# Review Article PDGF signaling in cancer progression

Feng Huang<sup>1</sup>, Dong Wang<sup>2</sup>, Yongliang Yao<sup>1</sup>, Mei Wang<sup>3</sup>

<sup>1</sup>Department of Clinical Laboratory, The First People's Hospital of Kunshan, Affiliated to Jiangsu University, Suzhou, Jiangsu Province, China; <sup>2</sup>Department of Burn and Plastic Surgery, Affiliated Hospital of Nantong University, Nantong, China; <sup>3</sup>School of Medicine, Jiangsu University, Zhenjiang, Jiangsu Province, China

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**Abstract:** Platelet-derived growth factor (PDGF) is a pro-angiogenic factor that was isolated from human platelets. PDGF family is comprised of four different polypeptide chains encoded by different genes, which have been identified: PDGF-A, PDGF-B, and recently discovered PDGF-C and PDGF-D. PDGF is a variety of strong mesenchymal cell mitogenic agents and growth chemokines. PDGF, acting as a vascular endothelial growth factor, is closely associated with tumor development. PDGF signaling pathway has been extensively studied and well characterized because PDGF can regulate many cellular processes, including cell proliferation, migration, invasion, angiogenesis and metastasis. According to the available data, PDGF plays an important role in the development of breast cancer, stomach cancer, gastric cancer, prostate cancer, lung cancer, colon cancer, and other cancers. Effects of PDGF on cancer are complex and diverse, and further analysis is needed to confirm the predictive and therapeutic values of PDGF and its receptor. This article reviews the recent progress of PDGF in tumor progression.

**Keywords:** Platelet-derived growth factor, signal pathway, proliferation, migration, invasion, angiogenesis, metastasis, tumor progression

#### Introduction

Platelet-derived growth factor (PDGF) is a proangiogenic factor that was isolated from human platelets [1-3]. PDGFs are a variety of strong mesenchymal cell mitogenic agents and growth chemokines; they are also important modifiers for the normal and pathological vascular development [4-6]. PDGF, acting as vascular endothelial growth factor, plays a pivotal role in the development and progression of human malignancies. The latest findings show that PDGFs regulate tumor growth and metastasis by targeting malignant cells, vascular cells, and stromal cells [7-10].

#### PDGF and cancer cells

#### Cancer cell proliferation

PDGF stimulates cancer cell proliferation through autocrine and paracrine manners, since some cancer cells express the PDGF receptor and some do not. For example, Ustach *et al.* [11, 12] demonstrated that LNCaP cells auto-activate latent PDGFD into the active PDGF domain, which can induce the phosphorylation of β-PDGF receptor and stimulate LNCaP cell proliferation in an autocrine manner. Additionally, LNCaP-PDGFD-conditioned medium induces migration of the prostate fibroblast cell line 1532-FTX, indicating LNCaP-processed PDGFD acts in a paracrine manner as well. The PDGF signaling pathway combines with a variety of substrate protein signaling molecules, including SRC tyrosine kinase subfamily, phospholipase C (PLC-y), phosphatidylinositol 3-kinase (PI3-K), GTPase activation protein (RAS-gap), growth factor binding protein (Grb2), tyrosine specific phosphatase (SYP), Src homology and cross-linked protein (SHC) and adaptation (Crk) protein, and non-receptor tyrosine kinase family (SRC) and so on, and forms a variety of signaling pathways, such as RAS-MAPK pathway, PI3-K pathway, the PLC pathway, and STAT pathway [13]. These signal molecules mediate cytoplasmic complex signaling networks, and play a role in cell membrane serine threonine residues, resulting in a variety of gene regulation and protein phosphorylation, thus promoting cancer cell growth and division

[14]. For example, PDGF has been proved to promote the proliferation of human meningiomas and mesothelioma cell through PI3-K pathway [15, 16]. Lu Y. demonstrated that hypoxia-induced PDGF-BB secretion by HCC cells stimulates HSCs to accumulate and proliferate in the tumor stroma [17]. Whereas angiogenesis inhibitor sorafenib downregulates the expression of PDGF-BB and TGF-B1 in the HSCs supernatant, and restrains the viability of the HSCs, resulting in suppressed proliferation and invasion in HepG2 cells [18]. Meanwhile, Platelet-derived growth factor receptor (PDGFR) signaling participates in different processes in solid tumors, including autocrine stimulation of tumor cell growth, recruitment of tumor stroma fibroblasts, and stimulation of tumor angiogenesis [19]. Moreover, malignant melanoma upregulates hyaluronan synthesis in fibroblasts by releasing PDGF-AA and PDGF-CC, which in turn stimulates the malignant melanoma cell proliferation in a paracrine manner [20]. PDGF also increases proliferation of Luminal breast cancer cells in the absence of estrogens [21]. In conclusion, PDGF induces proliferation and migration of tumor cells and inhibits apoptosis [21].

### Cancer cells metastasis

Cancer cell metastasis begins with detachment of metastatic cells from the primary tumor, traveling of the cells to different sites through blood/lymphatic vessels, settlement and growth of the cells at a distal site [22]. During the process, metastatic cells go through detachment, migration, invasion, and adhesion [22]. For tumor metastasis, cardiovascular generation is required, which provides oxygen and nutrients for tumor growth and metastasis. PDGF can promote connective tissue to produce extracellular matrix proteins, which directly or indirectly induces tumor angiogenesis, significantly affecting tumor metastasis. PDGF-D-overexpressing tumors express higher levels of MMP-9 and PDGFD to induce renal cancer cells proliferation and migration, thus providing a molecular mechanism for the higher incidence of lung metastasis and pericyte coverage observed in PDGF-D-overexpressing tumors [23, 24]. PDGFD plays an important role in breast tumor aggressiveness and this process is mechanistically linked with the activation of Notch and NF-kB signaling [25]. A grow-

ing number of researchers believe that cancer growth, metastasis, and invasion depend not only on the tumor cells themselves, but also on the growth of tumor microenvironment, which supports cancer behavior. Studies have shown that PDGF-C in breast cancer activates the receptor by binding to the receptor, and activation of the receptor leads to tyrosine kinase pathway activation, which mediates fibroblast activation and is involved in the secretion of a variety of cytokines and the formation of cellular microenvironment [26]. Activated fibroblasts, known as cancer-associated fibroblasts (CAFs), promote the formation of tumor metastasis and directional vessel by secreting SDF-1/ CXCR4. CAFs also can promote tumor growth and metastasis by secreting other factors such as hepatocyte growth factor (HGF), epidermal growth factor (EGF), basic fibroblast growth factor (b FGF), and insulin-like growth factor (IGF). PDGF-DD secreted by gastric cancer-derived mesenchymal stem cells (GC-MSCs) is capable of promoting gastric cancer cell proliferation and migration in vitro and in vivo. Jieqiong Liu et al. [27] reported that overexpression of PDGF-D promoted tumor growth and lymph node metastasis through increased proliferation, decreased apoptosis, and induction of CXCR4 expression [28]. A study reports a mechanism of the interaction between perivascular cells and tumor-associated macrophages (TAMs) that promotes metastasis through the IL-33-ST2-dependent pathway in xenograft mouse models of cancer. While PDGF-BB upregulates IL-33 gene though stimulating the activation of pericytes SOX7 transcription factor. Gain- and loss-of-function experiments validated that IL-33 promotes metastasis through recruitment of TAMs. Pharmacological inhibition of the IL-33-ST2 signaling by a soluble ST2 significantly inhibits TAMs and metastasis [29]. Genetic deletion of host IL-33 in mice blocks PDGF-BB-induced TAM recruitment and metastasis [29]. Yasuhiko Kitadai discovered that expression and phosphorylation of PDGF-Rb by stromal cells and pericytes was higher in orthotopic tumors than in ectopic tumors and, therefore, was associated with the metastatic potential of the neoplasms [30]. PDGFs may also have a role in determining the preferential organ of metastatization. Indeed, PDGFs released by the tumor cells are potent chemoattractants and mitogens for host mesenchymal cells and could mediate the interactions between cancer cells and the host environment of the preferred metastatic site [31]. PDGF plays an important role in epithelial mesenchymal transition. For example, it has been recently demonstrated that autocrine PDGF signaling maintains EMT and promotes metastatization in mouse mammary carcinoma and tumor dissemination [32]. PDGF promotes EMT via activation of STAT3 or PI3K pathway and PDGF-D over-expression was positively correlated with the expression of mesenchymal markers (vimentin and ZEB-2) in concomitant with expression of epithelial marker E-cadherin [25, 33, 34]. As is well known, epithelial-mesenchymal transition (EMT) enables the escape of epithelial cells from the rigid structural constraints of the tissue architecture to a phenotype more amenable to cell migration and, therefore, invasion and metastasis. All in all, PDGF affects tumor metastasis in many ways.

#### Chemotherapy resistance of cancer cells

Previous literature reported that PDGF is closely associated with chemotherapy resistance of cancer cells. A line of evidence demonstrates that chemo-resistance is associated with the acquisition of epithelial-mesenchymal transition (EMT) of cancer cells, and platelet-derived growth factor-D (PDGF-D) signaling pathway plays a critical role in the acquisition of EMT phenotype of GR HCC cells [35]. Rui Wang et al. demonstrates that overexpression of PDGF-D in gemcitabine-resistant (GR) HCC cells markedly inhibited miR-106a expression and subsequently upregulated Twist1 expression, whereas down-regulation of Twist1 reverses EMT to MET (mesenchymal-epithelial transition) in GR cells [36]. Some studies show that PDGF-C can promote tumor angiogenesis and tumor cell growth by upregulating VEGF level and activating VEGF independent angiogenesis pathway, which can produce VEGF-independent drug resistance [37]. VEGF inhibitors reduce the expression of VEGF and VEGFR, which can only inhibit VEGF-dependent angiogenic signal transduction pathways, while VEGF independent pathway is not suppressed, therefore anti-VEGF therapy can produce resistance. PDGFC plays an important role in drug resistance. It can promote angiogenesis through vascular cell pathway, inflammatory cell pathway, extracellular matrix pathway, but does not depend on VEGF. Exiting evidence demonstrates that the expression of platelet derived growth factor B can promote the cell recruitment in peripheral blood vessel wall and increase the stability of the structure [38]. The recruitment of peripheral blood cells increased blood vessel coverage which may lead to the resistance to anti-angiogenic drugs. In the study of Zhao Z *et al.*, PDGFD is one of the key candidate genes which may influence chemo sensitivity of glioblastoma (GBM) to Semustine (Me-CCNU) [39].

#### PDGF and cancer stroma or cancer microenvironment

### PDGF and extracellular matrix (ECM)

The occurrence, development, invasion and metastasis of malignant tumors are often accompanied by changes in the expression of extracellular matrix (ECM) and their cell surface receptors [40]. The extracellular matrix can provide the raw materials for tumor metastasis, and the transformation of extracellular matrix components [41]. The extracellular matrix plays an important role in tissue homeostasis, therefore, it is crucial in tumorigenesis, development and metastasis [42]. PDGF has a close relationship with extracellular matrix.

Growth factors and the extracellular matrix have been shown to play important developmental roles in many embryonic systems. In the study of PDGF-AA and -BB homodimer isoforms, M. Pekny found that the paracrine activity of PDGF-AA and PDGF-BB homodimers are significantly different, and the local effect of PDGF-BB on tumor growth has greatly increased in the tumor when compared with PDGF-AA, which is very likely associated to the components of the extracellular matrix [43]. According to the latest research results of Wright JH et al., receptors of PDGF-cc are localized on hepatic stellate cells (HSCS) that transform into myofibroblast-like cells that deposit in extracellular matrix (ECM) and promote the growth and transformation of the growth factor [44]. In different stages of tumor growth, the expression of extracellular matrix is enhanced and its distribution is different [45]. Extracellular matrix has a value in predicting the biological behavior of the tumor.

### PDGF and angiogenesis

PDGF regulates tumor growth in tumor development, and a variety of environmental factors that can induce cell expression of PDGF stimu-

lates tumor growth. In vivo experiments confirmed that PDGF-mediated tumor angiogenesis provides nourishment for tumor growth [46]. At the same time, tumor blood vessels also provide a convenient route for the tumor cells to transfer tumor development-related signals, thus angiogenesis is a hallmark of cancer. Angiogenesis is one of the signs of advanced cancer, promoting the invasion and metastasis of cancer. Lu Y et al. found that PDGF-BB expression on HepG2 cells increased significantly under hypoxia. They show that hypoxiainduced liver cancer cell PDGF-BB stimulates the accumulation of HSC substance in the tumor stroma, enhances the expression of VEGF-A in hematopoietic stem cells, and promote angiogenesis [47]. Existing evidence show that PDGF promote angiogenesis primarily by aggregating pericytes which is a kind of mural cells for microcirculation. Pericytes and endothelial cells have a specific contact, and pericytes and endothelial cells have specific contacts at the vascular basement membrane [48, 49]. Many evidences have showed that PDGF/PDGFR-β, involve in the regulation of pericytes recruitment, the mechanisms governing pericytes migration and regulating angiogenesis, especially in cancers [50]. PDGF-D is the most recently discovered member of the PDGF family, regulating systemic arterial blood pressure, and suggests a role in maintaining vascular homeostasis [51]. Sennino found that new DNA aptamer AX102 is capable of specifically inhibiting PDGF-BB signal, resulting in tumor vascular pericytes loss and degradation of the tumor vasculature [52]. The impact of PDGF on pericytes is great, and PDGFR kinase inhibitors can reduce the accumulation of pericytes, which leads to reduced pericyte coverage and tumor blood vessel growth [53].

### PDGF and stromal cells

Macrophages in the tumor microenvironment are key regulators of the immune response. The type of tumor associated macrophages was increasingly reported, and the research on the role of macrophages and PDGF are constantly advanced. Macrophage-derived PDGF can develop a series of factors to promote growth and development of tumor blood vessels. Dain Son have reported that pdgf-c-mediated signal pathway is involved in macrophage's anti-apoptotic effects, suggesting that tumor cells may promote enhanced malignancy via increasing tumor-associated macrophage survival [53]. Meanwhile, the anti-apoptotic role for macrophage signaling pathways mediated by PDGF-C was analyzed, and the researchers found that malignant human breast cancer cell line MDA-MB-231 produces high amount of PDGF-C; on the contrary, PDGF-C was not detected in benign MCF-7 cells. The conclusion is that the tumor cell-derived PDGF-C enhances the survival of tumor-associated macrophages, and promotes malignancy [54]. PDGF can be seen during the development of macrophages, which has a role in tumor growth.

Mesenchymal stem cells in bone marrow are a class of non-hematopoietic stem cells, which support and regulate hematopoiesis in vivo. Therefore, they have an effect on the occurrence and development of tumors. Recent studies show that PDGF-AA requires BMPRIA and PDGFRa receptor to activate the mesenchymal stem cells (MSC) bmp-smad1/5/8 channel [55]. Mesenchymal stem cells can be co-primary tumor lesion with other types of cells. A study showed that the presence of bone marrow mesenchymal stem cells/pericytes coverage of the target organ vasculature is necessary for effective melanoma metastasis to the bone marrow and liver [56]. The more interesting finding was that among patients with advanced breast cancer, bone marrow mesenchymal stem cells can change the migration of MCF-7 and MDA-MB231 cells. The levels of PDGF-AB, ICAM-1, and VCAM-1 in patient's bone marrow were significantly higher than those in healthy volunteers, suggesting that they may play a role in cancer cell extravasation, bone resorption, and cancer cell proliferation. This study shows that bone marrow mesenchymal stem cells may have similar function as PDGF-AB [57].

The relationship between platelet-derived growth factor and immune cells in the tumor is unclear. Agrawal *et al.* demonstrated that human PDGF inhibited dendritic cell (DCs) maturation and induced IL-10 secretion. They also found that PDGF induced the expression of C-type lectin-like receptor member 2 (CLEC-2) receptor on DCs, leading to the induction of regulatory T cells [58].

Neutrophils have dual role in tumor: they can inhibit the cancer development; on the other hand, they promote tumor growth and invasion [59]. Thus, it is necessary to investigate the relationship between neutrophils and PDGF. Platelets release Platelet-derived growth factor (PDGF) and attract inflammatory cells such as monocytes and neutrophils. MG Houghton et *al.* determined the effect of neutrophil elastase on tumor progression. Their results show that neutrophil elastase degradation of insulin receptor substrate 1 (IRS-1) can increase the interaction between phosphatidylinositol 3-kinase (PI3K) and platelet-derived growth factor interaction receptor (PDGFR), thus distorting the PI3K-axis toward tumor cell proliferation [60].

MDSC can suppress innate immune cell-NK cell and NKT cell-mediated tumor cytotoxicity, as well as CD4<sup>+</sup> CD8<sup>+</sup> T cell-mediated adaptive immunity, and will gradually become resistant to anti-tumor immune responses [61]. In the study of breast cancer in metastatic sites, Kaplan et al. found that VEGFR and bone marrow-derived haematopoietic progenitor cells are among the first to arrive at the pre-metastatic sites before the arrival of breast cancer cells to promote the formation of microenvironment called pre-metastatic niche (pre-metastatic niche) for tumor cell growth [62]. MDSC derived VEGF cytokine directly contribute to the formation of the pre-shift niche [63]. Research reports that MDSC, VEGF, and PDGF have similar functions, and they may all promote the formation of pre-metastatic niche.

Tumor associated CAF is an important participant in tumor proliferation. Desmoplastic malignant tumor such as cholangiocarcinoma is rich in tumor-associated fibroblasts [64]. CAF, as primary or metastatic tumor stroma, is an important component in tumor formation and progression [65]. Chu TY's team studied the impact of the crosstalk between cancer cells and cancer associated fibroblasts (CAFs) on the proliferation and survival of irradiated cancer cells, and found that CAF-cancer cell crosstalk has protective effect on radiated cancer cells. They identified various growth factors, including PDGF at CAF-cancer cell crosstalk which may affect the radiation protective effects of cancer cells [66]. The latest results show that PDGF-activated CAF upregulates puma, which causes the proapoptotic change of Bak, resulting in enhanced cell apoptosis in vivo [26]. PDGF is an important protumor factor that causes fibroblast transformation. Activation of Src and ERK can activate collagen integrin signaling pathway and improve the production of PDGF-A, a key regulator for fibroblast recruitment [67].

# Clinical application of PDGF detection in cancer

# Diagnosis

As a tumor growth factor, PDGF stimulates tumor growth, invasion, and metastasis and is also involved in chemotherapy resistance. A study has shown that cancer can cause increased serum PDGF content [68]. Cancer induced increase in the content of serum PDGF may be associated with a variety of factors: tumor cells release bone marrow stimulation substance which functions as thrombopoietin to stimulate the pluripotent stem cells, thus contributing to the increased platelet production following the increase of serum PDGF [68]. Their data also suggest that PDGF content in the serum of liver cancer patients was significantly higher than that in normal people, and benign liver disease patients' serum PDGF levels are slightly higher than that in the normal serum, but the difference did not reach statistical significance [68]. Therefore, the content of serum PDGF is not only an ideal marker for HCC diagnosis in liver cancer, but also a reference that is valuable in differential diagnosis among liver benign lesions such as liver cirrhosis, chronic hepatitis, and liver hemangioma [68]. Many studies have shown that cancer cells highly express PDGF. For example, Lei Xu et al. revealed that PDGF-D was homogenously strongly expressed in the tumor tissues of all histologic types, Jieqiong Liu et al. revealed that human breast cancers express high levels of PDGF-D, and Miguel Torres-Martin et al. identified that PDGFD was upregulated in meningiomas and schwannomas when compared with their respective healthy tissues [23, 30, 69]. Moreover, a study confirmed that the serum VEGF and PDGF levels in malignant ovarian tumors (MOT) increased compared with those in benign ovarian tumor (BOT) and the normal control group; serum VEGF and PDGF levels in BOT group were similar to those in the normal control group; and there was no significant difference between VEGF and PDGF. Thus, the serum VEGF and PDGF levels are associated with ovarian tumor malignant behavior and have certain value in the diagnosis of ovarian

tumors [70]. Overall, these results suggest that increased serum VEGF and PDGF levels may be associated with the formation of cancer. However, Zheng Li *et al.* found that the serum PDGF content slightly decreased in several cancers [68]. In addition, D Matei *et al.* claimed that measurable levels of PDGF were of no predictive value for the diagnosis of ovarian malignancy [71]. It can be seen that the application of PDGF in the diagnosis of cancer is still controversial and need to be further explored.

### Monitoring

PDGF plays an important role in tumor development. Monitoring the expression and sites of PDGF may monitor the tumor development. Studies have shown that there are differences in the expression of PDGF in cancers in different stages. For example, the levels of VEGF and PDGF in low differentiated tumors (G3) and late stage tumors (FIGO stage III and IV) were higher than those in highly differentiated tumors (G1, G2) and early stage tumors (FIGO I + II) in malignant ovarian tumors [70]. Moreover, VEGFA contributes to lung cancer progression by significantly induced the secretion of a variety of angiogenic factors, such as PDGFB, which might offer potential for monitor and therapeutic intervention [72]. NRASQ61 mutations are associated with hypomethylation of PDGFD, which consequently increases the gene expression of PDGFD and subsequently dysregulates downstream regulatory cascades. Hence, we can monitor the process of NRASQ61 mutations by monitoring the PDGF gene and downstream regulation genes [73]. In addition, a study has proved that in the liver metastasis of gastric cancer, serum PDGF levels were markedly increased to a level that was significantly higher than that of patients with primary gastric cancer and normal people, indicating that serum PDGF has a value in determining and monitoring the course of gastric cancer. In gastric cancer patients with serum PDGF levels increased significantly, the possibility of liver metastasis should be considered [70]. Yuan Wang et al. demonstrated that PDGF-D was commonly over-expressed in endometrial cancer, which was associated with late stage deep myometrium invasion and lymphoma vascular space invasion [74]. Both in vitro and in vivo experiments showed that PDGF-D could promote tumor growth and invasion through

up-regulating MMP2/9 and inducing EMT. Compared with that in matched normal endometrial cases, PDGF-D was up-regulated in endometrial cancer patients. Expression of PDGF-D protein, found in 78% of the cases, was associated with nonendometrioid histologic type (P=0.028), FIGO stage III/IV (P=0.039), >50% solid tumor growth (P=0.048), pelvic LN metastasis (P=0.035), and ER and PR negativity (P=0.04 and 0.002, respectively) [75]. PDGF-D expression was also significantly associated with the expression of VEGF-A (P=0.021) [75]. Collectively, these results suggest that increased serum VEGF and PDGF levels may be associated with the formation and malignant behavior of tumor and the serum VEGF and PDGF levels before the operation of ovarian tumor may have certain diagnostic value.

# Prognosis prediction

Prognosis (tumor size, lymph node status, and histological grading, etc.) and treatment-predictive factors are routinely used for the classification and guidance of subsequent treatment decisions in tumor. Staging of the cancer, determined by tumor-node-metastasis (TNM) staging system, can be divided into prognostic subgroups, and several markers in tumor cells are useful to predict the prognosis of the patients. Previous studies have pointed out that the PDGF receptor can be used as a prognostic and predictive marker of tumor. For example, Donnem et al. studied the prognostic value of PDGF in NSCLC tumor cells and stromal cells (including endothelial cells, immune cells, fibroblasts, etc.) and found that high tumor cell PDGF-B and PDGFR-β expression were independent negative prognostic factors for disease-specific survival, whereas in stromal cells high PDGF-A expression had an independent positive survival impact, which may due to the fact that high levels of PDGF-A in interstitial cells can activate adaptive anti-tumor immunity [76, 77]. High expression of PDGFRβ in stroma of pancreatic adenocarcinoma correlates with a worse prognosis [78]. Moreover, PDGF-B receptor expression in tumor stroma was correlated with HER-2 positivity, and the expression loss of PDGF-B receptor in tumor stroma was correlated with TNBC (triple negative breast cancer) [79]. Thus, stromal PDGF-β receptor expression significantly correlates with less favorable clinicopathological parameters and shorter survival in breast cancer [80].

Because tumor metastasis depends on the tumor microenvironment, which may be related to activation and differentiation of fibroblasts, pericytes recruitment, and extracellular matrix (ECM); Whereas PDGF induces fibroblast differentiation, pericyte recruitment, and ECM formation, thus PDGF can act as a reliable tumor prognostic marker [81]. It is demonstrated that the PDGF signature captures biological properties that are not captured by other stromaderived predictors, therefore is of prognostic significance [82]. Sigve Andersen et al. proved that PDGF expression in tumor emerged as an independent poor prognosticator for diseasespecific survival in non-small cell lung cancer, and Kawai T et al. also reported that immunohistochemistry for PDGF B-chain may predict the outcome for lung carcinoma patients [83, 84]. PDGF-D expression proved to be an independent prognostic factor in addition to histologic grade and FIGO stage, and patients with high expression levels of PDGF-D had a significantly poorer overall survival rate compared with patients with no expression [85]. Continued characterization of PDGFR expression in human tumors should present opportunities for improved accuracy in prognosis and also allow novel biomarker-based clinical studies exploring the efficacy of PDGFR-directed tumor therapies.

### Therapy

The fact that PDGF and/or PDGF receptors are overexpressed or mutated in different tumors makes it desirable to investigate whether PDGF or PDGF receptor antagonists can be used to treat patients with these diseases. Many studies have proved that PDGF ligands are expressed by cancer cells, whereas PDGF-Rs are expressed mainly by stromal cells. Directed therapy targets both PDGF and stromal PDGFR, which can be activated so as to be closely associated with recruitment and activation of fibroblasts and significant deposition of extracellular matrix (ECM) [80, 86]. A large number of studies have shown PDGF targeted therapy is effective in a variety of tumors, including thyroid nodules, recurrent goiter, pancreatic adenocarcinoma., leukemia, cholangiocarcinoma, lymphoma, gastric cancer, colon cancer, and prostate cancer etc. [78, 87-92]. Several potent PDGF receptor kinase inhibitors have been developed, including imatinib, sunitinib, sorafenib, pazopanib, and nilotinib, etc., which often get better results by combined use. For example, nilotinib and everolimus in combination reduced both the growth rate and stromal reaction in gastric cancer [91]. Also, monoclonal antibodies directed against PDGF or PDGFR have gradually been used to test whether they can delay and improve tumor development. For instance, IMC-3G3 specifically binds to human PDGF receptor  $\alpha$  (PDGFR $\alpha$ ) with high affinity and blocks PDGF ligand binding and PDGFRa activation; IMC-2C5 is directed against PDGFRβ from an antibody phage display library [93, 94]. However, a study shows that the resistance mechanisms limit the success of such treatments, and anti-PDGF receptor therapy is most likely to achieve lasting remission when combined with other signal transduction inhibitors, chemotherapy, or other treatments [11]. Marya F. et al. revealed that treatment of PDGF-BBoverexpressing tumors with imatinib mesylate (PDGFR inhibitor) resulted in increased growth and decreased total pericyte content compared with those in untreated PDGF-BBoverexpressing tumors [95]. Thus, Single-agent therapy targeting PDGF receptor must be used with caution when PDGFR is not the target on tumor cell itself. Further developments rely on clinical studies where systematic analyses of target status in malignant cells and in cells of the tumor stroma are included. It remains to be studied that how to further reduce the side effects of combination therapy with selective PDGFR inhibitors.

PDGF signaling has a key role in cancer progression, stimulating tumor cell proliferation, invasion, migration, and chemotherapy resistance. Through autocrine or paracrine secretion, PDGF binds to its receptor to activate series of protein signals to stimulate the growth of cancer cells, inhibit apoptosis, and directly or indirectly support cancer metastasis by acting on tumor cells or in tumor microenvironment. By promoting epithelial mesenchymal transition and VEGF dependent or independent angiogenesis, and recruitment of peripheral blood cells, PDGF signaling is involved in the chemotherapy resistance of cancers. A large number of studies have shown that due to the close relationship between PDGF and tumor progression, PDGF has certain clinical value in the diagnosis and monitoring of cancers. In addition, usually PDGF highly expression is typically associated with a poor prognosis. Finally, many researchers have studied the therapeutic

effects of PDGF signaling-targeting agents. However, effects of PDGF on cancer are complex and diverse, and further analysis is needed to confirm the predictive and therapeutic values of PDGF and its receptor. In addition, how to target PDGF signaling pathway to more effectively control the occurrence and development of cancer remains to be explored.

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#### Disclosure of conflict of interest

None.

Address correspondence to: Dr. Mei Wang, School of Medicine, Jiangsu University, 301 Xuefu Road, Zhenjiang 212013, Jiangsu Province, China. Tel: 86-13656135013; E-mail: wangmei8417@163.com

#### References

- Canalis E. Effect of platelet-derived growth factor on DNA and protein synthesis in cultured rat calvaria. Metabolism 1981; 30: 970-975.
- [2] Czyrski JA, Narczewska B and Inglot AD. New procedure for purification of human plateletderived growth factor. Arch Immunol Ther Exp (Warsz) 1984; 32: 589-598.
- [3] Niman HL, Houghten RA and Bowen-Pope DF. Detection of high molecular weight forms of platelet-derived growth factor by sequencespecific antisera. Sci 1984; 226: 701-703.
- [4] Buhl EM, Djudjaj S, Babickova J, Klinkhammer BM, Folestad E, Borkham-Kamphorst E, Weiskirchen R, Hudkins K, Alpers CE, Eriksson U, Floege J and Boor P. The role of PDGF-D in healthy and fibrotic kidneys. Kidney Int 2016; 89: 848-861.
- [5] Candilera V, Bouchè C, Schleef J and Pederiva F. Lung growth factors in the amniotic fluid of normal pregnancies and with congenital diaphragmatic hernia. J Matern Fetal Neonatal Med 2016; 29: 2104-2108.
- [6] Vera C, Tapia V, Vega M and Romero C. Role of nerve growth factor and its TRKA receptor in

normal ovarian and epithelial ovarian cancer angiogenesis. J Ovarian Res 2014; 7: 82.

- [7] Wang Z, Ahmad A, Li Y, Kong D, Azmi AS, Banerjee S and Sarkar FH. Emerging roles of PDGF-D signaling pathway in tumor development and progression. Biochim Biophys Acta 2010; 1806: 122-130.
- [8] Wang Z, Kong D, Li Y and Sarkar FH. PDGF-D signaling: a novel target in cancer therapy. Curr Drug Targets 2009; 10: 38-41.
- [9] Kong D, Wang Z, Sarkar SH, Li Y, Banerjee S, Saliganan A, Kim HR, Cher ML and Sarkar FH. Platelet-derived growth factor-D overexpression contributes to epithelial-mesenchymal transition of PC3 prostate cancer cells. Stem Cells 2008; 26: 1425-1435.
- [10] Carvalho I, Milanezi F, Martins A, Reis RM and Schmitt F. Overexpression of platelet-derived growth factor receptor alpha in breast cancer is associated with tumour progression. Breast Cancer Res 2005; 7: R788-795.
- [11] Westermark B and Heldin CH. Platelet-derived growth factor. Structure, function and implications in normal and malignant cell growth. Acta Oncol 1993; 32: 101-105.
- [12] Ustach CV, Taube ME, Hurst NJ Jr, Bhagat S, Bonfil RD, Cher ML, Schuger L and Kim HR. A potential oncogenic activity of platelet-derived growth factor d in prostate cancer progression. Cancer Res 2004; 64: 1722-1729.
- [13] Tian Y, Chu Q and Chen Y. Progress of platelet derived grow factor family in non-small cell lung cancer. Chin J Lung Cancer 2014; 17: 42-48.
- [14] Finlay GA, Thannickal VJ, Fanburg BL and Kwiatkowski DJ. Platelet-derived growth factor-induced p42/44 mitogen-activated protein kinase activation and cellular growth is mediated by reactive oxygen species in the absence of TSC2/tuberin. Cancer Res 2005; 65: 10881-10890.
- [15] Shamah SM, Alberta JA, Giannobile WV, Guha A, Kwon YK, Carroll RS, Black PM and Stiles CD. Detection of activated platelet-derived growth factor receptors in human memngioma. Cancer Res 1997; 57: 4141-4147.
- [17] Lu Y, Lin N, Chen Z and Xu R. Hypoxia-induced secretion of platelet-derived growth factor-BB by hepatocellular carcinoma cells increases activated hepatic stellate cell proliferation, migration and expression of vascular endothelial growth factor-A. Mol Med Rep 2015; 11: 691-697.

- [18] Geng ZM, Jha RK, Li B, Chen C, Li WZ, Zheng JB, Wang L and Huanchen S. Sorafenib inhibition of hepatic stellate cell proliferation in tumor microenvironment of hepatocellular carcinoma: a study of the sorafenib mechanisms. Cell Biochem Biophys 2014; 69: 717-724.
- [19] Furuhashi M, Sjoblom T, Abramsson A, Ellingsen J, Micke P, Li H, Bergsten-Folestad E, Eriksson U, Heuchel R, Betsholtz C, Heldin CH and Ostman A. Platelet-derived growth factor production by B16 melanoma cells leads to increased pericyte abundance in tumors and an associated increase in tumor growth rate. Cancer Res 2004; 64: 2725-2733.
- [20] Willenberg A, Saalbach A, Simon JC and Anderegg U. Melanoma cells control HA synthesisin peritumoral fibroblasts via PDGF-AA and PDGF-CC: impact on melanoma cell proliferation. J Invest Dermatol 2012; 132: 385-393.
- [21] Pinto MP, Dye WW, Jacobsen BM and Horwitz KB. Malignant stroma increases luminal breast cancer cell proliferation and angiogenesis through platelet-derived growth factor signaling. BMC Cancer 2014; 14: 735.
- [22] Guan X. Cancer metastases: challenges and opportunities. Acta Pharm Sin B 2015; 5: 402-418.
- [23] Xu L, Tong R, Cochran DM and Jain RK. Blocking platelet-derived growth factor-D/plateletderived growth factor receptor  $\beta$  signaling inhibits human renal cell carcinoma progression in an orthotopic mouse model. Cancer Res 2005; 65: 5711-5719.
- [24] Wagsaeter D, Zhu C, Björck H and Eriksson P. Effects of PDGF-C and PDGF-D on monocyte migration and MMP-2 and MMP-9 expression. Atherosclerosis 2009; 202: 415-423.
- [25] Ahmad A, Wang Z, Kong D, Ali R, Ali S, Banerjee S and Sarkar FH. Platelet-derived growth factor-D contributes to aggressiveness of breast cancer cells by up-regulating Notch and NF-κB signaling pathways. Breast Cancer Res Treat 2011; 126: 15-25.
- [26] Rizvi S, Mertens JC, Bronk SF, Hirsova P, Dai H, Roberts LR, Kaufmann SH and Gores GJ. Platelet-derived growth factor primes cancer-associated fibroblasts for apoptosis. J Biol Chem 2014; 289: 22835-22849.
- [27] Huang F, Wang M, Yang T, Cai J, Zhang Q, Sun Z, Wu X, Zhang X, Zhu W, Qian H and Xu W. Gastric cancer-derived MSC-secreted PDGF-DD promotes gastric cancer progression. J Cancer Res Clin Oncol 2014; 140: 1835-1848.
- [28] Liu J, Liao S, Huang Y, Samuel R, Shi T, Naxerova K, Huang P, Kamoun W, Jain RK, Fukumura D and Xu L. PDGF-D improves drug delivery and efficacy via vascular normalization, but promotes lymphatic metastasis by activating

CXCR4 in breast cancer. Clin Cancer Res 2011; 17: 3638-3648.

- [29] Yang Y, Andersson P, Hosaka K, Zhang Y, Cao R, Iwamoto H, Yang X, Nakamura M, Wang J, Zhuang R, Morikawa H, Xue Y, Braun H, Beyaert R, Samani N, Nakae S, Hams E, Dissing S, Fallon PG, Langer R and Cao Y. The PDGF-BB-SOX7 axis-modulated IL-33 in pericytes and stromal cells promotes metastasis through tumour-associated macrophages. Nat Commun 2016; 7: 11385.
- [30] Kitadai Y, Sasaki T, Kuwai T, Nakamura T, Bucana CD, Hamilton SR and Fidler IJ. Expression of activated platelet-derived growth factor receptor in stromal cells of human colon carcinomas is associated with metastatic potential. Int J Cancer 2006; 119: 2567-2574.
- [31] Yu J, Ustach C and Kim HR. Platelet-derived growth factor signaling and human cancer. J Biochem Mol Biol 2003; 36: 49-59.
- [32] Jechlinger M, Sommer A, Moriggl R, Seither P, Kraut N, Capodiecci P, Donovan M, Cordon-Cardo C, Beug H and Grunert S. Autocrine PDGFR signaling promotes mammary cancer metastasis. J Clin Invest 2006; 116: 1561-1570.
- [33] Jahn SC, Law ME, Corsino PE, Parker NN, Pham K, Davis BJ, Lu J and Law BK. An in vivo model of epithelial to mesenchymal transition reveals a mitogenic switch. Cancer Lett 2012; 326: 183-190.
- [34] Okui G, Tobiume K, Rizqiawan A, Yamamoto K, Shigeishi H, Ono S, Higashikawa K and Kamata N. AKT primes snail-induced EMT concomitantly with thecollective migration of squamous cell carcinoma cells. J Cell Biochem 2013; 114: 2039-2049.
- [35] Wu Q, Wang R, Yang Q, Hou X, Chen S, Hou Y, Chen C, Yang Y, Miele L, Sarkar FH, Chen Y and Wang Z. Chemoresistance to gemcitabine in hepatoma cells induces epithelial-mesenchymal transition and involves activation of PDGF-D pathway. Oncotarget 2013; 4: 1999-2009.
- [36] Wang R, Li Y, Hou Y, Yang Q, Chen S, Wang X, Wang Z, Yang Y, Chen C, Wang Z and Wu Q. The PDGF-D/miR-106a/Twist1 pathway orchestrates epithelial-mesenchymal transition in gemcitabine resistance hepatoma cells. Oncotarget 2015; 6: 7000-7010.
- [37] di Tomaso E, Snuderl M, Kamoun WS, Duda DG, Auluck PK, Fazlollahi L, Andronesi OC, Frosch MP, Wen PY, Plotkin SR, Hedley-Whyte ET, Sorensen AG, Batchelor TT and Jain RK. Glioblastoma recurrence after cediranib therapy in patients: lack of "rebound" revascularization as mode of escape. Cancer Res 2011; 71: 19-28.
- [38] Berg JT, Breen EC, Fu Z, Mathieu-Costello O and West JB. Alveolar hypoxia increases gene

expression of extracellular matrix proteins and platelet-derived growth factor-B in lung parenchyma. Am J Respir Crit Care Med 1988; 158: 1920-1928.

- [39] Zhao Z, Liu Y, He H, Chen X, Chen J and Lu YC. Candidate genes influencing sensitivity and resistance of human glioblastoma to Semustine. Brain Res Bull 2011; 86: 189-194.
- [40] Gallego-Muñoz P, Ibares-Frías L, Garrote JA, Valsero-Blanco MC, Cantalapiedra-Rodríguez R, Merayo-Lloves J and Martínez-García MC. Human corneal fibroblast migration and ECM synthesis during stromal repair: role played by PDGF-BB, bFGF, and TGFβ1. J Tissue Eng Regen Med 2016; [Epub ahead of print].
- [41] Li X, Yu X, Dai D, Song X and Xu W. The altered glucose metabolism in tumor and a tumor acidic microenvironment associated with extracellular matrix metalloproteinase inducer and monocarboxylate transporters. Oncotarget 2016; 7: 23141-23155.
- [42] Xu J, E C, Yao Y, Ren S, Wang G and Jin H. Matrix metalloproteinase expression and molecular interaction network analysis in gastric cancer. Oncol Lett 2016; 12: 2403-2408.
- [43] Pekny M, Ostman A, Hermansson A, Nistér M, Heldin CH and Westermark B. Differences in binding to the solid substratum and extracellular matrix may explain isoform-specific paracrine effects of platelet-derived growth factor. Growth Factors 1994; 10: 77-87.
- [44] Wright JH, Johnson MM, Shimizu-Albergine M, Bauer RL, Hayes BJ, Surapisitchat J, Hudkins KL, Riehle KJ, Johnson SC, Yeh MM, Bammler TK, Beyer RP, Gilbertson DG, Alpers CE, Fausto N and Campbell JS. Paracrine activation of hepatic stellate cells in platelet-derived growth factor C transgenic mice: evidence for stromal induction of hepatocellular carcinoma. Int J Cancer 2014; 134: 778-788.
- [45] Čunderlíková B. Clinical significance of immunohistochemically detected extracellular matrix proteins and their spatial distribution in primary cancer. Crit Rev Oncol Hematol 2016; 105: 127-144.
- [46] Cumpănas AA, Cimpean AM, Ferician O, Ceausu RA, Sarb S, Barbos V, Dema A and Raica M. The involvement of PDGF-B/PDGFRβ axis in the resistance to antiangiogenic and antivascular therapy in renal cancer. Anticancer Res 2016; 36: 2291-2295.
- [47] Lu Y, Lin N, Chen Z and Xu R. Hypoxia-induced secretion of platelet-derived growth factor-BB by hepatocellular carcinoma cells increases activated hepatic stellate cell proliferation, migration and expression of vascular endothelial growth factor-A. Mol Med Rep 2015; 11: 691-697.

- [48] Yang Y, Andersson P, Hosaka K, Zhang Y, Cao R, Iwamoto H, Yang X, Nakamura M, Wang J, Zhuang R, Morikawa H, Xue Y, Braun H, Beyaert R, Samani N, Nakae S, Hams E, Dissing S, Fallon PG, Langer R and Cao Y. The PDGF-BB-SOX7 axis-modulated IL-33 in pericytes and stromal cells promotes metastasis through tumour-associated macrophages. Nat Commun 2016; 7: 11385.
- [49] Song N, Huang Y, Shi H, Yuan S, Ding Y, Song X, Fu Y and Luo Y. Overexpression of platelet-derived growth factor-BB increases tumor pericyte content via stromal-derived factor-1alpha/ CXCR4 axis. Cancer Res 2009; 69: 6057-6064.
- [50] Chen Z, Xu XH and Hu J. Role of pericytes in angiogenesis: focus on cancer angiogenesis and anti-angiogenic therapy. Neoplasma 2016; 63: 173-182.
- [51] Gladh H, Folestad EB, Muhl L, Ehnman M, Tannenberg P, Lawrence AL, Betsholtz C and Eriksson U. Mice lacking platelet-derived growth factor D display a mild vascular phenotype. PLoS One 2016; 11: e0152276.
- [52] Sennino B, Falcón BL, Mccauley D, Le T, Mc-Cauley T, Kurz J, Haskell A, Epstein D and Mc-Donald DM. Sequential loss of tumor vessel pericytes and endot helial cells after inhibition of platelet-derived growth factor B by selective aptamer AX102. Cancer Res 2007; 67: 7358-7367.
- [53] Kitadai Y, Sasaki T, Kuwai T, Nakamura T, Bucana CD and Fidler IJ. Targeting the expression of Platelet-derived growth factor receptor by reactive stroma inhibits growth and metastasis of human colon carcinoma. Am Pathol 2006; 169: 2054-2065.
- [54] Son D, Na YR, Hwang ES and Seok SH. Platelet-derived growth factor-C (PDGF-C) induces anti-apoptotic effect son macrophages through akt and bad phosphorylation. J Biol Chem 2014; 289: 6225.
- [55] Li A, Xia X, Yeh J, Kua H, Liu H, Mishina Y, Hao A and Li B. PDGF-AA promotes osteogenic differentiation and migration of mesenchymal stem cell by down-regulating PDGFR $\alpha$  and derepressing BMP-Smad1/5/8 signaling. PLoS One 2014; 9: e113785.
- [56] Correa D, Somoza RA, Lin P, Schiemann WP and Caplan AI. Mesenchymal stem cells regulate melanoma cancer cells extravasation to bone and liver at their perivascular niche. Int J Cancer 2016; 138: 417-427.
- [57] Martinez LM, Vallone VB, Labovsky V, Choi H, Hofer EL, Feldman L, Bordenave RH, Batagelj E, Dimase F, Villafañe AR and Chasseing NA. Changes in the peripheral blood and bone marrow from untreated advanced breast can-

cer patients that are associated with the establishment of bone metastases. Clin Exp Metastasis 2014; 31: 213-32.

- [58] Agrawal S, Ganguly S, Hajian P, Cao J and Agrawal A. PDGF upregulates CLEC-2 to induce T regulatory cells. Oncotarget 2015; 6: 28621-28632.
- [59] Tazawa H, Okada F, Kobayashi T, Tada M, Mori Y, Une Y, Sendo F, Kobayashi M and Hosokawa M. Infiltration of neutrophils is required for acquisition of metastatic phenotype of benign murine fibrosarcoma cells: implication of inflammation-associated carcinogenesis and tumor progression. Am J Pathol 2003; 163: 2221-2232.
- [60] Houghton AM, Rzymkiewicz DM, Ji H, Gregory AD, Egea EE, Metz HE, Stolz DB, Land SR, Marconcini LA, Kliment CR, Jenkins KM, Beaulieu KA, Mouded M, Frank SJ, Wong KK and Shapiro SD. Neutrophil elastase-mediated degradation of IRS-1 accelerates lung tumor growth. Nat Med 2010; 16: 219-223.
- [61] Raber PL, Sierra RA, Thevenot PT, Shuzhong Z, Wyczechowska DD, Kumai T, Celis E and Rodriguez PC. T cells conditioned with MDSC show an increased anti-tumor activity after adoptive T cell based immunotherapy. Oncotarget 2016; 7: 17565-17578.
- [62] Kaplan RN, Riba RD, Zacharoulis S, Bramley AH, Vincent L, Costa C, MacDonald DD, Jin DK, Shido K, Kerns SA, Zhu Z, Hicklin D, Wu Y, Port JL, Altorki N, Port ER, Ruggero D, Shmelkov SV, Jensen KK, Rafii S and Lyden D. VEGFR1-positive haematopoietic bone marrow progenitors initiate the pre-metastatic niche. Nature 2005; 438: 820-827.
- [63] Tartour E, Pere H, Maillere B, Terme M, Merillon N and Taieb J. Angiogenesis and immunity: a bidirectional link potentially relevant for the monitoring of antiangiogenic therapy and the development of novel therapeutic combination with immunotherapy. Cancer Metastasis Rev 2011; 30: 83-95.
- [64] Heits N, Heinze T, Bernsmeier A, Kerber J, Hauser C, Becker T, Kalthoff H, Egberts JH and Braun F. Influence of mTOR-inhibitors and mycophenolic acid on human cholangiocellular carcinoma and cancer associated fibroblasts. BMC Cancer 2016; 16: 322.
- [65] Shan T, Chen S, Chen X, Lin WR, Li W, Ma J, Wu T, Ji H, Li Y, Cui X and Kang Y. Prometastatic mechanisms of CAF-mediated EMT regulation in pancreatic cancer cells. Int J Oncol 2017; 50: 121-128.
- [66] Chu TY, Yang JT, Huang TH and Liu HW. Crosstalk with cancer-associated fibroblasts increases the growth and radiation survival of cervical cancer cells. Radiat Res 2014; 181: 540-547.

- [67] Chen SY, Lin JS, Lina HC, Shan YS, Cheng YJ and Yang BC. Dependence of fibroblast infiltration in tumor stroma on type IV collagen-initiated integrin signal through induction of plateletderived growth factor. Biochim Biophys Acta 2015; 1853: 929-939.
- [68] Li Z and He J. Application of Patelet-derived growth factor in diagnosis of cancer. Chin J Modern Med 2011; 21: 2072.
- [69] Torres-Martin M, Lassaletta L, Isla A, De Campos JM, Pinto GR, Burbano R, Castresana JS, Melendez B and Rey JA. Global expression profile in low grade meningiomas and schwannomas shows upregulation of PDGFD, CDH1 and SLIT2 compared to their healthy tissue. Oncol Rep 2014; 32: 2327-2334.
- [70] Xie L, Pu HY, Lei HJ and Luo J. The Diagnostic values of assay of serum vascular endothelial growth factor and platelet-derived growth factor in ovarian tumors. West Chin Med J 2005; 20: 244.
- [71] Matei D, Emerson RE, Lai YC, Baldridge LA, Rao J, Yiannoutsos C and Donner DB. Autocrine activation of PDGFRα promotes the progression of ovarian cancer. Oncogene 2006; 25: 2060-2069.
- [72] Frezzetti D, Gallo M, Roma C, D'Alessio A, Maiello MR, Bevilacqua S, Normanno N and De Luca A. Vascular endothelial growth factor a regulates the secretion of different angiogenic factors in lung cancer cells. J Cell Physiol 2015; 231: 1514-1521.
- [73] Jiang W, Jia P, Hutchinson KE, Johnson DB, Sosman JA and Zhao Z. Clinically relevant genes and regulatory pathways associated with NRASQ61 mutations in melanoma through an integrative genomics approach. Oncotarget 2014; 6: 2496-2508.
- [74] Wang Y, Qiu H, Hu W, Li S and Yu J. Over-expression of platelet-derived growth Factor-D promotes tumor growth and invasion in endometrial cancer. Int J Mol Sci 2014; 15: 4370.
- [75] Ding J, Li XM, Liu SL, Zhang Y and Li T. Overexpression of platelet-derived growth Factor-D as a poor prognosticator in endometrial cancer. APJCP 2014; 15: 3741-3745.
- [76] Paulsson J, Ehnman M and Östman A. PDGF receptors in tumor biology: prognostic and predictive potential. Future Oncol 2014; 10: 1695-1708.
- [77] Donnem T, Al-Saad S, Al-Shibli K, Andersen S, Busund LT and Bremnes RM. Prognostic impact of platelet-derived growth factors in nonsmall cell lung cancer tumor and stromal cells. J Thorac Oncol 2008; 3: 963-970.
- [78] Yuzawa S, Kano MR, Einama T and Nishihara H. PDGFRβ expression in tumor stroma of pancreatic adenocarcinoma as a reliable prognostic marker. Med Oncol 2012; 29: 2824-2830.

- [79] Koo JS, Park S, Kim S, Lee S and Park BW. The impact of caveolin protein expression in tumor stroma on prognosis of breast cancer. Tumor Biol 2011; 32: 787-799.
- [80] Paulsson J, Sjoblom T, Micke P, Ponten F, Landberg G, Heldin CH, Bergh J, Brennan DJ, Jirstrom K and Ostman A. Prognostic Significance of stromal platelet-derived growth factorβreceptor expression in human breast cancer. Am J Pathol 2009; 175: 334-341.
- [81] Hamdan R, Zhou Z and Kleinerman ES. SDF-1α induces PDGF-B expression and the differentiation of bone marrow cells into pericytes. Mol Cancer Res 2011; 9: 1462-1470.
- [82] Frings O, Augsten M, Tobin NP, Carlson J, Paulsson J, Pena C, Olsson E, Veerla S, Bergh J, Ostman A and Sonnhammer EL. Prognostic significance in breast cancer of a gene signature capturing stromal PDGF signaling. Am J Pathol 2013; 182: 2037-2047.
- [83] Andersen S, Donnem T, Al-Saad S, Al-Shibli K, Busund LT and Bremnes RM. Angiogenic markers show high prognostic impacton survival in marginally operable non-small cell lung cancer patients treated with adjuvant radiotherapy. J Thorac Oncol 2009; 4: 463-471.
- [84] Kawai T, Hiroi S and Torikata C. Expression in lung carcinomas of platelet-derived growth factor and its receptors. Lab Invest 1997; 77: 431-436.
- [85] Ding J, Li XM, Liu SL, Zhang Y and Li T. Overexpression of platelet-derived growth Factor-D as a poor prognosticator in endometrial cancer. Asian Pac J Cancer Prev 2014; 15: 3741-3745.
- [86] Forsberg K, Valyi-Nagy I, Heldin CH and Westermark B. Platelet-derived growth factor (PDGF) in oncogenesis: development of a vascular connective tissue stroma in xenotransplanted human melanoma producing PDGF-BB. Proc Nati Acad Sci U S A 1993; 90: 393-397.
- [87] George D. Targeting PDGF receptors in cancerrationales and proof of concept clinical trials. Adv Exp Med Biol 2003; 532: 141-151.
- [88] Ding W, Knox TR, Tschumper RC, Wu W, Schwager SM, Boysen JC, Jelinek DF and Kay NE. Platelet-derived growth factor (PDGF)-PDGF receptor interaction activates bone marrow-derived mesenchymal stromal cells derived from chronic lymphocytic leukemia: implications for an angiogenic switch. Blood 2010; 116: 2984-2993.

- [89] Fingas CD, Mertens JC, Razumilava N, Bronk SF, Sirica AE and Gores GJ. Targeting PDGFR-β in cholangiocarcinoma. Liver Int 2012; 32: 400-409.
- [90] Kitadai Y, Kodama M and Shinagawa K. Stroma-directed molecular targeted therapy in gastric cancer. Cancers 2011; 3: 4245-4257.
- [91] Onoyama M, Kitadai Y, Tanaka Y, Yuge R, Shinagawa K, Tanaka S, Yasui W and Chayama K. Combining molecular targeted drugs to inhibit both cancer cells and activated stromal cells in gastric cancer. Neoplasia 2013; 15: 1391-1399.
- [92] Shinagawa K, Kitadai Y, Tanaka M, Sumida T, Onoyama M, Ohnishi M, Ohara E, Higashi Y, Tanaka S, Yasui W and Chayama K. Stroma-directed imatinib therapy impairs the tumor-promoting effect of bone marrow-derived mesenchymal stem cells in an ort hotopic transplantation model of colon cancer. Int J Cancer 2013; 132: 813-823.
- [93] Shah GD, Loizos N, Youssoufian H, Schwartz JD and Rowinsky EK. Rationale for the development of IMC-3G3, a fully human immunoglobulin G subclass 1 monoclonal antibody targeting the platelet-derived growth factor receptor alpha. Cancer 2010; 116: 1018-1026.
- [94] Shen J, Vil MD, Prewett M, Damoci C, Zhang H, Li H, Jimenez X, Deevi DS, Lacolina M, Kayas A, Bassi R, Persaud K, Rohoza-Asandi A, Balderes P, Loizos N, Ludwig DL, Tonra J, Witte L and Zhu Z. Development of a fully human anti-PDGFRβ antibody that suppresses growth of human tumor xenografts and enhances antitumor activity of an anti-VEGFR2 Antibody. Neoplasia 2009; 11: 594-604.
- [95] McCarty MF, Somcio RJ, Stoeltzing O, Wey J, Fan F, Liu W, Bucana C and Ellis LM. Overexpression of PDGF-BB decreases colorectal and pancreatic cancer growth by increasing tumor pericyte content. J Clin Invest 2007; 117: 2114-2122.