

Review Article

Mechanisms of COVID-19 thrombosis in an inflammatory environment and new anticoagulant targets

Huan Liu^{1,2}, Tianshui Hu², Cong Zhang², Xiaojing Chen², Shuoqi Zhang², Mengdi Li^{1,2}, Haijiao Jing², Chunxu Wang², Tenglong Hu¹, Jialan Shi^{2,3}

¹Department of Stomatology, The First Hospital of Harbin, Harbin Medical University, Harbin, China; ²Department of Hematology, The First Hospital of Harbin, Harbin Medical University, Harbin, China; ³Department of Research and Medicine, VA Boston Healthcare System, Brigham and Women's Hospital, Harvard Medical School, Boston, MA, USA

Received December 1, 2020; Accepted February 25, 2021; Epub May 15, 2021; Published May 30, 2021

Abstract: COVID-19 is widely epidemic in the world and poses a great threat to our life. Coagulopathy is one of the major characteristics in the COVID-19 patients. A growing number of studies have found that the severe COVID-19 patients have thrombotic microangiopathy and thromboembolism. Coagulopathy associated with increased risk of death in the patients. Unfortunately, the mechanism of coagulopathy is not clearly addressed. Understanding the pathophysiological mechanism of COVID-19 thrombosis and improving the coagulopathy through efficient treatment may help to stop disease progression, reduce mortality and sequelae. In severe COVID-19 patients, inflammation, cytokine storm, and coagulation are closely related, which together cause blood congestion and thrombosis. Many cytokines activate blood cells, expressing activating factors or releasing activated microparticles, and then accelerating thrombosis. However, the role of blood cells is not well understood in COVID-19 patients. In addition, cytokines stimulate endothelial cells, transforming them into a procoagulant phenotype. Therefore, determine their role and propose new strategies for the prevention and treatment of thrombosis in severe COVID-19 patients. We outline the major events of coagulopathies, discuss the role of blood and endothelial cells in thrombosis, to formulate a new anticoagulation protocol.

Keywords: COVID-19, thrombosis, cytokines, phosphatidylserine, neutrophil extracellular traps, anticoagulant therapy

Introduction

Since the outbreak of coronavirus disease-2019 (COVID-19) in December 2019, it has spread widely around the world. The disease, caused by severe acute respiratory syndrome coronavirus type 2 (SARS-CoV-2), shows flu-like symptoms and viral pneumonia, and develops into acute respiratory distress syndrome (ARDS), even multiple organ failure [1, 2]. The virus invasion stimulates the activation of the immune system, causing release of excessive pro-inflammatory factors, forming a cytokine storm in severe and critical patients, which is closely related to high mortality of COVID-19 patients [3-5]. There is a wide range of interactions between inflammation and

coagulation. The activation of one system may amplify the effect of another system, forming inflammatory-thrombosis exacerbating disease progression [1, 6, 7].

Studies have shown that thrombosis is one of the main causes of death in COVID-19 patients. In critically ill patients, D-dimer, fibrin degradation products, and fibrinogen are significantly increased, and D-dimer can be used as an indicator to predict patient mortality [8-10]. Early application of low-molecular-weight heparin (LMWH) improved the prognosis of patients, reduced mortality. Active anticoagulation measures were benefit to the clinical treatment of COVID-19 patients. Unfortunately, thrombotic events still occur in some patients with using

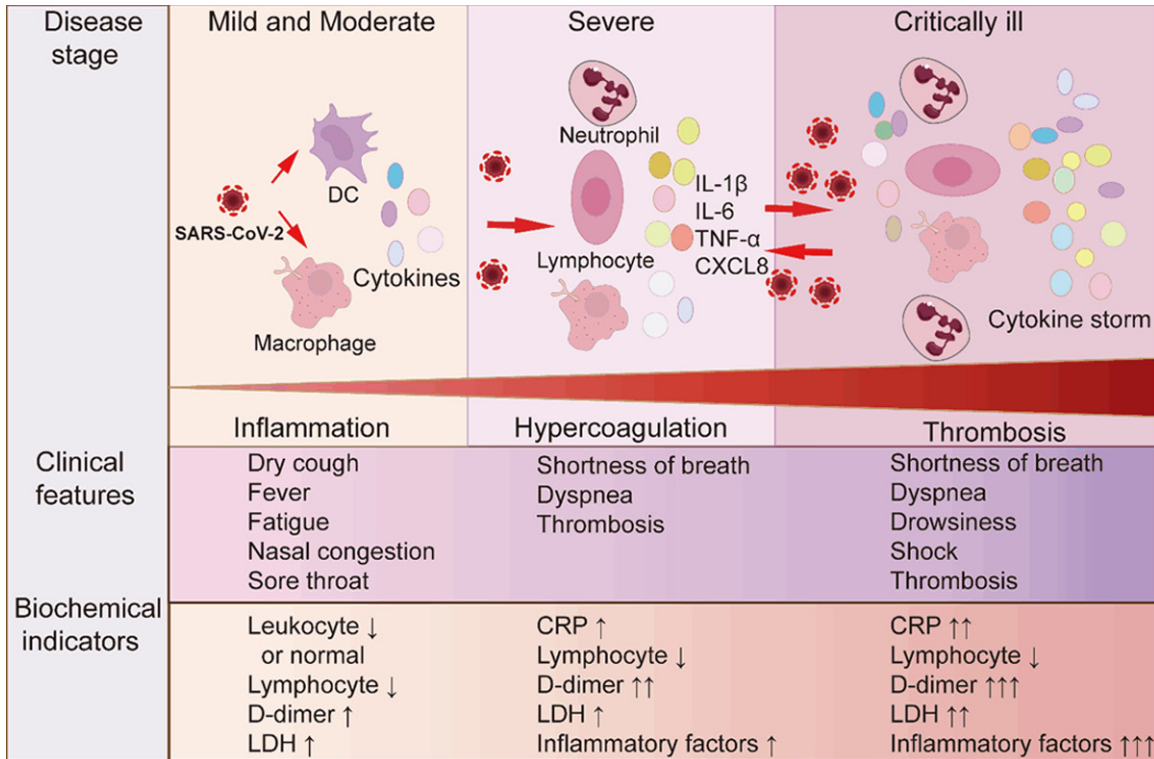


Figure 1. Clinical features and biochemical changes following COVID-19 infection. After virus invasion, active defense is made in response to the virus. With the progression of disease severity, patients show characteristic changes in clinical manifestations and hematological biochemical indicators. CRP: C-reactive protein; DC: Dendritic Cells; IL: interleukin; LDH: Lactate dehydrogenase; PMN: Polymorphonuclear neutrophil; TNF: tumor necrosis factor.

prophylactic anticoagulant [11-13]. This indicates that the thrombosis mechanism is complex, and other mechanisms are involved, so it is necessary to further explore the mechanism of coagulation disorders in COVID-19 patients. A variety of cells are involved in the process of thrombosis, and cytokines promote the activation of blood cells, thus we mainly explore the role of blood cells in thrombosis of COVID-19 patients. To expect multi-target blocking of thrombosis, improve disease prognosis, reduce patient sequelae and mortality.

Clinical feature of COVID-19

Coronavirus, a respiratory virus, is a coated RNA virus which induces the common cold to Middle East Respiratory Syndrome (MERS) and Severe Acute Respiratory Syndrome (SARS). Eventually, it will develop to fatal lower respiratory infections and extrapulmonary manifestations [4]. SARS-CoV-2, known as COVID-19, is a newly discovered coronavirus in 2019. It is a spherical particle with a diameter of about 50

nm, which consists of single-stranded RNA, the surface cell membrane, envelope protein, and nucleocapsid. The “spike” glycoprotein, also named S protein, has a high affinity for a receptor, angiotensin-converting enzyme 2 (ACE2). ACE2 is high expression mainly in alveolar cells, myocardial cells, endothelial cells (ECs) and others. COVID-19 can enter the lung, heart, blood vessels, kidney and gastrointestinal tract cells through the ACE2, leading to tissue injury, organ failure, such as ARDS, myocardial damage, kidney damage [14, 15].

COVID-19 is mainly spread by droplets, enters the lungs through the respiratory tract. Macrophages engulf the virus to secrete cytokines and promote the recruitment of white blood cells in the lung. In the early stage, the patient was in an asymptomatic stage or has a mild dry cough, fever, fatigue, and D-dimer increased slightly (Figure 1) [16]. With the virus proliferating, the immune system reacts strongly, appearing a series of clinical symptoms. The patient developed symptoms such

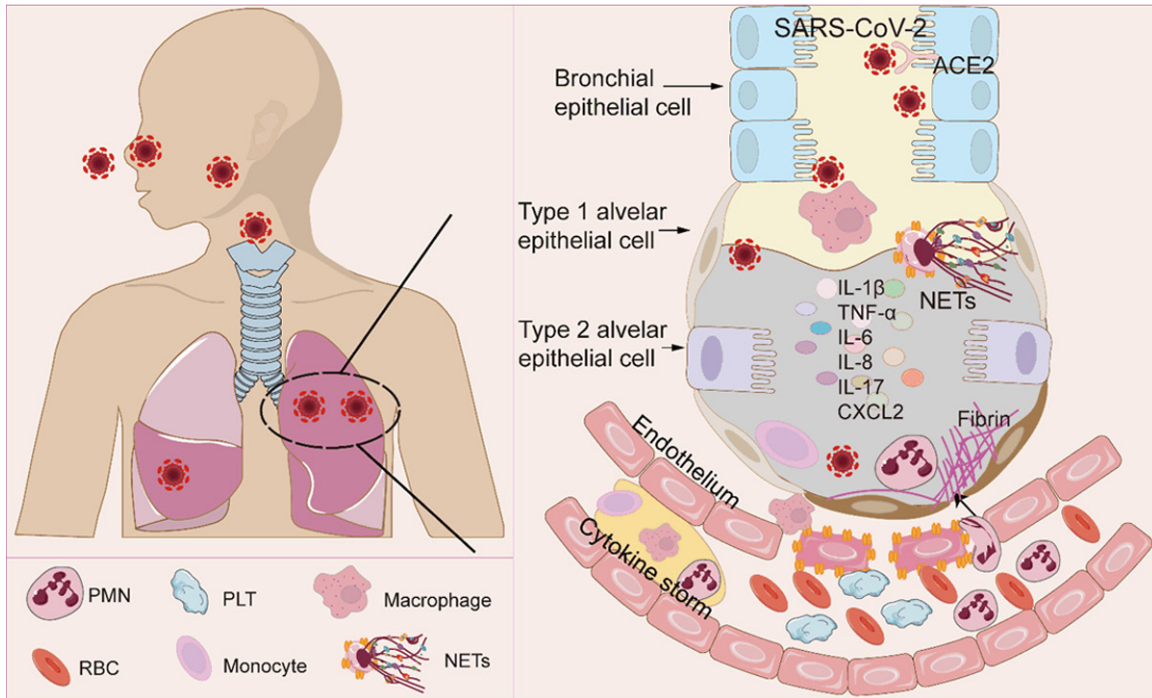


Figure 2. The mechanisms of COVID-19 damages to the lung. The COVID-19 invades the lung through the respiratory tract, enters the alveolar epithelial cells via the ACE2 receptor, activates the immune system, secretes pro-inflammatory factors and chemokines, which recruit leukocyte and promote virus clearance. However, as the number of virus increases, immune cells continue to secrete cytokines and promote the formation of cytokine storm. In patients with severe COVID-19, the virus invades blood vessels, damages vascular endothelial cells, increases vascular permeability, causes cells and proteins to enter the alveoli, accelerates alveolar endothelial cell damage and fiber deposition, and aggravates alveolar damage, forming a vicious circle. ACE2: angiotensin-converting enzyme 2; CXCL: Chemokines; IL: interleukin; TNF: tumor necrosis factor; NETs: neutrophil extracellular traps; PLT: Platelet; PMN: Polymorphonuclear neutrophil; RBC: Red blood cell; TNF: tumor necrosis factor.

as shortness of breath, dyspnea, shock and thrombosis. The levels of CRP, LDH, cytokines, D-dimer are increased significantly, which lead to coagulopathy [2, 17, 18].

The autopsy revealed that there was a large amount of inflammatory cells infiltration in the patient's lung, alveolar damage, and capillary congestion. Extensive fibers deposits and microthrombus formation were seen at the injury site, these conditions are more common in critically ill patients. In addition, quantities of infiltrating inflammatory cells secreted cytokines, which promoted the recruitment of inflammatory cells, then cytokines caused tissue and endothelial cells damage. The broken connection between damaged endothelial cells, coupled with vascular congestion, caused cells and proteins to infiltrate into the alveolar cavity, aggravating alveolar damage and fiber deposition, resulting in a worsening of the disease (Figure 2).

Cytokine storm

When SARS-CoV-2 invades, the immune system responds quickly, releasing pro-inflammatory cytokines and chemokines to promote defense against the virus [19]. Tamim et al. divided this process into stage I (early stage), stage II (IIa pulmonary symptom without hypoxia, IIb pulmonary symptom with hypoxia), stage III (hyper-inflammation period) [20]. He believed virus amplified and multiplied in the body, stimulating immune cells to release many pro-inflammatory cytokines, such as interleukin (IL)-1 β , IL-6 and tumor necrosis factor- α (TNF- α), and formed cytokine storm in severe and critically ill patients.

IL-6 was positively correlated with COVID-19 progression and significantly increased in critically ill patients, up to about 10 times compared with normal people or mild patients, which was closely related to the prognosis of

Thrombosis and anticoagulation of COVID-19

Table 1. Association between coagulation abnormalities or markers of thrombosis

| Study | Group | Leucocytes ($\times 10^9/L$) | Neutrophil ($\times 10^9/L$) | D-dimer ($\mu g/mL$) | Fibrinogen (g/L) | CRP (mg/L) | APTT (s) |
|--------------------------|-------------------------|--------------------------------|--------------------------------|------------------------|------------------|------------------|----------|
| Zhang et al. [9] (n=343) | D-dimer <2.0 | 6.5±4.4 | 3.5 (2.7, 4.8) | 0.4 (0.2, 0.7) | 4.1 (3.1, 5.1) | 1.7 (0.3, 16.6) | 29.6±4.3 |
| | D-dimer >2.0 | 7.4±3.7 | 4.7 (3.4, 7.3) | 4.8 (3.0, 11.9) | 4.3 (3.2, 5.6) | 13.6 (1.8, 62.8) | 28.8±5.2 |
| | P value | 0.09 | <0.001 | <0.001 | 0.54 | <0.001 | 0.21 |
| Cui et al. [33] (n=81) | VTE (n=20) | 7.8±3.1 | NR | 5.2±3.0 | NR | NR | 39.9±6.4 |
| | Non-VTE (n=61) | 6.6±2.6 | NR | 0.8±1.2 | NR | NR | 35.6±4.5 |
| | P value | 0.12 | NR | <0.001 | NR | NR | 0.001 |
| Wang et al. [34] (n=65) | Mild (n=30) | 5.2 (2.4) | 3.8 (2.4) | 1.6 (3.0) | NR | 53.6 (57.7) | NR |
| | Severe (n=20) | 6.9 (3.8) | 5.7 (3.7) | 4.7 (7.4) | NR | 91.8 (77.8) | NR |
| | Extremely severe (n=15) | 8.7 (4.1) | 7.7 (3.9) | 6.9 (8.4) | NR | 114.9 (62.5) | NR |
| | P value | <0.001 | <0.001 | <0.001 | NR | <0.01 | NR |
| Han et al. [35] (n=134) | Controls (n=40) | NR | NR | 0.3±0.2 | 2.9±0.5 | NR | 28.7±3.0 |
| | SARS-CoV-2 (n=94) | NR | NR | 10.4±25.3 | 5.0±1.5 | NR | 29.0±2.9 |
| | P value | NR | NR | <0.001 | <0.001 | NR | <0.001 |
| Yang et al. [36] (n=93) | non-severe (n=69) | 6.4±2.4 | 4.55±0.21 | 0.5±0.4 | NR | 20.1±24.5 | NR |
| | severe (n=24) | 9.1±5.6 | 7.73±5.4 | 16.6±23.1 | NR | 53.9±60.1 | NR |
| | P value | <0.01 | <0.01 | <0.01 | NR | <0.01 | NR |

APTT, activated partial thromboplastin time; CRP, C-reactive protein; NR, no report; VTE, Venous Thrombus Embolism.

the disease [5]. IL-6 can promote the macrophage activation, then leading to release multi-proinflammatory cytokines and recruit leucocyte and fibroblasts into the lung. This makes the deposition of fibrin, aggravating damage of the lung [21].

IL-17 is a member of the multifunctional cytokine family, mainly produced by activated T cells, neutrophils and mast cells, and participates in a variety of inflammatory diseases and autoimmune diseases [22-24]. Under physiological conditions, IL-17 is responsible for skin and mucosal immunity of the mouth, lung, airways, gastrointestinal tract, and vagina, where these organs widely express IL-17 receptors. It induces mucosal epithelia secrete antimicrobial peptides to maintain the epithelial barrier, directly removing pathogens or preventing microbial invasion [25, 26]. Regrettably, excessive IL-17 aggravates the inflammatory level and induces the airway epithelium to secrete chemokine ligand (CXCL) 1, CXCL5, IL-8, IL-6, granulocyte colony stimulating factor (G-CSF) and granulocyte and macrophage colony stimulating factor (GM-CSF), recruiting granulocytes to infiltrate into the lung [27, 28]. In the animal model of lipopolysaccharide (LPS) induced ARDS in mice, IL-17 was increased the severity of lung injury and closely related to the low survival rate of mice. Excessive IL-17 accelerated the severity of lung injury, which is closely related to the low survival rate of mice [29]. The level of IL-17 was

positively associated with mortality in severely ill COVID-19 patients [30]. IL-17 promotes the release of various proinflammatory factors, and inflammatory cytokines can magically promote the production of IL-17 [31]. The interaction of IL-17 and proinflammatory cytokines form a vicious circle, which may lead to accelerate the formation of COVID-19 cytokine storm. Due to the close relationship between inflammation and coagulation, we next explore the “contribution” of the cytokine storm formed by COVID-19 in thrombosis.

Abnormal coagulopathy in COVID-19

Most studies have found that D-Dimer, C-reactive protein (CRP) and fibrinogen are significantly increased in COVID-19 patients. D-dimer was positively related to the severity of the disease and can predict the risk of deep vein thrombosis and patient death [8, 9, 32]. **Table 1** summarizes the current specific changes in different coagulation parameters after infected [9, 33-36]. As the severity of the disease progress, the coagulation indicators were increased significantly in COVID-19 patients. This makes the clinical administration of COVID-19 patients a reasonable basis for using anticoagulation therapy, and by monitoring changes in laboratory indicators, timely adjustment of anticoagulant dose.

In severe COVID-19 patients, through comparing patients with or without LMWH for antico-

agulation treatment, and observing the 28-day mortality, they found that when critical patients meet the SIC score >4 or D-dimer >6 times the normal level, anticoagulant therapy could improve the prognosis and reduce their mortality in critically ill patients [11]. Unfortunately, some patients still have a high incidence of thrombosis during using of anticoagulants such as LMWH [37]. **Table 2** lists some studies of patients who experienced thrombosis with or without anticoagulation. Although most patients were treated with prophylactic anticoagulation, thromboembolism of varying degrees still occurred, and bleeding happened in some patients [12, 33, 37-41]. Consequently, the exploration of the thrombosis mechanism of COVID-19 may improve the effectiveness of anticoagulation, reduce the risk of bleeding.

The mechanism of thrombosis

Microparticle storm

Phosphatidylserine (PS), a negatively charged phospholipid, is one of the major phospholipid components, which makes up the cell membrane and usually locates in the intracellular membrane. When the cell is stimulated or activated or apoptosis, the cell membrane occurs to remodel, then PS is flipped from intracellular to extracellular [42]. It is the main cofactor for hemostasis and thrombosis. There are two PS-recognition motifs found among coagulation factors: the Gla domain and discoidin-like C2 domain [43, 44]. Factors VII, IX, X, and prothrombin, contain an N-terminal Gla domain and factors V and VIII have a C2 domain responsible for targeting to PS. The two domains will provide a binding site for activated factor X (FXa) and prothrombin complexes to promote thrombosis [43, 45]. Microparticles (MPs) are that cells remodel during apoptosis or activation, releasing a small vesicle, about 100-1000 nm, and contain large amounts of PS [46, 47]. Studies showed that MPs have a variety of biological activities and play a role in coagulation, inflammation, angiogenesis and intercellular communication [48-50]. In our previous study, we found that leukocytes, red blood cells, and monocytes release large amounts of MPs during the active phase in patients with inflammatory bowel disease, through labeling cell-specific expression molecules and using flow cytometry to determine the source of MPs in the blood [51]. In patients

with oral squamous cell carcinoma, especially patients with stage III/IV, many procoagulant MPs were detected in the circulation [52]. MPs plays an important role in the formation of procoagulant state in sepsis patients [53].

IL-6 and IL-8 is associated with changes in the membrane structure of red blood cells and platelets. Pre-blocking IL-6 and IL-8 would reduce the level of MPs [54]. Unfortunately, there are many cytokines in COVID-19 patients, and cytokine storm are formed in severe and critically ill patients. Abnormal hypercoagulability and thrombosis can be seen in ICU patients and non-surviving patients, which make us to guess the formation of thrombus may be due to a large number of cytokines stimulating circulating blood cells, causing the activation of blood cells and even apoptosis, which lead to the release of cell particles, forming a "Micro-particle storm" to promote the formation of inflammatory thrombus (**Figure 3**).

It was found that the membrane of MPs contains the tissue factor (TF). TF combines with activated factor VII (FVIIa) to effectively activate FX and FIX and promote the production of thrombin [55, 56]. Under normal circumstance, TF exists outside the blood vessel, and the presence of TF hardly is detected in circulation to avoid the occurrence of unnecessary coagulation reaction [57]. When the body is stimulated, large amounts of TF will be detected in the blood in a variety of diseases, such as sepsis and angina. Bogdanov VY and his colleagues believed that there were many TF in the blood circulation, and MPs was the main carrier of TF [58]. Normally TF, however, existed in encrypted form, and PS could decrypt the encrypted TF into active TF to participate in thrombosis [59]. In our experiment, the pro-coagulant state was improved after adding anti-TF antibody, and the pro-coagulant activity was significantly reduced when PS was blocked [60, 61]. Thus, we have reasons to believe that PS and TF play a major role in the procoagulant activity of MPs, and PS synergistically promotes the activation of the exogenous coagulation pathway of thrombosis by decrypting TF. Therefore, anticoagulant therapy for COVID-19 can inhibit the formation of thrombin complex by targeted blocking of PS on MPs and blood cells, reducing the occurrence of thrombosis.

Thrombosis and anticoagulation of COVID-19

Table 2. Studies and main findings for thrombosis events in COVID-19 patients

| Study | Patients | Thrombosis type | Main findings | Bleeding events | Therapy |
|---------------------|----------|--|---|---|--|
| Hanny et al. [37] | 400 | Venous thrombosis Arterial thrombosis | In critically ill COVID-19 patients, imaging confirmed VTE 10.4% (95% CI, 5.9-16.6%); The incidence in non-critically ill patients was 3.5% (95% CI, 1.6-6.6%). And arterial thrombosis: critical ill COVID-19 3.46%; Non-critical illness 1.3% | The incidence of massive hemorrhage in critically ill patients was 5.6% (2.4-10.7%) | Standard prophylactic dose of ordinary heparin or low molecular weight heparin |
| Cui et al. [33] | 81 | Venous thrombosis | Venous thromboembolism occurred in 25% of the 81 critically ill patients | NR | None |
| Klok et al. [38] | 184 | Venous thrombosis Arterial thrombosis | VTE accounts for 27% (95% CI 17-37%) of COVID-19 patients in ICU | NR | Use at least a prophylactic dose of anticoagulant |
| Corrado et al. [39] | 362 | Venous thrombosis | Incidence of thrombosis was 7.7% in all patients | NR | Prophylactic dose of LMWH |
| Jean-F et al. [12] | 26 | Venous thrombosis | Overall VTE rate was 69%. Among COVID-19 patients receiving therapeutic anticoagulation, the incidence of thromboembolic events was high, with 56% VTE, including 6 cases of pulmonary embolism | NR | 8 patients received preventive anticoagulation, 18 were treated with therapeutic anticoagulation |
| Tao et al. [40] | 1099 | Venous thrombosis | There were 407 patients with COVID-19 who were at high risk for Venous thromboembolism | 44 (11%) of 407 also had a high risk of bleeding | Only ten (7%) of 140 patients for whom anticoagulation data were available in our cohort (nine were given heparin and one rivaroxaban) |
| Bin et al. [41] | 48 | Venous thrombosis | Lower extremity DVT were detected in 41 patients (85.4%), with 36 (75%) isolated distal DVT and 5 (10.4%) proximal DVT | NR | 30-40 mg LMWH |

DVT, Deep Venous Thrombus; ICU, Intensive Care Unit; LMWH, Low molecular weight heparin; NR, no report; VTE, Venous Thrombus Embolism.

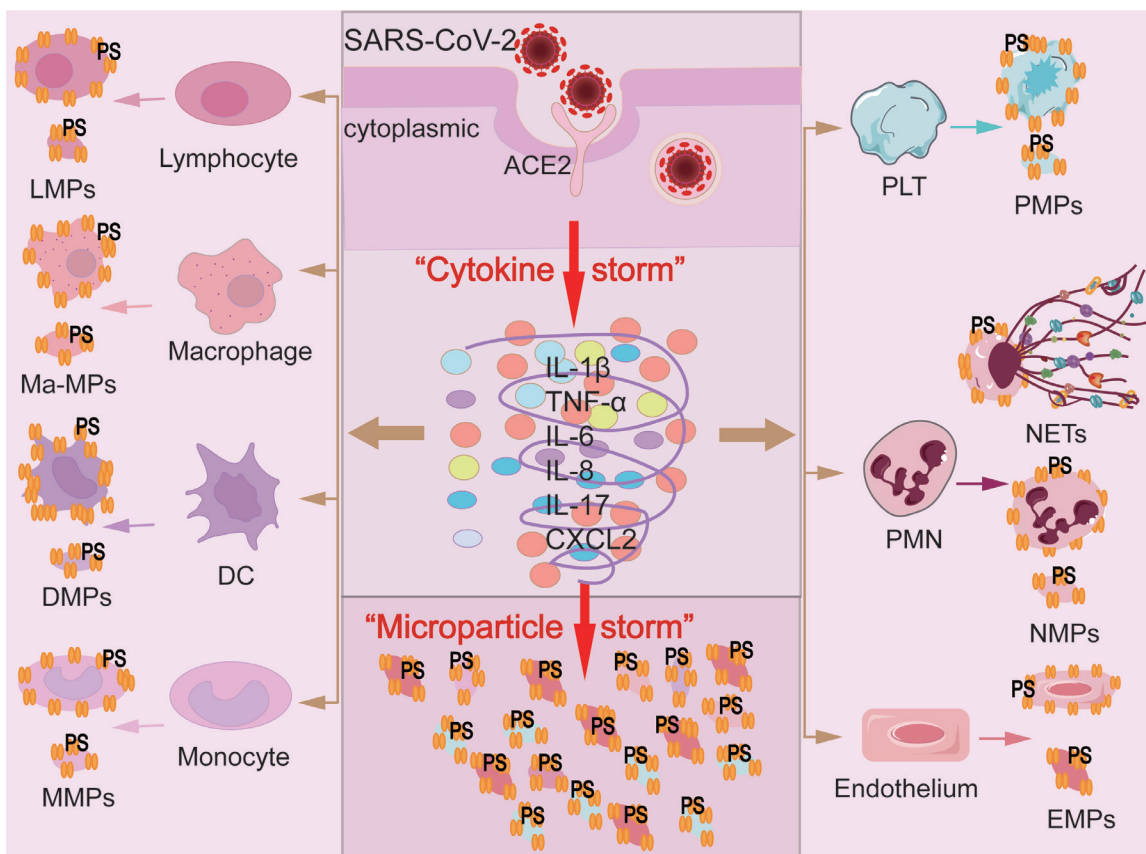


Figure 3. Cytokines trigger microparticle storm. After virus invasion, it stimulates the immune system to release a large number of cytokines and cytokine storm formation, cell factor effects on blood cells (lymphocytes, monocytes, macrophages, dendritic cells, neutrophils and platelets), endothelial cells, the cell activation, large numbers of cells release coagulant activated particles, forming similar cytokine storm “Microparticle storm”. ACE2: angiotensin-converting enzyme 2; DC: Dendritic Cells; MPs: Microparticles; PLT: Platelet; PMN: Polymorphonuclear neutrophil; PS: Phosphatidylserine.

NETs promote thrombosis

As a main member of the innate immune system, neutrophils are quickly recruited to the injury site when the body is attacked, playing a defensive role [62]. In 2004, it was first discovered that neutrophils were stimulated and then released a network structure outside the cell, called neutrophil extracellular traps (NETs) [63]. Bacteria, fungi and other microorganisms can stimulate neutrophils to release NETs. NETs consist of a DNA skeleton and a variety of granular enzymes, including histone, myeloperoxidase (MPO), elastase and matrix metalloproteinases [64]. It is a double-edged sword, not only exerts the defensive function of capturing and killing bacteria, but also activates the coagulation cascade reaction, such as the DNA skeleton activates the coagulation contact system to promote coagulation, and the histone inhibits the activation of protein C to promote coagu-

lation [65, 66]. In addition, NETs also cause damage to surrounding tissue, delay wound healing, and aggravate disease progression [67, 68]. Inflammatory factors stimulate neutrophils to release NETs, which are abundant in infection and inflammation sites [63, 68]. IL-8 induces the release of elastase in a concentration-dependent manner, and IL-1 β not only facilitate the process of IL-8 stimulating neutrophils, but also induce the release of MPO from neutrophils [69, 70]. TNF- α promotes the respiratory burst of neutrophils, and inflammatory factors stimulate neutrophils to release NETs, which are abundant in infection and inflammation sites [71]. The level of NETs was significantly reduced by neutralizing the effects of inflammatory cytokines through adding antibodies to the plasma of patients [72].

NETs are positively correlated with the severity and survival rate in COVID-19 patients. One

study found that plasma markers of NETs such as cell free-DNA (cf-DNA) and MPO-DNA were significantly elevated in severe and critically ill COVID-19 patients [73]. This study suggests that COVID-19 patients are more likely to produce NETs, and that overall levels of NETs increase with disease progression. The study of Elizabeth et al. reached the same conclusion and made further research. They found a large amount of neutrophil infiltration, NETs and platelet co-aggregation can be seen in the lung through autopsy. The interaction of NETs and platelets promotes the formation of immune thrombosis. Moreover, excessive NETs may further aggravate the lung damage [74]. In addition, the role of NETs in activating the coagulation pathway to promote thrombosis has been widely accepted, adding NETs inhibitors improved blood hypercoagulability and reduced thrombosis in animal models [75, 76]. Therefore, NETs may be involved in the hypercoagulable state and thrombosis in COVID-19 patients. Inhibiting the NETs production, or hydrolyzing its structure, or blocking the granular proteases attached to NETs, may be very helpful in reducing thrombosis in COVID-19 patients.

Endothelial cell damage

Endothelial dysfunction is the main factor of microvascular disease, which changes the balance of the blood vessels to promote vasoconstriction, subsequently leads to organ ischemia, inflammation, and tissue edema, forming a procoagulant state [77]. COVID-19 infects endothelial cells through ACE2 receptor to cause endothelial dysfunction. Autopsy of patients who died of COVID-19 revealed that the lung microvascular endothelial cells were damaged, and the intercellular connection was broken [78, 79]. Numerous of lymphocytes and macrophages infiltration and microthrombus were seen in the lung. In addition, abnormal neovascularization occurred in the lung, and surprisingly the structure of the neovascularization was disordered. Although endothelial cells proliferated in large quantities, they could not arrange in a normal manner, making many neovascularization disturbed and being unable to function normally. Endothelial integrity is an important factor in the prevention of thrombosis. When endothelial cells are injured, they express E-selectin and von Willebrand factor

(vWF), which promotes platelet adhesion and aggregation, then form thrombosis [80]. As COVID-19 causes endothelial cell damage and apoptosis, it may induce the release of MPs from endothelial cells, and the exposure of PS on the endothelial cell surface, and promote the binding of coagulation factors, leading to thrombosis. In addition, IL-6 and TNF- α motivate endothelial cells, which facilitate endothelial cells to release procoagulant soluble TF [81]. After COVID-19 infection, the release of pro-inflammatory factors causes endothelial cell damage, which may induce endothelial cells to release MPs, expose PS on the surface of endothelial cell, promote the binding of coagulation factors, and lead to thrombosis, further experiments are needed to confirm our speculation [82, 83].

Previous studies shown that NETs cause endothelial cell damage, especially attached histones, which have a toxic effect on endothelial cells, causing endothelial cells to shrink and activate. The deposition of coagulation factors Xa and Va and fibrin on endothelial cells was observed. By targeting inhibition of histone or hydrolyzation of NETs structure reversed endothelial cell damage on the influence of coagulant activity [72, 84]. Yu Zou and his colleagues compared the two groups of COVID-19 patients who were with mechanical ventilation and no mechanical ventilation, they found that the level of NETs in the mechanical ventilation group was significantly increased, indicating that NETs increased with the severity of the disease [75].

Hypoxia plays a vital role in promoting inflammation and endothelial damage [85]. It promotes the hypoxia-inducible factor (HIF) transcription in the nucleus [86]. HIF promotes the release of inflammatory factors by activating NF- κ B pathway and aggravates local inflammation [87]. Hypoxia damages endothelial cell, destroys intercellular connection and the endothelial barrier, then promotes microleakage, high viscous blood, and thrombosis [88]. Moreover, it not only activates the exogenous coagulation pathway, promotes the production of PAI-1 and inhibits the fibrinolytic system, but also inhibits the anticoagulation system [89, 90]. Regrettably, studies have shown that hypoxia is one of the important symptoms of COVID-19 [91, 92]. Although oxygen is used to

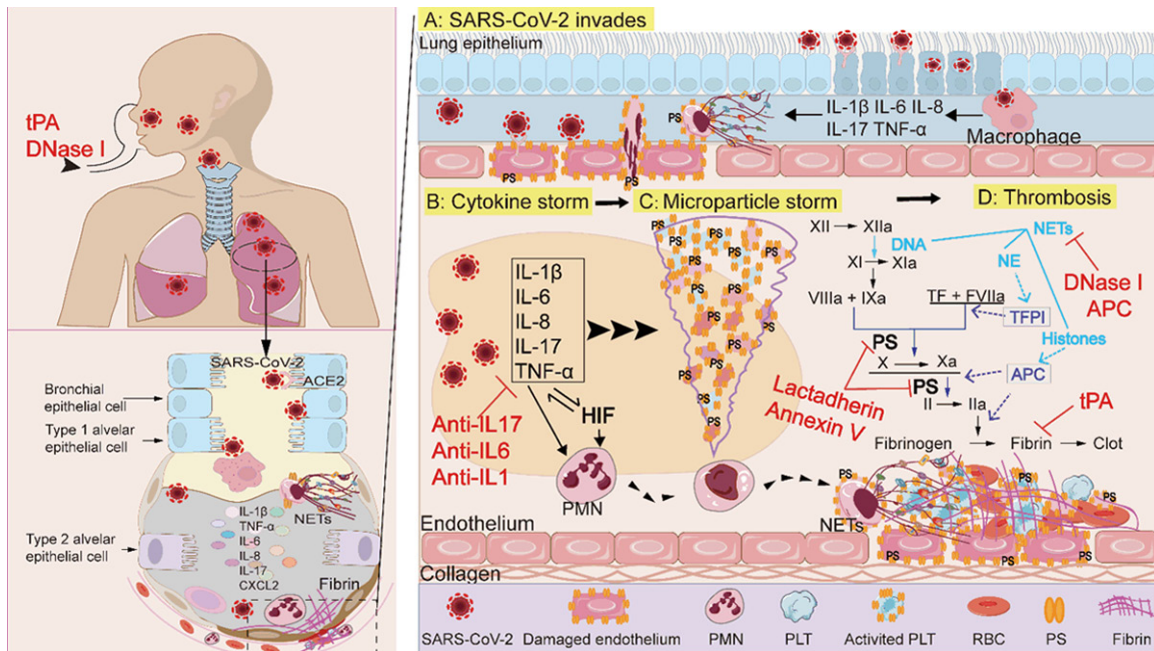


Figure 4. New targets for the treatment of thrombosis caused by COVID-19. Nebulized inhaled drugs improve pulmonary fibrosis, reduce lung tissue damage, and combine multiple targets to inhibit thrombosis, jointly reduce patient mortality and improve prognosis. Viruses damage the lung and promote the infiltration of inflammatory cells. Many neutrophils release NETs to further aggravate lung injury. The deposition of fibrin and decreased fibrinolytic capacity may lead to pulmonary fibrosis, which facilitates the deterioration of respiratory function. Inhaling tPA and NETs through atomization can reduce the degree of lung fibrosis and tissue damage, then improve respiratory function. Combining with inhibiting cytokines, PS on MPs and NETs, the activation of the coagulation pathway is suppressed, reduce the production of thrombin, and finally stop thrombosis. APC: activated protein C; NETs: neutrophil extracellular traps; PLT: Platelet; PMN: Polymorphonuclear neutrophil; PS: phosphatidylserine; tPA: tissue plasminogen activator.

increase oxygen saturation, it is still difficult to reverse. Accordingly, improving the hypoxic symptom of patients will reduce endothelial damage and decrease the risk of thrombosis.

In addition to the important role of blood cells in thrombosis, hemodynamics is also a mechanism of thrombosis formation in the Virchow's Triad [93, 94]. Bo Zhou reports a severe case of lower extremity deep venous thrombosis with arterial occlusion. Due to the endothelial injury of COVID-19 patients, if the activity is restricted, the shear rate of blood flow will be reduced. This condition will promote the adhesion of platelets, blood stasis, and thrombosis, especially the risk of venous thrombosis in the lower limbs is greatly increased [95]. Because severe COVID-19 patients are mainly bedridden, with limited voluntary activity, the blood flow shear rate is reduced. This condition will promote platelet adhesion, congestion and thrombosis, which will make the severe COVID-19 patients worse.

Therapy

According to the current situation of anticoagulant therapy for COVID-19 and our understanding of the mechanism of thrombosis, we propose our new target for anticoagulant therapy (Figure 4).

Cytokines inhibitor

As mentioned above, cytokine storm plays an important role for disease progression in COVID-19. The inflammatory response within 24 hours after admission may be related to the severity of the disease. So, suppression of inflammatory factors will delay disease progression [96]. Inhibition of IL-6 is a vigorous anti-inflammatory method because it accounts for a relatively large proportion. Tocilizumab, a commonly used inhibitor, has been shown to reduce inflammation levels and improve symptoms and prognosis in patients. In moderate COVID-19 patients, it was found that tocilizum-

ab was well tolerated and no adverse reactions [97]. Although tocilizumab can reduce inflammation, there are side effects with liver damage and gastrointestinal ulcers in critically ill patients [98, 99]. Kimmig LM et al. found inhibition of IL-6 may damage the clearance of the virus, secondary infection, leading to death in severe COVID-19 [100]. Therefore, it is important to carefully control the dosage and timing of use to maximize the benefits in severe and critically ill patients.

Anti-coagulant therapy

Heparin and LMWH, commonly anticoagulant drugs, bind to antithrombin III which mainly inhibits the activation of FX and the production of thrombin to achieve the purpose of anticoagulation [101]. Due to the high incidence of COVID-19 thrombosis, anticoagulation treatment is routinely administered for COVID-19. Once the diagnosis is confirmed, preventive anticoagulation is performed, and the dose of heparin is adjusted according to changes in the condition. However, its dosage is still controversial. Some patients have bleeding after medication, but compared with the serious consequences caused by thrombosis, the risk of bleeding can be temporarily ignored when used in a short time [32, 33]. Heparin also has anti-inflammatory effects through inhibiting the production of inflammatory factors, such as IL-6, IL-8, TNF- α [102, 103]. The heparin can counteract excessive inflammatory factors in the body and reduce inflammation and inflammation-promoted hypercoagulability in COVID-19. In addition, heparin can also protect the endothelium and reduce vascular leakage [101], playing a role in many ways to inhibit thrombosis. Therefore, the early use heparin will greatly reduce the occurrence of thrombosis.

Platelets counts varies in different COVID-19 studies. Studies reported thrombocytopenia in severe patients, although the decline was not serious, which is a sharp contrast to the disseminated intravascular coagulation [17, 104]. The reason for the difference may be the activation of platelets due to the excessive inflammatory environment [105]. Most anti-platelet drugs bind irreversibly to platelets, and the corresponding inhibitors have a relatively long half-life. In addition, the lack of relatively sensitive indicators can, quickly and effectively, detect

changes in platelets [106]. Therefore, when using platelet inhibitors, relevant indicators should be closely monitored.

NETs inhibitors

Inhibiting the NETs production and hydrolyzing the structure and components may decrease the activation of the coagulation pathway (**Figure 4**). Studies have shown that NADPH oxidase and peptidyl dearginase 4 inhibitors can reduce NETs production [107, 108]. However, it may increase the risk of infection. DNase hydrolyzes the NETs structure, destroying the ability of NETs bind cells and inhibiting DNA activating endogenous clotting pathways [109]. Histone antibody is used to block the cytotoxic histones, prevent tissue damage, promote the activation of protein C, and improve the anticoagulation ability [110]. In addition, due to the large amount of granular protein attached to NETs, the accumulation of local tissues aggravates tissue damage. Many neutrophils infiltrated in the lung, fibrin deposition, inhibit lung respiratory function. Inhalation of tPA and DNase I by atomization may promote pulmonary fibrinolysis, improve respiratory function [111]. Therefore, targeted NETs may alleviate the hypercoagulable state of COVID-19 and reduce thrombosis.

Targeting PS

The central point of the reaction is PS on the membrane of MPs and activated cells. With the invasion of the virus, the immune system overreacts and releases numbers of cytokines, which will cause quantities of cell activation, mutilation, apoptosis and even necrosis, then cells will produce PS on the surface, forming a microparticle storm [112]. Because MP storm provides an anchor platform for coagulation factors. Previous studies have confirmed that annexin V and lactadherin can bind PS interrupt the cascade of coagulation [74, 90]. Therefore, we believe that inhibitors, annexin V and lactadherin, will significantly reduce the incidence of thrombosis in COVID-19 patients without increasing the risk of bleeding. In addition, our previous research on the anticoagulant effect of lactadherin was mainly confirmed in vitro experiments. Whether its anticoagulant effect in vivo is affected by the microenvironment is still unknown. It is necessary to study the in vivo anticoagulation with lactadherin,

and we believe that inhibitors, annexin V and lactadherin, will significantly reduce the incidence of thrombosis in COVID-19 patients without increasing the risk of bleeding.

Improving the state of hypoxia via early oxygen absorption

Hypoxia is a common manifestation of severe or critically ill COVID-19 patients [113, 114]. Some studies have suggested that asymptomatic hypoxia occurs in mild patients, and the body is in a state of “happy hypoxia” or “silent hypoxia”. Chronic silent hypoxia may not only motivate circulating blood cells, but also cause chronic damage to endothelial cells, promote PS exposed, release microparticles, and promote thrombosis. Thus, we believe that low-flow oxygen inhalation will ameliorate the hypoxia state of the mild patient’s internal environment. According to the progress of patients, it will be increased the oxygen intake, and performed mechanical ventilation if necessary, to minimize the cell activation and endothelial cell damage caused by hypoxia, and diminish the risk factors of thrombosis.

Conclusion

In summary, we described the pathophysiological and the thrombotic mechanism in COVID-19. We proposed that the microparticle storm and the PS may play a significant role in thrombosis for the first time. Although a variety of mechanisms causing thrombosis have been analyzed, the combined effect of multiple factors will promote PS exposed of cells and eventually activate the coagulation pathway. By targeting PS, it can inhibit the generation of thrombin, reduce the hypercoagulable state and decrease thrombosis, which may provide a new treatment direction for the current inherent model of anticoagulation programs in COVID-19 patients. We believe that the assessment of different patients, combined with multi-target inhibition of the activation of the coagulation pathway, the formulation of a reasonable anticoagulant program, to decrease the occurrence of thrombosis events, will greatly diminish the mortality of patients and improve the prognosis.

Acknowledgements

This work was supported by grants from the National Natural Science Foundation of China

(81670128 and 81873433) and the Scientific Research Innovation Foundation of the First Hospital of Harbin Medical University (2020 M12).

Disclosure of conflict of interest

None.

Abbreviations

ACE2, Angiotensin-converting enzyme 2; APC, activated protein C; ARDS, Acute respiratory distress syndrome; cf-DNA, Cell free-DNA; COVID-19, Coronavirus disease-2019; CRP, C-reactive protein; ECs, endothelial cells; FVIIa, Activated factor VII; FXa, Activated factor X; GCSF, Granulocyte colony stimulating factor; GM-CSF, Granulocyte and macrophage colony stimulating factor; HIF, hypoxia-inducible factor; ICU, Intensive care unit; IL, Interleukin; LDH, Lactate dehydrogenase; LMWH, Low molecular weight heparin; LPS, Lipopolysaccharide; MERS, Middle East Respiratory syndrome; MPO, myeloperoxidase; MPs, Microparticles; NETs, Neutrophil extracellular traps; NF- κ B, Nuclear factor kappa-B; PLT, Platelet; PMN, Polymorphonuclear neutrophil; PS, Phosphatidylserine; RBC, Red blood cell; SARS, Severe Acute respiratory syndrome; SARS-CoV-2, Severe acute respiratory syndrome coronavirus type 2; TF, Tissue factor; TFPI, tissue factor pathway inhibitor; TNF- α , Tumor necrosis factor- α ; vWF, Von Willebrand factor.

Address correspondence to: Tenglong Hu, Department of Stomatology, The First Hospital of Harbin Medical University, Harbin, China. E-mail: Flylong_26@sina.com; Jialan Shi, Department of Hematology, The First Hospital of Harbin Medical University, Harbin, China; Department of Research and Medicine, VA Boston Healthcare System, Brigham and Women’s Hospital, Harvard Medical School, Boston, Massachusetts, USA. E-mail: jialan_shi@hms.harvard.edu; jialan_shi@dfci.harvard.edu

References

- [1] Zhou F, Yu T, Du R, Fan G, Liu Y, Liu Z, Xiang J, Wang Y, Song B, Gu X, Guan L, Wei Y, Li H, Wu X, Xu J, Tu S, Zhang Y, Chen H and Cao B. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *Lancet* 2020; 395: 1054-1062.
- [2] Wang D, Hu B, Hu C, Zhu F, Liu X, Zhang J, Wang B, Xiang H, Cheng Z, Xiong Y, Zhao Y, Li Y,

Thrombosis and anticoagulation of COVID-19

- Wang X and Peng Z. Clinical characteristics of 138 hospitalized patients with 2019 novel coronavirus-infected pneumonia in Wuhan, China. *JAMA* 2020; 323: 1061-1069.
- [3] Jose RJ and Manuel A. COVID-19 cytokine storm: the interplay between inflammation and coagulation. *Lancet Respir Med* 2020; 8: e46-e47.
- [4] Pedersen SF and Ho Y. SARS-CoV-2: a storm is raging. *J Clin Invest* 2020; 130: 2202-2205.
- [5] Chen X, Zhao B, Qu Y, Chen Y, Xiong J, Feng Y, Men D, Huang Q, Liu Y, Yang B, Ding J and Li F. Detectable serum severe acute respiratory syndrome coronavirus 2 viral load (RNAemia) is closely correlated with drastically elevated interleukin 6 level in critically ill patients with coronavirus disease 2019. *Clin Infect Dis* 2020; 71: 1937-1942.
- [6] José RJ, Williams AE and Chambers RC. Proteinase-activated receptors in fibroproliferative lung disease. *Thorax* 2014; 69: 190-2.
- [7] Chen X, Yu C, Jing H, Wang C, Zhao X, Zhang J, Zhang S, Liu H, Xie R and Shi J. COVID-19 associated thromboinflammation of renal capillary: potential mechanisms and treatment. *Am J Transl Res* 2020; 12: 7640-7656.
- [8] Panigada M, Bottino N, Tagliabue P, Grasselli G, Novembrino C, Chantarangkul V, Pesenti A, Peyvandi F and Tripodi A. Hypercoagulability of COVID-19 patients in intensive care unit: a report of thromboelastography findings and other parameters of hemostasis. *J Thromb Haemost* 2020; 18: 1738-1742.
- [9] Zhang L, Yan X, Fan Q, Liu H, Liu X, Liu Z and Zhang Z. D-dimer levels on admission to predict in-hospital mortality in patients with Covid-19. *J Thromb Haemost* 2020; 18: 1324-1329.
- [10] Tang N, Li D, Wang X and Sun Z. Abnormal coagulation parameters are associated with poor prognosis in patients with novel coronavirus pneumonia. *J Thromb Haemost* 2020; 18: 844-847.
- [11] Tang N, Bai H, Chen X, Gong J, Li D and Sun Z. Anticoagulant treatment is associated with decreased mortality in severe coronavirus disease 2019 patients with coagulopathy. *J Thromb Haemost* 2020; 18: 1094-1099.
- [12] Llitjos J, Leclerc M, Chochois C, Monsallier J, Ramakers M, Auvray M and Merouani K. High incidence of venous thromboembolic events in anticoagulated severe COVID-19 patients. *J Thromb Haemost* 2020; 18: 1743-1746.
- [13] Connors JM and Levy JH. COVID-19 and its implications for thrombosis and anticoagulation. *Blood* 2020; 135: 2033-2040.
- [14] Smadja DM, Guerin CL, Chocron R, Yatim N, Boussier J, Gendron N, Khider L, Hadjadj J, Goudot G, Debuc B, Juvin P, Hauw-Berlemont C, Augy J, Peron N, Messas E, Planquette B, Sanchez O, Charbit B, Gaussem P, Duffy D, Terrier B, Mirault T and Diehl J. Angiotensin-2 as a marker of endothelial activation is a good predictor factor for intensive care unit admission of COVID-19 patients. *Angiogenesis* 2020; 23: 611-620.
- [15] Wang Q, Zhang Y, Wu L, Niu S, Song C, Zhang Z, Lu G, Qiao C, Hu Y, Yuen KY, Wang Q, Zhou H, Yan J and Qi J. Structural and functional basis of SARS-CoV-2 entry by using human ACE2. *Cell* 2020; 181: 894-904, e9.
- [16] Rothe C, Schunk M, Sothmann P, Bretzel G, Froeschl G, Wallrauch C, Zimmer T, Thiel V, Janke C, Guggemos W, Seilmaier M, Drosten C, Vollmar P, Zwirgmaier K, Zange S, Wölfel R and Hoelscher M. Transmission of 2019-nCoV infection from an asymptomatic contact in Germany. *N Engl J Med* 2020; 382: 970-971.
- [17] Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, Zhang L, Fan G, Xu J, Gu X, Cheng Z, Yu T, Xia J, Wei Y, Wu W, Xie X, Yin Wen, Li H, Liu M, Xiao Y, Gao H, Guo L, Xie J, Wang G, Jiang R, Gao Z, Jin Q, Wang J and Cao B. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet* 2020; 395: 497-506.
- [18] Lauer SA, Grantz KH, Bi Q, Jones FK, Zheng Q, Meredith HR, Azman AS, Reich NG and Lessler J. The incubation period of coronavirus disease 2019 (COVID-19) from publicly reported confirmed cases: estimation and application. *Ann Intern Med* 2020; 172: 577-582.
- [19] Diao B, Wang C, Tan Y, Chen X, Liu Y, Ning L, Chen L, Li M, Liu Y, Wang G, Yuan Z, Feng Z, Zhang Y, Wu Y and Chen Y. Reduction and functional exhaustion of T cells in patients with coronavirus disease 2019 (COVID-19). *Front Immunol* 2020; 11: 827.
- [20] Alsuliman T, Alasadi L, Alkharat B, Srour M and Alrstom A. A review of potential treatments to date in COVID-19 patients according to the stage of the disease. *Curr Res Transl Med* 2020; 68: 93-104.
- [21] McGonagle D, Sharif K, O'Regan A and Bridgewood C. The role of cytokines including interleukin-6 in COVID-19 induced pneumonia and macrophage activation syndrome-like disease. *Autoimmun Rev* 2020; 19: 102537.
- [22] Schwarzenberger P, Huang W, Ye P, Oliver P, Manuel M, Zhang Z, Bagby G, Nelson S and Kolls JK. Requirement of endogenous stem cell factor and granulocyte-colony-stimulating factor for IL-17-mediated granulopoiesis. *J Immunol* 2000; 164: 4783-9.
- [23] Lin AM, Rubin CJ, Khandpur R, Wang JY, Riblett M, Yalavarthi S, Villanueva EC, Shah P, Kaplan MJ and Bruce AT. Mast cells and neutrophils release IL-17 through extracellular trap forma-

Thrombosis and anticoagulation of COVID-19

- tion in psoriasis. *J Immunol* 2011; 187: 490-500.
- [24] Reed M, Morris SH, Owczarczyk AB and Lukacs NW. Deficiency of autophagy protein Map1-LC3b mediates IL-17-dependent lung pathology during respiratory viral infection via ER stress-associated IL-1. *Mucosal Immunol* 2015; 8: 1118-30.
- [25] Veldhoen M. Interleukin 17 is a chief orchestrator of immunity. *Nat Immunol* 2017; 18: 612-621.
- [26] Liang SC, Tan X, Luxenberg DP, Karim R, Dunniss-Joannopoulos K, Collins M and Fouser LA. Interleukin (IL)-22 and IL-17 are coexpressed by Th17 cells and cooperatively enhance expression of antimicrobial peptides. *J Exp Med* 2006; 203: 2271-9.
- [27] Apostolidis SA, Crispín JC and Tsokos GC. IL-17-producing T cells in lupus nephritis. *Lupus* 2011; 20: 120-4.
- [28] Laan M, Cui ZH, Hoshino H, Lötval J, Sjöstrand M, Gruenert DC, Skoogh BE and Lindén A. Neutrophil recruitment by human IL-17 via C-X-C chemokine release in the airways. *J Immunol* 1999; 162: 2347-52.
- [29] Li Q, Gu Y, Tu Q, Wang K, Gu X and Ren T. Blockade of interleukin-17 restrains the development of acute lung injury. *Scand J Immunol* 2016; 83: 203-11.
- [30] Schett G, Sticherling M and Neurath MF. COVID-19: risk for cytokine targeting in chronic inflammatory diseases? *Nat Rev Immunol* 2020; 20: 271-272.
- [31] Pearce L, Davidson SM and Yellon DM. The cytokine storm of COVID-19: a spotlight on prevention and protection. *Expert Opin Ther Targets* 2020; 24: 723-730.
- [32] Spiezia L, Boscolo A, Poletto F, Cerruti L, Tiberio I, Campello E, Navalesi P and Simioni P. COVID-19-related severe hypercoagulability in patients admitted to intensive care unit for acute respiratory failure. *Thromb Haemost* 2020; 120: 998-1000.
- [33] Cui S, Chen S, Li X, Liu S and Wang F. Prevalence of venous thromboembolism in patients with severe novel coronavirus pneumonia. *J Thromb Haemost* 2020; 18: 1421-1424.
- [34] Wang F, Hou H, Luo Y, Tang G, Wu S, Huang M, Liu W, Zhu Y, Lin Q, Mao L, Fang M, Zhang H and Sun Z. The laboratory tests and host immunity of COVID-19 patients with different severity of illness. *JCI Insight* 2020; 5: e137799.
- [35] Han H, Yang L, Liu R, Liu F, Wu K, Li J, Liu X and Zhu C. Prominent changes in blood coagulation of patients with SARS-CoV-2 infection. *Clin Chem Lab Med* 2020; 58: 1116-1120.
- [36] Yang A, Li H, Tao W, Yang X, Wang M, Yang W and Liu J. Infection with SARS-CoV-2 causes abnormal laboratory results of multiple organs in patients. *Aging (Albany NY)* 2020; 12: 10059-10069.
- [37] Al-Samkari H, Karp Leaf RS, Dzik WH, Carlson JCT, Fogerty AE, Waheed A, Goodarzi K, Benda-pudi PK, Bornikova L, Gupta S, Leaf DE, Kuter DJ and Rosovsky RP. COVID-19 and coagulation: bleeding and thrombotic manifestations of SARS-CoV-2 infection. *Blood* 2020; 136: 489-500.
- [38] Klok FA, Kruip MJHA, van der Meer NJM, Arbous MS, Gommers DAMPJ, Kant KM, Kaptein FHJ, van Paassen J, Stals MAM, Huisman MV and Endeman H. Incidence of thrombotic complications in critically ill ICU patients with COVID-19. *Thromb Res* 2020; 191: 145-147.
- [39] Lodigiani C, Iapichino G, Carenzo L, Cecconi M, Ferrazzi P, Sebastian T, Kucher N, Studt J, Sacco C, Bertuzzi A, Sandri MT and Barco S; Humanitas COVID-19 Task Force. Venous and arterial thromboembolic complications in COVID-19 patients admitted to an academic hospital in Milan, Italy. *Thromb Res* 2020; 191: 9-14.
- [40] Wang T, Chen R, Liu C, Liang W, Guan W, Tang R, Tang C, Zhang N, Zhong N and Li S. Attention should be paid to venous thromboembolism prophylaxis in the management of COVID-19. *Lancet Haematol* 2020; 7: e362-e363.
- [41] Ren B, Yan F, Deng Z, Zhang S, Xiao L, Wu M and Cai L. Extremely high incidence of lower extremity deep venous thrombosis in 48 patients with severe COVID-19 in Wuhan. *Circulation* 2020; 142: 181-183.
- [42] Zwaal RF and Schroit AJ. Pathophysiologic implications of membrane phospholipid asymmetry in blood cells. *Blood* 1997; 89: 1121-32.
- [43] Stace CL and Ktistakis NT. Phosphatidic acid and phosphatidylserine-binding proteins. *Biochim Biophys Acta* 2006; 1761: 913-26.
- [44] Zwaal RF, Comfurius P and Bevers EM. Lipid-protein interactions in blood coagulation. *Biochim Biophys Acta* 1998; 1376: 433-53.
- [45] Yin Y, Li F, Li S, Cai J, Shi J and Jiang Y. TLR4 influences hepatitis B virus related hepatocellular carcinoma by regulating the Wnt/ β -catenin pathway. *Cell Physiol Biochem* 2017; 42: 469-479.
- [46] Mause SF and Weber C. Microparticles: protagonists of a novel communication network for intercellular information exchange. *Circ Res* 2010; 107: 1047-57.
- [47] Morel O, Jesel L, Freyssinet J and Toti F. Cellular mechanisms underlying the formation of circulating microparticles. *Arterioscler Thromb Vasc Biol* 2011; 31: 15-26.
- [48] Zhou W, Yu L, Fan J, Wan B, Jiang T, Yin J, Huang Y, Li Q, Yin G and Hu Z. Endogenous parathyroid hormone promotes fracture healing by increasing expression of BMP2 through

- cAMP/PKA/CREB pathway in mice. *Cell Physiol Biochem* 2017; 42: 551-563.
- [49] Leonetti D, Reimund J, Tesse A, Viennot S, Martinez MC, Bretagne AL and Andriantsitohaina R. Circulating microparticles from Crohn's disease patients cause endothelial and vascular dysfunctions. *PLoS One* 2013; 8: e73088.
- [50] Nieuwland R, Berckmans RJ, Rotteveel ERC, Maquelin KN, Roozendaal KJ, Jansen PG, ten Have K, Eijsman L, Hack CE and Sturk A. Cell-derived microparticles generated in patients during cardiopulmonary bypass are highly procoagulant. *Circulation* 1997; 96: 3534-41.
- [51] He Z, Si Y, Jiang T, Ma R, Zhang Y, Cao M, Li T, Yao Z, Zhao L, Fang S, Yu B, Dong Z, Thatte HS, Bi Y, Kou J, Yang S, Piao D, Hao L, Zhou J and Shi J. Phosphatidylserine exposure and neutrophil extracellular traps enhance procoagulant activity in patients with inflammatory bowel disease. *Thromb Haemost* 2016; 115: 738-51.
- [52] Liu Y, Li B, Hu TL, Li T, Zhang Y, Zhang C, Yu M, Wang C, Hou L, Dong Z, Hu TS, Novakovic VA and Shi J. Increased phosphatidylserine on blood cells in oral squamous cell carcinoma. *J Dent Res* 2019; 98: 763-771.
- [53] Meziani F, Delabranche X, Asfar P and Toti F. Bench-to bedside review: circulating microparticles—a new player in sepsis? *Crit Care* 2010; 14: 236.
- [54] Bester J and Pretorius E. Effects of IL-1 β , IL-6 and IL-8 on erythrocytes, platelets and clot viscoelasticity. *Sci Rep* 2016; 6: 32188.
- [55] Renné T, Pozgajová M, Grüner S, Schuh K, Pauer H, Burfeind P, Gailani D and Nieswandt B. Defective thrombus formation in mice lacking coagulation factor XII. *J Exp Med* 2005; 202: 271-81.
- [56] Tilley R and Mackman N. Tissue factor in hemostasis and thrombosis. *Semin Thromb Hemost* 2006; 32: 5-10.
- [57] Drake TA, Morrissey JH and Edgington TS. Selective cellular expression of tissue factor in human tissues. Implications for disorders of hemostasis and thrombosis. *Am J Pathol* 1989; 134: 1087-97.
- [58] Bogdanov VY, Cimmino G, Tardos JG, Tunstead JR and Badimon JJ. Assessment of plasma tissue factor activity in patients presenting with coronary artery disease: limitations of a commercial assay. *J Thromb Haemost* 2009; 7: 894-7.
- [59] Osterud B and Bjorklid E. Tissue factor in blood cells and endothelial cells. *Front Biosci (Elite Ed)* 2012; 4: 289-99.
- [60] Zhang Y, Meng H, Ma R, He Z, Wu X, Cao M, Yao Z, Zhao L, Li T, Deng R, Dong Z, Tian Y, Bi Y, Kou J, Thatte HS, Zhou J and Shi J. Circulating microparticles, blood cells, and endothelium, induce procoagulant activity in sepsis through phosphatidylserine exposure. *Shock* 2016; 45: 299-307.
- [61] Wang L, Bi Y, Yu M, Li T, Tong D, Yang X, Zhang C, Guo L, Wang C, Kou Y, Dong Z, Novakovic VA, Tian Y, Kou J, Shammam MA and Shi J. Phosphatidylserine-exposing blood cells and microparticles induce procoagulant activity in non-valvular atrial fibrillation. *Int J Cardiol* 2018; 258: 138-143.
- [62] Mantovani A, Cassatella MA, Costantini C and Jaillon S. Neutrophils in the activation and regulation of innate and adaptive immunity. *Nat Rev Immunol* 2011; 11: 519-31.
- [63] Brinkmann V, Reichard U, Goosmann C, Fauler B, Uhlemann Y, Weiss DS, Weinrauch Y and Zychlinsky A. Neutrophil extracellular traps kill bacteria. *Science* 2004; 303: 1532-5.
- [64] Cho JH, Fraser IP, Fukase K, Kusumoto S, Fujimoto Y, Stahl GL and Ezekowitz RA. Human peptidoglycan recognition protein S is an effector of neutrophil-mediated innate immunity. *Blood* 2005; 106: 2551-8.
- [65] von Brühl ML, Stark K, Steinhart A, Chandraratne S, Konrad I, Lorenz M, Khandoga A, Tircniceru A, Coletti R, Köllnberger M, Byrne RA, Laitinen I, Walch A, Brill A, Pfeiler S, Manukyan D, Braun S, Lange P, Riegger J, Ware J, Eckart A, Haidari S, Rudelius M, Schulz C, Echter K, Brinkmann V, Schwaiger M, Preissner KT, Wagner DD, Mackman N, Engelmann B and Massberg S. Monocytes, neutrophils, and platelets cooperate to initiate and propagate venous thrombosis in mice in vivo. *J Exp Med* 2012; 209: 819-35.
- [66] Ammollo CT, Semeraro F, Xu J, Esmon NL and Esmon CT. Extracellular histones increase plasma thrombin generation by impairing thrombomodulin-dependent protein C activation. *J Thromb Haemost* 2011; 9: 1795-803.
- [67] Silk E, Zhao H, Weng H and Ma D. The role of extracellular histone in organ injury. *Cell Death Dis* 2017; 8: e2812.
- [68] Keshari RS, Jyoti A, Dubey M, Kothari N, Kohli M, Bogra J, Barthwal MK and Dikshit M. Cytokines induced neutrophil extracellular traps formation: implication for the inflammatory disease condition. *PLoS One* 2012; 7: e48111.
- [69] Brandolini L, Bertini R, Bizzarri C, Sergi R, Caselli G, Zhou D, Locati M and Sozzani S. IL-1 beta primes IL-8-activated human neutrophils for elastase release, phospholipase D activity, and calcium flux. *J Leukoc Biol* 1996; 59: 427-34.
- [70] Dularay B, Elson CJ, Clements-Jewery S, Damais C and Lando D. Recombinant human interleukin-1 beta primes human polymorphonuclear leukocytes for stimulus-induced myeloperoxidase release. *J Leukoc Biol* 1990; 47: 158-63.

Thrombosis and anticoagulation of COVID-19

- [71] Ferrante A, Nandoskar M, Walz A, Goh DH and Kowanko IC. Effects of tumour necrosis factor alpha and interleukin-1 alpha and beta on human neutrophil migration, respiratory burst and degranulation. *Int Arch Allergy Appl Immunol* 1988; 86: 82-91.
- [72] Li B, Liu Y, Hu T, Zhang Y, Zhang C, Li T, Wang C, Dong Z, Novakovic VA, Hu T and Shi J. Neutrophil extracellular traps enhance procoagulant activity in patients with oral squamous cell carcinoma. *J Cancer Res Clin Oncol* 2019; 145: 1695-1707.
- [73] Zuo Y, Yalavarthi S, Shi H, Gockman K, Zuo M, Madison JA, Blair C, Weber A, Barnes BJ, Egeblad M, Woods RJ, Kanthi Y and Knight JS. Neutrophil extracellular traps in COVID-19. *JCI Insight* 2020; 5: e138999.
- [74] Middleton EA, He X, Denorme F, Campbell RA, Ng D, Salvatore SP, Mostyka M, Baxter-Stoltzfus A, Borczuk AC, Loda M, Cody MJ, Manne BK, Portier I, Harris ES, Petrey AC, Beswick EJ, Caulin AF, Iovino A, Abegglen LM, Weyrich AS, Rondina MT, Egeblad M, Schiffman JD and Yost CC. Neutrophil extracellular traps contribute to immunothrombosis in COVID-19 acute respiratory distress syndrome. *Blood* 2020; 136: 1169-1179.
- [75] Zhou P, Li T, Jin J, Liu Y, Li B, Sun Q, Tian J, Zhao H, Liu Z, Ma S, Zhang S, Novakovic VA, Shi J and Hu S. Interactions between neutrophil extracellular traps and activated platelets enhance procoagulant activity in acute stroke patients with ICA occlusion. *EBioMedicine* 2020; 53: 102671.
- [76] Tsourouktsoglou TD, Warnatsch A, Ioannou M, Hoving D, Wang Q and Papayannopoulos V. Histones, DNA, and citrullination promote neutrophil extracellular trap inflammation by regulating the localization and activation of TLR4. *Cell Rep* 2020; 31: 107602.
- [77] Bonetti PO, Lerman LO and Lerman A. Endothelial dysfunction: a marker of atherosclerotic risk. *Arterioscler Thromb Vasc Biol* 2003; 23: 168-75.
- [78] Pan F, Yang L, Li Y, Liang B, Li L, Ye T, Li L, Liu D, Gui S, Hu Y and Zheng C. Factors associated with death outcome in patients with severe coronavirus disease-19 (COVID-19): a case-control study. *Int J Med Sci* 2020; 17: 1281-1292.
- [79] Ackermann M, Verleden SE, Kuehnel M, Haverich A, Welte T, Laenger F, Vanstapel A, Werlein C, Stark H, Tzankov A, Li WW, Li VW, Mentzer SJ and Jonigk D. Pulmonary vascular endothelialitis, thrombosis, and angiogenesis in COVID-19. *N Engl J Med* 2020; 383: 120-128.
- [80] Iba T, Levy JH, Connors JM, Warkentin TE, Thachil J and Levi M. The unique characteristics of COVID-19 coagulopathy. *Crit Care* 2020; 24: 360.
- [81] Sztowski B, Antoniak S, Poller W, Schultheiss H and Rauch U. Procoagulant soluble tissue factor is released from endothelial cells in response to inflammatory cytokines. *Circ Res* 2005; 96: 1233-9.
- [82] Mehta P, McAuley DF, Brown M, Sanchez E, Tattersall RS and Manson JJ; HLH Across Speciality Collaboration, UK. COVID-19: consider cytokine storm syndromes and immunosuppression. *Lancet* 2020; 395: 1033-1034.
- [83] Sun X, Wang T, Cai D, Hu Z, Chen J, Liao H, Zhi L, Wei H, Zhang Z, Qiu Y, Wang J and Wang A. Cytokine storm intervention in the early stages of COVID-19 pneumonia. *Cytokine Growth Factor Rev* 2020; 53: 38-42.
- [84] Li T, Wang C, Liu Y, Li B, Zhang W, Wang L, Yu M, Zhao X, Du J, Zhang J, Dong Z, Jiang Tao, Xie R, Ma R, Fang S, Zhou J and Shi J. Neutrophil extracellular traps induce intestinal damage and thrombotic tendency in inflammatory bowel disease. *J Crohns Colitis* 2020; 14: 240-253.
- [85] Bartoszewski R, Moszyńska A, Serocki M, Cabaj A, Polten A, Ochocka R, Dell'Italia L, Bartoszewska S, Króliczewski J, Dąbrowski M and Collawn JF. Primary endothelial cell-specific regulation of hypoxia-inducible factor (HIF)-1 and HIF-2 and their target gene expression profiles during hypoxia. *FASEB J* 2019; 33: 7929-7941.
- [86] McElvaney OJ, McEvoy NL, McElvaney OF, Carroll TP, Murphy MP, Dunlea DM, Ní Choileáin O, Clarke J, O'Connor E, Hogan G, Ryan D, Sulaiman I, Gunaratnam C, Branagan P, O'Brien ME, Morgan RK, Costello RW, Hurley K, Walsh S, de Barra E, McNally C, McConkey S, Boland F, Galvin S, Kiernan F, O'Rourke J, Dwyer R, Power M, Geoghegan P, Larkin C, O'Leary RA, Freeman J, Gaffney A, Marsh B, Curley GF and McElvaney NG. Characterization of the inflammatory response to severe COVID-19 illness. *Am J Respir Crit Care Med* 2020; 202: 812-821.
- [87] Suresh MV, Balijepalli S, Zhang B, Singh VV, Swamy S, Panicker S, Dolgachev VA, Subramanian C, Ramakrishnan SK, Thomas B, Rao TC, Delano MJ, Machado AD, Shah YM and Raghavendran K. Hypoxia-inducible factor (HIF)-1 α promotes inflammation and injury following aspiration-induced lung injury in mice. *Shock* 2019; 52: 612-621.
- [88] Gong H, Rehman J, Tang H, Wary K, Mittal M, Chaturvedi P, Zhao YY, Komarova YA, Vogel SM and Malik AB. HIF2 α signaling inhibits adherens junctional disruption in acute lung injury. *J Clin Invest* 2015; 125: 652-64.

Thrombosis and anticoagulation of COVID-19

- [89] Thachil J. Hypoxia-an overlooked trigger for thrombosis in COVID-19 and other critically ill patients. *J Thromb Haemost* 2020; 18: 3109-3110.
- [90] Zhu J, Xie R, Piao X, Hou Y, Zhao C, Qiao G, Yang B, Shi J and Lu Y. Homocysteine enhances clot-promoting activity of endothelial cells via phosphatidylserine externalization and microparticles formation. *Amino Acids* 2012; 43: 1243-50.
- [91] Gupta N, Zhao YY and Evans CE. The stimulation of thrombosis by hypoxia. *Thromb Res* 2019; 181: 77-83.
- [92] Liu Y, Lv J, Liu J, Li M, Xie J, Lv Q, Deng W, Zhou N, Zhou Y, Song J, Wang P, Qin C, Tong WM and Huang B. Mucus production stimulated by IFN- α signaling triggers hypoxia of COVID-19. *Cell Res* 2020; 30: 1078-1087.
- [93] Wolberg AS, Aleman MM, Leiderman K and Machlus KR. Procoagulant activity in hemostasis and thrombosis: virchow's triad revisited. *Anesth Analg* 2012; 114: 275-85.
- [94] Varga Z, Flammer AJ, Steiger P, Haberecker M, Andermatt R, Zinkernagel AS, Mehra MR, Schuepbach RA, Ruschitzka F and Moch H. Endothelial cell infection and endotheliitis in COVID-19. *Lancet* 2020; 395: 1417-1418.
- [95] Zhou B, She J, Wang Y and Ma X. Venous thrombosis and arteriosclerosis obliterans of lower extremities in a very severe patient with 2019 novel coronavirus disease: a case report. *J Thromb Thrombolysis* 2020; 50: 229-232.
- [96] Potere N, Di NM, Rizzo G, La VM, Polilli E, Agostinone A, Spacone A, Di CS, Costantini AI, Abbate A, Porreca E and Parruti G. Low-dose subcutaneous tocilizumab to prevent disease progression in patients with moderate COVID-19 pneumonia and hyperinflammation. *Int J Infect Dis* 2020; 100: 421-424.
- [97] Qian S, An J, Qi F, Ye L, Chen Q, Liu X, Xie L and Li G. Tocilizumab exerts anti-inflammatory activity in six critically ill COVID-19 patients: a retrospective analysis. *Ann Transl Med* 2020; 8: 881.
- [98] Montesarchio V, Parrela R, Iommelli C, Bianco A, Manzillo E, Fraganza F, Palumbo C, Rea G, Murino P, De RR, Atripaldi L, D'Abbraccio M, Curvietto M, Mallardo D, Celentano E, Grimaldi AM, Palla M, Trojaniello C, Vitale MG, Million-Weaver SL and Ascierto PA. Outcomes and biomarker analyses among patients with COVID-19 treated with interleukin 6 (IL-6) receptor antagonist sarilumab at a single institution in Italy. *J Immunother Cancer* 2020; 8: e001089.
- [99] Choy EH, De BF, Takeuchi T, Hashizume M, John MR and Kishimoto T. Translating IL-6 biology into effective treatments. *Nat Rev Rheumatol* 2020; 16: 335-345.
- [100] Kimmig LM, Wu D, Gold M, Pettit NN, Pitrak D, Mueller J, Husain AN, Mutlu EA and Mutlu GM. IL-6 inhibition in critically ill COVID-19 patients is associated with increased secondary infections. *Front Med (Lausanne)* 2020; 7: 583897.
- [101] Garcia DA, Baglin TP, Weitz JI and Samama MM. Parenteral anticoagulants: antithrombotic therapy and prevention of thrombosis, 9th edition: American college of chest physicians evidence-based clinical practice guidelines. *Chest* 2012; 141: e24S-e43S.
- [102] Ludwig RJ. Therapeutic use of heparin beyond anticoagulation. *Curr Drug Discov Technol* 2009; 6: 281-9.
- [103] Shastri MD, Stewart N, Horne J, Peterson GM, Gueven N, Sohal SS and Patel RP. In-vitro suppression of IL-6 and IL-8 release from human pulmonary epithelial cells by non-anticoagulant fraction of enoxaparin. *PLoS One* 2015; 10: e0126763.
- [104] Tian R, Wu W, Wang C, Pang H, Zhang Z, Xu H, Luo Q, Gao P, Shi J, Li W, Qian H, Guo F, Li T, Liu Z, Wang J, Zhou X, Qin Y, Yan X and Zhang S. Clinical characteristics and survival analysis in critical and non-critical patients with COVID-19 in Wuhan, China: a single-center retrospective case control study. *Sci Rep* 2020; 10: 17524.
- [105] Amgalan A and Othman M. Exploring possible mechanisms for COVID-19 induced thrombocytopenia: unanswered questions. *J Thromb Haemost* 2020; 18: 1514-1516.
- [106] Hohl EL, Derhaschnig U, Firbas C, Schoergenhofer C, Schwameis M and Jilma B. Reversal strategy in antagonizing the P2Y₁₂-inhibitor ticagrelor. *Eur J Clin Invest* 2013; 43: 1258-61.
- [107] Campbell AM, Kashgarian M and Shlomchik MJ. NADPH oxidase inhibits the pathogenesis of systemic lupus erythematosus. *Sci Transl Med* 2012; 4: 157ra141.
- [108] Knight JS, Luo W, O'Dell AA, Yalavarthi S, Zhao W, Subramanian V, Guo C, Grenn RC, Thompson PR, Eitzman DT and Kaplan MJ. Peptidylarginine deiminase inhibition reduces vascular damage and modulates innate immune responses in murine models of atherosclerosis. *Circ Res* 2014; 114: 947-56.
- [109] Kannemeier C, Shibamiya A, Nakazawa F, Trusheim H, Ruppert C, Markart P, Song Y, Tzima E, Kennerknecht E, Niepmann M, von Bruehl ML, Sedding D, Massberg S, Günther A, Engelmann B and Preissner KT. Extracellular RNA constitutes a natural procoagulant cofactor in blood coagulation. *Proc Natl Acad Sci U S A* 2007; 104: 6388-93.
- [110] Varjú I, Longstaff C, Szabó L, Farkas ÁZ, Varga-Szabó VJ, Tanka-Salamon A, Machovich R and Kolev K. DNA, histones and neutrophil extracellular traps exert anti-fibrinolytic effects in a plasma environment. *Thromb Haemost* 2015; 113: 1289-98.

Thrombosis and anticoagulation of COVID-19

- [111] Whyte CS, Morrow GB, Mitchell JL, Chowdary P and Mutch NJ. Fibrinolytic abnormalities in acute respiratory distress syndrome (ARDS) and versatility of thrombolytic drugs to treat COVID-19. *J Thromb Haemost* 2020; **18**: 1548-1555.
- [112] Qin C, Zhou L, Hu Z, Zhang S, Yang S, Tao Y, Xie C, Ma K, Shang K, Wang W and Tian DS. Dysregulation of immune response in patients with coronavirus 2019 (COVID-19) in Wuhan, China. *Clin Infect Dis* 2020; **71**: 762-768.
- [113] Couzin-Frankel J. The mystery of the pandemic's 'happy hypoxia'. *Science* 2020; **368**: 455-456.
- [114] Tobin MJ, Laghi F and Jubran A. Why COVID-19 silent hypoxemia is baffling to physicians. *Am J Respir Crit Care Med* 2020; **202**: 356-360.