Original Article Hepatoprotective effects of Gentianella turkestanerum extracts on acute liver injury induced by carbon tetrachloride in mice

Jianghua Yang^{1*}, Dandan Zhu^{1*}, Bowei Ju¹, Xiangying Jiang², Junping Hu³

¹Department of Pharmacy, The First Affiliated Hospital, Xinjiang Medical University, Urumqi 830011, Xinjiang Uyghur Autonomous Region, China; ²Food and Drug Institute, Bainguoleng Mongolia Autonomous Region, Urumqi 830011, Xinjiang Uyghur Autonomous Region, China; ³Department of Natural Medicines, College of Pharmacy, Xinjiang Medical University, Urumqi 830011, Xinjiang Uyghur Autonomous Region, China. *Equal contributors.

Received July 29, 2016; Accepted January 12, 2017; Epub February 15, 2017; Published February 28, 2017

Abstract: Objective: To investigate the contents of secoiridoid compounds (i.e. sweroside, swertiamarin and gentiopicrin) from Gentianella turkestanerum extracts, and the potential effects of G. turkestanerum extracts against carbon tetrachloride (CCl₄) induced liver injury in mice. Methods: The contents of swertiamarin, gentiopicroside and sweroside from different G. turkestanerum extracts were determined with high performance liquid chromatography (HPLC). CCl, was used to induce acute liver injury in mice. The serum aspartate amino transferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP), total protein (TP), total bilirubin (TB), superoxide dismutase (SOD), malondialdehyde (MDA), glutathione transferase (GSH) and catalase (CAT) were measured. HE staining was performed to investigate the pathological changes of liver. Results: Iridoid glycoside showed the highest content in the product extracted by butanol (designated as GBA), but lower in the products extracted by ethyl acetate and water designated as GEA and GW, respectively. All G. turkestanerum extracts showed protective effects against CCI, induced acute liver injury in mice, among which GBA showed the maximal protective effects. G. turkestanerum extracts induced significant decrease in the serum ALT, AST, ALP and TB compared with those in the mice with acute lung injury (P < 0.01). Obvious increase was noticed in serum TP (P < 0.01). Moreover, such effects presented in a dose-dependent manner. Compared with the control group, the MDA was significantly elevated in the model group (P < 0.01), while significant decrease was observed in the levels of SOD, GSH and CAT in model group compared with the control group (P < 0.01). Whereas, such phenomenon was completely reversed by G. turkestanerum extracts in a dose-dependent manner. Conclusion: G. turkestanerum showed protective effects against CCl, induced acute liver injury in mice.

Keywords: Gentianella turkestanerum, different polar parts, iridoid glycoside, HPLC, carbon tetrachloride, hepatoprotective effect

Introduction

Liver is the major organ involved in xenobiotic metabolism and its damage could be caused by virus infiltration, drugs, and toxic chemicals from infection or ingestion [1, 2]. The risk of liver dysfunction has significantly increased due to the exposure to environmental pesticides, toxins and chemotherapeutics [3]. Nowadays, the management of acute and chronic hepatic injury is still a challenge worldwide despite the advances of modern medical technologies [4]. This leads us to identify any potential agents that may alleviate the hepatic injury effectively. Plants of *Gentiana* genus, a productive source in traditional Chinese medicine, are often used to ease pain, dispel rheumatism, treat liver jaundice with damp-heat pathogen, chronic pharyngitis and headache [5]. Recently, several studies have indicated that plants of *Gentianella* genus may have hepatoprotective effects [6-8]. For example, *G. cruciata and G. asclepiadea* could significantly decrease the levels of serum aspartate amino transferase (AST) and alanine aminotransferase (ALT) in rats with hepatic injury, and contribute to the attenuation of the inflammation and necrosis in hepatic tissue [6, 7]. In addition, *G. veitchiorum* was reported to exert hepatoprotective effects against

iridoid compounds in Gentianella turkestan- erum extracts by HPLC analysis							
Sample	Content (%)						
	Swertiamarin	Gentiopicrin	Sweroside				
GE	3.07	2.15	0.08				
GW	2.30	0.96	0.16				

5.69

1.54

ND

0.75

0.11

ND

Table 1. Quantitative determination of seco-

Note: ND, Not detected.

6.12

0.27

ND

GBA

GEA

GPE

carbon tetrachloride (CCl₄) induced hepatotoxicity in mice [8]. Gentianella turkestanerum, mainly distributed in Xinjiang and Inner Mongolia, has been frequently used for the management of diseases in these regions [9]. To our best knowledge, most of the studies on G. turkestanerum focused on the identification and quality control, but rare studies have been carried out to investigate the hepatoprotective effects of G. turkestanerum.

In this study, the effects and possible mechanisms of different G. turkestanerum extracts against CCI, induced hepatic injury were investigated. Our results showed these extracts presented protective effects on the hepatic injury in a dose dependent manner. Also, the safety of these extracts were approved with no toxicity in a dose of up to 2000 mg/Kg.

Materials and methods

Animals

ICR male mice (SPF grade, weighing 20±2 g) were obtained from the Experimental Animal Center of Xinjiang Medical University (Approve No.: SCXK 2011-0004). The animals were subject to adaptive feeding for 1 week on 12 h light/12 h dark cycle, at room temperature (24±2°C), with a relative humidity of 50%±15%. All animals were access to standard diet and water. The study protocols were approved by the Animal Ethics Committees of the First Affiliated Hospital, Xinjiang Medical University.

Chemicals and equipments

Aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP), total protein (TP), total bilirubin (TB) were purchased from the Mindray Bio-Medical Electronics Co., Ltd. (Shenzhen, China). Standard substances of swertiamarin (batch No. 2012-0129), gentiopicroside (batch No. 20120114) and sweroside (batch No. PN1125SA13) were purchased from the Yuanye Biotechnology Co. Ltd. (Shanghai, China). Silymarin was purchased from Sigma Aldrich (CA, USA). CCI, paraform (4%), ultrapure water, methanol were of chromatographic purity, and the other chemicals or reagents were of analytical grade.

Plant material and preparation of the extracts

G. turkestanerum was collected from the Gongnaisi and Bayinbuluke Forest Farm (Xinjiang, China). The collected plant material was authenticated by Professor Jiang XY in the Institute for Food and Drug Inspection (Xinjiang, China).

After grind, G. turkestanerum samples (0.5 kg) were extracted using 95% ethanol for 2 hrs. The extracts were concentrated under reduced pressure in a rotary evaporator to obtain ethanol extract (designated as GE). The GE was resuspended with sterile water, and extracted with petroleum benzene, ethyl acetate and butanol. Subsequently, the products extracted by petroleum benzene (GPE), ethyl acetate (GEA), butanol (GBA) and water (GW) were obtained, respectively.

HPLC analysis

The content of secoiridoides was analyzed using HPLC analysis according to the previous description [10]. Briefly, Cosmosil Packed Column 5C₁₈-PAQ (250 mm ×4.6 mm, 5 μm) was used for secoiridoid analysis. The temperature of the column was set at 25°C. The mobile phase was a mixture of methanol and 0.04% phosphoric acid (19:81, v/v). The flow rate was set to 1.0 mL/min, and a wavelength of 242 nm was used for the determination.

The standard substances swertiamarin, gentiopicroside and sweroside were dissolved in methanol and then diluted to the appropriate concentration ranges to establish the calibration curves. Linear regression analysis revealed an excellent linearity with correlation coefficient for swertiamarin (4.04-129.54 mg/L, r= 0.9994), gentiopicroside (3.96-126.73 mg/L, r=0.9993) and sweroside (0.93-29.70 mg/L, r=0.9995), respectively.



Figure 1. HPLC chromatogram of mixed reference substances (A) and the samples of different G. *turkestanorum* extracts including GE (B), GW (C), GBA (D), GEA (E) and GPE (F). The peaks of 1-3 stands for swertiamarin, gentiopicroside and sweroside, respectively.

horam on moc body weight							
Group	Sex	Base After		Body weight			
		level (g)	treatment (g)	increase (%)			
Control	F	20.620±1.195	27.940±0.513	40±0.513 35.50			
	Μ	21.140±0.999	29.680±0.672	40.40			
GE	F	20.600±0.639	27.700±0.982	34.47			
	Μ	21.200±0.678	27.720±0.847	30.75			
GW	F	20.820±0.887	26.560±0.654	27.57			
	Μ	21.340±0.713	27.420±0.540	28.50			
GBA	F	20.667±0.582	27.060±0.907	30.93			
	Μ	20.550±1.294	27.260±0.555	32.65			
GEA	F	20.517±1.057	26.860±0.673	30.92			
	Μ	21.100±0.725	27.520±0.531	30.43			

Table 2. Effect of different extracts from G. turkesta-					
norum on mice body weight					

Acute toxicity testing

Acute oral toxicity study was performed according to the OECD guidelines [11]. Mice (n=3) were randomly selected for the acute toxicity study using each agent. The animals were fasted overnight prior to the experiment and maintained under standard laboratory conditions. The extracts (i.e. GE, GW, GBA, GEA) were given via lavage in each mice (18-22 g) with a dose of 200 mg/mL body weight. Then the hair color, activity, respiration, urination and defecation, as well as toxicity or even death were observed.

Experimental design

The mice were randomly divided into 15 groups (n=12 in each group) as follows: (i) Group 1 served as the control, which was given normal saline (NS) daily for 2 weeks; (ii) Group 2: subject to hepatic injury using CCI_4 (10 mL/kg) by intraperitoneal (IP) injection; and (iii) After induction of hepatic

injury, the animals were subject to silymarin (100 mg/kg) (Group 3, positive control), GE extract dissolved in normal saline (NS) at 100 mg/kg (Group 4), 200 mg/kg (Group 5) and 400 mg/kg (Group 6), GW extract dissolved in NS at 100 mg/kg (Group 7), 200 mg/kg (Group 8) and 400 mg/kg (Group 9), GBA extract dis-



Figure 2. The effects of different extracts from *G. turkestanorum* on mice liver (A) and spleen index (B). Values are mean \pm S.E.M., n=10 in each group. **P* < 0.05, compared to the control group; ***P* < 0.01, compared to the control group; #*P* < 0.05, compared to the model group; #*P* < 0.01, compared to the model group.



Figure 3. Effects of *G. turkestanerum* extracts and silymarin on the levels of liver index (A) and spleen index (B) after CCl_4 treatment in mice. Group 1-Control group; Group 2-CCl₄ (0.1% in peanut oil) 10 ml/kg i.p; Group 3-Silymarin (100 mg/kg) + CCl_4 ; Group 4-GE 100 mg/kg + CCl_4 ; Group 5-GE 200 mg/kg + CCl_4 ; Group 6-GE 400 mg/kg + CCl_4 ; Group 7-GW 100 mg/kg + CCl_4 ; Group 8-GW 200 mg/kg + CCl_4 ; Group 9-GW 400 mg/kg + CCl_4 ; Group 10-GBA 100 mg/kg + CCl_4 ; Group 11-GBA 200 mg/kg + CCl_4 ; Group 12-GBA 400 mg/kg + CCl_4 ; Group 13-GEA 100 mg/kg + CCl_4 ; Group 14-GEA 200 mg/kg + CCl_4 ; Group 15-GEA 400 mg/kg + CCl_4 . Values are mean ± S.E.M., n=12 animals in each group. **P < 0.01, compared to the control group; "P < 0.05, compared to the model group; "#P < 0.01, compared to the model group.

solved in NS at 100 mg/kg (Group 10), 200 mg/kg (Group 11) and 400 mg/kg (Group 12), GEA extract dissolved in NS at 100 mg/kg (Group 13), 200 mg/kg (Group 14) and 400 mg/kg (Group 15), respectively. On the last day of the treatment, the animals in groups 2-15 were subject to a single dose of 0.1% CCl₄ dissolved in peanut oil at 10 mL/kg intraperitone-ally. Eighteen hours after CCl₄ administration, all the animals were sacrificed and blood samples were collected from the eyeballs. The liver and spleen were collected for biochemical analysis.

Biochemical assay

Collected blood samples were placed at room temperature for 30 min, and centrifuged at 3,000 rpm for 15 min at 4°C. Liver injury was evaluated by estimation of serum ALT, AST, ALP, TP and TB using automatic biochemistry analyzer.

Liver antioxidant markers analysis

Hepatic tissues (0.5 g) were homogenated in 0.9% NS, followed by centrifugation at 2,500 rpm for 15 min. Commercial kits were used for

Group	ALT (U/L)	AST (U/L)	ALP (U/L)	TP (g/L)	TB (µmoL/L)
1	50.44±7.26	129.91±19.35	48.82±6.38	48.98±4.50	2.66±0.47
2	283.80±26.91**	304.40±21.61**	68.76±5.74**	39.18±2.91**	5.37±0.63**
3	69.33±16.69**	149.31±34.33##	45.15±7.06##	49.20±4.18##	3.42±0.48 ^{##}
4	246.75±27.73##	250.39±25.91##	64.95±4.66	43.33±1.91#	4.72±0.41##
5	142.51±25.20##	196.95±16.95##	46.32±2.66##	47.23±2.52##	4.23±0.30##
6	99.82±16.15##	149.66±13.82##	36.11±3.07##	50.08±1.94##	3.75±0.28##
7	265.16±19.26#	261.89±18.34##	66.80±4.14	40.98±4.63	4.97±0.35#
8	172.63±22.89##	208.19±14.10##	49.61±5.52##	44.60±2.68##	4.06±0.33##
9	117.77±14.26##	179.34±14.92##	28.95±6.93##	46.97±2.32##	3.67±0.28##
10	211.33±15.43##	233.46±24.94##	62.74±7.66	45.40±3.74##	5.10±0.35
11	125.31±11.95##	193.47±22.27##	45.52±5.18##	47.94±2.14##	4.32±0.36##
12	58.54±14.00##	128.06±20.04##	33.93±7.29##	50.77±2.74##	3.57±0.44##
13	279.86±19.13	272.45±8.65##	65.44±5.21	39.51±2.57	5.30±0.33
14	188.56±12.61##	223.44±12.86##	49.90±5.77##	44.56±2.90##	4.77±0.38##
15	166.31±12.23##	163.94±9.12##	36.68±8.27**	46.95±2.01##	3.92±0.31##

Table 3. Effects of *G. turkestanerum* extracts and silymarin on serum biochemical parameters of CCl_4 intoxicated mice

Group 1-Control group; Group 2-CCl₄ (0.1% in peanut oil) 10 ml/kg i.p; Group 3-Silymarin (100 mg/kg) + CCl₄; Group 4-GE 100 mg/kg + CCl₄; Group 5-GE 200 mg/kg + CCl₄; Group 6-GE 400 mg/kg + CCl₄; Group 7-GW 100 mg/kg + CCl₄; Group 8-GW 200 mg/kg + CCl₄; Group 9-GW 400 mg/kg + CCl₄; Group 10-GBA 100 mg/kg + CCl₄; Group 11-GBA 200 mg/kg + CCl₄; Group 12-GBA 400 mg/kg + CCl₄; Group 13-GEA 100 mg/kg + CCl₄; Group 14-GEA 200 mg/kg + CCl₄; Group 15-GEA 400 mg/kg + CCl₄; Group 14-GEA 200 mg/kg + CCl₄; Group 15-GEA 400 mg/kg + CCl₄. Values are mean \pm S.E.M., n=12 animals in each group. ***P* < 0.01, compared to the control group; #*P* < 0.05, compared to the model group.

the determination of TP, superoxide dismutase (SOD), malondialdehyde (MDA), glutathione (GSH) and catalase (CAT), according to the manufacturer's instructions.

Hematoxylin-eosin staining

Liver slices were fixed in 4% paraform, and were embedded in paraffin after dehydration. The sections (4-5 μ m) were stained with hematoxy-lin-eosin as routinely described. The images were observed under a light microscope.

Statistical analysis

The data were expressed as mean \pm standard deviation. One-way analysis of variance (ANOVA) was used for the comparison among groups. Student's t test was used for comparison between groups. Data analysis was carried out using SPSS 19.0 software. P < 0.05 was considered to be statistically significant.

Results

HPLC analysis of secoiridoides

Table 1 showed the content of secoiridoid compounds (i.e. sweroside, swertiamarin and gen-

tiopicrin) in the GBA, GE, GW, GEA and GPE. Three secoiridoid compounds were identified in all the extracts except GPE. In addition, the content of these secoiridoid compound was much more higher in GBA than the others (**Figure 1**).

Acute toxicity testing

No mortality was noticed after administration of the extracts. The body weight was increased in each mice after administration of GBA, GE, GW, and GEA, but it showed no statistical difference compared with the control group (P >0.05, **Table 2**). Besides, the liver index and spleen index in these groups showed no statistical difference compared with the control group (P > 0.05, **Figure 2**). This indicated GBA, GE, GE and GEA showed no effects on the growth of the mice. All the extracts of *G. turkestanorum* were found to be safe in a dose of up to 2,000 mg/kg. Hence 1/20, 1/10 and 1/5 of the maximum dose (i.e., 100, 200 and 400 mg/kg) were suitable for the study.

Effects of G. turkestanorum extract on the liver and spleen index

As shown in **Figure 3**, compared with the control group, the liver and spleen index in mice



Figure 4. Effects of *G. turkestanerum* extracts and silymarin on the levels of hepatic MDA (A), SOD (B), GSH (C) and CAT (D) after CCl_4 treatment in mice. Group 1-Control group; Group 2- Ccl_4 (0.1% in peanut oil) 10 ml/kg i.p; Group 3-Silymarin (100 mg/kg) + Ccl_4 ; Group 4-GE 100 mg/kg + Ccl_4 ; Group 5-GE 200 mg/kg + Ccl_4 ; Group 6-GE 400 mg/kg + Ccl_4 ; Group 7-GW 100 mg/kg + Ccl_4 ; Group 8-GW 200 mg/kg + Ccl_4 ; Group 9-GW 400 mg/kg + Ccl_4 ; Group 10-GBA 100 mg/kg + Ccl_4 ; Group 11-GBA 200 mg/kg + Ccl_4 ; Group 12-GBA 400 mg/kg + Ccl_4 ; Group 13-GEA 100 mg/kg + Ccl_4 ; Group 14-GEA 200 mg/kg + Ccl_4 ; Group 15-GEA 400 mg/kg + Ccl_4 . Values are mean \pm S.E.M., n=12 animals in each group. ***P* < 0.01, compared to the control group; "*P* < 0.05, compared to the model group.

were significantly higher in the model group (P < 0.01). Silymarin and different *G. turkestanorum* extracts could significantly decrease the liver and spleen index induced by CCl₄ (P < 0.05).

Effects of G. turkestanerum extracts on serum ALT, AST, ALP, TP and TB

Significant increase was noticed in the serum ALT, AST, ALP and TB in the model group compared with the control group (P < 0.01). However, the level of TP was significantly lower in the model group than the control group (P < 0.01). In mice treated by silymarin, significant decrease was observed in the serum ALT, AST, ALP and TB compared with those in the mice treated by CCl₄ (P < 0.01). Whereas, silymarin induced obvious increase of serum TP (P < 0.01). Moreover, the effects presented in a dose-dependent manner, in which the GBA (400 mg/Kg) showed the maximal effects (P < 0.01, **Table 3**).

Effects of G. turkestanerum extracts on SOD, MDA, GSH and CAT in liver

As shown in **Figure 4**, compared with the control group, the level of MDA was significantly higher in the model group (P < 0.01), while the levels of SOD, GSH and CAT were significantly lower (P < 0.01). Silymarin could significantly inhibit the level of MDA, SOD, GSH and CAT compared with the model group (P < 0.01). Meanwhile, *G. turkestanerum* extracts showed similar effects, and presented in a dose-dependent manner. Among these extracts, GBA with a concentration of 400 mg/Kg showed the strongest effects (P < 0.01).

Histopathological examination

The hepatic lobe was clearly displayed in the control group, together with no aberrant changes in the cellular structure (**Figure 5A**). The liver sections in model group revealed extensive liver injuries, characterized by loss of cellular



Figure 5. Photomicrographs of liver sections from: Group 1-Control group (A) ; Group 2-CCl₄ (0.1% in peanut oil) 10 ml/kg i.p (B); Group 3-Silymarin (100 mg/kg) + CCl₄ (C); Group 4-GE 100 mg/kg + CCl₄ (D); Group 5-GE 200 mg/kg + CCl₄ (E); Group 6-GE 400 mg/kg + CCl₄ (F); Group 7-GW 100 mg/kg + CCl₄ (G); Group 8-GW 200 mg/kg + CCl₄ (H); Group 9-GW 400 mg/kg + CCl₄ (I); Group 10-GBA 100 mg/kg + CCl₄ (J); Group 11-GBA 200 mg/kg + CCl₄ (K); Group 12-GBA 400 mg/kg + CCl₄ (L); Group 13-GEA 100 mg/kg + CCl₄ (M); Group 14-GEA 200 mg/kg + CCl₄ (N); Group 15-GEA 400 mg/kg + CCl₄ (O). H&E, original magnification 200×. Arrow: CV- central vein; BD- ballooning degeneration; C- congestion; LI- lymphocyte infiltration; FD- fatty degeneration; HE- hepatocytes edema.

boundaries, irregular shape, obvious hepatocyte edema, ballooning degeneration, fatty degeneration, necrosis, and the inflammatory cell infiltration around the central vein (Figure 5B).

The silymarin could significantly ameliorate the liver injury, and the hepatocyte structure was normal in the silymarin treated group (**Figure 5C**). Pretreatment with *G. turkestanerum* extracts could ameliorate the hepatocyte denaturation and necrosis induced by CCl_4 (**Figure 5D-K**). The GBA extracts (400 mg/kg) could significantly ameliorate the hepatocyte edema, ballooning degeneration and inflammatory cell infiltration (**Figure 5L**). Other extracts of a concentration of 400 mg/kg may induce necrosis and inflammatory cell infiltration (**Figure 5M-O**).

Discussion

In this study, we investigated the effects and the possible mechanism of different *G. turkestanerum* extracts against CCI_4 induced hepatic injury in mice. Our results indicated that *G. turkestanerum* extracts showed protective effects against CCI_4 induced acute lung injury in mice. Besides, secoiridoides may play crucial roles in the hepatoprotective effects.

 CCl_4 has been widely used to induce liver injury in animal models [12, 13]. CCl_4 is metabolized to the trichloromethyl radical (CCl_3) and proxy trichloromethyl radical (CCl_3O_2) by cytochrome P4502E1 (CYP2E1), which initiates lipid peroxidation and destroys polyunsaturated fatty acid [14-16]. High levels of ROS damage cells are involved in several human pathologies, including liver cirrhosis and fibrosis [17]. These processes affect the permeability of endoplasmic reticulum, mitochondrial, and plasma membranes, which finally lead to leakage of liver enzymes into the blood [18]. Therefore, estimation of enzyme activity in the serum is a useful tool to assess the hepatocellular damages [19].

Elevation of serum ALT and AST levels contributed to liver damage as they were localized in the cytoplasm normally and released into the serum after the increase of membrane permeability [20, 21]. ALP and TB are indicators of pathological alteration in biliary flow [22], and serum ALP and TB were increased in the presence of liver damage [23, 24]. The TP level will be decreased under hepatotoxic conditions because of defective protein biosynthesis in liver [25, 26]. In our study, compared with the control group, the liver and spleen index, the levels of ALT, AST, ALP and TB were significantly higher in the model group (P < 0.01), while the level of TP was significantly lower (P < 0.01). Our results indicated that *G. turkestanerum* extracts could significantly decrease the liver and spleen index. Besides, these extracts induced significant decrease of ALT, AST, ALP and TB in a dose dependent manner. Moreover, it could significantly increase the level of TP (P < 0.05), especially in mice treated by GBA (400 mg/Kg, P < 0.01). Taken together, it is reasonable to conclude that *G. turkestanerum* shows protective effects against CCl₄ induced acute liver injury in mice.

Oxidative stress induced by reactive oxygen species (ROS) plays a key role in hepatotoxicity [27]. It could cause liver injury through initiating lipid peroxidation, changing biomembrane function, and destroying the enzyme activity [28]. MDA is the end product of lipid peroxidation, and the elevation of MDA in serum is an indicator of liver injury [29-32]. GSH is an important intracellular antioxidant that could protect cell from damage by peroxides, free radicals and toxins [33-36]. SOD and CAT were the major components of the antioxidant system, and the decreased levels represented the liver damage [2, 37]. In our study, G. turkestanerum extracts induced significant decrease of MDA and obvious increase of SOD, GSH and CAT in a dose depended manner, especially after treatment of GBA (400 mg/kg, P < 0.01). These findings suggested that G. turkestanerum may be useful as a hepatoprotective agent against CCl, induced liver injury. The mechanisms of protection may include the increase in antioxidant enzyme activity and the inhibition of lipid peroxidation process.

Liver pathological section could directly reflect the degree of liver damage. In our study, the liver sections after CCI_4 treatment revealed extensive liver injuries, characterized by obvious hepatocyte edema, ballooning degeneration, fatty degeneration, necrosis, and the inflammatory cell infiltration around the central vein. Pretreatment with *G. turkestanerum* extracts (400 mg/kg) could ameliorate the hepatocyte denaturation and necrosis induced by CCI_4 . Also, the effects of *G. turkestanerum* extracts on liver pathological section showed a dose depended manner, especially in mice treated with GBA (400 mg/Kg, P < 0.01).

In our study, toxicity studies showed that no mortality was observed up to a dose level of 2000 mg/kg body weight. Besides, no signs of

changes were found in the skins, furs and muscle of all animals. Central nerve response, respiration, behavior patterns, and sleep were similar to the normal group. The food intake was normal and no sialorrhea, lacrimation, rhinorrhea, dyspnea, diarrhea, constipation, and intestinal tympanites were noticed. Our results showed that all the extracts of *G. turkestanorum* were found to be safe in a dose of up to 2000 mg/kg.

Phytochemical studies revealed that secoiridoides was the mainly active constituent in gentian [5, 38, 39]. In our study, HPLC showed that the content sequence of the total secoiridoides in extracts was as follows: GBA (12%) > GE (5%) > GW (3.4%) > GEA (1.9%) > GPE (0). The sequence of the protective effects for extracts was as follows: GBA > GE > GW > GEA. Our results indicated that secoiridoides may be the main active constitute in *G. turkestanerum* playing hepatoprotective effects against lung injury. In future, further studies are still needed to investigate the exact protective effects of the *G. turkestanerum* extracts and the potential mechanisms.

In conclusion, *G. turkestanerum* played protective effects against CCI_4 induced acute liver injury in mice. The mechanism may be related with the inhibition of lipid peroxidation, and the improved anti-oxidant levels. Moreover, the hepatoprotective effects were correlated with the content of secoiridoides. The protective effects of *G. turkestanerum* against CCI_4 induced acute liver injury were presented in a dosedependent manner, especially the mice treated with GBA (400 mg/Kg). We conlcude that secoiridoides may play an important role in the hepatoprotective effects mediated by *G. turkestanerum*.

Acknowledgements

This study is supported by the National Nature Science Foundation of China (grant no. 8136-0670; grant no. 81560629), Xinjiang Scientific Innovation Program (No. 201472105) and Postgraduate Innovation Program of Xijiang Medical University (No. CXCY062).

Disclosure of conflict of interest

None.

Address correspondence to: Junping Hu, Department of Natural Medicines, College of Pharmacy, Xinjiang Medical University, 393 Xinyi Road, Urumqi 830011, Xinjiang Uyghur Autonomous Region, China. E-mail: hujp0551@yeah.net

References

- [1] Lee CP, Shih PH, Hsu CL and Yen GC. Hepatoprotection of tea seed oil (Camellia oleifera Abel.) against CCl_4 -induced oxidative damage in rats. Food Chem Toxicol 2007; 45: 888-895.
- [2] Jia XY, Zhang QA, Zhang ZQ, Wang Y, Yuan JF, Wang HY and Zhao D. Hepatoprotective effects of almond oil against carbon tetrachloride induced liver injury in rats. Food Chem 2011; 125: 673-678.
- [3] Cheshchevik V, Lapshina E, Dremza I, Zabrodskaya S, Reiter R, Prokopchik N and Zavodnik I. Rat liver mitochondrial damage under acute or chronic carbon tetrachlorideinduced intoxication: protection by melatonin and cranberry flavonoids. Toxicol Appl Pharmacol 2012; 261: 271-279.
- [4] Kumar KS, Liao JW, Xiao JH, Vani MG and Wang SY. Hepatoprotective effect of lucidone against alcohol-induced oxidative stress in human hepatic HepG2 cells through the up-regulation of HO-1/Nrf-2 antioxidant genes. Toxicol In Vitro 2012; 26: 700-708.
- [5] Cao XY and Wang ZZ. Simultaneous determination of four iridoid and secoiridoid glycosides and comparative analysis of radix gentianae macrophyllae and their related substitutes by HPLC. Phytochem Anal 2010; 21: 348-354.
- [6] Mihailović V, Katanić J, Mišić D, Stanković V, Mihailović M, Uskoković A, Arambašić J, Solujić S, Mladenović M and Stanković N. Hepatoprotective effects of secoiridoid-rich extracts from Gentiana cruciata L. against carbon tetrachloride induced liver damage in rats. Food Funct 2014; 5: 1795-1803.
- [7] Mihailović V, Mihailović M, Uskoković A, Arambašić J, Mišić D, Stanković V, Katanić J, Mladenović M, Solujić S and Matić S. Hepatoprotective effects of Gentiana asclepiadea L. extracts against carbon tetrachloride induced liver injury in rats. Food Chem Toxicol 2013; 52: 83-90.
- [8] Zhang ZF, Liu Y, Lu LY, Luo P. Hepatoprotective activity of Gentiana veitchiorum Hemsl. against carbon tetrachloride-induced hepatotoxicity in mice. Chin J Nat Med 2014; 12: 488-494.
- [9] Deng C, Yao N, Wang B and Zhang X. Development of microwave-assisted extraction followed by headspace single-drop microextraction for fast determination of paeonol in tradi-

tional Chinese medicines. J Chromatogr A 2006; 1103: 15-21.

- [10] Cao X and Wang Z. Simultaneous determination of four iridoid and secoiridoid glycosides and comparative analysis of radix gentianae macrophyllae and their related substitutes by HPLC. Phytochem Anal 2010; 21: 348-354.
- [11] Schlede E, Genschow E, Spielmann H, Stropp G and Kayser D. Oral acute toxic class method: a successful alternative to the oral LD 50 test. Regul Toxicol Pharmacol 2005; 42: 15-23.
- [12] Huang B, Ban X, He J, Tong J, Tian J and Wang Y. Hepatoprotective and antioxidant activity of ethanolic extracts of edible lotus (Nelumbo nucifera Gaertn.) leaves. Food Chem 2010; 120: 873-878.
- [13] Srivastava A and Shivanandappa T. Hepatoprotective effect of the root extract of decalepis hamiltonii against carbon tetrachlorideinduced oxidative stress in rats. Food Chem 2010; 118: 411-417.
- [14] Weber L, Boll M and Stampfl A. Hepatotoxicity and mechanism of action of haloalkanes: carbon tetrachloride as a toxicological model. Crit Rev Toxicol 2003; 33: 105-136.
- [15] Poli G, Albano E and Dianzani MU. The role of lipid peroxidation in liver damage. Chem Phys Lipids 1987; 45: 117-142.
- [16] Shyur LF, Huang CC, Lo CP, Chiu CY, Chen YP, Wang SY and Chang ST. Hepatoprotective phytocompounds from Cryptomeria japonica are potent modulators of inflammatory mediators. Phytochemistry 2008; 69: 1348-1358.
- [17] Wu D, Zhai Q and Shi X. Alcohol-induced oxidative stress and cell responses. J Gastroenterol Hepatol 2006; 21: S26-S29.
- [18] Zhou D, Ruan J, Cai Y, Xiong Z, Fu W and Wei A. Antioxidant and hepatoprotective activity of ethanol extract of Arachniodes exilis (Hance) Ching. J Ethnopharmacol 2010; 129: 232-237.
- [19] Jadon A, Bhadauria M and Shukla S. Protective effect of Terminalia belerica Roxb. and gallic acid against carbon tetrachloride induced damage in albino rats. J Ethnopharmacol 2007; 109: 214-218.
- [20] Huang Q, Zhang S, Zheng L, He M, Huang R and Lin X. Hepatoprotective effects of total saponins isolated from Taraphochlamys affinis against carbon tetrachloride induced liver injury in rats. Food Chem Toxicol 2012; 50: 713-718.
- [21] Ozturk IC, Ozturk F, Gul M, Ates B and Cetin A. Protective effects of ascorbic acid on hepatotoxicity and oxidative stress caused by carbon tetrachloride in the liver of Wistar rats. Cell Biochem Funct 2009; 27: 309-315.
- [22] Ploa G and Hewitt W. Principle and methods of toxicology. Wallace Hyes, A.(Ed.) 1989; 2: 399.

- [23] Pareek A, Godavarthi A, Issarani R and Nagori BP. Antioxidant and hepatoprotective activity of Fagonia schweinfurthii (Hadidi) Hadidi extract in carbon tetrachloride induced hepatotoxicity in HepG2 cell line and rats. J Ethnopharmacol 2013; 150: 973-981.
- [24] Huang B, Ban X, He J, Zeng H, Zhang P and Wang Y. Hepatoprotective and antioxidant effects of the methanolic extract from Halenia elliptica. J Ethnopharmacol 2010; 131: 276-281.
- [25] Gravela E, Albano E, Dianzani MU, Poli G and Slater TF. Effects of carbon tetrachloride on isolated rat hepatocytes. Inhibition of protein and lipoprotein secretion. Biochem J 1979; 178: 509-512.
- [26] Clawson GA. Mechanisms of carbon tetrachloride hepatotoxicity. Pathol limmuno Res 1989; 8: 104-112.
- [27] Kumar SS and Mishra S. Hepatoprotective effect of Pergularia daemia (Forsk) ethanol extract and its fraction. Indian J Exp Biol 2008; 46: 447.
- [28] Yu H, Zheng L, Yin L, Xu L, Qi Y, Han X, Xu Y, Liu K and Peng J. Protective effects of the total saponins from dioscorea nipponica makino against carbon tetrachloride-induced liver injury in mice through suppression of apoptosis and inflammation. Int Immunopharmacol 2014; 19: 233-244.
- [29] Li C, Yi LT, Geng D, Han YY and Weng LJ. Hepatoprotective effect of ethanol extract from Berchemia lineate against CCl4-induced acute hepatotoxicity in mice. Pharm Biol 2015; 53: 767-772.
- [30] Vaca C, Wilhelm J and Harms-Ringdahl M. Interaction of lipid peroxidation products with DNA. A review. Mutat Res 1988; 195: 137-149.
- [31] Naik S. Antioxidants and their role in biological functions: An overview. Ind Drugs 2003; 40: 501-516.
- [32] Ohkawa H, Ohishi N and Yagi K. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. Anal Biochem 1979; 95: 351-358.
- [33] Shen B, Chen H, Shen C, Xu P, Li J, Shen G, Yuan H and Han J. Hepatoprotective effects of lignans extract from Herpetospermum caudigerum against CCl₄-induced acute liver injury in mice. J Ethnopharmacol 2015; 164: 46-52.
- [34] Townsend DM, Tew KD and Tapiero H. The importance of glutathione in human disease. Biomed Pharmacother 2003; 57: 145-155.
- [35] Yuan L and Kaplowitz N. Glutathione in liver diseases and hepatotoxicity. Mol Aspects Med 2009; 30: 29-41.

- [36] Ai G, Liu Q, Hua W, Huang Z and Wang D. Hepatoprotective evaluation of the total flavonoids extracted from flowers of Abelmoschus manihot (L.) Medic: in vitro and in vivo studies. J Ethnopharmacol 2013; 146: 794-802.
- [37] Venukumar M and Latha M. Antioxidant activity ofcurculigo orchioides in carbon tetrachloride-induced hepatopathy in rats. Indian J Clin Biochem 2002; 17: 80-87.
- [38] Mustafa AM, Caprioli G, Ricciutelli M, Maggi F, Marín R, Vittori S and Sagratini G. Comparative HPLC/ESI-MS and HPLC/DAD study of different populations of cultivated, wild and commercial gentiana lutea L. Food Chem 2015; 174: 426-433.
- [39] Yang H, Que S, Wu X and Shi Y. Studies on glycosides from gentiana veitchiorum. Zhongguo Zhong Yao Za Zhi 2008; 33: 2505-2507.