### Original Article Effect of decitabine and thalidomide on the immunological effect and bone marrow mesenchymal stem cells of patients with myelodysplastic syndrome

Lina Xing, Jinhai Ren, Xiaonan Guo, Shukai Qiao, Tian Tian

Department of Hematology, Second Affiliated Hospital, Hebei Medical University, Shijiazhuang 050000, Hebei Province, China

Received June 11, 2020; Accepted November 22, 2020; Epub April 15, 2021; Published April 30, 2021

Abstract: Objective: This study intended to investigate the therapeutic effect of decitabine and thalidomide on myelodysplastic syndrome (MDS), immunological effect and effective mesenchymal stem cells (MSCs). Methods: Altogether 62 patients with MDS diagnosed in our hospital were selected. Patients who received 5-day treatment mainly and received decitabine from the 1<sup>st</sup> day to the 5<sup>th</sup> day were collected as group A (A), while patients who received thalidomide 1<sup>st</sup> to 5<sup>th</sup> day as in group A were collected as group B (B). The immunologic effects, blood and bone marrow index levels, clinical effects and adverse reactions of group A and group B before and after intervention were observed. Results: Th17 in the two groups after intervention were evidently lower than that before intervention, and the decrease of Th17 cells in group B after intervention was more obvious than that in group A (P<0.001). Th22 cells in the two groups after intervention were evidently down-regulated compared with those before intervention, and the down-regulation of Th17 cells in group B after intervention was more obvious than that in group A (P<0.001). However, compared with group A, the levels of CD3+, CD4+, CD4+/CD8+ in serum of group B increased more obviously and CD8+ decreased more obviously after intervention. The white blood cell count of group B after intervention was evidently higher than that of group A (P<0.001). The hemoglobin concentration after intervention in group B was evidently higher than that in group A (P<0.001). The platelet count after intervention in group A was evidently higher than that in group B (P<0.001). The total effective rate in group B was evidently higher than that in group A (P<0.05). Conclusion: The combination of decitabine and thalidomide has a better regulatory role in the immunological mechanism and bone marrow mesenchymal stem cells of patients with MDS than the single decitabine therapy on the premise of ensuring clinical efficacy.

**Keywords:** Decitabine, thalidomide, myelodysplastic syndrome, curative effect, immunological effect, bone marrow mesenchymal stem cells

#### Introduction

Myelodysplastic syndrome (MDS) is a heterogeneous disease composed of heterogeneous myeloma [1-3]. The risk of patients developing acute myeloid leukemia is very high due to low hematopoietic efficiency [4]. The incidence of MDS increases greatly with age [5, 6]. The epidemiological evaluation of MDS has been hindered by the continuous development of diagnostic criteria and the classification of MDS as cancer. Poor understanding of patients with early MDS will lead to the lack of timely and effective treatment [7, 8].

Chemotherapy and hematopoietic stem cell transplantation are currently the main clinical

treatments for MDS [9, 10]. Decitabine is a hypomethylating agent and can reactivate tumor suppressor genes by demethylating them [11]. In recent years, decitabine has become a common clinical drug for patients with MDS [12]. Decitabine has been mainly applied as monotherapy, but some studies have shown that the combination with various reagents such as vorinostat, thalidomide and decitabine gemtuzumab can effectively improve the prognosis of patients with MDS [13]. Thalidomide is a well-known angiogenesis inhibitor and immunomodulator, which has anti-angiogenic activity, can reduce tumor necrosis factor- $\alpha$ , and can be applied for the treatment of MDS, multiple myeloma and other diseases [14, 15]. The mechanism for promoting the development of

Group	Group A (n=32)	Group B (n=30)	X <sup>2</sup>	Р
Age (years)			0.022	0.881
<59.46	8 (25.00)	8 (26.67)		
>59.46	24 (75.00)	22 (73.33)		
Gender			0.625	0.429
Male	16 (50.00)	12 (40.00)		
Female	16 (50.00)	18 (60.00)		
Hypertension			1.245	0.265
Yes	28 (87.50)	23 (76.67)		
No	4 (12.50)	7 (23.33)		
Diabetes			0.389	0.533
Yes	20 (62.50)	21 (70.00)		
No	12 (37.50)	9 (30.00)		
Diagnostic type			0.453	0.767
Refractory anemia	10 (31.25)	10 (33.33)		
Refractory anemia with ring sideroblast	11 (34.38)	12 (40.00)		
Refractory anemia with increase of primitive cells	11 (34.38)	8 (26.67)		
Risk			0.230	0.632
Moderate risk I	13 (40.63)	14 (46.67)		
Moderate risk II	19 (59.38)	16 (53.33)		
High risk	0 (0.00)	0 (0.00)		

 Table 1. General clinical data

MDS diseases is complex. Relevant studies have shown that the occurrence of MDS is closely related to family genetic disease history, autoimmune effect abnormalities and changes in bone marrow micro-environment [16, 17].

This study aimed to analyze the therapeutic role of decitabine and thalidomide on patients with MDS, immunological effect and bone marrow mesenchymal stem cells through two different therapeutic schemes.

### General data and methods

### General data

Sixty-two patients with MDS diagnosed in our hospital were selected. Patients who received conventional MDS treatment (mainly with 5 d regimen and decitabine from the 1<sup>st</sup> day to the 5<sup>th</sup> day) were collected as group A (A), while patients who received thalidomide on the basis of group A were collected as group B (B). There were 32 cases in group A and 30 cases in group B. Inclusion criteria: (1) Patients were diagnosed as MDS and treated in our hospital, which was referred to the World Health Organization's Diagnostic Criteria for MDS [18]; participants had no abortion caused by chromosome, anatomy, endocrine abnormalities, reproductive system infection and autoimmune diseases. Exclusion criteria: (2) Patients with contraindications to the drugs applied in this study; patients had other primary diseases of morbid hematopoiesis and hematocytopenia; patients with hypertension, AIDS and various blood diseases. Participants and their families signed informed consent forms in advance. The study was approved by the Ethics Committee.

### Method

Group A: Patients were mainly treated with 5 d regimen, decitabine (SFDA Approval No. H20130067, Hansoh Pharma Co., Ltd., Jiangsu) was given intravenously from the  $1^{st}$  day to the  $5^{th}$  day, and the intravenous drip was completed within 1 h.

Group B: Patients were given thalidomide on the basis of group A (SFDA Approval No. H32026129, Changzhou Pharmaceutical Factory Co., Ltd.). After continuous oral administration of 50 mg/d thalidomide for 1 week, the dosage was adjusted to 100 mg/d, and the same medication was applied for 1 week.



**Figure 1.** Changes of Th17 cells before and after intervention in group A and group B. There was no evident difference in Th17 cells between group A and group B before intervention (P>0.05). The Th17 cells in the two groups after intervention were evidently lower than before intervention, and the Th17 cells in group B patients after intervention were evidently lower than those in group A patients. Note: a means P<0.001.

### Outcome measures

Immunological role was analyzed before and after intervention in group A and group B (fasting venous blood of elbow was collected before and after intervention in both groups, and Th17 cells, Th22 cells, CD3+, CD4+, CD8+, CD4+/ CD8+ levels were detected by flow cytometry). The blood and bone marrow index levels (including white blood cell count, hemoglobin concentration, platelet count, bone marrow mesenchymal stem cell ratio) of patients in group A and group B before and after intervention were compared. The clinical efficacy of group A and group B was compared [19]. The adverse reac-



**Figure 2.** Changes of Th22 cells before and after intervention in group A and group B. There was no evident difference in Th22 cells between group A and group B before intervention (P>0.05). The Th22 cells in the two groups after intervention were evidently lower than before intervention, and the Th22 cells in group B after intervention were evidently lower than those in group A. Note: a means P<0.001.

tions (including nausea and vomiting, fever, myelosuppression, thermal neutropenia, headache) of group A and group B were compared.

### Statistical methods

SPSS 17.0 (Beijing Bi Insight Information Technology Co., Ltd.) was applied for statistical analysis. The counting data were represented by [n (%)] and tested by X<sup>2</sup> test. The measurement data were represented as (x  $\pm$  s) and tested by independent sample t test. When P< 0.05, the difference was statistically evident.

### Result

General clinical data of A and B

There was no evident difference between A and B in general clinical data (P>0.05) (**Table 1**).



Figure 3. Levels of immune indexes before and after intervention in group A and group B. A. CD3+ (%) level of patients in group A and group B; B. CD4+ (%) level of patients in group A and group B; C. CD8+ (%) level of patients in group A and group B; D. CD4+/CD8+ (%) levels of patients in groups A and B; a means P<0.05.

## Analysis of immunological effects before and after intervention in groups A and B

Changes of Th17 cells in groups A and B before and after intervention: The Th17 cells of group A before and after intervention were  $(1.90 \pm 0.20)$ %,  $(1.79 \pm 0.20)$ %, while those of group B before and after intervention were  $(1.92 \pm 0.20)$ %,  $(1.18 \pm 0.20)$ %. There was no evident difference in Th17 cells between A and B before intervention (P>0.05). Th17 cells in the two groups after intervention were evident-

ly lower than before intervention, and the decrease of Th17 cells in group B after intervention was more obvious than that in group A (P<0.001). See **Figure 1**.

The changes of Th22 cells before and after intervention in group A and group B: The Th22 cells of group A patients before and after intervention were  $(4.45 \pm 0.30)\%$ , (3.67 ± 0.30)%, while the Th17 cells of group B patients before and after intervention were (4.52 ± 0.30)%, (2.89 ± 0.30)%. There was no evident difference in Th22 cells between A and B before intervention (P>0.05). Th22 cells in the two groups after intervention were evidently downregulated compared with those before intervention, and the down-regulation effect of Th17 cells in group B after intervention was more obvious than that in group A (P< 0.001). See Figure 2.

Immune index levels before and after intervention in group A and group B: CD3+, CD4+, CD8+, CD4+/CD8+ in A and B were tested by flow cytometry. The results showed that CD3+, CD4+, CD4+/ CD8+ in group A and group B after intervention increased (P<0.05), while CD8+ decreased. However, after intervention, CD3+, CD4+, CD4+/ CD8+ in B increased more

obviously and CD8+ decreased more obviously compared with A, but the difference was not statistically evident (**Figures 3**, **4**).

### Comparison of blood and myelogram indexes before and after intervention between group A and Group B patients

White blood cell count: There was no evident difference in white blood cell count between A and B before intervention (P>0.05). The white blood cell count of A and B after intervention

### Effect analysis of decitabine and thalidomide on MDS patients



Figure 4. Original figures of flow cytometry.

was evidently higher than that before intervention, and the white blood cell count of group B after intervention was evidently higher than that of group A (P<0.001). See **Figure 5**.

Hemoglobin concentration: There was no evident difference in hemoglobin concentration between A and B before intervention (P>0.05). The hemoglobin concentration after intervention in both groups was evidently higher than that before intervention, and the hemoglobin concentration after intervention in group B was evidently higher than that in group A (P<0.001). See **Figure 6**.

*Platelet count:* There was no evident difference in platelet count between A and B before intervention (P>0.05). The platelet count after intervention in both groups was evidently higher than that before intervention, and the platelet count after intervention in group A was evidently higher than that in group B (P<0.001). See Figure 7.

Proportion of bone marrow mesenchymal stem cells %: There was no evident difference in the proportion of bone marrow mesenchymal stem cells between A and B before intervention (P>0.05). The proportion of bone marrow mesenchymal stem cells after intervention in both groups was evidently lower than that before intervention, and the proportion of bone marrow mesenchymal stem cells after intervention in group B was evidently lower than that in group A (P<0.001). See **Figure 8**.

### Analysis of clinical efficacy and adverse reactions of groups A and B

Clinical efficacy: The curative role analysis of A and B showed that the total effective rate of B was evidently higher than that of A (P<0.05). See **Table 2**.





Figure 5. White blood cell count. The white blood cell count of group A and group B after intervention was evidently higher than that before intervention, and the white blood cell count of group B after intervention was evidently higher than that of group A. a means P<0.001.

Adverse reactions: The total adverse reaction rate of nausea, vomiting, fever, myelosuppression, thermal neutropenia and headache in A was 28.13%, while that of nausea, vomiting, fever, myelosuppression, thermal neutropenia and headache in B was 23.33%. The incidence of adverse reactions in B was lower than that in A (P>0.05). See Table 3.

### Discussion

MDS has a rising trend with the deterioration of diseases [20]. Patients with MDS have weakened immune surveillance due to abnormal immune mechanism, thus causing a series of diseased manifestationss [21]. Bone mesenchymal stem cells with immunoregulatory properties are important factors indicating the development of MDS. Clinically, it is very impor-

Figure 6. Hemoglobin concentration. The hemoglobin concentration after intervention in both groups was evidently higher than that before intervention, and the hemoglobin concentration after intervention in group B was evidently higher than that in group A. a means P<0.001.

tant to find a more safe and effective treatment scheme for relieving the physiological pain and syndromes of patients with MDS [22]. In this study, we analyzed the role of decitabine and thalidomide on the therapeutic efficacy, immunological effects and bone marrow mesenchymal stem cells of patients with MDS.

Clinical data suggested that decitabine had a good effect on promoting the stability of the disease, improving the prognosis of MDS, and prolonging the median time for MDS patients to transform into leukemia and die. However, the single use of decitabine is highly dosedependent, and the adverse drug effects caused by large doses are also important clinical problems yet to be solved [23]. In this study, through the analysis of immunologic effector cells before and after intervention, we found that Th17 cells after intervention in both



Figure 7. Platelet count. The platelet count after intervention in both groups was evidently higher than that before intervention, and the platelet count after intervention in group A was evidently higher than that in group B (P<0.001). a means P<0.001.

groups were evidently lower than that before intervention, and the down-regulation effect of Th17 and Th22 cells in patients with MDS after intervention with decitabine and thalidomide was evidently higher than that of patients with decitabine alone. Moreover, CD3+, CD4+, CD8+, CD4+/CD8+ measured by flow cytometry showed that those in patients with MDS after intervention with decitabine and thalidomide were increased to a greater extent. It was previously shown that thalidomide could correct immune abnormalities by improving CD3+, CD4+ and CD8+ levels [24]. Therefore, we believed that the combination of decitabine and thalidomide had a better regulatory effect on the immune suppression of patients.

Then, by comparing the blood and bone marrow index levels of group A and group B of patients, we found that the hemoglobin concentration,



Figure 8. Proportion of bone marrow mesenchymal stem cells. The proportion of bone marrow mesenchymal stem cells after intervention in both groups was evidently lower than that before intervention, and the proportion of bone marrow mesenchymal stem cells after intervention in group B was evidently lower than that in group A. a means P<0.001.

white blood cell count and platelet count of A and B after intervention were evidently higher than those before intervention. Among them, the hemoglobin concentration and white blood cell count of patients with MDS after intervention with decitabine combined with thalidomide were evidently higher than those of patients with decitabine alone, but the up-regulation of platelet count after intervention was lower than that of patients with decitabine alone. The proportion of bone marrow mesenchymal stem cells after intervention in both groups was evidently lower than that before intervention, and the proportion of bone marrow mesenchymal stem cells after intervention in patients treated with decitabine and thalidomide was evidently lower. MDS is caused by abnormal hematopoietic micro-environment of bone marrow, clonal proliferation of hematopoietic stem cells, and bone marrow mesenchymal stem cells are an

Table 2. The curative role analysis of A and B showed

Group	Group A (n=32)	Group B (n=30)	X <sup>2</sup>	Ρ
Complete remission	4 (12.50)	9 (30.00)	-	-
Partial remission	8 (25.00)	8 (26.67)	-	-
Progression	8 (25.00)	9 (30.00)	-	-
Ineffective	12 (37.50)	4 (13.33)	-	-
Total effective rate of treatment	20 (62.50)	26 (86.67)	4.723	0.030

Table 3	Analysis	of adverse	reactions	of A and B
	11019313	01 00 00 00	reactions	

Group	Group A (n=32)	Group B (n=30)	X <sup>2</sup>	Р
Nausea and vomiting	4 (12.50)	2 (6.67)	-	-
Fever	3 (9.38)	2 (6.67)	-	-
Bone marrow suppression	0 (0.00)	0 (0.00)	-	-
Thermal neutropenia	0 (0.00)	0 (0.00)		
Headache	2 (6.25)	3 (30.00)	-	-
Total adverse reaction rate	9 (28.13)	7 (23.33)	0.186	0.666

important component of hematopoietic microenvironment [25]. A large number of studies have proved that thalidomide can inhibit the abnormal proliferation of bone marrow cells, promote the recovery of hematopoietic function, and improve the body's bone and marrow images [26]. Moreover, Zhao et al. also found that decitabine and thalidomide treatment had better regulatory effect on blood and bone marrow indexes of patients [27]. Finally, at the end of the treatment course, we compared the clinical efficacy and adverse reactions of patients. The results showed that decitabine combined with thalidomide in the treatment of MDS reduced the total adverse reaction rate of nausea, vomiting, fever, myelosuppression, thermal neutropenia and headache to some extent.

In this study, there are still some deficiencies. For example, the data display of other blood indexes of patients still needs to be supplemented, and the time point of experimental design can be more specific. These deficiencies will produce certain errors in the data of this study. In view of the above deficiencies, we will continue to refer to the relevant research results on the treatment of MDS in order to enrich the actual clinical data in the later stage, and regularly follow up the patients to continuously improve the test.

To sum up, the combination of decitabine and thalidomide treatment has better regulatory effect on the immunological mechanism and effective bone marrow mesenchymal stem cells of patients with MDS than single decitabine treatment on the premise of ensuring clinical efficacy.

### Acknowledgements

This study is financially supported by Clinical study on the treatment of medullary tumors by desitabine (20180293).

# Disclosure of conflict of interest

None.

Address correspondence to:

Lina Xing, Department of Hematology, Second Affiliated Hospital, Hebei Medical University, NO. 215 Heping Xi Lu, Xinhua District, Shijiazhuang 050000, Hebei Province, China. Tel: +86-13831113199; E-mail: xinglina1023@163.com

### References

- [1] Moncharmont P, Quittancon E, Barday G and Benamara A; les Correspondants d'Hemovigilance et de sécuritétransfusionnelle Auvergne Rhône Alpes. Adverse transfusion reactions in patients with aplas tic anaemia or myelodysplastic syndromes. Vox Sang 2019; 114: 349-354.
- [2] Myers KC, Furutani E, Weller E, Siegele B, Galvin A, Arsenault V, Alter BP, Boulad F, Bueso-Ramos C, Burroughs L, Castillo P, Connelly J, Davies SM, DiNardo CD, Hanif I, Ho RH, Karras N, Manalang M, McReynolds LJ, Nakano TA, Nalepa G, Norkin M, Oberley MJ, Orgel E, Pastore YD, Rosenthal J, Walkovich K, Larson J, Malsch M, Elghetany MT, Fleming MD and Shimamura A. Clinical features and outcomes of patients with Shwachman-Diamond syndrome and myelodysplastic syndrome or acute myeloid leukaemia: a multicentre, retrospective, cohort study. Lancet Haematol 2020; 7: e238e246.
- [3] Girmenia C, Candoni A, Delia M, Latagliata R, Molteni A, Oliva EN, Palumbo GA, Poloni A, Salutari P, Santini V, Voso MT and Musto P. Infection control in patients with myelodysplastic syndromes who are candidates for active treatment: expert panel consensus-based recommendations. Blood Rev 2019; 34: 16-25.

- [4] Schaefer EJ and Lindsley RC. Significance of clonal mutations in bone marrow failure and inherited myelodysplastic syndrome/acute myeloid leukemia predisposition syndromes. Hematol Oncol Clin North Am 2018; 32: 643-655.
- [5] Brauninger A, Blau W, Kunze K, Desch AK, Brobeil A, Tur MK, Etschmann B, Gunther U, Korholz D, Schliesser G, Kabisch A, Kiehl M, Rummel M and Gattenlohner S. Targeted nextgeneration sequencing is a sensitive tool for differential diagnosis of myelodysplastic syndromes in bone marrow trephines. J Mol Diagn 2018; 20: 344-354.
- [6] Basiorka AA, McGraw KL, Abbas-Aghababazadeh F, McLemore AF, Vincelette ND, Ward GA, Eksioglu EA, Sallman DA, Ali NA, Padron E, Pinilla-Ibarz J, Komrokji R, Masala E, Santini V, Kosmider O, Fontenay M, Fenaux P, Sokol L, Wei S, Fridley B and List AF. Assessment of ASC specks as a putative biomarker of pyroptosis in myelodysplastic syndromes: an observational cohort study. Lancet Haematol 2018; 5: e393e402.
- Potter VT, Iacobelli S, van Biezen A, Maertens [7] J, Bourhis JH, Passweg JR, Yakhoub-Agha I, Tabrizi R, Bay JO, Chevallier P, Chalandon Y, Huynh A, Cahn JY, Ljungman P, Craddock C, Lenhoff S, Russell NH, Fegueux N, Socie G, Bruno B, Meijer E, Mufti GJ, de Witte T, Robin M and Kroger N. Comparison of intensive chemotherapy and hypomethylating agents before allogeneic stem cell transplantation for advanced myelodysplastic syndromes: a study of the myelodysplastic syndrome subcommittee of the chronic malignancies working party of the European society for blood and marrow transplant research. Biol Blood Marrow Transplant 2016; 22: 1615-1620.
- [8] Wesner N, Drevon L, Guedon A, Fraison JB, Trad S, Kahn JE, Aouba A, Gillard J, Ponsoye M, Hanslik T, Gourguechon C, Liozon E, Laribi K, Rossignol J, Hermine O, Ades L, Carrat F, Fenaux P, Mekinian A, Fain O and Gfm M. Inflammatory disorders associated with trisomy 8-myelodysplastic syndromes: French retrospective case-control study. Eur J Haematol 2019; 102: 63-69.
- [9] Stein EM, DiNardo CD, Fathi AT, Mims AS, Pratz KW, Savona MR, Stein AS, Stone RM, Winer ES, Seet CS, Dohner H, Pollyea DA, McCloskey J, Odenike O, Lowenberg B, Ossenkoppele GJ, Patel PA, Roshal M, Frattini MG, Lersch F, Franovic A, Nabhan S, Fan B, Choe S, Wang H, Wu B, Hua L, Almon C, Cooper M, Kantarjian HM and Tallman MS. Ivosidenib or enasidenib combined with intensive chemotherapy in patients with newly diagnosed AML: a phase 1 study. Blood 2020; blood.2020007233.

- [10] Bewersdorf JP and Zeidan AM. Evolving therapies for lower-risk myelodysplastic syndromes. Ann Hematol 2020; 99: 677-692.
- [11] Krevvata M, Shan X, Zhou C, Dos Santos C, Habineza Ndikuyeze G, Secreto A, Glover J, Trotman W, Brake-Silla G, Nunez-Cruz S, Wertheim G, Ra HJ, Griffiths E, Papachristou C, Danet-Desnoyers G and Carroll M. Cytokines increase engraftment of human acute myeloid leukemia cells in immunocompromised mice but not engraftment of human myelodysplastic syndrome cells. Haematologica 2018; 103: 959-971.
- [12] Gondek LP and DeZern AE. Assessing clonal haematopoiesis: clinical burdens and benefits of diagnosing myelodysplastic syndrome precursor states. Lancet Haematol 2020; 7: e73e81.
- [13] Oran B, de Lima M, Garcia-Manero G, Thall PF, Lin R, Popat U, Alousi AM, Hosing C, Giralt S, Rondon G, Woodworth G and Champlin RE. A phase 3 randomized study of 5-azacitidine maintenance vs observation after transplant in high-risk AML and MDS patients. Blood Adv 2020; 4: 5580-5588.
- [14] Kian W, Roisman LC, Wallach N, Levitas D, Yakobson A, Dudnik Y, Peled N and Rouvinov K. Programmed death-ligand 1 expression discrepancy between primary tumor and metastatic lymph nodes in non-small cell lung cancer. J Thorac Dis 2020; 12: 3918-3920.
- [15] Ludwig H, Ponisch W, Knop S, Egle A, Hinke A, Schreder M, Lechner D, Hajek R, Gunsilius E, Petzer A, Weisel K, Niederwieser D, Einsele H, Willenbacher W, Rumpold H, Pour L, Jelinek T, Krenosz KJ, Meckl A, Nolte S, Melchardt T, Greil R and Zojer N. Quality of life in patients with relapsed/refractory multiple myeloma during ixazomib-thalidomide-dexamethasone induction and ixazomib maintenance therapy and comparison to the general population. Leuk Lymphoma 2020; 61: 377-386.
- [16] Schratz KE and DeZern AE. Genetic predisposition to myelodysplastic syndrome in clinical practice. Hematol Oncol Clin North Am 2020; 34: 333-356.
- [17] Cao YG, He Y, Zhang SD, Liu ZX, Zhai WH, Ma QL, Pang AM, Wei JL, Yang DL, Huang Y, Feng SZ, Jiang EL and Han MZ. Conditioning regimen of 5-day decitabine administration for allogeneic stem cell transplantation in patients with myelodysplastic syndrome and myeloproliferative neoplasms. Biol Blood Marrow Transplant 2020; 26: 285-291.
- [18] Alkharabsheh O, Patnaik MM, Gangat N, Begna KH, Alkhateeb HB, Shah MV, Hogan WJ, He R, Greipp P, Nguyen PL, Litzow MR and Al-Kali A. Impact of marrow blasts percentage on high-grade myelodysplastic syndrome assess-

ed using revised international prognostic scoring system. Ann Hematol 2020; 99: 513-518.

- [19] Cluzeau T, McGraw KL, Irvine B, Masala E, Ades L, Basiorka AA, Maciejewski J, Auberger P, Wei S, Fenaux P, Santini V and List A. Pro-inflammatory proteins S100A9 and tumor necrosis factor-alpha suppress erythropoietin elaboration in myelodysplastic syndromes. Haematologica 2017; 102: 2015-2020.
- [20] Roupie AL, de Boysson H, Thietart S, Carrat F, Seguier J, Terriou L, Versini M, Queyrel V, Groh M, Benhamou Y, Maurier F, Decaux O, d'Aveni M, Rossignol J, Galland J, Solary E, Willems L, Schleinitz N, Ades L, Dellal A, Samson M, Aouba A, Fenaux P, Fain O and Mekinian A; On behalf MINHEMON (French Network of dysimmune disorders associated with hemopathies). Giant-cell arteritis associated with myelodysplastic syndrome: French multicenter case control study and literature review. Autoimmun Rev 2020; 19: 102446.
- [21] Montalban-Bravo G, Kanagal-Shamanna R, Guerra V, Ramos-Perez J, Hammond D, Shilpa P, Naqvi K, Sasaki K, Jabbour E, DiNardo C, Takahashi K, Konopleva M, Pemmaraju N, Kadia T, Ravandi F, Daver N, Borthakur G, Estrov Z, Khoury JD, Loghavi S, Sherry Pierce RN, Bueso-Ramos C, Patel K, Kantarjian H and Garcia-Manero G. Clinical outcomes and influence of mutation clonal dominance in oligomonocytic and classical chronic myelomonocytic leukemia. Am J Hematol 2020; [Epub ahead of print].
- [22] Short NJ, Kantarjian H and Jabbour E. Reply to ABCG2 overexpression and deoxyadenosine analogue activity in acute myeloid leukemia. Cancer 2017; 123: 4935-4936.

- [23] Garcia-Manero G, Pemmaraju N, Alvarado Y, Naqvi K, Ravandi F, Jabbour E, De Lumpa R, Kantarjian H, Advani A, Mukherjee S, Gerds A, Carraway HE, Nazha A, Iwamura H, Murase M, Bavisotto L, Kurman M, Maier G, Johansen M and Sekeres MA. Results of a phase 1/2a dose-escalation study of FF-10501-01, an IM-PDH inhibitor, in patients with acute myeloid leukemia or myelodysplastic syndromes. Leuk Lymphoma 2020; 61: 1943-1953.
- [24] Schuler E, Frank F, Hildebrandt B, Betz B, Strupp C, Rudelius M, Aul C, Schroeder T, Gattermann N, Haas R and Germing U. Myelodysplastic syndromes without peripheral monocytosis but with evidence of marrow monocytosis share clinical and molecular characteristics with CMML. Leuk Res 2018; 65: 1-4.
- [25] Kenealy M, Patton N, Filshie R, Nicol A, Ho SJ, Hertzberg M, Mills T, Prosser I, Link E, Cowan L, Zannino D and Seymour JF. Results of a phase II study of thalidomide and azacitidine in patients with clinically advanced myelodysplastic syndromes (MDS), chronic myelomonocytic leukemia (CMML) and low blast count acute myeloid leukemia (AML). Leuk Lymphoma 2017; 58: 298-307.
- [26] Tam CS, Seymour JF, Prince HM, Kenealy M, Wolf M, Januszewicz EH and Westerman D. Treatment-related myelodysplasia following fludarabine combination chemotherapy. Haematologica 2006; 91: 1546-1550.
- [27] Zhao WH, Huang BT, Zhang JY and Zeng QC. Distinct EphB4-mediated mechanisms of apoptotic and resistance to dasatinib in human chronic myeloid leukemia and K562 cell lines. Leuk Res 2017; 63: 28-33.