

Review Article

Treatment of bronchopulmonary dysplasia by vascular endothelial growth factor: the earlier the better?

Jinghua Luo^{1,2}, Yingjian Si¹, Jia Chen¹, Zhichun Feng¹

¹Affiliated Bayi Children's Hospital, Clinical Medical College in PLA Army General Hospital, Southern Medical University, Beijing, China; ²Department of Pediatric, The Second Affiliated Hospital of Nanchang University, Nanchang, Jiangxi, P.R. China

Received August 7, 2017; Accepted November 29, 2017; Epub February 15, 2018; Published February 28, 2018

Abstract: Bronchopulmonary dysplasia (BPD) is a chronic lung disease that most commonly occurs in premature infants who have needed mechanical ventilation and oxygen therapy for acute respiratory distress, but can also occur in immature infants who have had few signs of initial lung disease. Vascular endothelial growth factor (VEGF) has been shown to play a central role in vascular development. VEGF is a potent endothelial cell-specific mitogen and survival factor that stimulates angiogenesis, promotes vessel remodeling, and enhances endothelial survival. VEGF signaling is absolutely critical for vascular development and embryonic survival, and appears to protect the lung against hyperoxia or cytokine-induced endothelial cell injury. Whether disruption of VEGF signaling impairs lung vascular growth and contributes to the pathogenesis of BPD has been uncertain. Since the establishment of "vascular hypothesis of BPD", vascular endothelial growth factor (VEGF) has been adopted as one of the means for the treatments of BPD. However, the time of using VEGF is not unified. In this review, we firstly introduced the definition of BPD, and then explored the pathology, roles and mechanisms of VEGF in BPD, and finally briefly summarized the timing of using VEGF to treat BPD.

Keywords: Bronchopulmonary dysplasia, vascular endothelial growth factor, endothelial cells, treatment

Introduction

Bronchopulmonary dysplasia (BPD) is the most common disease among surviving premature infants and is associated with poor outcomes of long-term lung maturity and neuro-development [1-4]. Fortunately, various treatments for BPD have been developed, greatly increasing the survival rate of premature infants. The "old" BPD focuses mainly on lung injury resulting from oxygen therapy and mechanical ventilation, while the "new" BPD, on abnormalities in the lung development [1, 5, 6]. Recent reviews have shown that pulmonary vascular disease has become the new frontier of BPD research [5, 7]. According to the "vascular hypothesis of BPD", disruption of angiogenesis during lung maturity could impair lung development by decreasing alveolarization and pulmonary arterial density [8]. VEGF is a major mediator of vascular permeability, endothelial cell proliferation and migration, which is very important in vasculogenesis and angiogenesis [9, 10]. Previous studies have demonstrated that the expression of VEGF mRNA and protein decreased in alveolar lavage fluid or peripheral blood in children

with BPD or in animal models [11, 12]. Researchers have begun to explore the use of VEGF replacement therapy in BPD [13, 14]. However, the diagnosis of BPD is currently based on the need for supplemental oxygen for at least 28 days after birth, and BPD is classified into several grades according to the respiratory support required at 36 postmenstrual weeks [15, 16]. So will it be late to take VEGF at 36 postmenstrual weeks? Is it necessary to give VEGF treatment in 24 hours after birth? The earlier the better? With the above questions in mind, we reviewed the pathophysiological process of BPD, the mechanism of VEGF involved in angiogenesis and stabilization, and the status quo of VEGF application in the treatment of BPD, aiming to determine the right time of taking VEGF.

Definition of BPD

Bronchopulmonary dysplasia (BPD) was first defined by Northway and his coworkers in 1967 [17]. It was described as prolongation of the healing phase of respiratory-distress syndrome combined with a generalized pulmonary oxygen

toxicity involving mucosal, alveolar and vascular tissues. They stressed the need of neonates for the oxygen therapy 28 days after birth, presence of clinical symptoms and visible chest changes revealed by X-ray as the diagnostic criteria for BPD [18]. The incidence of BPD ranged from 6% to 57% between 1978 and 2015, depending on the definition chosen [19, 20]. "BPD" is an operational definition in which the treatment (oxygen therapy at 28th day or 36th week postmenstrual age) is used to define the disease [21], so with the improvement of treatment, the definition of BPD varies. According to Hine's review, the definition by Shennan and his coworkers was adopted in 45% of a total of 628 papers reviewed [22], the NICHD definition, in 30% [15] (**Table 1**), and the physiological definition indicated by the oxygen challenge test, in approximately 6%. BPD is associated with significant morbidity and mortality in the neonatal intensive care unit [23] and is also associated with worse long-term outcomes such as increased airway hyperresponsiveness in childhood, abnormal lung function in young adults, and potentially earlier come up of chronic obstructive pulmonary disease [21, 24, 25].

Fortunately, the introduction of antenatal steroids, natural surfactant therapy, lower supplemental oxygen concentrations and gentler ventilation techniques altered the clinical course and pathology exhibited by preterm infants [26, 27]. Vollsaeter and his coworkers compared preterm infants with a gestational age <28 weeks or a birth weight <1000 g in western Norway from 1999-2000 with those in 1991-1992. They found that for children with neonatal BPD, important lung function variables were better in EP1999-2000 than in EP1991-1992. In regression models, administration of antenatal corticosteroids and surfactant treatment improved the lung function in the EP1999-2000 [28] but failed to benefit others in a visible way [15]. This new kind of BPD focused more on the interruption of normal development than lung injury from oxygen therapy and mechanical ventilation. These consist of very low birth weight infants who initially have mild or no lung diseases but whose need for oxygen and ventilatory increase over the first several weeks of life [29]. Some authors have described that kind of BPD as a "new" BPD [15, 30].

Pathology of the new BPD

The lungs, together with the trachea, arise from the anterior foregut endoderm (the 4-7 week of

gestation in humans). From 7 to 16 weeks' gestation, evagination of these epithelial cells result in the formation of the trachea and two lung buds and the beginning of the lung development at the embryonic stage. At this stage, the trachea separates from the esophagus [31-34]. In the course of the lung development, first the trachea is formed, which then generates the bronchial tree and finally the airways which is largely in parallel with the vasculature of the pulmonary circulation [35]. Subsequent lung development at different stages including the canalicular, saccular and alveolar structures generate the alveolar-gas exchange units [36]. The lung at 26 weeks of gestation is just at the canalicular stage and is of the saccular structure without alveoli, which does not begin to develop in another 4 to 6 weeks [15]. About at 30 to 32 weeks, the lung is at the saccular stage. With the growth of terminal saccules, extensive vessels are generated, and then the secondary crests occur along with the loss and remodel of interstitial extracellular matrix [37]. Although alveoli appear in some infants at 32 weeks of gestation, they do not uniformly grow up to 36 weeks at the stage of alveolar, and they continue to grow at a slower rate during the first 2-3 years after birth [8]. Thus, premature births and the initiation of pulmonary gas exchange will interrupt the development of normal alveolar and distal vascular, thereby becoming the two major features of the new BPD [38]. The "old" BPD was characterized by severe lung injury, pronounced inflammation, lung edema, airway epithelial metaplasia, peribronchial fibrosis, and remarkable hypertrophy of airway and pulmonary vascular smooth muscle [17, 39]. However, the "new" BPD is characterized by alveolar hypoplasia (fewer and larger alveoli), thickened alveolar septa, dysmorphic pulmonary microvascular networks, mild hypertrophy of airway and vascular smooth muscle, accumulation of interstitial fluid, abnormal deposition of extracellular matrix components and an arrest of lung development at the late canalicular to early saccular stage [40].

We reviewed literatures over the past three decades and found the role of vascular dysplasia in new BPDs, which are receiving increasing attention. We summarized the pathological manifestations of the lungs in some animal models in the table (**Table 2**), to better show the pathological features of the new BPD. Soliman et al performed a prospective cohort

VEGF in bronchopulmonary dysplasia

Table 1. Definition of BPD (NICHD consensus 2001)

Gestational Age	<32 wk	≥32 wk
Time point of assessment	36 wk PMA or discharge to home, whichever comes first	Time point of assessment: >28 days but <56 days postnatal age or discharge to home, whichever comes first
Treatment with oxygen 21% for at least 28 d plus		
Mild BPD	Breathing room air at 36 weeks PMA or discharge, whichever comes first	Breathing room air at 56 days postnatal age or discharge, whichever comes first
Moderate BPD	Need for <30% oxygen at 36 weeks PMA or discharge, whichever comes first	Need for <30% oxygen at 56 days postnatal age or discharge, whichever comes first
Severe BPD	Need for ≥30% oxygen and/or positive pressure, (positive pressure ventilation or NCPAP) at 36 weeks PMA or discharge, whichever comes first	Need for ≥30% oxygen and/or positive pressure, (positive pressure ventilation or NCPAP) at 56 days PMA or discharge, whichever comes first

Definition of abbreviations: BPD bronchopulmonary dysplasia; NCPAP nasal continuous positive airway pressure; PMA postmenstrual age; PPV positive-pressure ventilation.

Table 2. Pathology of the BPD

Author	Model/human	Pathophysiological characteristics
Northway WH Jr et al. 1967 [17]	Human lung slices	Severe lung injury, pronounced inflammation, lung edema, airway epithelial metaplasia, peribronchial fibrosis, and marked airway and pulmonary vascular smooth muscle hypertrophy
Gorenflo M et al. 1991 [98]	Lung slices and barium angiogram	Decreased density of peripheral pulmonary arteries.
Cherukupalli K. et al. 1996 [99]	Human lung slices	a. Group I was a phase of acute lung injury; Alveolar hyaline membrane, alveolar epithelial necrosis. b. Group II the proliferative phase; Cell metaplasia, airway epithelium ulcer. c. Group III the phase of early repair; Extensive type II metaplasia, pulmonary fibroblasts rich in interstitial. d. Group IV the phase of late repair; Airway epithelium phosphorylation, bronchial smooth muscle fibrosis.
Husain A et al. 1998 [100]	Human lung slices	a. No surfactant therapy: alveolar septal fibrosis, partial to complete arrest in acinar development (alveolar saccular and alveolar). b. Use surfactant therapy: less phosphorus-like metaplasia.
Coalson JJ et al. 1999 [101]	Baboons appropriate oxygen (1-2 m)	Decreased pulmonary microvascular development and alveolarization
Bhatt AJ et al. 2001 [11]	Human lung slices	Alveolar capillaries were often located in the interior of thickened septa. dilated and lacked extensive network organization.
Coalson JJ 2003 [38]	Baboons and Clinical specimens	"Emphysematous" distal lung structure with fewer 51 lung units, areas of septal thickening, microvascular dysplasia/hypoplasia and inflammation.
De Paepe ME et al. 2006 [44]	Postmortem lung samples	The microvasculature of ventilated lungs appeared immature, retaining a saccular architectural pattern.
Velten M et al. 2010 [102]	C3H/HeN mice (85% O ₂ , 14 d pre-natal LPS)	Decreased alveolar number and increased size.
O'Reilly M et al. 2014 [103]	Mouse (65% O ₂ , 7 d)	More smooth muscle; no effect on bronchiolar epithelium or collagen.
Firsova AB et al. 2014 [104]	Mouse (95% O ₂ , 5 d)	Airspaces were significantly enlarged.
Belcastro R et al. 2015 [105]	Rat lung (60% O ₂ , 14 d)	Impairments of lung cell proliferation, secondary crest formation, and alveologensis.
Mankouski A et al. 2016 [2]	Sprague-Dawley rats (60% O ₂ , 14 d)	Decreased numbers of secondary crests and peripheral vessels.
Jiménez J et al. 2016 [106]	Rabbits	Fewer and larger alveoli with thicker walls, less developed distal airways and more inflammation.
Chou HC et al. 2016 [56]	Prenatal LPS (85% O ₂ , 14 d)	Reduced vascular density

study, from January 2007 to June 2010 at a single tertiary care center, with infants less than 32 weeks' gestation born to mothers with preeclampsia, and found that preeclampsia, an antiangiogenic state, is an independent risk factor of bronchopulmonary dysplasia (BPD) [41]. Baud et al found that angiogenesis blocked by vascular endothelial growth factor (VEGF)-Trap decreased the number of lung capillaries and enlarged the size of alveoli, which is similar to pathological manifestations of BPD [7, 42]. This suggests that angiogenesis plays an important role in alveolarization. It is noted that glucocorticoids are widely administered to accelerate the maturation of AEC2 cells and production of surfactant in premature babies, which appear to inhibit secondary septation and vascular development [33]. So far, the only consistent vascular findings in new BPD pathology are that the structural configuration of the distal microvasculature is abnormal, namely dysmorphic [37]. This kind of dysmorphism shows an abnormal distribution of alveolar capillaries in lungs, the vessels being far away from the air surface [43] and the dysmorphia being of a saccular architectural pattern [44].

The role of VEGF in BPD

VEGF family

Vascular endothelial growth factor (VEGF) is a multifunctional cytokine which plays a key role in many physiological (angiogenesis, growth and organ repair) and pathological (vascular disease) processes [45]. The VEGF gene is located on chromosome 6q21.3, and consists of eight exons and seven introns [46]. Multiple isoforms of VEGF, ranging from 121 to 206 amino acids [47], can be generated by alternative exon splicing, and these isoforms differ in their ability to bind heparin, which determines their bioavailability and may play distinct roles in angiogenesis during development [48-50]. In humans, VEGF is made up of five secreted glycoproteins which include VEGF-A, VEGF-B, VEGF-C, VEGF-D and placental growth factor (PlGF) [48]. VEGF-E is encoded by certain viruses and its gene is not contained within the human genome [51]. VEGF-A activates intracellular signaling pathways by binding to one of the two receptors: VEGF receptor-1 (VEGFR-1, previously termed fms-like tyrosine kinase-1 [Flt-1]) and VEGFR-2 (previously termed murine

fetal liver kinase-1 [Flk-1] or kinase domain region [KDR] in humans) [52, 53]; as well as two co-receptors: neuropilin-1 (NRP1) and neuropilin-2 (NRP2). NRP1 enhanced VEGF signaling has been shown to be important for p38/MAPK activation, and is thus central to vessel branching [54, 55]. The expression of VEGFR3 is mainly restricted by the lymphatic endothelium in adult tissues. It binds VEGF-C and VEGF-D but not VEGF-A. And VEGFR3 is considered to control lymphangiogenesis [56]. VEGF mRNA can be firstly detected in fetal tissues at 16 weeks of gestation [57]. The expression of VEGF is particularly high in the lung, where it is essential in lung development and maintaining the structure of lung [58]. In human fetal lung, VEGF is localized in alveolar epithelial cells and myocytes, which suggested that VEGF acts a paracrine in modulating the activity of adjacent vascular endothelium [57]. In patients with BPD, VEGF also arises in Type II pneumocytes.

Roles of VEGF in BPD

In a comparative study of the causes between infants dying with BPD and non-pulmonary diseases, Bhatt found the former group had lower VEGF mRNA level and VEGF immunostaining than did the latter group [11]. Another study which investigated the expression of VEGF in tracheal aspirates revealed that preterm infants who developed BPD had lower VEGF levels during the early postnatal days than those without BPD. That suggests a prolonged and more severe respiratory distress [57]. Administration of anti-angiogenic agents to neonatal rats impairs both pulmonary angiogenesis and alveolarization [59-63]. Over-expression of proangiogenic factors, such as vascular endothelial growth factor (VEGF), alleviates the adverse effects of hyperoxia on Alveolarization [7, 42]. Inactivation of the VEGF-A gene in respiratory epithelium results in an absence of pulmonary capillaries, suggesting that the development of pulmonary capillary is in a VEGF-A dependent manner [64]. As a matter of fact, previous treatments for BPD with inhibitors of VEGF-A have shown that inhibition of angiogenesis seriously affected the formation of alveolar [59, 65].

Expression of VEGF-A is regulated by many factors including hypoxia (hypoxia-inducible factors-1 α , HIF-1 α), oncogene and tumor suppres-

VEGF in bronchopulmonary dysplasia

Table 3. VEGF in BPD

Author	Model/human	Deal with	VEGF levels			Time of VEGF changing
			mRNA	Protein	Location	
Bhatt AJ et al. 2001 [11]	Human	Dead form BPD	↓	↓	Autopsy samples	P*65 ± 34 d
Tambunting F et al. 2005 [107]	Baboon	125 days gestation O ₂	↓	↓	Lung specimens	P14 d
Balasubramaniam V et al. 2007 [12]	Neonatal mice	80% O ₂ 10 d		↓	Blood, lung, and bone marrow	P10 d
Been JV et al. 2010 [93]	Preterm infants	—		↓	BALF concentrations	P0 d, P3 d
Grisafi D et al. 2013 [108]	Rats	60%O ₂ , 14 d	↓	↓	Lung sections	P14 d
Keenaghan M et al. 2013 [97]	Rats	10%, 21%, 30%, 40%, 50%, 60%, 70%, 80%, 90%, or 100% FiO ₂ for 2 h.		↓	Serum and lung	40% O ₂ 2 h
Firsova et al. 2014 [104]	Neonatal mice	95% O ₂ , 5 d		N [#]	Lung sections	p5, p28, and p56 d
Yang WC et al. 2015 [109]	Preterm infants	—		N	Cord blood	P0 d
Lajko M et al. 2016 [110]	Neonatal mice	75% o ₂ , P0-P14. room air 1 (P15), 7 (P21), or 14 days (P28)		↑	Retinal	p21 d
Kumar VH et al. 2016 [66]	Newborn mouse	85% O ₂ , P3-P15 Room air 15 weeks		↑	Lung sections	P15 w
Jin M et al. 2016 [111]	Newborn rats	21% or 85% O ₂ 7 d, room air 14 d	↓		Lung tissues	P7 d
Procianoy RS et al. 2016 [112]	Preterm neonates	72 h collected blood		↑	Peripheral blood	P72 h

Definition of abbreviations: P*: postnatal day; N[#]: normal.

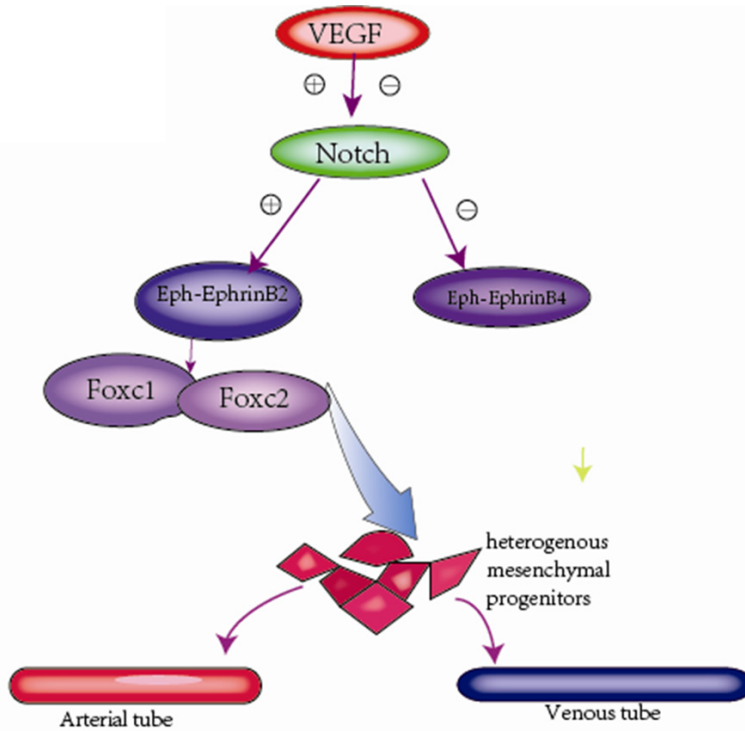


Figure 1. Angioblasts differentiate into endothelial cells which are prespecified to arterial or venous phenotypes by Notch signaling. Endothelial differentiation: Arterial and venous specification. when Notch increase, the Arterial tube conforming, and, when not decrease, the venous tube conforming.

Mechanisms of VEGF take part in vasculogenesis and angiogenesis

The formation of new blood vessels can be divided into two stages: vasculogenesis and angiogenesis [69]. Vasculogenesis starts from angioblasts or endothelial precursor cells which migrate and differentiate into local cues (growth factors, extracellular matrix), and further develop into vascular tubes, a process from nil to existence. Angiogenesis is the formation of new blood vessels from preexisting ones, which is a process from less to more [70]. However, VEGF is involved in many aspects of angiogenesis, including survival, proliferation, migration, tubulogenesis, remodeling and quiescence.

Differentiation of endothelial cells

...sor dysregulation, transcription factors (TGF- α , TGF- β), inflammatory mediators (IL-1 α , IL-1 β , IL-6, TNF α), and mechanical forces of shear stress [47]. Therefore, mechanical ventilation and hyperoxia will, theoretically, increase inflammatory factors [66] and shear stress, and consequentially, the level of VEGF. But most studies showed that expression of VEGF in histological sections of BPD patients or animal models decreased (Table 3). However, Tomanek's study on explanted embryonic quail hearts indicates that vascular formation can be enhanced by hypoxia (5-10% O₂) and inhibited by hyperoxia [67]. Nevertheless, why does not VEGF increase in BPD? We proposed three possible reasons according to previous reports: 1. Severe lung injury may render VEGF incapable of responding to the inflammatory stimuli. 2. An increased level of VEGF after birth is locally secreted, since VEGF acts as a mediator of paracrine. And when lungs were injured by post-partum ventilators, infections, and oxygen/nitrogen free radicals, locally increased VEGF do not well accelerate vascular development; 3. Hyperoxia (Postpartum oxygen or ventilator support) inhibited HIF-1 α which can enhance the expression of VEGF [68].

Angioblasts are differentiated into endothelial cells (ECs). ECs develop into the cords and form a lumen, whose phenotype can be distinguished into artery or vein. Arterial and venous ECs possess the ability of identifying specific molecules [71, 72]. Components of the notch signaling pathway which is activated by VEGF are highly expressed in arteries and are deficient in veins. Thus, inhibition of the Notch signaling pathway causes loss of arterial markers and re-expression of specific genes in veins [72, 73]. The Notch signaling pathway also regulates the expression of members of Eph-Ephrin family and Ephrin-B2. Ephrin-B2 is increased in response to Notch, whereas its receptor EphB4 in venous ECs is repressed by Notch (Figure 1).

Angiogenesis

Angiogenesis (neovascularization) occurs through a series of steps which consist of angiogenic stimulus, sprouting, elongation and branching, formation of vessel lumen, anastomosis and finally stabilization [9]. ECs become motile and invasive and protrude filopodia in response to VEGF released by matrix metallo-

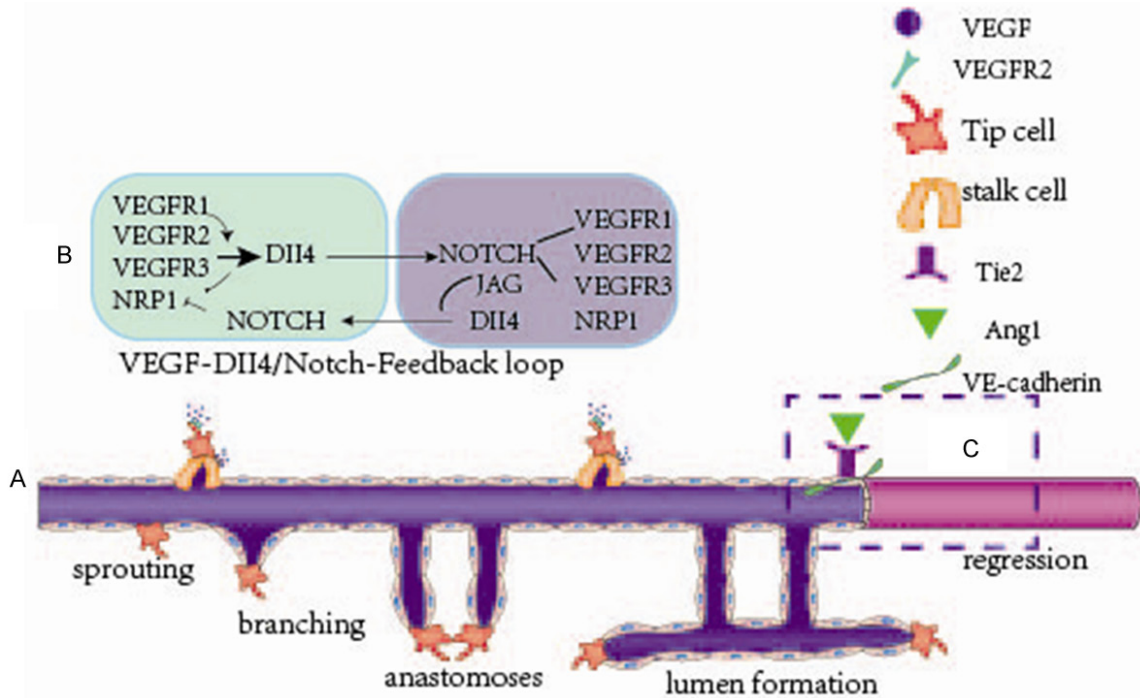


Figure 2. A: Steps of Angiogenesis; B: The feedback loop between VEGF and Notch; C: Ang1 activates Tie2 to stabilize vessels, promotes pericyte adhesion, and makes them leak.

proteinases (MMPs) [48], These so-called tip cells will sprout new ones; stalk cells seldom generate filopodia, but they establish a lumen and proliferate to support elongation of sprouts. Tip cells anastomose with cells from neighboring sprouts to set up vessel loops. Tip and stalk cells are affected by VEGF/Notch signaling [74] (Figure 2B). When blood begins to flow, the establishment of the basement membrane and the recruitment of mural cells stabilize new connections. The increase in oxygen and nutrient decreases the expression of VEGF and inactivates the sensors of endothelial oxygen with the blood perfusion, meanwhile the phenotype of endothelial behavior is shifted into a quiescent one (Figure 2A).

Maturation, stabilization, and quiescence of vessels

At the last stage of angiogenesis, the newly formed blood carries mural cells or pericytes to maintain stability of capillaries [75]. The role of pericytes in the function and angiogenesis of capillaries includes regulation of EC proliferation and migration, as well as production of basement membrane of capillary together with ECs [76]. Adherence junction molecules medi-

ate cell-cell adhesion, cytoskeletal reorganization, and intracellular signal transduction. VE-cadherin is one key component of EC junctions. In the case with VEGFR2 compound, VE-cadherin keeps EC static through dephosphorylate VEGFR2 to further inhibit VEGF signaling. Different types of VE-cadherin-based adherence junctions establish stable or transitory interactions with the cytoskeletons which can either solidify EC adhesion or facilitate EC separation and movement. Angiopoietin-1 (ANG1), produced by mural cells, activates its endothelial receptor TIE2 [77, 78] and plays a very important role in stabilizing the structure of vessels, promoting adhesions of pericytes, and tightening endothelial junctions (Figure 2C).

Exploring the application of VEGF in BPD treatment

Current methods of treating BPD include caffeine [79], nutrients, vitamin A [80], vitamin D [81], glucocorticoids [82], antibiotics [19], mesenchymal stromal cells (MSCs) [27, 83, 84] and BMSCs in combination with erythropoietin [85]. The VEGF gene was successfully used to treat limb after ischemia [86]. In recent years, researchers tried to promote angiogenesis of

bone tissue and ischemic myocardium through the VEGF gene therapy, and have made some satisfying achievements [87]. The application of VEGF in the treatment of BPD has also been investigated. Kunig et al [13] observed two-day-old Sprague-Dawley rats that were placed into hyperoxia or room air (RA) for 12 days. At 14 days, rats respectively received daily treatment with recombinant human VEGF (rhVEGF)-165 or saline. And they found rhVEGF treatment during the period of recovery accelerated vessels growth and alveolarization after hyperoxic lung injury in neonatal rats. He found fetal lung explants from eNOS(-/-) mice decreased the formation of terminal lung buds, it was restored with rhVEGF treatment [14], a finding similar to that of Seedorf's study. The postnatal intratracheal adenovirus-mediated VEGF gene therapy remarkably improves the survival, promotes the formation of lung capillaries, and preserves the development of alveolars in BPD model of irreversible lung injury [70]. To determine whether disruption of vascular endothelial growth factor receptor (VEGFR) signaling in the newborn has long-term effects on lung structure and function, Le Cras et al injected 1-day-old newborn rat pups with a single dose of Su-5416, a VEGFR inhibitor, or vehicle (controls). Lungs from infant (3-wk-old) and adult (3- to 4-mo-old) rats treated with Su-5416 showed reductions in arterial density (82 and 31%, respectively) and alveolar counts (45 and 29%) compared with the controls. Treatment for neonatals with Su-5416 increased right ventricle weight to body wt ratios (4.2-fold and 2.0-fold) and pulmonary arterial wall thickness measurements (2.7-fold and 1.6-fold) in infant and adult rats, respectively, indicating marked pulmonary hypertension. We conclude that treatment of newborn rats with the VEGFR inhibitor Su-5416 impairs the pulmonary vascular growth and postnatal alveolarization and causes pulmonary hypertension and that these are long-term effects lasting well into adulthood [88]. As the expression of HLA class I and II molecules are very low, MSCs cannot trigger an immune response once administered to animals or humans in an allogeneic MSCs [84, 89]. Moreover, MSCs have been shown to effectively ameliorate experimental BPD when administered in a preventive or therapeutic way [90, 91]. Chang studied intratracheal MSC transplantation which was performed in 9 preterm infants, with a mean gestational age of $25.3 \pm$

0.9 weeks and a mean birth weight of 793 ± 127 g, at a mean of 10.4 ± 2.6 days after birth. The first 3 patients were given a low dose (1×10^7 cells/kg) of cells, and the other 6 were given a high dose (2×10^7 cells/kg). Having compared their adverse outcomes, including BPD severity, with those of the historical case-matched comparison group, they conclude that intratracheal transplantation of allogeneic hUCB-derived MSCs in preterm infants is safe and feasible, and warrants a larger and controlled phase II study [92]. Several phase 1 and phase 2 trials are in progress (NCT02443961, NCT02381366, NCT01828957) [84]. MSCs, derived from bone marrow stroma with the ability of self-renewal, can be divided into mesodermal stem cells, and a variety of cells such as endothelial cells and endothelial progenitor cells. These cells can conjugate with VEGF and better to promote the formation of pulmonary vessels.

The timing of using VEGF to treat BPD

We found that infants diagnosed with BPD after birth did not have lower levels of VEGF in umbilical cord blood than infants without BPD (Table 3). Been's view [93] was different from other researcher's. He believes that VEGF in newborns with BPD decreased in the first day after birth. The possible reason is that the patients in his study have basic characteristics different from those in other studies, a lower gestational age of patients, for example. The results would be inconsistent. Higher lavage VEGF levels on days 1 and 3 were also correlated with a lower gestational age after birth [94]. The accumulation of VEGF may aggravate the body injury. Zeng's study found that over-expression of VEGF in fetal murine lungs not only enhanced pulmonary vasculogenesis but also resulted in an abnormal alveolar development [95]. It is not necessary to administer VEGF in the first day after birth. We summarized from previous studies that with the increase of the oxygen concentration, VEGF decreases sooner (Table 3). In the condition of moderate oxygen (60%) [12, 96], VEGF in mice decreases on the 14th day after birth. Here are two key points: Firstly, there exist difference between the models of mouse with BPD and humans with BPD, We still do not monitor the changes of VEGF in infants with BPD before they died. Whereas the autopsy materials from non-survivors with BPD present one avenue for the exploration of pathogen-

ic mechanisms at play in the lungs of affected patients. These materials are increasingly rare and difficult to obtain, because the survival rate of BPD patients has steadily increased over time. Both mice and rats are delivered at term in the saccular stage of lung development, and this fact is often used to justify the superiority of mice and rats as model animals for BPD, since preterm infants that develop BPD are also delivered in the saccular stage of lung development [1]; Secondly, the time of decline of VEGF is one key point in establishing the model of mouse with BPD. The amount of VEGF in lung sections still cannot be continuously monitored; whether VEGF is declined or not before BPD needs further researches. Keenaghan used FiO_2 exposed rats for 2 hours, and found VEGF decreased on 40% in 2 h [97].

Conclusion

VEGF signaling pathway acts as one key mechanism in the pathology of BPD, and treatment for infants with BPD by VEGF improves the outcome. We summarized that treatment of VEGF for infants with BPD before preterm infants 14 days after birth may effectively prevent BPD, but the exact time of treating for BPD still needs further researching.

Disclosure of conflict of interest

None.

Address correspondence to: Zhichun Feng, Affiliated Bayi Children's Hospital, Clinical Medical College in PLA Army General Hospital, Southern Medical University, Beijing, China. E-mail: zhjfengzc@126.com

References

- [1] Nardiello C, Mizikova I, Morty RE. Looking ahead: where to next for animal models of bronchopulmonary dysplasia? *Cell Tissue Res* 2017; 367: 457-468.
- [2] Mankouski A, Kantores C, Wong MJ, Ivanovska J, Jain A, Benner EJ, Mason SN, Tanswell AK, Auten RL, Jankov RP. Intermittent hypoxia during recovery from neonatal hyperoxic lung injury causes long-term impairment of alveolar development: a new rat model of BPD. *Am J Physiol Lung Cell Mol Physiol* 2017; 312: L208-L216.
- [3] Bancalari E, Claure N. Definitions and diagnostic criteria for bronchopulmonary dysplasia. *Semin Perinatol* 2006; 30: 164-170.
- [4] Short EJ, Klein NK, Lewis BA, Fulton S, Eisen-gart S, Kercksmar C, Baley J, Singer LT. Cognitive and academic consequences of broncho-pulmonary dysplasia and very low birth weight: 8-year-old outcomes. *Pediatrics* 2003; 112: e359.
- [5] Day CL, Ryan RM. Bronchopulmonary dysplasia: new becomes old again! *Pediatr Res* 2017; 81: 210-213.
- [6] Chao CM, Yahya F, Moiseenko A, Tiozzo C, Shrestha A, Ahmadvand N, El AE, Quantius J, Dilai S, Kheirollahi V, Jones M, Wilhem J, Carraro G, Ehrhardt H, Zimmer KP, Barreto G, Ahlbrecht K, Morty RE, Herold S, Abellar RG, Seeger W, Schermuly R, Zhang JS, Minoo P, Bellusci S. Fgf10 deficiency is causative for lethality in a mouse model of bronchopulmonary dysplasia. *J Pathol* 2017; 241: 91-103.
- [7] Alvira CM. Aberrant pulmonary vascular growth and remodeling in bronchopulmonary dysplasia. *Front Med (Lausanne)* 2016; 3: 21.
- [8] Abman SH. Bronchopulmonary dysplasia: "a vascular hypothesis". *Am J Respir Crit Care Med* 2001; 164: 1755-1756.
- [9] Potente M, Gerhardt H, Carmeliet P. Basic and therapeutic aspects of angiogenesis. *Cell* 2011; 146: 873-887.
- [10] Fujioka K, Shibata A, Yokota T, Koda T, Nagasaka M, Yagi M, Takeshima Y, Yamada H, Iijima K, Morioka I. Association of a vascular endothelial growth factor polymorphism with the development of bronchopulmonary dysplasia in Japanese premature newborns. *Sci Rep* 2014; 4: 4459.
- [11] Bhatt AJ, Pryhuber GS, Huyck H, Watkins RH, Metlay LA, Maniscalco WM. Disrupted pulmonary vasculature and decreased vascular endothelial growth factor, Flt-1, and TIE-2 in human infants dying with bronchopulmonary dysplasia. *Am J Respir Crit Care Med* 2001; 164: 1971-1980.
- [12] Balasubramaniam V, Mervis CF, Maxey AM, Markham NE, Abman SH. Hyperoxia reduces bone marrow, circulating, and lung endothelial progenitor cells in the developing lung: implications for the pathogenesis of bronchopulmonary dysplasia. *Am J Physiol Lung Cell Mol Physiol* 2007; 292: L1073-L1084.
- [13] Kunig AM, Balasubramaniam V, Markham NE, Morgan D, Montgomery G, Grover TR, Abman SH. Recombinant human VEGF treatment enhances alveolarization after hyperoxic lung injury in neonatal rats. *Am J Physiol Lung Cell Mol Physiol* 2005; 289: L529-L535.
- [14] Seedorf G, Metoxen AJ, Rock R, Markham N, Ryan S, Vu T, Abman SH. Hepatocyte growth factor as a downstream mediator of vascular endothelial growth factor-dependent preservation of growth in the developing lung. *Am J Physiol Lung Cell Mol Physiol* 2016; 310: L1098-L1110.

VEGF in bronchopulmonary dysplasia

- [15] Jobe AH, Bancalari E. Bronchopulmonary dysplasia. *Am J Respir Crit Care Med* 2001; 163: 1723-1729.
- [16] Jobe AH. What is BPD in 2012 and what will BPD become? *Early Hum Dev* 2012; 88 Suppl 2: S27-S28.
- [17] Northway WJ, Rosan RC, Porter DY. Pulmonary disease following respirator therapy of hyaline-membrane disease. Bronchopulmonary dysplasia. *N Engl J Med* 1967; 276: 357-368.
- [18] Bancalari E, Abdenour GE, Feller R, Gannon J. Bronchopulmonary dysplasia: clinical presentation. *J Pediatr* 1979; 95: 819-823.
- [19] Rudloff I, Cho SX, Bui CB, McLean C, Veldman A, Berger PJ, Nold MF, Nold-Petry CA. Refining anti-inflammatory therapy strategies for bronchopulmonary dysplasia. *J Cell Mol Med* 2017; 21: 1128-1138.
- [20] Hines D, Modi N, Lee SK, Isayama T, Sjors G, Gagliardi L, Lehtonen L, Vento M, Kusuda S, Bassler D, Mori R, Reichman B, Hakansson S, Darlow BA, Adams M, Rusconi F, San FL, Lui K, Morisaki N, Musrap N, Shah PS. Scoping review shows wide variation in the definitions of bronchopulmonary dysplasia in preterm infants and calls for a consensus. *Acta Paediatr* 2017; 106: 366-374.
- [21] Ambalavanan N, Morty RE. Searching for better animal models of BPD: a perspective. *Am J Physiol Lung Cell Mol Physiol* 2016; 311: L924-L927.
- [22] Shennan AT, Dunn MS, Ohlsson A, Lennox K, Hoskins EM. Abnormal pulmonary outcomes in premature infants: prediction from oxygen requirement in the neonatal period. *Pediatrics* 1988; 82: 527-532.
- [23] Hilgendorff A, O'Reilly MA. Bronchopulmonary dysplasia early changes leading to long-term consequences. *Front Med (Lausanne)* 2015; 2: 2.
- [24] Landry JS, Tremblay GM, Li PZ, Wong C, Benedetti A, Taivassalo T. Lung function and bronchial hyperresponsiveness in adults born prematurely. A cohort study. *Ann Am Thorac Soc* 2016; 13: 17-24.
- [25] Islam JY, Keller RL, Aschner JL, Hartert TV, Moore PE. Understanding the short- and long-term respiratory outcomes of prematurity and bronchopulmonary dysplasia. *Am J Respir Crit Care Med* 2015; 192: 134-156.
- [26] Strueby L, Thebaud B. Advances in bronchopulmonary dysplasia. *Expert Rev Respir Med* 2014; 8: 327-338.
- [27] O'Reilly M, Thebaud B. The promise of stem cells in bronchopulmonary dysplasia. *Semin Perinatol* 2013; 37: 79-84.
- [28] Vollaeter M, Skromme K, Satrell E, Clemm H, Roksund O, Oymar K, Markestad T, Halvorsen T. Children born preterm at the turn of the millennium had better lung function than children born similarly preterm in the early 1990s. *PLoS One* 2015; 10: e144243.
- [29] Bancalari E, Claire N, Sosenko IR. Bronchopulmonary dysplasia: changes in pathogenesis, epidemiology and definition. *Semin Neonatol* 2003; 8: 63-71.
- [30] D'Angio CT, Ambalavanan N, Carlo WA, McDonald SA, Skogstrand K, Hougaard DM, Shankaran S, Goldberg RN, Ehrenkranz RA, Tyson JE, Stoll BJ, Das A, Higgins RD. Blood cytokine profiles associated with distinct patterns of bronchopulmonary dysplasia among extremely low birth weight infants. *J Pediatr* 2016; 174: 45-51.
- [31] Herriges M, Morrisey EE. Lung development: orchestrating the generation and regeneration of a complex organ. *Development* 2014; 141: 502-513.
- [32] Morrisey EE, Cardoso WV, Lane RH, Rabinovitch M, Abman SH, Ai X, Albertine KH, Bland RD, Chapman HA, Checkley W, Epstein JA, Kintner CR, Kumar M, Minoo P, Mariani TJ, McDonald DM, Mukoyama YS, Prince LS, Reese J, Rossant J, Shi W, Sun X, Werb Z, Whitsett JA, Gail D, Blaisdell CJ, Lin QS. Molecular determinants of lung development. *Ann Am Thorac Soc* 2013; 10: S12-S16.
- [33] Morrisey EE, Hogan BL. Preparing for the first breath: genetic and cellular mechanisms in lung development. *Dev Cell* 2010; 18: 8-23.
- [34] Warburton D, Bellusci S, De Langhe S, Del MP, Fleury V, Mailleux A, Tefft D, Unbekandt M, Wang K, Shi W. Molecular mechanisms of early lung specification and branching morphogenesis. *Pediatr Res* 2005; 57: 26R-37R.
- [35] Rawlins EL. The building blocks of mammalian lung development. *Dev Dyn* 2011; 240: 463-476.
- [36] Morty RE, Konigshoff M, Eickelberg O. Transforming growth factor-beta signaling across ages: from distorted lung development to chronic obstructive pulmonary disease. *Proc Am Thorac Soc* 2009; 6: 607-613.
- [37] Coalson JJ. Pathology of bronchopulmonary dysplasia. *Semin Perinatol* 2006; 30: 179-184.
- [38] Coalson JJ. Pathology of new bronchopulmonary dysplasia. *Semin Neonatol* 2003; 8: 73-81.
- [39] Hilgendorff A, Reiss I, Ehrhardt H, Eickelberg O, Alvira CM. Chronic lung disease in the preterm infant. Lessons learned from animal models. *Am J Respir Cell Mol Biol* 2014; 50: 233-245.
- [40] Laube M, Stolzing A, Thome UH, Fabian C. Therapeutic potential of mesenchymal stem cells for pulmonary complications associated with preterm birth. *Int J Biochem Cell Biol* 2016; 74: 18-32.

VEGF in bronchopulmonary dysplasia

- [41] Soliman N, Chaput K, Alshaikh B, Yusuf K. Pre-eclampsia and the risk of bronchopulmonary dysplasia in preterm infants less than 32 weeks' gestation. *Am J Perinatol* 2017; 34: 585-592.
- [42] Thebaud B, Ladha F, Michelakis ED, Sawicka M, Thurston G, Eaton F, Hashimoto K, Harry G, Haromy A, Korbitt G, Archer SL. Vascular endothelial growth factor gene therapy increases survival, promotes lung angiogenesis, and prevents alveolar damage in hyperoxia-induced lung injury: evidence that angiogenesis participates in alveolarization. *Circulation* 2005; 112: 2477-2486.
- [43] Thibeault DW, Mabry SM, Norberg M, Truog WE, Ekekezie II. Lung microvascular adaptation in infants with chronic lung disease. *Biol Neonate* 2004; 85: 273-282.
- [44] De Paepe ME, Mao Q, Powell J, Rubin SE, DeKoninck P, Appel N, Dixon M, Gundogan F. Growth of pulmonary microvasculature in ventilated preterm infants. *Am J Respir Crit Care Med* 2006; 173: 204-211.
- [45] Debette S, Visvikis-Siest S, Chen MH, Ndiaye NC, Song C, Destefano A, Safa R, Azimi NM, Sawyer D, Marteau JB, Xanthakis V, Siest G, Sullivan L, Pfister M, Smith H, Choi SH, Lamont J, Lind L, Yang Q, Fitzgerald P, Ingelsson E, Vasan RS, Seshadri S. Identification of cis- and trans-acting genetic variants explaining up to half the variation in circulating vascular endothelial growth factor levels. *Circ Res* 2011; 109: 554-563.
- [46] Ruggiero D, Dalmaso C, Nutile T, Sorice R, Dionisi L, Aversano M, Broet P, Leutenegger AL, Bourgain C, Ciullo M. Genetics of VEGF serum variation in human isolated populations of Cilento: importance of VEGF polymorphisms. *PLoS One* 2011; 6: e16982.
- [47] Ferrara N, Henzel WJ. Pituitary follicular cells secrete a novel heparin-binding growth factor specific for vascular endothelial cells. *Biochem Biophys Res Commun* 1989; 161: 851-858.
- [48] Vempati P, Popel AS, Mac GF. Extracellular regulation of VEGF: isoforms, proteolysis, and vascular patterning. *Cytokine Growth Factor Rev* 2014; 25: 1-19.
- [49] Crawford Y, Ferrara N. VEGF inhibition: insights from preclinical and clinical studies. *Cell Tissue Res* 2009; 335: 261-269.
- [50] Meller S, Bhandari V. VEGF levels in humans and animal models with RDS and BPD: temporal relationships. *Exp Lung Res* 2012; 38: 192-203.
- [51] Majumder S, Advani A. VEGF and the diabetic kidney: more than too much of a good thing. *J Diabetes Complications* 2017; 31: 273-279.
- [52] Robinson CJ, Stringer SE. The splice variants of vascular endothelial growth factor (VEGF) and their receptors. *J Cell Sci* 2001; 114: 853-865.
- [53] LeCouter J, Moritz DR, Li B, Phillips GL, Liang XH, Gerber HP, Hillan KJ, Ferrara N. Angiogenesis-independent endothelial protection of liver: role of VEGFR-1. *Science* 2003; 299: 890-893.
- [54] Kawamura H, Li X, Goishi K, van Meeteren LA, Jakobsson L, Cebe-Suarez S, Shimizu A, Edholm D, Ballmer-Hofer K, Kjellen L, Klagsbrun M, Claesson-Welsh L. Neuropilin-1 in regulation of VEGF-induced activation of p38MAPK and endothelial cell organization. *Blood* 2008; 112: 3638-3649.
- [55] Fantin A, Vieira JM, Plein A, Denti L, Fruttiger M, Pollard JW, Ruhrberg C. NRP1 acts cell autonomously in endothelium to promote tip cell function during sprouting angiogenesis. *Blood* 2013; 121: 2352-2362.
- [56] Chou HC, Li YT, Chen CM. Human mesenchymal stem cells attenuate experimental bronchopulmonary dysplasia induced by perinatal inflammation and hyperoxia. *Am J Transl Res* 2016; 8: 342-353.
- [57] Lassus P, Turanlahti M, Heikkila P, Andersson LC, Nupponen I, Sarnesto A, Andersson S. Pulmonary vascular endothelial growth factor and Flt-1 in fetuses, in acute and chronic lung disease, and in persistent pulmonary hypertension of the newborn. *Am J Respir Crit Care Med* 2001; 164: 1981-1987.
- [58] Voelkel NF, Vandivier RW, Tuder RM. Vascular endothelial growth factor in the lung. *Am J Physiol Lung Cell Mol Physiol* 2006; 290: L209-L221.
- [59] Jakkula M, Le Cras TD, Gebb S, Hirth KP, Tuder RM, Voelkel NF, Abman SH. Inhibition of angiogenesis decreases alveolarization in the developing rat lung. *Am J Physiol Lung Cell Mol Physiol* 2000; 279: L600-L607.
- [60] Le Cras TD, Markham NE, Tuder RM, Voelkel NF, Abman SH. Treatment of newborn rats with a VEGF receptor inhibitor causes pulmonary hypertension and abnormal lung structure. *Am J Physiol Lung Cell Mol Physiol* 2002; 283: L555-L562.
- [61] Kasahara Y, Tuder RM, Taraseviciene-Stewart L, Le Cras TD, Abman S, Hirth PK, Waltenberger J, Voelkel NF. Inhibition of VEGF receptors causes lung cell apoptosis and emphysema. *J Clin Invest* 2000; 106: 1311-1319.
- [62] Kasahara Y, Tuder RM, Cool CD, Lynch DA, Flores SC, Voelkel NF. Endothelial cell death and decreased expression of vascular endothelial growth factor and vascular endothelial growth factor receptor 2 in emphysema. *Am J Respir Crit Care Med* 2001; 163: 737-744.
- [63] Tang JR, Karumanchi SA, Seedorf G, Markham N, Abman SH. Excess soluble vascular endothelial growth factor receptor-1 in amniotic fluid impairs lung growth in rats: linking pre-

VEGF in bronchopulmonary dysplasia

- eclampsia with bronchopulmonary dysplasia. *Am J Physiol Lung Cell Mol Physiol* 2012; 302: L36-L46.
- [64] Yamamoto H, Yun EJ, Gerber HP, Ferrara N, Whittsett JA, Vu TH. Epithelial-vascular cross talk mediated by VEGF-A and HGF signaling directs primary septae formation during distal lung morphogenesis. *Dev Biol* 2007; 308: 44-53.
- [65] McGrath-Morrow SA, Cho C, Cho C, Zhen L, Hicklin DJ, Tudor RM. Vascular endothelial growth factor receptor 2 blockade disrupts postnatal lung development. *Am J Respir Cell Mol Biol* 2005; 32: 420-427.
- [66] Kumar VH, Lakshminrusimha S, Kishkurno S, Paturi BS, Gugino SF, Nielsen L, Wang H, Ryan RM. Neonatal hyperoxia increases airway reactivity and inflammation in adult mice. *Pediatr Pulmonol* 2016; 51: 1131-1141.
- [67] Tomanek RJ, Lund DD, Yue X. Hypoxic induction of myocardial vascularization during development. *Adv Exp Med Biol* 2003; 543: 139-149.
- [68] Vogel ER, Britt RJ, Trinidad MC, Faksh A, Martin RJ, MacFarlane PM, Pabelick CM, Prakash YS. Perinatal oxygen in the developing lung. *Can J Physiol Pharmacol* 2015; 93: 119-127.
- [69] Bhatt AJ, Amin SB, Chess PR, Watkins RH, Maniscalco WM. Expression of vascular endothelial growth factor and Flk-1 in developing and glucocorticoid-treated mouse lung. *Pediatr Res* 2000; 47: 606-613.
- [70] Thebaud B, Ladha F, Michelakis ED, Sawicka M, Thurston G, Eaton F, Hashimoto K, Harry G, Haromy A, Korbitt G, Archer SL. Vascular endothelial growth factor gene therapy increases survival, promotes lung angiogenesis, and prevents alveolar damage in hyperoxia-induced lung injury: evidence that angiogenesis participates in alveolarization. *Circulation* 2005; 112: 2477-2486.
- [71] Adams RH, Alitalo K. Molecular regulation of angiogenesis and lymphangiogenesis. *Nat Rev Mol Cell Biol* 2007; 8: 464-478.
- [72] Swift MR, Weinstein BM. Arterial-venous specification during development. *Circ Res* 2009; 104: 576-588.
- [73] Gridley T. Notch signaling in the vasculature. *Curr Top Dev Biol* 2010; 92: 277-309.
- [74] Eilken HM, Adams RH. Dynamics of endothelial cell behavior in sprouting angiogenesis. *Curr Opin Cell Biol* 2010; 22: 617-625.
- [75] Logsdon EA, Finley SD, Popel AS, Mac GF. A systems biology view of blood vessel growth and remodelling. *J Cell Mol Med* 2014; 18: 1491-1508.
- [76] Ribatti D, Nico B, Crivellato E. The role of pericytes in angiogenesis. *Int J Dev Biol* 2011; 55: 261-268.
- [77] Augustin HG, Koh GY, Thurston G, Alitalo K. Control of vascular morphogenesis and homeostasis through the angiopoietin-Tie system. *Nat Rev Mol Cell Biol* 2009; 10: 165-177.
- [78] Huang YS, Chang CW, Chen YM, Lee YH, Chen MC, Shih NL. Investigating expression profiles of VEGF-Flk, and Angpt1 during development of gas glands in Japanese eel (*Anguilla japonica*). *Comp Biochem Physiol A Mol Integr Physiol* 2010; 155: 350-360.
- [79] Taha D, Kirkby S, Nawab U, Dysart KC, Genen L, Greenspan JS, Aghai ZH. Early caffeine therapy for prevention of bronchopulmonary dysplasia in preterm infants. *J Matern Fetal Neonatal Med* 2014; 27: 1698-1702.
- [80] Ma L, Zhou P, Neu J, Lin HC. Potential nutrients for preventing or treating bronchopulmonary dysplasia. *Paediatr Respir Rev* 2017; 22: 83-88.
- [81] Mandell E, Seedorf G, Gien J, Abman SH. Vitamin D treatment improves survival and infant lung structure after intra-amniotic endotoxin exposure in rats: potential role for the prevention of bronchopulmonary dysplasia. *Am J Physiol Lung Cell Mol Physiol* 2014; 306: L420-L428.
- [82] Yeh TF, Chen CM, Wu SY, Husan Z, Li TC, Hsieh WS, Tsai CH, Lin HC. Intratracheal administration of budesonide/surfactant to prevent bronchopulmonary dysplasia. *Am J Respir Crit Care Med* 2016; 193: 86-95.
- [83] Aslam M, Baveja R, Liang OD, Fernandez-Gonzalez A, Lee C, Mitsialis SA, Kourembanas S. Bone marrow stromal cells attenuate lung injury in a murine model of neonatal chronic lung disease. *Am J Respir Crit Care Med* 2009; 180: 1122-1130.
- [84] Mobius MA, Rudiger M. Mesenchymal stromal cells in the development and therapy of bronchopulmonary dysplasia. *Mol Cell Pediatr* 2016; 3: 18.
- [85] Zhang ZH, Pan YY, Jing RS, Luan Y, Zhang L, Sun C, Kong F, Li KL, Wang YB. Protective effects of BMSCs in combination with erythropoietin in bronchopulmonary dysplasia-induced lung injury. *Mol Med Rep* 2016; 14: 1302-1308.
- [86] Baumgartner I, Pieczek A, Manor O, Blair R, Kearney M, Walsh K, Isner JM. Constitutive expression of phVEGF165 after intramuscular gene transfer promotes collateral vessel development in patients with critical limb ischemia. *Circulation* 1998; 97: 1114-1123.
- [87] Jabbarzadeh E, Starnes T, Khan YM, Jiang T, Wirtel AJ, Deng M, Lv Q, Nair LS, Doty SB, Laurencin CT. Induction of angiogenesis in tissue-engineered scaffolds designed for bone repair: a combined gene therapy-cell transplantation approach. *Proc Natl Acad Sci U S A* 2008; 105: 11099-11104.

VEGF in bronchopulmonary dysplasia

- [88] Le Cras TD, Markham NE, Tudor RM, Voelkel NF, Abman SH. Treatment of newborn rats with a VEGF receptor inhibitor causes pulmonary hypertension and abnormal lung structure. *Am J Physiol Lung Cell Mol Physiol* 2002; 283: L555-L562.
- [89] Heise RL, Link PA, Farkas L. From here to there, progenitor cells and stem cells are everywhere in lung vascular remodeling. *Front Pediatr* 2016; 4: 80.
- [90] Pierro M, Ciarmoli E, Thebaud B. Bronchopulmonary dysplasia and chronic lung disease: stem cell therapy. *Clin Perinatol* 2015; 42: 889-910.
- [91] van Haaften T, Byrne R, Bonnet S, Rochefort GY, Akabutu J, Bouchentouf M, Rey-Parra GJ, Galipeau J, Haromy A, Eaton F, Chen M, Hashimoto K, Abley D, Korbutt G, Archer SL, Thebaud B. Airway delivery of mesenchymal stem cells prevents arrested alveolar growth in neonatal lung injury in rats. *Am J Respir Crit Care Med* 2009; 180: 1131-1142.
- [92] Chang YS, Ahn SY, Yoo HS, Sung SI, Choi SJ, Oh WI, Park WS. Mesenchymal stem cells for bronchopulmonary dysplasia: phase 1 dose-escalation clinical trial. *J Pediatr* 2014; 164: 966-972.
- [93] Been JV, Debeer A, van Iwaarden JF, Kloosterboer N, Passos VL, Naulaers G, Zimmermann LJ. Early alterations of growth factor patterns in bronchoalveolar lavage fluid from preterm infants developing bronchopulmonary dysplasia. *Pediatr Res* 2010; 67: 83-89.
- [94] D'Angio CT, Maniscalco WM, Ryan RM, Avissar NE, Basavegowda K, Sinkin RA. Vascular endothelial growth factor in pulmonary lavage fluid from premature infants: effects of age and postnatal dexamethasone. *Biol Neonate* 1999; 76: 266-273.
- [95] Zeng X, Wert SE, Federici R, Peters KG, Whitsett JA. VEGF enhances pulmonary vasculogenesis and disrupts lung morphogenesis in vivo. *Dev Dyn* 1998; 211: 215-227.
- [96] Yi M, Masood A, Ziino A, Johnson BH, Belcastro R, Li J, Shek S, Kantores C, Jankov RP, Tanswell AK. Inhibition of apoptosis by 60% oxygen: a novel pathway contributing to lung injury in neonatal rats. *Am J Physiol Lung Cell Mol Physiol* 2011; 300: L319-L329.
- [97] Keenaghan M, Cai CL, Kumar D, Valencia GB, Rao M, Aranda JV, Beharry KD. Response of vascular endothelial growth factor and angiogenesis-related genes to stepwise increases in inspired oxygen in neonatal rat lungs. *Pediatr Res* 2013; 73: 630-638.
- [98] Gorenflo M, Vogel M, Obladen M. Pulmonary vascular changes in bronchopulmonary dysplasia: a clinicopathologic correlation in short- and long-term survivors. *Pediatr Pathol* 1991; 11: 851-866.
- [99] Cherukupalli K, Larson JE, Rotschild A, Thurlbeck WM. Biochemical, clinical, and morphologic studies on lungs of infants with bronchopulmonary dysplasia. *Pediatr Pulmonol* 1996; 22: 215-229.
- [100] Husain AN, Siddiqui NH, Stocker JT. Pathology of arrested acinar development in postsurfactant bronchopulmonary dysplasia. *Hum Pathol* 1998; 29: 710-717.
- [101] Coalson JJ, Winter VT, Siler-Khodr T, Yoder BA. Neonatal chronic lung disease in extremely immature baboons. *Am J Respir Crit Care Med* 1999; 160: 1333-1346.
- [102] Velten M, Heyob KM, Rogers LK, Welty SE. Deficits in lung alveolarization and function after systemic maternal inflammation and neonatal hyperoxia exposure. *J Appl Physiol* (1985) 2010; 108: 1347-1356.
- [103] O'Reilly M, Harding R, Sozo F. Altered small airways in aged mice following neonatal exposure to hyperoxic gas. *Neonatology* 2014; 105: 39-45.
- [104] Firsova AB, Cole TJ, Mollard R. Transient vascular and long-term alveolar deficits following a hyperoxic injury to neonatal mouse lung. *BMC Pulm Med* 2014; 14: 59.
- [105] Belcastro R, Lopez L, Li J, Masood A, Tanswell AK. Chronic lung injury in the neonatal rat: up-regulation of TGFbeta1 and nitration of IGF-R1 by peroxynitrite as likely contributors to impaired alveologenesis. *Free Radic Biol Med* 2015; 80: 1-11.
- [106] Jimenez J, Richter J, Nagatomo T, Salaets T, Quarck R, Wagennar A, Wang H, Vanoirbeek J, Deprest J, Toelen J. Progressive vascular functional and structural damage in a bronchopulmonary dysplasia model in preterm rabbits exposed to hyperoxia. *Int J Mol Sci* 2016; 17.
- [107] Tambunting F, Beharry KD, Waltzman J, Modanlou HD. Impaired lung vascular endothelial growth factor in extremely premature baboons developing bronchopulmonary dysplasia/chronic lung disease. *J Investig Med* 2005; 53: 253-262.
- [108] Grisafi D, Pozzobon M, Dedja A, Vanzo V, Tomanin R, Porzionato A, Macchi V, Salmaso R, Scarpa M, Cozzi E, Fassina A, Navaglia F, Maran C, Onisto M, Caenazzo L, De Coppi P, De Caro R, Chiandetti L, Zaramella P. Human amniotic fluid stem cells protect rat lungs exposed to moderate hyperoxia. *Pediatr Pulmonol* 2013; 48: 1070-1080.
- [109] Yang WC, Chen CY, Chou HC, Hsieh WS, Tsao PN. Angiogenic factors in cord blood of preterm infants predicts subsequently developing bronchopulmonary dysplasia. *Pediatr Neonatol* 2015; 56: 382-385.
- [110] Lajko M, Cardona HJ, Taylor JM, Shah RS, Farrow KN, Fawzi AA. Hyperoxia-induced proliferative retinopathy: early interruption of retinal

VEGF in bronchopulmonary dysplasia

- vascular development with severe and irreversible neurovascular disruption. *PLoS One* 2016; 11: e166886.
- [111] Jin M, Lee J, Lee KY, Jin Z, Pak JH, Kim HS. Alteration of TGF-beta-ALK-Smad signaling in hyperoxia-induced bronchopulmonary dysplasia model of newborn rats. *Exp Lung Res* 2016; 42: 354-364.
- [112] Procianoy RS, Hentges CR, Silveira RC. Vascular endothelial growth factor/placental growth factor heterodimer levels in preterm infants with bronchopulmonary dysplasia. *Am J Perinatol* 2016; 33: 480-485.