Original Article miR-21-5p inhibits inflammation injuries in LPS-treated H9c2 cells by regulating PDCD4

Jianhua Xue^{1*}, Jiajia Liu^{1*}, Bo Xu⁵, Junbo Yu¹, Aixian Zhang², Lili Qin³, Chun Liu⁴, Yang Yang¹

Departments of ¹Trauma Center, ²General Practice Medicine, ³Endoscopic Center, ⁴Emergency Medicine, Affiliated Hospital of Nantong University, Nantong, Jiangsu Province, China; ⁵Department of Orthopaedics, Qidong Hospital of Traditional Chinese Medicine, Nantong, Jiangsu Province, China. ^{*}Equal contributors and co-first authors.

Received May 14, 2021; Accepted August 31, 2021; Epub October 15, 2021; Published October 30, 2021

Abstract: Objectives: To explore the expression levels and the potential regulatory mechanism of miR-21-5p in LPS-treated H9c2 cells. Methods: The secretions of the inflammatory cytokines induced by LPS in H9c2 cells were evaluated using ELISA. We used RT-RCR and western blot to measure the relative mRNA and protein expression levels in LPS-treated H9c2 cells. CCK-8 and EdU assays showed the viability and proliferation profiles of the H9c2 cells. TUNEL assays demonstrated the apoptotic behaviors of the H9c2 cells, and a luciferase reporter analysis was used to investigate the interactions between miR-21-5p and programmed cell death protein 4 (PDCD4). Results: LPS induced damage to the H9c2 cells by reducing the cell viability and down-regulating miR-21-5p. On the other hand, miR-21-5p overexpression inhibited the LPS-induced inflammatory damage in the H9c2 cells. Moreover, PDCD4 was verified as a downstream target gene of miR-21-5p, and its expression rescued the miR-21-5p effect in the LPS-treated H9c2 cells. Conclusion: Our findings suggest that miR-21-5p inhibits the LPS-induced progression of sepsis in H9c2 cells. Additionally, PDCD4 is a downstream target gene of miR-21-5p, and both molecules serve as potential therapeutic targets for heart sepsis patients.

Keywords: miR-21-5p, programmed cell death protein 4, LPS, heart sepsis, H9c2 cells

Introduction

A syndrome of physiologic, pathological, and biochemical abnormalities, sepsis is induced by infection [1]. It is a major public health concern and a leading cause of death in hospitals worldwide [2, 3]. Severe sepsis and septic shock lead to organ dysfunctions by elevating oxidative stress and exacerbating inflammation in organs like the brain, heart, kidneys, liver, and lungs [4]. It has been reported that the mortality of sepsis is related to cardiac dysfunction [5, 6]. However, the specific regulatory mechanism with the progression of septic cardiac dysfunction remain elusive.

Lipopolysaccharide (LPS) can activate the excessive production of pro-inflammatory mediators, leading to myocardial damage, heart failure, and even death [7]. Since one initial feature of sepsis is acute inflammation, H9c2 cells have been used to mock how cardio-

myocytes respond to such a situation by exposing them to LPS [8].

Non-coding RNAs that contain 21 to 24 nucleotides are defined as microRNAs (miRNAs). miR-NAs regulate gene expression during translation. They have been studied as regulators in various physiological and inflammatory processes in human diseases [9]. Specifically, three miRNAs (miR-21-5p, miR-125-5p, and miR-18b) have been shown to act as major regulators in immune-inflammatory responses [10-12]. Furthermore, numerous studies have revealed miRNAs as modulators of sepsisinduced damage in inflamed organs [13-15]. Specifically, miR-21-5p suppresses inflammation and cell apoptosis in the injured brain microvascular endothelial barrier, as well as the progression of aged intracerebral hemorrhages [12, 16]. In this study, we explored the role miR-21-5p plays in LPS-treated H9c2 cells.

Table 1.	The sec	nuences	of the	miRNA	constructs
10010 1	1110 000	1001000		11111 (1 1/ 1	0011011010

-	
miR-21-5p mimics NC	5'-GUGUAACACGUCUAUACGCCCA-3'
miR-21-5p mimics	5'-UAGCUUAUCAGACUGAUGUUGA-3'
miR-21-5p inhibitors NC	5'-UCACAACCUCCUAGAAAGAGU-3'
miR-21-5p inhibitors	5'-UCAACAUCAGUCUGAUAAGCUA-3'

Programmed cell death protein 4 (PDCD4) is a tumor suppressor that is normally down-regulated in various cancers [17, 18]. Many studies have found that PDCD4 participates in many cellular functions, especially transcription, translation, and proliferation [19]. Furthermore, miR-21 was found to target the PDCD4 gene at the 3'-UTR region [20], and the inhibition of miR-21 blocks the LPS-induced down-regulation of PDCD4 in periodontitis, suggesting that miR-21 is vital in inflammatory responses and cancers by targeting PDCD4 [21]. Silencing miR-21-5p, a member of the miR-21 family, will improve paclitaxel (PTX) resistance and inhibit PTX-resistant processes in breast cancer cells partly by targeting PDCD4 [22]. Nevertheless, the underlying regulatory mechanisms between miR-21-5p and PDCD4 in septic H9c2 cells remain to be elucidated, which is precisely the topic of this article.

In this study, a model of LPS-induced sepsis in H9c2 cells was established. Subsequently, we measured the expression levels and clarified the regulatory mechanisms of miR-21-5p in septic H9c2. The results show that miR-21-5p promotes cell viability and inhibits apoptosis in LPS-treated H9c2 cells. Our study provides experimental evidence to clarify the molecular mechanisms of sepsis in H9c2 cells, as well as information about potential therapeutic targets against sepsis.

Materials and methods

Cell treatments and transfection

The cardiomyoblast culture medium (H9c2 cells; ATCC, Manassas, VA, USA) was DMEM (Gibco Laboratories, USA) with 10% FBS (Gibco Laboratories, USA) and the antibiotics 1% penicillin and streptomycin (Gibco Laboratories, USA), which were incubated at 5% CO_2 , 37°C.

The H9c2 cells were respectively co-cultured with a concentration (0.5, 1, 2 μ g/mL) of LPS for 24 h (Sigma, Missouri, USA). Cells or cell supernatants were extracted and collected for the analysis.

The miR-21-5p mimics, the miR-21-5p mimic negative controls (NC), the miR-21-5p inhibitor negative controls (NC), the pcDNA3.1 vector, and the pcDNA3.1-programmed cell death 4 (PDCD4) were purchased from Shanghai GenePharma Co., Ltd. The

cells were seeded in a 6-well plate with a density of 10^5 cells/ml and pre-incubated for 12 h at 37°C, before being transfected with 50 nM miR-21-5p mimics or miR-21-5p mimics NC, 2 µg pc DNA3.1 or pcDNA3.1-PDCD4, 50 nM miR-21-5p inhibitors or miR-21-5p inhibitors NC with 20 nmol/I Lipofectamine[®] 2000 (Invitrogen; Thermo Fisher Scientific, Inc.) then incubated at 37°C with 5% CO₂ for 48 h. The cells were harvested 48 h after the transfection for the subsequent experiments. The sequences of the miRNA constructs are shown in **Table 1**.

ELISA assay

The supernatants of the LPS-treated H9c2 cells were harvested and the IL-1 β , TNF- α , and IL-6 releases were measured ELISA Kits (TaKaRa, Dalian, China).

CCK-8 cell viability assays

The cells were seeded into a 96-well plate and treated with LPS at 37° C and 5% CO₂ for 24 h. The cell proliferation was measured CCK-8 Kits (Abcam, Cambridge, UK). The optical density values were measured at 450 nm using an ultraviolet spectrophotometer (Thermo Fisher Scientific, Inc.).

EdU assays

The cells were seeded into a 96-well plate and treated with LPS at 37°C and 5% CO_2 for 24 h. The cell proliferation was measured using EdU assays (Solarbio, Beijing, China). The EdU was added for 2 h. After fixation, 1× Apollo was added to stain the cells for 0.5 h without light. Next, we DAPI stained the nuclei for 5 min. Finally, the cells were viewed under an inverted fluorescence microscope (Olympus IX51, Japan).

TUNEL assays

The apoptosis of the LPS-treated cells was assessed using a commercial Cell Death Detection kit (Roche, Germany). The labelled

	•
Rno-miR-21-5pforward	GACAAGCTTGCGGCCGCCCTTTAGGAGCATTATGAGCAT
Rno-miR-21-5p reverse	ATCCTCTAGAGTCGACGAAGGTCAAGTAACAGTCATAC
Rno-PDCD4 forward	TTGAGCACGGAGATACGAAC
Rno-PDCD4 reverse	GTCCCGCAAAGGTCAGAAAG
Rno-U6 forward	CTCGCTTCGGCAGCACACATA
Rno-U6 reverse	ACGCTTCACGAATTTGCGTGTC
Rno-β-actin forward	GTTGGAGCAAACATCCCCCA
Rno-β-actin reverse	CGCGACCATCCTCCTCTTAG

 Table 2. The primers for qPCR

Note: The $2^{-\Delta\Delta CT}$ relative quantification method was used to quantify the qPCR data. PDCD4: programmed cell death protein 4.

cells were viewed using a fluorescence microscope (Olympus IX51, Japan).

Western blotting

The protein extracted from the H9c2 cells was lysed in an ice-cold RIPA buffer plus PMSF on ice (Beyotime Institute of Biotechnology, Shanghai, China).

The primary antibodies were anti-Bax (Abcam, 1:1,000), anti-Bcl-2 (Abcam, 1:500), anti-procleaved Caspase-3 (Proteintech, 1:500), anticleaved Caspase-3 (Proteintech, 1:500), antipro-cleaved Caspase-9 (Proteintech, 1:300), anti-cleaved Caspase-9 (Proteintech, 1:300), anti- β -actin (Abcam, 1:1,000), and anti-PDCD4 antibody (Proteintech, 1:1,000).

qRT-PCR assays

The RNA was isolated using TRIzol (Takara, Otsu, Japan) and reversed transcribed into cDNA using Superscript II reverse transcriptase (Thermo Fisher Scientific, Inc.). The conditions for the RT reaction were incubation at 25°C for 10 min, 50°C for 15 min, and heated to 85°C for 5 min to terminate the reaction. A real-time PCR machine (Bio-Rad) was used to conduct the quantitative gPCR (Table 2). The gRT-PCR was subsequently performed using SYBR Green I Super mix (Takara, Otsu, Japan) according to the manufacturer's protocol. The thermocycling conditions for the RT-qPCR were 95°C for 10 min, followed by 35 cycles at 95°C and at 60°C for 1 min. The RT-qPCR results were analyzed using the $2^{-\Delta\Delta CT}$ method.

Statistical analysis

All the results are presented as the means \pm sd from three independent experiments. The com-

parisons were determined using unpaired/paired Student's *t* tests, and Bonferroni, Tukey, two-way ANOVA. P<0.05 was considered to indicate a statistically significant difference.

Results

The miR-21-5p expressions in the LPS-treated H9c2 cells

H9c2 cells were co-cultured with low, medium, and high concentrations of LPS for 24 h. respectively. The LPS-induced secretions of the inflammatory cytokines in the H9c2 cells, including TNF- α , IL-1 β , and IL-6, were evaluated using ELISA, and the results demonstrated dose-dependent secretions of these cytokines with respect to the LPS concentrations (Figure 1A-C). Moreover, the CCK-8 and EdU assays revealed that the viability and proliferation of the H9c2 cells were decreased (Figure 1D, 1E). Furthermore, the number of TUNELpositive H9c2 cells increased in a dose-dependent manner concerning the LPS concentrations, indicating the apoptosis of the H9c2 cells is promoted by LPS (Figure 1F).

Moreover, the Bcl-2 expressions were found to drop, but the Bax, Cleaved caspase-3, and Cleaved caspase-9 levels were elevated in the LPS-treated H9c2 cells in an LPS-dosedependent manner (**Figure 1G**). Notably, the miR-21-5p levels were down-regulated in the H9c2 cells (**Figure 1H**). Judging from the data, LPS can induce damage in H9c2 cells by reducing the cell viability and promoting apoptosis, and the power of such an induction was positively correlated to the LPS concentration. Moreover, miR-21-5p was downregulated by LPS in H9c2 cells.

Overexpression of miR-21-5p suppresses the inflammatory damage in H9c2 cells

To investigate the functions of miR-21-5p in the LPS-treated H9c2 cells, miR-21-5p mimics and NC mimics were transfected into these cells. The qRT-PCR results indicated that the miR-21-5p mimics can upregulate the miR-21-5p contents in the H9c2 cells (**Figure 2A**), while downregulating the secretions of TNF- α , IL-1 β ,



miR-21-5p inhibits the inflammatory damage caused by LPS in H9c2 cells

Figure 1. The expression of miR-21-5p is down-regulated in LPS-treated H9c2 cells. H9c2 cells were treated with different concentrations of LPS for 24 h, respectively. A-C. The secretions of the cytokines were determined using ELISA. D. The cell viability was measured using CCK-8 assays. E. We used EdU assays to analyze the cell proliferation (400×). F. We used TUNEL assays to analyze the apoptosis (400×). G. We used Western blot to analyze the expression levels of the apoptosis-associated proteins. H. We used qRT-PCR to test the expressions of miR-21-5p. The values are the mean \pm sd, *P<0.05, **P<0.01 vs. the control, n=3 per group.

and IL-6 (Figure 2B-D). Moreover, the CCK-8 and EdU assays showed that the miR-21-5p mimics can significantly promote the viability and proliferation of the LPS-treated H9c2 cells (Figure 2E, 2F). Then, the proportion of the TUNEL-positive cells dropped in the group transfected with the miR-21-5p mimics (Figure 2G). Moreover, the miR-21-5p mimics elevated the expression level of Bcl-2 and inhibited the expressions of Bax, Cleaved caspase-3, and Cleaved caspase-9 (Figure 2H). All these results confirmed that the overexpression of miR-21-5p can inhibit LPS-induced inflammatory damage in H9c2 cells.

PDCD4 is a downstream target gene of miR-21-5p

To explore the regulatory mechanisms of miR-21-5p in the LPS-treated H9c2 cells, we used miRwalk and TargetScan to search the target genes and found 14 intersection target genes (Figure 3A). To specify the target gene that interacts with miR-21-5p, the miR-21-5p mimics and the miR-21-5p inhibitors were transfected into the LPS-treated H9c2 cells. Among the 14 target genes, the qRT-PCR analysis found that PDCD4 exhibited an expression pattern that was significantly affected by the mimics and the miR-21-5p inhibitors in the LPS-treated H9c2 cells (Figure 3A). Subsequently, we predicted the potential interaction sites between miR-21-5p and PDCD4 (Figure 3B) and performed a luciferase reporter analysis to verify them. The results demonstrated that the luciferase activity at the 3'UTR region in PDCD4 dropped significantly when the H9c2 cells were co-transfected with the PDCD4-WT and miR-21-5p mimics. However, the luciferase activity of the PDCD4-Mut didn't change significantly between the groups of the miR-21-5p mimics and the control (Figure **3C**). Moreover, the miR-21-5p inhibitors were found to increase the expressions of the PDCD4 mRNA and protein in the H9c2 cells (Figure 3D, 3E). In general, PDCD4 was confirmed to be a downstream target gene of miR- 21-5p and its expression is negatively correlated with the miR-21-5p expression.

Pc-PDCD4 reversed the effects of the miR-21-5p inhibitors in the LPS-treated H9c2 cells

Next, we performed a rescue experiment to investigate the functions of the PDCD4 and miR-21-5p mimics in the LPS-treated H9c2 cells. First, the PDCD4 expressions were upregulated in the H9c2 cells transfected with pc-PDCD4 compared with the pc-DNA3.1 group (Figure 4A). Then, the pc-PDCD4 was found to reverse the down-regulation of TNF- α , IL-1 β , and IL-6 by the overexpression of miR-21-5p (Figure 4B-D). Moreover, the CCK-8 and EdU assays revealed that the PDCD4 up-regulation could reverse the promotive effects of the miR-21-5p mimics on the LPS-treated H9c2 cells (Figure 4E, 4F). Meanwhile, pc-PDCD4 increased the proportion of the TUNEL-positive cells, indicating that the overexpression of PDCD4 aggravated the apoptosis of the H9c2 cells that were treated with LPS and transfected with the miR-21-5p mimics (Figure 4G). Finally, a western blot analysis proved that pc-PDCD4 inhibited the up-regulation of Bcl-2 and reversed the inhibitory effects on the expressions of Bax, Cleaved caspase-3, and Cleaved caspase-9 seen in the miR-21-5p group (Figure 4H). In general, miR-21-5p can inhibit the septic process, and such an effect was rescued by the PDCD4 overexpression of PDCD4 in the LPS-treated H9c2 cells.

Discussion

LPS is an essential component of the outer membrane of gram-negative bacteria [23]. As a molecule responsible for the systemic inflammatory response, it is routinely used as a sepsis model in studies [15, 24]. LPS-induced sepsis is defined as a systemic inflammatory response process that is related to many cardiac diseases [25]. Moreover, LPS is involved in various molecular metabolism mechanisms, mitochondrial dysfunction, apoptosis, and inflammation [15]. In this study, LPS was used

miR-21-5p inhibits the inflammatory damage caused by LPS in H9c2 cells



Figure 2. The overexpression of miR-21-5p inhibits inflammatory injuries in LPS-treated H9c2 cells. miR-NC and miR-21-5p were transfected in the LPS-treated H9c2 cells. A. A qRT-PCR analysis of the expression of miR-21-5p. B-D. The cytokines were measured using ELISA. E. The cell viability was measured using CCK-8 assays. F. EdU assays were used to analyze the cell proliferation (400×). G. TUNEL assays were used to analyze the apoptosis (400×). H. Western blot was used to analyze the expression levels of the apoptosis associated proteins. The values are the mean ± sd, *P<0.05, **P<0.01, ***P<0.001 vs. LPS + miR-NC, n=3 per group.

miR-21-5p inhibits the inflammatory damage caused by LPS in H9c2 cells





miR-21-5p inhibits the inflammatory damage caused by LPS in H9c2 cells

Figure 4. The function of the miR-21-5p inhibitor is reversed by pc-PDCD4 in the LPS-treated H9c2 cells. A. The protein levels of PDCD4 were measured using western blot. The values are the mean \pm SD, **P<0.01 vs. pc-DNA3.1. n=3 per group. The LPS-treated H9c2 cells transfected with miR-NC, miR-21-5p mimics or miR-21-5p mimics + pc-PDCD4. B-D. We used ELISA to measure the secretions of cytokines TNF- α , IL-1 β , and IL-6. **P<0.01 vs. miRNA-NC group, ##P<0.01 vs. miRNA-21-5p group. E. The cell viability was measured using CCK-8 assays. **P<0.01 vs. the miRNA-NC group, ##P<0.01 vs. the miRNA-NC group, #P<0.05 vs. miRNA-21-5p group. G. We used a TUNEL assay to analyze the apoptosis (400×). **P<0.01 vs. the miRNA-NC group, #P<0.05 vs. miRNA-21-5p group. G. We used a TUNEL assay to analyze the apoptosis (400×). **P<0.01 vs. the miRNA-NC group, ##P<0.01 vs. the mean \pm SD. **P<0.01 vs. the miRNA-NC group, #P<0.05, ##P<0.01 vs. miRNA-21-5p group. n=3 per group.



Figure 5. A schematic diagram of relationship between LPS and miR-21-5p. The overexpression of miR-21-5p cannot protect the cells from LPS-induced damage in the LPS-treated H9c2 cells by promoting the viability of the cells, inhibiting the cell apoptosis or the secretions of the inflammatory-related cytokines.

to induce damage in H9c2 cells and to create an experimental model of sepsis *in vitro*. Our results indicated that the viability of the H9c2 cells was significantly inhibited by LPS. On the other hand, the proportion of apoptotic cells and the expressions of the apoptosis-related proteins increased when LPS was administered in the cell model. Moreover, LPS contributed to the overexpressions of IL-6, IL-1 β , and TNF- α in the H9c2 cells, suggesting that an experimental model of sepsis was established.

MiRNAs are regulatory molecules that are vital in disease diagnosis and treatment, and they usually interact with the target gene at the 3' untranslated region (3'-UTR) [12, 26]. Previous studies have shown that miRNAs participate in regulating cell metabolism and are involved in LPS pathophysiology [21, 27, 28]. Therefore, miRNAs may become new potential therapeutic targets for LPS-induced sepsis. Specifically, human NLRP3 inflammasomes are found to be inhibited by miR-21 knockdown in septic shock [29]. Moreover, the inhibition of miR-21 suppresses snail expression and prevents LPSinduced podocyte injuries [30].

As a member of the miR-21 family, miR-21-5p has been shown to be involved in many diseases [31, 32]. Although scientists have conductive extensive research about the inflammation inhibition of miR-21-5p across lots of diseases, the upstream regulatory mechanisms of miR-21-5p in sepsis have remained almost

unknown until now. In this paper, we confirmed that LPS downregulates the expression of miR-21-5p in H9c2 cells while upregulating the concentrations of the inflammatory cytokines and the apoptosis-related proteins. When LPStreated H9c2 cells are transfected with the miR-21-5p mimics, the expressions of the inflammatory cytokines and the apoptosisrelated proteins dropped, and the cell viability increased. Therefore, miR-21-5p is verified to inhibit inflammation in LPS-treated H9c2 cells.

With the bioinformatics analysis, PDCD4 was predicted as one of the potential targets of miR-21-5p. Moreover, PDCD4 has been shown to significantly suppress tumor growth and progression. Moreover, miR-21-5p has been found to play a role in disease development by targeting PDCD4 [20]. In our study, PDCD4 was experimentally confirmed as a target of miR-21-5p, and its expression was found to be negatively correlated with miR-21-5p expression in H9c2 cells. Furthermore, pc-PDCD4 was found to partially antagonize the inhibitory effects on the inflammation of the miR-21-5p mimics in LPS-treated H9c2 cells (**Figure 5**).

However, there are still some limitations to the present study. The *in vivo* animal model experiments were lacking. The roles of miR-21-5p and PDCD4 in the sepsis animal model should be further determined. We will further confirm our conclusions by performing animal model experiments in our future studies.

In summary, the overexpression of miR-21-5p can protect cells from LPS-induced damage by promoting cell viability and inhibiting apoptosis and the secretions of the inflammation-related cytokines. Meanwhile, the relationship between miR-21-5p and PDCD4 in LPS-treated H9c2 cells is clarified *in vitro*, and a novel therapeutic target (PDCD4) for sepsis is proposed. Such findings should be verified further by *in vivo* studies.

Acknowledgements

This work was supported by the Science and Technology Project of Nantong City (JCZ19116) and the Science and Technology Project of Nantong Municipal Health Commission (MA2-020023).

Disclosure of conflict of interest

None.

Address correspondence to: Yang Yang, Department of Trauma Center, Affiliated Hospital of Nantong University, No. 20 Xisi Road, Chongchuan District, Nantong 226001, Jiangsu Province, China. Tel: +86-0513-85052222; Fax: +86-0513-850-52222; E-mail: Yangyang286228@163.com; Chun Liu, Department of Emergency Medicine, Affiliated Hospital of Nantong University, No. 20 Xisi Road, Chongchuan District, Nantong 226001, Jiangsu Province, China. Tel: +86-0513-85052232; E-mail: Liuchun.nt@163.com

References

- [1] Singer M, Deutschman CS, Seymour CW, Shankar-Hari M, Annane D, Bauer M, Bellomo R, Bernard GR, Chiche JD, Coopersmith CM, Hotchkiss RS, Levy MM, Marshall JC, Martin GS, Opal SM, Rubenfeld GD, van der Poll T, Vincent JL and Angus DC. The third international consensus definitions for sepsis and septic shock (Sepsis-3). JAMA 2016; 315: 801-810.
- [2] Hartman ME, Linde-Zwirble WT, Angus DC and Watson RS. Trends in the epidemiology of pediatric severe sepsis*. Pediatr Crit Care Med 2013; 14: 686-693.
- [3] Haileselassie B, Mukherjee R, Joshi AU, Napier BA, Massis LM, Ostberg NP, Queliconi BB, Monack D, Bernstein D and Mochly-Rosen D. Drp1/Fis1 interaction mediates mitochondrial dysfunction in septic cardiomyopathy. J Mol Cell Cardiol 2019; 130: 160-169.
- [4] Shankar-Hari M, Bertolini G, Brunkhorst FM, Bellomo R, Annane D, Deutschman CS and

Singer M. Judging quality of current septic shock definitions and criteria. Crit Care 2015; 19: 445.

- [5] Antonucci E, Fiaccadori E, Donadello K, Taccone FS, Franchi F and Scolletta S. Myocardial depression in sepsis: from pathogenesis to clinical manifestations and treatment. J Crit Care 2014; 29: 500-511.
- [6] Court O, Kumar A, Parrillo JE and Kumar A. Clinical review: myocardial depression in sepsis and septic shock. Crit Care 2002; 6: 500-508.
- [7] Chen J, Lai J, Yang L, Ruan G, Chaugai S, Ning Q, Chen C and Wang DW. Trimetazidine prevents macrophage-mediated septic myocardial dysfunction via activation of the histone deacetylase sirtuin 1. Br J Pharmacol 2016; 173: 545-561.
- [8] Xu J, Lin C, Wang T, Zhang P, Liu Z and Lu C. Ergosterol attenuates LPS-induced myocardial injury by modulating oxidative stress and apoptosis in rats. Cell Physiol Biochem 2018; 48: 583-592.
- [9] Ambros V. The functions of animal microRNAs. Nature 2004; 431: 350-355.
- [10] Sun X, Icli B, Wara AK, Belkin N, He S, Kobzik L, Hunninghake GM, Vera MP, Blackwell TS, Baron RM and Feinberg MW. MicroRNA-181b regulates NF-κB-mediated vascular inflammation. J Clin Invest 2012; 122: 1973-1990.
- [11] Martinez-Fierro ML, Garza-Veloz I, Gutierrez-Arteaga C, Delgado-Enciso I, Barbosa-Cisneros OY, Flores-Morales V, Hernandez-Delgadillo GP, Rocha-Pizaña MR, Rodriguez-Sanchez IP, Badillo-Almaraz JI, Ortiz-Rodriguez JM, Castañeda-Miranda R, Solis-Sanchez LO and Ortiz-Castro Y. Circulating levels of specific members of chromosome 19 microRNA cluster are associated with preeclampsia development. Arch Gynecol Obstet 2018; 297: 365-371.
- [12] Ouyang Y, Li D, Wang H, Wan Z, Luo Q, Zhong Y, Yin M, Qing Z, Li Z, Bao B, Chen Z, Yin X and Zhu LQ. MiR-21-5p/dual-specificity phosphatase 8 signalling mediates the anti-inflammatory effect of haem oxygenase-1 in aged intracerebral haemorrhage rats. Aging Cell 2019; 18: e13022.
- [13] Funahashi Y, Kato N, Masuda T, Nishio F, Kitai H, Ishimoto T, Kosugi T, Tsuboi N, Matsuda N, Maruyama S and Kadomatsu K. miR-146a targeted to splenic macrophages prevents sepsis-induced multiple organ injury. Lab Invest 2019; 99: 1130-1142.
- [14] Yao Y, Xu K, Sun Y, Tian T, Shen W, Sun F, Yuan W, Wu H, Chen G, Yuan L, Zhang W, Lu M and Lei M. MiR-215-5p inhibits the inflammation injury in septic H9c2 by regulating ILF3 and LRRFIP1. Int Immunopharmacol 2020; 78: 106000.

- [15] An R, Feng J, Xi C, Xu J and Sun L. miR-146a attenuates sepsis-induced myocardial dysfunction by suppressing IRAK1 and TRAF6 via targeting ErbB4 expression. Oxid Med Cell Longev 2018; 2018: 7163057.
- [16] Ge X, Huang S, Gao H, Han Z, Chen F, Zhang S, Wang Z, Kang C, Jiang R, Yue S, Lei P and Zhang J. miR-21-5p alleviates leakage of injured brain microvascular endothelial barrier in vitro through suppressing inflammation and apoptosis. Brain Res 2016; 1650: 31-40.
- [17] Manirujjaman M, Ozaki I, Murata Y, Guo J, Xia J, Nishioka K, Perveen R, Takahashi H, Anzai K and Matsuhashi S. Degradation of the tumor suppressor PDCD4 is impaired by the suppression of p62/SQSTM1 and autophagy. Cells 2020; 9: 218.
- [18] Fehler O, Singh P, Haas A, Ulrich D, Müller JP, Ohnheiser J and Klempnauer KH. An evolutionarily conserved interaction of tumor suppressor protein Pdcd4 with the poly(A)-binding protein contributes to translation suppression by Pdcd4. Nucleic Acids Res 2014; 42: 11107-11118.
- [19] Zhao M, Zhu N, Hao F, Song Y, Wang Z, Ni Y and Ding L. The regulatory role of non-coding RNAs on programmed cell death four in inflammation and cancer. Front Oncol 2019; 9: 919.
- [20] Lu Z, Liu M, Stribinskis V, Klinge CM, Ramos KS, Colburn NH and Li Y. MicroRNA-21 promotes cell transformation by targeting the programmed cell death 4 gene. Oncogene 2008; 27: 4373-4379.
- [21] Zhou W, Su L, Duan X, Chen X, Hays A, Upadhyayula S, Shivde J, Wang H, Li Y, Huang D and Liang S. MicroRNA-21 down-regulates inflammation and inhibits periodontitis. Mol Immunol 2018; 101: 608-614.
- [22] Tao L, Wu YQ and Zhang SP. MiR-21-5p enhances the progression and paclitaxel resistance in drug-resistant breast cancer cell lines by targeting PDCD4. Neoplasma 2019; 66: 746-755.
- [23] Zhang K, Liu J, You X, Kong P, Song Y, Cao L, Yang S, Wang W, Fu Q and Ma Z. P2X7 as a new target for chrysophanol to treat lipopolysaccharide-induced depression in mice. Neurosci Lett 2016; 613: 60-65.

- [24] Chen L, Liu P, Feng X and Ma C. Salidroside suppressing LPS-induced myocardial injury by inhibiting ROS-mediated PI3K/Akt/mTOR pathway in vitro and in vivo. J Cell Mol Med 2017; 21: 3178-3189.
- [25] Lyu Y, Jiang X and Dai W. The roles of a novel inflammatory neopterin in subjects with coronary atherosclerotic heart disease. Int Immunopharmacol 2015; 24: 169-172.
- [26] Hu YL, Wang H, Huang Q, Wang G and Zhang HB. MicroRNA-23a-3p promotes the perihematomal edema formation after intracerebral hemorrhage via ZO-1. Eur Rev Med Pharmacol Sci 2018; 22: 2809-2816.
- [27] Yang S, Wang Y, Gao H and Wang B. MicroR-NA-30a-3p overexpression improves sepsis-induced cell apoptosis in vitro and in vivo via the PTEN/PI3K/AKT signaling pathway. Exp Ther Med 2018; 15: 2081-2087.
- [28] Zhao J, Li X, Zou M, He J, Han Y, Wu D, Yang H and Wu J. miR-135a inhibition protects A549 cells from LPS-induced apoptosis by targeting Bcl-2. Biochem Biophys Res Commun 2014; 452: 951-957.
- [29] Xue Z, Xi Q, Liu H, Guo X, Zhang J, Zhang Z, Li Y, Yang G, Zhou D, Yang H, Zhang L, Zhang Q, Gu C, Yang J, Da Y, Yao Z, Duo S and Zhang R. miR-21 promotes NLRP3 inflammasome activation to mediate pyroptosis and endotoxic shock. Cell Death Dis 2019; 10: 461.
- [30] Wang S, Wang J, Zhang Z and Miao H. Decreased miR-128 and increased miR-21 synergistically cause podocyte injury in sepsis. J Nephrol 2017; 30: 543-550.
- [31] Moghaddam AS, Afshari JT, Esmaeili SA, Saburi E, Joneidi Z and Momtazi-Borojeni AA. Cardioprotective microRNAs: lessons from stem cell-derived exosomal microRNAs to treat cardiovascular disease. Atherosclerosis 2019; 285: 1-9.
- [32] Paiva RM, Zauli DAG, Neto BS and Brum IS. Urinary microRNAs expression in prostate cancer diagnosis: a systematic review. Clin Transl Oncol 2020; 22: 2061-2073.