

## Original Article

# ODAM is a predictor for biomedical recurrence and inhibits the migration and invasion of prostate cancer

Yun Luo<sup>1\*</sup>, Jie-Ying Wu<sup>1\*</sup>, Guo-Liang Hou<sup>3</sup>, Min-Hua Lu<sup>1</sup>, Zhi Shi<sup>2</sup>, Jin-Ming Di<sup>1</sup>

<sup>1</sup>Department of Urology, The 3rd Affiliated Hospital of Sun Yat-Sen University, Guangzhou, Guangdong, China; <sup>2</sup>Department of Cell Biology & Institute of Biomedicine, National Engineering Research Center of Genetic Medicine, Guangdong Provincial Key Laboratory of Bioengineering Medicine, College of Life Science and Technology, Jinan University, Guangzhou, Guangdong, China; <sup>3</sup>Department of Urology, Foshan First Municipal People's Hospital, Foshan 528000, China. \*Equal contributors.

Received December 7, 2015; Accepted January 4, 2016; Epub February 15, 2016; Published February 29, 2016

**Abstract:** Odontogenic ameloblast associated protein (ODAM) is a protein contributed to cell adhesion and has been shown to express in normal prostate tissue, but the expression and significance of ODA in prostate cancer remain unknown. In this study, we detected the protein expressions of ODA in 88 prostate cancer tissues with immunohistochemical staining, and found that 53 cases (60.2%) was high expression of ODA, which was shown in the cytoplasm and paranuclear regions. Furthermore, low expression of ODA was significantly correlated with lymph node metastasis, preoperative PSA and Gleason score, but not with mean age, follow-up duration, PSM rate and distribution of pathological T stage. Additionally, our results of multivariate analysis showed that low ODA expression was an independent predictor of biomedical recurrence, while the positive lymph node metastasis, Gleason score, and preoperative PSA were not the independent risks for biomedical recurrence. Overexpression of ODA did not inhibit the growth of prostate cancer cells PC3, but significant suppressed their invasion and migration with decrease of the protein levels of MMP-2. These results suggest that ODA is a predictor for biomedical recurrence and inhibits the migration and invasion of prostate cancer.

**Keywords:** ODA, migration, invasion, prostate cancer

## Introduction

Prostate cancer is one of the most common male tumors in the worldwide, and invasion and metastasis of prostate cancer is the main reason for death [1-3]. However, the mechanisms of invasion and metastasis of prostate cancer are complicated and remain to be further understood. In our previous study, we analyzed TLR9 signaling network in regulation of migration and invasion by whole-genome microarray and found that odontogenic ameloblast-associated protein (ODAM) might be associated with migration and invasion in prostate cancer cells [4]. ODA is first identified as the protein constituent of calcifying epithelial odontogenic/Pindborg tumors and subsequent studies revealed that it was expressed in a wide range of normal odontogenic, glandular, and epithelial tissues [5-10] as well as in malignancies including odontogenic cancer, breast cancer, gastric cancer, and melanoma [11, 12]. The

previous study have reported that ODA was highly expressed in normal prostate tissue [13]. However, the expression and significance of ODA in prostate cancer remain unknown.

In this study, we determined the expression of ODA in the tissues of patients with prostate cancer and analyzed the correlation of ODA expression with lymph node metastasis, biomedical recurrence, Gleason score, PSM, preoperative PSA, pathological T stage and Age. Furthermore, we overexpressed ODA in human prostate cancer cells to gain further insight into the function of ODA in regulating migration and invasion of prostate cancer.

## Material and methods

### Patients and specimens

A total of 88 formalin-fixed, paraffin-embedded specimens were collected from prostate cancer

## ODAM inhibits the migration and invasion of prostate cancer

patients who underwent radical prostatectomy without neoadjuvant hormonal therapy or transurethral resection of prostate at the Department of Urology, 3rd Affiliated Hospital of Sun Yat-Sen University, Guangzhou, China from July 2007 to June 2014. Patients with histologically confirmed prostate cancer with diagnostic prostate biopsies and no metastatic disease diagnosed by pelvic computed tomography and bone scan were eligible. The radical prostatectomy was performed with standard pelvic lymph node dissection as previously described [14, 15]. The clinical characteristics, preoperative serum PSA level, surgical margin state, the state of lymph node metastasis, pathological T stage, and Gleason score were obtained from patient records. The clinical follow-up data include biochemical recurrence (defined as PSA  $\geq$  0.2 ng/ml on 2 successive measurements in 3 months after radical prostatectomy), clinical recurrence (defined as identification of metastases or histologically confirmed local recurrence) and tumor specific death. Progression-free survival rates were calculated from the date of radical prostatectomy to either biochemical recurrence leading to second line treatments, clinical recurrence or the last day of follow-up. The study was conducted with the ethical approval of the hospital human ethics committee.

### *Immunohistochemical staining*

Immunohistochemistry assay was performed using two micrometer thick formalin-fixed paraffin embedded archived tissue sections as previously described [16, 17]. Anti-ODAM antibodies were purchased from Proteintech Group, Inc., Chicago, USA. Staining was assessed by two independent investigators in a blind manner to reach a consensus. Staining intensity was recorded as negative (0), weakly positive (+1), positive (+2), and strongly positive (+3). Samples with +2 and +3 staining of ODA were classified as 'high expression group', and those with 0 and +1 were assigned as 'low expression group'.

### *Cell culture and transfection*

The human prostate cancer cell lines PC3 were obtained from the American Type Culture Collection and maintained in DMEM/F12 supplemented with 10% fetal bovine serum (FBS, Invitrogen), penicillin (100 units/mL), and strep-

tomycin (100  $\mu$ g/mL) at 37°C under 5% CO<sub>2</sub> in a humidified incubator. PC3 cells were transfected with pcDNA5T/O (Invitrogen) plasmid containing full-length human ODA cDNA or the empty vector using Lipofectamine LTX reagent (Invitrogen, Carlsbad, CA) according to the manufacturer's protocol. Cell clones with stable ODA overexpression were selected with hygromycin. ODA overexpression and control cell clones were designated PC3-ODA or PC3-CON, respectively.

### *Matrigel invasion assay*

Matrigel invasion assay was performed as previously described [18]. Briefly, PC3-ODA or PC3-CON cells were trypsinized, washed and suspended ( $1 \times 10^5$  cells/ml) in serum-free DMEM/F12 medium. One hundred  $\mu$ l cell suspension was added into the upper chamber of a Costar Transwell permeable support (8- $\mu$ m pore size, Thermo-Fisher) coated with Matrigel at 5 mg/ml, and the lower chamber was filled with medium containing 10% FBS. After incubation for 24 h at 37°C, non-migrating cells were swabbed from the upper surface and those that passed through to the lower surface were fixed in 90% ethanol for 10 min, stained with 0.1% crystal violet, counted and photographed.

### *Western blotting*

Western blotting was carried out according to previously reported [19, 20]. In brief, cells were harvested, lysed and centrifuged. Supernatants were collected, and protein concentration was quantified using Bradford assay. Proteins (30  $\mu$ g) were separated by 10% SDS-polyacrylamide gel electrophoresis and transferred to nitrocellulose membranes. Membranes were blocked and incubated with primary antibodies. Corresponding horseradish peroxidase-conjugated secondary antibodies were used against the primary antibody. GAPDH was used as an internal control (anti-GAPDH, 1:1000; Cell Signaling Technology, Inc., Beverly, MA, USA). Immunoreactive proteins were then visualized using the ECL western blotting system (Pierce Biotechnology, Rockford, IL, USA).

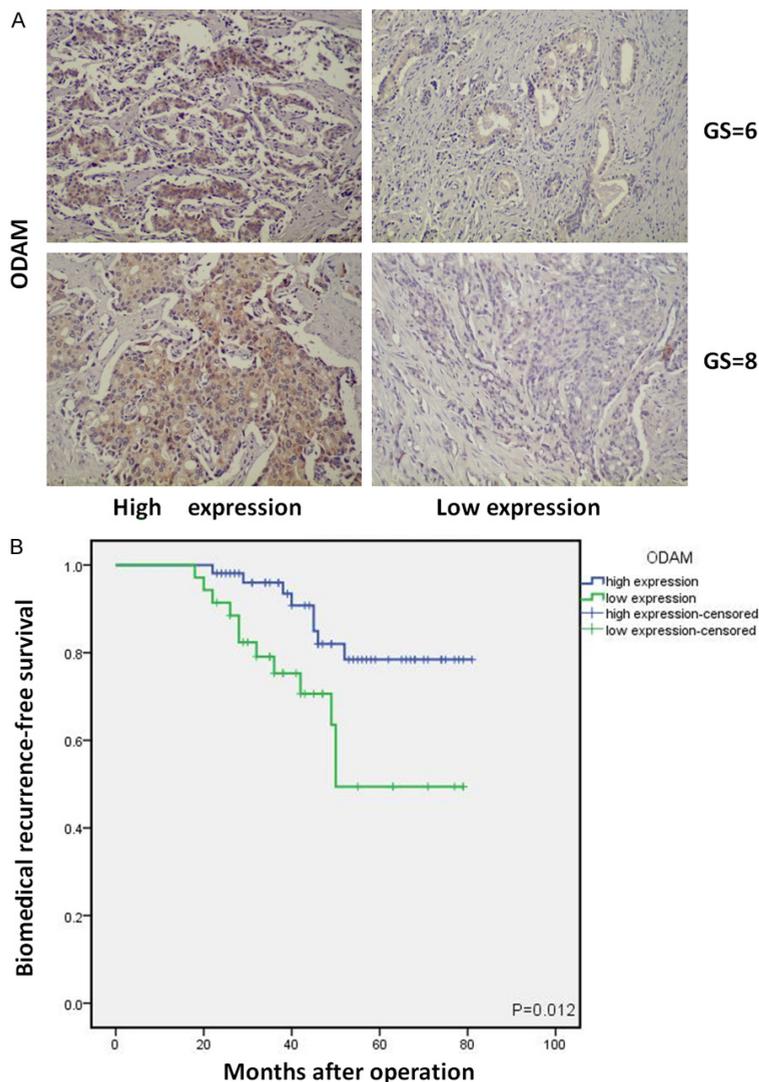
### *Wound healing assay*

Wound Healing Assay was performed as previously reported [21, 22]. PC3-ODA and PC3-

## ODAM inhibits the migration and invasion of prostate cancer

**Table 1.** Summary of clinical characteristics in 88 patients with differential expression of ODAM

Characteristics	ODAM expression			P value
	Total	Low expression (N=35)	High expression (N=52)	
Age	58.9±6.5	58.4±6.9	59.2±6.4	0.597
Preoperative PSA	15.3±6.3	20.2±5.2	12.1±4.7	<0.001
Gleason score	6.67±1.72	7.4±1.4	6.17±1.76	0.001
Follow up (M)	51.6±17.1	51.1±17.8	52.0±16.8	0.808



**Figure 1.** ODAM is an independent risk factor for biochemical recurrence. The expression of ODAM in human prostate cancer tissues were examined by immunohistochemistry staining. The representative samples (GS=6 and GS=8) with low and high expression of ODAM (A) and progression-free survival rate of patients with prostate cancer (B) were shown.

CON cells were seeded in the six-well plate until the cells reach a confluence of 80%. The

cell monolayer was scratched using a sterile 10  $\mu$ l pipette tip, and the detached cells were washed with PBS three times. Cells were allowed to migrate for 24 h and 48 h, and the scratches were carefully observed and photographed. The experiments were repeated 3 times independently.

### Statistical analysis

All quantitative data are presented as the mean  $\pm$  SD. Student's t test (two-tailed) was used to compare two groups. Correlation of ODAM expression and clinicopathological variables was analyzed using the  $\chi^2$  test or, in the case of low expected frequencies, by the Fisher's exact test. Correlation analysis was performed with spearman correlation analysis. Biomedical recurrence-free survival was analyzed using Kaplan-Meier method with the log-rank test for significance. Multivariate survival analysis was carried out using the Cox proportional hazards model. Hazard ratios (HRs) were assessed using Cox univariate analysis.  $P < 0.05$  was considered statistically significant.

### Results

*Low expression of ODAM is correlation with a higher probability of lymph node metastasis, preoperative PSA and Gleason score*

To investigate the expression of ODAM in human prostate cancer tissues, a total of 88 prostate cancer specimens were collected to detect their ODAM expressions with immunohistochemical staining. The clinical characteristics of patient were summarized in **Table 1**. The mean age at surgery, pre-

## ODAM inhibits the migration and invasion of prostate cancer

**Table 2.** Correlation of ODAM expression with age, preoperative PSA, Gleason score, lymph node metastasis, biomedical recurrence, PSM and T stage

Characteristics	ODAM expression (No. patients)		P value
	Low expression	High expression	
Age			0.456
≤60	19	33	
>60	16	20	
Preoperative PSA			<0.001
≤10	2	21	
>10	33	32	
Gleason score			<0.001
≤7	11	38	
>7	24	15	
Lymph node metastasis			<0.001
Negative	22	48	
Positive	13	5	
Biomedical recurrence			0.036
No	23	45	
Yes	12	8	
PSM			0.535
Negative	31	49	
Positive	4	4	
Pathological T stage			0.217
≤T2	26	45	
>T2	9	8	

operative PSA value, Gleason score and follow-up duration was 58.9±6.5 years, 15.3±6.3 ng/ml, 6.67±1.72, and 51.6±17.1 months, respectively. Positive surgical margin (PSM) was observed in 8 cases (9.1%), and 18 cases (20.4%) were found to have lymph node metastasis after surgery. After a mean of 51.6 months of follow-up, biochemical recurrence occurred in 20 cases (22.7%), clinical recurrence was observed in no patients. The representative results of low and high expressions were shown in **Figure 1A**. The ODAM expression was found to reside in the cytoplasm and paranuclear regions. Thirty-five cases (39.8%) with low expression of ODAM were observed. As shown in **Tables 1** and **2**, there was no significant difference between patients with low and high ODAM expression for the mean age (58.4±6.9 VS 59.2±6.4 years, P=0.597), follow-up duration (51.1±17.8 VS 52.0±16.8, p=0.808), PSM rate (11.4% VS 7.5%, p>0.05) and distribution of pathological T stage

(P=0.371), respectively. However, significant differences were observed in preoperative PSA values (20.2±5.2 VS 12.1±4.7 ng/ml, p<0.001) and Gleason score (7.4±1.4 VS 6.17±1.76, P=0.001) between low and high ODAM expression group. In patients with low and high ODAM expression, positive lymph node metastasis after surgery was observed in 13 (37.1%) and 5 (9.4%) patients, respectively. There was a statistically significant correlation between lymph node metastasis and low expression of ODAM (p<0.001). Biochemical recurrence occurred in 12 cases (34.3%) of patients with low ODAM expression, while in 8 cases (15.1%) of patients with high ODAM expression. Low level of ODAM expression was shown to be significantly correlated with biomedical recurrence (p=0.036) (**Table 2**).

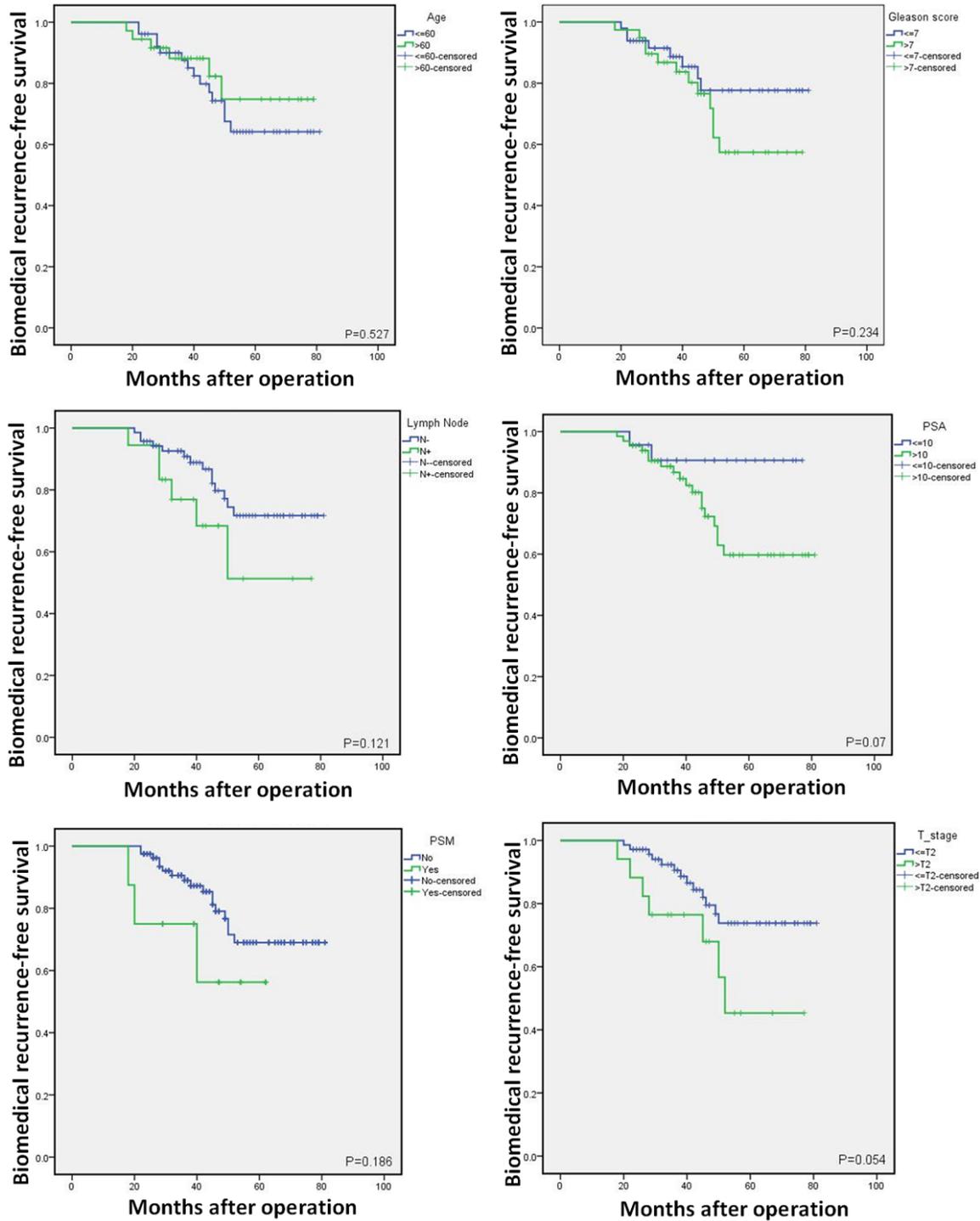
### *ODAM is an independent risk factor for biochemical recurrence*

To further determine whether ODAM expression and other clinic pathological characteristics are associated with prognosis of patients with prostate cancer, biomedical recurrence-free survival was compared in patients. As shown in **Figure 1B**, biomedical recurrence-free survival of patients with high ODAM expression was significantly larger than that of patients with low ODAM expression (P=0.012). No other characteristics showed statistical difference in biomedical recurrence-free survival of patients (**Figure 2**). Multivariate Cox regression analysis showed that low expression of ODAM (hazard ratio 3.902, 95% CI: 1.19-12.799, P=0.025) was the independent risk factor of biomedical recurrence during follow-up. No significant association was detected between biomedical recurrence and age, PSM, PSA level, Gleason score, T stage and lymph node metastasis (**Table 3**).

### *ODAM inhibits the invasion and migration of prostate cancer cells*

To understand the role of ODAM in prostate cancer progression, we examined the expression of ODAM in high-invasive PC3 cancer cells. Western blot was performed to determine the expression of ODAM in PC3 cells, and the result showed that PC3 cell lines did not express detectable ODAM protein. Next, we generated

## ODAM inhibits the migration and invasion of prostate cancer



**Figure 2.** Kaplan-Meier analysis of biochemical recurrence-free survival in patients based on age, preoperative PSA, Gleason score, lymph node metastasis, PSM and T stage respectively.

PC3 cells with stable overexpression of ODAM (**Figure 3A**). The data of cell growth assays did not show significant growth inhibitory effect in PC3-ODAM cells compared to PC3-CON cells (data not shown). To further determine whether

ODAM could affect PC3 cell migration, we conducted wound healing assay and the results revealed significant suppression of cell migration in PC3-ODAM cells compared to PC3-CON cells (**Figure 3B**). Consistently, the inhibitory

## ODAM inhibits the migration and invasion of prostate cancer

**Table 3.** Univariate and multivariate analyses of various prognostic parameters of biomedical recurrence.

Variables	No. patients	Univariate Analysis			Multivariate Analysis		
		P	HR	95% CI	P	HR	95% CI
Age		0.530	0.736	0.282-1.918			
≤60	52						
>60	36						
Preoperative PSA		0.091	3.524	0.817-15.208			
≤10	23						
>10	65						
Gleason score		0.242	1.707	0.697-4.179			
≤7	49						
>7	39						
Lymph node metastasis		0.13	2.104	0.803-5.514			
Negative	70						
Positive	18						
ODAM		0.017	2.987	1.215-7.346	0.025	3.902	1.19-12.799
High expression	53						
Low expression	35						
PSM		0.20	2.234	0.654-7.635			
Negative	80						
Positive	8						
T stage		0.063	2.395	0.955-6.007			
≤T2	71						
>T2	17						

effect of ODAM on cell invasion was confirmed by the results of Transwell invasion assay (**Figure 3C**). Furthermore, we found that the protein levels of MMP-2 were significantly inhibited after ODAM over-expression in PC3 cells (**Figure 3D**).

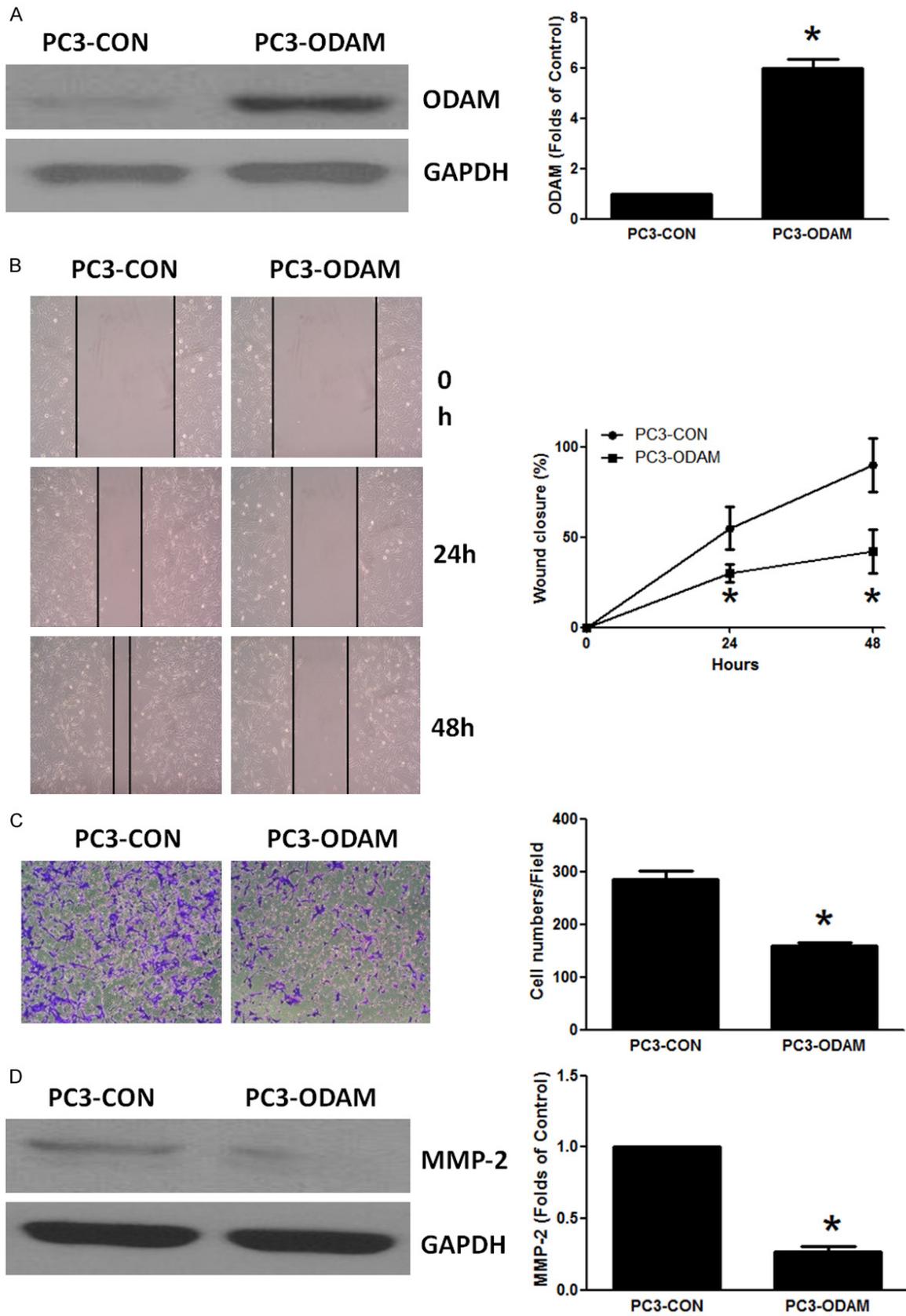
### Discussion

Prior studies of breast cancer patient biopsies suggested that ODAM served as a favorable prognostic biomarker and these patients with nuclear ODAM expression had an survival benefit regardless of tumor stage [23, 24]. In the present study, we analyzed 88 specimens including a series of Gleason score from 4-10 and the result showed that 53 cases (60.2%) was high expression of ODAM, which was found in the cytoplasm and paranuclear regions. Furthermore, low expression of ODAM was significantly correlated with lymph node metastasis, biomedical recurrence and progression-free survival rate, but not with mean age, follow-up duration, PSM rate and distribution of pathological T stage. Additionally, our results of multivariate analysis showed that low ODAM

expression was an independent predictor of biomedical recurrence, while the positive lymph node metastasis, Gleason score, and preoperative PSA were not the independent risks for biomedical recurrence. These data indicates ODAM may play a critical role in the metastasis and prognostic implications of prostate cancer.

Cell motility, which is essential for metastasis, is a complex, multistep process that integrates multiple proteins and cell signaling pathways. Tight junction proteins, such as VMP-1, JAM-A, PARD6B and Claudin-4, have been shown to take an important part on tumor invasion and metastasis [25, 26]. ODAM belongs to the family of tight junction proteins and have been shown to suppress the invasion and metastasis in breast cancer cells MCF-7 and MDA-MB-231, melanoma cells A375 and C8161 by activation of PTEN and inhibition of the phosphorylation of AKT [13, 27]. Moreover, ODAM expression could enhanced adhesion and apoptosis of the transfected breast cancer cells MDA-MB-231 and inhibited their growth rate, migratory and invasive activity *in vitro* and *in vivo* [23]. In our

ODAM inhibits the migration and invasion of prostate cancer



**Figure 3.** Overexpression of ODAM inhibits the migration and invasion of prostate cancer cells PC3. PC3 cells were transfected with the human ODAM pcDNA5T/O plasmid construct or the empty vector. After stably expressing cells

## ODAM inhibits the migration and invasion of prostate cancer

were generated, proteins were detected by Western blot with the indicated antibodies (A). Cell migration were analyzed by wound healing assays in PC3-CON and PC3-ODAM. The scratch-wounds were photographed at the indicated time points (0, 24 and 48 h) after cell scratching and the wound sizes were measured (B). Cells were seeded into the upper chamber of transwell with matrigel. Cell invasion were induced by FBS-containing media in the lower chamber. Invaded cells in the lower surface of the filters were stained and microphotographed after serum induction 24 h (C). Proteins were detected by Western blot with the indicated antibodies (D). The representative figures and quantified data from three individual experiments were shown. \*Represents  $P < 0.05$ .

case, overexpression of ODA M did not inhibit the growth of prostate cancer cells PC3, but significant suppress their invasion and migration. Additionally, the protein levels of MMP-2 were significantly inhibited after ODA M overexpression in PC3 cells. It is well known that tumor invasion and metastasis require increased expressions of matrix metalloproteinases (MMPs), where MMP-2 degrades type IV collagen in extracellular matrix and have been thought to be key enzymes [28-30]. Several groups have revealed that high expression of MMP-2 was correlated with lymph node metastasis of cancer [31, 32]. However, the molecular mechanisms of ODA M decreasing MMP-2 protein level and inhibiting the invasion and migration of prostate cancer need to be further investigated.

In summary, our study demonstrates that ODA M expression is significantly correlated with lymph node metastasis, preoperative PSA and Gleason score and serves as an independent risk for predicting biomedical recurrence in prostate cancer. Furthermore, our results also provide strong evidence for a functional role of ODA M in suppressing the migration and invasion of prostate cancer cells.

### Acknowledgements

The authors acknowledge the financial support from the Guangdong Natural Science Funds No. 2014A030313180 (JM. Di), National Natural Science Foundation of China No. 81201694 (Y. Luo), Specialized Research Fund for the Doctoral Program of Higher Education of China No. 20120171120059 (Y. Luo), Science and Technology Planning Project of Guangdong Province No. 2014A020212160 (Y. Luo), Young teacher training program of Sun Yat-sen University No. 15ykpy27 (Y. Luo) and Foundation of the 3rd Affiliated Hospital of Sun Yat-sen University (Y. Luo).

### Disclosure of conflict of interest

None.

**Address correspondence to:** Dr. Jin-Ming Di, Department of Urology, The 3rd Affiliated Hospital of Sun Yat-Sen University, No 600, Tianhe Road, Tianhe District, Guangzhou, Guangdong, China. Tel: +86-20-85252052; Fax: +86-20-85252678; E-mail: dijinm@mail.sysu.edu.cn

### References

- [1] Gudem G, Van Loo P, Kremeyer B, Alexandrov LB, Tubio JM, Papaemmanuil E, Brewer DS, Kallio HM, Hognas G, Annala M, Kivinummi K, Goody V, Latimer C, O'Meara S, Dawson KJ, Isaacs W, Emmert-Buck MR, Nykter M, Foster C, Kote-Jarai Z, Easton D, Whitaker HC, Neal DE, Cooper CS, Eeles RA, Visakorpi T, Campbell PJ, McDermott U, Wedge DC and Bova GS. The evolutionary history of lethal metastatic prostate cancer. *Nature* 2015; 520: 353-357.
- [2] Di JM, Pang J, Sun QP, Zhang Y, Fang YQ, Liu XP, Zhou JH, Ruan XX and Gao X. Toll-like receptor 9 agonists up-regulates the expression of cyclooxygenase-2 via activation of NF-kappaB in prostate cancer cells. *Mol Biol Rep* 2010; 37: 1849-1855.
- [3] Di JM, Zhou J, Zhou XL, Gao X, Shao CQ, Pang J, Sun QP, Zhang Y and Ruan XX. Cyclooxygenase-2 expression is associated with vascular endothelial growth factor-C and lymph node metastases in human prostate cancer. *Arch Med Res* 2009; 40: 268-275.
- [4] Luo Y, Jiang QW, Wu JY, Qiu JG, Zhang WJ, Mei XL, Shi Z and Di JM. Regulation of migration and invasion by Toll-like receptor-9 signaling network in prostate cancer. *Oncotarget* 2015; 6: 22564-22574.
- [5] Moffatt P, Wazen RM, Dos Santos Neves J and Nanci A. Characterisation of secretory calcium-binding phosphoprotein-proline-glutamine-rich 1: a novel basal lamina component expressed at cell-tooth interfaces. *Cell Tissue Res* 2014; 358: 843-855.
- [6] Murphy CL, Kestler DP, Foster JS, Wang S, Macy SD, Kennel SJ, Carlson ER, Hudson J, Weiss DT and Solomon A. Odontogenic ameloblast-associated protein nature of the amyloid found in calcifying epithelial odontogenic tumors and unerupted tooth follicles. *Amyloid* 2008; 15: 89-95.
- [7] Lee HK, Park JT, Cho YS, Bae HS, Cho MI and Park JC. Odontogenic ameloblasts-associated

## ODAM inhibits the migration and invasion of prostate cancer

- protein (ODAM), via phosphorylation by bone morphogenetic protein receptor type IB (BMPRI-B), is implicated in ameloblast differentiation. *J Cell Biochem* 2012; 113: 1754-1765.
- [8] Lee HK, Ji S, Park SJ, Choung HW, Choi Y, Lee HJ, Park SY and Park JC. Odontogenic Ameloblast-associated Protein (ODAM) Mediates Junctional Epithelium Attachment to Teeth via Integrin-ODAM-Rho Guanine Nucleotide Exchange Factor 5 (ARHGGEF5)-RhoA Signaling. *J Biol Chem* 2015; 290: 14740-14753.
- [9] Lee HK, Lee DS, Ryoo HM, Park JT, Park SJ, Bae HS, Cho MI and Park JC. The odontogenic ameloblast-associated protein (ODAM) cooperates with RUNX2 and modulates enamel mineralization via regulation of MMP-20. *J Cell Biochem* 2010; 111: 755-767.
- [10] Holcroft J and Ganss B. Identification of amelotin- and ODAAM-interacting enamel matrix proteins using the yeast two-hybrid system. *Eur J Oral Sci* 2011; 119 Suppl 1: 301-306.
- [11] Kestler DP, Foster JS, Macy SD, Murphy CL, Weiss DT and Solomon A. Expression of odontogenic ameloblast-associated protein (ODAM) in dental and other epithelial neoplasms. *Mol Med* 2008; 14: 318-326.
- [12] Lee HK, Park SJ, Oh HJ, Kim JW, Bae HS and Park JC. Expression pattern, subcellular localization, and functional implications of ODAAM in ameloblasts, odontoblasts, osteoblasts, and various cancer cells. *Gene Expr Patterns* 2012; 12: 102-108.
- [13] Lee HK, Choung HW, Yang YI, Yoon HJ, Park IA and Park JC. ODAAM inhibits RhoA-dependent invasion in breast cancer. *Cell Biochem Funct* 2015; 33: 451-461.
- [14] Hou GL, Luo Y, Di JM, Lu L, Yang Y, Pang J, Si-Tu J and Gao X. Predictors of urinary continence recovery after modified radical prostatectomy for clinically high-risk prostate cancer. *Urol J* 2015; 12: 2021-2027.
- [15] Li K, Pang J, Cheng H, Liu WP, Di JM, Xiao HJ, Luo Y, Zhang H, Huang WT, Chen MK, Li LY, Shao CK, Feng YH and Gao X. Manipulation of prostate cancer metastasis by locus-specific modification of the CRMP4 promoter region using chimeric TALE DNA methyltransferase and demethylase. *Oncotarget* 2015; 6: 10030-10044.
- [16] Shi Z, Park HR, Du Y, Li Z, Cheng K, Sun SY, Fu H and Khuri FR. Cables1 complex couples survival signaling to the cell death machinery. *Cancer Res* 2015; 75: 147-158.
- [17] Shi Z, Li Z, Li ZJ, Cheng K, Du Y, Fu H and Khuri FR. Cables1 controls p21/Cip1 protein stability by antagonizing proteasome subunit alpha type 3. *Oncogene* 2015; 34: 2538-2545.
- [18] Li Z, Park HR, Shi Z, Pham CD, Du Y, Khuri FR, Zhang Y, Han Q and Fu H. Pro-oncogenic function of HIP-55/Drebrin-like (DBNL) through Ser269/Thr291-phospho-sensor motifs. *Oncotarget* 2014; 5: 3197-3209.
- [19] Mei XL, Yang Y, Zhang YJ, Li Y, Zhao JM, Qiu JG, Zhang WJ, Xue YQ, Zheng DW, Chen Y, Qin WM, Wei MN and Shi Z. Sildenafil inhibits the growth of human colorectal cancer *in vitro* and *in vivo*. *Am J Cancer Res* 2015; 5: 3311-24.
- [20] Xie FF, Pan SS, Ou RY, Zheng ZZ, Huang XX, Jian MT, Qiu JG, Zhang WJ, Yang Y, Li WF, Shi Z and Yan XJ. Volasertib Suppresses Tumor Growth and Potentiates the Activity of Cisplatin in Cervical Cancer. *Am J Cancer Res* 2015; 5: 3548-59.
- [21] Gao X, Pang J, Li LY, Liu WP, Di JM, Sun QP, Fang YQ, Liu XP, Pu XY, He D, Li MT, Su ZL and Li BY. Expression profiling identifies new function of collapsin response mediator protein 4 as a metastasis-suppressor in prostate cancer. *Oncogene* 2010; 29: 4555-4566.
- [22] Lei Y, Li B, Tong S, Qi L, Hu X, Cui Y, Li Z, He W, Zu X, Wang Z and Chen M. miR-101 suppresses vascular endothelial growth factor C that inhibits migration and invasion and enhances cisplatin chemosensitivity of bladder cancer cells. *PLoS One* 2015; 10: e0117809.
- [23] Kestler DP, Foster JS, Bruker CT, Prenshaw JW, Kennel SJ, Wall JS, Weiss DT and Solomon A. ODAAM Expression Inhibits Human Breast Cancer Tumorigenesis. *Breast Cancer (Auckl)* 2011; 5: 73-85.
- [24] Siddiqui S, Bruker CT, Kestler DP, Foster JS, Gray KD, Solomon A and Bell JL. Odontogenic ameloblast associated protein as a novel biomarker for human breast cancer. *Am Surg* 2009; 75: 769-775; discussion 775.
- [25] Martin TA and Jiang WG. Loss of tight junction barrier function and its role in cancer metastasis. *Biochim Biophys Acta* 2009; 1788: 872-891.
- [26] Cunliffe HE, Jiang Y, Fornace KM, Yang F and Meltzer PS. PAR6B is required for tight junction formation and activated PKCzeta localization in breast cancer. *Am J Cancer Res* 2012; 2: 478-491.
- [27] Foster JS, Fish LM, Phipps JE, Bruker CT, Lewis JM, Bell JL, Solomon A and Kestler DP. Odontogenic ameloblast-associated protein (ODAM) inhibits growth and migration of human melanoma cells and elicits PTEN elevation and inactivation of PI3K/AKT signaling. *BMC Cancer* 2013; 13: 227.
- [28] Egeblad M and Werb Z. New functions for the matrix metalloproteinases in cancer progression. *Nat Rev Cancer* 2002; 2: 161-174.
- [29] Wang Y, Liao H, Zheng HC, Li L, Jia L, Zhang Z and Zheng W. Effect of luteinizing hormone-induced prohibitin and matrix metalloproteinases on ovarian epithelial tumor cell proliferation. *Am J Cancer Res* 2015; 5: 114-124.

## ODAM inhibits the migration and invasion of prostate cancer

- [30] Li YY, Zhou CX and Gao Y. Podoplanin promotes the invasion of oral squamous cell carcinoma in coordination with MT1-MMP and Rho GTPases. *Am J Cancer Res* 2015; 5: 514-529.
- [31] de Vicente JC, Fresno MF, Villalain L, Vega JA and Hernandez Vallejo G. Expression and clinical significance of matrix metalloproteinase-2 and matrix metalloproteinase-9 in oral squamous cell carcinoma. *Oral Oncol* 2005; 41: 283-293.
- [32] Katayama A, Bando N, Kishibe K, Takahara M, Ogino T, Nonaka S and Harabuchi Y. Expressions of matrix metalloproteinases in early-stage oral squamous cell carcinoma as predictive indicators for tumor metastases and prognosis. *Clin Cancer Res* 2004; 10: 634-640.