Original Article Decreased expression of stromal estrogen receptor α and β in prostate cancer

Garrett Daniels^{1*}, Lan Lin Gellert^{1*}, Jonathan Melamed¹, David Hatcher¹, Yirong Li¹, Jianjun Wei¹, Jinhua Wang³, Peng Lee^{1,2,3,4}

Departments of ¹Pathology, ²Urology, ³NYU Cancer Institute, ⁴New York Harbor Healthcare System, New York University School of Medicine, New York, NY, USA. *Equal contributors.

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Abstract: Background: Recently there has been an increased interest in the role of tumor-associated stroma in prostate tumorigenesis, but little is known about the respective roles of stomal ERa and ERB in prostate cancer (PCa). This study characterizes the expression patterns of ER α and ER β in tumor-associated stroma in association with various clinicopathological factors of importance in PCa prognosis and treatment. Design: Immunohistochemistry was performed using antibodies against $ER\alpha$ and $ER\beta$ to characterize their expression patterns in PCa tissue. Stromal ER levels (ER α and ER β) on tissue sections (n=47), were compared between tumor associated stroma and adjacent benign associated stroma. Immunohistochemistry was also performed on a PCa tissue microarray (TMA) (n=177) to correlate stromal expression with various clinicopathological parameters. The levels of ER nuclear expression were scored semi-quantitatively. Results: The expression levels of both ERα and ERβ were significantly lower in tumor-associated stroma than stroma surrounding benign prostatic glands on the same tissue section (ERa: p<0.01; ERB: p=0.01). When correlated with clinicopathological factors, the level of ERa expression in tumorassociated stroma showed a positive correlation with Gleason score ($R^2=0.8638$). The expression of ER α was higher in PCa with advanced tumor stage (p=0.05) and not significantly different in extraprostatic extension (p>0.05). The level of ERB expression in tumor-associated stroma was decreased in patients older than 60 years compared to younger patients (p=0.01). Conclusion: This study demonstrates significant down-regulation of ER α and ER β expression in the tumor-associated stroma of PCa. However, the level of ERα expression in tumor-associated stroma shows a positive correlation with cancer differentiation and tumor stage.

Keywords: Estrogen receptors, prostate cancer, stromal

Introduction

Prostate cancer (PCa) is the most common cancer among men, as well as one of the leading causes of cancer related deaths [1]. Curative therapy is a challenge because late stage disease often evolves to a state that is refractory to therapy, developing the androgen-independent disease [2]. A better understanding of PCa progression is crucial to the discovery of reliable prognostic markers and novel therapies for the disease.

Recently, there has been an increased interest in role of tumor-associated stroma in neoplastic progression, including in PCa. Stromal cells in the surrounding matrix play an important role in cancer, communicating with nearby tumor cells via either direct or paracrine signaling [3-6]. The introduction of tumor-associated stroma to normal epithelial cells has been shown to cause alterations of the epithelium leading to hyperplasia and tumorigenesis [7]. Though non-cancerous themselves, these stromal cells often show different patterns of expression for specific factors as compared to stromal cells distant from cancer [8]. A recent study has been performed to identify key factors specific to PCa tumor-associated stroma, including CAV1 a predictor of early PCa tumor recurrence [9]. Importantly, some of these stromal factors may be potential predictive and prognostic markers of cancer.

Nuclear hormone receptors, including androgen receptor (AR), ER and progesterone receptor (PR), have previously been reported to be

tion on TMA		
Characteristic	No. of Patients	%
PSA Recurrence Status		
PSA Recurrence	43	21.5
Post-Prostatectomy Residual Tumor	48	24
None	60	30
Unknown	49	24.5
Vital Status		
Alive	157	78.5
Dead	36	18
Lost to Follow-Up	7	3.5
Extraprostatic Extension		
Focal	47	23.5
Established	20	10
Multifocal	23	11.5
None	109	54.5
Unknown	1	0.5
pT Stage		
pT1b	1	0.5
pT2	1	0.5
pT2+	4	2
pT2a	23	11.5
pT2b	77	38.5
рТЗ	1	0.5
рТЗа	67	33.5
pT3b	26	13
pN Stage		
pNO	166	83
pN1	2	1
pNX	32	16
pM Stage		
рМО	178	89
pMX	22	11

 Table 1. Clinical characteristics of the study population on TMA

important modulators of prostate growth and differentiation. Once activated by their respective hormones, they regulate gene expression, modifying differentiation and proliferation pathways [10-13]. ER, activated by estrogen, has activity as both a DNA binding transcription factor regulating gene expression as well as nongenomic functions, including membrane signaling leading to post-translational modifications of many existing proteins [14]. ER is expressed in two separate forms, α and β , that are regulated separately and have different expression patterns in PCa [15-19]. ER plays a role in cell proliferation in the prostate, both as a stimulatory factor and a growth inhibitor, via activation of its two separate isoforms [20, 21]. ERß may have anti-proliferative effects to counter the proliferative effects of ER α [22, 23]. One study found that the inability to activate ER β by estrogen in tissue recombinant mice leads to prostate hyperplasia, which can be resolved by an ER β specific agonist [24].

The roles of androgen and estrogen receptors in PCa have been primarily focused on epithelial cells [22, 25], while their roles in stromal cells (i.e. effects on prostate tumorigenesis and cancer progression) have been far less studied. ER α and β are both expressed in PCa associated stromal cells; however, prior to this study, no correlation between stromal ER expression and clinicopathological factors of PCa have been reported. We previously reported an association between decreased stromal AR and PCa differentiation and androgen-independence [26]. Additionally, PR expression is reduced in tumor associated stroma when compared to benign associated stroma [27]. The purpose of this study was to evaluate the expression of both isoforms of ER in tumor-associated stroma compared to benign stroma to determine any link between stromal ER expression and clinicopathological factors of disease.

Material and methods

Case selection and TMA construction

Two sets of PCa cases were used in this study. In the first study set, the tissue sections from forty-seven PCa cases were

studied to compare stromal ER (ER α and ER β) levels in PCa and adjacent benign prostate on the same slide. Gleason score ranges from 5 to 10, stages from T2 to T3b and PSA ranges from 0.05 to 50 (including both recurrent and non-recurrent tumor) were used. In the second study set, ER α and ER β expression patterns were studied in 177 samples of PCa on a tissue microarray (TMA) (177 out of 200 with IHC scores available). The samples on the TMA were stratified by various clinicopathological factors including age at diagnosis, preoperative PSA, grade, and stage (Table 1).

Immunohistochemistry (IHC)

Archival formalin-fixed, paraffin-embedded tissue sections and TMAs of PCa cases were used



Figure 1. The expression of ER α is significantly lower in tumor-associated stroma (B) compared with stroma around benign glands (A) (p<0.01). Upper inset: ER α in breast cancer as positive control. Lower inset: Negative control for ER α in prostate.



Figure 2. The expression of ER β is significantly lower in tumor-associated stroma (B) compared with stroma around benign glands (A) (p=0.01). Upper inset: ER β in breast cancer as positive control. Lower inset: Negative control for ER β in prostate.



Figure 3. The level of ER α expression in tumor-associated stroma shows a positive correlation with Gleason score (r=0.93, p=0.05). No significant correlation was detected between the expression of ER β in tumor-associated stroma and Gleason score.

in this study with NYU IRB approval. The IHC was performed using single label immunohistochemistry by the NexES automated immunostainer and detection system (Ventana Medical Systems, Tucson, AZ, USA). 4 micron sections were deparaffinized in xylene, rehydrated through graded alcohols, and rinsed in distilled water. All incubations were carried out at 37°C unless otherwise noted. After deparaffinization, heat induced epitope retrieval was performed by



Figure 4. The expression level of ER α is higher in PCa with advanced tumor stage (n (pT2): 108, n (pT3): 86, p=0.05). No significant correlation was detected between the expression of ER β in tumor-associated stroma and tumor stage.



Figure 5. The level of ER β expression in tumor-associated stroma is decreased in older patients (p=0.01). No significant correlation was detected between the expression of ER α in tumor-associated stroma and age.

microwaving sections with 0.01 M, pH 6.0 citrate buffer for 20 min in a 1200 watt microwave oven. Endogenous peroxidase was blocked by application of hydrogen peroxide for 4 minutes. ER α and ER β specific antibodies (Cell Signaling) were detected by the application of a biotinylated goat anti-rabbit for 8 minutes, followed by the application of streptavidin-horseradish peroxidase for 8 minutes. The chromogen, 3,3'-diaminobenzidine/hydrogen peroxide mix was applied for 8 minutes and then enhanced with copper sulfate for 4 minutes. Slides were then counterstained with

hematoxylin, dehydrated, and mounted in permanent media. The levels of ER nuclear expression were scored semi-quantitatively: 0 as negative, 1 as weak, 2 as moderate, and 3 as strong expression. Breast cancer cases were used as positive control for ER α and ER β expression. No primary antibody controls were used to serve as a negative control.

Statistical analysis

Statistical analyses of the above results were performed by Student's *t*-test and linear regression and correlation analysis. Differences were considered statistically significant if $p \le 0.05$.

Results

Characteristics of study population

Analyses of the correlation of stromal ER α and ER β expression with clinicopathological factors were performed on 177 samples of PCa and compared with race, age at diagnosis, Gleason score, grade and stage, extraprostatic extension, preoperative PSA, and

PSA recurrence. The demographic and clinicopathological data from the 177 patients on the TMA are summarized in **Table 1**.

The expression levels of both ER α and ER β are significantly lower in tumor-associated stroma

First, we compared the expression levels of ER α and ER β between tumor-associated stroma and the stroma surrounding benign prostatic glands. The expression levels of ER in the stroma were scored based on the intensity of immunostaining and the percentage of stromal fibroblasts and myofibroblasts adjacent to

either the benign or malignant prostatic glands. The staining patterns of the infiltrating lymphocytes, macrophages, and other cells in the stromal compartment were not taken into account. On the whole tissue section of 47 PCa cases, ER α and ER β are expressed in 40% of stromal cells in benign prostate tissue. The expression levels of both ER α and ER β were significantly lower in tumor-associated stroma (ER α : p<0.01; ER β : p=0.01) (**Figures 1** and **2**).

Increased expression of stromal ER $\!\alpha$ is associated with advanced disease

Since the expression of stromal ER α and ER β was decreased in PCa in general, we performed a correlation analysis between stromal ER expression and various clinicopathological factors of PCa (including race, age at diagnosis, Gleason score, grade and stage, extraprostatic extension, preoperative PSA, and status of PSA recurrence). The expression of ERa in tumorassociated stroma showed a positive correlation with the Gleason score (r=0.93, p=0.05) (Figure 3) and tumor stage (p=0.05) (Figure 4). In addition, the cases with extraprostatic extension showed a trend of higher stromal expression of ERa, but the difference was not statistically significant (p>0.05). No significant correlation was detected between the expression levels of ER α and the other clinicopathological factors. The level of ERß expression in tumorassociated stroma was significantly decreased in patients older than 60 years compared to younger patients (p=0.01) (Figure 5). No significant correlation was detected between the expression level of ERß in tumor-associated stroma and any of the other clinicopathological factors.

Discussion

The stromal environment in the prostate is heterogeneous in nature, consisting of macrophages, lymphocytes, smooth muscle, and fibroblasts in benign tissue, while myofibroblasts predominate in the tumor-associated stroma of cancer [28]. The prostate stroma is also heterogeneous in its expression of steroid hormone receptors (including AR, ER and PR). Low levels of stromal AR have been correlated with more aggressive cancer and loss of stromal AR facilitates growth in vitro and in vivo and invasion [26, 29]. Decreased PR expression has also been observed in PCa associated stromal cells [27]. Estrogen and ER have also been shown to have carcinogenic effects on the prostate [30, 31]. Both ER α and ER β show proliferative and anti proliferative properties in the prostate in mouse models [24, 32]. These models shed light on ER α and β -specific effects that might otherwise be masked, such as by the indirect effects of estrogen. However, little is known about the expression and function of stromal ER and how it affects the neighboring epithelium. In benign prostate, ERB is the primary isoform expressed in the epithelium, whereas $ER\alpha$ is primarily found in the stromal and basal cells. However, $ER\alpha$ levels increase in epithelial cells in cancer [17]. More recently, it has been reported that TMPRSS2-ERG, a commonly identified fusion protein under the regulation by androgen in PCa, is regulated by estrogen signaling in a subclass of PCa [22]. ER α is also significantly lower in the stroma of Caucasian and African American men, who are at higher risk for PCa, compared to Hispanic and Asian men [33]. In this study, we showed that the expression of stromal ER is distinct from that found in epithelial cells. There is a significant reduction in both ERα and ERβ in tumor-associated stroma compared with adjacent benign prostate. Thus, down-regulation of stromal ER may be a critical step in the early stages of tumorigenesis. Although both isoforms are down-regulated in cancer compared to benign prostate, higher levels of ER α was observed in high grade and advanced stage tumors in tumor-associated stroma compared to low grade and stage cancer. This data was consistent with a recent report showing a trend of higher stromal ERa expression with high grade PCa [34]. This data is in direct contrast to stromal AR expression patterns where the AR levels are decreased in relation with grade [26]. Thus, an eventual rise of ERa levels may actually be critical in the transition into more advanced disease, or conversely, this could represent a passive change in advanced cancers that no longer require down-regulation of stromal ERa for tumor growth. It would be of great interest to determine whether the same relationship is found in metastatic and androgen-independent PCa.

ER β levels were found to be markedly reduced in benign prostate stroma in older patients (>60), though ER β levels did not show the same correlation with Gleason score and advanced tumor stage in this study. This observation may be of significance for the development and progression of PCa. It is reported that low stromal AR levels in normal and tumor prostate tissue are related to poor outcome in PCa patients [29]. ER β may have similar effects. ER β -specific agonists have shown promising results in PCa treatment [23]. Taken together, these observations suggest that stromal ER β may have anti proliferative effects in the prostate as well as an inhibitory role in the progression to aggressive cancers.

We previously showed that stromal AR inhibits proliferation and invasion of malignant prostate epithelial cells. On the other hand, loss of AR in the stroma facilitated growth and invasion both in vitro and in vivo [26, 29]. In light of a similar stromal ER expression pattern, ER may also have inhibitory effects in the prostate. It would be of great interest to assess the direct effects of stromal ER (ER α and ER β) on prostate growth in future studies.

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Disclosure of conflict of interest

None.

Address correspondence to: Dr. Peng Lee, Department of Pathology and Urology, New York University School of Medicine, 423 E. 23rd Street, Room 6140N, New York, NY 10010. E-mail: peng. lee@nyumc.org

References

- Jemal A, Siegel R, Ward E, Murray T, Xu J and Thun MJ. Cancer statistics, 2007. CA Cancer J Clin 2007; 57: 43-66.
- [2] Freedland SJ, Isaacs WB, Platz EA, Terris MK, Aronson WJ, Amling CL, Presti JC Jr and Kane CJ. Prostate size and risk of high-grade, advanced prostate cancer and biochemical progression after radical prostatectomy: a search database study. J Clin Oncol 2005; 23: 7546-7554.

- [3] Cunha GR, Hayward SW, Wang YZ and Ricke WA. Role of the stromal microenvironment in carcinogenesis of the prostate. Int J Cancer 2003; 107: 1-10.
- [4] Joesting MS, Cheever TR, Volzing KG, Yamaguchi TP, Wolf V, Naf D, Rubin JS and Marker PC. Secreted frizzled related protein 1 is a paracrine modulator of epithelial branching morphogenesis, proliferation, and secretory gene expression in the prostate. Dev Biol 2008; 317: 161-173.
- [5] Karagiannis GS, Poutahidis T, Erdman SE, Kirsch R, Riddell RH and Diamandis EP. Cancer-associated fibroblasts drive the progression of metastasis through both paracrine and mechanical pressure on cancer tissue. Mol Cancer Res 2012; 10: 1403-1418.
- [6] Cunha GR. Mesenchymal-epithelial interactions: past, present, and future. Differentiation 2008; 76: 578-586.
- [7] Olumi AF, Grossfeld GD, Hayward SW, Carroll PR, Tlsty TD and Cunha GR. Carcinoma-associated fibroblasts direct tumor progression of initiated human prostatic epithelium. Cancer Res 1999; 59: 5002-5011.
- [8] Dakhova O, Ozen M, Creighton CJ, Li R, Ayala G, Rowley D and Ittmann M. Global gene expression analysis of reactive stroma in prostate cancer. Clin Cancer Res 2009; 15: 3979-3989.
- [9] Orr B, Riddick AC, Stewart GD, Anderson RA, Franco OE, Hayward SW and Thomson AA. Identification of stromally expressed molecules in the prostate by tag-profiling of cancerassociated fibroblasts, normal fibroblasts and fetal prostate. Oncogene 2012; 31: 1130-1142.
- [10] Linja MJ and Visakorpi T. Alterations of androgen receptor in prostate cancer. J Steroid Biochem Mol Biol 2004; 92: 255-264.
- [11] Taplin ME and Balk SP. Androgen receptor: a key molecule in the progression of prostate cancer to hormone independence. J Cell Biochem 2004; 91: 483-490.
- [12] Ho SM. Estrogens and anti-estrogens: key mediators of prostate carcinogenesis and new therapeutic candidates. J Cell Biochem 2004; 91: 491-503.
- [13] Bonkhoff H, Fixemer T, Hunsicker I and Remberger K. Progesterone receptor expression in human prostate cancer: correlation with tumor progression. Prostate 2001; 48: 285-291.
- [14] Levin ER. Integration of the extranuclear and nuclear actions of estrogen. Mol Endocrinol 2005; 19: 1951-1959.
- [15] Fixemer T, Remberger K and Bonkhoff H. Differential expression of the estrogen receptor beta (ERbeta) in human prostate tissue, premalignant changes, and in primary, metastat-

ic, and recurrent prostatic adenocarcinoma. Prostate 2003; 54: 79-87.

- [16] Bonkhoff H, Fixemer T, Hunsicker I and Remberger K. Estrogen receptor gene expression and its relation to the estrogen-inducible HSP27 heat shock protein in hormone refractory prostate cancer. Prostate 2000; 45: 36-41.
- [17] Bonkhoff H, Fixemer T, Hunsicker I and Remberger K. Estrogen receptor expression in prostate cancer and premalignant prostatic lesions. Am J Pathol 1999; 155: 641-647.
- [18] Prins GS and Korach KS. The role of estrogens and estrogen receptors in normal prostate growth and disease. Steroids 2008; 73: 233-244.
- [19] Ricke WA, Wang Y and Cunha GR. Steroid hormones and carcinogenesis of the prostate: the role of estrogens. Differentiation 2007; 75: 871-882.
- [20] Cunha GR, Wang YZ, Hayward SW and Risbridger GP. Estrogenic effects on prostatic differentiation and carcinogenesis. Reprod Fertil Dev 2001; 13: 285-296.
- [21] Bonkhoff H and Berges R. The evolving role of oestrogens and their receptors in the development and progression of prostate cancer. Eur Urol 2009; 55: 533-542.
- [22] Setlur SR, Mertz KD, Hoshida Y, Demichelis F, Lupien M, Perner S, Sboner A, Pawitan Y, Andren O, Johnson LA, Tang J, Adami HO, Calza S, Chinnaiyan AM, Rhodes D, Tomlins S, Fall K, Mucci LA, Kantoff PW, Stampfer MJ, Andersson SO, Varenhorst E, Johansson JE, Brown M, Golub TR and Rubin MA. Estrogen-dependent signaling in a molecularly distinct subclass of aggressive prostate cancer. J Natl Cancer Inst 2008; 100: 815-825.
- [23] Ellem SJ and Risbridger GP. The dual, opposing roles of estrogen in the prostate. Ann N Y Acad Sci 2009; 1155: 174-186.
- [24] McPherson SJ, Ellem SJ, Simpson ER, Patchev V, Fritzemeier KH and Risbridger GP. Essential role for estrogen receptor beta in stromal-epithelial regulation of prostatic hyperplasia. Endocrinology 2007; 148: 566-574.
- [25] Horvath LG, Henshall SM, Lee CS, Head DR, Quinn DI, Makela S, Delprado W, Golovsky D, Brenner PC, O'Neill G, Kooner R, Stricker PD, Grygiel JJ, Gustafsson JA and Sutherland RL. Frequent loss of estrogen receptor-beta expression in prostate cancer. Cancer Res 2001; 61: 5331-5335.

- [26] Li Y, Li CX, Ye H, Chen F, Melamed J, Peng Y, Liu J, Wang Z, Tsou HC, Wei J, Walden P, Garabedian MJ and Lee P. Decrease in stromal androgen receptor associates with androgen-independent disease and promotes prostate cancer cell proliferation and invasion. J Cell Mol Med 2008; 12: 2790-2798.
- [27] Yu Y, Liu L, Xie N, Xue H, Fazli L, Buttyan R, Wang Y, Gleave M and Dong X. Expression and function of the progesterone receptor in human prostate stroma provide novel insights to cell proliferation control. J Clin Endocrinol Metab 2013; 98: 2887-2896.
- [28] Tuxhorn JA, Ayala GE and Rowley DR. Reactive stroma in prostate cancer progression. J Urol 2001; 166: 2472-2483.
- [29] Wikstrom P, Marusic J, Stattin P and Bergh A. Low stroma androgen receptor level in normal and tumor prostate tissue is related to poor outcome in prostate cancer patients. Prostate 2009; 69: 799-809.
- [30] Santti R, Newbold RR, Makela S, Pylkkanen L and McLachlan JA. Developmental estrogenization and prostatic neoplasia. Prostate 1994; 24: 67-78.
- [31] Singh PB, Matanhelia SS and Martin FL. A potential paradox in prostate adenocarcinoma progression: oestrogen as the initiating driver. Eur J Cancer 2008; 44: 928-936.
- [32] Prins GS, Birch L, Couse JF, Choi I, Katzenellenbogen B and Korach KS. Estrogen imprinting of the developing prostate gland is mediated through stromal estrogen receptor alpha: studies with alphaERKO and betaERKO mice. Cancer Res 2001; 61: 6089-6097.
- [33] Haqq C, Li R, Khodabakhsh D, Frolov A, Ginzinger D, Thompson T, Wheeler T, Carroll P and Ayala G. Ethnic and racial differences in prostate stromal estrogen receptor alpha. Prostate 2005; 65: 101-109.
- [34] Hetzl AC, Montico F, Lorencini RM, Kido L, Candido E, Billis A, Ferreira U and Cagnon VH. Fibroblast growth factor, estrogen, and prolactin receptor features in different grades of prostatic adenocarcinoma in elderly men. Microsc Res Tech 2013; 76: 321-330.