Review Article HSF1: a potential target for therapeutic intervention in cancer

Shu-Yue Wu^{3#}, Peng Guo^{4#}, Tao Peng¹, Jing Xu¹, Qing-Qing Hou², Xing Sun², Zhi Zhang^{2*}, Hai Huang^{2*}

¹Department of Hepatobiliary Surgery, The First Affiliated Hospital of Guangxi Medical University, Nanning, China; Departments of ²Hepatobiliary Surgery, ³Pediatrics, The Fifth Affiliated Hospital of Guangxi Medical University, Nanning, China; ⁴Department of Oral Surgery, The Dental Hospital of Guangxi Medical University, Nanning, China. [#]Equal contributors and co-first authors. ^{*}Equal contributors.

Received September 5, 2016; Accepted February 4, 2017; Epub March 15, 2017; Published March 30, 2017

Abstract: Heat-Shock Transcription Factor 1 (HSF1) is an evolutionarily highly conserved transcription factor, which plays a key role in the heat-shock response of stressed cells and increases their survival by protecting them against environmental stressors. The last decade of research has revealed that HSF1 is chronically activated or overexpressed in a wide range of cancers, in which it facilitates cancer cell survival, malignant transformation, and cancer proliferation in model systems. HSF1 has a role in cellular homeostasis to transactive genes that encode heat-shock proteins. Apart from its role in reprogramming transcriptions, HSF1 acts as a remarkably potent modifier of signal modulation, stimulating kinase activity and regulating energy metabolism. In addition, HSF1 plays a pivotal role in the development of cancer tumor-free survival in whole animals. The high incidence of HSF1 overexpression in human tumors suggests that the protein is important in the carcinogenic process and therefore a potential candidate for therapeutic intervention. In the present review, we will summarize the current knowledge and highlights in the HSF1 field, discuss disrupting the influence of HSF1 physiological condition, received stimuli and the organismal control over HSF1 in cancer.

Keywords: HSF1, carcinogen, cancer

Introduction

The last century has witnessed a remarkable evolution of insights in the cellular activity and physiology. Among numerous changes most remarkable activity in stressed cells is the production of heat shock of stress proteins, which are a highly conserved set of proteins. The heat shock response is mediated by the heat shock element (HSE), characterized by increased expression of heat shock proteins (HSPs), which is an important homeostatic mechanism that maintains protein homeostasis in all organisms from bacteria to humans [1-4]. This protein has been proven to be essential for survival of all these organisms under stressful conditions. The heat shock factor (HSF) is an activator protein, and can be specifically bind to the HSE and regulate the HSP expression on stress stimulation [5]. In vertebrates and plants, HSF are grouped into four major families according to their biological characteristics as follows:

HSF1, HSF2, HSF3 and HSF4. In addition, HSF1 and HSF2 are currently known to be existed in all vertebrates, and it was found that HSF3 is specific for avian species and HSF4 for mammals [6]. Among the HSF family, HSF1 has been considered a master regulator of the heat shock response in eukaryotes [7]. Work over the last three decades has further revealed the importance of HSF1 as a transcriptional activator of chaperones, ubiquitin and cochaperones as well as translational regulator and also as a coordinator of the expression of many transcriptional, mitotic determinants and signaling molecules [8-11]. This review summarizes the current knowledge regarding the potential of influence of HSF1 in cancer and evaluates the usefulness of using HSF1 as a biomarker in clinical practice.

The structure and activation of HSF1

A number of studies have shown that HSF1 is highly conserved in eukaryotic species. The

16	120	137	212	310 300	37	78 4	07 503
HI	н	HR	A/B			HR-C	

Figure 1. Graphic of the HSF1 protein structure. HTH, helix-turn-helix; HR, hydrophobic heptad repeats.

functional properties of HSF1 are linked to a highly conserved structural domains in these proteins (Figure 1): they contain an N-terminal DNA-binding helix-turn-helix (HTH) domain and an adjacent oligomerization domain that consists of an adjacent bipartite oligomerization domain (HR-A/B) [12-16]. It has been reported that spontaneous trimerization of HSF1 is allosterically inhibited by a C-terminal heptad repeat (HR-C), which folds back and forms intramolecular contacts with the oligomerization domain HR-A/B [15]. The transactivation domain of HSF1 is targeted by several proteins to link directly to the HSF1 activation and to direct HSF1 to specific target genes [17]. The activity of HSF1 is known to be complex mechanisms protein-protein interactions and post-translational modifications [18]. Currently, our understanding of the phosphorylated, trimerized, and translocation to the nucleus, induces chaperone gene, expression by binding to DNA sequence motifs known as HSEs [19]. Several reports have shown that phosphorylation of S326 and S230 have significantly increased the transcriptional capacity of HSF1 [20, 21]. Studies suggested the importance of HSF1 in transcriptional activation of chaperones, ubiquitin, cochaperones, translational regulators. It also coordinates the expression of many transcriptional, mitotic determinants and signaling molecules [8-11].

The remarkable property of HSF1 activation by stress, such as: heavy metals, exposure to oxidants, elevated temperatures, and bacterial or viral infections, and is found on the promoters of target genes within a few seconds of heat shock [22]. After the stresses, HSPs are expressed abundantly [23], and to deter unfolding of client proteins in the stress and to mediate refolding [24]. The proteome is constantly vulnerable to protein stress due to replicative, mitotic, proteotoxic, metabolic, oxidative stress and the expression of proteins with dominant aggregation prone conformations [25]. Tumors, is a consequence of the multitude of stressful conditions involves the opposite scenario-elevated HSP levels that closely related to malignancy [26]. Furthermore, cancer cells are mutation prone that correlate with metabolism, cell-cycle regulation, signaling, adhesion and translation. It is well known that HSF1 is an essential factor in the heat-shock response, in which it causes facilitates malignant transformation, proliferation, and cancer cell survival in model systems, however, many are remain elusive in malignancy.

Calderwood et al. and Ciocca et al. revealed that HSF1 can bind to 5'-promoter regions of all HSP genes and trigger rapid and abundant transcription of these stress protein genes [27, 28]. Importantly, HSP genes involve stressinduced formation of a HSF1 homotrimer. Furthermore, the posttranslational modifications (PTM) that convert the factor into an active form that lead to a nuclear localization and binds the promoter of HSP genes in a fruitful manner [29]. There are five main HSP families that is organized by molecular size and functional class, include the Hsp90 (HSPC), Hsp70 (HSPA), Hsp60 (HSPD), small HSP family (HS-PB), and large HSP (HSPH) families [30], many of the families are thought to play key roles in cancer [27, 28]. HSF1 has been shown to involved in etiology of cancer by its multiple effects in: (i) Increased transcription and translation of HSPs; (ii) Regulation of translation [28]. However, the pathways of the induction of HSF1 in cancer are still under intense investigation.

The role of HSF1 in cell cycle

It has been shown that the transcriptional responses are involved in all phases of the cell cycle [31], and that the HSF1 is involved in the regulation of mitosis [32]. Additionally, the depletion of HSF1 in established human cancer lines strongly impaired the cell proliferation and survival [33-37]. Huang et al. [38] shown that HSF1 promotes polyploidy in p53 deficient cells, and expressions of dominant negative HSF1 delays the decay in cyclin B1. In addition, studies have suggested that silencing HSF1 by short hairpin RNA decreases cell proliferation in human melanoma cell lines [39]. Interestingly, there are several lines of evidence to suggest that Cdc20 binds directly to HSF1 and this interaction may thus be involved in the cell cycling. In a recent review, it has been suggested that HSF1 impacts a large array of healthy and malignant cells [9, 11, 40]. It has been shown that HSF1 can bind to a number of protein kinases to modulate a range of signal transduction pathways [38, 41, 42], leading to phosphorylation of HSF1 and potentially other substrates in cancer cells.

HSF1 relationship with cancer and therapeutic potential

A few recent studies have shown that elevated expression of the HSF1 can be associated with advanced stages in hepatocellular carcinoma, oral squamous cell carcinoma and breast cancer. suggesting HSF1 activity as a mechanistic link between stages in tumor progression and cancer cell invasion and metastasis [43-46]. Others have reported that HSF1 overexpression may contribute to mammary carcinoma with an Epithelial-mesenchymal transition (EMT) phenotype and ability to grow under anchorage independent conditions [47]. In a recent pilot study, increasing expressions of HSF1 in tumors was associated with the tumorigenesis in multiple animal models [33, 45, 481. This may provide further mechanistic information on HSF1 activity and explain why expressions a higher level of HSF1 in tumors is a predictor of advanced stage events. This section summarizes the important roles of HSF1 in all reported types of cancer to date.

Breast cancer: Several recent studies have suggested that over-expression of HSF1 led to massive cell death of human breast cancer Bcap37 cells by promoting apoptosis induced by heat shock [49]. In addition, over-expression of HSF1 enhances the Adel55 killing Bcap37 cell line potential through increasing the viral replication both in vitro and in vivo [50]. Wang et al. [51] showed that HSF1 over-expression augment Breast cancer stem cells (CSC) phenotype in breast cancer cell lines, suggested that HSF1 may be one route to target CSCs in breast cancer. Interestingly, HSF1 depletion decreases the MICB expression leads to a reduction in the natural killer (NK) cell-mediated cytotoxicity, promotes tumor development, metastasis and therapy resistance [52]. Schilling et al. [53] demonstrated that indicated the Hsp90 inhibitor NVPAUY922 induced Hsp70 expression by inhibition of HSF1 activity and suppressed invasion and migration in human breast tumor cells. In in vitro tests, withaferin A induces apoptosis breast cancer cell lines down-regulation of protein expression [54]. There is increasing evidence that elevation of HSF1 expression elevated the risk of breast cancer among patients with Triple-Negative Breast Cancer (TNBC) and poor prognosis of HER2-/ER+ subtypes [55].

Schulz et al. [56] reported that over-expressed HER2 constitutively activates HSF1 promoted tumor proliferation via the PI3K-AKT-mTOR signaling pathway. Xi et al. [57] have shown that deletion of HSF1 in mice over-expressing ErbB2/Neu significantly reduces mammary metastasis and tumorigenesis via RAS/RAF/ MEK/ERK1/2 signaling pathway with suppressed levels of HSP90 in complex with threonine-protein kinase (RAF1). In addition, HSF1 has been shown to be essential for HER2induced tumorigenesis, might be a target of rs4919510:C.G in mature miR-608 and may influence HER2+breast cancer risk and tumor proliferation [58]. Zhao et al. [37] found that trastuzumab inhibits glycolysis induced tumor growth via down-regulation of HSF1in ErbB2+ breast cancer cells. Chou et al. [59] experiments suggested that activated HSF1 is a key factor in inducer of gene expression at the post transcriptional level in mammary cancer with influence translation of a range of proteins through its effects on RNA-binding protein HuR. microRNAs, and lincRNA-p21.

Studies demonstrated that HSF1 plays an important role in the pathogenesis of ErbB2overexpressing cells by increased the expression of glycolysis-regulating molecules lactate dehydrogenase A (LDH-A), and inhibited human breast cancer MCF7 cells growth [48]. Kim et al. [60] found that it could be a more effective therapeutic approach for use in combination with Hsp90 inhibitor and silent information regulator two homologue one (SIRT1) inhibitor, and the approach would be aimed at the treatment of Hsp90 inhibitor-resistant multi-drug resistance (MDR) human breast cancer cell lines via down-regulation of HSF1. Yang et al. [61] reported that HSF1 plays an important role in breast cancer epithelial cells cycle progression via ERK1/2 MAPK and PI3K/Akt signaling pathways. Interestingly, another recently published study report that the natural isothiocyanate significantly inhibited the expression of HSF1, accompanied by cell-cycle arrest at G2/M phase and apoptosis in breast cancer cell lines [62]. Studies have reported that during HSF1overexpression, the autophagy-related 7 (ATG7) regulates the cytoprotective autophagy in breast cancer cells [63, 64]. In addition, breast cancer cell lines was transfected with HSF1d-202-316 exhibiting a highly resistant phenotype undergo apoptosis [65]. Wang et al. [66] suggested that mHSF1 sensitizes breast cancer cell line Bcap37 to hyperthermia by promoting the apoptotic process via enhancing JNK and caspase-3 pathways.

Studies evaluating the HSF1-MTA1 complex formation were induced to assemble on the chromatin of breast carcinoma cell MCF7, participate in repression of estrogen-dependent transcription [47, 67]. It was reported that HSF1 protein levels are up-regulated in response to expressions of JWA protein expression and enhance intracellular defenses against H₂O₂induced oxidative damage in MCF7 cell lines [68]. Hansen et al. [69] showed that guercetin slightly affected HSF1 expression Hansen n in MDA-MB-231 breast cancer cells through regulation of HSF transcriptional activity. Thus, when taken together, these findings reinforce the carcinogenic and therapeutic potential of HSF1 in breast cancer, which could have important clinical implications for HSF1 use as a therapeutic target in breast cancer.

Colorectal cancers: HSF1 is highly expressed in colorectal cancer tissues, and its expression level correlates with the carcinogenesis of colorectal cancer [70]. Moreover, pHSF1 (Ser230) protein is strongly expressed in human colorectal carcinoma tissues and overexpression of HSF1 promotes an increase in DAPK mRNA level and induced apoptosis colorectal cancer cells [71]. Additionally, overexpression of HSF1 in Colo205 cells enhanced sensitivity to NK cell killing following mild thermal stress via up-regulation of molecularly imprinted colloidal array (MICA) expression [72]. Wales et al. [73] study showed that silencing HSF1 results in a significant enhancement of drug potency, which dependent on activation of ERK-1/2 mitogenactivated protein kinase pathway. Additionally, in vitro assays revealed that HCT-116 colorectal cancer cells were treated with the natural compound cantharidin, significantly suppressed the expression of HSF1 downstream target proteins: HSP70 and BCL2-associated athanogene 3 (BAG3) by blocking HSF1 binding to promoters [74]. It was observed that glutamine

protects the ethanol induced intestinal barrier function in human epithelial colorectal carcinoma cells (Caco-2), by modulating HSF1-mediated Hsp70 protein expression [75].

It was reported that intra-dermal murine models of colorectal carcinoma (CT26) were treated with Adel55-cHSF1 showed sustained resistance upon re-challenge with autologous tumor cells, suggesting that cHSF1 is a better gene beneficial to prevent tumor recurrence for neoadjuvant immunotherapy [76]. Moreover, fisetin abolished HSF1 target proteins including: HSP70, HSP27 and BAG3 activity with an IC₅₀ of 14 µM, led to HCT-116 cancer cells proliferation and apoptosis [77]. Additionally, mixed-lineage leukemia 1 (MLL1) depletion is combined with HSP90 inhibition through block a HSF1mediated feedback mechanism inhibits HCT-116 cell proliferation [78]. Observations by Jacobs et al. [79] indicated that HSF1-mediated BAG3 expression impairs apoptosis treatment with 4-hydroxynonenal through stabilization of Bcl-2 proteins in colon cancer cells. These findings imply that target HSF1 gene attracted considerable interest for further preclinical studies to challenge the current paradigm available for treatment of colon cancer which is characterized by elevated levels of HSF1 protein.

Skin tumor: HSF1 has been shown to be essential for the proliferation and survival of the immortalized Hsf1+/+ MEF cells [33]. Kourtis et al. [80] have shown that the ubiquitin ligase FBXW7 α interacts with HSF1 correlates with increased metastatic potential and disease progression in human melanoma. Additionally, MLL1 regulates HSF1-target genes upon HSP90 inhibition inhibits A375 and A2058 melanoma cell line proliferation [78]. The results indicate that HSF1 have shown its mechanism of action as a carcinogenic in skin tumor.

Glioblastoma: HSF1 has also been investigated in correlated with forkhead box transcription factor (FoxM1) overexpression in human glioblastoma specimens. Additionally, study showed that HSF1 bound to FoxM1 promoter induced FoxM1 promoter activity enhanced cell survival under lethal heat shock stress condition in mouse embryo fibroblast cells. It was reported that Ectopic expression of BAG3 leads to the activation of an HSF1-driven stress response, results in a remarkable increase in colony for-

mation capacity in glioblastoma cells [81]. Recently Oh et al. [82] demonstrated that HSF1 plays an important role in induces HSP72 expression thus activating the Rac1-NADPH oxidase-independent ROS production pathway in C6 glioma cell. Liu et al. [83] assumed that glioma cells were treated with PS-341-induced cell damage by upregulated HSF1 expression. Grogan et al. [84] discovered that withaferin A prevented glioblastoma multiforme cell proliferation by dose-dependent G2/M cell cycle arrest and cell death through the Akt/mTOR apoptotic pathway, through decreased the expression of HSF1. Additionally the nature of H-7 (1-(5-isoquinolinesulfonyl)-2-methylpiperazine dihydrochloride) was significantly suppressed HSF1 gene expression, which dependent on cyclic AMP-dependent PK, calcium-dependent PK, and cyclic GMP-dependent PK, in in a human glioblastoma cell line (A-172) [85]. The results demonstrated that HSF1 as a potential of carcinogenic in glioblastoma, suggested that HSF1 can act as therapeutic targets for glioblastoma.

Head and neck cancers: It was reported that p-HSF1 is highly expressed in oral squamous cell cancer (OSCC) tissues, and its expression level correlates with advanced clinical stage, metastasis and recurrence of the patients [86]. This suggests that HSF1 mediates the metastatic potential of OSCC cells and also points to HSF1 as a potential therapeutic target for the clinical management of head and neck cancer.

Hepatic cancers: A more recent study showed a high prevalence of total and phosphorylated HSF1 protein, mRNA expression, induces DNA-PKcs upregulation via the activation of the MAPK/JNK/AP-1 axis, which is significantly correlated with aggressive features and unfavorable prognosis in hepatocellular carcinoma (HCC). Additionally, HSF1 high expression in peritumoral tissue and significantly correlate with poorer overall survival and early recurrence [87]. Recently, in vitro tests reported that HSF1 is overexpressed in HCC at both mRNA and protein level, and HSF1 knockdown could inhibit the proliferation of Hep3B [88]. It was report that HSF1 is over-expressed in HCC with adverse pathologic and clinical features, and was capable of promoting HepG2, MHCC97-L and HCCLM3 cell lines migration, invasion in vitro and in vivo by facilitating the expression of p-Hsp 27 [43]. In addition, overexpression of HSF1 led to induction of insulin sensitivity and activation of AMP-activated protein kinase, suppressed cancer progression, mitigating adverse effects of carcinogens on hepatic metabolism [45]. Interestingly, down-regulated of heat-induced p53/HSF1 led to the inhibition of Hsp70 protein expression, induced carboplatintriggered HepG2 cells death by restoring the sensitivity of heat-stressed [89].

Chuma et al. [90] have shown that knockdown HSF1 reduced tumorigenesis and appeared attributable to increased apoptosis and decreased proliferation in KYN2 HCC cells. In vitro tests, the collaboration of HSF1 and APOBEC3B cytidine deaminase promoted the growth of neoplastic human HepG2 liver cells [91]. It was demonstrated that, HSF1 activated miR-135b expression via kazal motifs and ecotropic viral integration site 5 pathway, promoted HCC cell motility and invasiveness [92]. Lee et al. [93] results indicate that mitochondrial respiratory defects induced CIn-1-mediated SNU hepatoma cells invasiveness through reactive oxygen species (ROS)-mediated HSF1 activation. Additionally, the phosphorylation of HSF1/S326 can be activated by glucose-mTOR pathway via upregulate the expression of HSF1-s downstream alpha B-crystallin and Hsp70, promoted plc/prf5 cell proliferation [94]. Above in vitro and in vivo trials have demonstrated that HSF1 acts as an oncoprotein can be expected to be expanded as research progresses, and target HSF1 therapy may be an important modality to treat HCC.

Leukemia: Previous description has shown that knockdown of HSF1 abrogated the colony formation capacity of the acute myeloid leukemia (AML) cancer stem cells (CSCs), through HSP90-mediated AKT activation [95]. Additionally, inhibition of HSF1 resulted in reduced the expression level of FOP2-FGFR1 could lead to block the oncoprotein induced proliferation of KG-1a leukemic cells [96]. Interestingly, resveratrol down-regulation of Hsp70 correlated with a diminished presence of HSF1 induced apoptosis in K562 cells [97]. Takaki et al. [98] reported that leukemia inhibitory factor (LIF) expression are lacking in HSF1-null mice. It was demonstrated that overexpression of HSF1 in CLL sensitive to triptolide treatment induces apoptosis in cultured through reduced association of HSP90 with co-chaperone cell division cycle [99]. There is also some evidence suggesting that silent information regulator two homologue one (SIRT1) depletion caused significant down-regulation of HSF1 led to the sensitization of human chronic myeloid leukemia K562 cells to Hsp90 inhibitor by SIRT1 inhibitor [100]. It has been shown that geldanamycin induced the phosphorylation of HSF1 stimulates HSP70 protein expression in the human erythroleukemic cell line K562 [101, 102]. Sarkar et al. [67] showed that overexpression of HSF1, HSP27, HSP70, HSP90, and histone deacetylase 6 (HDAC6) were down-regulated by curcumin, which resulted in cell cycle arrest at the G2/M stage, leading to apoptosis in two different leukemia cell lines (K-562 and HL-60) [103]. It can be suggested that HSF1 is involved in leukemia metastasis, and HSF1 treatment may be used as an alternative treatment against leukemia.

Lung cancers: It has been shown that HSF1 is highly expressed in non-small cell lung cancer tissues, and its expression level correlates with node metastasis of the patients [104]. Both in vitro and in vivo tests, Coxsackievirus B3 (CVB3) replication and leads to phosphorylation of HSF1 enhanced Hsp70-1 transcription [105]. Chang et al. [106] shown that human non-small cell lung cancer H460 cells were treated with geldanamycin induces HSF1 activation and HSP70 protein expression. Kim et al. [107] reported that Coniferyl aldehyde protected normal lung tissues from the therapeutic irradiation by increased expression of the HSF1 protein, but not in A549 lung orthotopic lung tumor model. Ma et al. [108] demonstrated that HSF1 as a regulator of energy metabolism through activation of the PGC1 α -dependent metabolic program, represents a potential strategy for treating obesity and metabolic syndrome. In addition, it has been shown that 2,4-Bis(4-hydroxybenzyl)phenol inhibited HSF1 activity with decreased levels of hsp27 and hsp70, which induced growth arrest and apoptosis of NCI-H460 human lung cancer cells [109]. These results indicate that HSF1 is a potential anti-metastatic and anti-invasive agent, and may be useful in gene therapy strategies for the treatment lung cancer.

Prostate cancers: It was also shown that prostate cancer shows a high prevalence of HSF1

expression, which is significantly correlated with aggressive cell growth, differentiation, or apoptosis [110]. In addition, HSF1 plays an important role in the human prostate cancer PC-3 cells influences cell cycle behavior and progression via mitosis and promotes the development of the aneuploid state, by regulation of cyclin B1 degradation [111]. Other authors [112], demonstrating that in in vivo experiments, severe hypothermia and rewarming increased expression of HSF1 mRNA level in rat ventral prostate were quantified hypothermia and in rewarming, which promote proliferation of cells in healthy rat prostate tissue through ErbB signaling pathway. This suggests that targeting the destruction of HSF1 expression maybe provides a novel specific lead into prostate cancer therapy.

Lymphoma: Recent studies have reported that AG490 (JAK/STAT inhibitor) induced a complete autophagy by reduction of HSP70 and HSF1 protein expression in primary effusion lymphoma (PEL) cells [113]. Moreover, chelerythrine reductions the expression of HSF1 and hsp70 in Dalton's lymphoma (DL) cells by PKC phosphorylation [114]. This finding also provides a significant cue to lymphoma treatment, by targeting the destruction of HSF1 expression.

Pancreatic cancer: Recently, it has been demonstrated that downregulation of HSF1 expression induces apoptosis via caspase-3 activation in pancreatic cancer (MIA PaCa-2 and S2-013) and cholangiocarcinoma (KMBC and KMCH) cell lines [115]. In addition, triptolide induced apoptotic cell death through decreased levels of hsp70 and HSF1 in pancreatic cancer cell lines [116, 117]. Furthermore, studies shown that active hexose-correlated compound down-regulated hsp27 via reduction of the HSF1 in human pancreatic cancer cells [118]. There is also some evidence suggesting that HSF1 reinforce the carcinogenic in pancreatic cancer, and target HSF1 may be a potent and novel therapy for treatment of patients with pancreatic cancer.

Cervical cancer: It was shown that HSF1 was essential for the transcriptional regulator of dystrophin Dp71 expression in human cervical carcinoma HeLa cells, suggesting that target HSF1 could be effective in future therapeutic strategies in cervical cancer [119].

Perspectives

In summary, HSF1 is involved in a multitude of physiological processes and associated with the development of a multitude of cancer types. Undoubtedly, these effects include: HSF1 controls the cell cycle, the signaling and the mediation of tumor progression; therefore, the role of pleiotropic and essential properties in carcinogenesis and tumor progression of HSF1 as a predictor of beneficial target in therapy should be thoroughly evaluated for different cancer types and treatment regimens.

Finally, although our understanding of the importance of HSF1 activity has driven a transcriptional program to support highly malignant human cancers, however, a number of essential aspects of HSF1 remain to be elucidated, and investigations regarding HSF1 cannot yet be applied in the clinic. In addition, there is still a long road ahead before HSF1 determination can leave the laboratory bench and can be deployed in cancer therapy. At the moment, HSF1 as a carcinogen that may prove to be clinically relevant in the future, however a large number of well-designed studies to test for suitable in vivo properties are needed before this can become a reality.

Acknowledgements

This research was supported by Youth Science Foundation of Guangxi Medical University (No. GXMUYSF201542), the scientific research and technology development plan of Nanning (No. 20163130).

Disclosure of conflict of interest

None.

Address correspondence to: Hai Huang and Zhi Zhang, Department of Hepatobiliary Surgery, The Fifth Affiliated Hospital of Guangxi Medical University, Nanning, China. E-mail: nnsyy2016@aliyun. com (HH); zzhndx@aliyun.com (ZZ)

References

- Balch WE, Morimoto RI, Dillin A and Kelly JW. Adapting proteostasis for disease intervention. Science 2008; 319: 916-919.
- [2] Morimoto RI. Regulation of the heat shock transcriptional response: cross talk between a family of heat shock factors, molecular chaper-

ones, and negative regulators. Genes Dev 1998; 12: 3788-3796.

- [3] Parsell DA and Lindquist S. The function of heat-shock proteins in stress tolerance: degradationand reactivation of damaged proteins. Annu Rev Genet 1993; 27: 437-496.
- [4] Westerheide SD and Morimoto RI. Heat shock response modulators as therapeutic tools for diseases of protein conformation. J Biol Chem 2005; 280: 33097-33100.
- [5] Akerfelt M, Trouillet D, Mezger V and Sistonen L. Heat shock factors at a crossroad between stress and development. Ann N Y Acad Sci 2007; 1113: 15-27.
- [6] Nakai A. New aspects in the vertebrate heat shock factor system: Hsf3 and Hsf4. Cell Stress Chaperones 1999; 4: 86-93.
- [7] Koizumi S, Suzuki K and Yamaguchi S. Heavy metal response of the heat shock protein 70 gene is mediated by duplicated heat shock elements and heat shock factor 1. Gene 2013; 522: 184-191.
- [8] Hahn JS, Hu Z, Thiele DJ and Iyer VR. Genomewide analysis of the biology of stress responses through heat shock transcription factor. Mol Cell Biol 2004; 24: 5249-5256.
- [9] Mendillo ML, Santagata S, Koeva M, Bell GW, Hu R, Tamimi RM, Fraenkel E, Ince TA, Whitesell L and Lindquist S. HSF1 drives a transcriptional program distinct from heat shock to support highly malignant human cancers. Cell 2012; 150: 549-562.
- [10] Trinklein ND, Murray JI, Hartman SJ, Botstein D and Myers RM. The role of heat shock transcription factor 1 in the genome-wide regulation of the mammalian heat shock response. Mol Biol Cell 2004; 15: 1254-1261.
- [11] Vihervaara A, Sergelius C, Vasara J, Blom MA, Elsing AN, Roos-Mattjus P and Sistonen L. Transcriptional response to stress in the dynamic chromatin environment of cycling and mitotic cells. Proc Natl Acad Sci U S A 2013; 110: E3388-3397.
- [12] Fujimoto M and Nakai A. The heat shock factor family and adaptation to proteotoxic stress. FEBS J 2010; 277: 4112-4125.
- [13] Nover L, Bharti K, Döring P, Mishra SK, Ganguli A and Scharf KD. Arabidopsis and the heat stress transcription factor world: how many heat stress transcription factors do we need? Cell Stress Chaperones 2001; 6: 177-189.
- [14] Peteranderl R and Nelson HC. Trimerization of the heat shock transcription factor by a triplestranded alpha-helical coiled-coil. Biochemistry 1992; 31: 12272-12276.
- [15] Rabindran SK, Haroun RI, Clos J, Wisniewski J and Wu C. Regulation of heat shock factor trimer formation: role of a conserved leucine zipper. Science 1993; 259: 230-234.

- [16] Westwood JT and Wu C. Activation of Drosophila heat shock factor: conformational change associated with a monomer-to-trimer transition. Mol Cell Biol 1993; 13: 3481-3486.
- [17] Eastmond DL and Nelson HC. Genome-wide analysis reveals new roles for the activation domains of the saccharomyces cerevisiae heat shock transcription factor (Hsf1) during the transient heat shock response. J Biol Chem 2006; 281: 32909-32921.
- [18] Anckar J and Sistonen L. Regulation of HSF1 function in the heat stress response: implications in aging and disease. Annu Rev Biochem 2011; 80: 1089-1115.
- [19] Sakurai H and Enoki Y. Novel aspects of heat shock factors: DNA recognition, chromatin modulation and gene expression. FEBS J 2010; 277: 4140-4149.
- [20] Guettouche T, Boellmann F, Lane WS and Voellmy R. Analysis of phosphorylation of human heat shock factor 1 in cells experiencing a stress. BMC Biochem 2005; 6: 4.
- [21] Holmberg Cl, Hietakangas V, Mikhailov A, Rantanen JO, Kallio M, Meinander A, Hellman J, Morrice N, MacKintosh C, Morimoto Rl, Eriksson JE and Sistonen L. Phosphorylation of serine 230 promotes inducible transcriptional activity of heat shock factor 1. EMBO J 2001; 20: 3800-3810.
- [22] Ni Z, Schwartz BE, Werner J, Suarez JR and Lis JT. Coordination of transcription, RNA processing, and surveillance by P-TEFb kinase on heat shock genes. Mol Cell 2004; 16: 55-65.
- [23] Ellis RJ. Protein misassembly: macromolecular crowding and molecular chaperones. Adv Exp Med Biol 2007; 594: 1-13.
- [24] Bukau B, Weissman J and Horwich A. Molecular chaperones and protein quality control. Cell 2006; 125: 443-451.
- [25] Whitesell L and Lindquist S. Inhibiting the transcription factor HSF1 as an anticancer strategy. Expert Opin Ther Targets 2009; 13: 469-478.
- [26] Calderwood SK and Gong J. Molecular chaperones in mammary cancer growth and breast tumor therapy. J Cell Biochem 2012; 113: 1096-1103.
- [27] Calderwood SK, Xie Y, Wang X, Khaleque MA, Chou SD, Murshid A, Prince T and Zhang Y. Signal transduction pathways leading to heat shock transcription. Sign Transduct Insights 2010; 2: 13-24.
- [28] Ciocca DR, Arrigo AP and Calderwood SK. Heat shock proteins and heat shock factor 1 in carcinogenesis and tumor development: an update. Arch Toxicol 2013; 87: 19-48.
- [29] Westwood JT and Wu C. Activation of drosophila heat shock factor: conformational change

associated with a monomer-to-trimer transition. Mol Cell Biol 1993; 13: 3481-6.

- [30] Kampinga HH, Hageman J, Vos MJ, Kubota H, Tanguay RM, Bruford EA, Cheetham ME, Chen B and Hightower LE. Guidelines for the nomenclature of the human heat shock proteins. Cell Stress Chaperones 2009; 14: 105-111.
- [31] Alabert C and Groth A. Chromatin replication and epigenome maintenance. Nat Rev Mol Cell Biol 2012; 13: 153-167.
- [32] Lee YJ, Lee HJ, Lee JS, Jeoung D, Kang CM, Bae S, Lee SJ, Kwon SH, Kang D and Lee YS. A novel function for HSF1-induced mitotic exit failure and genomic instability through direct interaction between HSF1 and Cdc20. Oncogene 2008; 27: 2999-3009.
- [33] Dai C, Whitesell L, Rogers AB and Lindquist S. Heat shock factor 1 is a powerful multifaceted modifier of carcinogenesis. Cell 2007; 130: 1005-1018.
- [34] Meng L, Gabai VL and Sherman MY. Heatshock transcription factor HSF1 has a critical role in human epidermal growth factor receptor-2-induced cellular transformation and tumorigenesis. Oncogene 2010; 29: 5204-5213.
- [35] Min JN, Huang L, Zimonjic DB, Moskophidis D and Mivechi NF. Selective suppression of lymphomas by functional loss of Hsf1 in a p53deficient mouse model for spontaneous tumors. Oncogene 2007; 26: 5086-5097.
- [36] Santagata S, Xu YM, Wijeratne EM, Kontnik R, Rooney C, Perley CC, Kwon H, Clardy J, Kesari S, Whitesell L, Lindquist S and Gunatilaka AA. Using the heat-shock response to discover anticancer compounds that target protein homeostasis. ACS Chem Biol 2012; 7: 340-349.
- [37] Zhao Y, Liu H, Liu Z, Ding Y, Ledoux SP, Wilson GL, Voellmy R, Lin Y, Lin W, Nahta R, Liu B, Fodstad O, Chen J, Wu Y, Price JE and Tan M. Overcoming trastuzumab resistance in breast cancer by targeting dysregulated glucose metabolism. Cancer Res 2011; 71: 4585-4597.
- [38] Huang J, Nueda A, Yoo S and Dynan WS. Heat shock transcription factor 1 binds selectively in vitro to Ku protein and the catalytic subunit of the DNA-dependent protein kinase. J Biol Chem 1997; 272: 26009-26016.
- [39] Nakamura Y, Fujimoto M, Hayashida N, Takii R, Nakai A and Muto M. Silencing HSF1 by short hairpin RNA decreases cell proliferation and enhances sensitivity to hyperthermia in human melanoma cell lines. J Dermatol Sci 2010; 60: 187-192.
- [40] Santagata S, Mendillo ML, Tang YC, Subramanian A, Perley CC, Roche SP, Wong B, Narayan R, Kwon H, Koeva M, Amon A, Golub TR, Porco JA Jr, Whitesell L and Lindquist S. Tight coordination of protein translation and HSF1 activa-

tion supports the anabolic malignant state. Science 2013; 341: 1238303.

- [41] Murshid A, Chou SD, Prince T, Zhang Y, Bharti A and Calderwood SK. Protein kinase A binds and activates heat shock factor 1. PLoS One 2010; 5: e13830.
- [42] Nueda A, Hudson F, Mivechi NF and Dynan WS. DNA-dependent protein kinase protects against heat-induced apoptosis. J Biol Chem 1999; 274: 14988-14996.
- [43] Fang F, Chang R and Yang L. Heat shock factor 1 promotes invasion and metastasis of hepatocellular carcinoma in vitro and in vivo. Cancer 2012; 118: 1782-1794.
- [44] Ishiwata J, Kasamatsu A, Sakuma K, Iyoda M, Yamatoji M, Usukura K, Ishige S, Shimizu T, Yamano Y, Ogawara K, Shiiba M, Tanzawa H and Uzawa K. State of heat shock factor 1 expression as a putative diagnostic marker for oral squamous cell carcinoma. Int J Oncol 2012; 40: 47-52.
- [45] Jin X, Moskophidis D and Mivechi NF. Heat shock transcription factor 1 is a key determinant of HCC development by regulating hepatic steatosis and metabolic syndrome. Cell Metab 2011; 14: 91-103.
- [46] Santagata S, Hu R, Lin NU, Mendillo ML, Collins LC, Hankinson SE, Schnitt SJ, Whitesell L, Tamimi RM, Lindquist S and Ince TA. High levels of nuclear heat-shock factor 1 (HSF1) are associated with poor prognosis in breast cancer. Proc Natl Acad Sci U S A 2011; 108: 18378-18383.
- [47] Khaleque MA, Bharti A, Gong J, Gray PJ, Sachdev V, Ciocca DR, Stati A, Fanelli M and Calderwood SK. Heat shock factor 1 represses estrogen-dependent transcription through association with MTA1. Oncogene 2008; 27: 1886-1893.
- [48] Zhao YH, Zhou M, Liu H, Ding Y, Khong HT, Yu D, Fodstad O and Tan M. Upregulation of lactate dehydrogenase A by ErbB2 through heat shock factor 1 promotes breast cancer cell glycolysis and growth. Oncogene 2009; 28: 3689-3701.
- [49] Wang JH, Yao MZ, Gu JF, Sun LY, Shen YF and Liu XY. Blocking HSF1 by dominant-negative mutant to sensitize tumor cells to hyperthermia. Biochem Biophys Res Commun 2002; 290: 1454-1461.
- [50] Wang C, Dai Z, Fan R, Deng Y, Lv G and Lu G. HSF1 overexpression enhances oncolytic effect of replicative adenovirus. J Transl Med 2010; 8: 44.
- [51] Wang B, Lee CW, Witt A, Thakkar A and Ince TA. Heat shock factor 1 induces cancer stem cell phenotype in breast cancer cell lines. Breast Cancer Res Treat 2015; 153: 57-66.
- [52] Schilling D, Kühnel A, Tetzlaff F, Konrad S and Multhoff G. NZ28-induced inhibition of HSF1,

SP1 and NF-kappaB triggers the loss of the natural killer cell-activating ligands MICA/B on human tumor cells. Cancer Immunol Immuno-ther 2015; 64: 599-608.

- [53] Schilling D, Kühnel A, Konrad S, Tetzlaff F, Bayer C, Yaglom J and Multhoff G. Sensitizing tumor cells to radiation by targeting the heat shock response. Cancer Lett 2015; 360: 294-301.
- [54] Zhang X, Mukerji R, Samadi AK and Cohen MS. Down-regulation of estrogen receptor-alpha and rearranged during transfection tyrosine kinase is associated with withaferin a-induced apoptosis in MCF-7 breast cancer cells. BMC Complement Altern Med 2011; 11: 84.
- [55] Cheng Q, Chang JT, Geradts J, Neckers LM, Haystead T, Spector NL and Lyerly HK. Amplification and high-level expression of heat shock protein 90 marks aggressive phenotypes of human epidermal growth factor receptor 2 negative breast cancer. Breast Cancer Res 2012; 14: R62.
- [56] Schulz R, Streller F, Scheel AH, Rüschoff J, Reinert MC, Dobbelstein M, Marchenko ND and Moll UM. HER2/ErbB2 activates HSF1 and thereby controls HSP90 clients including MIF in HER2-overexpressing breast cancer. Cell Death Dis 2014; 5: e980.
- [57] Xi C, Hu Y, Buckhaults P, Moskophidis D and Mivechi NF. Heat shock factor Hsf1 cooperates with ErbB2 (Her2/Neu) protein to promote mammary tumorigenesis and metastasis. J Biol Chem 2012; 287: 35646-35657.
- [58] Huang AJ, Yu KD, Li J, Fan L and Shao ZM. Polymorphism rs4919510:C>G in mature sequence of human microRNA-608 contributes to the risk of HER2-positive breast cancer but not other subtypes. PLoS One 2012; 7: e35252.
- [59] Chou SD, Murshid A, Eguchi T, Gong J and Calderwood SK. HSF1 regulation of betacatenin in mammary cancer cells through control of HuR/elavL1 expression. Oncogene 2015; 34: 2178-2188.
- [60] Kim HB, Lee SH, Um JH, Oh WK, Kim DW, Kang CD and Kim SH. Sensitization of multidrug-resistant human cancer cells to Hsp90 inhibitors by down-regulation of SIRT1. Oncotarget 2015; 6: 36202-36218.
- [61] Yang X, Wang J, Liu S and Yan Q. HSF1 and Sp1 regulate FUT4 gene expression and cell proliferation in breast cancer cells. J Cell Biochem 2014; 115: 168-178.
- [62] Sarkars R, Mukherjee S and Roy M. Targeting heat shock proteins by phenethyl isothiocyanate results in cell-cycle arrest and apoptosis of human breast cancer cells. Nutr Cancer 2013; 65: 480-493.
- [63] Desai S, Liu Z, Yao J, Patel N, Chen J, Wu Y, Ahn EE, Fodstad O and Tan M. Heat shock factor 1

(HSF1) controls chemoresistance and autophagy through transcriptional regulation of autophagy-related protein 7 (ATG7). J Biol Chem 2013; 288: 9165-9176.

- [64] Sarkar R, Mukherjee S, Biswas J and Roy M. Sulphoraphane, a naturally occurring isothiocyanate induces apoptosis in breast cancer cells by targeting heat shock proteins. Biochem Biophys Res Commun 2012; 427: 80-85.
- [65] Xia W, Hardy L, Liu L, Zhao S, Goodman M, Voellmy R and Spector NL. Concurrent exposure to heat shock and H7 synergizes to trigger breast cancer cell apoptosis while sparing normal cells. Breast Cancer Res Treat 2003; 77: 233-243.
- [66] Wang JH, Yao MZ, Zhang ZL, Zhang YH, Wang YG and Liu XY. HSF1 blockade-induced tumor thermotolerance abolishment is mediated by JNK-dependent caspase-3 activation. Biochem Biophys Res Commun 2004; 321: 736-745.
- [67] Ciocca DR, Gago FE, Fanelli MA and Calderwood SK. Co-expression of steroid receptors (estrogen receptor alpha and/or progesterone receptors) and Her-2/neu: clinical implications. J Steroid Biochem Mol Biol 2006; 102: 32-40.
- [68] Zhu T, Chen R, Li AP, Gu DA, Liu QZ and Zhou JW. Expression of novel environmental responsive protein JWA involved in the oxidative stress responsiveness in MCF-7 cells. Zhonghua Lao Dong Wei Sheng Zhi Ye Bing Za Zhi 2005; 23: 122-124.
- [69] Hansen RK, Oesterreich S, Lemieux P, Sarge KD and Fuqua SA. Quercetin inhibits heat shock protein induction but not heat shock factor DNA-binding in human breast carcinoma cells. Biochem Biophys Res Commun 1997; 239: 851-856.
- [70] Fang YM, Dong Q, Cen H, Tang XP and Zheng S. Induction of HSF1 expression and sporadic colorectal cancer. Zhejiang Da Xue Xue Bao Yi Xue Ban 2004; 33: 390-394.
- [71] Benderska N, Ivanovska J, Rau TT, Schulze-Luehrmann J, Mohan S, Chakilam S, Gandesiri M, Ziesché E, Fischer T, Söder S, Agaimy A, Distel L, Sticht H, Mahadevan V and Schneider-Stock R. DAPK-HSF1 interaction as a positivefeedback mechanism stimulating TNF-induced apoptosis in colorectal cancer cells. J Cell Sci 2014; 127: 5273-5287.
- [72] Dayanc BE, Bansal S, Gure AO, Gollnick SO and Repasky EA. Enhanced sensitivity of colon tumour cells to natural killer cell cytotoxicity after mild thermal stress is regulated through HSF1mediated expression of MICA. Int J Hyperthermia 2013; 29: 480-490.
- [73] Wales CT, Taylor FR, Higa AT, McAllister HA and Jacobs AT. ERK-dependent phosphorylation of HSF1 mediates chemotherapeutic resistance

to benzimidazole carbamates in colorectal cancer cells. Anticancer Drugs 2015; 26: 657-666.

- [74] Kim JA, Kim Y, Kwon BM and Han DC. The natural compound cantharidin induces cancer cell death through inhibition of heat shock protein 70 (HSP70) and Bcl-2-associated athanogene domain 3 (BAG3) expression by blocking heat shock factor 1 (HSF1) binding to promoters. J Biol Chem 2013; 288: 28713-28726.
- [75] Akagi R, Ohno M, Matsubara K, Fujimoto M, Nakai A and Inouye S. Glutamine protects intestinal barrier function of colon epithelial cells from ethanol by modulating Hsp70 expression. Pharmacology 2013; 91: 104-111.
- [76] Fan R, Wang C, Wang Y, Ren P, Gan P, Ji H, Xia Z, Hu S, Zeng Q, Huang W, Jiang Y and Huang X. Enhanced antitumoral efficacy and immune response following conditionally replicative adenovirus containing constitutive HSF1 delivery to rodent tumors. J Transl Med 2012; 10: 101.
- [77] Kim JA, Lee S, Kim DE, Kim M, Kwon BM and Han DC. Fisetin, a dietary flavonoid, induces apoptosis of cancer cells by inhibiting HSF1 activity through blocking its binding to the hsp70 promoter. Carcinogenesis 2015; 36: 696-706.
- [78] Chen Y, Chen J, Yu J, Yang G, Temple E, Harbinski F, Gao H, Wilson C, Pagliarini R and Zhou W. Identification of mixed lineage leukemia 1(MLL1) protein as a coactivator of heat shock factor 1(HSF1) protein in response to heat shock protein 90 (HSP90) inhibition. J Biol Chem 2014; 289: 18914-18927.
- [79] Jacobs AT and Marnett LJ. HSF1-mediated BAG3 expression attenuates apoptosis in 4-hydroxynonenal-treated colon cancer cells via stabilization of anti-apoptotic Bcl-2 proteins. J Biol Chem 2009; 284: 9176-9183.
- [80] Kourtis N, Moubarak RS, Aranda-Orgilles B, Lui K, Aydin IT, Trimarchi T, Darvishian F, Salvaggio C, Zhong J, Bhatt K, Chen EI, Celebi JT, Lazaris C, Tsirigos A, Osman I, Hernando E and Aifantis I. FBXW7 modulates cellular stress response and metastatic potential through HSF1 posttranslational modification. Nat Cell Biol 2015; 17: 322-332.
- [81] Gentilella A and Khalili K. BAG3 expression in glioblastoma cells promotes accumulation of ubiquitinated clients in an Hsp70-dependent manner. J Biol Chem 2011; 286: 9205-9215.
- [82] Oh SY, Kim JH, Park MJ, Kim SM, Yoon CS, Joo YM, Park JS, Han SI, Park HG and Kang HS. Induction of heat shock protein 72 in C6 glioma cells by methyl jasmonate through ROSdependent heat shock factor 1 activation. Int J Mol Med 2005; 16: 833-839.
- [83] Liu Y, Zheng T, Zhao S, Liu H, Han D, Zhen Y, Xu D, Wang Y, Yang H, Zhang G, Wang C, Wu J and Ye Y. Inhibition of heat shock protein response

enhances PS-341-mediated glioma cell death. Ann Surg Oncol 2012; 19 Suppl 3: S421-429.

- [84] Grogan PT, Sarkaria JN, Timmermann BN and Cohen MS. Oxidative cytotoxic agent withaferin A resensitizes temozolomide-resistant glioblastomas via MGMT depletion and induces apoptosis through Akt/mTOR pathway inhibitory modulation. Invest New Drugs 2014; 32: 604-617.
- [85] Ohnishi K, Wang X, Takahashi A, Matsumoto H and Ohnishi T. The protein kinase inhibitor, H-7, suppresses heat induced activation of heat shock transcription factor 1. Mol Cell Biochem 1999; 197: 129-135.
- [86] Zhao J, Wang S, Liu N and Tang X. Correlation between the expression of Id-1 and hyperthermia-associated molecules in oral squamous cell carcinoma. J Clin Pathol 2013; 66: 758-763.
- [87] Zhang JB, Guo K, Sun HC, Zhu XD, Zhang B, Lin ZH, Zhang BH, Liu YK, Ren ZG and Fan J. Prognostic value of peritumoral heat-shock factor-1 in patients receiving resection of hepatocellular carcinoma. Br J Cancer 2013; 109: 1648-1656.
- [88] Chen Y, Chen J, Loo A, Jaeger S, Bagdasarian L, Yu J, Chung F, Korn J, Ruddy D, Guo R, McLaughlin ME, Feng F, Zhu P, Stegmeier F, Pagliarini R, Porter D and Zhou W. Targeting HSF1 sensitizes cancer cells to HSP90 inhibition. Oncotarget 2013; 4: 816-829.
- [89] Sharma A, Meena AS and Bhat MK. Hyperthermia-associated carboplatin resistance: differential role of p53, HSF1 and Hsp70 in hepatoma cells. Cancer Sci 2010; 101: 1186-1193.
- [90] Chuma M, Sakamoto N, Nakai A, Hige S, Nakanishi M, Natsuizaka M, Suda G, Sho T, Hatanaka K, Matsuno Y, Yokoo H, Kamiyama T, Taketomi A, Fujii G, Tashiro K, Hikiba Y, Fujimoto M, Asaka M and Maeda S. Heat shock factor 1 accelerates hepatocellular carcinoma development by activating nuclear factor-kappaB/mitogen-activated protein kinase. Carcinogenesis 2014; 35: 272-281.
- [91] Xu R, Zhang X, Zhang W, Fang Y, Zheng S and Yu XF. Association of human APOBEC3 cytidine deaminases with the generation of hepatitis virus B x antigen mutants and hepatocellular carcinoma. Hepatology 2007; 46: 1810-1820.
- [92] Li Y, Xu D, Bao C, Zhang Y, Chen D, Zhao F, Ding J, Liang L, Wang Q, Liu L, Li J, Yao M, Huang S and He X. MicroRNA-135b, a HSF1 target, promotes tumor invasion and metastasis by regulating RECK and EVI5 in hepatocellular carcinoma. Oncotarget 2015; 6: 2421-2433.
- [93] Lee JH, Lee YK, Lim JJ, Byun HO, Park I, Kim GH, Xu WG, Wang HJ and Yoon G. Mitochondrial respiratory dysfunction induces claudin-1

expression via reactive oxygen species-mediated heat shock factor 1 activation, leading to hepatoma cell invasiveness. J Biol Chem 2015; 290: 21421-21431.

- [94] Ma W, Zhang Y, Mu H, Qing X, Li S, Cui X, Lou Q, Ma Y, Pu H and Hu Y. Glucose regulates heat shock factor 1 transcription activity via mTOR pathway in HCC cell lines. Cell Biol Int 2015; 39: 1217-1224.
- [95] Newman B, Liu Y, Lee HF, Sun D and Wang Y. HSP90 inhibitor 17-AAG selectively eradicates lymphoma stem cells. Cancer Res 2012; 72: 4551-4561.
- [96] Jin Y, Zhen Y, Haugsten EM and Wiedlocha A. The driver of malignancy in KG-1a leukemic cells, FGFR10P2-FGFR1, encodes an HSP90 addicted oncoprotein. Cell Signal 2011; 23: 1758-1766.
- [97] Chakraborty PK, Mustafi SB, Ganguly S, Chatterjee M and Raha S. Resveratrol induces apoptosis in K562 (chronic myelogenous leukemia) cells by targeting a key survival protein, heat shock protein 70. Cancer Sci 2008; 99: 1109-1116.
- [98] Takaki E, Fujimoto M, Sugahara K, Nakahari T, Yonemura S, Tanaka Y, Hayashida N, Inouye S, Takemoto T, Yamashita H and Nakai A. Maintenance of olfactory neurogenesis requires HSF1, a major heat shock transcription factor in mice. J Biol Chem 2006; 281: 4931-4937.
- [99] Ganguly S, Home T, Yacoub A, Kambhampati S, Shi H, Dandawate P, Padhye S, Saluja AK, Mc-Guirk J and Rao R. Targeting HSF1 disrupts HSP90 chaperone function in chronic lymphocytic leukemia. Oncotarget 2015; 6: 31767-31779.
- [100] Kim HB, Lee SH, Um JH, Kim MJ, Hyun SK, Gong EJ, Oh WK, Kang CD and Kim SH. Sensitization of chemo-resistant human chronic myeloid leukemia stem-like cells to Hsp90 inhibitor by SIRT1 inhibition. Int J Biol Sci 2015; 11: 923-934.
- [101] Kim HR, Kang HS and Kim HD. Geldanamycin induces heat shock protein expression through activation of HSF1 in K562 erythroleukemic cells. IUBMB Life 1999; 48: 429-433.
- [102] Yoshima T, Yura T and Yanagi H. Heat shock factor 1 mediates hemin-induced hsp70 gene transcription in K562 erythroleukemia cells. J Biol Chem 1998; 273: 25466-25471.
- [103] Sarkar R, Mukherjee A, Mukherjee S, Biswas R, Biswas J and Roy M. Curcumin augments the efficacy of antitumor drugs used in leukemia by modulation of heat shock proteins via HDAC6. J Environ Pathol Toxicol Oncol 2014; 33: 247-263.
- [104] Cui J, Tian H and Chen G. Upregulation of nuclear heat shock factor 1 contributes to tumor angiogenesis and poor survival in patients with

non-small cell lung cancer. Ann Thorac Surg 2015; 100: 465-472.

- [105] Qiu Y, Ye X, Hanson PJ, Zhang HM, Zong J, Cho B and Yang D. Hsp70-1: upregulation via selective phosphorylation of heat shock factor 1 during coxsackieviral infection and promotion of viral replication via the AU-rich element. Cell Mol Life Sci 2016; 73: 1067-1084.
- [106] Chang YS, Lee LC, Sun FC, Chao CC, Fu HW and Lai YK. Involvement of calcium in the differential induction of heat shock protein 70 by heat shock protein 90 inhibitors, geldanamycin and radicicol, in human non-small cell lung cancer H460 cells. J Cell Biochem 2006; 97: 156-165.
- [107] Kim SY, Lee HJ, Nam JW, Seo EK and Lee YS. Coniferyl aldehyde reduces radiation damage through increased protein stability of heat shock transcriptional factor 1 by phosphorylation. Int J Radiat Oncol Biol Phys 2015; 91: 807-816.
- [108] Ma X, Xu L, Alberobello AT, Gavrilova O, Bagattin A, Skarulis M, Liu J, Finkel T and Mueller E. Celastrol protects against obesity and metabolic dysfunction through activation of a HSF1-PGC1alpha transcriptional axis. Cell Metab 2015; 22: 695-708.
- [109] Yoon T, Kang GY, Han AR, Seo EK and Lee YS. 2,4-Bis(4-hydroxybenzyl)phenol inhibits heat shock transcription factor 1 and sensitizes lung cancer cells to conventional anticancer modalities. J Nat Prod 2014; 77: 1123-1129.
- [110] Hoang AT, Huang J, Rudra-Ganguly N, Zheng J, Powell WC, Rabindran SK, Wu C and Roy-Burman P. A novel association between the human heat shock transcription factor 1 (HSF1) and prostate adenocarcinoma. Am J Pathol 2000; 156: 857-864.
- [111] Wang Y, Theriault JR, He H, Gong J and Calderwood SK. Expression of a dominant negative heat shock factor-1 construct inhibits aneuploidy in prostate carcinoma cells. J Biol Chem 2004; 279: 32651-32659.
- [112] Kaija H, Pakanen L, Kortelainen ML and Porvari K. Hypothermia and rewarming induce gene expression and multiplication of cells in healthy rat prostate tissue. PLoS One 2015; 10: 127854.

- [113] Granato M, Chiozzi B, Filardi MR, Lotti LV, Di Renzo L, Faggioni A and Cirone M. Tyrosine kinase inhibitor tyrphostin AG490 triggers both apoptosis and autophagy by reducing HSF1 and Mcl-1 in PEL cells. Cancer Lett 2015; 366: 191-197.
- [114] Kumar S, Deepak P, Kumar S, Gautam PK and Acharya A. A benzophenanthridine alkaloid, chelerythrine induces apoptosis in vitro in a Dalton's lymphoma. J Cancer Res Ther 2013; 9: 693-700.
- [115] Dudeja V, Chugh RK, Sangwan V, Skube SJ, Mujumdar NR, Antonoff MB, Dawra RK, Vickers SM and Saluja AK. Prosurvival role of heat shock factor 1 in the pathogenesis of pancreatobiliary tumors. Am J Physiol Gastrointest Liver Physiol 2011; 300: 948-955.
- [116] Banerjee S, Sangwan V, McGinn O, Chugh R, Dudeja V, Vickers SM and Saluja AK. Triptolideinduced cell death in pancreatic cancer is mediated by O-GlcNAc modification of transcription factor Sp1. J Biol Chem 2013; 288: 33927-33938.
- [117] Sangwan V, Banerjee S, Jensen KM, Chen Z, Chugh R, Dudeja V, Vickers SM and Saluja AK. Primary and liver metastasis-derived cell lines from KrasG12D; Trp53R172H; Pdx-1 Cre animals undergo apoptosis in response to triptolide. Pancreas 2015; 44: 583-589.
- [118] Tokunaga M, Baron B, Kitagawa T, Tokuda K and Kuramitsu Y. Active hexose-correlated compound down-regulates heat shock factor 1, a transcription factor for HSP27, in gemcitabine-resistant human pancreatic cancer cells. Anticancer Res 2015; 35: 6063-6067.
- [119] Tan J, Tan S, Zheng H, Liu M, Chen G, Zhang H, Wang K, Tan S, Zhou J and Xiao XZ. HSF1 functions as a transcription regulator for Dp71 expression. Cell Stress Chaperones 2015; 20: 371-379.