Original Article

Circulating levels of adiponectin and leptin in patients with prostate cancer

Ligang Zhang^{1*}, Meng Zhang^{1,2*}, Jun Zhou^{1,2}, Guangyue Luo^{1,2}, Xianguo Chen^{1,2}, Li Zhang^{1,2}, Chaozhao Liang^{1,2}

¹Department of Urology, The First Affiliated Hospital of Anhui Medical University, Hefei 230022, China; ²Institute of Urology, Anhui Medical University, Hefei 230022, China. *Equal contributors.

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Abstract: Prostate cancer (PCa) is one of the major health care problems in males. It is known that red and processed meat consumption, fat intake and obesity are risk factors for PCa development. Adiponectin and leptin are adipokines that are synthesized in visceral adipose tissue and associated with obesity. Up to date, the association of serum adiponectin and leptin with PCa largely remains unexplored. Therefore, we studied the concentration of adiponectin and leptin in PCa patients in Chinese population. 92 prospective cases of prostate cancer and 92 matched healthy controls were enrolled in this study. Serum adiponectin and leptin levels were detected by enzymelinked immunosorbent assays (ELISA) technique. No statistically significant differences were observed in age, bodymass index (BMI), prostate specific antigen (PSA), fasting blood glucose (FBG), high density lipoprotein cholesterol (HDL-C), low density lipoprotein cholesterol (LDL-C), total cholesterol (TC), triglycerides (TG), creatinine (CRE), and blood urea nitrogen (BUN) in the paired groups. Both serum adiponectin and leptin levels were significantly higher in patients with PCa compared to healthy controls (P<0.001 for both). Subsequently, there was a positive correlation between adiponectin and PSA (r=0.285, P<0.001). Positive significant correlations between BMI, PSA, TG and leptin were also observed in whole group (r=0.270, P<0.001; r=0.348, P<0.001; r=0.170, P=0.021, respectively). However, the serum adiponectin and leptin levels were not related to the Gleason score of PCa. Receiver operating characteristic curves analysis of the investigated serum adiponectin differentiated cancer patients from the healthy individuals with a sensitivity of 87%, specificity of 56%. Leptin levels also distinguished patients from the healthy controls with a sensitivity of 69%, specificity of 68%. Our study shows that the serum levels of adiponectin and leptin in PCa patients were higher than healthy controls. Adiponectin and leptin may be important markers of PCa. For validation, further studies including large cohort studies would be required.

Keywords: Prostate cancer, circulating biomarkers, adiponectin, leptin, ELISA

Introduction

Prostate cancer (PCa) is becoming an increasingly noticeable public health problem, especially for those countries with an aging society. As it was described in the very recent cancer statistics, PCa is estimated to account for about 21% of all cases of cancers newly diagnosed and 8% of cancer-related deaths in men in America [1]. Although China was reported to have a lower incidence rate of PCa, the incidence in China has already increased steadily over the past few decades [2]. The etiology of PCa is largely inconclusive. Accumulative proofs have verified that genetic alternations and other etiology risk factors such as red and pro-

cessed meat consumption, fat intake, related nutrients, and obesity are considered to be risk factors to develop PCa in a complex manner [3]. Over the past few decades, the potential role of obesity in promoting the process of carcinogenesis has been gradually discovered. It has been estimated that roughly 20% of all cancers were caused by excess weight gain, and this percent may be underestimated [4, 5] and the relationship between rectal cancer, renal cancer and obesity has been revealed [6, 7]. Although individual studies were conflicted regarding the association between obesity and PCa, obesity is believed to be an increasingly prevalent factor in contributing to the high incidence of PCa, and some large meta-analyses

have reported that obesity was related to a modestly increasing incidence of PCa [8, 9]. However, the potential mechanisms between cancer development and obesity are largely undiscovered. Several studies have elucidated that a variety of physiological and pathological processes such as insulin resistance, hyperinsulinemia, sustained hyperglycemia, glucose intolerance, oxidative stress, inflammation and/or adipokine production were well-recognized risk factors contributing to the link of obesity to cancer association and determining the patients' risks [10]. Among the various factors, the participation of adipokines has been proposed recently. Adipokines are secreted by adipose tissue which is currently considered as a complex and crucial endocrine organ in cancer development. It has been indicated the aberrant adipokines production of adipose tissue may result in chronic inflammation in the microenvironment and thereby initiate or promote carcinogenesis [11]. Several adipokines such as omentin-1, adiponectin and leptin are biologically active polypeptides produced by adipocytes and have been shown to be involved in obesity's association with PCa [12, 13].

Adiponectin is secreted exclusively from adipose tissue, which is encoded by the gene AdipoQ and makes a protein 244 amino acids in length that is 30 kDa in weight [14]. It has been shown that adiponectin played vital roles in anti-atherosclerosis, anti-insulin resistance [14-16]. Leptin is a protein that is 16 kDa in weight and 167 amino acids in length [17]. Indeed, research suggests that leptin played a role in the progression of mammary tissue tumorigenesis via its function as a growth hormone [18]. Importantly, an inverse correlation between reduced adiponectin and obesity and a positive correlation between leptin levels and obesity had been reported [19, 20].

In addition, several studies have reported increased or decreased circulating adiponectin and leptin levels in colorectal, breast, pancreatic, ovarian, and lung cancer patients [15, 19, 21, 22], indicating a underlying role with tumorigenesis. Currently, to our knowledge, circulating levels of adiponectin and leptin in PCa patients are largely unexplored in Asian area, especially in Chinese population. Herein, we conduct this matched case-control study to determine the serum levels of adiponectin and

leptin in patients with PCa in Chinese population.

Materials and methods

Patients and healthy controls

Between June 2014 and June 2015, 92 patients newly diagnosed with PCa and who underwent trans-rectal prostate biopsy at the first affiliated hospital of Anhui Medical University were enrolled in this study. Patients with PCa were divided into three groups according to grades (low, intermediate, and high grade determined by a Gleason score of less than 7, 7 and more than 7, respectively). Meanwhile, 92 age-matched volunteers were selected as healthy controls from people who confirmed their fitness at the health examination center of the first affiliated hospital of Anhui Medical University. This study was approved by the Ethics Committee of the first affiliated hospital of Anhui Medical University, Hefei, Anhui, People's Republic of China. All candidates provided written informed consent to allow analysis of data for research purposes.

All patients were recruited using the following criteria: no curative medication for prostate diseases; no history of malignancy or prostate operations; no diagnosis of acute infectious diseases, and no impairment of heart, liver or kidney. Venous blood samples were drawn from all patients and controls after fasting for at least 12 hours. Hemolytic, lipaemic or icteric samples were discarded. Samples were then centrifuged with 8000 rpm at 4°C for 4 minutes and the supernatant was collected. All the serum were kept in polypropylene tubes and stored at -80°C until detection.

Physical and biochemical measurements

Anthropometric measurements obtained in this study included height, weight. Body mass index (BMI) was calculated as body weight divided by height squared (kg/m²). Biochemical parameters were measured in the stored serum samples. The serum levels of total cholesterol (TC), triacylglycerol (TG), high density lipoprotein-cholesterol (HDL-C), low density lipoprotein-cholesterol (LDL-C) levels, fasting blood glucose (FBG), creatinine (CRE), blood urea nitrogen (BUN) and prostate specific antigen (PSA) were detected using the standard

Table 1. Comparison of general characteristics and biochemical parameters

Variables	Prostate cancer group (n=92)	Healthy control group (n=92)	T or Z [∆]	P value
Age (years)	72.20±6.15	70.88±5.45	1.535	0.126
BMI (kg/m ²)	24.08±3.01	24.42±2.87	-0.793	0.430
PSA (ng/mL)	36.80±35.44	1.13±0.44	-11.719△	<0.001
FBG (mmol/L)	5.92±1.18	6.04±1.67	-0.588	0.557
HDL-C (mmol/L)	1.48±0.36	1.37±0.32	2.187	0.030
LDL-C (mmol/L)	2.86±0.80	2.70±0.82	1.381	0.169
TC (mmol/L)	4.41±0.94	4.54±0.91	-1.002	0.318
TG (mmol/L)	1.51±0.55	1.36±0.58	1.749	0.082
CRE (µmol/L)	80.08±22.91	77.87±14.76	0.777	0.438
BUN (mmol/L)	6.04±1.40	5.81±1.34	1.119	0.264
Adiponectin (µg/mL)	20.32±12.09	14.56±11.72	-3.758△	<0.001
Leptin (ng/mL)	6.28±7.28	2.44±2.52	-5.177△	<0.001

Bold values are statistically significant P<0.05. Abbreviations: BMI body mass index, PSA prostate-specific antigen, FBG fasting blood glucose, HDL-C high density lipoprotein-cholesterol, LDL-C low density lipoprotein-cholesterol, TC total cholesterol, TG triacylglycerol, CRE creatinine, BUN blood urea nitrogen.

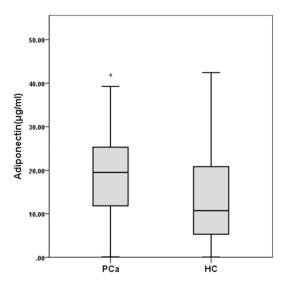


Figure 1. The adiponectin levels of PCa patients and healthy control people (n=92 for each group).

methods by a clinical chemistry analyzer (Shimadzu, cl8000, Japan).

Serum adiponectin and leptin concentration determination

Concentrations of adiponectin and leptin were detected in serum samples reserved using an enzyme-linked immunosorbent assay (ELISA) according to the user manual (adiponectin: Cusabio, CSB-E07270h, China; leptin: Cusabio,

CSB-E04649h, China). The linear ranges of the assay were 1.562 ng/mL-100 ng/mL for adiponectin and 0.156 ng/mL-10 ng/mL for leptin. The inter-assay and intra-assay coefficients of variation were less than 8% and 10%, respectively.

Statistical analysis

IBM SPSS Statistics for Windows version 20 (IBM Corp, Armonk, NY, USA) was used for statistical analyses. A twotailed *P*<0.05 was considered as significant for all analyses. Variables were investigated using visual (histograms, probability plots) and analytical methods (Kolmogorov-Smirnov test) to determine whe-

ther they were normally distributed. Quantitative characteristics were reported as mean ± standard deviation (SD). If they were normallydistributed variables, the t-test was introduced for comparison within the group and between groups; if not, we selected the nonparametric Wilcoxon Mann-Whitney test to assess the significant differences between different groups. Subsequently, Spearman's correlation analyses were conducted to test for associations between the level of adipokines and general clinical characteristics and biochemical parameters. Non-parametric Kruskal-Wallis test was used to explore the associations between PCa grades and circulating levels of the adipokines. Receiver operating characteristic (ROC) curve analysis was conducted to evaluate the sensitivity and specificity of adiponectin and leptin in the prediction of malignancy and to differentiated cancer patients from the healthy individuals.

Results

The general characteristics including anthropometric measurements and biochemical parameters of patients with PCa (92 subjects) compared with the healthy control group (92 subjects) were listed in **Table 1**. No significant differences were observed in age and BMI between the PCa patients and the healthy controls (*P*>0.05). In addition, we did not find sta-

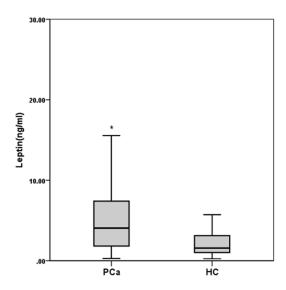


Figure 2. The leptin levels of PCa patients and healthy control people (n=92 for each group).

tistically significant differences in FBG, LDL-C, TC, TG, CRE and BUN levels between PCa patients and healthy controls (P>0.05). Patients with PCa had a higher level of PSA (P<0.001) and HDL-C (P=0.030) than healthy individuals. The adiponectin and leptin levels in both groups were shown in **Table 1**. PCa patients had significantly higher serum adiponectin (P<0.001) and leptin (P<0.001) levels compared with the healthy controls (**Figures 1, 2**).

Spearman's correlation analyses of adiponectin and leptin levels with clinical and biochemical parameters

Then, we analyzed the correlation between the serum adiponectin and leptin levels with various clinical characteristics and biochemical parameters in PCa group, healthy control group and whole group respectively via Spearman's rank correlation coefficient analysis. The result showed that serum levels of adiponectin was not significantly correlated with age, BMI, serum FBG, HDL-C, LDL-C, TC, TG, CRE, and BUN levels in each group (Table 2). Whereas, adiponectin correlated positively with PSA (r= 0.282, P<0.001) in the whole group (**Table 2**). For leptin, we found a positive correlation between leptin and BMI (r=0.269, P<0.001), TG (r=0.205, P=0.005), PSA (r=0.335, P< 0.001) in the whole group. We also found a positive association between leptin and BMI (r=0.461, P<0.001), and FBG (0.345, P=0.001)in healthy control group (Table 3).

Associations between adiponectin and leptin levels with grades

We have demonstrated no significant correlation between serum adiponectin levels with PCa grades (χ^2 =0.047, P=0.964) (**Table 4**). We also found no significant association between circulating leptin levels with grades (χ^2 =1.407, p=0.495) in PCa patients (**Table 5**).

ROC curve of adiponectin and leptin in PCa patients and healthy controls

We showed the ROC curves of the investigated serum adiponectin in **Figure 3**, Using a cutoff point of $8.28 \, \mu g/mL$ for serum levels of adiponectin, we were able to differentiate patients with PCa patients from healthy controls with a sensitivity of 87%, specificity of 56%. In **Figure 4**, leptin serum levels differentiated PCa patients from the healthy controls with a sensitivity of 69%, specificity of 68%, using a cutoff point of $2.40 \, ng/mL$.

Discussion

Prostate cancer (PCa) is the most common urological cancer, which accounts for more than 20% of men's malignant neoplasms. There has been a gradual increase in the frequencies of PCa and obesity in developed countries, with increasingly western eating habits represented by high fat and high cholesterol [23]. Therefore, the parallel morbidity trends of PCa and obesity indicated the possible relevance between them. However, the association between obesity and PCa incidence is complex and has yielded inconsistent conclusions. Park et al. reported that obesity was associated with a higher risk of PCa detection as an independent factor [24], which was supported by several other studies [25, 26]. Among them, Barrington et al. elucidated that obesity was more strongly associated with increased PCa risk among African American than non-Hispanic white men and reducing obesity among African American men could reduce the racial disparity in cancer incidence [25]. Moreover, previous meta-analysis reported associations between a higher BMI and a higher risk of PCa [27]. Conversely, a large age-matched casecontrol study revealed that elevated BMI was associated with a lower risk of PCa [28]. And interestingly, there were also some other studies argued that BMI was not associated with the risk of PCa [29-31].

Altered adipokines levels in prostate cancer patients

Table 2. Spearman's correlation analysis of adiponectin levels with clinical parameters and biochemical parameters

	Prostate can	Prostate cancer group		Normal control group		Whole group	
Variables	Correlation coefficient (r)	P value	Correlation coefficient (r)	P value	Correlation coefficient (r)	P value	
Age (years)	-0.182	0.082	-0.151	0.150	-0.103	0.164	
BMI (kg/m²)	-0.170	0.105	0.132	0.210	-0.143	0.052	
PSA (ng/mL)	0.136	0.196	0.084	0.424	0.285	<0.001	
FBG (mmol/L)	-0.163	0.121	-0.025	0.814	-0.096	0.193	
HDL-C (mmol/L)	0.079	0.457	0.029	0.782	0.096	0.193	
LDL-C (mmol/L)	-0.028	0.788	0.046	0.666	0.055	0.458	
TC (mmol/L)	0.171	0.103	-0.048	0.651	0.036	0.629	
TG (mmol/L)	-0.083	0.434	-0.004	0.969	0.028	0.784	
CRE (µmol/L)	-0.039	0.715	-0.146	0.164	-0.100	0.178	
BUN (mmol/L)	0.000	0.997	0.076	0.473	0.072	0.334	

Bold values are statistically significant P<0.05. Abbreviations: BMI body mass index, PSA prostate-specific antigen, FBG fasting blood glucose, HDL-C high density lipoprotein-cholesterol, LDL-C low density lipoprotein-cholesterol, TC total cholesterol, TG triacylglycerol, CRE creatinine, BUN blood urea nitrogen.

Table 3. Spearman's correlation analysis of leptin levels with clinical parameters and biochemical parameters

	Prostate can	Prostate cancer group		Normal control group		Whole group	
Variables	Correlation coefficient (r)	Pyalue		P value	Correlation coefficient (r)	P value	
Age (years)	0.187	0.075	0.047	0.658	0.183	0.013	
BMI (kg/m ²)	0.189	0.071	0.401	<0.001	0.270	<0.001	
PSA (ng/mL)	0.176	0.093	-0.116	0.273	0.348	<0.001	
FBG (mmol/L)	0.041	0.699	0.346	0.001	0.144	0.050	
HDL-C (mmol/L)	0.009	0.934	-0.088	0.402	0.015	0.845	
LDL-C (mmol/L)	0.061	0.564	-0.073	0.489	0.046	0.536	
TC (mmol/L)	0.069	0.512	-0.066	0.534	-0.030	0.686	
TG (mmol/L)	0.078	0.459	0.103	0.329	0.170	0.021	
CRE (µmol/L)	0.078	0.462	0.083	0.430	0.086	0.243	
BUN (mmol/L)	0.069	0.512	0.027	0.795	0.093	0.211	

Bold values are statistically significant P<0.05. Abbreviations: BMI body mass index, PSA prostate-specific antigen, FBG fasting blood glucose, HDL-C high density lipoprotein-cholesterol, LDL-C low density lipoprotein-cholesterol, TC total cholesterol, TG triacylglycerol, CRE creatinine, BUN blood urea nitrogen.

Table 4. Non-parametric Kruskal-Wallis test on adiponectin levels with PCa status

PCa Grades	N	Mean rank	χ²	P value
Low (Gleason<7)	27	1.96	0.047	0.964
Intermediate (Gleason=7)	29	2.04		
High (Gleason>7)	36	2.00		

Among the different hypothesis that described the relationship between obesity and tumor development, the impact of adipokines on carcinogenesis has been widely discussed. Abnormal synthesis of adipokines such as

omentin-1, resistin, apelin, adiponectin and leptin, results in chronic low-grade inflammation in the microenvironment, which may contribute to tumor initiation or progression [11, 32]. For instance, it was showed that omentin-1 significantly inhibited the proliferation of hepatocellular carcinoma cells by inducing apoptosis in these cells [33].

Adiponectin is synthesized in white adipose tissue and exerts function of anti-atherosclerosis, anti-inflammation or anti-insulin resistance [15]. Adiponectin is involved in glucose and lipid homeostasis and is therefore implicated

Table 5. Non-parametric Kruskal-Wallis test on leptin levels with PCa status

PCa Grades	N	Mean rank	χ^2	P value
Low (Gleason<7)	27	1.81	1.407	0.495
Intermediate (Gleason=7)	29	2.07		
High (Gleason>7)	36	2.11		

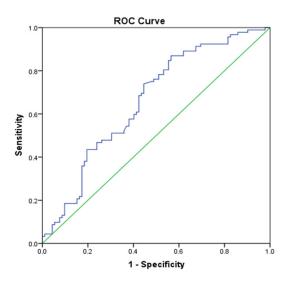


Figure 3. Receiver operation characteristic (ROC) Curve of adiponectin in PCa patients and Healthy controls.

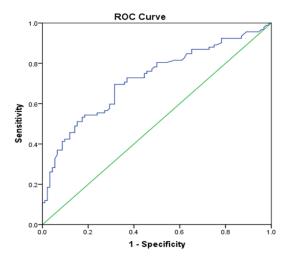


Figure 4. Receiver operation characteristic (ROC) Curve of leptin in PCa patients and Healthy controls.

in the pathogenesis of insulin resistance and diabetes [34]. Altered concentrations of adiponectin have been reported in PCa patients. Although some of these studies reported that cancer patients had significantly lower adipo-

nectin levels than controls [35-37], some other studies did not find any difference in concentrations of adiponectin between patients and controls [33, 38]. Our study demonstrated that patients with PCa had a higher level of adiponectin than healthy controls in Chinese population, which was supported by Al Khaldi et al. [39]. In addition, it has

been shown that low adiponectin levels were related to high colorectal cancer risk [40]. Also, some reported circulated levels of adiponectin were inversely correlated with renal cancer incidence [41]. In PCa. Michalakis et al. reported an inverse relationship between adiponectin levels and risk of PCa in an epidemiological study [36], which was supported by another study [42]. However, there were other studies demonstrated that adiponectin concentrations were not associated with risk of prostate cancer [38]. In our study, the circulating adiponectin levels of 20.32±12.09 µg/mL were obviously higher within PCa patients than the healthy people of 14.56±11.72 µg/mL (P<0.05). In addition, we found no correlation between the adiponectin level and PCa grades, indicating that adiponectin might not be associated with the histological grade and disease stage.

Leptin is a kind of peptide hormone which is secreted from adipose tissue in proportion to an individual's fat mass and exerts its effects via blood circulation with targets such as the central nervous system, muscle, liver and adipose tissue [43]. Similar to adiponectin, the leptin concentrations were correlated with obesity and altered concentrations of leptin have been reported in cancer patients. Epidemiological studies indicated increased circulating levels of leptin, as occurs during obesity, were associated with cancers, such as breast and colorectal cancer [19]. Research suggested that leptin played a role in the progression of mammary tissue tumorigenesis via its function as a growth hormone [18]. In addition, leptin had also been studied in vitro on cancer cells and was concerned with proliferation of ovarian, breast, lung pancreatic, and colorectal cancers [22], indicating a crucial effect of leptin in cancer development. In prostate cancer, serum leptin was reported significantly higher in patients with prostate cancer as compared to controls [44], which was supported by other studies [13]. Another study reported although mean serum levels of leptin in case patients

were 10% higher than those in control subjects, the difference was not statistically significant [38]. However, studies reported no significant association between plasma leptin levels and PCa risk was found [38, 45, 46]. Interestingly, there were also studies reported that PCa was associated with raised serum leptin, which was independent of obesity and serum PSA [44]. In our study, the leptin level in PCa patients at 6.28±7.28 ng/ml was higher than healthy controls at 2.44 ± 2.52 ng/ml (p<0.001). Furthermore, the serum levels were not correlated with different clinical and biochemical parameters, such as age, FBG, HDL-C, HDL-C, TC, CRE and BUN. In addition, the serum levels were positively associated with BMI and PSA. suggesting leptin may be a marker of PCa related to BMI. Thus, obesity-related leptin exhibits a tumorigenic role in prostate cancer. Similar to adiponectin, we also did not find a correlation between the leptin level and PCa grades, suggesting that leptin also might not be correlated with the histological grades and stages of PCa.

ROC analysis is a standard methodology to evaluate the performance of a classification system, which is applied extensively within clinical medicine. The ROC curve is a two-dimensional plot which illustrates the relationship between the true positive rate (sensitivity) and the false positive rate (1-specificity) of a binary classifier [47-49]. In our study, ROC analysis of the investigated serum adiponectin differentiated cancer patients from the healthy individuals with a sensitivity of 87%, specificity of 56%; leptin levels also distinguished patients from healthy controls with a sensitivity of 69%, specificity of 68%, indicating the diagnostic role of the adipokines.

In conclusion, we found significantly elevated adiponectin and leptin levels in patients with PCa, which were independent of most of the clinical and biochemical parameters. Adiponectin and leptin may be important markers of PCa. In the future, more large cohort-based researches will be necessary to elucidate the possible mechanisms underlying the deregulated adipokines levels and the interactions between adipose tissues with PCa. Such efforts will shed novel insights into effective and efficient therapy and diagnosis of prostate cancer.

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Disclosure of conflict of interest

None.

Address correspondence to: Chaozhao Liang and Li Zhang, Department of Urology, The First Affiliated Hospital of Anhui Medical University, Hefei 230022, China; Institute of Urology, Anhui Medical University, No. 218 Jixi Road, Hefei 230022, China. Tel: +86-551-62923932; Fax: +86-551-63633742; E-mail: Izhang@ahmu.edu.cn (LZ); Iiang_chaozhao@ahmu.edu.cn (CZL)

References

- Siegel RL, Miller KD, Jemal A. Cancer statistics, 2016. CA Cancer J Clin 2017; 67: 7-30.
- [2] Chen W, Zheng R, Baade PD, Zhang S, Zeng H, Bray F, Jemal A, Yu XQ, He J. Cancer statistics in China, 2015. CA Cancer J Clin 2016; 66: 115-132.
- [3] Stefani ED, Boffetta PL, Ronco A, Deneo-Pellegrini H. Meat consumption, related nutrients, obesity and risk of prostate cancer: a casecontrol study in uruguay. Asian Pac J Cancer Prev 2016; 17: 1937-1945.
- [4] Calle EE, Rodriguez C, Walker-Thurmond K, Thun MJ. Overweight, obesity, and mortality from cancer in a prospectively studied cohort of U.S. adults. N Engl J Med 2003; 348: 1625-1638.
- [5] Wolin KY, Carson K, Colditz GA. Obesity and cancer. Oncologist 2010; 15: 556-565.
- [6] Shen XD, Zhang L, Che H, Zhang YY, Yang C, Zhou J, Liang CZ. Circulating levels of adipocytokine omentin-1 in patients with renal cell cancer. Cytokine 2016; 77: 50-55.
- [7] Fazeli MS, Dashti H, Akbarzadeh S, Assadi M, Aminian A, Keramati MR, Nabipour I. Circulating levels of novel adipocytokines in patients with colorectal cancer. Cytokine 2013; 62: 81-85.
- [8] Bergstrom A, Pisani P, Tenet V, Wolk A, Adami HO. Overweight as an avoidable cause of cancer in Europe. Int J Cancer 2001; 91: 421-430.
- [9] Hu MB, Liu SH, Jiang HW, Bai PD, Ding Q. Obesity affects the biopsy-mediated detection of prostate cancer, particularly high-grade pros-

- tate cancer: a dose-response meta-analysis of 29,464 patients. PLoS One 2014; 9: e106677.
- [10] De Pergola G, Silvestris F. Obesity as a major risk factor for cancer. J Obes 2013; 2013: 291546.
- [11] Liao LM, Schwartz K, Pollak M, Graubard BI, Li Z, Ruterbusch J, Rothman N, Davis F, Wacholder S, Colt J, Chow WH, Purdue MP. Serum leptin and adiponectin levels and risk of renal cell carcinoma. Obesity (Silver Spring) 2013; 21: 1478-1485.
- [12] Uyeturk U, Sarici H, Kin Tekce B, Eroglu M, Kemahli E, Uyeturk U, Gucuk A. Serum omentin level in patients with prostate cancer. Med Oncol 2014; 31: 923.
- [13] Arisan ED, Arisan S, Atis G, Palavan-Unsal N, Ergenekon E. Serum adipocytokine levels in prostate cancer patients. Urol Int 2009; 82: 203-208.
- [14] Scherer PE, Williams S, Fogliano M, Baldini G, Lodish HF. A novel serum protein similar to C1q, produced exclusively in adipocytes. J Biol Chem 1995: 270: 26746-26749.
- [15] Barb D, Williams CJ, Neuwirth AK, Mantzoros CS. Adiponectin in relation to malignancies: a review of existing basic research and clinical evidence. Am J Clin Nutr 2007; 86: s858-866.
- [16] Ziemke F, Mantzoros CS. Adiponectin in insulin resistance: lessons from translational research. Am J Clin Nutr 2010; 91: 258S-261S.
- [17] Friedman J. 20 years of leptin: leptin at 20: an overview. J Endocrinol 2014; 223: T1-8.
- [18] Roberts DL, Dive C, Renehan AG. Biological mechanisms linking obesity and cancer risk: new perspectives. Annu Rev Med 2010; 61: 301-316.
- [19] Tessitore L, Vizio B, Jenkins O, De Stefano I, Ritossa C, Argiles JM, Benedetto C, Mussa A. Leptin expression in colorectal and breast cancer patients. Int J Mol Med 2000; 5: 421-426.
- [20] Arita Y, Kihara S, Ouchi N, Takahashi M, Maeda K, Miyagawa J, Hotta K, Shimomura I, Nakamura T, Miyaoka K, Kuriyama H, Nishida M, Yamashita S, Okubo K, Matsubara K, Muraguchi M, Ohmoto Y, Funahashi T, Matsuzawa Y. Paradoxical decrease of an adipose-specific protein, adiponectin, in obesity. Biochem Biophys Res Commun 1999; 257: 79-83.
- [21] Drew JE. Molecular mechanisms linking adipokines to obesity-related colon cancer: focus on leptin. Proc Nutr Soc 2012; 71: 175-180.
- [22] Garofalo C, Surmacz E. Leptin and cancer. J Cell Physiol 2006; 207: 12-22.
- [23] Yang L, Colditz GA. Prevalence of overweight and obesity in the united states, 2007-2012. JAMA Intern Med 2015; 175: 1412-1413.
- [24] Park J, Cho SY, Lee SB, Son H, Jeong H. Obesity is associated with higher risk of prostate cancer detection in a biopsy population in Korea. BJU Int 2014; 114: 891-895.

- [25] Barrington WE, Schenk JM, Etzioni R, Arnold KB, Neuhouser ML, Thompson IM Jr, Lucia MS, Kristal AR. Difference in association of obesity with prostate cancer risk between US African American and bon-hispanic white men in the Selenium and Vitamin E Cancer Prevention Trial (SELECT). JAMA Oncol 2015; 1: 342-349.
- [26] Hu MB, Bai PD, Wu YS, Zhang LM, Xu H, Na R, Jiang HW, Ding Q. Higher body mass index increases the risk for biopsy-mediated detection of prostate cancer in Chinese men. PLoS One 2015; 10: e0124668.
- [27] Bhaskaran K, Douglas I, Forbes H, dos-Santos-Silva I, Leon DA, Smeeth L. Body-mass index and risk of 22 specific cancers: a populationbased cohort study of 5.24 million UK adults. Lancet 2014; 384: 755-765.
- [28] Boehm K, Sun M, Larcher A, Blanc-Lapierre A, Schiffmann J, Graefen M, Sosa J, Saad F, Parent ME, Karakiewicz Pl. Waist circumference, waist-hip ratio, body mass index, and prostate cancer risk: results from the North-American case-control study prostate cancer & environment study. Urol Oncol 2015; 33: 494, e1-7.
- [29] Benn M, Tybjaerg-Hansen A, Smith GD, Nordestgaard BG. High body mass index and cancer risk-a Mendelian randomisation study. Eur J Epidemiol 2016; 31: 879-92
- [30] Bonn SE, Sjolander A, Tillander A, Wiklund F, Gronberg H, Balter K. Body mass index in relation to serum prostate-specific antigen levels and prostate cancer risk. Int J Cancer 2016; 139: 50-57.
- [31] Baillargeon J, Platz EA, Rose DP, Pollock BH, Ankerst DP, Haffner S, Higgins B, Lokshin A, Troyer D, Hernandez J, Lynch S, Leach RJ, Thompson IM. Obesity, adipokines, and prostate cancer in a prospective population-based study. Cancer Epidemiol Biomarkers Prev 2006; 15: 1331-1335.
- [32] Aprahamian TR, Sam F. Adiponectin in cardiovascular inflammation and obesity. Int J Inflam 2011; 2011: 376909.
- [33] Zhang YY, Zhou LM. Omentin-1, a new adipokine, promotes apoptosis through regulating Sirt1-dependent p53 deacetylation in hepatocellular carcinoma cells. Eur J Pharmacol 2013; 698: 137-144.
- [34] Duncan BB, Schmidt MI, Pankow JS, Bang H, Couper D, Ballantyne CM, Hoogeveen RC, Heiss G. Adiponectin and the development of type 2 diabetes: the atherosclerosis risk in communities study. Diabetes 2004; 53: 2473-2478.
- [35] Goktas S, Yilmaz MI, Caglar K, Sonmez A, Kilic S, Bedir S. Prostate cancer and adiponectin. Urology 2005; 65: 1168-1172.
- [36] Michalakis K, Williams CJ, Mitsiades N, Blakeman J, Balafouta-Tselenis S, Giannopoulos A, Mantzoros CS. Serum adiponectin concentra-

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- tions and tissue expression of adiponectin receptors are reduced in patients with prostate cancer: a case control study. Cancer Epidemiol Biomarkers Prev 2007; 16: 308-313.
- [37] Tewari R, Rajender S, Natu SM, Goel A, Dalela D, Goel MM, Tondon P. Significance of obesity markers and adipocytokines in high grade and high stage prostate cancer in North Indian men - a cross-sectional study. Cytokine 2013; 63: 130-134.
- [38] Li H, Stampfer MJ, Mucci L, Rifai N, Qiu W, Kurth T, Ma J. A 25-year prospective study of plasma adiponectin and leptin concentrations and prostate cancer risk and survival. Clin Chem 2010; 56: 34-43.
- [39] Al Khaldi RM, Al Mulla F, Al Awadhi S, Kapila K, Mojiminiyi OA. Associations of single nucleotide polymorphisms in the adiponectin gene with adiponectin levels and cardio-metabolic risk factors in patients with cancer. Dis Markers 2011; 30: 197-212.
- [40] Wei EK, Giovannucci E, Fuchs CS, Willett WC, Mantzoros CS. Low plasma adiponectin levels and risk of colorectal cancer in men: a prospective study. J Natl Cancer Inst 2005; 97: 1688-1694.
- [41] Spyridopoulos TN, Petridou ET, Skalkidou A, Dessypris N, Chrousos GP, Mantzoros CS, Obesity, Cancer Oncology G. Low adiponectin levels are associated with renal cell carcinoma: a case-control study. Int J Cancer 2007; 120: 1573-1578.
- [42] Schenk JM, Kristal AR, Neuhouser ML, Tangen CM, White E, Lin DW, Thompson IM. Serum adiponectin, C-peptide and leptin and risk of symptomatic benign prostatic hyperplasia: results from the prostate cancer prevention trial. Prostate 2009; 69: 1303-1311.

- [43] Margetic S, Gazzola C, Pegg GG, Hill RA. Leptin: a review of its peripheral actions and interactions. Int J Obes Relat Metab Disord 2002; 26: 1407-1433.
- [44] Singh SK, Grifson JJ, Mavuduru RS, Agarwal MM, Mandal AK, Jha V. Serum leptin: a marker of prostate cancer irrespective of obesity. Cancer Biomark 2010; 7: 11-15.
- [45] Stattin P, Kaaks R, Johansson R, Gislefoss R, Soderberg S, Alfthan H, Stenman UH, Jellum E, Olsson T. Plasma leptin is not associated with prostate cancer risk. Cancer Epidemiol Biomarkers Prev 2003; 12: 474-475.
- [46] Hsing AW, Chua S Jr, Gao YT, Gentzschein E, Chang L, Deng J, Stanczyk FZ. Prostate cancer risk and serum levels of insulin and leptin: a population-based study. J Natl Cancer Inst 2001; 93: 783-789.
- [47] Toh KA, Kim J, Lee S. Maximizing area under ROC curve for biometric scores fusion. Pattern Recognit 2008; 41: 3373-3392.
- [48] Berrar D, Flach P. Caveats and pitfalls of ROC analysis in clinical microarray research (and how to avoid them). Brief Bioinform 2012; 13: 83-97.
- [49] Wang SJ, Li D, Petrick N, Sahiner B, Linguraruc MG, Summersa RM. Optimizing area under the ROC curve using semi-supervised learning. Pattern Recognit 2015; 48: 276-287.