### Original Article

# Plasma hydrogen sulfide as a promising predictor of collateral cerebral circulation in acute ischemic stroke

Haiyan Chen<sup>1,2,3</sup>, Jianmin Huang<sup>2</sup>, Pin Zheng<sup>4</sup>, Guixin Yang<sup>2</sup>, Bingbing Qin<sup>2</sup>, Mengxue Zang<sup>2</sup>, Jie Wang<sup>5</sup>, Xuebin Li<sup>1,2,3,6</sup>

¹The First Clinical Medical College of Jinan University, Guangzhou 510000, Guangdong, China; ²Department of Neurology, The Affiliated Hospital of Youjiang Medical University for Nationalities, Baise 533000, Guangxi, China; ³Key Laboratory of Research on Clinical Molecular Diagnosis for High Incidence Diseases in Western Guangxi of Guangxi Higher Education Institutions, Baise 533000, Guangxi, China; ⁴Department of Gastroenterology, The Affiliated Hospital of Youjiang Medical University for Nationalities, Baise 533000, Guangxi, China; ⁵Department of Nephrology, The Affiliated Hospital of Youjiang Medical University for Nationalities, Baise 533000, Guangxi, China; ⁵Youjiang Medical University for Nationalities, Baise 533000, Guangxi, China

Received May 19, 2025; Accepted July 23, 2025; Epub July 25, 2025; Published July 30, 2025

Abstract: Objectives: To investigate the relationship between plasma hydrogen sulfide (H2S) levels and collateral circulation in patients with acute ischemic stroke (AIS), and to explore the effects of vascular endothelial growth factor (VEGF), homocysteine (Hcy), folic acid (FA), vitamin B<sub>12</sub> (VB<sub>12</sub>), and vitamin B<sub>6</sub> (VB<sub>6</sub>). Methods: A total of 68 AIS patients were enrolled and classified into two groups based on collateral vessel grading: the Good Collateral Circulation (GCC) group (n = 37) and the Poor Collateral Circulation (PCC) group (n = 31). Plasma levels of  $H_2S$ , VEGF, Hcy, FA, VB<sub>12</sub>, and VB<sub>8</sub> were measured on the  $2^{nd}$  and  $7^{th}$  days after admission using microassays and ELISA. Results: Compared to the PCC group, patients in the GCC group had significantly lower National Institutes of Health Stroke Scale (NIHSS) and Modified Rankin Scale scores (mRS) scores at both admission and discharge (all P < 0.05). On both the  $2^{nd}$  and  $7^{th}$  days, plasma levels of  $H_2S$ , VEGF, FA,  $VB_{12}$ , and  $VB_6$  were significantly higher in the GCC group, while Hcy levels were significantly lower (all P < 0.05). Within group comparisons between the two time points also showed significant changes (all P < 0.001). Correlation analysis revealed that plasma H<sub>2</sub>S levels were positively correlated with collateral circulation, VEGF, FA, VB<sub>1.7</sub>, and VB<sub>8</sub>, and negatively correlated with Hcy levels (all P < 0.001). Plasma H<sub>2</sub>S levels demonstrated high predictive value for collateral circulation (area under the curve, AUC = 0.943). An interaction between time and collateral circulation on HaS levels was also observed. Conclusions: Plasma HaS levels may serve as a valuable biomarker for predicting good collateral circulation in patients with acute ischemic stroke.

**Keywords:** Plasma hydrogen sulfide, collateral circulation, acute ischemic stroke, vascular endothelial growth factor, homocysteine

#### Introduction

Stroke is a common acute cerebrovascular disorder with significant sequelae, resulting from ischemia or hypoxia in brain tissue due to vascular obstruction or rupture. It primarily manifests in two forms: ischemic stroke (cerebral infarction) and hemorrhagic stroke (cerebral hemorrhage), with ischemic stroke being the more prevalent. Stroke is the second leading cause of death globally, with approximately 15 million people experiencing their first stroke each year [1]. According to the 2021 Report on Stroke Prevention and Treatment in China, the

overall incidence of stroke in China continues to rise [2]. Although the rate of hemorrhagic stroke has declined, the incidence of ischemic stroke has increased in recent years, with a five year recurrence rate of 41% and a mortality rate of 16%. Stroke is now the leading cause of death and disability among adults in China [2]. Current treatments, such as intravenous thrombolysis and endovascular thrombectomy, aim to restore vascular patency, improving cerebral perfusion, and limit irreversible neurological damage in patients with acute ischemic stroke (AIS) [3, 4]. However, these therapies are constrained by narrow therapeutic time windows,

high costs, and associated complications, thus benefiting only a minority of patients.

Emerging research has shown that cerebral collateral circulation (CCC) is activated as a secondary compensatory mechanism following acute cerebral ischemia. CCC can provide supplementary perfusion to ischemic areas, making it a promising target for therapeutic intervention [5, 6]. CCC refers to alternative blood flow pathways that form when major cervical or intracranial vessels are stenosed or occluded. These collaterals bypass the obstructed vessels and deliver oxygen and nutrients to the ischemic penumbra, improving tissue viability. Studies have demonstrated a strong correlation between the presence of CCC and improved clinical outcomes after cerebral infarction [7-9]. Patients with well-developed CCC have been shown to exhibit lower NIH Stroke Scale (NIHSS) scores, smaller ischemic penumbra volumes, and reduced infarct core size compared to those with poor CCC [10]. Therefore, accurate assessment of CCC and its relationship to prognosis is of significant clinical importance.

Hydrogen sulfide (H<sub>2</sub>S) is the third endogenous gaseous signaling molecule identified, primarily sunthesized in the central nervous system via the degradation of L-cysteine by cystathionine β-synthase (CBS) [11]. L-cysteine itself is a product of homocysteine (Hcy) metabolism. Hyperhomocysteinemia (HHcy), characterized by elevated plasma Hcy levels, is an established independent risk factor for cerebrovascular disease. HHcy may result from reduced CBS activity, genetic mutations in the 5,10methylene tetrahydrofolate reductase (MTHFR) gene, and deficiencies in vitamin cofactors such as folic acid (FA), vitamin B<sub>12</sub> (VB<sub>12</sub>), and vitamin B<sub>6</sub> (VB<sub>6</sub>). Additionally, reduced plasma H<sub>2</sub>S concentrations may contribute to the development of HHcy in AIS [12, 13]. Previous studies have shown that serum HaS levels peak within 24 hours of AIS onset, then decline between days 1 to 4 before partially rebounding on days 4 to 5, though still remaining below initial levels [14]. Moreover, serum HaS levels have been inversely correlated with cerebral infarct volume. In an MCAO (middle cerebral artery occlusion) mouse model, Tao et al. [15] reported that low-dose sodium hydrosulfide (NaHS) administration reduced inflammatory responses and improved neurological function. Researchers Jiang et al. [16] found that NaHS injection in MCAO rats reduced infarct volume and the expression of apoptosis-related proteins, demonstrating a neuroprotective effect. Furthermore. Another study in hemorrhagic stroke models showed that NaHS promoted angiogenesis and improved neurological outcomes [17]. Despite these neuroprotective roles of HaS, its involvement in the development of collateral circulation after cerebral infarction remains insufficiently understood. We hypothesize that H<sub>2</sub>S may facilitate angiogenesis following cerebral ischemia and serve as an useful biomarker for evaluating CCC status.

Vascular endothelial growth factor (VEGF) plays a key role in neovascularization after cerebral ischemia [18]. Clinical studies have demonstrated elevated serum VEGF levels in stroke patients compared to healthy controls, with higher levels observed in those with good CCC compared to poor CCC [19]. In stroke model mice, intraventricular injection of VEGF increased microvessel density in the ischemic penumbra and reduced infarct size [20]. Zhang et al. [21] further reported that NaHS enhanced Akt phosphorylation, endothelial cell migration. and tube formation, while also upregulating VEGFR2 expression, thereby promoting angiogenesis and facilitating recovery from hypoxiareoxygenation repair. These findings suggest that VEGF expression contributes to vascular remodeling and neuronal repair, potentially improving neurological outcomes. However, the relationship between plasma H<sub>2</sub>S levels, VEGF concentrations, and the status of collateral circulation in AIS patients has not been fully elucidated. Therefore, this study aims to examine the correlation between peripheral blood H<sub>a</sub>S levels and CCC in AIS, and to explore how VEGF, Hcy, FA, VB<sub>6</sub>, and VB<sub>12</sub> influence plasma H<sub>2</sub>S levels. The goal is to identify reliable biomarkers for predicting CCC status and to provide novel insights into targeted therapeutic strate-

#### Materials and methods

#### Subjects

A total of 68 inpatients diagnosed with cerebral infarction were retrospectively screened between January 2023 and May 2024 at the Department of Neurology, Affiliated Hospital of Youjiang Medical University for Nationalities. Based on digital subtraction angiography (DSA) findings, patients were clssified into two groups: the Good Collateral Circulation (GCC) group (n = 37) and the Poor Collateral Circulation (PCC) group (n = 31). This study was approved by the Ethics Committee of the Affiliated Hospital of Youjiang Medical University for Nationalities (Approval Number: YYFY-LL-2023-039).

Inclusion criteria: (1) First episode of cerebral infarction within  $\leq 3$  days of symptom onset. Diagnosis was based on the Chinese Guidelines for Diagnosis and Treatment of Acute Ischemic Stroke [22], which defines acute onset with focal neurological deficits (e.g., unilateral facial or limb weakness/numbness, speech impairment) lasting over 24 hours, with cerebral infarction confirmed by cranial magnetic resonance diffusion-weighted imaging. Non-vascular etiologies were excluded. (2) DSA examination confirming middle cerebral artery stenosis or occlusion  $\geq$  70% on one side of the brain. (3) Classification into the large-artery atherosclerosis subtype based on the Trial of Org 10172 in Acute Stroke Treatment classification [23]. (4) Patients who exceeded the time window for intravenous thrombolysis and arterial thrombectomy at admission and did not receive endovascular treatment. (5) Age between 18 and 80 years. (6) Complete clinical data.

Exclusion criteria: (1) History of cerebral infarction, cerebral hemorrhage, craniocerebral trauma, brain tumor, or similar conditions. (2) Cerebral infarction due to cardiogenic embolism, intracranial infection, vasculitis, subarachnoid hemorrhage, intracranial tumor, or blood disorders. (3) Serious concurrent diseases such as heart, lung, or liver diseases, or complications like renal insufficiency, severe infections, or tumors. (4) Failure to cooperate with blood reexamination, cranial MRI, or DSA examination. (5) Incomplete clinical data or refusal to participate.

## Assessment of collateral cerebral circulation and grouping

On the day after admission, patients underwent DSA examination after providing informed consent. The degree of cerebral artery stenosis was calculated using the formula:

intracranial artery stenosis rate = (1- the diameter of the narrowest intracranial artery/the diameter of the normal part) × 100% [24].

CCC was evaluated by two interventional physicians using the collateral vessel grading system from the American Society of Interventional and Therapeutic Neuroradiology/Society of Interventional Radiology [25]. The CCC state was categorized as "good" (grades 0-1) or "poor" (grades 2-4), and patients were assigned to the Good Collateral Circulation (GCC) group (n = 37) and the Poor Collateral Circulation (PCC) group (n = 31) accordingly. A typical DSA image of patients in each group is shown in **Figure 1**.

#### Treatment plan

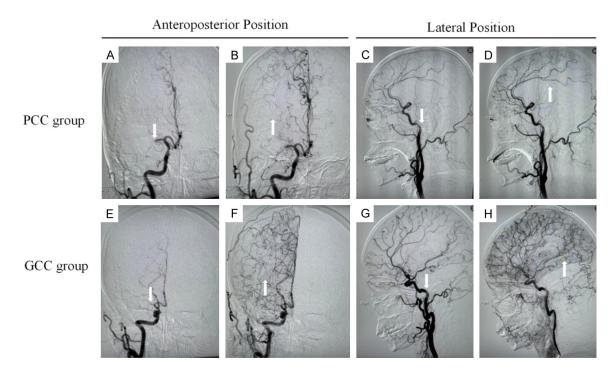
All patients with cerebral infarction were treated with aspirin and/or clopidogrel hydrogen chloride, atorvastatin, butylphthalide, urinary kallikreinogenase, idebenone, and other drugs according to the Chinese Guidelines for Diagnosis and Treatment of Acute Ischemic Stroke. Patients with hypertension and diabetes received appropriate antihypertensive and hypoglycemic treatments. Edaravone, FA, VB<sub>6</sub>, and VB<sub>12</sub> were not administered to any patient before or during hospital treatment.

#### Clinical data collection

Age, gender, blood glucose, blood pressure, medical history (diabetes, hypertension, hyperlipidemia, stroke, coronary heart disease, etc.), smoking history, drinking history, and other relevant information were collected for all included subjects. The National Institutes of Health Stroke Scale (NIHSS) and Modified Rankin Scale (mRS) scores were recorded by the attending physician at the time of admission, taking the patient's condition into consideration, and the prognosis was assessed by the neurologist using the NIHSS and mRS scores again at the time of discharge.

#### Sample collection

Fasting venous blood (3 mL) was collected from patients with cerebral infarction on the 2<sup>nd</sup> and 7<sup>th</sup> days of admission, using an EDTA anticoagulation tube. After collection, the blood was gently mixed by inversion, left to stand for 20 minutes, and then centrifuged at 3000 rpm for 20



**Figure 1.** DSA results of different collateral circulation states. Panels (A-D) show the DSA results on the day after admission for a patient in the PCC group. Panels (A and C): Right middle cerebral artery occlusion (anteroposterior and lateral views). Panels (B and D): The right anterior cerebral artery leptomeningeal vessels compensating for the pericortical blood supply to the ipsilateral middle cerebral artery, with sparse and slow blood flow, and a collateral circulation score of 1 (anteroposterior and lateral views). Panels (E-H) show the DSA results on the day after admission for a patient in the GCC group. Panels (E and G): Right middle cerebral artery occlusion (anteroposterior and lateral views). Panels (F and H): Formation of leptomeningeal vessels and capillary networks from the right external carotid artery and right anterior cerebral artery, compensating for the blood supply to the entire region of the ipsilateral middle cerebral artery, with a collateral circulation score of 4 (anteroposterior and lateral views). DSA, digital subtraction angiography; PCC, poor collateral circulation; GCC, good collateral circulation.

minutes. The supernatant was transferred into a sterile high-pressure 1.5 mL EP tube for subsequent analysis. The remaining samples were aliquoted and stored at -80°C to avoid repeated freezing and thawing.

Detection of plasma H<sub>2</sub>S level by microassay

A 150  $\mu$ L plasma sample was added to the sample tube, and a blank tube was prepared. Samples were processed according to the instructions provided with the H $_2$ S kit (Enzymelinked Biological, mI076943). OD values for each well were measured using an enzymelabeled instrument, and  $\Delta$ A values (A determination - A blank) for each well were calculated. H $_2$ S concentration was then calculated using the following formula:

$$H_2S \text{ (nmol/mL)} = \frac{\Delta A}{0.0022} \times V_{\text{total}} \div V_{\text{sample}} = 681.8 \times \Delta A$$

 $V_{\text{total}}$ : total reaction volume;  $V_{\text{sample}}$ : volume of sample in reaction.

Detection of plasma VEGF,  $VB_{6'}$ ,  $VB_{12'}$ , FA and HCY levels by ELISA

A 50  $\mu$ L plasma sample was added to the reaction well, with blank wells, control group wells, and standard curve wells prepared. The assays for VEGF (Quanzhou Ruixin Biotechnology Co., Ltd., RX105003H), VB $_{6}$  (Quanzhou Ruixin Biotechnology Co., Ltd., RXJ105135H), VB $_{12}$  (Quanzhou Ruixin Biotechnology Co., Ltd., RXJ105137H), FA (Quanzhou Ruixin Biotechnology Co., Ltd., RXJ106799H), and Hcy (Quanzhou Ruixin Biotechnology Co., Ltd., RXJ105166H) were conducted according to the manufacturers' instructions. Enzyme-labeled antibodies were added for incubation. After incubation, the wells were washed with PBS, followed by the addition of substrate

**Table 1.** Comparison of clinical data (n (%),  $\overline{X} \pm s$ , M ( $P_{25}$ ,  $P_{75}$ ))

|                          | GCC group (n = 37) | PCC group (n = $31$ ) | $t/Z/\chi^2$ | Р       |
|--------------------------|--------------------|-----------------------|--------------|---------|
| Gender (Male/Female)     | 25/12              | 23/8                  | 0.466        | 0.495   |
| Age (years)              | 56 (49.00, 66.00)  | 63 (55.00, 70.00)     | 1.782        | 0.075   |
| Diabetes history         | 9 (23.1%)          | 4 (12.9%)             | 0.605        | 0.432   |
| Hypertension history     | 32 (82.1%)         | 17 (54.8%)            | 6.090        | 0.014   |
| Smoking history          | 21 (53.8%)         | 9 (29.0%)             | 4.342        | 0.037   |
| Alcohol history          | 21 (53.8%)         | 10 (32.3%)            | 3.262        | 0.071   |
| mRS score at admission   | 2.0 (2.0, 3.0)     | 3.0 (2.0, 4.0)        | 3.841        | < 0.001 |
| mRS score at discharge   | 1.0 (1.0, 2.0)     | 2.0 (1.0, 2.0)        | 3.701        | < 0.001 |
| NIHSS score at admission | 10.0 (8.0, 14.0)   | 16.0 (12.0, 20.0)     | 4.454        | < 0.001 |
| NIHSS score at discharge | 6.36 ± 2.87        | 10.26 ± 3.45          | -5.159       | < 0.001 |

Note: GCC, Good Collateral Circulation; PCC, Poor Collateral Circulation; mRS, Modified Rankin Scale; NIHSS, National Institutes of Health Stroke Scale.

developing solution for color reaction. The reaction was stopped by adding the corresponding stop solution, and optical density was measured using a microplate reader. The concentrations of VEGF,  ${\rm VB}_{\rm 6}$ ,  ${\rm VB}_{\rm 12}$ , FA, and Hcy in each plasma sample were determined from the standard curve values.

#### Statistical analysis

Data were analyzed and visualized using GraphPad Prism 9.0 software. Normality testing was performed on all measurement data. Data conforming to a normal distribution were expressed as mean ± standard deviation  $(\bar{x} \pm sd)$ . An independent sample t-test was used for comparisons between two groups, while a paired sample t-test was used for comparisons at different time points within a group. Data that were not normally distributed were expressed as M (P<sub>25</sub>, P<sub>75</sub>), and the Mann-Whitney U test was used for comparisons between two samples. Count data were presented as frequency (rate) and analyzed using the  $\chi^2$  test. Spearman rank correlation was used to assess the correlation between HaS concentration and collateral circulation state, as well as between H<sub>2</sub>S and VEGF, Hcy, FA, VB<sub>6</sub>, and VB<sub>12</sub>. Differences were considered statistically significant at P < 0.05.

#### Results

#### Comparison of clinical data

No significant differences were observed between the groups regarding age, gender, diabetes, or drinking history (P > 0.05). The NIHSS

and mRS scores on admission and discharge were significantly higher in the PCC group compared to the GCC group (all P < 0.001), as shown in **Table 1**.

#### Comparison of plasma H<sub>2</sub>S levels

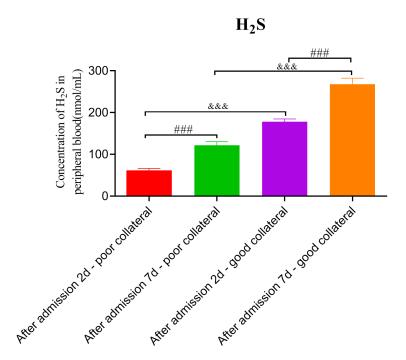
To evaluate the plasma  $\rm H_2S$  levels in patients under different CCC states,  $\rm H_2S$  was measured using a microassay method. Results showed that plasma  $\rm H_2S$  levels in the GCC group were significantly higher than those in the PCC group on both the  $\rm 2^{nd}$  and  $\rm 7^{th}$  days of admission (t = 13.875 and 8.776, both P < 0.001). Additionally, plasma  $\rm H_2S$  levels on the  $\rm 7^{th}$  day were significantly higher than on the  $\rm 2^{nd}$  day for both groups (t = 8.271 and 6.817, both P < 0.001). See **Figure 2**.

#### Comparison of plasma VEGF levels

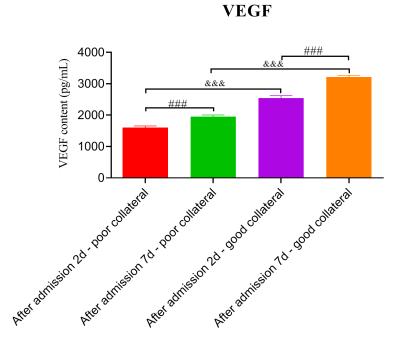
Plasma VEGF levels were measured by ELISA to assess the VEGF concentrations under different collateral circulation states. The results indicated that plasma VEGF levels were significantly higher in the GCC group compared to the PCC group on both the  $2^{nd}$  and  $7^{th}$  days of admission (t = 43.894 and 64.245, both P < 0.001). The plasma VEGF levels in both groups were higher on the  $7^{th}$  day than on the  $2^{nd}$  day (t = 18.021 and 38.339, both P < 0.001). See **Figure 3** 

Comparison of plasma HCY, FA,  ${\rm VB_6}$  and  ${\rm VB_{12}}$  levels

Hcy generates L-cysteine and H<sub>2</sub>S through CBS, and its level is influenced by cofactors FA, VB<sub>6</sub>,



**Figure 2.** Levels of H<sub>2</sub>S in peripheral blood in different collateral circulation. Note: Paired t test \*\*#P < 0.001; independent t test, \*&&P < 0.001. Plasma hydrogen sulfide (H<sub>2</sub>S).



**Figure 3.** Peripheral blood VEGF levels in different collateral circulation. Note: Paired t test \*\*\*P < 0.001; independent t test \*&\*P < 0.001. VEGF, vascular endothelial growth factor.

and  $VB_{12}$ . Plasma levels of Hcy, FA,  $VB_{6}$ , and  $VB_{12}$  were measured by ELISA to explore their

relationship in the two groups. The results showed that plasma Hcy levels in the PCC group were significantly higher than in the GCC group on both the 2<sup>nd</sup> and 7<sup>th</sup> days of admission (t = 16.637 and 13.264, both P < 0.001). Plasma Hcy levels on the 7th day in both groups were significantly lower than on the  $2^{nd}$  day (t = 6.371 and 13.090, both P < 0.001). Regarding plasma VB<sub>6</sub>, VB<sub>12</sub>, and FA, the PCC group had significantly lower levels than the GCC group on both the 2<sup>nd</sup> and  $7^{\text{th}}$  days (t = 18.626, 7.306 for  $VB_{e}$ ; t = 19.400, 19.630 for  $VB_{12}$ ; t = 13.464, 7.791 for FA; all P < 0.001). On the  $7^{th}$ day, plasma VB<sub>6</sub>, VB<sub>12</sub>, and FA levels were higher than on the  $2^{nd}$  day in both groups (t =18.969, 11.547 for  $VB_a$ ; t =11.674, 19.630 for  $VB_{12}$ ; t =21.048, 17.977 for FA; all P < 0.001). See Figure 4.

Correlation analysis of H<sub>2</sub>S and collateral circulation state

To explore the correlation between plasma  $\rm H_2S$  levels and collateral circulation status, patients' collateral circulation scores were assigned, with collateral circulation dysfunction = 1 and GCC = 2. Bivariate Spearman's correlation analysis showed that plasma  $\rm H_2S$  levels were positively correlated with collateral circulation status, with a correlation coefficient of 0.855 (P < 0.001), as shown in **Table 2**.

Correlation analysis between  $H_2S$  and levels of VEGF, Hcy, FA,  $VB_8$  and  $VB_{12}$ 

To explore the correlation between plasma H<sub>2</sub>S levels and VEGF, Hcy, FA, VB<sub>6</sub>, and VB<sub>12</sub>

levels in the two patient groups, a bivariate Spearman's correlation analysis was per-

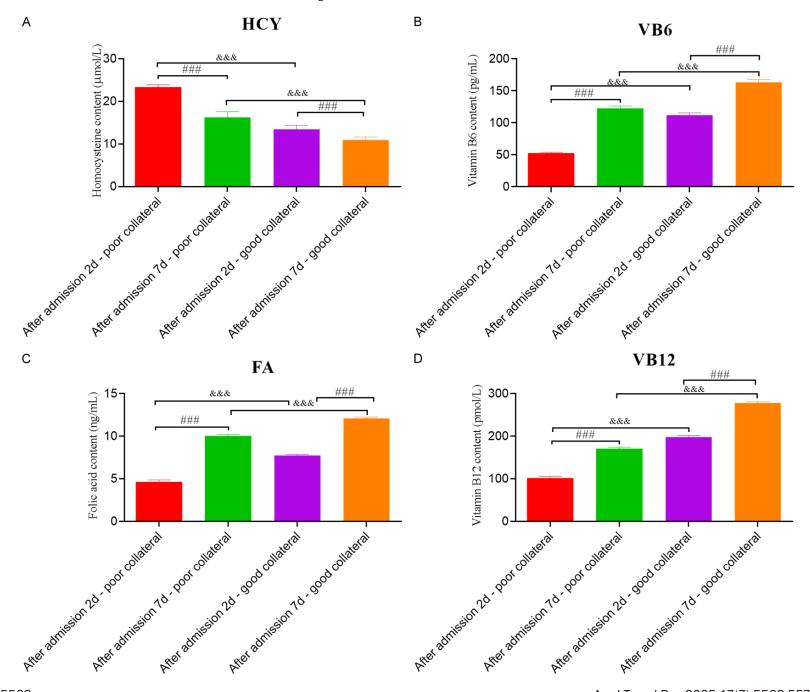


Figure 4. Levels of Hcy, FA, VB<sub>6</sub> and VB<sub>12</sub> in peripheral blood in different collateral circulation. Note: Paired t test \*\*#P < 0.001; independent t test \*&&P < 0.001. Hcy, homocysteine; VB<sub>6</sub>, vitamin B<sub>6</sub>; FA, folic acid; VB<sub>12</sub>, vitamin B<sub>12</sub>.

**Table 2.** Correlation analysis between H<sub>2</sub>S and collateral circulation score

|   | r     | Р       |
|---|-------|---------|
| collateral circulation & H <sub>2</sub> S | 0.855 | < 0.001 |

Note: valuation of collateral circulation: collateral circulation dysfunction = 1; GCC =2, PCC group n = 31, GCC group n = 37.

**Table 3.** Correlation analysis between H<sub>2</sub>S and each index

|                  | r      | Р       |
|------------------|--------|---------|
| VEGF             | 0.696  | < 0.001 |
| Hcy              | -0.664 | < 0.001 |
| FA               | 0.759  | < 0.001 |
| VB <sub>12</sub> | 0.610  | < 0.001 |
| VB <sub>6</sub>  | 0.543  | < 0.001 |

Note: VEGF, vascular endothelial growth factor; Hcy, homocysteine; FA, folic acid;  $VB_{12}$ , vitamin  $B_{12}$ ;  $VB_{6}$ , vitamin  $B_{e}$ .

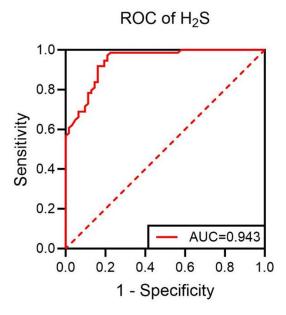
formed. The results showed that plasma  $\rm H_2S$  levels were positively correlated with VEGF, FA, VB $_{\rm e}$ , and VB $_{\rm 12}$ , with correlation coefficients of 0.696, 0.759, 0.610, and 0.543, respectively (all P < 0.001). Conversely, plasma  $\rm H_2S$  levels were negatively correlated with Hcy, with a correlation coefficient of -0.664, which was also statistically significant (P < 0.001). Detailed results are presented in **Table 3**.

Predictive value of H<sub>2</sub>S for collateral circulation

Receiver Operating Characteristic (ROC) curve analysis revealed that the Area Under the Curve (AUC) for plasma  $\rm H_2S$  level as a predictor of collateral circulation was 0.943 (95% Confidence Interval [CI]: 0.907-0.978), with a diagnostic sensitivity of 60.81% and specificity of 98.39% (P < 0.001, **Figure 5**).

Interaction of time and collateral circulation on  $H_aS$  change

Interactive analysis demonstrated that after adjusting for related variables (including VEGF, Hcy, FA,  $VB_6$ , and  $VB_{12}$  levels), the change in  $H_2S$  levels between the two groups was significantly different (all P < 0.001, Figure 6).

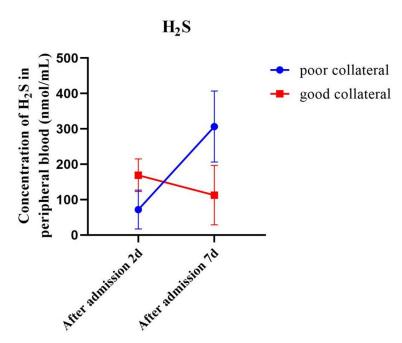


**Figure 5.** ROC curve of H<sub>2</sub>S expression on collateral circulation. ROC, receiver operator characteristic curve; AUC, Area Under The Curve. Plasma hydrogen sulfide (H<sub>2</sub>S).

#### Discussion

Cerebral infarction is a prevalent cerebrovascular condition characterized by high rates of mortality and recurrence, posing a substantial economic burden on patients and families and increasing psychological stress [26]. The formation of collateral circulation plays a pivotal role in both the treatment and prognosis of cerebral infarction. Upon onset, collateral vessels facilitate blood flow to ischemic regions, thereby enhancing vascular recanalization and maintaining perfusion in the ischemic penumbra. This can ultimately help reduce infarct size [27]. Good collateral circulation (GCC) is also a key factor in selecting candidates for endovascular intervention [28]. Notably, the recanalization rate in patients with GCC after thrombolysis reaches 61.80%, compared to only 28.10% in those with poor collateral circulation (PCC) [29], highlighting the critical value of GCC in endovascular treatment strategies.

In this study, we used the ASITN/SIR collateral grading system based on DSA imaging to assess collateral circulation. This system is



**Figure 6.** Interaction of time and collateral circulation on  $\rm H_2S$  change. Plasma hydrogen sulfide ( $\rm H_2S$ ).

widely utilized due to its reproducibility and diagnostic consistency [30, 31]. A prospective study from the Interventional Management of Stroke III trial showed that patients with ASITN/ SIR grades 3 or 4 had significantly better rates of recanalization, reperfusion, and functional recovery [32]. Collateral circulation development is influenced by age, genetics, and various cerebrovascular risk factors or biomarkers [33]. Homocysteine (Hcy) has been identified as a key risk factor; vitamins B<sub>6</sub>, B<sub>12</sub>, and folic acid (FA) are integral to Hcy metabolism [12]. Additionally, vascular endothelial growth factor (VEGF) is a well-established regulator of collateral formation in ischemic stroke [34]. Thus, we explored peripheral blood levels of H2S, VEGF, VB<sub>6</sub>, VB<sub>12</sub>, Hcy, and FA in cerebral infarction patients to assess their associations with collateral circulation.

Hydrogen sulfide  $({\rm H_2S})$ , a gaseous signaling molecule, has attracted growing attention in stroke research. Inflammatory responses following cerebral infarction may impact cerebral hemodynamics and collateral vessel formation. Accumulating evidence indicates that  ${\rm H_2S}$  exerts anti-inflammatory, antioxidant, and neuroprotective effects in neurological disorders [35]. It has been shown to protect against cerebral ischemia-reperfusion injury, potentially

by suppressing excessive autophagy activation [36]. Thus, increased H<sub>2</sub>S levels may improve cerebral blood flow, enhance microcirculation, and facilitate collateral development. Plasma H<sub>2</sub>S levels have been implicated in stroke pathophysiology [14]; in a study of 71 elderly stroke patients, plasma H<sub>a</sub>S levels initially declined and later increased as the disease progressed, with a negative correlation observed between H<sub>2</sub>S levels and infarct size. Our results were consistent with this trend-plasma H<sub>o</sub>S levels on days 2 and 7 postadmission were significantly higher in the GCC group versus the PCC group, and levels rose from day 2 to day 7 in both groups. Additionally, NIHSS and mRS scores at admission

and discharge were significantly lower in the GCC group. These findings suggest that cerebral ischemia may stimulate  $\rm H_2S$  production, offering neuroprotection. Plasma  $\rm H_2S$  may thus serve as a potential biomarker for predicting GCC. One study [37] indicated that the neuroprotective effects of  $\rm H_2S$  are dose-dependent: low concentrations confer benefits, while high concentrations may cause neurotoxicity. Our data suggest that levels between 61-268  $\rm \mu mol/L$  are within the protective range and may facilitate collateral formation, potentially through  $\rm H_2S$ -mediated VEGF upregulation.

VEGF is a key factor in angiogenesis and collateral vessel formation after stroke [19, 38]. It promotes endothelial cell proliferation and migration, enhancing vascular regeneration and perfusion of ischemic areas [39]. Animal studies revealed that VEGF mRNA peaks between days 1-3 following hypoxia and declines after day 7 [40], suggesting a time-sensitive regulatory role in ischemia. In this study, VEGF treatment of cortical neurons increased Bcl-2 while reducing Bax and TREK protein levels, suggesting a neuroprotective mechanism via TWIK channels. Li et al. [41] reported that VEGF levels rise early after AIS onset and peak at 506.68 ng/L between days 3-7, correlating with angiogenesis and collateral formation. In

our study, VEGF levels were significantly higher in the GCC group on both day 2 and day 7, with levels rising over time in both groups. These results affirm the role of VEGF in collateral development and suggest that elevated VEGF may serve as a marker of GCC.

Our findings further suggest a synergistic relationship between  $\rm H_2S$  and VEGF in collateral circulation.  $\rm H_2S$  modulates angiogenesis, endothelial proliferation, and vascular tone, partly by stimulating VEGFR2 signaling and reducing intracellular calcium [42].  $\rm H_2S$  may directly act on VEGF receptors by disrupting disulfide bonds (Cys1045-Cys1024) in VEGFR2, enhancing endothelial activity [43]. Spearman correlation analysis showed a positive correlation between  $\rm H_2S$  and VEGF levels, supporting a regulatory interaction that may promote vascular remodeling and collateral development.

Hcy impairs endothelial function by generating reactive oxygen species (ROS), promoting apoptosis, and inhibiting VEGF expression, thereby hindering collateral formation [44]. Elevated Hcy increases platelet aggregation and blood viscosity, further limiting perfusion [45]. Our study found that Hcy levels were significantly higher in the PCC group on both day 2 and day 7, consistent with these mechanisms.

Previous studies have demonstrated a negative correlation between H<sub>2</sub>S and Hcy levels in stroke patients [12], a finding replicated in our study. This suggests that H<sub>2</sub>S may attenuate Hcy toxicity, promoting collateral formation. Moreover, Hcy is metabolized via remethylation and transsulfuration pathways, requiring VB<sub>12</sub>, FA, and VB<sub>6</sub>. Deficiencies in these vitamins result in elevated Hcy levels, a known independent risk factor for stroke [46]. Studies have shown that low FA and VB<sub>12</sub> increase stroke risk [47], and lower levels of these vitamins are observed in stroke patients with cognitive impairment [48]. Clinical intervention with FA and VB reduces Hcy and improves neurological scores in AIS [49]. In our study, levels of VB<sub>6</sub>, VB<sub>12</sub>, and FA were significantly lower in the PCC group and positively correlated with HaS, while Hcy levels showed an inverse trend. These results suggest that low vitamin levels contribute to PCC, and the H<sub>2</sub>S-Hcy-vitamin axis may influence collateral status.

This study has limitations. Its retrospective, single-center design and small sample size may affect generalizability and introduce bias. Furthermore, some relevant markers could not be evaluated due to the retrospective nature. Future multicenter, prospective studies are needed to confirm our findings and elucidate the underlying mechanisms.

In conclusion, plasma  $\rm H_2S$  levels are positively associated with GCC in cerebral infarction and may serve as a predictive biomarker.  $\rm H_2S$  correlates positively with VEGF, FA, VB $_{\rm e}$ , and VB $_{\rm 12}$  levels, and negatively with Hcy, suggesting its pivotal role in regulating collateral circulation and neurological outcomes in stroke.

#### Acknowledgements

The author wishes to express her profound gratitude to all the study participants. Special thanks for Xuebin Li's research guidance. This study was supported by a grant from National Natural Science Foundation of China (No. 81860226); Guangxi Medical and Health Appropriate Technology Development and Application Project (No. S2018073); Young and Middle-aged Backbone Talent Scientific Research Projects of the Affiliated Hospital of Youjiang Medical University for Nationalities in 2021 (No. Y20212610); and Guangxi Health Commission Self-Financed Research Project (No. Z-L20230898).

#### Disclosure of conflict of interest

None.

Address correspondence to: Xuebin Li, Youjiang Medical University for Nationalities, No. 98 of Chengxiang Road, Baise 533000, Guangxi, China. Tel: +86-0776-2836742; E-mail: 00025@ymun.edu. cn; Jie Wang, Department of Nephrology, The Affiliated Hospital of Youjiang Medical University for Nationalities, No. 18 of Zhongshan Second Road, Baise 533000, Guangxi, China. Tel: +86-0776-2855284; E-mail: jie.wang@ymun.edu.cn

#### References

[1] Martin SS, Aday AW, Allen NB, Almarzooq ZI, Anderson CAM, Arora P, Avery CL, Baker-Smith CM, Bansal N, Beaton AZ, Commodore-Mensah Y, Currie ME, Elkind MSV, Fan W, Generoso G, Gibbs BB, Heard DG, Hiremath S, Johansen MC, Kazi DS, Ko D, Leppert MH, Magnani JW,

- Michos ED, Mussolino ME, Parikh NI, Perman SM, Rezk-Hanna M, Roth GA, Shah NS, Springer MV, St-Onge MP, Thacker EL, Urbut SM, Van Spall HGC, Voeks JH, Whelton SP, Wong ND, Wong SS, Yaffe K and Palaniappan LP; American Heart Association Council on Epidemiology, Prevention Statistics Committee and Stroke Statistics Committee. 2025 heart disease and stroke statistics: a report of US and global data from the American heart association. Circulation 2025; 151: e41-e660.
- [2] The Writing Committee Of The Report On Cardiovascular Health And Diseases In China; Hu SS. Report on cardiovascular health and diseases in China 2021: an updated summary. J Geriatr Cardiol 2023; 20: 399-430.
- [3] Renu Jornet A, Urra X, Laredo C, Montejo C, Rudilosso S, Llull L, Blasco J, Amaro S, Torne R, Obach V, Macho J and Chamorro A. Benefit from mechanical thrombectomy in acute ischemic stroke with fast and slow progression. J Neurointery Surg 2020; 12: 132-135.
- [4] Yoshimoto T, Inoue M, Tanaka K, Kanemaru K, Koge J, Shiozawa M, Kamogawa N, Kimura S, Chiba T, Satow T, Takahashi JC, Toyoda K, Koga M and Ihara M. Identifying large ischemic core volume ranges in acute stroke that can benefit from mechanical thrombectomy. J Neurointerv Surg 2021; 13: 1081-1087.
- [5] Shuaib A, Butcher K, Mohammad AA, Saqqur M and Liebeskind DS. Collateral blood vessels in acute ischaemic stroke: a potential therapeutic target. Lancet Neurol 2011; 10: 909-921.
- [6] Ginsberg MD. The cerebral collateral circulation: relevance to pathophysiology and treatment of stroke. Neuropharmacology 2018; 134: 280-292.
- [7] Liu Y, Tian X, Leung TW, Liu L, Liebeskind DS and Leng X. Good collaterals and better outcomes after EVT for basilar artery occlusion: a systematic review and meta-analysis. Int J Stroke 2023; 18: 917-926.
- [8] Ban M, Han X, Bao W, Zhang H and Zhang P. Evaluation of collateral status and outcome in patients with middle cerebral artery stenosis in late time window by CT perfusion imaging. Front Neurol 2022; 13: 991023.
- [9] Kwak HS and Park JS. Mechanical thrombectomy in basilar artery occlusion: clinical outcomes related to posterior circulation collateral score. Stroke 2020; 51: 2045-2050.
- [10] Sperti M, Arba F, Acerbi A, Busto G, Fainardi E and Sarti C. Determinants of cerebral collateral circulation in acute ischemic stroke due to large vessel occlusion. Front Neurol 2023; 14: 1181001.
- [11] Kabil O and Banerjee R. Enzymology of H2S biogenesis, decay and signaling. Antioxid Redox Signal 2014; 20: 770-782.

- [12] Kumar M, Modi M and Sandhir R. Hydrogen sulfide attenuates homocysteine-induced cognitive deficits and neurochemical alterations by improving endogenous hydrogen sulfide levels. Biofactors 2017; 43: 434-450.
- [13] Aschner M, Skalny AV, Ke T, da Rocha JB, Paoliello MM, Santamaria A, Bornhorst J, Rongzhu L, Svistunov AA, Djordevic AB and Tinkov AA. Hydrogen sulfide (H(2)S) signaling as a protective mechanism against endogenous and exogenous neurotoxicants. Curr Neuropharmacol 2022; 20: 1908-1924.
- [14] Lv L, Xi HP, Huang JC and Zhou XY. LncRNA SNHG1 alleviated apoptosis and inflammation during ischemic stroke by targeting miR-376a and modulating CBS/H(2)S pathway. Int J Neurosci 2021; 131: 1162-1172.
- [15] Tao L, Yu Q, Zhao P, Yang Q, Wang B, Yang Y, Kuai J and Ding Q. Preconditioning with hydrogen sulfide ameliorates cerebral ischemia/reperfusion injury in a mouse model of transient middle cerebral artery occlusion. Chem Biol Interact 2019; 310: 108738.
- [16] Jiang WW, Huang BS, Han Y, Deng LH and Wu LX. Sodium hydrosulfide attenuates cerebral ischemia/reperfusion injury by suppressing overactivated autophagy in rats. FEBS Open Bio 2017; 7: 1686-1695.
- [17] Chen Y, Chen S, Wu M, Chen F, Guan Q, Zhang S, Wen J, Sun Z and Chen Z. Hydrogen sulfide protects against rat ischemic brain injury by promoting rhoa phosphorylation at serine 188. ACS Omega 2024; 9: 13227-13238.
- [18] Zajkowska M, Lubowicka E, Fiedorowicz W, Szmitkowski M, Jamiolkowski J and Lawicki S. Human plasma levels of VEGF-A, VEGF-C, VEGF-D, their soluble receptor - VEGFR-2 and applicability of these parameters as tumor markers in the diagnostics of breast cancer. Pathol Oncol Res 2019; 25: 1477-1486.
- [19] Lu M, Liu Y, Xian Z, Yu X, Chen J, Tan S, Zhang P and Guo Y. VEGF to CITED2 ratio predicts the collateral circulation of acute ischemic stroke. Front Neurol 2022; 13: 1000992.
- [20] Zechariah A, ElAli A, Doeppner TR, Jin F, Hasan MR, Helfrich I, Mies G and Hermann DM. Vascular endothelial growth factor promotes pericyte coverage of brain capillaries, improves cerebral blood flow during subsequent focal cerebral ischemia, and preserves the metabolic penumbra. Stroke 2013; 44: 1690-1697.
- [21] Zhang S, Cheng Y, Guan Y, Wen J and Chen Z. Hydrogen sulfide exerted a pro-angiogenic role by promoting the phosphorylation of VEGFR2 at Tyr797 and Ser799 sites in hypoxia-reoxygenation injury. Int J Mol Sci 2024; 25: 4340.
- [22] Liu L, Chen W, Zhou H, Duan W, Li S, Huo X, Xu W, Huang L, Zheng H, Liu J, Liu H, Wei Y, Xu J and Wang Y; Chinese Stroke Association Stroke Council Guideline Writing Committee. Chinese

- stroke association guidelines for clinical management of cerebrovascular disorders: executive summary and 2019 update of clinical management of ischaemic cerebrovascular diseases. Stroke Vasc Neurol 2020; 5: 159-176.
- [23] Adams HP Jr, Bendixen BH, Kappelle LJ, Biller J, Love BB, Gordon DL and Marsh EE 3rd. Classification of subtype of acute ischemic stroke. Definitions for use in a multicenter clinical trial. TOAST. Trial of Org 10172 in acute stroke treatment. Stroke 1993; 24: 35-41.
- [24] Feng Y, Wu T, Wang T, Li Y, Li M, Li L, Yang B, Bai X, Zhang X, Wang Y, Gao P, Chen Y, Ma Y and Jiao L. Correlation between intracranial vertebral artery stenosis diameter measured by digital subtraction angiography and cross-sectional area measured by optical coherence tomography. J Neurointerv Surg 2021; 13: 1002-1006.
- [25] Sheth SA, Sanossian N, Hao Q, Starkman S, Ali LK, Kim D, Gonzalez NR, Tateshima S, Jahan R, Duckwiler GR, Saver JL, Vinuela F and Liebeskind DS; UCLA Collateral Investigators. Collateral flow as causative of good outcomes in endovascular stroke therapy. J Neurointerv Surg 2016; 8: 2-7.
- [26] Barthels D and Das H. Current advances in ischemic stroke research and therapies. Biochim Biophys Acta Mol Basis Dis 2020; 1866: 165260.
- [27] Binder NF, El Amki M, Gluck C, Middleham W, Reuss AM, Bertolo A, Thurner P, Deffieux T, Lambride C, Epp R, Handelsmann HL, Baumgartner P, Orset C, Bethge P, Kulcsar Z, Aguzzi A, Tanter M, Schmid F, Vivien D, Wyss MT, Luft A, Weller M, Weber B and Wegener S. Leptomeningeal collaterals regulate reperfusion in ischemic stroke and rescue the brain from futile recanalization. Neuron 2024; 112: 1456-1472, e6.
- [28] Liu L, Ding J, Leng X, Pu Y, Huang LA, Xu A, Wong KSL, Wang X and Wang Y. Guidelines for evaluation and management of cerebral collateral circulation in ischaemic stroke 2017. Stroke Vasc Neurol 2018; 3: 117-130.
- [29] Kim BM, Baek JH, Heo JH, Nam HS, Kim YD, Yoo J, Kim DJ, Jeon P, Baik SK, Suh SH, Lee KY, Kwak HS, Roh HG, Lee YJ, Kim SH, Ryu CW, Ihn YK, Kim B, Jeon HJ, Kim JW, Byun JS, Suh S, Park JJ, Lee WJ, Roh J, Shin BS and Bang OY. Collateral status affects the onset-to-reperfusion time window for good outcome. J Neurol Neurosurg Psychiatry 2018; 89: 903-909.
- [30] Topcu A, Ozkul A, Yilmaz A, Yi HJ, Shin DS and Kim B. The impact of collateral status on cerebral vasospasm and delayed cerebral ischemia in subarachnoid hemorrhage. J Cerebrovasc Endovasc Neurosurg 2023; 25: 288-296.

- [31] Consoli A, Pizzuto S, Sgreccia A, Di Maria F, Coskun O, Rodesch G, Lapergue B, Felblinger J, Chen B and Bracard S. Angiographic collateral venous phase: a novel landmark for leptomeningeal collaterals evaluation in acute ischemic stroke. J Neurointerv Surg 2023; 15: e323-e329.
- [32] Liebeskind DS, Tomsick TA, Foster LD, Yeatts SD, Carrozzella J, Demchuk AM, Jovin TG, Khatri P, von Kummer R, Sugg RM, Zaidat OO, Hussain SI, Goyal M, Menon BK, Al Ali F, Yan B, Palesch YY and Broderick JP; IMS III Investigators. Collaterals at angiography and outcomes in the interventional management of stroke (IMS) III trial. Stroke 2014; 45: 759-764.
- [33] Fukuda KA and Liebeskind DS. Evaluation of collateral circulation in patients with acute ischemic stroke. Radiol Clin North Am 2023; 61: 435-443.
- [34] Yang B, Ding Y, Liu X, Cai Y, Yang X, Lu Q, Gu W, Liu L and Pu Y. Matrix metallopeptidase 9 and placental growth factor may correlate with collateral status based on whole-brain perfusion combined with multiphase computed tomography angiography. Neurol Res 2021; 43: 838-845.
- [35] Lobov GI, Sokolova IB, Gorshkova OP, Shvetsova ME and Dvoretskii DP. Contribution of hydrogen sulfide to dilation of rat cerebral arteries after ischemia/reperfusion injury. Bull Exp Biol Med 2020; 168: 597-601.
- [36] Lv S, Wang Z, Wang J and Wang H. Exogenous hydrogen sulfide plays an important role through regulating autophagy in ischemia/reperfusion injury. Front Mol Biosci 2021; 8: 681676.
- [37] Qu K, Chen CP, Halliwell B, Moore PK and Wong PT. Hydrogen sulfide is a mediator of cerebral ischemic damage. Stroke 2006; 37: 889-893.
- [38] Saikia Q, Reeve H, Alzahrani A, Critchley WR, Zeqiraj E, Divan A, Harrison MA and Ponnambalam S. VEGFR endocytosis: Implications for angiogenesis. Prog Mol Biol Transl Sci 2023; 194: 109-139.
- [39] Moon S, Chang MS, Koh SH and Choi YK. Repair mechanisms of the neurovascular unit after ischemic stroke with a focus on VEGF. Int J Mol Sci 2021; 22: 8543.
- [40] Liu J, Tian L and Li N. Treatment efficacy of arterial urokinase thrombolysis combined with mechanical thrombectomy for acute cerebral infarction and its influence on neuroprotective factors and factors for neurological injury. Am J Transl Res 2021; 13: 3380-3389.
- [41] Li P, Lu H, Shi X, Yan J, Zhou L, Yang J, Wang B, Zhao Y, Liu L, Zhu Y, Xu L, Yang X, Su X, Yang Y, Zhang T, Guo L and Liu X. Protective effects of human urinary kallidinogenase against corti-

#### H<sub>2</sub>S & AIS collateral circulation

- cospinal tract damage in acute ischemic stroke patients. Neuroreport 2024; 35: 431-438.
- [42] Wang MJ, Cai WJ, Li N, Ding YJ, Chen Y and Zhu YC. The hydrogen sulfide donor NaHS promotes angiogenesis in a rat model of hind limb ischemia. Antioxid Redox Signal 2010; 12: 1065-1077.
- [43] Tao BB, Liu SY, Zhang CC, Fu W, Cai WJ, Wang Y, Shen Q, Wang MJ, Chen Y, Zhang LJ, Zhu YZ and Zhu YC. VEGFR2 functions as an H2S-targeting receptor protein kinase with its novel Cys1045-Cys1024 disulfide bond serving as a specific molecular switch for hydrogen sulfide actions in vascular endothelial cells. Antioxid Redox Signal 2013; 19: 448-464.
- [44] Kaplan P, Tatarkova Z, Sivonova MK, Racay P and Lehotsky J. Homocysteine and mitochondria in cardiovascular and cerebrovascular systems. Int J Mol Sci 2020; 21: 7698.
- [45] Santilli F, Davi G and Patrono C. Homocysteine, methylenetetrahydrofolate reductase, folate status and atherothrombosis: a mechanistic and clinical perspective. Vascul Pharmacol 2016; 78: 1-9.

- [46] Modi M, Prabhakar S, Majumdar S, Khullar M, Lal V and Das CP. Hyperhomocysteinemia as a risk factor for ischemic stroke: an Indian scenario. Neurol India 2005; 53: 297-301; discussion 301-292.
- [47] Hou HM, Qin XJ and Zhao HY. MTHFR C677T polymorphism, homocysteine, burden, and location of AMI and ACI. Eur Rev Med Pharmacol Sci 2023; 27: 1427-1435.
- [48] Olaso-Gonzalez G, Inzitari M, Bellelli G, Morandi A, Barcons N and Vina J. Impact of supplementation with vitamins B(6), B(12), and/or folic acid on the reduction of homocysteine levels in patients with mild cognitive impairment: a systematic review. IUBMB Life 2022; 74: 74-84.
- [49] Lu W and Wen J. Role and relationship between homocysteine and H(2)S in ischemic stroke. Mol Neurobiol 2025; [Epub ahead of print].