## Original Article Association between ERCC1 and XPF polymorphisms and risk of extrahepatic cholangiocarcinoma

Kai Sun, Wentao Wang, Fuhai Wang, Peng Xiu, Haochen Wang, Feng Liu

Department of Hepatobiliary Surgery, Qianfoshan Hospital, Shandong University, No. 16766, Jingshi Road, Jinan 250014, Shandong Province, China

Received July 24, 2017; Accepted September 13, 2018; Epub December 15, 2018; Published December 30, 2018

Abstract: Background: Several risk factors, including primary sclerosing cholangitis, liver fluke infection, HBV/HCV infection, biliary malformations, and hepatolithiasis have been identified for developing cholangiocarcinoma (CCA). However, more than 85% of patients with extrahepatic cholangiocarcinoma (ECCA) have no explicit risk factors. Polymorphisms in excision repair cross-complementing group 1 (ERCC1) and xeroderma pigmentosum group F (XPF) could affect DNA repair capability. In this study, we studied the influence of ERCC1-XPF polymorphisms on ECCA incidence. Methods: The present study included 127 patients diagnosed of ECCA and 145 normal controls. The Genotypes of ERCC1-XPF were detected by polymerase chain reaction restriction fragment length polymorphism (PCR-RFLP) method, and then the products were sent for sequencing. Results: The ERCC1 rs3212986 C > A genotype AC+AA frequency was significantly different between the cases and controls (AC+AA, OR: 1.68, 95% CI: 1.04-2.72) comparing with genotype CC. The ERCC1 rs2298881 A > C genotype CC frequency was significantly different between the cases and controls (CC. OR: 2.15, 95% CI: 1.01-4.56) comparing with genotype AA. No associations with risk of ECCA were found for other three SNPs (ERCC1 rs11615, XPF rs6498486 and XPF rs2276466). Subgroup analysis showed that an extra increased risk in smokers was observed both in ERCC1 rs3212986 AC+AA genotype (OR: 2.75, 95% CI: 1.04-7.30) and rs229888 AC+CC genotype in smokers (OR: 3.22, 95% CI: 1.19-8.71). Conclusions: The present study indicated that rs3212986 C > A and rs2298881 A > C polymorphisms of ERCC1 were associated with an increased risk of ECCA, especially in smokers. It would be necessary to confirm these findings in a large sample size and multiethnic population study in future.

Keywords: Excision repair cross complementing group 1, xeroderma pigmentosum group F, polymorphism, cholangiocarcinoma, risk

#### Introduction

Extrahepatic cholangiocarcinoma (ECCA) is a rare but vicious tumor which originates from the epithelial cells of bile duct [1, 2]. Most patients usually present late and are often difficult to diagnose in most cases, so radical resection, the only curative option, is applicable in few patients. However, the recurrence rate after resection is extremely high. Even though chemotherapy regiment of gemcitabine and cisplatin is often used for advanced ECCA, the 5-year survival rate is very low [3, 4]. Several potential risk factors have been clarified, which include primary sclerosing cholangitis (PSC), parasitic infection, cholelithiasis, viral hepatitis, smoking, obesity and diabetes mellitus [5, 6], only a small percentage of patients have explicit risk factors. More than 85% of patients have no identifiable risk factors. Understanding of ECCA biology, oncogenic landscape and its complex interactions with tumor environment [7, 8] could lead to early diagnose and optimum therapies of this disease.

Human genomic DNA is continuously under attack by endogenous and exogenous mutagens. However, tumors only occur in a few people because DNA damage is spontaneously repaired by highly effective DNA repair pathways, which include base excision repair (BER), mismatch repair (MMR) and nucleotide excision repair (NER) [9]. Single-nucleotide polymorphisms (SNP) in DNA repair pathways might affect the quantity and quality of the encoding protein and the DNA repair capacity, conse-

	010			
Variables	Case (%) N = 127	Control (%) N = 145	X <sup>2</sup>	Р
Age				
≤ 65	77 (60.6)	84 (57.9)	0.20	0.65
> 65	50 (39.4)	61 (42.1)		
Gender				
Male	68 (53.5)	85 (58.6)	0.71	0.40
Female	59 (46.5)	60 (41.4)		
Smoking				
No	91 (71.7)	112 (77.2)	1.12	0.29
Yes	36 (28.3)	33 (22.8)		
Alcohol consumption				
No	102 (80.3)	126 (86.9)	2.16	0.14
Yes	25 (19.7)	19 (13.1)		
BMI (kg/m²)				
≤ 18.5	10 (7.9)	9 (6.2)	0.73	0.87
18.5-22.9	51 (40.2)	60 (41.4)		
23.0-24.9	46 (36.2)	49 (33.8)		
> 25	20 (15.7)	27 (18.6)		
Family history of cancer				
No	103 (81.1)	129 (89.0)	3.34	0.07
Yes	24 (18.9)	16 (11.0)		

Table 1. Characteristics of extrahepatic cholangiocarci-
noma cases and controls

quently increasing the susceptibility to carcinogens [10]. Excision repair cross-complementing group 1 (ERCC1) is located in chromosome 19q13.2-13.3, and takes part in the significant step of NER. Together with xeroderma pigmentosum group F (XPF), ERCC1 forms the ERCC1-XPF enzyme complex that participates in DNA repair and DNA recombination [11].

Previously, a lot of studies have investigated the association between ERCC1-XPF polymorphisms and the risk of breast cancer [12, 13], colorectal cancer [14-16], gastric cancer [17] and glioma [18], except cholangiocarcinoma. We here carried out a hospital-based case-control study to comprehensively investigate the association between ERCC1-XPF polymorphisms and the risk of developing ECCA in a Chinese population.

#### Materials and methods

#### Ethics

The present study was approved by the Ethics Committee of Qianfoshan Hospital of Shandong University (ethics approval number 2015013). All participants signed the informed consent.

#### Materials

A hospital-based case-control study was performed. 127 patients newly diagnosed of ECCA were recruited at Qianfoshan Hospital of Shandong University between March 2009 and January 2015. We included subjects that met the following criterion: (1) patients newly diagnosed with ECCA according to the clinical presentation and image examination, including computerized tomography (CT), magnetic resonance imaging (MRI), magnetic resonance cholangiopancreatography (MRCP) and endoscopic retrograde cholangiopancreatography (ERCP); (2) those with no previous history of other cancers or precancerous lesions; (3) those did not receive chemotherapy or radiotherapy previously; (4) those with their signed informed consent for the use of human blood and the current study protocol. 145 normal controls were randomly selected from healthy volunteers who visited the hospital for general health check-up. We excluded subjects that met the following criterion: (1) patients diagnosed with malignancy within

one year after blood draw in controls; (2) no blood specimens were available for analysis. After written informed consent was obtained, demographic data and environmental exposure history were obtained from the past digital records. All the subjects were the Han nationality without immediate family relations. This manuscript did not contain any individual person's data in any form.

#### Biochemical analysis

A total 5 ml venous blood samples were collected in an EDTA tube and stored at 4°C within 24 hours before DNA genome extracted. The genomic DNA was extracted by a routine phenol-chloroform method.

#### ERCC1 and XPF genotyping

The genotypes of ERCC1 and XPF polymorphisms were detected by polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) technique. We selected ERCC1 SNP rs2298881 A > C which might affect the binding site activity of transcription factor, and other two widely investigated functional SNPs (rs3212986 C > A and rs11615 G > A).

risk of extrahepat	ic cholangic	carcinoma		
Genotypes	Case (%) N = 127	Control (%) N = 145	OR (95% CI)	Ρ
ERCC1 rs3212986				
CC	59 (46.5)	86 (59.3)	1.00 (Ref.)	-
AC	52 (40.9)	49 (33.8)	1.55 (0.93-2.58)	0.09
AA	16 (12.6)	10 (6.9)	2.33 (0.99-5.49)	0.05
AC+AA	68 (53.5)	59 (40.7)	1.68 (1.04-2.72)	0.03
С	170 (66.9)	221 (76.2)	1.00 (Ref.)	-
А	84 (33.1)	69 (23.8)	1.58 (1.09-2.31)	0.02
ERCC1 rs2298881				
AA	57 (44.9)	78 (53.8)	1.00 (Ref.)	-
AC	48 (37.8)	53 (36.6)	1.24 (0.74-2.08)	0.42
CC	22 (17.3)	14 (9.6)	2.15 (1.01-4.56)	0.05
AC+CC	70 (55.1)	67 (46.2)	1.43 (0.89-2.31)	0.14
А	162 (63.8)	209 (72.1)	1.00 (Ref.)	-
С	92 (36.2)	81 (27.9)	1.47 (1.02-2.11)	0.04
ERCC1 rs11615				
GG	68 (53.5)	83 (57.2)	1.00 (Ref.)	-
AG	50 (39.4)	52 (35.9)	1.17 (0.71-1.94)	0.53
AA	9 (7.1)	10 (6.9)	1.10 (0.42-2.86)	0.85
AG+AA	59 (46.5)	62 (42.8)	1.16 (0.72-1.88)	0.54
G	186 (73.0)	218 (75.2)	1.00 (Ref.)	-
А	68 (27.0)	72 (24.8)	1.11 (0.75-1.63)	0.60
XPF rs6498486				
AA	70 (55.1)	84 (57.9)	1.00 (Ref.)	-
AC	44 (34.7)	52 (35.9)	1.02 (0.61-1.69)	0.95
CC	13 (10.2)	9 (6.2)	1.73 (0.70-4.29)	0.23
AC+CC	57 (44.9)	61 (42.1)	1.12 (0.69-1.81)	0.64
А	184 (72.4)	220 (75.9)	1.00 (Ref.)	-
С	70 (27.6)	70 (24.1)	1.20 (0.81-1.76)	0.36
XPF rs2276466				
CC	73 (57.5)	82 (56.6)	1.00 (Ref.)	-
CG	43 (33.9)	49 (33.8)	0.99 (0.59-1.65)	0.96
GG	11 (8.6)	14 (9.6)	0.88 (0.38-2.07)	0.77
CG+GG	54 (42.5)	63 (43.4)	0.96 (0.60-1.56)	0.88
С	189 (74.4)	213 (73.4)	1.00 (Ref.)	-
G	65 (25.6)	77 (26.6)	0.95 (0.65-1.40)	0.80

**Table 2.** Association of ERCC1 and XPF polymorphisms with

 risk of extrahepatic cholangiocarcinoma

Meanwhile, we selected XPF SNP rs2276466 C > G and rs6498486 A > C, the former one might affect the miRNA binding site activity, while the latter one might affect the binding site activity of transcription factor.

After the PCR finished, the amplified fragments were identified by electrophoresis on 2% agarose gels and the PCR products were sent for sequencing by the Shanghai Sangon Biotech Corp (Shanghai, China). All assays were repea-

### Population characteristics

Results

#### The distributions of demographic characteristics of the subjects were presented in **Table 1**. The case and control groups were not statistically different with respect to age ( $\chi^2 = 0.20$ , P= 0.65) and gender ( $\chi^2 = 0.71$ , P = 0.40). Other confirmed risk factors were also matched well between two groups (smoking, $\chi^2 = 1.12$ , P =

cago, IL, USA).

ted at least once by the same indi-

vidual. 10% of all samples were

randomly selected to verify the results by repeating the tests.

Mean and standard deviations were used to summarize the continuous variables. Differences  $(\alpha = 0.05)$  between two continuous variables were evaluated by the student's t test, while the  $x^2$  test was used to determine whether the frequencies between cases and controls were significantly different ( $\alpha = 0.05$ ). The  $\chi^2$  test was also used to compare distribution differences in haplotype and genotype. The Pearson's goodness-of-fit  $\chi^2$  test was used to assess Hardy-Weinberg equilibrium for genotype frequency in controls with one degree of freedom. Odds ratios (ORs) with 95% confidence intervals (CI) were used to detect the associations between these ERCC1-XPF polymorphisms and ECCA risk. Furthermore, we calculated crude ORs with 95% CIs by univariate logistic regression models to access the associations between the ERCC1-XPF genotypes and ECCA risk with and without adjustment for age, gender, smoking, alcohol consumption, body mass index (BMI) and family history of cancer. All statistical tests were two sided, considered statistically significant with P < 0.05. All analyses were conducted by SPSS version 16.0 software (SPSS, Chi-

Statistical analysis

tion within the Hardy-Weinberg equilibrium				
SNPs	χ <sup>2</sup>	Р		
ERCC1 rs3212986	0.673	0.412		
ERCC1 rs2298881	1.230	0.267		
ERCC1 rs11615	0.223	0.637		

0.063

2.588

0.802

0.108

 Table 3. ERCC1 and XPF genotype distribution within the Hardy-Weinberg equilibrium

SNPs: single-nucleotide polymorphisms.

XPF rs6498486

XPF rs2276466

0.29, alcohol consumption,  $\chi^2 = 2.16$ , P = 0.14, BMI,  $\chi^2 = 0.73$ , P = 0.87, and family history of cancer,  $\chi^2 = 3.34$ , P = 0.07).

Association of ERCC1 and XPF polymorphisms with the risk of ECCA

The result of ERCC1 polymorphisms with the risk of ECCA was shown in Table 2. For ERCC1 rs3212986, the genotype frequencies of CC, AC and AA were 46.5, 40.9 and 12.6%, respectively, in the ECCA cases compared with 59.3, 33.8 and 6.9%, respectively, in the controls. The genotype distribution in the controls was within the Hardy-Weinberg equilibrium ( $\chi^2$  = 0.67, P = 0.41) (**Table 3**). The genotype AC+AA frequency was significantly different between the cases and controls (AC+AA, OR: 1.68, 95% CI: 1.04-2.72) comparing with genotype CC, but not for individual genotype AC and AA frequencies (AC, OR: 1.55, 95% CI: 0.93-2.58, AA, OR: 2.33, 95% CI: 0.99-5.49). The allele frequencies of rs3212986 C > A between the two groups (OR: 1.58, 95% CI: 1.09-2.31) was also significantly different. For ERCC1 rs2298881, the genotype frequencies of AA, AC and CC were 44.9, 37.8 and 17.3%, respectively, in the ECCA cases compared with 53.8, 36.6 and 9.6%, respectively, in the controls. The genotype distribution in the controls was within the Hardy-Weinberg equilibrium ( $\chi^2$  = 1.23, P = 0.27) (Table 3). The genotype CC frequency was significantly different between the cases and controls (CC, OR: 2.15, 95% CI: 1.01-4.56), but not for genotype AC and AC+CC frequencies (AC, OR: 1.24, 95% CI: 0.74-2.08, AC+CC, OR: 1.43, 95% CI: 0.89-2.31). The allele frequencies of rs2298881 A > C between the two groups (OR: 1.47, 95% CI: 1.02-2.11) was also significantly different. No association with risk of ECCA was found for other three SNPs (ERCC1 rs11615, XPF rs6498486 and XPF rs2276466).

# Subgroup analysis for associations between ERCC1 variant genotypes with the risk of ECCA

**Table 4** showed the association between variant genotypes of two selected SNPs of ERCC1 and risk of ECCA by subgroup analysis considering age, gender, smoking, alcohol consumption, BMI and family history of cancer. The ERCC1 rs3212986 variant AC+AA genotype was associated with an extra increased risk in smokers (OR: 2.75, 95% Cl: 1.04-7.30). Quite similar result was observed for ERCC1 rs229888 variant AC+CC genotype in smokers (OR: 3.22, 95% Cl: 1.19-8.71). However the other common risk factors, such as alcohol consumption status, BMI and family history of cancer, did not show an extra increased risk.

#### Discussion

Radical surgical resection is the only curative treatment for ECCA. Although chemotherapy regiment of gemcitabine and cisplatin is often used for advanced ECCA, the 5-year survival rate is very low [19]. Patients with positive margins are no better than those who receive only palliative therapy [20]. The development of diagnostic tools (genetic change and tumor markers) may be an important way of identifying early patients who can benefit from RO resection [5]. The mechanism of cholangiocarcinogenesis is not yet clear. Environmental and genetic factors are thought to play an important role in the development of cancer. Previous studies showed that several environmental factors were identified as risk factors, including primary sclerosing cholangitis, liver fluke infection, HBV/HCV infection, biliary malformations, and cholelithiasis [21-23]. However, not all individuals who have been exposed to the environmental risk factors actually develop ECCA, and up to 90% of patients presenting with ECCA have no identifiable risk factors, suggesting that genetic susceptibility might contribute to the individual risk of ECCA. It is widely accepted that ERCC1-XPF enzyme complex was required for the nucleotide excision repair [24], DNA double-strand break repair [25, 26] and interstrand crosslink repair [27-29] pathways. Polymorphisms in ERCC1 and XPF could affect DNA repair capability. Previous meta-analyses have indicated that ERCC1 polymorphisms were associated with the risk of different kinds of cancers [30].

	rs321	2986		rs2298881				
Variables	(Cases/	Controls)	OR (95% CI)	Р	(Cases/Controls)		OR (95% CI)	Р
	CC	AC+AA			AA	AC+CC		
Age								
≤ 65	36/49	41/35	1.59 (0.85-2.97)	0.14	35/45	42/39	1.38 (0.74-2.58)	0.30
> 65	23/37	27/24	1.81 (0.85-3.86)	0.12	22/33	28/28	1.50 (0.71-3.18)	0.29
Gender								
Male	33/53	35/32	1.76 (0.92-3.36)	0.09	32/47	36/38	1.39 (0.73-2.64)	0.31
Female	26/33	33/27	1.55 (0.75-3.20)	0.23	25/31	34/29	1.45 (0.71-3.00)	0.31
Smoking								
No	45/65	46/47	1.41 (0.81-2.47)	0.22	42/55	49/57	1.13 (0.65-1.96)	0.68
Yes	14/21	22/12	2.75 (1.04-7.30)	0.04	15/23	21/10	3.22 (1.19-8.71)	0.02
Alcohol								
No	48/74	54/52	1.60 (0.95-2.71)	0.08	47/68	55/58	1.37 (0.81-2.32)	0.24
Yes	11/12	14/7	2.18 (0.64-7.40)	0.21	10/10	15/9	1.67 (0.50-5.56)	0.41
BMI (kg/m²)								
≤ 18.5	4/4	6/5	1.20 (0.19-7.44)	0.84	6/6	4/3	1.33 (0.20-8.71)	0.76
18.5-22.9	27/38	24/22	1.54 (0.72-3.28)	0.27	18/31	33/29	1.96 (0.91-4.21)	0.09
23.0-24.9	21/29	25/20	1.73 (0.77-3.89)	0.19	24/28	22/21	1.22 (0.54-2.75)	0.63
> 25	7/15	13/12	2.32 (0.70-7.64)	0.17	9/13	11/14	1.13 (0.36-3.62)	0.83
Family history of cancer								
No	50/76	53/53	1.52 (0.90-2.56)	0.12	46/71	57/58	1.52 (0.90-2.55)	0.12
Yes	9/10	15/6	2.78 (0.75-10.26)	0.13	11/7	13/9	0.92 (0.26-3.28)	0.90

Table 4. Subgroup analysis for associations between ERCC1 variant genotypes and risk of extrahe-
patic cholangiocarcinom

Our data showed that ERCC1 rs3212986 genotype AC+AA frequency and rs2298881 genotype CC frequency were significant association with increased risk of ECCA, especially in smokers. To date, only a few studies have addressed the contribution of genetic variants of so called 'susceptibility' genes to ECCA risk. Glutathione S-transferase omega 1 (GSTO1) [31], 5, 10-methylenetetrahydrofolate reductase (MTHFR) plus thymidylate synthase enhancer region (TSER) [32], X-ray repair crosscomplementing group 1 (XRCC1), apurinic/apyrimidinic endonuclease (APEX1) [33] and MutY homolog (MYH) polymorphisms were reported with an increased susceptibility to CCA, while N-acetyltransferase 2 [34] might significantly decrease the cancer risk. To our knowledge, this study is the first one which providing data on ERCC1 and XPF genetic variant and ECCA risk. Compared with subjects carrying the ERCC1 rs3212986 genotype CC, those with dominant model AC+AA had a 1.68-fold risk of ECCA (OR: 1.68, 95% CI: 1.04-2.72), while in smokers an extra increased risk was observed with a 2.75-fold risk (OR: 2.75, 95% CI: 1.047.30). Similarly, the ERCC1 rs2298881 genotype CC was associated with a 2.15-fold risk of ECCA (OR: 2.15, 95% CI: 1.01-4.56) comparing with genotype AA and an extra 3.22-fold increased risk of AC+CC genotype in smokers (OR: 3.22, 95% CI: 1.19-8.71). The relationship between ERCC1 polymorphism and ECCA risk needs to be clarified by more large size studies.

In this study, all our control subjects were under Hardy-Weinberg equilibrium minimizing population stratification. We conducted quality control strictly throughout the whole study. The controls were frequency matched and the investigators were unified-trained rigorously. Moreover, we sequenced the five SNPs duplicated and verified them by repeated 10% of randomly selected samples, making the results credible. We entirely noticed that our findings were based on a small number of cases and, therefore, the biologic significance of the results might be limited. However, considering the low incidence of ECCA, well-characterized cohorts are difficult to obtain. Several limitations and sources of bias of this study should be addressed. Firstly, like all other case-control studies, inherent biases like selection bias and recall bias in the present study might have led to some spurious results. Secondly, the present study only investigated the ERCC1 and XPF gene polymorphism and ECCA risk. Many popular gene variants reported in other cancers were not investigated here. Thirdly, the present study only adjusted age, gender, smoking, BMI and family history of cancer. Other known risk factors, such as liver fluke infection, HBV/HCV infection, and cholelithiasis were not controlled which might present a bias in the results. Although the relatively small sample size of our study showed significant results, a more comprehensive approach including environmental factors might improve the results.

#### Conclusions

The present study suggested that rs3212986 C > A and rs2298881 A > C polymorphisms of ERCC1 were associated with an increased risk of ECCA, especially in smokers. It would be necessary to confirm these findings in a large sample size and multiethnic population study in future, because of the relatively small sample size in this study and limited gene-environment interaction analysis. The underlying mechanism of cholangiocarcinogenesis needs to be further investigated.

#### Acknowledgements

This work was supported by Shandong Provincial Medical Science & Technology Development Program (No. 2015WSB04031) awarded to Liu Feng.

All participants signed the informed consent. This study did not involve the use of any animal.

#### Disclosure of conflict of interest

None.

Address correspondence to: Feng Liu, Department of Hepatobiliary Surgery, Qianfoshan Hospital, Shandong University, No. 16766, Jingshi Road, Jinan 250014, Shandong Province, China. Tel: +86-183-02045970; E-mail: qfsliufeng@126.com

#### References

- [1] Isomoto H. Epigenetic alterations associated with cholangiocarcinoma (review). Oncol Rep 2009; 22: 227-232.
- [2] Petrowsky H, Hong JC. Current surgical management of hilar and intrahepatic cholangiocarcinoma: the role of resection and orthotopic liver transplantation. Transplant Proc 2009; 41: 4023-4035.
- [3] Stein A, Arnold D, Bridgewater J, Goldstein D, Jensen LH, Klumpen HJ, Lohse AW, Nashan B, Primrose J, Schrum S, Shannon J, Vettorazzi E, Wege H. Adjuvant chemotherapy with gemcitabine and cisplatin compared to observation after curative intent resection of cholangiocarcinoma and muscle invasive gallbladder carcinoma (ACTICCA-1 trial) - a randomized, multidisciplinary, multinational phase III trial. BMC Cancer 2015; 15: 564.
- [4] Valle JW, Wasan H, Johnson P, Jones E, Dixon L, Swindell R, Baka S, Maraveyas A, Corrie P, Falk S, Gollins S, Lofts F, Evans L, Meyer T, Anthoney A, Iveson T, Highley M, Osborne R, Bridgewater J. Gemcitabine alone or in combination with cisplatin in patients with advanced or metastatic cholangiocarcinomas or other biliary tract tumours: a multicentre randomised phase II study - The UK ABC-01 Study. Br J Cancer 2009; 101: 621-627.
- [5] Gatto M, Bragazzi MC, Semeraro R, Napoli C, Gentile R, Torrice A, Gaudio E, Alvaro D. Cholangiocarcinoma: update and future perspectives. Dig Liver Dis 2010; 42: 253-260.
- [6] Tran B, Whiteman DC, Webb PM, Fritschi L, Fawcett J, Risch HA, Lucas R, Pandeya N, Schulte A, Neale RE. Association between ultraviolet radiation, skin sun sensitivity and risk of pancreatic cancer. Cancer Epidemiol 2013; 37: 886-892.
- [7] Shields PG, Harris CC. Cancer risk and lowpenetrance susceptibility genes in gene-environment interactions. J Clin Oncol 2000; 18: 2309-2315.
- [8] Galvan A, Ioannidis JP, Dragani TA. Beyond genome-wide association studies: genetic heterogeneity and individual predisposition to cancer. Trends Genet 2010; 26: 132-141.
- [9] Martin SA, Hewish M, Lord CJ, Ashworth A. Genomic instability and the selection of treatments for cancer. J Pathol 2010; 220: 281-289.
- [10] Jiang J, Zhang X, Yang H, Wang W. Polymorphisms of DNA repair genes: ADPRT, XRCC1, and XPD and cancer risk in genetic epidemiology. Methods Mol Biol 2009; 471: 305-333.
- [11] Westerveld A, Hoeijmakers JH, van Duin M, de Wit J, Odijk H, Pastink A, Wood RD, Bootsma D. Molecular cloning of a human DNA repair gene. Nature 1984; 310: 425-429.

- [12] Zhu G, Wang L, Guo H, Lu L, Yang S, Wang T, Guo H, Wang H, Min J, Yang K, Chen X, Liu Y, Wang Z, Su H. DNA repair genes XRCC1 and ERCC1 polymorphisms and the risk of sporadic breast cancer in Han women in the Gansu Province of China. Genet Test Mol Biomarkers 2015; 19: 387-393.
- [13] Pei XH, Yang Z, Lv XQ, Li HX. Genetic variation in ERCC1 and XPF genes and breast cancer risk. Genet Mol Res 2014; 13: 2259-2267.
- [14] Yang H, Li G, Li WF. Association between ERCC1 and XPF polymorphisms and risk of colorectal cancer. Genet Mol Res 2015; 14: 700-705.
- [15] Ni M, Zhang WZ, Qiu JR, Liu F, Li M, Zhang YJ, Liu Q, Bai J. Association of ERCC1 and ERCC2 polymorphisms with colorectal cancer risk in a Chinese population. Sci Rep 2014; 4: 4112.
- [16] Dai Q, Luo H, Li XP, Huang J, Zhou TJ, Yang ZH. XRCC1 and ERCC1 polymorphisms are related to susceptibility and survival of colorectal cancer in the Chinese population. Mutagenesis 2015; 30: 441-449.
- [17] He J, Xu Y, Qiu LX, Li J, Zhou XY, Sun MH, Wang JC, Yang YJ, Jin L, Wei QY, Wang Y. Polymorphisms in ERCC1 and XPF genes and risk of gastric cancer in an eastern Chinese population. PLoS One 2012; 7: e49308.
- [18] Hui L, Yue S, Gao G, Chang H, Li X. Association of single-nucleotide polymorphisms in ERCC1 and ERCC2 with glioma risk. Tumour Biol 2014; 35: 7451-7457.
- [19] Khan SA, Davidson BR, Goldin R, Pereira SP, Rosenberg WM, Taylor-Robinson SD, Thillainayagam AV, Thomas HC, Thursz MR, Wasan H. Guidelines for the diagnosis and treatment of cholangiocarcinoma: consensus document. Gut 2002; 51 Suppl 6: VI1-9.
- [20] Jarnagin WR, Fong Y, DeMatteo RP, Gonen M, Burke EC, Bodniewicz BJ, Youssef BM, Klimstra D, Blumgart LH. Staging, resectability, and outcome in 225 patients with hilar cholangiocarcinoma. Ann Surg 2001; 234: 507-517, 517-519.
- [21] Khan SA, Toledano MB, Taylor-Robinson SD. Epidemiology, risk factors, and pathogenesis of cholangiocarcinoma. HPB (Oxford) 2008; 10: 77-82.
- [22] Kubo S, Kinoshita H, Hirohashi K, Hamba H. Hepatolithiasis associated with cholangiocarcinoma. World J Surg 1995; 19: 637-641.
- [23] Okuda K, Nakanuma Y, Miyazaki M. Cholangiocarcinoma: recent progress. Part 1: epidemiology and etiology. J Gastroenterol Hepatol 2002; 17: 1049-1055.
- [24] Sijbers AM, de Laat WL, Ariza RR, Biggerstaff M, Wei YF, Moggs JG, Carter KC, Shell BK, Evans E, de Jong MC, Rademakers S, de Rooij J, Jaspers NG, Hoeijmakers JH, Wood RD. Xeroderma pigmentosum group F caused by a defect in a structure-specific DNA repair endonuclease. Cell 1996; 86: 811-822.

- [25] Ahmad A, Robinson AR, Duensing A, van Drunen E, Beverloo HB, Weisberg DB, Hasty P, Hoeijmakers JH, Niedernhofer LJ. ERCC1-XPF endonuclease facilitates DNA double-strand break repair. Mol Cell Biol 2008; 28: 5082-5092.
- [26] Niedernhofer LJ, Essers J, Weeda G, Beverloo B, de Wit J, Muijtjens M, Odijk H, Hoeijmakers JH, Kanaar R. The structure-specific endonuclease Ercc1-Xpf is required for targeted gene replacement in embryonic stem cells. EMBO J 2001; 20: 6540-6549.
- [27] Wood RD. Mammalian nucleotide excision repair proteins and interstrand crosslink repair. Environ Mol Mutagen 2010; 51: 520-526.
- [28] Sargent RG, Rolig RL, Kilburn AE, Adair GM, Wilson JH, Nairn RS. Recombination-dependent deletion formation in mammalian cells deficient in the nucleotide excision repair gene ERCC1. Proc Natl Acad Sci U S A 1997; 94: 13122-13127.
- [29] Klein DD, Boonen RA, Long DT, Szypowska AA, Raschle M, Walter JC, Knipscheer P. XPF-ERCC1 acts in Unhooking DNA interstrand crosslinks in cooperation with FANCD2 and FANCP/SLX4. Mol Cell 2014; 54: 460-471.
- [30] Zhang L, Wang J, Xu L, Zhou J, Guan X, Jiang F, Wu Y, Fan W. Nucleotide excision repair gene ERCC1 polymorphisms contribute to cancer susceptibility: a meta-analysis. Mutagenesis 2012; 27: 67-76.
- [31] Marahatta SB, Punyarit P, Bhudisawasdi V, Paupairoj A, Wongkham S, Petmitr S. Polymorphism of glutathione S-transferase omega gene and risk of cancer. Cancer Lett 2006; 236: 276-281.
- [32] Ko KH, Kim NK, Yim DJ, Hong SP, Park PW, Rim KS, Kim S, Hwang SG. Polymorphisms of 5, 10-methylenetetrahydrofolate reductase (MT-HFR C677T) and thymidylate synthase enhancer region (TSER) as a risk factor of cholangiocarcinoma in a Korean population. Anticancer Res 2006; 26: 4229-4233.
- [33] Huang WY, Gao YT, Rashid A, Sakoda LC, Deng J, Shen MC, Wang BS, Han TQ, Zhang BH, Chen BE, Rosenberg PS, Chanock SJ, Hsing AW. Selected base excision repair gene polymorphisms and susceptibility to biliary tract cancer and biliary stones: a population-based case-control study in China. Carcinogenesis 2008; 29: 100-105.
- [34] Prawan A, Kukongviriyapan V, Tassaneeyakul W, Pairojkul C, Bhudhisawasdi V. Association between genetic polymorphisms of CYP1A2, arylamine N-acetyltransferase 1 and 2 and susceptibility to cholangiocarcinoma. Eur J Cancer Prev 2005; 14: 245-250.