## Original Article

# Bioinformatics analysis of gene expression profile data to screen key genes involved in cardiac ischemia-reperfusion injury

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Abstract: In the present study, whole genome mRNA expression profiles in cardiac ischemia regions of rats with cardiac ischemia/reperfusion injury were analyzed to identify unique target genes. A total of 544 differentially expressed genes were identified between normal and cardiac ischemia samples. Our results identified ten key genes CXCR1, GLP1R, CCL21, GRK1, oxidative stress induced growth inhibitor 1 (OSGIN1), iroquois homeobox 1 (IRX1), opioid binding protein/cell adhesion molecule-like (OPCML), gi|672075288|ref|XR\_595614.1|, NON-RATT008949, XR\_601753.1, NONRATT008080 were related to cardiac ischemia/reperfusion injury. These candidate genes were further examined in a cardiac ischemia/reperfusion model so as to provide important therapeutic targets for the treatment of local cardiac injury.

**Keywords:** Cardiac ischemia/reperfusion injury, differentially expressed genes, functional enrichment analysis, pathway analysis

#### Introduction

There is a complex community of specific molecules in cardiac ischemia regions, and these important genes may profoundly influence many aspects of development for ischemia/reperfusion (I/R) injury. Cardiac I/R injury is an alarming global public health problem [1-3]. Accumulating evidence suggests that I/R injury can increase the risk of many other life-threatening diseases, including certain types of heart disease. Further, I/R-induced autophagy increases the severity of cardiomyocyte injury [4].

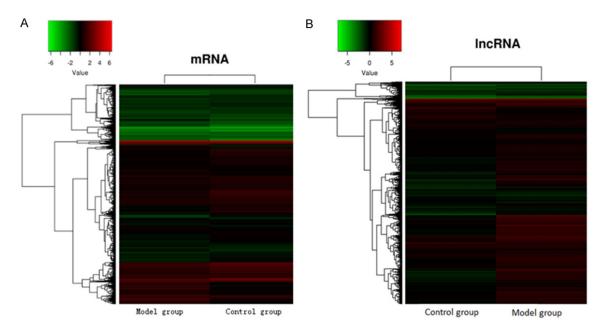
Advances in high-throughput RNA sequencing techniques have rapidly expanded our knowledge about many biological processes [5-10]. The aim of this study was to identify gene expression profile data for screening key genes

involved in cardiac I/R injury. This work helped us to understand the potential biological functions of mRNAs and IncRNAs and the pathophysiological mechanism involved in cardiac I/R injury.

#### Materials and methods

Animals and ethic statement

Male adult SD rats (250-300 g) were obtained from the Tongji Laboratory Animal Center. All rats were housed under a 12 h light/dark cycle with food and water provided *ad libitum*. The current study was performed in accordance with the guidelines of the National Institutes of Health. Animal care and experimental protocols were approved by the local Committee on Animal Care.



**Figure 1.** Heat maps and hierarchical clustering of expression ratios (log<sub>2</sub> scale) of mRNAs (A) and IncRNAs (B) in rat hearts 2 h after reperfusion. "Red" denotes high relative expression and "blue" denotes low relative expression. (For interpretation of the references to color in this figure legend, the reader is referred to the web version of this article).

Experimental protocol and myocardial ischemia/reperfusion injury model

SD rats were randomly divided into two groups: control group (Sham, n = 3) and I/R group (I/R, n = 3). Myocardial ischemia/reperfusion injury model was performed as previously described [11-14]. Briefly, rats were anesthetized and completely sedated, then tracheal intubation was set up. The left anterior descending coronary artery was then exposed and ligatured to form a reversible trap until myocardial ischemia occurred. After 30 min of myocardial ischemia, the trap was opened and myocardial reperfusion appeared. 2 h after reperfusion, the myocardial ischemic tissues were collected. For control group, animals, sutures were not tied undergoing a sham operation.

#### Tissue collection and RNA extraction

Following decapitation, the myocardial ischemic tissues were immediately dissected and washed thoroughly with sterile normal saline, and finally stored in liquid nitrogen for total RNA extraction. Total RNA was isolated using TRIzol (Invitrogen) according to manufacturer's instructions [6, 15-17]. RNA qualities were measured and RNA integrity was determined. The OD A260/A230 ratio was identified.

#### Microarray studies

Three myocardial tissues from each group were analyzed by IncRNAs + mRNAs Rat Gene Expression microarray (Agilent 8 × 60 K chips). After RNA quantities were measured, the samples were labeled and hybridized according to the manufacturer's guideline [18-20]. Significantly differentially expressed IncRNAs + mRNAs were then identified. Finally, hierarchical clustering was performed among the samples.

#### Bioinformatics analysis

For investigating the target genes of IncRNAs + mRNAs, some data were used. The IncRNAs and mRNAs expression profiles from the myocardial ischemic regions were screened by volcano plot filtering. The target genes were analyzed by GO (http://www.geneontology.org) and KEGG (http://www.genome.jp/dbget-bin).

#### Statistical analyses

Results are expressed as the mean  $\pm$  SEM. The statistical analyses and graphs were performed using GraphPad Prism (GraphPad Software, Inc., La Jolla, CA). P < 0.05 was considered statistically significant.

### Key genes involved in cardiac ischemia-reperfusion injury

**Table 1.** Detailed information of the top 50 up-regulated mRNAs in the ischemic region 2 h after reperfusion

Gene ID	Fold change (R/N)	GENE_SYMBOL	Gene name
NM_001134481	25.97118	Plcxd2	Phosphatidylinositol-specific phospholipase C, X domain containing 2
NM_001106165	25.46518	Car7	Carbonic anhydrase 7
XM_002727399	21.23528	Pilra	Paired immunoglobin-like type 2 receptor alpha
NM_001008339	21.07919	Nelfa	Negative elongation factor complex member A
XM_001081804	15.23514	Pde6g	Phosphodiesterase 6G, cGMP-specific, rod, gamma
XM_006249194	14.28584	RGD1561143	Similar to cell surface receptor FDFACT
NM_138504	13.01391	Osgin1	Oxidative stress induced growth inhibitor 1
NM 001107331	10.5432	Irx1	iroquois homeobox 1
_ NM_053848	10.10015	Opcml	Opioid binding protein/cell adhesion molecule-like
XM_006228512	8.814655	RGD1563034	Similar to ETS domain transcription factor ERF (Ets2 repressor factor)
NM_001109372	8.515041	Fam150b	Family with sequence similarity 150, member B
NM_001012224	8.093776	Nfe2	Nuclear factor, erythroid derived 2
NM_021688	8.069224	Kcnk1	Potassium channel, subfamily K, member 1
NM_020104	7.85615	Myl1	Myosin, light chain 1
XM 001074323	7.737585	Tnfsf18	Tumor necrosis factor (ligand) superfamily, member 18
NM_182952	7.403613	Cxcl11	Chemokine (C-X-C motif) ligand 11
NM_017019	7.245778	II1a	Interleukin 1 alpha
NM_134372	7.220482	Acmsd	Aminocarboxymuconate semialdehyde decarboxylase
XM_006226563	7.156018	Cdhr4	Cadherin-related family member 4
	6.972716	L0C690326	Hypothetical protein LOC690326
NM_001109578			Cysteine-rich, angiogenic inducer, 61
NM_031327	6.843192	Cyr61	
NM_001108195	6.718838	Klhl40	Kelch-like family member 40
NM_181479	6.621698	Kir3dl1	Killer cell immunoglobulin-like receptor, three domains, long cytoplasmic tail, 1
NM_001047866	6.594324	Cntnap5c	Contactin associated protein-like 5C
NM_001134481	6.215656	Plcxd2	Phosphatidylinositol-specific phospholipase C, X domain containing 2
NM_001106165	6.109064	Car7	Carbonic anhydrase 7
XM_002727399	5.92121	Pilra	Paired immunoglobin-like type 2 receptor alpha
NM_001008339	5.667461	Nelfa	Negative elongation factor complex member A
XM_001081804	5.640686	Pde6g	Phosphodiesterase 6G, cGMP-specific, rod, gamma
XM_006249194	5.442043	RGD1561143	Similar to cell surface receptor FDFACT
NM_138504	5.379033	Osgin1	Oxidative stress induced growth inhibitor 1
NM_001107331	5.352544	lrx1	iroquois homeobox 1
NM_053848	5.347504	Opcml	Opioid binding protein/cell adhesion molecule-like
XM_006228512	5.144021	RGD1563034	Similar to ETS domain transcription factor ERF (Ets2 repressor factor)
NM_001109372	5.078389	Fam150b	Family with sequence similarity 150, member B
NM_001012224	5.060686	Nfe2	Nuclear factor, erythroid derived 2
NM_021688	5.022726	Kcnk1	Potassium channel, subfamily K, member 1
NM_020104	4.914793	Myl1	Myosin, light chain 1
XM_001074323	4.900176	Tnfsf18	Tumor necrosis factor (ligand) superfamily, member 18
NM_182952	4.871627	Cxcl11	Chemokine (C-X-C motif) ligand 11
NM_017019	4.866426	II1a	Interleukin 1 alpha
NM_134372	4.727727	Acmsd	Aminocarboxymuconate semialdehyde decarboxylase
XM_006226563	4.721964	Cdhr4	Cadherin-related family member 4
NM_001109578	4.706878	L0C690326	Hypothetical protein LOC690326
NM_031327	4.699767	Cyr61	Cysteine-rich, angiogenic inducer, 61
NM_001108195	4.689268	Klhl40	Kelch-like family member 40
NM_181479	4.649013	Kir3dl1	$ \hbox{Killer cell immunoglobulin-like receptor, three domains, long cytoplasmic tail, 1} \\$
NM_001047866	4.638201	Cntnap5c	Contactin associated protein-like 5C
NM_001029927	4.525999	CpvI	Carboxypeptidase, vitellogenic-like
NM_017358	4.451633	Cdon	Cell adhesion associated, oncogene regulated

Values are fold changes (FC) in the reperfusion groups (reperfusion 2 h) over the control group (N > F2.0-fold; P < 0.05 by analysis of variance).

**Table 2.** Detailed information of the top 50 down-regulated mRNAs in the ischemic region 2 h after reperfusion

Gene ID	Fold change (R/N)	GENE_SYMBOL	Gene name
XM_001065496	-38.1959	Mmrn1	Multimerin 1
NM_031737	-33.0016	Nkx6-1	NK6 homeobox 1
XM_006223594	-29.4102	L0C688778	Similar to fatty aldehyde dehydrogenase-like
NM_001000011	-23.3124	Olr1428	Olfactory receptor 1428
NM_001008513	-22.6481	Ccl21	Chemokine (C-C motif) ligand 21
NM_001000856	-21.9947	0lr386	Olfactory receptor 386
NM_022544	-19.1504	Defb4	Defensin beta 4
NM_019310	-15.8884	Cxcr1	Chemokine (C-X-C motif) receptor 1
NM_012728	-14.7924	Glp1r	Glucagon-like peptide 1 receptor
NM_012535	-14.7229	Prl3b1	"Prolactin family 3, subfamily b, member 1"
NM_019329	-14.5408	Cntn3	Contactin 3 (plasmacytoma associated)
NM_001000693	-13.3859	Olr24	Olfactory receptor 24
NM_001106017	-13.1395	Myl7	Myosin, light chain 7, regulatory
NM_001134799	-13.0844	Tmem95	Transmembrane protein 95
NM_001109370	-11.8416	Dgat2I6	Diacylglycerol 0-acyltransferase 2-like 6
KM_001065496	-11.6631	Mmrn1	Multimerin 1
NM_031737	-11.5115	Nkx6-1	NK6 homeobox 1
KM_006223594	-11.3629	LOC688778	Similar to fatty aldehyde dehydrogenase-like
NM_00100011	-11.1664	Olr1428	Olfactory receptor 1428
NM_001008513	-11.023	Ccl21	Chemokine (C-C motif) ligand 21
_	-10.8804	0lr386	, , ,
NM_001000856			Olfactory receptor 386 Defensin beta 4
NM_022544	-10.5916	Defb4	
NM_019310	-10.3024	Cxcr1	Chemokine (C-X-C motif) receptor 1
NM_012728	-9.81794	Glp1r	Glucagon-like peptide 1 receptor
NM_012535	-9.79548	Prl3b1	Prolactin family 3, subfamily b, member 1
NM_019329	-9.69249	Cntn3	Contactin 3 (plasmacytoma associated)
VM_001109370	-9.24467	Dgat2l6	Diacylglycerol 0-acyltransferase 2-like 6
(M_001065496	-9.23634	Mmrn1	Multimerin 1
NM_031737	-9.13645	Nkx6-1	NK6 homeobox 1
(M_006223594	-9.11597	LOC688778	Similar to fatty aldehyde dehydrogenase-like
NM_001134799	-9.39616	Tmem95	Transmembrane protein 95
NM_001017501	-9.09525	Hormad2	HORMA domain containing 2
NM_001109520	-8.92488	Kcnk16	Potassium channel, subfamily K, member 16
NM_173333	-8.92021	Olr1361	Olfactory receptor 1361
NM_001105884	-8.89532	Hoxd1	Homeo box D1
KM_002726648	-8.74563	Pramef17	PRAME family member 17
NM_001000019	-8.73589	Olr1448	Olfactory receptor 1448
KM_002726602	-8.37797	Catsper4	Cation channel, sperm associated 4
NM_001134845	-8.26427	LOC688613	Hypothetical protein LOC688613
KM_003751638	-8.03944	LOC100912252	Uncharacterized LOC100912252
NM_001161846	-7.99213	Nipsnap3a	Nipsnap homolog 3A (C. elegans)
NM_022274	-7.92157	Birc5	Baculoviral IAP repeat-containing 5
NM_001271453	-7.89264	Gbx1	Gastrulation brain homeobox 1
KM_001081217	-7.84544	RGD1564031	Similar to transcription elongation factor B (SIII), polypeptide 2 $$
NM_013098	-7.82849	G6pc	Glucose-6-phosphatase, catalytic subunit
NM_176078	-7.78195	Clic6	Chloride intracellular channel 6
NM_001004079	-7.77497	Врі	Bactericidal/permeability-increasing protein
- NM_001000112	-7.76414	Olr5	Olfactory receptor 5
_ NM_198790	-7.76106	Rgsl2h	Regulator of G-protein signaling like 2 homolog (mouse)
_ NM_001105776	-7.71202	Foxi1	Forkhead box I1
NM_001109089	-7.69987	Msantd1	Myb/SANT-like DNA-binding domain containing 1

Values are fold changes (FC) in the reperfusion groups (reperfusion 2 h) over the control group (N > 2.0-fold; P < 0.05 by analysis of variance).

**Table 3.** Detailed information of the top 50 up-regulated IncRNAs in the ischemic regions 2 h after reperfusion

ProbeName	FC (abs)	Log <sub>2</sub> FC	Length (bp)
gi 672030539 ref XR_598272.1	11.25695	3.492744	823
gi 672022169 ref XR_601936.1	11.02217	3.462337	9453
gi 672075288 ref XR_595614.1	10.97383	3.455996	1023
NONRATT008949	10.93034	3.450266	593
gi 672021685 ref XR_601753.1	10.49752	3.391977	3534
NONRATTO08080	10.44272	3.384425	639
NONRATT016029	10.30865	3.365783	856
NONRATT016422	10.15852	3.344618	876
NONRATT028588	10.14481	3.34267	345
gi 672075969 ref XR_359418.2	9.935386	3.312576	2073
NONRATTO10658	9.540987	3.254139	737
NONRATTO13997	9.101868	3.186163	584
NONRATTO10660	9.099189	3.185738	739
gi 672029961 ref XR_341569.2	8.762763	3.131386	1557
NONRATTO23075	8.715472	3.123579	746
gi 672084663 ref XR_597212.1	8.47595	3.083375	344
NONRATTO09756	8.330487	3.058401	500
NONRATT017646	8.206457	3.036759	1112
NONRATT002658	7.978861	2.996183	720
NONRATT006492	7.693932	2.943721	345
gi 672017126 ref XR_600286.1	7.499788	2.90685	931
gi 672056839 ref XR_593095.1	7.364812	2.880649	4960
gi 672040804 ref XR_590723.1	7.343456	2.876459	1718
NONRATTO27869	7.312231	2.870312	617
NONRATTO10300	7.309803	2.869833	504
NONRATT019811	7.211921	2.850384	344
NONRATT027180	7.191667	2.846326	352
gi 672083774 ref XR_361470.2	7.186728	2.845335	473
gi 672029852 ref XR_598075.1	7.15444	2.838839	323
gi 672024868 ref XR_338678.2	7.136673	2.835252	2991
gi 672016311 ref XR_599992.1	6.644287	2.732114	374
NONRATT025404	6.484052	2.696896	420
NONRATT025357	6.268275	2.648069	266
gi 564335313 ref XR_351412.1	6.241109	2.641802	589
NONRATT012880	6.239899	2.641523	322
NONRATT024390	6.190577	2.630074	385
NONRATT013400	6.115241	2.612409	373
NONRATT006836	6.071511	2.602056	772
gi 672028592 ref XR_340666.2	5.80575	2.537482	1887
NONRATT014969	5.718843	2.515723	315
NONRATT001619	5.707868	2.512952	1341
gi 672029992 ref XR_598146.1	5.476558	2.453269	1793
gi 672032741 ref XR_589772.1	5.468528	2.451153	517
NONRATT013679	5.439823	2.44356	1082
NONRATT030203	5.389799	2.430231	1073
uc.284	5.379776	2.427546	209
NONRATT001867	5.283186	2.401408	323

#### Results

Hierarchical clustering analysis of mRNAs and IncRNAs

After extracting the expression values of the differentially expressed genes, hierarchical clustering analysis was conducted for the mRNAs and IncRNAs 2 h after I/R-induced cardiac injury. As shown in the heat maps and hierarchical clustering (Figure 1), the differentially expressed genes could clearly distinguish the myocardial ischemic samples from the normal myocardial samples. "Red" denotes high relative expression and "blue" denotes low relative expression. In myocardial ischemic samples, there were more down-regulated genes than up-regulated genes (Figure 1).

The up-regulated mRNAs in the myocardial ischemic regions 2 h after I/R-induced cardiac injury

mRNA expression was quantified using RPKM value to identify the gene expression changes in T1-T4 spinal cord upon I/R-induced cardiac injury. The top 50 up-regulated mRNAs are listed in Table 1. Our results show that the upregulated mRNAs in the myocardial ischemic regions include phosphatidylinositol-specific phospholipase C, X domain containing 2 (PLCXD2), carbonic anhydrase 7 (CAR7), paired immunoglobin-like type 2 receptor alpha (PILRA), negative elongation factor complex member A (Nelfa), oxidative stress induced growth inhibitor 1 (OSGIN1), iroquois homeobox 1 (IRX1), and opioid binding protein/ cell adhesion molecule-like (OPCML).

gi 672014436 ref XR_343345.2	5.281367	2.400912	1039
NONRATTO12717	5.280564	2.400692	426
NONRATT017465	5.255965	2.393956	292
gi 672030539 ref XR_598272.1	11.25695	3.492744	823
gi 672022169 ref XR_601936.1	11.02217	3.462337	9453

Values are fold changes (FC) in the reperfusion groups (reperfusion 2 h) over the control group (N > 2.0-fold; P < 0.05 by analysis of variance).

**Table 4.** Detailed information of the top 50 down-regulated IncRNAs in the ischemic regions 2 h after reperfusion

ProbeName	FC (abs)	Log <sub>2</sub> FC	Length (bp)
NONRATT004082	-23.196	-4.53581	470
NONRATT000894	-23.0546	-4.52698	540
NONRATT028709	-22.9795	-4.52228	995
NONRATT015658	-21.8799	-4.45153	755
gi 672070370 ref XR_358071.2	-21.5694	-4.43092	498
NONRATT001791	-21.1837	-4.40488	735
NONRATT023521	-21.1414	-4.4020	833
gi 672082871 ref XR_596918.1	-20.6647	-4.3691	1655
NONRATT014802	-20.3956	-4.35019	1014
NONRATT008376	-20.178	-4.33471	858
NONRATT028780	-20.0827	-4.32788	390
uc.367	-19.8568	-4.31156	298
NONRATT030839	-19.6054	-4.29318	305
NONRATT018033	-19.5257	-4.2873	840
NONRATT014624	-19.5141	-4.28644	744
NONRATT013112	-19.0972	-4.25529	438
NONRATT014824	-19.0196	-4.24942	579
NONRATT014480	-18.956	-4.24459	300
NONRATT010526	-18.6505	-4.22114	744
NONRATT009061	-18.6214	-4.21889	545
NONRATT013001	-18.4618	-4.20647	528
NONRATT008744	-18.3621	-4.19866	477
uc.437	-18.3618	-4.19864	215
NONRATT005800	-18.2419	-4.18918	720
NONRATT010464	-18.2309	-4.18831	288
gi 672033376 ref XR_589489.1	-18.1298	-4.1803	778
NONRATT010825	-17.9918	-4.16927	891
gi 672089372 ref XR_597918.1	-17.8159	-4.15509	506
gi 672031982 ref XR_589619.1	-17.6334	-4.14024	4919
NONRATT022292	-17.5927	-4.13691	387
NONRATT023015	-17.5174	-4.13072	418
NONRATT007424	-17.4914	-4.12857	251
NONRATT027491	-17.3819	-4.11952	1137
NONRATT024410	-17.2341	-4.1072	371
NONRATT005822	-17.2261	-4.10652	923
gi 672023024 ref XR_348601.2	-17.0555	-4.09216	1125

Expression profiling of downregulated mRNAs in the myocardial ischemic regions 2 h after I/R-induced cardiac injury

Expression profiling of downregulated mRNAs was identified in myocardial ischemic samples relative to control samples. The top 50 downregulated mRNAs are listed in Table 2. Our results showed that the down-regulated mRNAs in the myocardial ischemic regions included multimerin 1 (MMRN1), NK6 homeobox 1 (NKX6-1), olfactory receptor 1428 (OLR1428), chemokine (C-C motif) ligand 21 (CCL21), olfactory receptor 386 (OLR386), defensin beta 4 (DEFB4), chemokine (C-X-C motif) receptor 1 (CXCR1), and glucagon-like peptide 1 receptor (GLP1R).

Expression profiling of upregulated IncRNAs in the myocardial ischemic regions 2 h after I/R-induced cardiac injury

High-throughput RNA sequencing technique for the expression profiling of up-regulated IncRNAs from rats with I/R-induced cardiac injury was analyzed to acquire key Inc-RNAs associated with I/R. The top 50 up-regulated IncRNAs are listed in Table 3. Our results showed that the upregulated IncRNAs in the myocardial ischemic regions mainly included XR\_598272.1, XR\_601936.1, NONRATT016-029, gi|672075288|ref|XR\_ 595614.1|, NONRATT0089-49, XR\_601753.1, NONRATT-008080, NONRATT016422, NONRATT028588.

uc.391	-16.9984	-4.08733	311
NONRATT003963	-16.9707	-4.08498	352
NONRATTO10147	-16.9354	-4.08197	565
NONRATT009885	-16.8779	-4.07706	345
NONRATT013237	-16.668	-4.05901	693
NONRATT030311	-16.6182	-4.0547	948
NONRATT013031	-16.3546	-4.03162	416
gi 672034555 ref XR_350029.2	-16.3519	-4.03138	2403
NONRATT007621	-16.3285	-4.02932	725
NONRATT007868	-16.2052	-4.01839	896
NONRATT001032	-16.1868	-4.01675	721
NONRATT022178	-16.154	-4.01382	368
NONRATT004973	-16.1106	-4.00994	630
NONRATT006935	-16.1068	-4.00959	334
NONRATT004082	-23.196	-4.53581	470
NONRATT000894	-23.0546	-4.52698	540

Values are fold changes (FC) in the reperfusion groups (reperfusion 2 h) over the control group (N > 2.0-fold; P < 0.05 by analysis of variance).

Expression profiling of down-regulated IncRNAs in the myocardial ischemic regions 2 h after I/R-induced cardiac injury

Expression profiling of down-regulated IncRNAs from rats with I/R-induced cardiac injury was analyzed by high-throughput RNA sequencing techniques to acquire key IncRNAs associated with I/R. The top 50 down-regulated IncRNAs are listed in **Table 4**. Our results showed that the down-regulated IncRNAs in the myocardial ischemic regions mainly included NONRATT004082, NONRATT000894, NONRATT028709, NONRATT015658, gi|67207037-0|ref|XR\_358071.2|, NONRATT001791, NONRATT023521, gi|672082871|ref|XR\_59691-8.1|, NONRATT014802, NONRATT008376.

Gene ontology and KEGG Pathway enrichment analysis

The top six enriched GO biological processes (**Table 5**) in the I/R group mainly included neurological system process (*P*-Value = 3.94E-13), sensory perception (*P*-Value = 5.80E-13), system process (*P*-Value = 5.20E-12), G-protein coupled receptor signaling pathway (*P*-Value = 5.62E-12), cell surface receptor signaling pathway (*P*-Value = 2.79E-11), and detection of stimulus (*P*-Value = 1.25E-10). The top six enriched GO molecular function (**Table 6**) in the I/R group included G-protein coupled receptor activity (*P*-Value = 4.45e-12), signaling receptor

activity (P-Value = 5.17e-12), transmembrane signaling receptor activity (P-Value = 1.91e-11), receptor activity (P-Value = 1.04e-10), signal transducer activity (P-Value = 1.50e-10), and molecular transducer activity (P-Value = 1.93e-09). The top six enriched GO cellular components (Table 7) in the I/R group included integral component of membrane (P-Value = 2.02e-08), cell periphery (P-Value = 1.23e-07), plasma membrane (P-Value = 1.74e-07), extracellular space (P-Value = 2.51e-07), membrane part (P-Value = 1.80e-05), and intrinsic component of plasma membrane (P-Value =

2.27e-05). The top five enriched KEGG pathways (**Table 8**) of STRING database in the I/R group mainly included olfactory transduction (*P*-Value = 7.03e-11), neuroactive ligand-receptor interaction (*P*-Value = 0.0002), cytokine-cytokine receptor interaction (*P*-Value = 0.0009), and steroid hormone biosynthesis (*P*-Value = 0.0015), retinol metabolism (*P*-Value = 0.0015). The top five enriched KEGG pathways (**Table 9**) of PANTHER database in the I/R group included Nicotine pharmacodynamics pathway (*P*-Value = 0.042), and heterotrimeric G-protein signaling pathway-rod outer segment phototransduction (*P*-Value = 0.045).

#### Discussion

In this study, chip data for the myocardial tissue samples from rats with I/R-induced cardiac injury was analysis. We found that the top 50 up-regulated mRNAs/IncRNAs and top 50 down-regulated mRNAs/IncRNAs were identified in I/R cardiac samples relative to normal cardiac samples. Among which, ten key genes CXCR1, GLP1R, CCL21, GRK1, oxidative stress induced growth inhibitor 1 (OSGIN1), iroquoishomeobox 1 (IRX1), opioid binding protein/cell adhesion molecule-like (OPCML), gi|67-2075288|ref|XR\_595614.1|, NONRATT0089-49, XR\_601753.1, and NONRATT008080 were related to cardiac ischemia/reperfusion injury.

**Table 5.** Biological process analyses of the Gene Ontology (GO) terms enriched for the differentially expressed genes (DEGs) involved in the protein-protein interaction (PPI) network constructed by STRING database

Term		Gene number	P-value
G0:0050877	Neurological system process	272	3.94e-13
GO:0007600	Sensory perception	235	5.80e-13
G0:0003008	System process	321	5.20e-12
GO:0007186	G-protein coupled receptor signaling pathway	277	5.62e-12
GO:0007166	Cell surface receptor signaling pathway	431	2.79e-11
GO:0051606	Detection of stimulus	194	1.25e-10
GO:0050906	Detection of stimulus involved in sensory perception	182	2.59e-10
G0:0007606	Sensory perception of chemical stimulus	181	2.19e-09
GO:0050907	Detection of chemical stimulus involved in sensory perception	171	3.75e-09
G0:0007608	Sensory perception of smell	169	7.47e-09
GO:0050911	Detection of chemical stimulus involved in sensory perception of smell	165	8.26e-09
G0:0009593	Detection of chemical stimulus	173	8.93e-09
G0:0032501	Multicellular organismal process	691	2.88e-08
GO:0044707	Single-multicellular organism process	669	4.86e-08
G0:0042221	Response to chemical	458	4.97e-07
GO:0044700	Single organism signaling	576	2.66e-06
G0:0023052	Signaling	576	2.71e-06
GO:0009605	Response to external stimulus	219	2.91e-06
GO:0007165	Signal transduction	540	3.66e-06
G0:0007154	Cell communication	584	4.62e-06
G0:0050877	Neurological system process	272	3.94e-13

**Table 6.** Molecular function analyses of the Gene Ontology (GO) terms enriched for the differentially expressed genes (DEGs) involved in the protein-protein interaction (PPI) network constructed by STRING database

Term		Gene number	P-value
G0:0004930	G-protein coupled receptor activity	245	4.45e-12
G0:0038023	Signaling receptor activity	283	5.17e-12
G0:0004888	Transmembrane signaling receptor activity	270	1.91e-11
G0:0004872	Receptor activity	291	1.04e-10
G0:0004871	Signal transducer activity	301	1.50e-10
G0:0060089	Molecular transducer activity	309	1.93e-09
G0:0004984	Olfactory receptor activity	165	8.26e-09
G0:0046873	Metal ion transmembrane transporter activity	59	2.86e-05
G0:0005125	Cytokine activity	33	5.10e-05
G0:0042379	Chemokine receptor binding	15	7.88e-05
GO:0005261	Cation channel activity	45	8.35e-05
G0:0008009	Chemokine activity	13	0.0001
G0:0005216	lon channel activity	54	0.0001
G0:0022843	Voltage-gated cation channel activity	27	0.0001
G0:0022838	Substrate-specific channel activity	55	0.0001
G0:0022832	Voltage-gated channel activity	32	0.0002
G0:0005244	Voltage-gated ion channel activity	32	0.0002
G0:0022836	Gated channel activity	45	0.0003
G0:0022803	Passive transmembrane transporter activity	56	0.0003
G0:0015267	Channel activity	56	0.0003

Previous studies showed that the inflammatory response was likely to be the main factor for I/R-induced cardiac injury [21]. It is well-known that chemokine family was related to inflammatory response. A study of Zouggari et al. [22] identified a crucial interaction between mature B lymphocytes and monocytes after acute myocardial ischemia, and showed that high circulating concentrations of CCL7 and BAFF in patients with acute myocardial infarction predicted increased risk

**Table 7.** Cellular component analyses of the Gene Ontology (GO) terms enriched for the differentially expressed genes (DEGs) involved in the protein-protein interaction (PPI) network constructed by STRING database

Term		Gene number	<i>P</i> -value
G0:0031224	Intrinsic component of membrane	549	3.50e-09
G0:0016021	Integral component of membrane	533	2.02e-08
G0:0071944	Cell periphery	494	1.23e-07
G0:0005886	Plasma membrane	484	1.74e-07
G0:0005615	Extracellular space	147	2.51e-07
G0:0044425	Membrane part	607	1.80e-05
G0:0031226	Intrinsic component of plasma membrane	100	2.27e-05
G0:0005887	Integral component of plasma membrane	92	6.22e-05
G0:0034702	Ion channel complex	39	0.0006
G0:1902495	Transmembrane transporter complex	40	0.0018
G0:0034703	Cation channel complex	24	0.0019
G0:1990351	Transporter complex	40	0.002
G0:0032281	Alpha-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid selective glutamate receptor complex	9	0.002
G0:0045095	Keratin filament	14	0.002
G0:0005882	Intermediate filament	20	0.003
G0:0009986	Cell surface	78	0.003
G0:0005891	Voltage-gated calcium channel complex	8	0.004
GO:0044459	Plasma membrane part	181	0.004
G0:0031225	Anchored component of membrane	15	0.011
G0:0043235	Receptor complex	35	0.015

**Table 8.** The Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways enriched for the differentially expressed genes (DEGs) involved in the protein-protein interaction (PPI) network constructed by STRING database

Term		Gene number	<i>P</i> -value
rno04740	Olfactory transduction	194	7.03e-11
rno04080	Neuroactive ligand-receptor interaction	49	0.0002
rno04060	Cytokine-cytokine receptor interaction	37	0.0009
rno00140	Steroid hormone biosynthesis	18	0.0015
rno00830	Retinol metabolism	18	0.0015
rno05321	Inflammatory bowel disease (IBD)	15	0.0021
rno05323	Rheumatoid arthritis	17	0.007
rno04913	Ovarian steroidogenesis	12	0.012
rno04514	Cell adhesion molecules (CAMs)	26	0.018
rno04010	MAPK signaling pathway	36	0.02
rno05150	Staphylococcus aureus infection	11	0.02
rno05204	Chemical carcinogenesis	15	0.027
rno05322	Systemic lupus erythematosus	20	0.030
rno05310	Asthma	7	0.035
rno05332	Graft-versus-host disease	12	0.036
rno04940	Type I diabetes mellitus	13	0.040
rno05410	Hypertrophic cardiomyopathy (HCM)	14	0.043
rno04668	TNF signaling pathway	17	0.044
rno00270	Cysteine and methionine metabolism	8	0.045
rno05030	Cocaine addiction	9	0.048

of death or recurrent myocardial infarction, suggesting CCL7 may be new therapeutic targets for acute myocardial infarction [23]. By DNA microarray data for four acute coronary syndrome patients' samples and four normal samples, Zhang et al. [24] reported that ten upregulated genes belonging to chemokine family (CCL2, CCR1, CXCL3, CXCL2, CCL8, CXCL11, CCL7, IL-10, CCL22 and CC-L20) were related to inflammatory response, speculating that these ge-nes might be related to acute coronary syndrome. Kimura et al. indicated that

**Table 9.** The Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways enriched for the differentially expressed genes (DEGs) involved in the protein-protein interaction (PPI) network constructed by PANTHER database

Term	Gene number	P-value	Gene symbol
P06587: Nicotine pharmacodynamics pathway	4	0.042	rno:54239, Chrnb2, 54239:down
			rno:24318, Drd2, 24318:down
			rno:304081, Clic6, 304081:down
			rno:29238, Drd3, 29238:down
P00028: Heterotrimeric G-protein signaling pathway-rod outer segment phototransduction	5	0.045	rno:245986, Gng8, 245986:down
			rno:25539, Sag, 25539:down
			rno:363143, Gnat1, 363143:down
			rno:81760, Grk1, 81760:down
			rno:25343, Pdc, 25343:down

macrophage-derived chemokine (CCL22) was involved centrally in the development of human atherosclerotic lesions and was a characteristic of the monocytes/macrophages migrating into atherosclerotic lesions [25].

The results from function and pathway analysis indicated that the differentially expressed genes were most significantly associated with inflammation mediated by chemokine and cytokine signaling pathway, e.g. CXCR1, GLP1R, CCL21, GRK1. Opfermann et al. [26] reported a pilot study on a CXCR1/2 antagonist reparixin to assess safety and efficacy in attenuating I/R injury and inflammation after on-pump coronary artery bypass graft surgery, and fou-nd that administration of reparixin in CABG patients appeared to be feasible and safe. Leonard et al. [27] indicated that the expression of the interleukin-8 receptors CXCR1 and CXCR2 in peripheral blood cells were increased in obstructive coronary artery disease and decreased in patients with improved perfusion, suggesting that these genes may serve as markers of disease severity and progression. Xu et al. showed that overexpression of CXCR1/CXCR2 on mesenchymal stromal cells might be an effective treatment for acute myocardial infarction [28]. A study of Basalay et al. suggested that glucagon-like peptide-1 (GLP1) mediated cardioprotection by remote ischemic conditioning [29]. DeNicola et al. demonstrated that stimulation of glucagonlike peptide-1 receptor through exendin-4 preserved myocardial performance and prevented cardiac remodeling in infarcted myocardium, suggesting that GLP-1R serves as a novel approach to eliciting cardioprotection and mitigating oxidative stress-induced injury. Cervia et al. [30] reported the modulation of the neuronal response to ischemia by somatostatin analogues acting at G protein-coupled receptor kinase 1 (GRK1). Nihei et al. examined whether Rho-kinase activity in circulating leukocytes, and found that Rho-kinase activity exhibited distinct circadian variation associated with alterations in coronary vasomotor responses and autonomic activity in VSA patients [31].

Overall, the gene expression profiles of cardiac ischemia/reperfusion injury were provided and differentially expressed genes were screened out. These results will be beneficial for understanding the mechanism and potential drug targets of cardiac ischemia/reperfusion injury.

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#### Disclosure of conflict of interest

None.

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#### References

- [1] Bei Y, Xu T, Lv D, Yu P, Xu J, Che L, Das A, Tigges J, Toxavidis V, Ghiran I, Shah R, Li Y, Zhang Y, Das S and Xiao J. Exercise-induced circulating extracellular vesicles protect against cardiac ischemia-reperfusion injury. Basic Res Cardiol 2017; 112: 38.
- [2] Xu AJ, Song ZP, Peng YW and Xiang HB. Change of mitochondrial function in the early stage after cardiac ischemia-reperfusion injury in mice. Int J Clin Exp Med 2016; 9: 2549-2554.
- [3] Murphy E and Steenbergen C. Mechanisms underlying acute protection from cardiac ischemia-reperfusion injury. Physiol Rev 2008; 88: 581-609.
- [4] Huang Z, Han Z, Ye B, Dai Z, Shan P, Lu Z, Dai K, Wang C and Huang W. Berberine alleviates cardiac ischemia/reperfusion injury by inhibiting excessive autophagy in cardiomyocytes. Eur J Pharmacol 2015; 762: 1-10.
- [5] Liu QQ, Liu H, He ZG, Zhang SJ, Liu BW, Wang L, Qiu WH, Xu Q, Xiang HB and Lv YM. Differential gene and IncRNA expression in the lower thoracic spinal cord following ischemia/reperfusion-induced acute kidney injury in rats. Oncotarget 2017; 8: 53465-53481.
- [6] Liu BW, Li ZX, He ZG, Liu C, Xiong J and Xiang HB. Altered expression of target genes of spinal cord in different itch models compared with capsaicin assessed by RT-qPCR validation. Oncotarget 2017; 8: 74423-74433.
- [7] Wang S, Xu H, Zou L, Xie J, Wu H, Wu B, Yi Z, Lv Q, Zhang X, Ying M, Liu S, Li G, Gao Y, Xu C, Zhang C, Xue Y and Liang S. LncRNA uc.48+ is involved in diabetic neuropathic pain mediated by the P2X receptor in the dorsal root ganglia. Purinergic Signal 2016; 12: 139-48.
- [8] Boon RA, Jae N, Holdt L and Dimmeler S. Long noncoding RNAs: from clinical genetics to therapeutic targets? J Am Coll Cardiol 2016; 67: 1214-1226.
- [9] Zhao X, Tang Z, Zhang H, Atianjoh FE, Zhao JY, Liang L, Wang W, Guan X, Kao SC, Tiwari V, Gao YJ, Hoffman PN, Cui H, Li M, Dong X and Tao YX. A long noncoding RNA contributes to neuropathic pain by silencing Kcna2 in primary afferent neurons. Nat Neurosci 2013; 16: 1024-1031.
- [10] Jiang BC, Sun WX, He LN, Cao DL, Zhang ZJ and Gao YJ. Identification of IncRNA expression profile in the spinal cord of mice following spinal nerve ligation-induced neuropathic pain. Mol Pain 2015; 11: 43.
- [11] Murry CE, Jennings RB and Reimer KA. Preconditioning with ischemia: a delay of lethal cell

- injury in ischemic myocardium. Circulation 1986; 74: 1124-1136.
- [12] Huang CH, Lai CC, Yang AH and Chiang SC. Myocardial preconditioning reduces kidney injury and apoptosis induced by myocardial ischaemia and reperfusion. Eur J Cardiothorac Surg 2015; 48: 382-391.
- [13] Borst O, Ochmann C, Schonberger T, Jacoby C, Stellos K, Seizer P, Flogel U, Lang F and Gawaz M. Methods employed for induction and analysis of experimental myocardial infarction in mice. Cell Physiol Biochem 2011; 28: 1-12.
- [14] Pisarenko OI, Shulzhenko VS, Studneva IM, Serebryakova LI, Pelogeykina YA and Veselova OM. Signaling pathways of a structural analogue of apelin-12 involved in myocardial protection against ischemia/reperfusion injury. Peptides 2015; 73: 67-76.
- [15] Liu T, He Z, Tian X, Kamal GM, Li Z, Liu Z, Liu H, Xu F, Wang J and Xiang H. Specific patterns of spinal metabolites underlying alpha-Me-5-HTevoked pruritus compared with histamine and capsaicin assessed by proton nuclear magnetic resonance spectroscopy. Biochim Biophys Acta 2017; 1863: 1222-1230.
- [16] He ZG, Liu BW, Li ZX, Liu C and Xiang HB. Altered expression profiling of spinal genes modulated by compound 48/80 in a mouse itch model. J Anesth Perioper Med 2017; 4: 220-224.
- [17] Liu TT, Liu BW, He ZG, Feng L, Liu SG and Xiang HB. Delineation of the central melanocortin circuitry controlling the kidneys by a virally mediated transsynaptic tracing study in transgenic mouse model. Oncotarget 2016; 7: 69256-69266.
- [18] Orom UA, Derrien T, Beringer M, Gumireddy K, Gardini A, Bussotti G, Lai F, Zytnicki M, Notredame C, Huang Q, Guigo R and Shiekhattar R. Long noncoding RNAs with enhancer-like function in human cells. Cell 2010; 143: 46-58.
- [19] Patterson TA, Lobenhofer EK, Fulmer-Smentek SB, Collins PJ, Chu TM, Bao W, Fang H, Kawasaki ES, Hager J, Tikhonova IR, Walker SJ, Zhang L, Hurban P, de Longueville F, Fuscoe JC, Tong W, Shi L and Wolfinger RD. Performance comparison of one-color and two-color platforms within the MicroArray quality control (MAQC) project. Nat Biotechnol 2006; 24: 1140-1150.
- [20] Bottomly D, Walter NA, Hunter JE, Darakjian P, Kawane S, Buck KJ, Searles RP, Mooney M, McWeeney SK and Hitzemann R. Evaluating gene expression in C57BL/6J and DBA/2J mouse striatum using RNA-Seq and microarrays. PLoS One 2011; 6: e17820.
- [21] Chandran S, Watkins J, Abdul-Aziz A, Shafat M, Calvert PA, Bowles KM, Flather MD, Rushworth SA and Ryding AD. Inflammatory differences in

- plaque erosion and rupture in patients with STsegment elevation myocardial infarction. J Am Heart Assoc 2017; 6.
- [22] Zouggari Y, Ait-Oufella H, Bonnin P, Simon T, Sage AP, Guerin C, Vilar J, Caligiuri G, Tsiantoulas D, Laurans L, Dumeau E, Kotti S, Bruneval P, Charo IF, Binder CJ, Danchin N, Tedgui A, Tedder TF, Silvestre JS and Mallat Z. B lymphocytes trigger monocyte mobilization and impair heart function after acute myocardial infarction. Nat Med 2013; 19: 1273-1280.
- [23] Kim ND and Luster AD. To B or not to B-that is the question for myocardial infarction. Nat Med 2013; 19: 1208-1210.
- [24] Zhang L, Li J, Liang A, Liu Y, Deng B and Wang H. Immune-related chemotactic factors were found in acute coronary syndromes by bioinformatics. Mol Biol Rep 2014; 41: 4389-4395.
- [25] Kimura S, Tanimoto A, Wang KY, Shimajiri S, Guo X, Tasaki T, Yamada S and Sasaguri Y. Expression of macrophage-derived chemokine (CCL22) in atherosclerosis and regulation by histamine via the H2 receptor. Pathol Int 2012; 62: 675-683.
- [26] Opfermann P, Derhaschnig U, Felli A, Wenisch J, Santer D, Zuckermann A, Dworschak M, Jilma B and Steinlechner B. A pilot study on reparixin, a CXCR1/2 antagonist, to assess safety and efficacy in attenuating ischaemiareperfusion injury and inflammation after onpump coronary artery bypass graft surgery. Clin Exp Immunol 2015; 180: 131-142.

- [27] Leonard DA, Merhige ME, Williams BA and Greene RS. Elevated expression of the interleukin-8 receptors CXCR1 and CXCR2 in peripheral blood cells in obstructive coronary artery disease. Coron Artery Dis 2011; 22: 491-496.
- [28] Xu J, Chen Q, Shi C and Yin Z. Overexpression of CXCR1/CXCR2 on mesenchymal stromal cells may be an effective treatment for acute myocardial infarction. Cytotherapy 2009; 11: 990-991.
- [29] Basalay MV, Mastitskaya S, Mrochek A, Ackland GL, Del Arroyo AG, Sanchez J, Sjoquist PO, Pernow J, Gourine AV and Gourine A. Glucagonlike peptide-1 (GLP-1) mediates cardioprotection by remote ischaemic conditioning. Cardiovasc Res 2016: 112: 669-676.
- [30] Cervia D, Martini D, Ristori C, Catalani E, Timperio AM, Bagnoli P and Casini G. Modulation of the neuronal response to ischaemia by somatostatin analogues in wild-type and knockout mouse retinas. J Neurochem 2008; 106: 2224-2235.
- [31] Nihei T, Takahashi J, Tsuburaya R, Ito Y, Shiroto T, Hao K, Takagi Y, Matsumoto Y, Nakayama M, Miyata S, Sakata Y, Ito K and Shimokawa H. Circadian variation of Rho-kinase activity in circulating leukocytes of patients with vasospastic angina. Circ J 2014; 78: 1183-1190.