi and mDNAsi sets were APOBEC3C. C1orf116, GABBR2, and TFCP2L1. In the filtered ICGC cohort, 190 DEGs were observed, with 16 genes downregulated and 174 genes upregulated in the mRNAsi-high group compared with the mRNAsi-low group (Figure 5C). The only DEG in common between the CGA and ICGC cohorts was upregulated ALDH1A3. These differences in gene expression reflect indexvalue differences (high versus low) among HCC samples.

Functional analyses of differentially expressed genes

To explore the possible roles of these DEGs in the CGA HCC samples for tumorigenesis, me-

C _{0.55} В _{0.55} A _{0.55 1} 0.45 0.45 0.45 .iskNAsi 0.32 .18NAsi 0.32 HBV Hegative Negative Positive 0.25 0.25 0.25 0.15 L iCluster 0.15 iCluster 1 0.15 HBV Negative Positive 2 2 3 3 1 E _{0.55} D F 0.55 1.00 0.45 0.45 0.75 . ISV 0.35 0.35 iskNAsi 0.32 HBV .is 0.50 MRNAsi 0.25 Hegative Negative Positive 📄 0.25 0.25 0.00 0.15 Grade G1 0.15 G2 Ġ3 TNM Stage I Π Ш iv Grade G1 G2 G3 G Н 0.6 Т $R^2 = 0.09$ p= 0.00012 1.00 $R^2 = 0.064$ p = 0.0012 0 0 0 4000 3000 2000 1000 0 0.5 0.75 .is 0.4 0.3 mRNAsi 0.20 0.2 0.25 -1000-0.1 -2000æ ۰ 0.00 HBV Negative Positive 0.2 0.3 0.5 0.4 0.2 0.3 0.4 mRNAsi 0.5 mRNAsi

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Figure 2. Relationships between the mRNAsi values and clinical characteristics. A. mRNAsi scores were higher in samples infected with HBV in the CGA HCC samples. B. mRNAsi scores were higher in iCluster3 compared to scores in iCluster1 and iCluster2 for all CGA HCC samples. C. mRNAsi scores were highest in iCluster3, and lowest in iCluster1, for CGA HCC samples without HBV infection. D. mRNAsi scores were highest for grade 3, and lowest for grade 1, for all CGA HCC samples. E. mRNAsi scores were highest for grade 3, and lowest for grade 1, for CGA HCC samples. E. mRNAsi scores were highest for grade 3, and lowest for grade 1, for CGA HCC samples. E. mRNAsi scores were highest for grade 3, and lowest for grade 1, for CGA HCC samples. E. mRNAsi scores were highest increased with TNM staging only for the first three staging levels. G. There was no significant correlation between mRNAsi values and HBV infection for ICGC HCC samples. H. Immune scores were inversely related to mRNAsi scores in the CGA HCC samples. I. mRNAsi and mDNAsi values were positively correlated for CGA HCC samples. *P < 0.05. mRNAsi: mRNA gene expression-based stemness index; HBV: hepatitis B virus; CGA: Cancer Genome Atlas. ICGC: International Cancer Genome Consortium.

tastasis, recurrence, and drug resistance, we performed Gene Ontology (GO) functional analyses for the 136 mRNAsi DEGs that showed no significant enrichments (FDR < 0.05). GO functional annotations for the 569 DEGs identified using mDNAsi values indicated that the "metal ion transmembrane transporter activity" pathway, and "monovalent inorganic action transmembrane transporter activity" were significantly enriched (Figure 5D). KEGG-pathway enrichment analysis of mRNAsi DEGs found no significantly enriched pathways (FDR < 0.05), while KEGG-pathway enrichment analysis of mDNAsi DEGs demonstrated that the "cAMP signaling pathway" was significantly enriched (Figure 5E). GO functional annotations of the 190 mRNAsi-grouped DEGs in the ICGC HCC samples showed significant enrichment for "extracellular matrix structural constituents" and "glycosaminoglycan binding" (Figure 5F). KEGG-pathway enrichment analysis of mRNAsi DEGs in the ICGC HCC samples found three significantly enriched pathways, including the PI3K-AKT signaling pathway (Figure 5G). Taken together, we found that the DEGs from highand low-stemness indices were enriched for pathways related to tumor formation, metastasis and recurrence. These enrichments should aid in our understanding of tumorigenesis mechanisms and may guide the development of targeted therapies for HCC.

PPI and hub-gene analyses of differentially expressed genes

To identify hub genes related to these stemness-index values, PPI analyses were performed based on DEGs. Ten genes (SNAP25, EPCAM, CALB2, SOX2, KRT19, GABBR1, MUC1, AFP, GRIN1, and GAD1) were identified as hub genes among mDNAsi DEGs (Figure 6A). Correlation analyses between these hub genes and mDNAsi values were performed, and significant negative correlations were found be-

tween SNAP25, KPT19, GABBR1, and EPCAM and these index values (**Figure 6B-E**) while six other hub genes had no prominent connections (**Figure 6F-K**). These findings suggested that tumor stemness-index values could be utilized as new prognosticators for patients with HCC. In addition, SNAP25, KPT19, GABBR1, and EPCAM could be key genes involved in HCC tumorigenesis, metastasis and recurrence.

Discussion

In this study, we found that stemness indices were related to pathology grading, HBV infection, immune-cell score, iCluster typing, and TNM staging. Consistent with our HCC findings, several other studies have demonstrated that stemness-index values were associated with pathological characteristics in many cancers. For example, Pan et al. [24] observed that mRNAsi values in bladder cancer increased as tumor stage increased, with T3 staging having the most stem-cell characteristics. Lower mR-NAsi scores also had better overall survival and treatment outcomes. Another recent study reported that the expression of CSC marker OCT4 was correlated with poor differentiation, tumor size, and N stage in patients with rectal cancer, and concluded that OCT4 could be an independent prognostic biomarker [29]. These results suggest that stemness-index scores could play an essential role in defining tumor progression.

An increasing number of studies have also linked stemness indices with patient prognosis. In our study, we constructed a novel model to predict overall survival in HCC patients, and observed a relationship between HCC stemness-index values, clinical characteristics, and survival. Our results showed that patients with worse clinical phenotypes usually possessed higher stemness-index scores. Furthermore, patients with higher mRNAsi scores tended to



Figure 3. Kaplan-Meier survival analyses of the CGA HCC samples. A. Survival probabilities for sex-grouped samples showed no significant differences. B. Survival probabilities for age-grouped samples showed no significant differences. C. Survival probabilities for samples grouped by HCV infection showed no significant differences. D. Survival probabilities for samples grouped by pathology grading showed no significant differences. E. Survival probabilities for iCluster-grouped samples showed no significant differences. F. The high immune-score group showed higher survival probabilities than the low immune-score group. G. The survival probabilities for HBV-infected samples was lower than for HBV-negative samples. H. The survival probabilities for the mRNAsi-low group were higher than for the mRNAsi-high group in iCluster1 were not significantly different. J. The survival probabilities for the mRNAsi-low group were higher than those for the mRNAsi-high group in iCluster2. K. The survival probabilities for the mRNAsi-low group were higher than those for the mRNAsi-high group in iCluster3 were not significantly different. *P < 0.05. mRNAsi: mRNA gene expression-based stemness index; CGA: Cancer Genome Atlas; HCV: hepatitis C virus.

	mRNAsi						
iCluster	HR			Cut p			
	HR	Lower	Upper	р	Cut	p (Cut)	
C1	1.80	0.96	3.40	6.77E-02	0.37	0.23	
C2	3.61	1.20	10.84	2.21E-02	0.35	0.08	
C3	1.60	0.79	3.23	0.19	0.43	0.77	
All	1.68	1.11	2.53	0.0145	0.38	0.08	
	mDNAsi						
iCluster	HR			Cut p			
	HR	Lower	Upper	р	Cut	p (Cut)	
C1	0.69	0.37	1.31	0.26	0.12	0.87	
C2	0.52	0.20	1.34	1.75E-01	0.20	0.67	
C3	1.85	0.91	3.76	0.09	0.30	0.48	
All	0.62	0.39	0.98	4.04E-02	0.12	0.51	

Table 3. Hazard ratio (HR) analysis and optimal

 thresholds for the CGA stem-cell index grouping

have lower overall survival. However, we observed the opposite result in mDNAsi-grouped samples which may have been due to small sample size. Consistent with our findings, previous studies have shown that increased expression of CSC marker CD133 confers a poor prognosis for invasive breast cancer [30], and Qin et al. [31] reported that an mRNAsi score was an independent prognostic factor in lung squamous-cell carcinoma. Additionally, Zhao et al. [32] reported that TBX21 was both a patient prognosis predictor and a CSC maintenance driver via the TBX21-IL-4 pathway in lung adenocarcinoma, suggesting that TBX21 may serve as a novel predictive biomarker and therapeutic target. Lastly, Lian et al. determined that a prognostic signature based on mRNAsi values may predict sonic-hedgehog medulloblastoma prognosis and such a signature could be a potential biomarker for informing treatmentoptions in clinical practice [23]. These findings demonstrate that stemness-index scores may be reliable predictors for HCC prognoses.

Our investigation also examined DEGs between stemness indices and their possible functions. The functional pathways for "metal ion transmembrane transporter activity", "monovalent inorganic action transmembrane transporter activity", "cAMP signaling pathway", "extracellular matrix structural constituent", "glycosaminoglycan binding", and "PI3K/AKT signaling pathway" were significantly enriched. Previously, Roberto et al. found that synergistic inhibition of HCC and liver CSC proliferation could occur by targeting RAS/RAF/MAPK and WNT/β-Catenin pathways [33], and microRNA-28-5p was reported to regulate liver CSC expansion via the IGF-1 pathway [34]. Moreover, Si et al. identified the miR219/E-cadherin axis as a potential therapeutic target against liver CSCs and as a predictor for sorafenib treatment in HCC patients [35]. Based on these results, stem cell-related DEGs correlated with a variety of tumorigenesis-related pathways, offering mechanistic insight into how to target CSC maintenance in patients with HCC.

Finally, we performed PPI analyses to identify HCC hub genes based on the identified DEGs. We found that SNAP25, KPT19, GABBR1, and EPCAM were significantly correlated with mDNAsi towards negative outcomes. Similar to our results, prior work showed that SENP1 activity sustained CSCs in hypoxic HCC [36]. Li et al. observed that activated STAT3 played a pivotal role in maintaining stemness in HCC CSCs [37], and Ritu et al. found that miR-26b-5p imparted metastatic properties and helped maintain Ep+ CSCs via HSPA8. Thus, miR-26b-5p and HSPA8 may serve as molecular targets for selectively eliminating the Ep+ CSC population in human HCC [38]. Collectively, these findings further support the idea that identifying hub genes related to CSCs in HCC could inform about its pathogenesis and indicate novel targets for HCC therapy.

The stem-cell index results of this study may advance the development of objective diagnostic tools for quantitating HCC stemness, prognosticating judgments, and treatment-response predictions. This provides new directions for clinical treatment, especially for developing personalized treatment protocols. Further research is needed to explore stemness indexrelated genes and their roles in tumor proliferation and metastasis to advance this promising area.

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Figure 4. Kaplan-Meier survival analyses of ICGC HCC samples. A. Survival probabilities for samples grouped by gender showed no significant differences. B. Survival probabilities for age-grouped samples showed no significant differences. C. Survival probabilities for samples grouped by immune-cell scores showed no significant differences. D. Survival probabilities for samples grouped by HBV infection showed no significant differences. E. The survival probabilities for HCV-infected samples were lower than for HCV-negative samples. F. Survival probabilities were lower with increasing TNM staging. G. The survival probabilities for the mRNAsi-low group were higher than those for the mRNAsi-high group. *P < 0.05. mRNAsi: mRNA gene expression-based stemness index. ICGC: International Cancer Genome Consortium. TNM: Tumor Node Metastasis.



Figure 5. Volcano plots, and GO and KEGG annotation maps, for differentially expressed genes (DEGs). A. 110 genes were downregulated, and 26 genes were upregulated, in the mRNAsi-high group compared to the mRNAsi-low group for CGA HCC samples. B. 546 genes were downregulated, and 23 genes were upregulated, in the mDNAsi-high group compared to the mDNAsi-low group for CGA HCC samples. C. 16 genes were downregulated, and 174 genes were upregulated in the mDNAsi-high group compared to the mRNAsi-low group for CGA HCC samples. C. 16 genes were downregulated, and 174 genes were upregulated in the mDNAsi-high group compared to the mRNAsi-low group for ICGC HCC samples. D. The GO annotation map for the top 5 (out of 569) mDNAsi DEGs in the CGA HCC samples. E. The KEGG annotation map for the top 5 mDNAsi DEGs in the CGA HCC samples. F. The GO annotation map for the top 5 (out of 569) mRNAsi DEGs in the ICGC HCC samples. G. The KEGG annotation map for the top 3 mRNAsi DEGs in the ICGC HCC samples. *FDR < 0.05. GO: Gene Ontology; KEGG: Kyoto Encyclopedia of Genes and Genomes; DEGs: differentially expressed genes.

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Figure 6. PPI analyses and correlations between hub genes and mDNAsi scores. A. Protein interactions between the 10 hub genes; darker colors indicate higher scores that represent more network involvement and significance. B. EPCAM had a significant negative correlation with mDNAsi scores. C. GABBR1 has a significant negative correlation with mDNAsi scores. D. KRT19 had a significant negative correlation with mDNAsi scores. E. SNAP25 has a significant negative correlation with mDNAsi scores. F. AFP was not significantly correlated with mDNAsi scores. G. CALB2 was not significantly correlated with mDNAsi scores. H. GAD1 was not significantly correlated with mDNAsi scores. J. MUC1 was not significantly correlated with mDNAsi scores. K. SOX2 was not significantly correlated with mDNAsi scores. *P < 0.05. mDNAsi: DNA methylation-based stemness index; PPI: protein-protein interaction.

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Disclosure of conflict of interest

None.

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Figure S1. Relationship between mDNAsi values and clinical characteristics. A. There were no significant correlations between mDNAsi scores and HBV infections in CGA HCC samples. B. mDNAsi values were highest in iCluster3 and lowest in iCluster1 for CGA HCC samples without HBV infections. C. There were no significant correlations between mDNAsi scores and pathology grading in CGA HCC samples without HBV infections. D. mDNAsi scores were highest in iCluster3 and lowest in iCluster1 for all CGA HCC samples. E. There were no significant correlations between mDNAsi values and pathology grading for all CGA HCC samples. F. Immune scores were inversely related to mDNAsi values.



Figure S2. Kaplan-Meier survival analyses for CGA HCC samples. A. Survival probabilities for the mDNAsi-high group were higher than those of the mDNAsi-low group for all GCA HCC samples. B. Survival probabilities for the mDNAsi-low group and the mDNAsi-high group in iCluster1 were not significantly different. C. Survival probabilities for the mDNAsi-low group and the mDNAsi-high group in iCluster2 were not significantly different. D. Survival probabilities for the mDNAsi-low group and the mDNAsi-high group in iCluster3 were not significantly different.