

Review Article

Illustrative and historic cases of phenoconversion

Veronique Michaud^{1,2}, Pamela Dow¹, Jacques Turgeon^{1,2}

¹TRHC Precision Pharmacotherapy Research and Development Institute, 13485 Veterans Way, Suite 410, Orlando, FL 32827, USA; ²Université de Montréal, Faculty of Pharmacy, Montreal, Quebec, H3T 1J4, Canada

Received June 24, 2021; Accepted October 11, 2021; Epub December 15, 2021; Published December 30, 2021

Abstract: Intersubject variability in drug response, whether related to efficacy or toxicity, is well recognized clinically. Over the years, drug selection from our pharmacologic armamentarium has moved from providers' preferred choices to more personalized treatments as clinicians' decisions are guided by data from clinical trials. Since the advent of more accessible and affordable pharmacogenomic (PGx) testing, the promise of precise pharmacotherapy has been made. Results have accumulated in the literature with numerous examples demonstrating the value of PGx to improve drug response or prevent drug toxicity. Unfortunately, limited availability of reimbursement policies has dampened the enthusiasm of providers and organizations. The clinical application of PGx knowledge remains difficult for most clinicians under real-world conditions in patients with numerous chronic conditions and polypharmacy. This may be due to phenoconversion, a condition where there is a discrepancy between the genotype-predicted phenotype and the observed phenotype. This condition complicates the interpretation of PGx results and may lead to inappropriate recommendations and clinical decision making. For this reason, regulatory agencies have limited the transfer of information from PGx laboratories directly to consumers, especially recommendations about the use of certain drugs. This mini-review presents cases (mexiletine, propafenone, clopidogrel, warfarin, codeine, procainamide) from historical observations where drug response was modified by phenoconversion. The cases illustrate, from decades ago, that we are still in great need of advanced clinical decision systems that cope with conditions associated with phenoconversion, especially in patients with polypharmacy.

Keywords: Phenoconversion, genotype, phenotype, pharmacogenomics

Introduction

Phenoconversion is a phenomenon by which there is a mismatch between an individual's genotype-based predicted phenotype and the observed phenotype [1]. Phenoconversion can be caused by extrinsic factors such as environment, food, drugs, or patient- or disease-related factors [1-3]. A simple example is the genotype associated with an individual predicting hair color and the actual hair color observed on that person. Like phenoconversion, this mismatch between the observed phenotype and the genotype-predicted phenotype can be due to extrinsic factors, such as the use of hair coloring agents, or due to patient-specific conditions, such as aging.

There are numerous conditions associated with phenoconversion when predicting drug response (efficacy and toxicity), especially in patients with polypharmacy. This phenomenon

is not new and we purposely use older examples - several initiated from our research activities - to demonstrate how our knowledge has evolved and to show how difficult it has been to translate pharmacogenetic results into applied clinical interventions. This topic has recently gained interest with more laboratories interested in promoting pharmacogenomic (PGx) testing, as molecular biology technologies are now more readily accessible and affordable. This review presents examples of increased or decreased drug efficacy or toxicity due to phenoconversion (see **Table 1** for a summary of examples discussed).

Increased efficacy

In 1991, Duff *et al.* reviewed data supporting the use of selected drug combinations to enhance antiarrhythmic activity, specifically the mexiletine-quinidine combination [4]. They had previously found that the co-administration of

Cases of phenoconversion

Table 1. Examples of conditions and drugs susceptible to phenoconversion

Condition	Victim drug subjected to phenoconversion	Enzyme involved	Perpetrator drug causing phenoconversion	Mechanism	Refs
Increased efficacy	Mexiletine	CYP2D6	Quinidine or other potent CYP2D6 inhibitors such as paroxetine	Quinidine is a potent CYP2D6 inhibitor leading to increased plasma concentrations of mexiletine when the two drugs are co-administered. Improved efficacy at smaller doses of mexiletine and decreased side-effects (gastric irritation).	[4-11]
Increased efficacy	Propafenone	CYP2D6	Quinidine or other potent CYP2D6 inhibitors such as paroxetine	Quinidine is a potent CYP2D6 inhibitor leading to increased plasma concentrations of propafenone but decreased concentrations of its active 5-hydroxymetabolite. Due to different electrophysiological effects, this condition improves overall propafenone efficacy for the maintenance of sinus rhythm in patients with atrial fibrillation.	[12-15]
Decreased efficacy	Codeine	CYP2D6	Quinidine or other potent CYP2D6 inhibitors such as paroxetine	Codeine is a prodrug that needs to be activated into morphine to produce analgesic effects. Inhibition of CYP2D6 decreases morphine formation and codeine analgesic effects.	[16-22]
Decreased efficacy	Clopidogrel	CYP2C19	Omeprazole	Clopidogrel is a prodrug that needs to be activated mostly by CYP2C19 to inhibit platelet aggregation. Inhibition of CYP2C19 decreases clopidogrel efficacy.	[23-31]
Increased toxicity	Warfarin	CYP2C9/CYP3As	CYP2C9 and/or CYP3A4 inhibitors such as amiodarone, macrolide antibiotics, antifungals	Decreases CYP2C9 and/or CYP3A4 activities increases plasma levels of S-warfarin and/or R-warfarin, respectively. These inhibitions lead to greater inhibition of the Vitamin K epoxide reductase complex and increased risk of bleeding.	[38-52]
Decreased toxicity	Procainamide	NAT2/CYP2D6	Quinidine	Inhibition of CYP2D6 by potent inhibitors such as quinidine prevents the formation of N-oxidized, reactive toxic metabolites of procainamide.	[53-56]

CYP2D6, cytochrome P450 2D6; CYP2C19, cytochrome P450 2C19; CYP2C9, cytochrome P450 2C9; CYP3A, cytochrome P450 3A; NAT2, polymorphic N-acetyltransferase 2.

these two class I antiarrhythmic agents was more effective at suppressing spontaneous ventricular tachycardia with fewer side effects than high-dose monotherapy [5]. Duff *et al.* also conducted several electrophysiologic studies suggesting the potentiation of drug effects by various parameters, such as the prolongation of the refractory periods of extra stimuli and the prolongation of conduction into the dyskinetic zone of the ventricle [6].

In the 1980s, Turgeon and collaborators conducted several studies on the metabolism, disposition, and electrophysiologic effects of mexiletine [7-10]. Later, in 1991, they reported on the involvement of debrisoquine hydroxylase (CYP2D6) on the disposition of mexiletine and demonstrated different pharmacokinetic profiles between poor metabolizers (PMs) and extensive (EMs) metabolizers [11]. They also used quinidine to inhibit CYP2D6 and convert

EMs into PMs, demonstrating that inhibition of CYP2D6 could be associated with a 4-fold increase in mexiletine plasma levels under steady state conditions, as this CYP450 isoform contributes to 75% of the partial metabolic clearance of the drug. Hence, phenoconversion of EMs into PMs by the co-administration of quinidine could explain, at least in part, the potentiation of drug effects observed with the mexiletine-quinidine combination as reported by Duff *et al.* in the same year [4].

The involvement of debrisoquine 4-hydroxylase (CYP2D6) in the metabolism and disposition of propafenone was also well characterized in the 1980s [12]. Funck-Brentano *et al.* demonstrated how low-dose quinidine was able to convert CYP2D6 EMs into PMs when interacting with propafenone [13]. Propafenone is metabolized into an active 5-hydroxymetabolite; the electrophysiologic effects of propafenone include the

Cases of phenoconversion

blocking of sodium, calcium, and potassium channels, while the 5-hydroxymetabolite mostly exhibits electrophysiological effects through potent inhibition of sodium channels [12, 14]. Taking advantage of the more comprehensive and favorable electrophysiological effects of propafenone over its 5-hydroxymetabolite, O'Hara *et al.* demonstrated the superior efficacy of propafenone when combined with quinidine in patients with atrial fibrillation (CAQ-PAF study) [15]. Recurrence of atrial fibrillation was observed in 22 patients (n=23) with low propafenone levels (<1,000 ng/mL; mostly due to extensive and unblocked CYP2D6 metabolism), while 80% of patients with propafenone levels >1,500 ng/mL (associated with a genetically-determined or quinidine-induced PM phenotype) were in sinus rhythm at one year. In fact, phenoconversion induced by quinidine persisted for the entire study period (one year).

Decreased efficacy

In 1988, Dayer *et al.* demonstrated that codeine was bioactivated into morphine by the debrisoquine 4-hydroxylase (CYP2D6) [16]. In a double-blind randomized cross-over study, Dayer *et al.* also demonstrated that virtually no morphine was observed in PMs or after administration of quinidine to EMs [17]. In EMs, codeine significantly increased subjective (VAS) and objective (R-III reflex) pain thresholds in response to selective transcutaneous nerve stimulation, whereas no significant analgesia was detected after quinidine pretreatment in PMs. The Clinical Pharmacogenetics Implementation Consortium guidelines for selected opioid therapy clearly recommend choosing alternative treatments to codeine for analgesia in patients with a CYP2D6 PM genotype or phenotype (due to phenoconversion and inhibition of CYP2D6) [18]. We have reviewed and reported clinical cases of poor response in phenotypic PMs due to phenoconversion while being treated with codeine [19-21]. Further, in a study performed with data from more than 50,000 adults, we demonstrated the economic burden associated with opioid treatment in patients with polypharmacy causing inhibition of CYP2D6 [22].

Clopidogrel is another example of decreased efficacy due to phenoconversion. Clopidogrel is a prodrug that undergoes sequential oxida-

tions - mediated mostly by CYP2C19 and CYP3A5 - leading to the formation of 2-oxo-clopidogrel and its active metabolite (5-thiol clopidogrel) to produce antiplatelet effects [23]. In the IGNITE study, carrying a variant allele of CYP2C19 was associated with worse clinical outcomes in patients [24]. A meta-analysis conducted by Mega *et al.* demonstrated that the concomitant administration of clopidogrel and a proton pump inhibitor - especially the CYP2C19 mechanism-based inhibitor omeprazole - was associated with poor clinical outcomes [25]. We have shown that a chronic inflammatory status associated with type 2 diabetes causes a significant (two-fold) decrease in CYP2C19 activity, triggering a phenoconversion like phenomenon [3, 26]. Decreased plasma levels of the 5-thiolactive metabolite, as well as poor clinical response to clopidogrel, has been observed in patients with type 2 diabetes. However, these patients appear to respond well to other antiplatelet agents, such as prasugrel or ticagrelor, that do not require bioactivation by CYP2C19 [27-31].

Increased toxicity

Warfarin has a narrow therapeutic index and therefore the dose required to achieve therapeutic anticoagulation is marginally different to the dose that leads to over-anticoagulation. Additionally, the maintenance dose varies between different individuals and ranges from 0.5 mg/day to more than 10 mg/day [32]. Warfarin is a racemic mixture of two enantiomers: S-warfarin and R-warfarin. Most pharmacological activity resides at the level of S-warfarin, which is two- to five- times more potent than the R-isomer [33, 34]. Under steady-state conditions, R-warfarin predominates in the plasma of patients at concentrations approximately double those of S-warfarin [34, 35]. However, the pharmacokinetics and pharmacodynamics of warfarin are modulated by numerous factors, including age, sex, genetic variants, illnesses, and drug interactions [36, 37].

CYP2C9 is principally responsible for the metabolism of S-warfarin, while CYP3A4/5, CYP1A2, and CYP2C19 are responsible for the metabolism of R-warfarin. Two of the variant alleles identified for CYP2C9 (*3 and *6) are associated with a loss of activity, whereas *2, *4, *5, and *11 are associated with weaker

Cases of phenoconversion

enzyme activities [38-42]. Scordo *et al.* have demonstrated that the free clearance of S-warfarin showed large variability in subjects with *CYP2C9**1/*1, *1/*2 or *1/*3 genotypes, such that it becomes impossible to predict the dose requirement in these subjects [43]. Only in a fraction of their subjects (8.6%), *i.e.* those with a *2/*2, *2/*3 or *3/*3, could a precise warfarin dose requirement be derived.

The contribution of other genetic polymorphisms in the Vitamin K epoxide reductase complex (*VKORC1*) or in *CYP4F2* (rs2108622) involved in the metabolism of Vitamin K1 are associated with warfarin maintenance dose requirements [44, 45]. However, several extrinsic factors including food, such as green vegetables with high Vitamin K content, or concomitant administration of drugs metabolized by or inhibiting *CYP2C9*, have been associated with phenoconversion and modulation of warfarin dose requirements. We conducted a study to determine the value of genotype-derived (*CYP2C9*, *VKORC1*) or phenotype-derived (using losartan as a *CYP2C9* probe drug) determination of warfarin dose requirement in patients with polypharmacy (receiving 11±4 drugs daily). In multivariate analyses, the dose-adjusted international normalized ratio (INR) at day four explained 31% of variability observed in warfarin doses at day 14, whereas genotypic measures (*CYP2C9-VKORC1*) contributed only 6.5%. Some, but not all, studies have shown an association between bleeding and genetic factors, such as *CYP2C9* polymorphisms. They clearly identify the role of extrinsic factors and phenoconversion on the risk of warfarin toxicity [46-52].

Decreased toxicity

Most pharmacogenomics textbooks have demonstrated the value of pharmacogenomics using a positive association with the production of antinuclear antibodies as indicators of procainamide-induced systemic lupus erythematosus. In a study conducted by Woosley *et al.*, it was demonstrated that the rate at which procainamide induces antinuclear antibodies - and therefore lupus erythematosus - was dependent on the acetylator (*NAT2*) genotype; slow acetylators required on average 12±5 months to develop lupus vs. 48±22 months in rapid acetylators [53]. Uetrecht *et al.* demon-

strated that a rapid acetylator phenotype and extensive formation of N-acetyl-procainamide was associated with a lower incidence of procainamide-induced lupus while the N-oxidation of procainamide lead to the formation of a reactive metabolite, causing revertants in the Ames test [54]. However, the N-acetylation status was not protective of procainamide-induced toxicity, but was predictive of the exposure time associated with toxicity. Hence, the real question remains unanswered: which enzymatic system is responsible for the formation of the toxic N-oxidized reactive metabolites and can their formation be prevented?

We then undertook drug metabolism studies to characterize enzymes involved in the N-oxidation of procainamide. We demonstrated that this metabolic pathway was mediated by *CYP2D6* and conducted pharmacokinetic studies in subjects receiving procainamide either alone or following the concomitant administration of quinidine [55, 56]. No N-oxidized metabolites could be measured in the urine of *CYP2D6* PM subjects; however, they were present in the urine of EMs. Coadministration of quinidine with procainamide caused a phenoconversion of EMs to PMs, modulated procainamide pharmacokinetics, and prevented the formation of the N-oxidized metabolites. Therefore, we postulated that patients who did not present with lupus upon exposure to procainamide in the studies conducted by Woosley *et al.* were PMs of *CYP2D6* - as those were observed in both rapid and slow acetylator groups - and that the concomitant administration of low-dose quinidine with procainamide could prevent the formation of the toxic metabolite due to the phenoconversion of patients into a PM phenotype.

Conclusion

Pharmacogenomic testing has evolved significantly in the last 20 years, with most laboratories using next-generation sequencing techniques. Pharmacogenomic results are now highly reliable and deemed appropriate to predict an expected phenotype. Additionally, testing costs have been reduced significantly, making pharmacogenomics more affordable and attractive for precise pharmacotherapy. In the past decade, polypharmacy has reached epidemic proportions with >40% of older adults taking

Cases of phenoconversion

five or more prescription drugs a day and nearly 20% take more than 10 drugs a day [57]. Under these conditions, clinicians must have access to advanced clinical decision support systems that allow consideration of phenoconversion to make appropriate medication choices within a drug regimen. Recent studies have demonstrated the value of endogenous markers, such as 4 β -hydroxycholesterol (CYP3As) or metabolomic products (CYP2D6), to determine a patient's actual phenotype [58, 59]. This strategy may represent a way forward to clinically understand patients overall drug metabolism capacities, response, and phenoconversion.

Disclosure of conflict of interest

None.

Address correspondence to: Dr. Jacques Turgeon, TRHC Precision Pharmacotherapy Research and Development Institute, 13485 Veterans Way, Orlando, FL 32827, USA. Tel: (407) 454-9933; E-mail: jturgeon@trhc.com

References

- [1] Klomp SD, Manson ML, Guchelaar HJ and Swen JJ. Phenoconversion of cytochrome P450 metabolism: a systematic review. *J Clin Med* 2020; 9: 2890.
- [2] Deodhar M, Al Rihani SB, Arwood MJ, Darakjian L, Dow P, Turgeon J and Michaud V. Mechanisms of CYP450 inhibition: understanding drug-drug interactions due to mechanism-based inhibition in clinical practice. *Pharmaceutics* 2020; 12: 846.
- [3] Darakjian L, Deodhar M, Turgeon J and Michaud V. Chronic inflammatory status observed in patients with type 2 diabetes induces modulation of cytochrome P450 expression and activity. *Int J Mol Sci* 2021; 22: 4967.
- [4] Duff HJ, Mitchell LB, Wyse DG, Gillis AM and Sheldon RS. Mexiletine/quinidine combination therapy: electrophysiologic correlates of antiarrhythmic efficacy. *Clin Invest Med* 1991; 14: 476-483.
- [5] Duff HJ, Roden D, Primm RK, Oates JA and Woosley RL. Mexiletine in the treatment of resistant ventricular arrhythmias: enhancement of efficacy and reduction of dose-related side effects by combination with quinidine. *Circulation* 1983; 67: 1124-1128.
- [6] Duff HJ, Mitchell LB, Manyari D and Wyse DG. Mexiletine-quinidine combination: electrophysiologic correlates of a favorable antiarrhythmic interaction in humans. *J Am Coll Cardiol* 1987; 10: 1149-1156.
- [7] Grech-Bélanger O, Gilbert M, Turgeon J and LeBlanc PP. Effect of cigarette smoking on mexiletine kinetics. *Clin Pharmacol Ther* 1985; 37: 638-643.
- [8] Turgeon J, Uprichard AC, Bélanger PM, Harron DW and Grech-Bélanger O. Resolution and electrophysiological effects of mexiletine enantiomers. *J Pharm Pharmacol* 1991; 43: 630-635.
- [9] Labbé L and Turgeon J. Clinical pharmacokinetics of mexiletine. *Clin Pharmacokinet* 1999; 37: 361-384.
- [10] Turgeon J. Métabolisme stéréosélectif de la mexiletine: implications des isoenzymes du cytochrome P-450. Ecole des gradués Université Laval. 1987. Thèse de doctorat. pp 301.
- [11] Turgeon J, Fiset C, Giguère R, Gilbert M, Morige K, Rouleau JR, Kroemer HK, Eichelbaum M, Grech-Bélanger O and Bélanger PM. Influence of debrisoquine phenotype and of quinidine on mexiletine disposition in man. *J Pharmacol Exp Ther* 1991; 259: 789-798.
- [12] Kroemer HK, Mikus G, Kronbach T, Meyer UA and Eichelbaum M. In vitro characterization of the human cytochrome P-450 involved in polymorphic oxidation of propafenone. *Clin Pharmacol Ther* 1989; 45: 28-33.
- [13] Funck-Brentano C, Kroemer H, Pavlou H, Woosley R and Roden D. Genetically-determined interaction between propafenone and low dose quinidine: role of active metabolites in modulating net drug effect. *Br J Clin Pharmacol* 1989; 27: 435-444.
- [14] Thompson KA, Iansmith DH, Siddoway LA, Woosley RL and Roden DM. Potent electrophysiologic effects of the major metabolites of propafenone in canine Purkinje fibers. *J Pharmacol Exp Ther* 1988; 244: 950-955.
- [15] O'Hara GE, Philippon F, Gilbert M, Champagne J, Michaud V, Charbonneau L, Pruneau G, Hamelin BA, Geelen P and Turgeon J. Combined administration of quinidine and propafenone for atrial fibrillation: the CAQ-PAF study. *J Clin Pharmacol* 2012; 52: 171-179.
- [16] Dayer P, Desmeules J, Leemann T and Striberini R. Bioactivation of the narcotic drug codeine in human liver is mediated by the polymorphic monooxygenase catalyzing debrisoquine 4-hydroxylation (cytochrome P-450 db1/bufl). *Biochem Biophys Res Commun* 1988; 152: 411-416.
- [17] Desmeules J, Gascon MP, Dayer P and Magistris M. Impact of environmental and genetic factors on codeine analgesia. *Eur J Clin Pharmacol* 1991; 41: 23-26.
- [18] Crews KR, Monte AA, Huddart R, Caudle KE, Kharasch ED, Gaedigk A, Dunnenberger HM, Leeder JS, Callaghan JT, Samer CF, Klein TE, Haidar CE, Van Driest SL, Ruano G, Sangkuhl

Cases of phenoconversion

- K, Cavallari LH, Müller DJ, Prows CA, Nagy M, Somogyi AA and Skaar TC. Clinical pharmacogenetics implementation consortium guideline for CYP2D6, OPRM1, and COMT genotypes and select opioid therapy. *Clin Pharmacol Ther* 2021; 110: 888-896.
- [19] Matos A, Bankes DL, Bain KT, Ballinghoff T and Turgeon J. Opioids, polypharmacy, and drug interactions: a technological paradigm shift is needed to ameliorate the ongoing opioid epidemic. *Pharmacy (Basel)* 2020; 8: 154.
- [20] Ballinghoff T, Bain KT, Matos A, Bardolia C, Turgeon J and Amin NS. Opioid response in an individual with altered cytochrome P450 2D6 activity: implications of a pharmacogenomics case. *Clin Case Rep J* 2020; 1: 1-4.
- [21] Deodhar M, Dow P, Al Rihani SB, Turgeon J and Michaud V. An illustrative case of phenoconversion due to multi-drug interactions. *Clin Case Rep J* 2020; 1: 1-6.
- [22] Michaud V, Bikmetov R, Smith MK, Dow P, Darrakjian L, Deodhar M, Cicali B, Bain KT and Turgeon J. Use of drug claims data and a medication risk score to assess the impact of CYP2D6 drug interactions among opioid users on healthcare costs. *J Pers Med* 2021; 11: 1174.
- [23] Kazui M, Nishiya Y, Ishizuka T, Hagihara K, Farid NA, Okazaki O, Ikeda T and Kurihara A. Identification of the human cytochrome P450 enzymes involved in the two oxidative steps in the bioactivation of clopidogrel to its pharmacologically active metabolite. *Drug Metab Dispos* 2010; 38: 92-99.
- [24] Cavallari LH, Lee CR, Beitelshes AL, Cooper-DeHoff RM, Duarte JD, Voora D, Kimmel SE, McDonough CW, Gong Y, Dave CV, Pratt VM, Alestock TD, Anderson RD, Alsip J, Ardani AK, Brott BC, Brown L, Chumnumwat S, Clare-Salzler MJ, Coons JC, Denny JC, Dillon C, Elsey AR, Hamadeh IS, Harada S, Hillegass WB, Hines L, Horenstein RB, Howell LA, Jeng LJB, Kelemen MD, Lee YM, Magvanjav O, Montasser M, Nelson DR, Nutescu EA, Nwaba DC, Pakyz RE, Palmer K, Peterson JF, Pollin TI, Quinn AH, Robinson SW, Schub J, Skaar TC, Smith DM, Sripramoju VB, Starostik P, Stys TP, Stevenson JM, Varunok N, Vesely MR, Wake DT, Weck KE, Weitzel KW, Wilke RA, Willig J, Zhao RY, Kreutz RP, Stouffer GA, Empey PE, Limdi NA, Shuldiner AR, Winterstein AG and Johnson JA. Multi-site investigation of outcomes with implementation of CYP2C19 genotype-guided antiplatelet therapy after percutaneous coronary intervention. *JACC Cardiovasc Interv* 2018; 11: 181-191.
- [25] Mega JL, Simon T, Collet JP, Anderson JL, Antman EM, Bliden K, Cannon CP, Danchin N, Giusti B, Gurbel P, Horne BD, Hulot JS, Kastrati A, Montalescot G, Neumann FJ, Shen L, Sibbing D, Steg PG, Trenk D, Wiviott SD and Sabatine MS. Reduced-function CYP2C19 genotype and risk of adverse clinical outcomes among patients treated with clopidogrel predominantly for PCI: a meta-analysis. *JAMA* 2010; 304: 1821-1830.
- [26] Gravel S, Chiasson JL, Turgeon J, Grangeon A and Michaud V. Modulation of CYP450 activities in patients with type 2 diabetes. *Clin Pharmacol Ther* 2019; 106: 1280-1289.
- [27] Hall HM, Banerjee S and McGuire DK. Variability of clopidogrel response in patients with type 2 diabetes mellitus. *Diab Vasc Dis Res* 2011; 8: 245-253.
- [28] Angiolillo DJ, Fernandez-Ortiz A, Bernardo E, Ramirez C, Sabaté M, Jimenez-Quevedo P, Hernández R, Moreno R, Escaned J, Alfonso F, Bañuelos C, Costa MA, Bass TA and Macaya C. Platelet function profiles in patients with type 2 diabetes and coronary artery disease on combined aspirin and clopidogrel treatment. *Diabetes* 2005; 54: 2430-2435.
- [29] Sweeny JM, Angiolillo DJ, Franchi F, Rollini F, Waksman R, Raveendran G, Dangas G, Khan ND, Carlson GF, Zhao Y, Teng R and Mehran R. Impact of diabetes mellitus on the pharmacodynamic effects of ticagrelor versus clopidogrel in troponin-negative acute coronary syndrome patients undergoing Ad Hoc percutaneous coronary intervention. *J Am Heart Assoc* 2017; 6: e005650.
- [30] Schuette C, Steffens D, Witkowski M, Stellabaum C, Bobbert P, Schultheiss HP and Rauch U. The effect of clopidogrel on platelet activity in patients with and without type-2 diabetes mellitus: a comparative study. *Cardiovasc Diabetol* 2015; 14: 15.
- [31] Erlinge D, Varenhorst C, Braun OÖ, James S, Winters KJ, Jakubowski JA, Brandt JT, Sugidachi A, Siegbahn A and Wallentin L. Patients with poor responsiveness to thienopyridine treatment or with diabetes have lower levels of circulating active metabolite, but their platelets respond normally to active metabolite added ex vivo. *J Am Coll Cardiol* 2008; 52: 1968-1977.
- [32] Wadelius M and Pirmohamed M. Pharmacogenetics of warfarin: current status and future challenges. *Pharmacogenomics J* 2007; 7: 99-111.
- [33] Fasco MJ and Principe LM. R- and S-Warfarin inhibition of vitamin K and vitamin K 2,3-epoxide reductase activities in the rat. *J Biol Chem* 1982; 257: 4894-4901.
- [34] Wingard LB Jr, O'Reilly RA and Levy G. Pharmacokinetics of warfarin enantiomers: a search

Cases of phenoconversion

- for intrasubject correlations. *Clin Pharmacol Ther* 1978; 23: 212-217.
- [35] Chan E, McLachlan AJ, Pegg M, MacKay AD, Cole RB and Rowland M. Disposition of warfarin enantiomers and metabolites in patients during multiple dosing with rac-warfarin. *Br J Clin Pharmacol* 1994; 37: 563-569.
- [36] James AH, Britt RP, Raskino CL and Thompson SG. Factors affecting the maintenance dose of warfarin. *J Clin Pathol* 1992; 45: 704-706.
- [37] Voora D, McLeod HL, Eby C and Gage BF. Use of pharmacogenetics to guide warfarin therapy. *Drugs Today (Barc)* 2004; 40: 247-257.
- [38] Sullivan-Klose TH, Ghanayem BI, Bell DA, Zhang ZY, Kaminsky LS, Shenfield GM, Miners JO, Birkett DJ and Goldstein JA. The role of the CYP2C9-Leu359 allelic variant in the tolbutamide polymorphism. *Pharmacogenetics* 1996; 6: 341-349.
- [39] Ieiri I, Tainaka H, Morita T, Hadama A, Mamiya K, Hayashibara M, Ninomiya H, Ohmori S, Kitada M, Tashiro N, Higuchi S and Otsubo K. Catalytic activity of three variants (Ile, Leu, and Thr) at amino acid residue 359 in human CYP2C9 gene and simultaneous detection using single-strand conformation polymorphism analysis. *Ther Drug Monit* 2000; 22: 237-244.
- [40] Dickmann LJ, Rettie AE, Kneller MB, Kim RB, Wood AJ, Stein CM, Wilkinson GR and Schwarz UI. Identification and functional characterization of a new CYP2C9 variant (CYP2C9*5) expressed among African Americans. *Mol Pharmacol* 2001; 60: 382-387.
- [41] Kidd RS, Curry TB, Gallagher S, Edeki T, Blaisdell J and Goldstein JA. Identification of a null allele of CYP2C9 in an African-American exhibiting toxicity to phenytoin. *Pharmacogenetics* 2001; 11: 803-808.
- [42] Tai G, Farin F, Rieder MJ, Dreisbach AW, Veenstra DL, Verlinde CL and Rettie AE. In-vitro and in-vivo effects of the CYP2C9*11 polymorphism on warfarin metabolism and dose. *Pharmacogenet Genomics* 2005; 15: 475-481.
- [43] Scordo MG, Pengo V, Spina E, Dahl ML, Gusella M and Padriani R. Influence of CYP2C9 and CYP2C19 genetic polymorphisms on warfarin maintenance dose and metabolic clearance. *Clin Pharmacol Ther* 2002; 72: 702-710.
- [44] Sun X, Yu WY, Ma WL, Huang LH and Yang GP. Impact of the CYP4F2 gene polymorphisms on the warfarin maintenance dose: A systematic review and meta-analysis. *Biomed Rep* 2016; 4: 498-506.
- [45] Rieder MJ, Reiner AP, Gage BF, Nickerson DA, Eby CS, McLeod HL, Blough DK, Thummel KE, Veenstra DL and Rettie AE. Effect of VKORC1 haplotypes on transcriptional regulation and warfarin dose. *N Engl J Med* 2005; 352: 2285-2293.
- [46] Higashi MK, Veenstra DL, Kondo LM, Wittkowsky AK, Srinouanprachanh SL, Farin FM and Rettie AE. Association between CYP2C9 genetic variants and anticoagulation-related outcomes during warfarin therapy. *Jama* 2002; 287: 1690-1698.
- [47] Ogg MS, Brennan P, Meade T and Humphries SE. CYP2C9* 3 allelic variant and bleeding complications. *Lancet* 1999; 354: 1124.
- [48] Margaglione M, Colaizzo D, D'Andrea G, Braccaccio V, Ciampa A, Grandone E and Di Minno G. Genetic modulation of oral anticoagulation with warfarin. *Thromb Haemost* 2000; 84: 775-778.
- [49] Aithal GP, Day CP, Kesteven PJ and Daly AK. Association of polymorphisms in the cytochrome P450 CYP2C9 with warfarin dose requirement and risk of bleeding complications. *Lancet* 1999; 353: 717-719.
- [50] Wadelius M, Sörlin K, Wallerman O, Karlsson J, Yue QY, Magnusson PK, Wadelius C and Melhus H. Warfarin sensitivity related to CYP2C9, CYP3A5, ABCB1 (MDR1) and other factors. *Pharmacogenomics J* 2004; 4: 40-48.
- [51] Taube J, Halsall D and Baglin T. Influence of cytochrome P-450 CYP2C9 polymorphisms on warfarin sensitivity and risk of over-anticoagulation in patients on long-term treatment. *Blood* 2000; 96: 1816-1819.
- [52] Joffe HV, Xu R, Johnson FB, Longtine J, Kucher N and Goldhaber SZ. Warfarin dosing and cytochrome P450 2C9 polymorphisms. *Thromb Haemost* 2004; 91: 1123-1128.
- [53] Woosley RL, Drayer DE, Reidenberg MM, Nies AS, Carr K and Oates JA. Effect of acetylator phenotype on the rate at which procainamide induces antinuclear antibodies and the lupus syndrome. *N Engl J Med* 1978; 298: 1157-1159.
- [54] Uetrecht JP, Freeman RW and Woosley RL. The implications of procainamide metabolism to its induction of lupus. *Arthritis Rheum* 1981; 24: 994-1003.
- [55] Lessard E, Fortin A, Bélanger PM, Beaune P, Hamelin BA and Turgeon J. Role of CYP2D6 in the N-hydroxylation of procainamide. *Pharmacogenetics* 1997; 7: 381-390.
- [56] Lessard E, Hamelin BA, Labbé L, O'Hara G, Bélanger PM and Turgeon J. Involvement of CYP2D6 activity in the N-oxidation of procainamide in man. *Pharmacogenetics* 1999; 9: 683-696.
- [57] Garber J and Brownlee S. Medication overload: America's other drug problem. 2019; <https://lowinstitute.org/reports/medication-overload-americas-other-drug-problem/>.

Cases of phenoconversion

- [58] Gravel S, Chiasson JL, Gaudette F, Turgeon J and Michaud V. Use of 4 β -Hydroxycholesterol plasma concentrations as an endogenous biomarker of CYP3A activity: clinical validation in individuals with type 2 diabetes. *Clin Pharmacol Ther* 2019; 106: 831-840.
- [59] Magliocco G, Matthey A, Bararpour N, Joye T, Gloor Y, Desmeules J, Thomas A and Daali Y. Metabolomics reveals five endogenous biomarkers in human urine and plasma to predict CYP2D6 activity. *Br J Pharmacol* 2021; 178: 4708-4725.