# Review Article Hormonal therapy in traumatic spinal cord injury

Parker E Ludwig<sup>1</sup>, Arun A Patil<sup>1,2</sup>, Andrea J Chamczuk<sup>2</sup>, Devendra K Agrawal<sup>1</sup>

Departments of <sup>1</sup>Clinical and Translational Science, <sup>2</sup>Neurosurgery, Creighton University School of Medicine, Omaha, NE, USA

Received May 9, 2017; Accepted August 25, 2017; Epub September 15, 2017; Published September 30, 2017

Abstract: Traumatic spinal cord injuries are major health problems and the underlying pathophysiological events and treatment strategies are currently under investigation. In this article, we critically reviewed the literature investigating the effects of estrogen, progesterone, and human chorionic gonadotropin on spinal cord damage or preservation following traumatic spinal cord injury. The National Library of Medicine database was searched through December 2016 using PubMed for articles addressing the clinical relevance of the hormones to improve neural structural integrity following traumatic spinal cord injury. It was found that each of these hormones, through varied mechanisms, could serve to reduce the harmful effects associated with spinal cord injury, and could aid in restoring some function to the injured spinal cord in the animal models. The most striking effects of human chorionic gonadotropin administration following spinal cord injury have received far less attention than those of either estrogen or progesterone, and additional inquiry could be of general benefit. In this article, we discussed the outstanding questions and suggested future directions for further investigation.

Keywords: Estrogen, progesterone, human chorionic gonadotropin, spinal cord injury, remyelination, inflammation

#### Introduction

Traumatic spinal cord injuries (SCI), both complete and incomplete can have devastating physiological consequences. Depending on the severity of injury, patients incur neurological deficits ranging from paralysis, loss of sensation, impaired bowel, bladder and sexual function, autonomic dysfunction and even death [1-4], and the sequelae of impairment carry with them systemic complications, including impaired wound healing, pneumonia, and ventilator dependence, to name a few. The irreversibility of SCI can be ascribed largely to the relative rarity with which axonal regeneration occurs in the adult spinal cord [5]. This lack of regeneration is attributable to glial scar formation, inflammation and cell death, dominance of inhibitory growth components, and the loss of substrates that support growth [2, 6-8].

The incidence of SCI has been demonstrated, through systematic reviews to be more prevalent in developing countries than in developed nations [9, 10], however, in the United States alone, the staggering incidence of SCI approximates to 40 injuries per million annually or about 12,400 injuries across the general population as reported in 2010 [11]. The occurrence of traumatic SCIs show a bimodal age distribution with one peak being between the ages of 15 and 29 years, and the second being above age 65 years [11, 12]. Injuries occur more often in men. The leading cause of SCI is motor vehicle accidents, followed by falls, violence (particularly gunshot wounds), and sports accidents, in descending order [13, 14]. The pronounced mobility of the cervical spine causes it to be the most commonly damaged spinal region followed by the thoracolumbar junction, which has greater mobility than that of the thoracic spine; the additional stability conferred by the ribs results in fewer traumatic thoracic injuries [10, 12].

The underlying causes of SCI vary dramatically including crush injuries, piercing injuries, vertebral herniation, and gunshot wounds. Many injuries are associated with compression, flexion, extension, distraction, axial loading or rotation of the spinal cord or column.

Hormone	Direct effect	Implications
Estrogen	Microglial activation, increased VEGF expression along with increased blood flow to site of injury, reduced calpain and caspase 3 expression, attenuated cellular calcium influx, decreased TNF- $\alpha$ and iNOS expression	Reduction in neuronal death, anti- apoptotic effect leading the increased cell survival, decreased inflammatory response; overall increase in preser- vation of function following injury
Progesterone	Downregulation of inflammatory cytokines including TNF- $\alpha$ and iNOS, NOS2, MCP-1, and IL-1 $\beta$ ; downregulation of caspase 3 and GFAP	Neuroprotection due to attenuated inflammation and reduction of apop- tosis; improved motor function and increased preservation of neuronal structural and functional integrity
Human chorionic gonadotropin	Reduced lesion volume in experimental stroke models	Potential improvement in functional and structural recovery following spinal cord injury

 Table 1. Effects of hormones in relation to spinal cord injury

In recent years, studies have been conducted on various substances and biomolecules that could be effective in enhancing repair of the spinal cord following trauma. As a result, many promising advances have come forth in understanding the pathophysiological and biological processes involved in SCI and developing potential treatments.

Cell transplantation has been explored as a useful technique in minimizing the damage associated with SCI, and in regenerating the injured spinal cord; astrocyte transplantation has been studied in particular detail [5]. All astrocyte transplantation attempts have not provided significant benefit [15, 16]. However, recent work with embryonic glial-restricted precursor-derived astrocytes (GDAs) has proven to be effective in limiting lesion size, while also preserving white matter [17-24].

Furthermore, there have been promising advances in the delivery methods of cell and biomolecules that could improve tissue repair and regeneration in the central nervous system. Methods include encapsulated cell therapy, the use of implanted scaffolds, and biomolecule delivery in polymeric nano/microspheres and hydrogels [25]. Ji et al. observed that maintaining the integrity of the blood-spinal cord barrier following spinal cord ischemia reperfusion injury could lead to improved outcomes [26, 27].

Potential effects of immune modulatory therapies on SCI have also been examined [28]. The steroid methylprednisolone has been shown to be neuroprotective; it provides functional ben-

efit in SCI through its action on glucocorticoid receptors, its interaction with NF-kB, and its antioxidant activity [29-35]. Inhibiting neutrophil recruitment and adhesion through targeting of the ICAM-1 and/or P-selectin has been observed to improve results in some cases of SCI [36-39], however, one study involving antineutrophil serum did not offer corroboratively compelling results [40]. While macrophage depletion could modestly improve outcomes following SCI [41, 42], there is a relative lack of specific methods for targeting macrophages which complicates the determination that macrophage impairment, and not another mechanism, is responsible. Despite some conflicting results [43-45] with anti-T cell methods, likewise therapies aimed at inhibiting T- and B-cells have been seen to offer some benefit [46-50]. Several anti-inflammatory therapies have been shown to provide benefit following SCI; these include the administration of anti-CD11d monoclonal antibody [51-53], the inhibition of monocytes and neutrophils together (as potential collaborators) [54], and the use of intravenous immunoglobulin [55, 56]. The antibiotic, minocycline, provides neuroprotective benefit through its ability to attenuate inflammation and apoptosis [57-69]. Some plant-derived substances such as allicin and gastrodin have also been observed to mitigate the effects of SCI through anti-inflammatory mechanisms [70, 71]. Current research is attempting to elucidate the potential benefits to be derived from mediators of inflammation.

Hormonal therapies such as those involving the administration of estrogen, progesterone, and



Figure 1. Schematic diagram showing underlying causes of SCI and immune response activation. Some of the major transcription factors involved in inflammatory injury following spinal cord trauma are shown. The complete signaling pathways are not depicted. Causes of SCI vary and include motor vehicle accidents, falls, violent encounters, and sports injuries. SCI is generally marked by an increase in cytokines such as TNF- $\alpha$ , IL-1 $\beta$ , and IL-6 that lead to upregulation of inflammatory and/or apoptotic agents including NF- $\kappa$ B, AP-1, JNK, p38 MAPK, and PGE<sub>2</sub>. The upregulation of these factors often contributes to secondary damage which worsens the outcome following the initial injury, typically presenting as increased lesion size or increased loss of cells.

human chorionic gonadotropin (HCG) have been shown to improve outcomes following SCI (**Table 1**). These hormones act through various mechanisms that will be reviewed in this paper. The use of endogenous hormones as SCI therapy is attractive because associated results and side effects may be limited or more readily anticipated as compared to the use of some exogenous therapies. These hormones are also relatively accessible and inexpensive which improve their potential for wide-spread use.

## Hormonal therapy

Historically, regeneration of peripheral nerves has been considered plausible in certain situations, however, the dogma persists that SCIs are permanently debilitating without chance of recovery. While the reality largely confirms this impression, there are areas of research in spinal cord regeneration that are demonstrating tremendous promise. Areas that have been recently explored include cell transplantation, steroid hormone administration, immune modulatory therapies, and the administration of inflammation suppressants and other biomolecules among others. This review will focus on the hormones, estrogen, progesterone, and HCG and their effects on the injured spinal cord.

## Estrogen

Many pharmacological agents and their effects following traumatic spinal cord injury have been studied [72-75]. Among these, estrogen has been shown to exhibit a neuroprotective effect [76-79]. This effect is a result of anti-inflammatory processes and activation of varied serine proteases by estrogen [76]. Letaif et al. [80] note that this anti-inflammatory activity is perpetrated through microglial activation, increased blood flow to the site of injury, increased levels of anti-apoptotic proteins, attenuated cellular influx of calcium following injury and administration of estrogen [81-83]. In fact, increased estrogen levels may be partially responsible for improved out-

comes in females relative to males following SCI [84].

The overexpression of cytokines has been observed following injury of the central nervous system [85, 86]. Upregulation of genes expressing cytokines such as tumor necrosis factor-a (TNF- $\alpha$ ), interleukin-1 (IL-1), and interleukin-6 (IL-6) has been observed shortly following SCI [87-92] (Figure 1). TNF-α mediates inflammatory processes through its activation of NF-kB, which in turn upregulates other proinflammatory cytokines [93-97]. Pishva et al. [98], determined that administration of estrogen twice daily following SCI in rats significantly reduced the gene expression of TNF- $\alpha$  and its downstream cytokine iNOS and this likely accounts for at least some of the inhibitory effect estrogen has on inflammation following SCI (Figure 1).

While estrogen may not be effective in returning complete function and mobility to an individual following traumatic spinal cord injury, it has been shown to improve functional scores in studies performed in rats [80]. However, between weeks four and six following the induced injury, the group administered a single dose of 17b-estradiol intraperitoneally scored significantly higher than their control counterparts with mean BBB scores being reported as 15.1 for the estradiol group and 9.3 for the control by week 6, which was statistically significant [80]. Hubscher et al. [99] likewise reported improved scores through the sixth



**Figure 2.** Compared to SCI controls, subjects receiving estrogen injections following injury exhibit reduction in TNF- $\alpha$ , increased microglial activity, reduced calpain and caspase 3 levels, increased levels of anti-apoptotic protein, decreased cellular calcium influx, increased serine protease activation, and increased VEGF expression. This, in turn, leads to attenuated inflammation, decreased apoptosis, and increased blood flow to the site of injury, resulting in improved scores on the Basso-Bresnahan-Beattie (BBB) scale for locomotion, improved conduction velocity on motor evoked potential (MEP) monitoring, and increased quantity and diameter of axons.

week of their experiment. Other studies reviewed cover a short period and only provide insight into the initial effects of estrogen following SCI [76]. While Ritz and Hausmann reported lower BBB scores after the fourth week [77], other groups have reported significant improvements in BBB scores in estrogen-treated rats earlier than the fourth week [76, 78]. The conflicting findings underscores the importance of a potential benefit with additional long-term studies extending well beyond the fourth week to clarify our understanding of the effects of estrogen on functional nervous system scores, beyond the short-term. Moreover, some investigators have administered estradiol in multiple doses [76, 78, 83] and observed good cellular and physiological results. Olsen et al. [83] administered estrogen for the first 21 days following SCI. The studies employing multiple dosing schedules [76, 78] reported earlier improvements in BBB scores compared to those that administered a single dose [80] (Figure 2).

In addition, large differences between treatment and control animals have been seen using motor evoked potential monitoring (MEP) [80]. In this test, latency, or the time taken for an electrical impulse to travel from the head to the limbs. and the amplitude of the transmitted impulse were measured. Letaif et al. [80] reported a 17-fold increase in the speed of travel of the electrical impulse in estrogentreated rats as compared to the control, and a 7-fold increase in the amplitude of same impulse. The strictly objective nature of this test makes it a valuable and evaluative tool for measuring neural function. Thus, MEP proves indispensable in future studies involving injuries of the nervous system in evaluating the effect of treatment (Figure 2).

No significant improvement has been observed from a histological perspective in rats treated with estrogen following SCI when analyzed for necrosis, hemorrhage, hyper-

emia, axon degeneration, and cellular infiltration. However, there has been a marked increase in quantity and diameter of axons observed in rats treated with estrogen compared to control [80]. The mean number of fibers for the estradiol group was reported as 92.6 and that of the control group was 56.9 which was statistically significant. Whereas the mean diameter of axons for the estradiol group was 92.4 compared to a mean of 55.1 for the control group, which was again, statistically significant [80].

So far, studies examining the effects of estrogen injection following SCI have administered the treatment directly following the injury. This provides useful insight into the results of early treatment with estrogen. However, it would be of benefit to directly compare the effects of early administration with later administration to ascertain any differences in benefit that exist. In fact, Sribnick et al. found that chronic cases of SCI are also amenable to estrogen therapy and exhibit improvements in motor function following treatment [78]. The effect on chronic cases could be further explored.

It has been shown that some molecules that bind the estrogen receptors such as G-1, tamoxifen, and other estrogen receptor agonists, can have neuroprotective functions similar or identical to those of estrogen following SCI [100-108]. High dose estrogen administration has not become a standard of care for SCI patients in large part due to the adverse effects associated with estrogen levels well above normal physiological levels. These side effects include increased rates of deep vein thrombosis and cancer [109, 110], and the development of feminine physical traits in males. Samantaray et al. [1] reported that low dose estrogen administration attenuates gliosis and provides protection for neurons in the caudal penumbra following traumatic SCI in rats. It was observed that low dose estrogen administration (5-10  $\mu$ g/kg) 48 hours following the injury resulted in the attenuation of inflammatory events, reduced calpain expression which induced an anti-apoptotic effect, reduced caspase-3 expression also inhibiting apoptosis, increased expression of the estrogen receptors ER- $\alpha$  and ER- $\beta$  (suggesting effects relate to increased receptor signaling), attenuated neuronal death, and increased expression of VEGF which is a potent stimulator of vasculogenesis and angiogenesis. Apart from the increased expression of estrogen receptors, the observations failed to exhibit any significant difference between the 10 µg and 100 µg doses of estrogen.

Thus, the research findings to date suggest that low dose estrogen administration and estrogen receptor agonists exhibit potential to be further explored in animals, and in clinical trials in humans following additional knowledge on potentially adverse effects. Estrogen has been shown to ameliorate SCI, but the undesirable effects associated with high-dose estrogen administration limit its potential as a standalone therapy.

## Progesterone

Progesterone (PROG) is a steroid hormone produced by the ovaries and placenta in females, and by the adrenal glands in both females and males. The nervous system has the capacity to locally synthesize PROG and convert PROG into its active metabolite, allopregnanolone [111]. As with estrogen, PROG has been shown to be promyelinating, anti-inflammatory, and neuroprotective in cases of nervous system injury [112-115]. When studied in relation to brain trauma, PROG was shown to prevent neuron loss and mitochondrial dysfunction, reduce edema and inhibit inflammatory cytokines, as well as, improve motor function on diagnostic scales [116-118]. Moreover, PROG has also been tested in two Phase II clinical trials which suggested its efficacy as a treatment option for traumatic brain injury patients [119-121]. With respect to SCI, PROG has been shown to prevent chromatolysis, preserve motor neuron structure, upregulate the expression of choline acetyltransferase and brain-derived neurotrophic factor (BDNF), which increase production of acetylcholine and help to support preservation, growth, and differentiation of neurons, respectively. PROG has also been shown to reduce the proliferation and activation of astrocytes and microglia, and increase the production of oligodendrocyte progenitor cells [113, 122-124]. PROG could be a target for therapies aimed at improving neural function following injury by modulation of astrocytes and their pathogenesis [125]. As was observed with estrogen, 10 µg/kg/12 h PROG administration significantly reduced the expression of TNF- $\alpha$ and iNOS genes following SCI, which lead to production of inflammatory mediators and nitric oxide (NO) which can contribute to reactive radical damage [126]. Garcia-Ovejero et al. [127] observed comparable effects following SCI in rats administered PROG subcutaneously each day. After 60 days, there was a marked increase in spared white matter (SWM) preservation in PROG-treated rats compared to control with white matter measurements of 58.60 ± 4.06% and 22.99 ± 3.03% volume, respectively, as measured 2.5 mm rostrally and caudally from the epicenter of contusion. However, there was no significant difference in the volume of spared gray matter (SGM) observed between control rats and those treated with PROG. Increased oligodendrocytes, decreased myelin damage, improved axonal preservation, and improved locomotor function were all demonstrated following the administration of PROG after SCI.

While another study recapitulated the improved motor and histological outcome with PROG administration [128], the beneficial results are not universal: Fee et al. [129] found no improvement with PROG injection following SCI. Part of this discrepancy could be a result of



**Figure 3.** Compared to SCI controls, subjects receiving progesterone injections following injury exhibit reduced expression of TNF- $\alpha$  and subsequently iNOS, increased acetylcholine production and neuron growth through upregulation of choline acetyltransferase and brain-derived growth factor (BDNF), downregulation of NOS2, MCP-1, IL-1 $\beta$ , caspase-3, and GFAP. Observations have been noted of decreased chromatolysis, increased preservation of neuron structure, and a reduction of mitochondrial dysfunction in animals administered progesterone following SCI. It has been noted that progesterone and its metabolites 5a-dihydroprogesterone and allopregnanolone act on those receptors depicted to contribute to the mediation of the effects listed. In this schematic diagram, complete pathways are not depicted, but some of the major components are shown.

differences in the duration of study, as reported observations in the study of Fee et al. [129] were limited to 5 days as opposed to 60 days and 6 weeks in the studies producing positive results.

Guennoun et al. [111] found the binding of progesterone to intracellular progesterone receptors (PR) via its classical pathway, and it may also bind to specific membrane sites (mPR/ PGRMC1 complex) to activate intracellular signaling pathways (Figure 3). Moreover, allopregnanolone may act on GABA, receptors following its conversion from PROG [111, 130-135]. They also determined that PROG acted on the effectors PR, mPRs, and PGRMC1. After conversion to 5a-dihydroprogesterone through a mechanism involving 5a-reductases, action occurred on the effectors PR and mPRa (Figure 3). Following conversion via 3a-HSOR to allopregnanolone (3a5a-THPROG), it can bind to GABA, receptors, PXR, and mPRd to induce neuroprotective effects [111] (Figure 3). Further understanding of the receptors upon which PROG and related molecules act could prove useful in developing treatments aimed at neuroprotection. Future work is required to ascertain potential side effects associated with synthetic progestins and allopregnanolone as many synthetic progestins may also bind androgen and glucocorticoid receptors producing undesirable effects [136, 137], and allopregnanolone has been associated with some cognitive impairment and symptoms such as anxiety, irritability, aggressiveness, seizure, and increased pain [138-142]. Of note, it has been determined that PR reduces reactive gliosis and preserves oligodendrocyte precursor cells in the injured spinal cord in rats [143].

PROG shows potential as a modulator of neuropathic pain following SCI [144-146]. While there may be many mechanisms by which pain is transmitted following central nervous system injury [147, 148], neuro-inflammation and reactive gliosis are primary

causes of chronic pain following SCI [149-151]. Cytokines are involved in the modulation of neuronal function and pain transmission [151-156]. Coronel et al. [144] explored the effects of PROG on IL-1ß and its receptors IL-1RI and IL-1RII, antagonist IL-1ra, IL-6, TNF-α, and NR1 subunit of N-methyl-D-aspartate receptor (NM-DAR) following SCI. IL-1 $\beta$ , IL-6, and TNF- $\alpha$  mRNA (and protein) levels were significantly lower in rats receiving PROG than the placebo, on the first day status post injury: however, there was only a significant difference in mRNA levels observed on day 14 in IL-1β, and no significant difference on day 28 of the study. There was no significant difference in IL-1RI mRNA levels between the groups on days 1 or 14, but by day 28, the PROG-treated group had significantly lower levels than the placebo. IL-1RII mRNA levels were seen to be markedly higher in the PROG group on day one, with no significant difference thereafter. No differences were observed between the two groups in IL-1ra mRNA levels. These data as well as those showing fewer IL-1RI positive neurons in the spinal cords of PROG-treated rats compared to those receiving placebo suggest that PROG may provide benefit to those experiencing chronic pain following SCI, but may not have the same effect or mediate it in the same way in the immediate aftermath of the injury. Additional research on the effect of PROG following SCI, with a focus on IL-1RI, and its utility as a treatment option in cases of chronic pain following SCI, is warranted. There is indication that the effect of PROG could differ in acute and chronic settings.

Yang et al. [157] observed that PROG significantly reduces axonal dieback and neuronal death in mice following SCI when observed at intervals of 24 h, 48 h, and 72 h from the initial injury. This was mediated via down-regulation of inflammatory cytokines, including NOS2, MCP-1, and IL-1 $\beta$  as well as activated caspase-3 and GFAP. Interestingly, upregulation of myelin basic protein (MBP) was also noted. It was also suggested that PROG improved behavioral function following SCI. Further studies of a longer duration would help elucidate the effects of PROG on axon and neuron preservation beyond the acute stages.

PROG is effective as an anti-inflammatory agent that can improve motor function and histological outcomes following SCI. To date, the side-effects associated with PROG are not as immediately apprehensible as those of some others, and it continues to exhibit potential as an effective treatment following injury of the central nervous system.

## HCG

Human chorionic gonadotropin (HCG) is a heterodimer consisting of an  $\alpha$  and a  $\beta$  subunit, which are non-covalently linked. The hCGa subunit constitutes part of other hormones such as luteinizing hormone, follicle-stimulating hormone, and thyroid-stimulating hormone. HCG is produced in the adrenal pituitary gland by gonadotropin cells [158, 159].

HCG presents potential as a treatment in the case of central nervous system injury. It was observed in a preliminary study that HCG helped to increase the amount of adrenal medulla tissue survival following autologous transplant in the lateral ventricles of rat brains [160]. Proliferation of endogenous stem cells in the subgranular zone and the subventricular zone has been observed to increase with administration of HCG [161]. Meng et al. [162] observed that HCG induced neuronal differentiation of PC12 cells by activating the stably expressed lutropin/choriogonadotropin receptor in vitro. In a study of rats with experimental strokes, treatment with HCG + erythropoietin

(EPO) significantly reduced the lesion volumes (by 82-89%) and significantly improved neurological scores compared to three other treatment groups including HCG + saline, saline + EPO, and saline + saline [163]. This would also suggest that EPO may play a role in the reduction of lesion size following stroke.

Extensive literature reviews illustrate a relative void in research performed using HCG in association with SCI as compared with studies using estrogen and progesterone. Patil and Nagaraj [164] found that 12 rats receiving HCG injections following SCI exhibited a significant improvement in functional recovery (assessed by measurement and grading of the return of bladder function and the ability to climb up an inclined plane) within 6 weeks as compared to 10 rats serving as control. In a later study, Patil et al. [165] transected the spinal cords of 21 rats at the midthoracic level; the 11 rats administered HCG exhibited significantly increased amplitudes of the cortical evoked motor action potentials after six weeks compared to the control group. These findings suggest that the administration of HCG may serve to improve spinal cord function following traumatic injury. However, the relatively small number of animals studied and the limited number of retrievable studies of the effects of HCG on individuals with SCI preclude any conclusive determinations of universal effectiveness or possible side effects until further research is undertaken.

HCG may act to improve motor function and minimize neuronal damage following spinal cord lesion. The minimal side effects associated with HCG, and the relative lack of research that has been performed on its effect on SCI mandate further investigation as a potential therapeutic option.

## Conclusion

Estrogen, progesterone, and HCG are hormones with diverse functions that could serve to attenuate the harmful consequences of SCI through their abilities to interact with the GABA system, reduce excitotoxicity, free radicals, edema, and apoptosis, inhibit inflammatory cytokines, induce increased angiogenesis, mitochondrial recoupling, remyelination [166], and induce stem cell migration to the site of injury. These functions have potential to improve prognoses for individuals suffering after SCI with promise of increased motor function, preserved structure, and a reduction of neuropathic pain. Further investigation into each of these methods is of paramount importance.

### Acknowledgements

This work was supported by research grants R01 HL112597, R01 HL116042, and R01 HL120659 to DK Agrawal from the National Heart, Lung and Blood Institute, National Institutes of Health, USA. The content of this review article is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

### Disclosure of conflict of interest

None.

Address correspondence to: Dr. Devendra K Agrawal, Department of Clinical and Translational Science, Creighton University School of Medicine, CRISS II Room 510, 2500 California Plaza, Omaha, NE, 68178, USA. Tel: (402) 280-2938; Fax: (402) 280-1421; E-mail: dkagr@creighton.edu

#### References

- [1] Samantaray S, Das A, Matzelle DC, Yu SP, Wei L, Varma A, Ray SK, Banik NL. Administration of low dose estrogen attenuates gliosis and protects neurons in acute spinal cord injury in rats. J Neurochem 2016; 136: 1064-1073.
- [2] Cao HQ, Dong ED. An update on spinal cord injury research. Neurosci Bull 2013; 29: 94-102.
- [3] Donovan WH. Spinal cord injury-past, present, and future. J Spinal Cord Med 2007; 30: 85-100.
- [4] Pickelsimer E, Shiroma EJ, Wilson DA. Statewide investigation of medically attended adverse health conditions of persons with spinal cord injury. J Spinal Cord Med 2010; 33: 221-231.
- [5] Chu T, Zhou H, Li F, Wang T, Lu L, Feng S. Astrocyte transplantation for spinal cord injury: current status and perspective. Brain Res Bull 2014; 107: 18-30.
- [6] Maier IC, Schwab ME. Sprouting, regeneration and circuit formation in the injured spinal cord: factors and activity. Philos Trans R Soc Lond B Biol Sci 2006; 361: 1611-1634.
- [7] Oyinbo CA. Secondary injury mechanisms in traumatic spinal cord injury: a nugget of this

multiply cascade. Acta Neurobiol Exp (Warsz) 2011; 71: 281-299.

- [8] Smith GM, Miller RH, Silver J. Changing role of forebrain astrocytes during development, regenerative failure, and induced regeneration upon transplantation. J Comp Neurol 1986; 251: 23-43.
- [9] Chiu WT, Lin HC, Lam C, Chu SF, Chiang YH, Tsai SH. Review paper: epidemiology of traumatic spinal cord injury: comparisons between developed and developing countries. Asia Pac J Public Health 2010; 22: 9-18.
- [10] van den Berg MEL, Castellote JM, Mahillo-Fernandez I, de Pedro-Cuesta J. Incidence of spinal cord injury worldwide: a systematic review. Neuroepidemiology 2010; 34: 184-192; discussion 192.
- [11] Devivo MJ. Epidemiology of traumatic spinal cord injury: trends and future implications. Spinal Cord 2012; 50: 365-372.
- [12] Wolf M, Weber MA. Neuroimaging of the traumatic spine. Magn Reson Imaging Clin N Am 2016; 24: 541-561.
- [13] NSCISC National Spinal Cord Injury Statistical Center: Spinal Cord Injury Model Systems: 2014 Annual Report. 2015.
- [14] Jakoi A, Iorio J, Howell R, Zampini JM. Gunshot injuries of the spine. Spine J 2015; 15: 2077-2085.
- [15] Olby NJ, Blakemore WF. Reconstruction of the glial environment of a photochemically induced lesion in the rat spinal cord by transplantation of mixed glial cells. J Neurocytol 1996; 25: 481-498.
- [16] Hayashi K, Hashimoto M, Koda M, Naito AT, Murata A, Okawa A, Takahashi K, Yamazaki M. Increase of sensitivity to mechanical stimulus after transplantation of murine induced pluripotent stem cell-derived astrocytes in a rat spinal cord injury model. J Neurosurg Spine 2011; 15: 582-593.
- [17] Davies JE, Huang C, Proschel C, Noble M, Mayer-Proschel M, Davies SJ. Astrocytes derived from glial-restricted precursors promote spinal cord repair. J Biol 2006; 5: 7.
- [18] Davies JE, Pröschel C, Zhang N, Noble M, Mayer-Pröschel M, Davies SJ. Transplanted astrocytes derived from BMP- or CNTF-treated glialrestricted precursors have opposite effects on recovery and allodynia after spinal cord injury. J Biol 2008; 7: 24.
- [19] Davies SJ, Shih CH, Noble M, Mayer-Proschel M, Davies JE, Proschel C. Transplantation of specific human astrocytes promotes functional recovery after spinal cord injury. PLoS One 2011; 6: e17328.
- [20] Jin Y, Neuhuber B, Singh A, Bouyer J, Lepore A, Bonner J, Himes T, Campanelli JT, Fischer I. Transplantation of human glial restricted pro-

genitors and derived astrocytes into a contusion model of spinal cord injury. J Neurotrauma 2011; 28: 579-594.

- [21] Haas C, Neuhuber B, Yamagami T, Rao M, Fischer I. Phenotypic analysis of astrocytes derived from glial restricted precursors and their impact on axon regeneration. Exp Neurol 2012; 233: 717-732.
- [22] Haas C, Fischer I. Human astrocytes derived from glial restricted progenitors support regeneration of the injured spinal cord. J Neurotrauma 2013; 30: 1035-1052.
- [23] Fan C, Zheng Y, Cheng X, Qi X, Bu P, Luo X, Kim DH, Cao Q. Transplantation of D15A-expressing glial-restricted-precursor-derived astrocytes improves anatomical and locomotor recovery after spinal cord injury. Int J Biol Sci 2013; 9: 78-93.
- [24] Wu L, Li J, Chen L, Zhang H, Yuan L, Davies SJ. Combined transplantation of GDAsBMP and hr-decorin in spinal cord contusion repair. Neural Regen Res 2013; 8: 2236-2248.
- [25] Elliott Donaghue I, Tam R, Sefton MV, Shoichet MS. Cell and biomolecule delivery for tissue repair and regeneration in the central nervous system. J Control Release Off J Control Release Soc 2014; 190: 219-227.
- [26] Hu J, Yu Q, Xie L, Zhu H. Targeting the bloodspinal cord barrier: a therapeutic approach to spinal cord protection against ischemia-reperfusion injury. Life Sci 2016; 158: 1-6.
- [27] Yu Q, Huang J, Hu J, Zhu H. Advance in spinal cord ischemia reperfusion injury: Blood-spinal cord barrier and remote ischemic preconditioning. Life Sci 2016; 154: 34-38.
- [28] Plemel JR, Wee Yong V, Stirling DP. Immune modulatory therapies for spinal cord injurypast, present and future. Exp Neurol 2014; 258: 91-104.
- [29] Hall ED, Springer JE. Neuroprotection and acute spinal cord injury: a reappraisal. NeuroRx J Am Soc Exp Neurother 2004; 1: 80-100.
- [30] Coutinho AE, Chapman KE. The anti-inflammatory and immunosuppressive effects of glucocorticoids, recent developments and mechanistic insights. Mol Cell Endocrinol 2011; 335: 2-13.
- [31] Rhen T, Cidlowski JA. Antiinflammatory action of glucocorticoids--new mechanisms for old drugs. N Engl J Med 2005; 353: 1711-1723.
- [32] De Bosscher K, Vanden Berghe W, Haegeman G. The interplay between the glucocorticoid receptor and nuclear factor-kappaB or activator protein-1: molecular mechanisms for gene repression. Endocr Rev 2003; 24: 488-522.
- [33] McKay LI, Cidlowski JA. Molecular control of immune/inflammatory responses: interactions between nuclear factor-kappa B and steroid receptor-signaling pathways. Endocr Rev 1999; 20: 435-459.

- [34] Braughler JM, Hall ED. Correlation of methylprednisolone levels in cat spinal cord with its effects on (Na+ + K+)-ATPase, lipid peroxidation, and alpha motor neuron function. J Neurosurg 1982; 56: 838-844.
- [35] Hall ED, Braughler JM. Acute effects of intravenous glucocorticoid pretreatment on the in vitro peroxidation of cat spinal cord tissue. Exp Neurol 1981; 73: 321-324.
- [36] Hamada Y, Ikata T, Katoh S, Nakauchi K, Niwa M, Kawai Y, Fukuzawa K. Involvement of an intercellular adhesion molecule 1-dependent pathway in the pathogenesis of secondary changes after spinal cord injury in rats. J Neurochem 1996; 66: 1525-1531.
- [37] Taoka Y, Okajima K, Uchiba M, Murakami K, Kushimoto S, Johno M, Naruo M, Okabe H, Takatsuki K. Role of neutrophils in spinal cord injury in the rat. Neuroscience 1997; 79: 1177-1182.
- [38] Farooque M, Isaksson J, Olsson Y. Improved recovery after spinal cord trauma in ICAM-1 and P-selectin knockout mice. Neuroreport 1999; 10: 131-134.
- [39] Farooque M, Isaksson J, Olsson Y. White matter preservation after spinal cord injury in ICAM-1/P-selectin-deficient mice. Acta Neuropathol (Berl) 2001; 102: 132-140.
- [40] Holtz A, Nyström B, Gerdin B. Spinal cord injury in rats: inability of nimodipine or anti-neutrophil serum to improve spinal cord blood flow or neurologic status. Acta Neurol Scand 1989; 79: 460-467.
- [41] Iannotti CA, Clark M, Horn KP, van Rooijen N, Silver J, Steinmetz MP. A combination immunomodulatory treatment promotes neuroprotection and locomotor recovery after contusion SCI. Exp Neurol 2011; 230: 3-15.
- [42] Horn KP, Busch SA, Hawthorne AL, van Rooijen N, Silver J. Another barrier to regeneration in the CNS: activated macrophages induce extensive retraction of dystrophic axons through direct physical interactions. J Neurosci Off J Soc Neurosci 2008; 28: 9330-9341.
- [43] Hauben E, Agranov E, Gothilf A, Nevo U, Cohen A, Smirnov I, Steinman L, Schwartz M. Posttraumatic therapeutic vaccination with modified myelin self-antigen prevents complete paralysis while avoiding autoimmune disease. J Clin Invest 2001; 108: 591-599.
- [44] Hauben E, Butovsky O, Nevo U, Yoles E, Moalem G, Agranov E, Mor F, Leibowitz-Amit R, Pevsner E, Akselrod S, Neeman M, Cohen IR, Schwartz M. Passive or active immunization with myelin basic protein promotes recovery from spinal cord contusion. J Neurosci Off J Soc Neurosci 2000; 20: 6421-6430.
- [45] Hauben E, Nevo U, Yoles E, Moalem G, Agranov E, Mor F, Akselrod S, Neeman M, Cohen IR,

Schwartz M. Autoimmune T cells as potential neuroprotective therapy for spinal cord injury. Lancet Lond Engl 2000; 355: 286-287.

- [46] Luchetti S, Beck KD, Galvan MD, Silva R, Cummings BJ, Anderson AJ. Comparison of immunopathology and locomotor recovery in C57BL/6, BUB/BnJ, and NOD-SCID mice after contusion spinal cord injury. J Neurotrauma 2010; 27: 411-421.
- [47] Wu B, Matic D, Djogo N, Szpotowicz E, Schachner M, Jakovcevski I. Improved regeneration after spinal cord injury in mice lacking functional T- and B-lymphocytes. Exp Neurol 2012; 237: 274-285.
- [48] Gonzalez R, Glaser J, Liu MT, Lane TE, Keirstead HS. Reducing inflammation decreases secondary degeneration and functional deficit after spinal cord injury. Exp Neurol 2003; 184: 456-463.
- [49] Ankeny DP, Guan Z, Popovich PG. B cells produce pathogenic antibodies and impair recovery after spinal cord injury in mice. J Clin Invest 2009; 119: 2990-2999.
- [50] Ankeny DP, Lucin KM, Sanders VM, McGaughy VM, Popovich PG. Spinal cord injury triggers systemic autoimmunity: evidence for chronic B lymphocyte activation and lupus-like autoantibody synthesis. J Neurochem 2006; 99: 1073-1087.
- [51] Ditor DS, Bao F, Chen Y, Dekaban GA, Weaver LC. A therapeutic time window for anti-CD 11d monoclonal antibody treatment yielding reduced secondary tissue damage and enhanced behavioral recovery following severe spinal cord injury. J Neurosurg Spine 2006; 5: 343-352.
- [52] Gris D, Marsh DR, Oatway MA, Chen Y, Hamilton EF, Dekaban GA, Weaver LC. Transient blockade of the CD11d/CD18 integrin reduces secondary damage after spinal cord injury, improving sensory, autonomic, and motor function. J Neurosci 2004; 24: 4043-4051.
- [53] Oatway MA, Chen Y, Bruce JC, Dekaban GA, Weaver LC. Anti-CD11d integrin antibody treatment restores normal serotonergic projections to the dorsal, intermediate, and ventral horns of the injured spinal cord. J Neurosci 2005; 25: 637-647.
- [54] Lee SM, Rosen S, Weinstein P, van Rooijen N, Noble-Haeusslein LJ. Prevention of both neutrophil and monocyte recruitment promotes recovery after spinal cord injury. J Neurotrauma 2011; 28: 1893-1907.
- [55] Gok B, Sciubba DM, Okutan O, Beskonakli E, Palaoglu S, Erdamar H, Sargon MF. Immunomodulation of acute experimental spinal cord injury with human immunoglobulin G. J Clin Neurosci 2009; 16: 549-553.
- [56] Nguyen DH, Cho N, Satkunendrarajah K, Austin JW, Wang J, Fehlings MG. Immunoglobulin

G (IgG) attenuates neuroinflammation and improves neurobehavioral recovery after cervical spinal cord injury. J Neuroinflammation 2012; 9: 224.

- [57] Stirling DP, Koochesfahani KM, Steeves JD, Tetzlaff W. Minocycline as a neuroprotective agent. Neuroscientist 2005; 11: 308-322.
- [58] Lee SM, Yune TY, Kim SJ, Kim YC, Oh YJ, Markelonis GJ, Oh TH. Minocycline inhibits apoptotic cell death via attenuation of TNF-alpha expression following iNOS/NO induction by lipopolysaccharide in neuron/glia co-cultures. J Neurochem 2004; 91: 568-578.
- [59] Lee SM, Yune TY, Kim SJ, Park DW, Lee YK, Kim YC, Oh YJ, Markelonis GJ, Oh TH. Minocycline reduces cell death and improves functional recovery after traumatic spinal cord injury in the rat. J Neurotrauma 2003; 20: 1017-1027.
- [60] Sanchez Mejia RO, Ona VO, Li M, Friedlander RM. Minocycline reduces traumatic brain injury-mediated caspase-1 activation, tissue damage, and neurological dysfunction. Neurosurgery 2001; 48: 1393-1399-1401.
- [61] Yrjänheikki J, Keinänen R, Pellikka M, Hökfelt T, Koistinaho J. Tetracyclines inhibit microglial activation and are neuroprotective in global brain ischemia. Proc Natl Acad Sci U S A 1998; 95: 15769-15774.
- [62] Yrjänheikki J, Tikka T, Keinänen R, Goldsteins G, Chan PH, Koistinaho J. A tetracycline derivative, minocycline, reduces inflammation and protects against focal cerebral ischemia with a wide therapeutic window. Proc Natl Acad Sci U S A 1999; 96: 13496-13500.
- [63] Kim SS, Kong PJ, Kim BS, Sheen DH, Nam SY, Chun W. Inhibitory action of minocycline on lipopolysaccharide-induced release of nitric oxide and prostaglandin E2 in BV2 microglial cells. Arch Pharm Res 2004; 27: 314-318.
- [64] Kremlev SG, Roberts RL, Palmer C. Differential expression of chemokines and chemokine receptors during microglial activation and inhibition. J Neuroimmunol 2004; 149: 1-9.
- [65] Kobayashi K, Imagama S, Ohgomori T, Hirano K, Uchimura K, Sakamoto K, Hirakawa A, Takeuchi H, Suzumura A, Ishiguro N, Kadomatsu K. Minocycline selectively inhibits M1 polarization of microglia. Cell Death Dis 2013; 4: e525.
- [66] Suk K. Minocycline suppresses hypoxic activation of rodent microglia in culture. Neurosci Lett 2004; 366: 167-171.
- [67] Tikka T, Fiebich BL, Goldsteins G, Keinanen R, Koistinaho J. Minocycline, a tetracycline derivative, is neuroprotective against excitotoxicity by inhibiting activation and proliferation of microglia. J Neurosci 2001; 21: 2580-2588.
- [68] Zhu S, Stavrovskaya IG, Drozda M, Kim BYS, Ona V, Li M, Sarang S, Liu AS, Hartley DM, Wu

DC, Gullans S, Ferrante RJ, Przedborski S, Kristal BS, Friedlander RM. Minocycline inhibits cytochrome c release and delays progression of amyotrophic lateral sclerosis in mice. Nature 2002; 417: 74-78.

- [69] Wang J, Wei Q, Wang CY, Hill WD, Hess DC, Dong Z. Minocycline up-regulates Bcl-2 and protects against cell death in mitochondria. J Biol Chem 2004; 279: 19948-19954.
- [70] Du F, Wang X, Shang B, Fang J, Xi Y, Li A, Diao Y. Gastrodin ameliorates spinal cord injury via antioxidant and anti-inflammatory effects. Acta Biochim Pol 2016; 63: 589-593.
- [71] Wang S, Ren D. Allicin protects traumatic spinal cord injury through regulating the HSP70/ Akt/iNOS pathway in mice. Mol Med Rep 2016; 14: 3086-3092.
- [72] Cristante AF, de Barros Filho TEP, Marcon RM, Letaif OB, da Rocha ID. Therapeutic approaches for spinal cord injury. Clinics 2012; 67: 1219-1224.
- [73] Cristante AF, Filho TEPB, Oliveira RP, Marcon RM, Ferreira R, Santos GB. Effects of antidepressant and treadmill gait training on recovery from spinal cord injury in rats. Spinal Cord 2013; 51: 501-507.
- [74] Marcon RM, Cristante AF, de Barros Filho TEP, de Oliveira RP, dos Santos GB. Potentializing the effects of GM1 by hyperbaric oxygen therapy in acute experimental spinal cord lesion in rats. Spinal Cord 2010; 48: 808-813.
- [75] Tator CH. Review of treatment trials in human spinal cord injury: issues, difficulties, and recommendations. Neurosurgery 2006; 59: 957-982-987.
- [76] Sribnick EA, Wingrave JM, Matzelle DD, Wilford GG, Ray SK, Banik NL. Estrogen attenuated markers of inflammation and decreased lesion volume in acute spinal cord injury in rats. J Neurosci Res 2005; 82: 283-293.
- [77] Ritz MF, Hausmann ON. Effect of 17beta-estradiol on functional outcome, release of cytokines, astrocyte reactivity and inflammatory spreading after spinal cord injury in male rats. Brain Res 2008; 1203: 177-188.
- [78] Sribnick EA, Samantaray S, Das A, Smith J, Matzelle DD, Ray SK, Banik NL. Postinjury estrogen treatment of chronic spinal cord injury improves locomotor function in rats. J Neurosci Res 2010; 88: 1738-1750.
- [79] Yune TY, Kim SJ, Lee SM, Lee YK, Oh YJ, Kim YC, Markelonis GJ, Oh TH. Systemic administration of 17beta-estradiol reduces apoptotic cell death and improves functional recovery following traumatic spinal cord injury in rats. J Neurotrauma 2004; 21: 293-306.
- [80] Letaif OB, Cristante AF, de Barros Filho TEP, Ferreira R, dos Santos GB, da Rocha ID, Marcon RM. Effects of estrogen on functional and

neurological recovery after spinal cord injury: an experimental study with rats. Clinics 2015; 70: 700-705.

- [81] Sribnick EA, Wingrave JM, Matzelle DD, Ray SK, Banik NL. Estrogen as a neuroprotective agent in the treatment of spinal cord injury. Ann N Y Acad Sci 2003; 993: 125-133-160.
- [82] Brann DW, Dhandapani K, Wakade C, Mahesh VB, Khan MM. Neurotrophic and neuroprotective actions of estrogen: basic mechanisms and clinical implications. Steroids 2007; 72: 381-405.
- [83] Olsen ML, Campbell SC, McFerrin MB, Floyd CL, Sontheimer H. Spinal cord injury causes a wide-spread, persistent loss of Kir4.1 and glutamate transporter 1: benefit of 17β-oestradiol treatment. Brain 2010; 133: 1013-1025.
- [84] Datto JP, Yang J, Dietrich WD, Pearse DD. Does being female provide a neuroprotective advantage following spinal cord injury? Neural Regen Res 2015; 10: 1533-1536.
- [85] Hsu CY, Doster K, Hu ZY. Cell-mediated injury. In: Narayan K, Wilberger JE Jr, Polishock PT, editors. Neurotrauma: a comprehensive textbook of head and spinal cord injury. St. Louis, MO: McGraw-Hill; 1996: p.1433-44. In: no date.
- [86] Rothwell NJ, Luheshi G, Toulmond S. Cytokines and their receptors in the central nervous system: physiology, pharmacology, and pathology. Pharmacol Ther 1996; 69: 85-95.
- [87] Bartholdi D, Schwab ME. Expression of pro-inflammatory cytokine and chemokine mRNA upon experimental spinal cord injury in mouse: an in situ hybridization study. Eur J Neurosci 1997; 9: 1422-1438.
- [88] Hayashi M, Ueyama T, Nemoto K, Tamaki T, Senba E. Sequential mRNA expression for immediate early genes, cytokines, and neurotrophins in spinal cord injury. J Neurotrauma 2000; 17: 203-218.
- [89] Wang CX, Olschowka JA, Wrathall JR. Increase of interleukin-1beta mRNA and protein in the spinal cord following experimental traumatic injury in the rat. Brain Res 1997; 759: 190-196.
- [90] Wang CX, Nuttin B, Heremans H, Dom R, Gybels J. Production of tumor necrosis factor in spinal cord following traumatic injury in rats. J Neuroimmunol 1996; 69: 151-156.
- [91] Yakovlev AG, Faden AI. Sequential expression of c-fos protooncogene, TNF-alpha, and dynorphin genes in spinal cord following experimental traumatic injury. Mol Chem Neuropathol 1994; 23: 179-190.
- [92] Streit WJ, Semple-Rowland SL, Hurley SD, Miller RC, Popovich PG, Stokes BT. Cytokine mRNA profiles in contused spinal cord and axotomized facial nucleus suggest a beneficial role for inflammation and gliosis. Exp Neurol 1998; 152: 74-87.

- [93] Schreck R, Rieber P, Baeuerle PA. Reactive oxygen intermediates as apparently widely used messengers in the activation of the NF-kappa B transcription factor and HIV-1. EMBO J 1991; 10: 2247-2258.
- [94] Scheinman RI, Gualberto A, Jewell CM, Cidlowski JA, Baldwin AS. Characterization of mechanisms involved in transrepression of NF-kappa B by activated glucocorticoid receptors. Mol Cell Biol 1995; 15: 943-953.
- [95] Mukaida N, Morita M, Ishikawa Y, Rice N, Okamoto S, Kasahara T, Matsushima K. Novel mechanism of glucocorticoid-mediated gene repression. Nuclear factor-kappa B is target for glucocorticoid-mediated interleukin 8 gene repression. J Biol Chem 1994; 269: 13289-13295.
- [96] Grell M, Zimmermann G, Hülser D, Pfizenmaier K, Scheurich P. TNF receptors TR60 and TR80 can mediate apoptosis via induction of distinct signal pathways. J Immunol Baltim Md 1950 1994; 153: 1963-1972.
- [97] Caldenhoven E, Liden J, Wissink S, Van de Stolpe A, Raaijmakers J, Koenderman L, Okret S, Gustafsson JA, Van der Saag PT. Negative cross-talk between RelA and the glucocorticoid receptor: a possible mechanism for the antiinflammatory action of glucocorticoids. Mol Endocrinol Baltim Md 1995; 9: 401-412.
- [98] Amini Pishva A, Akbari M, Farahabadi A, Arabkheradmand A, Beyer C, Dashti N, Moradi F, Hassanzadeh G. Effect of estrogen therapy on TNF- $\alpha$  and iNOS Gene Expression in Spinal Cord Injury Model. Acta Med Iran 2016; 54: 296-301.
- [99] Hubscher CH, Fell JD, Gupta DS. Sex and hormonal variations in the development of At-level Allodynia In a Rat chronic spinal cord injury model. Neurosci Lett 2010; 477: 153-156.
- [100] Cheng Q, Meng J, Wang X, Kang W, Tian Z, Zhang K, Liu G, Zhao J. G-1 exerts neuroprotective effects through G protein-coupled estrogen receptor 1 following spinal cord injury in mice. Biosci Rep 2016; 36.
- [101] Chen J, Hu R, Ge H, Duanmu W, Li Y, Xue X, Hu S, Feng H. G-protein-coupled receptor 30-mediated antiapoptotic effect of estrogen on spinal motor neurons following injury and its underlying mechanisms. Mol Med Rep 2015; 12: 1733-1740.
- [102] Colón JM, Miranda JD. Tamoxifen: an FDA approved drug with neuroprotective effects for spinal cord injury recovery. Neural Regen Res 2016; 11: 1208-1211.
- [103] Ray SK, Samntaray S, Banik NL. Future directions for using estrogen receptor agonists in the treatment of acute and chronic spinal cord injury. Neural Regen Res 2016; 11: 1418-1419.

- [104] Chakrabarti M, Haque A, Banik NL, Nagarkatti P, Nagarkatti M, Ray SK. Estrogen receptor agonists for attenuation of neuroinflammation and neurodegeneration. Brain Res Bull 2014; 109: 22-31.
- [105] Guptarak J, Wiktorowicz JE, Sadygov RG, Zivadinovic D, Paulucci-Holthauzen AA, Vergara L, Nesic O. The cancer drug tamoxifen: a potential therapeutic treatment for spinal cord injury. J Neurotrauma 2014; 31: 268-283.
- [106] Mosquera L, Colón JM, Santiago JM, Torrado Al, Meléndez M, Segarra AC, Rodríguez-Orengo JF, Miranda JD. Tamoxifen and estradiol improved locomotor function and increased spared tissue in rats after spinal cord injury: their antioxidant effect and role of estrogen receptor alpha. Brain Res 2014; 1561: 11-22.
- [107] Colón JM, Torrado AI, Cajigas Á, Santiago JM, Salgado IK, Arroyo Y, Miranda JD. Tamoxifen administration immediately or 24 hours after spinal cord injury improves locomotor recovery and reduces secondary damage in female rats. J Neurotrauma 2016; 33: 1696-1708.
- [108] Li X, Yang Q, Li X, Cheng Q, Zhang K, Han J, Zhao J, Liu G, Zhao M. Estrogen-like neuroprotection of isopsoralen against spinal cord injury through estrogen receptor ER $\alpha$ . Metab Brain Dis 2017; 32: 259-265.
- [109] Manson JE, Chlebowski RT, Stefanick ML, Aragaki AK, Rossouw JE, Prentice RL, Anderson G, Howard BV, Thomson CA, LaCroix AZ, Wactawski-Wende J, Jackson RD, Limacher M, Margolis KL, Wassertheil-Smoller S, Beresford SA, Cauley JA, Eaton CB, Gass M, Hsia J, Johnson KC, Kooperberg C, Kuller LH, Lewis CE, Liu S, Martin LW, Ockene JK, O'Sullivan MJ, Powell LH, Simon MS, Van Horn L, Vitolins MZ, Wallace RB. Menopausal hormone therapy and health outcomes during the intervention and extended poststopping phases of the Women's Health Initiative randomized trials. JAMA 2013; 310: 1353-1368.
- [110] Nelson HD, Humphrey LL, Nygren P, Teutsch SM, Allan JD. Postmenopausal hormone replacement therapy: scientific review. JAMA 2002; 288: 872-881.
- [111] Guennoun R, Labombarda F, Gonzalez Deniselle MC, Liere P, De Nicola AF, Schumacher M. Progesterone and allopregnanolone in the central nervous system: response to injury and implication for neuroprotection. J Steroid Biochem Mol Biol 2015; 146: 48-61.
- [112] Ciriza I, Carrero P, Frye CA, Garcia-Segura LM. Reduced metabolites mediate neuroprotective effects of progesterone in the adult rat hippocampus. The synthetic progestin medroxyprogesterone acetate (Provera) is not neuroprotective. J Neurobiol 2006; 66: 916-928.

- [113] De Nicola AF, Labombarda F, Gonzalez Deniselle MC, Gonzalez SL, Garay L, Meyer M, Gargiulo G, Guennoun R, Schumacher M. Progesterone neuroprotection in traumatic CNS injury and motoneuron degeneration. Front Neuroendocrinol 2009; 30: 173-187.
- [114] Giatti S, Caruso D, Boraso M, Abbiati F, Ballarini E, Calabrese D, Pesaresi M, Rigolio R, Santos-Galindo M, Viviani B, Cavaletti G, Garcia-Segura LM, Melcangi RC. Neuroprotective effects of progesterone in chronic experimental autoimmune encephalomyelitis. J Neuroendocrinol 2012; 24: 851-861.
- [115] Schumacher M, Hussain R, Gago N, Oudinet JP, Mattern C, Ghoumari AM. Progesterone synthesis in the nervous system: implications for myelination and myelin repair. Front Neurosci 2012; 6: 10.
- [116] Pettus EH, Wright DW, Stein DG, Hoffman SW. Progesterone treatment inhibits the inflammatory agents that accompany traumatic brain injury. Brain Res 2005; 1049: 112-119.
- [117] Robertson CL, Puskar A, Hoffman GE, Murphy AZ, Saraswati M, Fiskum G. Physiologic progesterone reduces mitochondrial dysfunction and hippocampal cell loss after traumatic brain injury in female rats. Exp Neurol 2006; 197: 235-243.
- [118] Stein DG. Progesterone exerts neuroprotective effects after brain injury. Brain Res Rev 2008; 57: 386-397.
- [119] Stein DG. Progesterone in the treatment of acute traumatic brain injury: a clinical perspective and update. Neuroscience 2011; 191: 101-106.
- [120] Wright DW, Kellermann AL, Hertzberg VS, Clark PL, Frankel M, Goldstein FC, Salomone JP, Dent LL, Harris OA, Ander DS, Lowery DW, Patel MM, Denson DD, Gordon AB, Wald MM, Gupta S, Hoffman SW, Stein DG. ProTECT: a randomized clinical trial of progesterone for acute traumatic brain injury. Ann Emerg Med 2007; 49: 391-402, 402-2.
- [121] Xiao G, Wei J, Yan W, Wang W, Lu Z. Improved outcomes from the administration of progesterone for patients with acute severe traumatic brain injury: a randomized controlled trial. Crit Care Lond Engl 2008; 12: R61.
- [122] González SL, Labombarda F, González Deniselle MC, Guennoun R, Schumacher M, De Nicola AF. Progesterone up-regulates neuronal brain-derived neurotrophic factor expression in the injured spinal cord. Neuroscience 2004; 125: 605-614.
- [123] Labombarda F, González S, Lima A, Roig P, Guennoun R, Schumacher M, De Nicola AF. Progesterone attenuates astro- and microgliosis and enhances oligodendrocyte differentiation following spinal cord injury. Exp Neurol 2011; 231: 135-146.

- [124] Labombarda F, González SL, Lima A, Roig P, Guennoun R, Schumacher M, de Nicola AF. Effects of progesterone on oligodendrocyte progenitors, oligodendrocyte transcription factors, and myelin proteins following spinal cord injury. Glia 2009; 57: 884-897.
- [125] Arbo BD, Bennetti F, Ribeiro MF. Astrocytes as a target for neuroprotection: modulation by progesterone and dehydroepiandrosterone. Prog Neurobiol 2016; 144: 27-47.
- [126] Farahabadi A, Akbari M, Amini Pishva A, Zendedel A, Arabkheradmand A, Beyer C, Dashti N, Hassanzadeh G. Effect of progesterone therapy on TNF- $\alpha$  and iNOS gene expression in spinal cord injury model. Acta Med Iran 2016; 54: 345-351.
- [127] Garcia-Ovejero D, González S, Paniagua-Torija B, Lima A, Molina-Holgado E, De Nicola AF, Labombarda F. Progesterone reduces secondary damage, preserves white matter, and improves locomotor outcome after spinal cord contusion. J Neurotrauma 2014; 31: 857-871.
- [128] Thomas AJ, Nockels RP, Pan HQ, Shaffrey CI, Chopp M. Progesterone is neuroprotective after acute experimental spinal cord trauma in rats. Spine 1999; 24: 2134-2138.
- [129] Fee DB, Swartz KR, Joy KM, Roberts KN, Scheff NN, Scheff SW. Effects of progesterone on experimental spinal cord injury. Brain Res 2007; 1137: 146-152.
- [130] Schumacher M, Mattern C, Ghoumari A, Oudinet JP, Liere P, Labombarda F, Sitruk-Ware R, De Nicola AF, Guennoun R. Revisiting the roles of progesterone and allopregnanolone in the nervous system: resurgence of the progesterone receptors. Prog Neurobiol 2014; 113: 6-39.
- [131] Mani SK, O'Malley BW. Mechanism of Progesterone Receptor Action in the Brain. In: D.W. Pfaff, P. Arnold, A.M. Etgen (Eds.). molecular mechanisms of hormone actions on behavior. Amsterdam: Academic Press; 2009. pp. 375-411.
- [132] Schumacher M, Zhu X, Guennoun R. Progesterone: synthesis, metabolism, mechanism of action, and effects in the nervous system. in: hormones, brain and behavior. Elsevier; 2017. p. 215-244.
- [133] Li X, Wong J, Tsai SY, Tsai MJ, O'Malley BW. Progesterone and glucocorticoid receptors recruit distinct coactivator complexes and promote distinct patterns of local chromatin modification. Mol Cell Biol 2003; 23: 3763-3773.
- [134] Lösel R, Wehling M. Nongenomic actions of steroid hormones. Nat Rev Mol Cell Biol 2003; 4: 46-56.
- [135] Mani SK. Signaling mechanisms in progesterone-neurotransmitter interactions. Neuroscience 2006; 138: 773-781.

- [136] Stanczyk FZ, Hapgood JP, Winer S, Mishell DR. Progestogens used in postmenopausal hormone therapy: differences in their pharmacological properties, intracellular actions, and clinical effects. Endocr Rev 2013; 34: 171-208.
- [137] Moore NL, Hickey TE, Butler LM, Tilley WD. Multiple nuclear receptor signaling pathways mediate the actions of synthetic progestins in target cells. Mol Cell Endocrinol 2012; 357: 60-70.
- [138] Rabinowitz A, Cohen SJ, Finn DA, Stackman RW. The neurosteroid allopregnanolone impairs object memory and contextual fear memory in male C57BL/6J mice. Horm Behav 2014; 66: 238-246.
- [139] Johansson IM, Birzniece V, Lindblad C, Olsson T, Bäckström T. Allopregnanolone inhibits learning in the Morris water maze. Brain Res 2002; 934: 125-131.
- [140] Matthews DB, Morrow AL, Tokunaga S, McDaniel JR. Acute ethanol administration and acute allopregnanolone administration impair spatial memory in the Morris water task. Alcohol Clin Exp Res 2002; 26: 1747-1751.
- [141] Turkmen S, Lundgren P, Birzniece V, Zingmark E, Backstrom T, Johansson IM. 3beta-20betadihydroxy-5alpha-pregnane (UC1011) antagonism of the GABA potentiation and the learning impairment induced in rats by allopregnanolone. Eur J Neurosci 2004; 20: 1604-1612.
- [142] Bäckström T, Haage D, Löfgren M, Johansson IM, Strömberg J, Nyberg S, Andréen L, Ossewaarde L, van Wingen GA, Turkmen S, Bengtsson SK. Paradoxical effects of GABA-A modulators may explain sex steroid induced negative mood symptoms in some persons. Neuroscience 2011; 191: 46-54.
- [143] Labombarda F, Jure I, Gonzalez S, Lima A, Roig P, Guennoun R, Schumacher M, De Nicola AF. A functional progesterone receptor is required for immunomodulation, reduction of reactive gliosis and survival of oligodendrocyte precursors in the injured spinal cord. J Steroid Biochem Mol Biol 2015; 154: 274-284.
- [144] Coronel MF, Raggio MC, Adler NS, De Nicola AF, Labombarda F, González SL. Progesterone modulates pro-inflammatory cytokine expression profile after spinal cord injury: implications for neuropathic pain. J Neuroimmunol 2016; 292: 85-92.
- [145] Coronel MF, Sánchez Granel ML, Raggio MC, Adler NS, De Nicola AF, Labombarda F, González SL. Temporal changes in the expression of the translocator protein TSPO and the steroidogenic enzyme 5α-reductase in the dorsal spinal cord of animals with neuropathic pain: effects of progesterone administration. neurosci Lett 2016; 624: 23-28.

- [146] González SL, Coronel MF. Beyond reproduction: the role of progesterone in neuropathic pain after spinal cord injury. Neural Regen Res 2016; 11: 1238-1240.
- [147] Hulsebosch CE, Hains BC, Crown ED, Carlton SM. Mechanisms of chronic central neuropathic pain after spinal cord injury. Brain Res Rev 2009; 60: 202-213.
- [148] Yezierski RP. Spinal cord injury pain: spinal and supraspinal mechanisms. J Rehabil Res Dev 2009; 46:95–107.
- [149] Ji RR, Berta T, Nedergaard M. Glia and pain: is chronic pain a gliopathy? Pain 2013; 154 Suppl 1: S10-28.
- [150] Walters ET. Neuroinflammatory contributions to pain after SCI: roles for central glial mechanisms and nociceptor-mediated host defense. Exp Neurol 2014; 258: 48-61.
- [151] Gwak YS, Kang J, Unabia GC, Hulsebosch CE. Spatial and temporal activation of spinal glial cells: role of gliopathy in central neuropathic pain following spinal cord injury in rats. Exp Neurol 2012; 234: 362-372.
- [152] Viviani B, Boraso M, Marchetti N, Marinovich M. Perspectives on neuroinflammation and excitotoxicity: a neurotoxic conspiracy? Neurotoxicology 2014; 43: 10-20.
- [153] Vezzani A, Viviani B. Neuromodulatory properties of inflammatory cytokines and their impact on neuronal excitability. Neuropharmacology 2015; 96: 70-82.
- [154] Ji RR, Xu ZZ, Gao YJ. Emerging targets in neuroinflammation-driven chronic pain. Nat Rev Drug Discov 2014; 13: 533-548.
- [155] Guo W, Wang H, Watanabe M, Shimizu K, Zou S, LaGraize SC, Wei F, Dubner R, Ren K. Glialcytokine-neuronal interactions underlying the mechanisms of persistent pain. J Neurosci Off J Soc Neurosci 2007; 27: 6006-6018.
- [156] Tiwari V, Guan Y, Raja SN. Modulating the delicate glial-neuronal interactions in neuropathic Pain: promises and potential caveats. Neurosci Biobehav Rev 2014; 45: 19-27.
- [157] Yang Z, Xie W, Ju F, Khan A, Zhang S. In vivo two-photon imaging reveals a role of progesterone in reducing axonal dieback after spinal cord injury in mice. Neuropharmacology 2017; 116: 30-37.
- [158] Matsuura S, Ohashi M, Chen HC, Shownkeen RC, Hartree AS, Reichert LE, Stevens VC, Powell JE. Physicochemical and immunological characterization of an HCG-like substance from human pituitary glands. Nature 1980; 286: 740-741.
- [159] Cole LA, Sasaki Y, Muller CY. Normal Production of Human Chorionic Gonadotropin in Menopause. N Engl J Med 2007; 356: 1184-1186.

- [160] Patil A, Fillmore K, Valentine J, Hill D. The study of the effect of human chorionic gonadotrophic (HCG) hormone on the survival of adrenal medulla transplant in brain. Preliminary study. Acta Neurochir (Wien) 1987; 87: 76-78.
- [161] Lei ZM, Rao CV. Neural actions of luteinizing hormone and human chorionic gonadotropin. Semin Reprod Med 2001; 19: 103-109.
- [162] Meng XL, Rennert OM, Chan WY. Human chorionic gonadotropin induces neuronal differentiation of PC12 cells through activation of stably expressed lutropin/choriogonadotropin receptor. Endocrinology 2007; 148: 5865-5873.
- [163] Belayev L, Khoutorova L, Zhao KL, Davidoff AW, Moore AF, Cramer SC. A novel neurotrophic therapeutic strategy for experimental stroke. Brain Res 2009; 1280: 117-123.

- [164] Patil AA, Nagaraj MP. The effect of human chorionic gonadotropin (HCG) on functional recovery of spinal cord sectioned rats. Acta Neurochir (Wien) 1983; 69: 205-218.
- [165] Patil AA, Filmore K, Hill D. The effect of human chorionic gonadotropin (HCG) on restoration of physiological continuity of the spinal cord. A preliminary report. Int Surg 1990; 75: 54-57.
- [166] Brotfain E, Gruenbaum SE, Boyko M, Kutz R, Zlotnik A, Klein M. Neuroprotection by estrogen and progesterone in traumatic brain injury and spinal cord injury. Curr Neuropharmacol 2016; 14: 641-653.