

## Original Article

# A crucial role for the long non-coding RNA *CASC11* in the pathogenesis of human cancers

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**Abstract:** Long non-coding RNAs (lncRNAs) are non-coding RNAs more than 200 nucleotides in length. Although they do not encode proteins, lncRNAs can regulate gene expression at the transcriptional, post-transcriptional, and epigenetic levels. Emerging data show that lncRNAs are important for tumorigenesis and cancer progression. Cancer susceptibility candidate 11 (*CASC11*) is a prominent lncRNA that is upregulated in various types of cancers. Moreover, its overexpression correlates with larger tumor size, more advanced cancer stages, cancer metastasis, and poor overall survival for most types of cancer. Functionally, the knockdown of *CASC11* can inhibit cell proliferation, invasion, and migration, while enhancing apoptosis through its regulation of gene expression and signaling pathways and its interactions with functional proteins. Here, we discuss the identification, expression, and function of *CASC11*. Additionally, we discuss the potential roles of *CASC11* as a diagnostic biomarker, prognostic biomarker, and therapeutic target in various cancers.

**Keywords:** Cancer, lncRNA, *CASC11*, prognosis, treatment

## Introduction

According to 2015 estimates from the World Health Organization, cancer remains a leading cause of death worldwide [1, 2], and cancer incidence and mortality rates remain high [3]. In recent years, although the understanding of cancer-related molecular mechanisms has increased substantially, and the therapeutic methods for treating cancer have greatly improved [4-6], treatments for advanced-stage cancers still remain unsatisfactory, and early-stage cancers tend to go undiagnosed, in part, due to the lack of useful biomarkers for their detection. Therefore, it is urgent to identify biomarkers of early disease while continuing to investigate the mechanisms that contribute to cancer growth, invasion, and metastasis, to develop new cancer therapies.

In humans, although a large portion of the genome is transcribed, less than 2% of the genome encodes proteins [7]. The remaining transcripts represent various classes of non-coding RNAs (ncRNAs) [8, 9]. One class of ncRNAs is long non-coding RNAs (lncRNAs),

which are defined as RNA transcripts exceeding 200 nucleotides in length [10, 11]. In recent years, lncRNAs have received much attention because they can regulate gene expression at the transcriptional, post-transcriptional, and epigenetic levels [12-14]. At the transcriptional level lncRNAs can affect the cofactors, transcription factors, and RNA polymerase II, at the post-transcriptional level they can influence protein interactions, alternative splicing, miRNA sponging, and mRNA stability and translation, and at the epigenetic level they can impact DNA methylation, histone modifications, and chromatin remodeling [10, 15-18].

The development of new genome sequencing technologies and bioinformatics tools has led to more and more lncRNAs being implicated as vital regulators of many physiological and pathological processes, including cancer occurrence and progression [19-21]. lncRNAs have been reported to affect multiple cellular behaviors, such as growth, differentiation, and apoptosis, which are generally dysregulated in cancer [22, 23]. Further, many lncRNAs are aberrantly expressed in cancer tissues and are

present at higher levels in cancer patient plasma compared with controls, suggesting lncRNAs might be promising diagnostic biomarkers for cancer [24-27].

A growing body of evidence shows that the lncRNA cancer susceptibility candidate 11 (CASC11) promotes oncogenesis and is upregulated in many cancers. CASC11 has attracted much attention, not only for its value in diagnosing cancer and predicting prognosis, but also for its potential as a therapeutic target in various cancers. Here, we summarize recent progress toward defining the molecular function of CASC11 and how it relates to cancer. We also evaluate the potential prognostic, diagnostic, and therapeutic value of this lncRNA in various cancer types.

### Approaches to detecting lncRNA CASC11

Unlike protein-coding genes, lncRNAs generally lack conserved sequence motifs that aid in their detection. At present, quantitative reverse-transcription polymerase chain reaction (qRT-PCR) is considered the gold standard for quantifying lncRNAs, and it is the most commonly used method for measuring the expressions of CASC11 in tumor tissues, plasma, and cells. Liu et al. used qRT-PCR to show that CASC11 is overexpressed in hepatocellular carcinoma (HCC) by comparing its expression in 69 HCC tumor samples to adjacent healthy tissue controls [28]. Likewise, Luo et al. used qRT-PCR to detect the presence of CASC11 in the plasma of bladder cancer patients [29].

In addition to qRT-PCR, new methods are being developed for the detection of lncRNAs, such as, chemiluminescent, electrochemical techniques, next-generation sequencing, and third generation sequencing. Some studies have used microarray-based analyses to compare CASC11 expression in tumors vs. controls. For example, microarrays were used in a study showing that CASC11 is upregulated in gastric cancer tissues compared with normal adjacent tissues [30]. Others have used RNA-sequencing data downloaded from The Cancer Genome Atlas (TCGA) to examine CASC11 expressions [31].

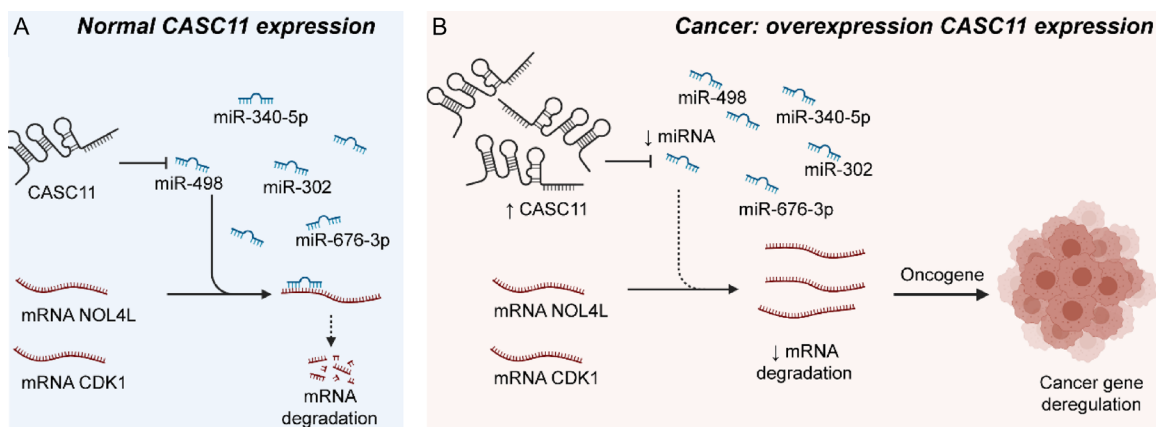
### The expression of CASC11 in cancer

Transcriptional profiling has revealed increased CASC11 expressions in tumor tissues com-

pared to adjacent non-tumor tissues in patients with various cancers, including colorectal cancer, hepatocellular carcinoma, bladder cancer, lung cancer, glioma, ovarian carcinomas including ovarian squamous cell carcinoma, cervical cancer, gastric cancer, neuroblastoma, and esophageal carcinoma. Cell lines representing these malignancies also show the trend of CASC11 upregulation. Importantly, elevated CASC11 expression is associated with increased tumor size, invasion, lymph metastasis, more advanced TNM stage, and decreased differentiation [30, 32-34]. Importantly, Kaplan-Meier survival analyses indicate that cancer patients with higher CASC11 expression had poorer prognoses than those with lower CASC11 expression.

It remains unclear how CASC11 is upregulated in tumors; however, transcriptional activation is a common mechanism used to upregulate lncRNA expression. Zhang et al. explored the promoter region of CASC11 for potential transcription factor binding sites using the University of California Santa Cruz (UCSC) Genome Browser [33]. They identified an E-box region in the CASC11 promoter that might be a potential c-Myc binding site. They performed in vitro experiments to demonstrate that the transfection of c-Myc into SW480 and HEK293T cells promotes luciferase expression from reporter constructs bearing the E-box region of the CASC11 promoter. Additional chromatin immunoprecipitation (ChIP) assays and c-Myc gain- and c-Myc chromatin immunoprecipitation can directly bind to the E-box element and regulate the expression of CASC11 [33]. The effects of c-Myc on transcription are likely due to the role of c-Myc in recruiting proteins important for histone H3 lysine acetylation. The UCSC Genome Browser was used to identify a region in the promoter of CASC11 which was likely to bear the histone H3 lysine 27 acetylation (H3K27Ac) mark. The knockdown of c-Myc in SW620 cells combined with treatment with the histone deacetylase (HDAC) inhibitor trichostatin A demonstrated that preventing histone deacetylation facilitates the expression of CASC11 in SW620 cells knocked-down for c-Myc. These results indicate that c-Myc enhances promoter histone acetylation to increase CASC11 expression [33] (**Figure 1**). Another study showed that STAT3 can also upregulate CASC11 expression in the context of HCC [35]. In this study, online bioinformatics tools from

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**Figure 1.** Transcriptional activation mechanisms leading to the elevated expression of *CASC11*. A. The E-box region of the *CASC11* promoter might serve as a c-Myc binding site. Once bound, c-Myc can recruit other proteins to enhance promoter histone acetylation and increase *CASC11* expression. B. *CASC11* expression can be increased in the presence of the transcriptional activator STAT3, and the transcription factors FOXO3 and SP1 have both been shown to bind to the *CASC11* promoter region to promote *CASC11* expression.

**Table 1.** The expression patterns of *CASC11* in cancers

Cancer types	Expression	Detecting method	Sample types	Prognostic relevance	Refs
colorectal cancer	upregulated	RT-qPCR	tissues and cells	/	27012187
hepatocellular carcinoma	upregulated	RT-qPCR and IHC	Serum and tissues	poor survival	33252856
	upregulated	RT-qPCR	tissues	/	32194694
	upregulated	RT-qPCR	tissues and cell lines	poor survival	30503497
	upregulated	RT-qPCR	tissues	poor survival	30910841
bladder cancer	upregulated	RT-qPCR	tissues	/	30916832
lung cancer	upregulated	RT-qPCR	tissues and cell lines	/	31378894
	upregulated	RT-qPCR	tissues	poor survival	31537383
	upregulated/plasma	RT-qPCR	plasma	poor survival	30965130
glioma	upregulated	RT-qPCR	tissues and cell lines	poor survival	31121483
ovarian carcinoma	upregulated	RT-qPCR	plasma	poor survival	32158258
	upregulated	RT-qPCR	plasma		31181314
cervical cancer	upregulated	RT-qPCR	tissues and cell lines	poor survival	31255182
gastric cancer	upregulated	RT-qPCR	tissues	/	30200804
	upregulated	RT-qPCR	tissues	/	31632064
neuroblastoma	upregulated	RT-qPCR	tissues and cell lines	poor survival	31645055
esophageal carcinoma	upregulated	RT-qPCR	tissues	poor survival	31696474

JASPAR (<http://jaspar.genereg.net>) and UCSC (<http://genome.ucsc.edu/>) were used to predict that STAT3 is a transcriptional activator of *CASC11*. Luciferase reporter assays and ChIP assays confirmed that STAT3 occupies the *CASC11* promoter [35]. Other transcription factors also reported to promote *CASC11* expression include FOXO3 and SP1 [32, 36] (Figure 1). Taken together, it is clear that transcription factors regulate the expression of *CASC11*, but the contributions of each of these factors in different contexts is not yet clear.

### Functional activities of *CASC11* in cancer

Emerging data show that many lncRNAs have important roles in tumorigenesis and cancer progression [37-40]. lncRNAs can promote malignant features, such as sustaining growth, inducing invasion and metastasis, inhibiting death, and enhancing chemotherapeutic resistance [19, 41, 42]. Strikingly, *CASC11* expression is consistently elevated in most cancer types and generally enhances malignant cell characteristics (Table 1).

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### *CASC11 in cell proliferation*

Increasing evidence indicates that *CASC11* overexpression promotes cancer cell growth in vitro and vivo. Colony formation and cell proliferation assays show that the knockdown of *CASC11* in HCC cells significantly inhibits cell proliferative potential compared with control cells [31, 43]. *CASC11* also promotes tumor cell growth in other tumor cell lines, including colorectal cancer [33], bladder cancer [29], lung cancer [32, 44], glioma [36], ovarian carcinoma [45], cervical cancer [46], gastric cancer [30, 47], neuroblastoma [48] and esophageal carcinoma [49], suggesting it may act as an oncogene. Consistently, the effects of silencing and overexpressing *CASC11* in xenograft animal models also indicate that *CASC11* plays a role in oncogenesis [33, 46].

### *CASC11 in cell apoptosis*

Apoptosis is a finely-balanced, ordered, orchestrated mechanism of cellular death, and tumor formation and cancer progression can result from too little apoptosis [50, 51]. Several studies have revealed that *CASC11* suppresses apoptosis in multiple types of cancer tumor cells. For example, the overexpression of *CASC11* inhibits apoptosis in ovarian carcinoma cells [45], but the silencing of *CASC11* enhances apoptosis in HCC cells [31]. Additionally, in cervical cancer [46], gastric cancer [30], and esophageal carcinoma [49], the knockdown of *CASC11* was reported to remarkably increase tumor cell apoptosis and inhibit cancer progression.

### *CASC11 in cell invasion and migration*

The invasion and migration of cancer cells into surrounding tissue and vasculature is an essential step in cancer metastasis [52, 53], which greatly contributes to cancer-related death. Transwell experiments using HeLa cervical cancer cells showed that silencing *CASC11* decreases cell migration and invasion compared to controls, but the overexpression of *CASC11* increases cell migration and invasion [46]. Similar results were demonstrated in other cancer cell lines [29-31, 33, 36]. Consistent with these in vitro findings, *CASC11* also promotes tumor metastasis in vivo [33]. Together, these findings indicate *CASC11* may have an important role in cancer cell invasion and migration.

### *CASC11 in chemotherapeutic resistance*

Chemotherapeutic resistance is one of the major hindrances in the fight against cancer, and *CASC11* expression may contribute to this resistance [54]. The *CASC11* plasma levels were significantly increased at both 3 and 6 months following the initiation of chemotherapy in ovarian squamous cell carcinoma patients [55]. Similarly, ovarian carcinoma cells treated with either cisplatin, oxaliplatin, tetraplatin or carboplatin displayed higher *CASC11* expressions than the untreated controls, and when *CASC11* was overexpressed in treated cells, it led to increased cell proliferation, but when *CASC11* was silenced, it inhibited proliferation [55]. Consistent with these observations, the plasma levels of *CASC11* were also increased in HCC patients at 3 and 6 months after the initiation of chemotherapy, and the knockdown of *CASC11* decreased the viability of carboplatin-treated HCC cells [28]. Together, these findings are consistent with a model in which elevated levels of *CASC11* contribute to chemotherapy resistance.

### **Mechanisms of *CASC11* action in cancer**

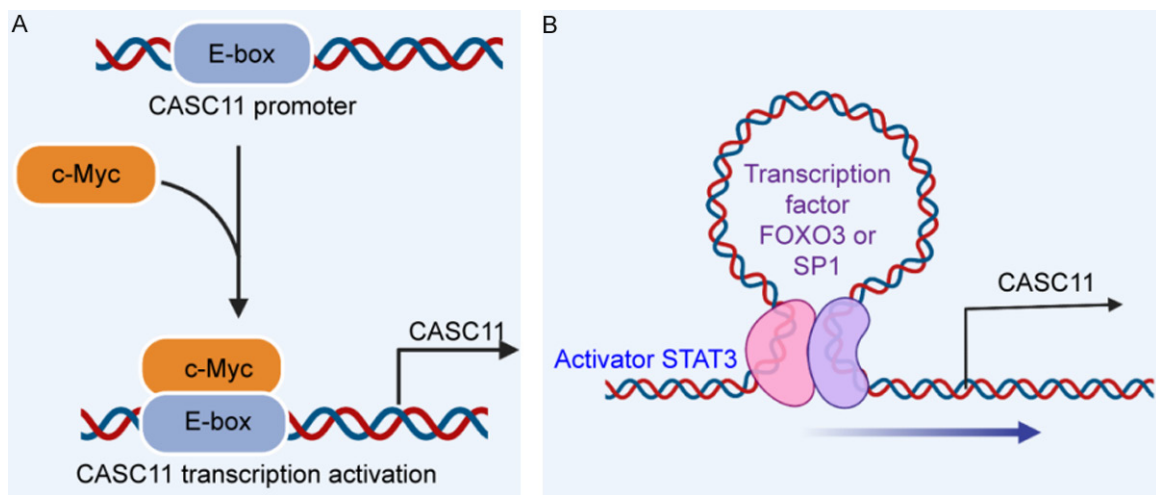
Although most lncRNAs have not been functionally annotated, and lncRNAs remain challenging to characterize [56], accumulating evidence indicates that lncRNAs interact with other RNAs, DNA, and proteins to participate in processes ranging from transcription and intracellular trafficking to chromosome remodeling [57]. Although it is clear that *CASC11* has important functions related to cancer, the specific mechanisms and pathways involved are extremely complex, multifactorial, and poorly understood (**Table 2**).

One mechanism, the regulation of target gene expression via competing endogenous RNA (ceRNA) interplay, might be the major mechanism underlying the oncogenic functions of *CASC11*. CeRNAs are non-coding RNAs that compete with messenger RNAs (mRNAs) for their microRNA (miRNAs) regulators, to modulate gene expression post-transcriptionally [58]. Many RNA species can act as ceRNAs, including other miRNAs, lncRNAs, circular RNAs, and pseudogenes [59, 60]. As a ceRNA, *CASC11* might inhibit related miRNAs to promote the translation of their miRNA-targeted mRNAs by

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**Table 2.** The regulatory mechanisms of lncRNA CASC11 in various cancers

Cancer types	Interacting proteins or genes	Related signaling pathways	Refs
colorectal cancer	hnRNP-K	WNT/P-Ktal ca	27012187
hepatocellular carcinoma	EIF4A3	NF-κF signaling and PI3K/AKT/mTOR pathway	33252856
hepatocellular carcinoma	EZH2	PI3K/AKT signaling pathway	30503497
hepatocellular carcinoma	miRNA-188-5p	/	30910841
bladder cancer	miRNAer c	/	30916832
lung cancer	microRNA-302/CDK1	/	31378894
non-small cell lung cancer	miR-498	/	31537383
glioma	miR-498/FOXK1	/	31121483
cervical cancer	/	Wnt/β-catenin	31255182
gastric cancer	miR-340-5p/CDK1	cell cycle pathway	30200804
neuroblastoma	microRNA-676-3p/nucleolar protein 4like (NOL4L)	/	31645055
esophageal carcinoma	KLF6	/	31696474

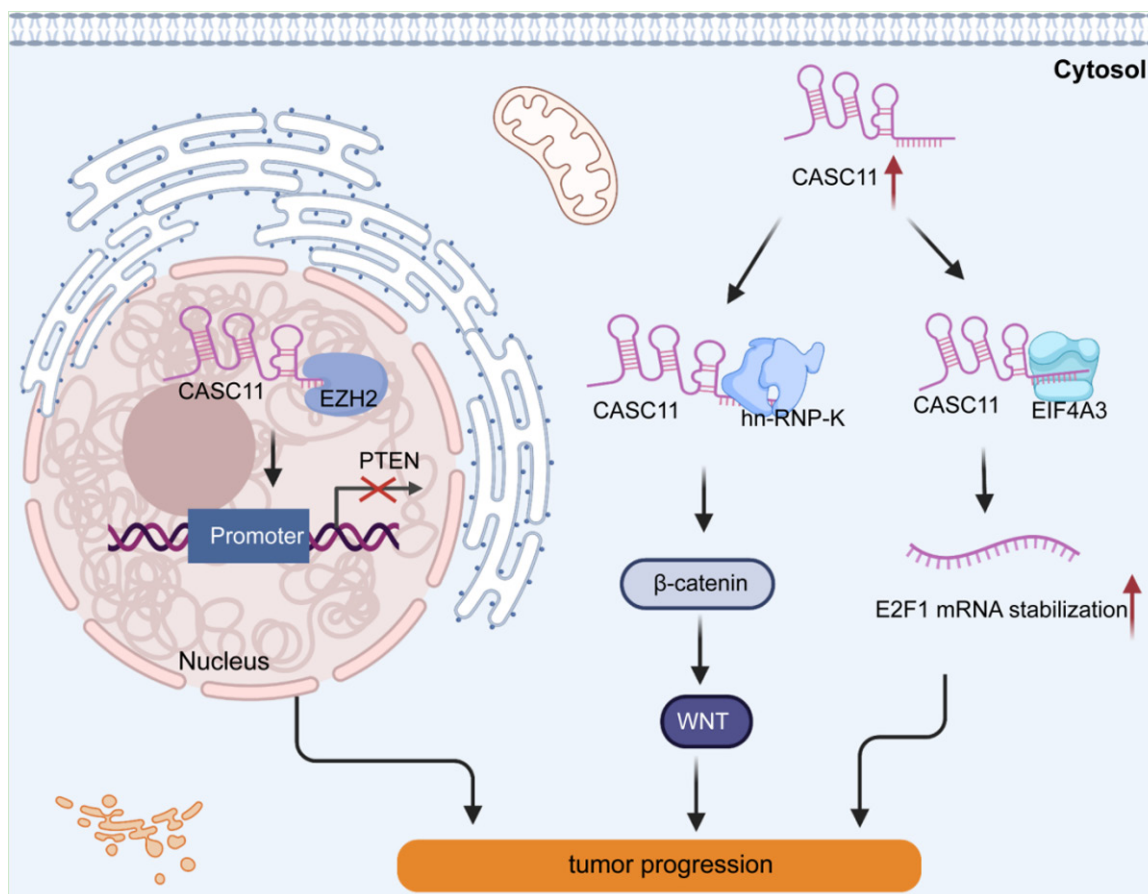


**Figure 2.** CASC11 may act as a ceRNA in cancer. CASC11 can inhibit specific miRNAs to promote protein expression by regulating the level of the miRNA's target gene's mRNA degradation. A. When CASC11 is normally expressed, the CASC11 levels are insufficient to compete with mRNA for binding to specific miRNAs, permitting the miRNA to promote degradation of its mRNA target. B. When CASC11 is overexpressed, it can compete with mRNA for binding to specific miRNAs, thus inhibiting the ability of the miRNA to promote the degradation of its mRNA target as has been shown for MIR302, MIR498, MIR340-5p, and MIR676-3p.

modulating the miRNA-mediated mRNA degradation level. Indeed, CASC11 has been reported to act as a ceRNA for several miRNA/mRNA combinations: MIR302/CDK1 [44], MIR498/FOXO3 [32], MIR340-5p/CDK1 [30], and MIR-676-3p/NOL4L [48]. Dual-luciferase reporter gene data combined with other experimental data show that CASC11 binds MIR302, which targets cyclin dependent kinase 1 (CDK1) mRNA in lung cancer cells [44]. Additional functional assays indicate that high expressions of CASC11 may accelerate lung cancer progression through a MIR302/CDK1 axis. Similarly, CASC11, promotes tumor cell proliferation in non-small cell lung cancer through targeting MIR498/forkhead box O3 (FOXO3), and Zhang

et al. linked the CASC11/MIR340-5p/CDK1 network to gastric cancer [30, 32]. In addition, CASC11 was found to serve as a molecular sponge for MIR676-3p, relieving its inhibition of nucleolar protein 4-like mRNA (NOL4L) in neuroblastoma cells [48] (Figure 2).

Several studies suggest that in addition to acting as a ceRNA that influences protein levels, CASC11 can exert its function by interacting with proteins directly. For example, the histone methyltransferase enhancer of zeste homolog 2 (EZH2) epigenetically represses the expression of the tumor suppressor phosphatase and tensin homolog deleted on chromosome ten (PTEN) [35]. CASC11 binds to EZH2 to enhance



**Figure 3.** CASC11 exerts some of its functions through protein interactions. CASC11 directly binds to EZH2 to enhance the epigenetic silencing of the PTEN promoter region and suppress PTEN expression. The overexpression of CASC11 activates the WNT signaling pathway in colorectal cancer cells by increasing  $\beta$ -catenin nuclear accumulation through its interaction with hnRNP-K. Additionally, the upregulation of CASC11 promotes tumor progression by enhancing the stability of *E2F1* mRNA by recruiting eukaryotic translation initiation factor 4A3 (EIF4A3).

ce epigenetic silencing in the PTEN promoter region. CASC11 also interacts with heterogeneous nuclear riboprotein K (hnRNP-K), a protein that shuttles between the nucleus and cytoplasm and interacts with RNA and DNA [61, 62]. The overexpression of CASC11 activates WNT signaling in colorectal cancer cells by increasing the nuclear accumulation of  $\beta$ -catenin by interacting with hnRNP-K [33]. Additionally, another recent study demonstrated that CASC11 promotes HCC progression by enhancing the stability of E2F transcription factor 1 (*E2F1*) mRNA by binding to and recruiting eukaryotic translation initiation factor 4A3 (EIF4A3) [31] (**Figure 3**). Given these many examples of functional CASC11-protein interactions, it will be important to determine, whether CASC11 has other regulatory functions mediated by direct interactions with other proteins.

### CASC11 as a diagnostic and prognostic biomarker in cancer

Molecular biomarkers for cancer have improved dramatically in recent years [63-65]. The primacy of specificity as a requirement for diagnostic and prognostic biomarkers points to lncRNAs, because the expression of most lncRNAs is strikingly tissue-specific compared with protein-coding genes [66]. CASC11 is upregulated in the sera and tissues of HCC patients, and HCC patients with higher CASC11 expressions exhibit larger tumors and have poorer prognoses than those with lower CASC11 expressions. Receiver operating characteristic (ROC) curve analyses show that CASC11 may be a good candidate as a prognostic biomarker for HCC (AUC=0.8) [31]. The diagnostic potential of CASC11 has also been evaluated

using ROC curve analyses in patients with bladder cancer vs. healthy controls (AUC=0.899) indicating *CASC11* expression might be a predictor for this cancer [29]. *CASC11* plasma levels are also upregulated in ovarian carcinoma patients compared with healthy controls and ROC curve analyses show that *CASC11* levels can predict early-stage ovarian carcinoma [45]. The results of a survival curve analysis demonstrated that *CASC11* overexpression predicts poor prognosis in ovarian carcinoma and ovarian squamous cell carcinoma [45, 55]. Taken together, these studies strongly indicate the potential of *CASC11* expression as a biomarker that can be detected in less-invasive blood tests.

### **CASC11 as a therapeutic target in cancer**

Accumulating evidence indicates that lncRNAs might be promising therapeutic targets [67-70]. *CASC11* has attracted much attention, not only for its value in diagnosing cancer and predicting prognosis, but also for its potential as a therapeutic target. Several studies show that the expression of *CASC11* is altered in tumor tissues, plasma from cancer patients, and in cancer cell lines, indicating it has a regulatory role in cancer. The overexpression of *CASC11* is closely associated with tumor size, stage, and prognosis [31, 32, 35]. Gain- and loss-of-function experiments with *CASC11* indicate that *CASC11* may function as an oncogene to promote tumor cell proliferation, invasion, migration, and chemotherapeutic resistance, while suppressing tumor cell apoptosis through the modulation of the WNT/ $\beta$ -catenin, NF- $\kappa$ B signaling, and PI3K/AKT/mTOR pathways [31, 33, 35]. Thus, these findings suggest that *CASC11* plays an oncogenic role in a variety of cancer types and that targeting this lncRNA may provide a novel approach toward improved cancer therapy.

### **Conclusions and perspectives**

A growing amount of evidence supports lncRNAs as biomarkers for the diagnosis and prognosis of patients with cancer [71-74]. *CASC11* is a novel lncRNA that acts as an oncogene in several different types of cancer. Most studies indicate that *CASC11* is overexpressed in multiple cancers, and its overexpression correlates with larger tumor size, more advanced cancer

stages, cancer metastasis, and poor overall survival for most types of cancer. Functionally, the knockdown of *CASC11* can inhibit cell proliferation, invasion and migration, while promoting apoptosis through its regulation of gene expression and signaling pathways, and its interactions with functional proteins. Overall, there is substantial evidence that *CASC11* may have immediate clinical potential for diagnosing cancer and predicting patient prognosis, as well as future potential as a therapeutic target. Although the wide diversity of molecular interactions that contribute to cancer tumorigenesis and progression complicate the development of optimal cancer treatments, *CASC11* may provide a new target for cancer therapy. Currently, the development of methods to inhibit *CASC11* to target cancer is in the early stages [30, 32, 35], and further in-depth studies of *CASC11* and the molecular mechanisms it affects are both needed and worthy of attention.

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### **Disclosure of conflict of interest**

None.

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### **References**

- [1] Siegel RL, Miller KD and Jemal A. Cancer statistics, 2019. *CA Cancer J Clin* 2019; 69: 7-34.
- [2] Miller KD, Nogueira L, Mariotto AB, Rowland JH, Yabroff KR, Alfano CM, Jemal A, Kramer JL and Siegel RL. Cancer treatment and survivorship statistics, 2019. *CA Cancer J Clin* 2019; 69: 363-385.
- [3] Granátová M and Svoboda I. Detection of bovine leukemia virus antibodies using the cytotoxicity test in comparison with other serologic methods. *Vet Med (Praha)* 1986; 31: 451-458.

## CASC11 role in human cancers

- [4] Pan Y, Kadash-Edmondson KE, Wang R, Phillips J, Liu S, Ribas A, Aplenc R, Witte ON and Xing Y. RNA dysregulation: an expanding source of cancer immunotherapy targets. *Trends Pharmacol Sci* 2021; 42: 268-282.
- [5] Wang C, Zhang J, Yin J, Gan Y, Xu S, Gu Y and Huang W. Alternative approaches to target Myc for cancer treatment. *Signal Transduct Target Ther* 2021; 6: 117.
- [6] Wang Y, Li S, Wang X, Chen Q, He Z, Luo C and Sun J. Smart transformable nanomedicines for cancer therapy. *Biomaterials* 2021; 271: 120737.
- [7] ENCODE Project Consortium, Birney E, Stamatoyannopoulos JA, Dutta A, Guigó R, Gingeras TR, Margulies EH, Weng Z, Snyder M, Dermitzakis ET, Thurman RE, Kuehn MS, Taylor CM, Neph S, Koch CM, Asthana S, Malhotra A, Adzhubei I, Greenbaum JA, Andrews RM, Flicek P, Boyle PJ, Cao H, Carter NP, Clelland GK, Davis S, Day N, Dhami P, Dillon SC, Dorschner MO, Fiegler H, Giresi PG, Goldy J, Hawrylycz M, Haydock A, Humbert R, James KD, Johnson BE, Johnson EM, Frum TT, Rosenzweig ER, Karnani N, Lee K, Lefebvre GC, Navas PA, Neri F, Parker SC, Sabo PJ, Sandstrom R, Shafer A, Vetrie D, Weaver M, Wilcox S, Yu M, Collins FS, Dekker J, Lieb JD, Tullius TD, Crawford GE, Sunyaev S, Noble WS, Dunham I, Denoeud F, Reymond A, Kapranov P, Rozowsky J, Zheng D, Castelo R, Frankish A, Harrow J, Ghosh S, Sandelin A, Hofacker IL, Baertsch R, Keefe D, Dike S, Cheng J, Hirsch HA, Sekinger EA, Lagarde J, Abril JF, Shahab A, Flamm C, Fried C, Hackermüller J, Hertel J, Lindemeyer M, Missal K, Tanzer A, Washietl S, Korb J, Emanuelsson O, Pedersen JS, Holroyd N, Taylor R, Swarbreck D, Matthews N, Dickson MC, Thomas DJ, Weirauch MT, Gilbert J, Drenkow J, Bell I, Zhao X, Srinivasan KG, Sung WK, Ooi HS, Chiu KP, Foissac S, Alioto T, Brent M, Pachter L, Tress ML, Valencia A, Choo SW, Choo CY, Ucla C, Manzano C, Wyss C, Cheung E, Clark TG, Brown JB, Ganesh M, Patel S, Tammanna H, Chrast J, Henrichsen CN, Kai C, Kawai J, Nagalakshmi U, Wu J, Lian Z, Lian J, Newburger P, Zhang X, Bickel P, Mattick JS, Carninci P, Hayashizaki Y, Weissman S, Hubbard T, Myers RM, Rogers J, Stadler PF, Lowe TM, Wei CL, Ruan Y, Struhl K, Gerstein M, Antonarakis SE, Fu Y, Green ED, Karaöz U, Siepel A, Taylor J, Liefer LA, Wetterstrand KA, Good PJ, Feingold EA, Guyer MS, Cooper GM, Asimenos G, Dewey CN, Hou M, Nikolaev S, Montoya-Burgos JI, Löytynoja A, Whelan S, Pardi F, Massingham T, Huang H, Zhang NR, Holmes I, Mullikin JC, Ureta-Vidal A, Paten B, Seringhaus M, Church D, Rosenbloom K, Kent WJ and Stone EA; NISC Comparative Sequencing Program, Baylor College of Medicine Human Genome Sequencing Center, Washington University Genome Sequencing Center, Broad Institute, Children's Hospital Oakland Research Institute, Batzoglou S, Goldman N, Hardison RC, Haussler D, Miller W, Sidow A, Trinklein ND, Zhang ZD, Barrera L, Stuart R, King DC, Ameer A, Enroth S, Bieda MC, Kim J, Bhinge AA, Jiang N, Liu J, Yao F, Vega VB, Lee CW, Ng P, Shahab A, Yang A, Moqtaderi Z, Zhu Z, Xu X, Squazzo S, Oberley MJ, Inman D, Singer MA, Richmond TA, Munn KJ, Rada-Iglesias A, Wallerman O, Komorowski J, Fowler JC, Couttet P, Bruce AW, Dovey OM, Ellis PD, Langford CF, Nix DA, Euskirchen G, Hartman S, Urban AE, Kraus P, Van Calcar S, Heintzman N, Kim TH, Wang K, Qu C, Hon G, Luna R, Glass CK, Rosenfeld MG, Aldred SF, Cooper SJ, Halees A, Lin JM, Shulha HP, Zhang X, Xu M, Haidar JN, Yu Y, Ruan Y, Iyer VR, Green RD, Wadelius C, Farnham PJ, Ren B, Harte RA, Hinrichs AS, Trumbower H, Clawson H, Hillman-Jackson J, Zweig AS, Smith K, Thakkapallayil A, Barber G, Kuhn RM, Karolchik D, Armengol L, Bird CP, de Bakker PI, Kern AD, Lopez-Bigas N, Martin JD, Stranger BE, Woodroffe A, Davydov E, Dimas A, Eyras E, Hallgrímsson IB, Huppert J, Zody MC, Abecasis GR, Estivill X, Bouffard GG, Guan X, Hansen NF, Idol JR, Maduro VV, Maskeri B, McDowell JC, Park M, Thomas PJ, Young AC, Blakesley RW, Muzny DM, Sodergren E, Wheeler DA, Worley KC, Jiang H, Weinstock GM, Gibbs RA, Graves T, Fulton R, Mardis ER, Wilson RK, Clamp M, Cuff J, Gnerre S, Jaffe DB, Chang JL, Lindblad-Toh K, Lander ES, Koriabine M, Nefedov M, Osoegawa K, Yoshinaga Y, Zhu B and de Jong PJ. Identification and analysis of functional elements in 1% of the human genome by the ENCODE pilot project. *Nature* 2007; 447: 799-816.
- [8] ENCODE Project Consortium. An integrated encyclopedia of DNA elements in the human genome. *Nature* 2012; 489: 57-74.
- [9] Hangauer MJ, Vaughn IW and McManus MT. Pervasive transcription of the human genome produces thousands of previously unidentified long intergenic noncoding RNAs. *PLoS Genet* 2013; 9: e1003569.
- [10] Rinn JL and Chang HY. Genome regulation by long noncoding RNAs. *Annu Rev Biochem* 2012; 81: 145-166.
- [11] Chi Y, Wang D, Wang J, Yu W and Yang J. Long non-coding RNA in the pathogenesis of cancers. *Cells* 2019; 8: 1015.
- [12] Dykes IM and Emanueli C. Transcriptional and post-transcriptional gene regulation by long non-coding RNA. *Genomics Proteomics Bioinformatics* 2017; 15: 177-186.
- [13] Panni S, Lovering RC, Porras P and Orchard S. Non-coding RNA regulatory networks. *Biochim*



## CASC11 role in human cancers

- Biophys Acta Gene Regul Mech 2020; 1863: 194417.
- [14] Wei JW, Huang K, Yang C and Kang CS. Non-coding RNAs as regulators in epigenetics (review). *Oncol Rep* 2017; 37: 3-9.
- [15] Ponting CP, Oliver PL and Reik W. Evolution and functions of long noncoding RNAs. *Cell* 2009; 136: 629-641.
- [16] Su M, Xiao Y, Ma J, Cao D, Zhou Y, Wang H, Liao Q and Wang W. Long non-coding RNAs in esophageal cancer: molecular mechanisms, functions, and potential applications. *J Hematol Oncol* 2018; 11: 118.
- [17] Geisler S and Collier J. RNA in unexpected places: long non-coding RNA functions in diverse cellular contexts. *Nat Rev Mol Cell Biol* 2013; 14: 699-712.
- [18] Quinn JJ and Chang HY. Unique features of long non-coding RNA biogenesis and function. *Nat Rev Genet* 2016; 17: 47-62.
- [19] Peng L, Yuan X, Jiang B, Tang Z and Li GC. LncRNAs: key players and novel insights into cervical cancer. *Tumour Biol* 2016; 37: 2779-2788.
- [20] Li J, Meng H, Bai Y and Wang K. Regulation of lncRNA and its role in cancer metastasis. *Oncol Res* 2016; 23: 205-217.
- [21] Kong X, Duan Y, Sang Y, Li Y, Zhang H, Liang Y, Liu Y, Zhang N and Yang Q. LncRNA-CDC6 promotes breast cancer progression and function as ceRNA to target CDC6 by sponging microRNA-215. *J Cell Physiol* 2019; 234: 9105-9117.
- [22] Hu W, Alvarez-Dominguez JR and Lodish HF. Regulation of mammalian cell differentiation by long non-coding RNAs. *EMBO Rep* 2012; 13: 971-983.
- [23] Rossi MN and Antonangeli F. LncRNAs: new players in apoptosis control. *Int J Cell Biol* 2014; 2014: 473857.
- [24] Lu Y, Sha H, Sun X, Zhang Y, Wu Y, Zhang J, Zhang H, Wu J and Feng J. CRNDE: an oncogenic long non-coding RNA in cancers. *Cancer Cell Int* 2020; 20: 162.
- [25] Gu X, Chu Q, Zheng Q, Wang J and Zhu H. The dual functions of the long noncoding RNA CASC15 in malignancy. *Biomed Pharmacother* 2021; 135: 111212.
- [26] Gu X, Zheng Q, Chu Q and Zhu H. HAND2-AS1: A functional cancer-related long non-coding RNA. *Biomed Pharmacother* 2021; 137: 111317.
- [27] Ghafouri-Fard S, Omrani MD and Taheri M. Long noncoding RNA PVT1: a highly dysregulated gene in malignancy. *J Cell Physiol* 2020; 235: 818-835.
- [28] Liu H, Liu T, Zhou Y, Song X and Wei R. Overexpression of long non-coding RNA cancer susceptibility 11 is involved in the development of chemoresistance to carboplatin in hepatocellular carcinoma. *Oncol Lett* 2020; 19: 1993-1998.
- [29] Luo H, Xu C, Le W, Ge B and Wang T. lncRNA CASC11 promotes cancer cell proliferation in bladder cancer through miRNA-150. *J Cell Biochem* 2019; 120: 13487-13493.
- [30] Zhang L, Kang W, Lu X, Ma S, Dong L and Zou B. LncRNA CASC11 promoted gastric cancer cell proliferation, migration and invasion in vitro by regulating cell cycle pathway. *Cell Cycle* 2018; 17: 1886-1900.
- [31] Song H, Liu Y, Li X, Chen S, Xie R, Chen D, Gao H, Wang G, Cai B and Yang X. Long noncoding RNA CASC11 promotes hepatocarcinogenesis and HCC progression through EIF4A3-mediated E2F1 activation. *Clin Transl Med* 2020; 10: e220.
- [32] Yan R, Jiang Y, Lai B, Lin Y and Wen J. The positive feedback loop FOXO3/CASC11/miR-498 promotes the tumorigenesis of non-small cell lung cancer. *Biochem Biophys Res Commun* 2019; 519: 518-524.
- [33] Zhang Z, Zhou C, Chang Y, Zhang Z, Hu Y, Zhang F, Lu Y, Zheng L, Zhang W, Li X and Li X. Long non-coding RNA CASC11 interacts with hnRNP-K and activates the WNT/ $\beta$ -catenin pathway to promote growth and metastasis in colorectal cancer. *Cancer Lett* 2016; 376: 62-73.
- [34] Fu Y, Zhang P, Nan H, Lu Y, Zhao J, Yang M and Song Q. LncRNA CASC11 promotes TGF- $\beta$ 1, increases cancer cell stemness and predicts postoperative survival in small cell lung cancer. *Gene* 2019; 704: 91-96.
- [35] Han Y, Chen M, Wang A and Fan X. STAT3-induced upregulation of lncRNA CASC11 promotes the cell migration, invasion and epithelial-mesenchymal transition in hepatocellular carcinoma by epigenetically silencing PTEN and activating PI3K/AKT signaling pathway. *Biochem Biophys Res Commun* 2019; 508: 472-479.
- [36] Jin J, Zhang S, Hu Y, Zhang Y, Guo C and Feng F. SP1 induced lncRNA CASC11 accelerates the glioma tumorigenesis through targeting FOXK1 via sponging miR-498. *Biomed Pharmacother* 2019; 116: 108968.
- [37] Zhang YX, Yuan J, Gao ZM and Zhang ZG. LncRNA TUC338 promotes invasion of lung cancer by activating MAPK pathway. *Eur Rev Med Pharmacol Sci* 2018; 22: 443-449.
- [38] Cao HL, Liu ZJ, Huang PL, Yue YL and Xi JN. lncRNA-RMRP promotes proliferation, migration and invasion of bladder cancer via miR-206. *Eur Rev Med Pharmacol Sci* 2019; 23: 1012-1021.
- [39] Zhuang C, Ma Q, Zhuang C, Ye J, Zhang F and Gui Y. LncRNA GClnc1 promotes proliferation and invasion of bladder cancer through activation of MYC. *Faseb j* 2019; 33: 11045-11059.
- [40] Li T, Chen Y, Zhang J and Liu S. LncRNA TUG1 promotes cells proliferation and inhibits cells apoptosis through regulating AURKA in epithelial

## CASC11 role in human cancers

- lial ovarian cancer cells. *Medicine (Baltimore)* 2018; 97: e12131.
- [41] Zhou S, He Y, Yang S, Hu J, Zhang Q, Chen W, Xu H, Zhang H, Zhong S, Zhao J and Tang J. The regulatory roles of lncRNAs in the process of breast cancer invasion and metastasis. *Biosci Rep* 2018; 38: BSR20180772.
- [42] Xiong HG, Li H, Xiao Y, Yang QC, Yang LL, Chen L, Bu LL, Zhang WF, Zhang JL and Sun ZJ. Long noncoding RNA MYOSLID promotes invasion and metastasis by modulating the partial epithelial-mesenchymal transition program in head and neck squamous cell carcinoma. *J Exp Clin Cancer Res* 2019; 38: 278.
- [43] Cheng N, Wu J, Yin M, Xu J, Wang Y, Chen X, Nie Z and Yin J. LncRNA CASC11 promotes cancer cell proliferation in hepatocellular carcinoma by inhibiting miRNA-188-5p. *Biosci Rep* 2019; 39: BSR20190251.
- [44] Tong W, Han TC, Wang W and Zhao J. LncRNA CASC11 promotes the development of lung cancer through targeting microRNA-302/CDK1 axis. *Eur Rev Med Pharmacol Sci* 2019; 23: 6539-6547.
- [45] Cui Y, Shen G, Zhou D and Wu F. CASC11 overexpression predicts poor prognosis and regulates cell proliferation and apoptosis in ovarian carcinoma. *Cancer Manag Res* 2020; 12: 523-529.
- [46] Hsu W, Liu L, Chen X, Zhang Y and Zhu W. LncRNA CASC11 promotes the cervical cancer progression by activating Wnt/beta-catenin signaling pathway. *Biol Res* 2019; 52: 33.
- [47] Su X, Zhang J, Luo X, Yang W, Liu Y, Liu Y and Shan Z. LncRNA LINC01116 promotes cancer cell proliferation, migration and invasion in gastric cancer by positively interacting with lncRNA CASC11. *Onco Targets Ther* 2019; 12: 8117-8123.
- [48] Yu Z, Zhang J and Han J. Silencing CASC11 curbs neonatal neuroblastoma progression through modulating microRNA-676-3p/nucleolar protein 4 like (NOL4L) axis. *Pediatr Res* 2020; 87: 662-668.
- [49] Chen SG, Wang CH, He RQ, Xu RY and Ji CB. LncRNA CASC11 promotes the development of esophageal carcinoma by regulating KLF6. *Eur Rev Med Pharmacol Sci* 2019; 23: 8878-8887.
- [50] Wong RS. Apoptosis in cancer: from pathogenesis to treatment. *J Exp Clin Cancer Res* 2011; 30: 87.
- [51] Goldar S, Khaniani MS, Derakhshan SM and Baradaran B. Molecular mechanisms of apoptosis and roles in cancer development and treatment. *Asian Pac J Cancer Prev* 2015; 16: 2129-2144.
- [52] Duff D and Long A. Roles for RACK1 in cancer cell migration and invasion. *Cell Signal* 2017; 35: 250-255.
- [53] van de Merbel AF, van der Horst G, Buijs JT and van der Pluijm G. Protocols for migration and invasion studies in prostate cancer. *Methods Mol Biol* 2018; 1786: 67-79.
- [54] Nyongesa CO and Park S. Chemotherapeutic resistance: a nano-mechanical point of view. *Biol Chem* 2018; 399: 1433-1446.
- [55] Shen F, Feng L, Zhou J, Zhang H, Xu Y, Jiang R, Zhang H and Chen Y. Overexpression of CASC11 in ovarian squamous cell carcinoma mediates the development of cancer cell resistance to chemotherapy. *Gene* 2019; 710: 363-366.
- [56] Bartonicek N, Maag JL and Dinger ME. Long noncoding RNAs in cancer: mechanisms of action and technological advancements. *Mol Cancer* 2016; 15: 43.
- [57] Zhang H, Chen Z, Wang X, Huang Z, He Z and Chen Y. Long non-coding RNA: a new player in cancer. *J Hematol Oncol* 2013; 6: 37.
- [58] Tay Y, Rinn J and Pandolfi PP. The multilayered complexity of ceRNA crosstalk and competition. *Nature* 2014; 505: 344-352.
- [59] Zhong Y, Du Y, Yang X, Mo Y, Fan C, Xiong F, Ren D, Ye X, Li C, Wang Y, Wei F, Guo C, Wu X, Li X, Li Y, Li G, Zeng Z and Xiong W. Circular RNAs function as ceRNAs to regulate and control human cancer progression. *Mol Cancer* 2018; 17: 79.
- [60] Kartha RV and Subramanian S. Competing endogenous RNAs (ceRNAs): new entrants to the intricacies of gene regulation. *Front Genet* 2014; 5: 8.
- [61] Bomsztyk K, Denisenko O and Ostrowski J. hnRNP K: one protein multiple processes. *Bioessays* 2004; 26: 629-638.
- [62] Bomsztyk K, Van Seuning I, Suzuki H, Denisenko O and Ostrowski J. Diverse molecular interactions of the hnRNP K protein. *FEBS Lett* 1997; 403: 113-115.
- [63] Wang X, Kaczor-Urbanowicz KE and Wong DT. Salivary biomarkers in cancer detection. *Med Oncol* 2017; 34: 7.
- [64] Hristova VA and Chan DW. Cancer biomarker discovery and translation: proteomics and beyond. *Expert Rev Proteomics* 2019; 16: 93-103.
- [65] Wu L and Qu X. Cancer biomarker detection: recent achievements and challenges. *Chem Soc Rev* 2015; 44: 2963-2997.
- [66] Cabili MN, Trapnell C, Goff L, Koziol M, Tazon-Vega B, Regev A and Rinn JL. Integrative annotation of human large intergenic noncoding RNAs reveals global properties and specific subclasses. *Genes Dev* 2011; 25: 1915-1927.
- [67] Rivandi M, Pasdar A, Hamzadeh L, Tajbakhsh A, Seifi S, Moetamani-Ahmadi M, Ferns GA and Avan A. The prognostic and therapeutic values of long noncoding RNA PANDAR in colorectal cancer. *J Cell Physiol* 2019; 234: 1230-1236.
- [68] Kumar MM and Goyal R. LncRNA as a therapeutic target for angiogenesis. *Curr Top Med Chem* 2017; 17: 1750-1757.

## CASC11 role in human cancers

- [69] Pan X, Zheng G and Gao C. LncRNA PVT1: a novel therapeutic target for cancers. *Clin Lab* 2018; 64: 655-662.
- [70] Botti G, Marra L, Malzone MG, Anniciello A, Botti C, Franco R and Cantile M. LncRNA HOTAIR as prognostic circulating marker and potential therapeutic target in patients with tumor diseases. *Curr Drug Targets* 2017; 18: 27-34.
- [71] Liang Z, Zhu B, Meng D, Shen X, Li X, Wang Z and Li L. Down-regulation of lncRNA-NEF indicates poor prognosis in intrahepatic cholangiocarcinoma. *Biosci Rep* 2019; 39: BSR20181573.
- [72] Wang D, Dai J, Hou S and Qian Y. LncRNA SNHG20 predicts a poor prognosis and promotes cell progression in epithelial ovarian cancer. *Biosci Rep* 2019; 39: BSR20182186.
- [73] Yin Q, Shen X, Cui X and Ju S. Elevated serum lncRNA TUG1 levels are a potential diagnostic biomarker of multiple myeloma. *Exp Hematol* 2019; 79: 47-55, e42.
- [74] Garzon R, Volinia S, Papaioannou D, Nicolet D, Kohlschmidt J, Yan PS, Mrózek K, Bucci D, Carroll AJ, Baer MR, Wetzler M, Carter TH, Powell BL, Kolitz JE, Moore JO, Eisfeld AK, Blachly JS, Blum W, Caligiuri MA, Stone RM, Marcucci G, Croce CM, Byrd JC and Bloomfield CD. Expression and prognostic impact of lncRNAs in acute myeloid leukemia. *Proc Natl Acad Sci U S A* 2014; 111: 18679-18684.