Original Article Elevated expression of LIMK2 is an independent prognostic indicator in breast cancer

Peng Sun^{1,2*}, Lin Ren^{1*}, Qing-Qiu Chen¹, Xuan-Ni Tan¹, Ting Zhang¹, Yan-Ling Li¹, Fan Zhang¹, Jun Jiang¹

¹Breast Disease Center, Southwest Hospital, Third Military Medical University, Chongqing 400038, China; ²Department of Breast, Thyroid Surgery, Chengdu Military General Hospital, Chengdu, Sichuan Province, China. *Equal contributors.

Received June 15, 2017; Accepted March 13, 2018; Epub June 15, 2018; Published June 30, 2018

Abstract: Background: LIM domain kinase 2 (LIMK2) plays an important role in cell proliferation, adhesion, migration, differentiation and inflammation. Abnormal expression of LIMK2 is implicated in various malignancies, but little is known about its expression and prognostic value in breast cancer. Methods: Specimens were collected from 212 patients for the analysis of LIMK2 expression by immunohistochemistry. Kaplan-Meier analysis and Cox regression analysis were performed to evaluate the prognostic significance of LIMK2 in breast cancer. The same analyses were conducted using METABRIC and the Kaplan Meier plotter databases to validate the expression pattern and prognostic value of LIMK2. Results: LIMK2 was located in the cytoplasm, and the positive rate of LIMK2 expression in the breast cancer tissues was significantly higher than that in normal breast tissues (P<0.0001). Elevated LIMK2 expression correlated with large tumor size and high histological grade (P<0.05 for each). Kaplan-Meier analysis showed elevated expression of LIMK2 predicted worse disease-free survival (DFS) (HR: 3.295, 95% CI: 2.251-7.156, P<0.0001) and overall survival (OS) (HR: 6.251, 95% CI: 3.874-16.260, P<0.0001). Multivariable Cox regression analysis indicated elevated expression of LIMK2 was an independent prognostic factor for both DFS and OS (for DFS, HR: 2.707, 95% CI: 1.546-4.740, P=0.0005; for OS, HR: 5.241, 95% CI: 2.436-11.277, P<0.0001). The expression pattern and prognostic significance of LIMK2 in breast cancer patients were validated by bioinformatics analysis of public databases. Conclusion: In conclusion, our results suggest that LIMK2 is an independent poor prognostic factor for breast cancer patients, and LIMK2 might play an important role in the progression of breast cancer.

Keywords: LIMK2, breast cancer, expression, prognosis

Introduction

Invasion and metastasis are the main causes of death in breast cancer. Mounting evidence suggested that LIM domain kinases (LIMK), key regulators of the actin cytoskeleton, was prominently associated with tumorcell invasion and metastasis [1, 2]. Typical structure of LIMKs is comprised with two LIM domains at the Nterminus, a PDZ domain connected to proline/ serine-rich regions and a C-terminal kinase domain [3, 4]. LIMK family has two members, LIMK-1 and LIMK-2 [4]. Though they share about 50% of overall identical sequence, different expression profile, subcellular and organic location and function were observed between them [5-7].

LIMKs are play an important role in cell proliferation, adhesion, migration, differentiation and inflammation [7-13]. The overexpression of LIMK1 was detected in melanoma, prostate and breast cancer [14-17]. Recently, LIMK2 was reported to be required for the formation of invadopodia, matrix degradation and invasion as well as migration of breast cancer cells [13], which suggested that LIMK2 might be involved in the progression of breast cancer; however, the expression pattern of LIMK2 in breast cancer cer and its correlations with prognosis of breast patients is poorly understood.

To investigate the prognostic value of LIMK2 in breast cancer patients, specimens from 212 patients were collected for the analysis of LIMK2 expression by immunohistochemistry (IHC). The relationship between LIMK2 expression and clinicopathological parameters of breast cancer was evaluated. Meanwhile, the prognostic roles of LIMK2 in breast cancer were

Characteristics	Number of patients (%)
Total	212
Age (years)	
<50	99 (46.70%)
≥50	113 (53.30%)
Location	, , , , , , , , , , , , , , , , , , ,
Left	122 (57.55%)
Right	90 (42.45%)
pT Stage	
0	3 (1.42%)
1	65 (30.66%)
2	121 (57.08%)
3	8 (3.77%)
4	15 (7.08%)
pN stage	20 (1100/07)
0	99 (46,70%)
1	84 (39.62%)
2	10 (4 72%)
-	19 (8 96%)
M stage	10 (0.0070)
0	203 (95 75%)
1	9 (4 25%)
	5 (4.2570)
0	3 (1 42%)
1	34 (16 04%)
2	125 (58 96%)
2	A1 (10 34%)
3	41(19.34%)
4 Histological type	9 (4.25%)
	170 (04 420/)
	12 (6 4.45%)
	13 (0.13%)
Othere	10 (4.72%)
Uners	11 (3.19%)
Histological grade	42 (00 00%)
1	43 (20.28%)
2	144 (67.92%)
3	25 (11.79%)
ER	1.10 (00.0.10)
Positive	140 (66.04%)
Negative	72 (33.96%)
PR	
Positive	109 (51.42%)
Negative	103 (48.58%)
HER2	
Positive	33 (15.57%)
Negative	179 (84.43%)

Table 1. Clinical and pathological characteris-
tics of all 212 breast cancer patients

analyzed with Cox regression and Kaplan-Meier analysis. To validate the expression pattern and prognostic value of LIMK2, the same analyses were conducted using METABRIC and the Kaplan Meier plotter databases. Our data implicated that elevated LIMK2 expression is an independent poor prognostic factor for breast cancer patients.

Materials and methods

Patients and tissue samples

The present study was conducted with the approval of the Ethical and Scientific Committees of Southwest Hospital, Third Military Medical University (Chongqing, China). Patients were informed that the specimens would be used for scientific research, and their privacy would be maintained.

A number of 212 breast cancer patients who treated with surgery at the Southwest Hospital between February 2006 and June 2009 were identified. No treatment were received before surgery. The patients' age ranged between 25 and 79 years with a mean age of 51.4 years. The mean follow-up time is 66 months. Patients' characteristics are shown in **Table 1**. Two experienced pathologists observed the hematoxylin and eosin-stained slides of the different biopsies according to the World Health Organization classification guidelines. Additionally, 17 samples of normal breast tissue were used as normal controls.

Immunohistochemical staining

The ChemMate[™] Envision[™] Detection Kit (Dako, Carpinteria, CA, USA) was used for IHC according to the manufacturer's instructions. Briefly, the samples were fixed by 10% formaldehyde and embedded by paraffin, and then 4-um section-cut specimens was performed. All of the sections were dewaxing and hydration with dimethylbenzene and a gradient concentration of alcohol. Deionized water and phosphate-buffered saline washed the sections, and then an antigen retrieval process was performed at high temperature and high pressure with citrate buffer (pH 6.0). The endogenous peroxidase was locking with 0.3% (v/v) H₀O₂ solution. For reducing nonspecific reaction, the sections were then incubated with goat serum for 60 min Then, the sections were



Figure 1. LIMK2 expression in breast cancer and adjacent normal specimens. (A and B) IHC staining performed for LIMK2 in normal breast tissues, and LIMK2 negative (A) and positive (B) images were shown. Bar, 50 um. (C-E) Weak (C), moderate (D) and strong (E) staining of LIMK2 in breast cancer tissues. Positively reactive substance of LIMK2 was mainly localized in the cytoplasm. Bar, 50 um. (F) The positive rate of LIMK2 expression in breast cancer tissues and normal breast tissues (33.02% vs. 5.882%, **P<0.0001).

incubated with LIMK2 antibody (1:50 dilutions; HPA008183, Sigma-Aldrich, St. Louis, MO, USA), overnight at 4°C. After thawing, the sections were rinsed five times with PBS, then incubated with ChemMate[™] Envision[™]/HRP, rabbit/mouse reagent as a secondary antibody. Subsequently, the sections were treated using ChemMate[™] DAB+ Chromogen (Dako, Carpinteria, CA, USA) and counterstained with hematoxylin. After dehydration and transparency with a gradient concentration of alcohol and dimethylbenzene, all of the specimens were finally fixed using neutral balsam.

Evaluation of staining

Slides were reviewed under a light microscope three time by two observers, who did not know the identity of the specimens between evaluations. Brown-yellow or brown granular deposits at the corresponding antibody expression sites indicated a positive expression result. LIMK2 is expressed in the cytoplasm, and rarely positive expression in the nucleus. The percentage of positive cells was evaluated and scored according to the following categories: 0, less than 5%; 1, 5-25%; 2, 25-50%; 3, 50-75%; 4, greater than 75%. The intensity of staining cells was recorded in the following categories: 0: no staining; 1: weak staining; 2: strong staining. The two scores were summed to obtain an immunoreactivity score (IRS) value ranging from 0 to 6. To evaluate the association of LIMK2 expression with clinical and pathological parameters, the patients were then grouped into two categories based on IRS values: low-expression (IRS 0-5) and high-expression (IRS 6).

Bioinformatics analysis

To validate the expression pattern and prognostic significance of LIMK2, retrospective analyses were conducted based on METABRIC database, which includes genomic profiles of 2,509 breast cancer patients through OASIS platform [18], and Kaplan Meier plotter database [19], which includes information on survival of 5,143 breast cancer patients.

Statistical analysis

Statistical calculations were performed using SAS software (version 9.3; SAS Institute, Cary,

Characteristics	LIMK2 expression (N, %)		- D	
Characteristics	Negative	Positive	— Р	
Total	142 (66.98%)	70 (33.02%)		
Age (years)				
<50	73 (73.74%)	26 (26.26%)	0.0502	
≥50	69 (61.06%)	44 (38.94%)		
Location				
Left	82 (67.21%)	40 (32.79%)	0.9334	
Right	60 (66.67%)	30 (33.33%)		
T stage				
0+1+2	53 (77.94%)	15 (22.06%)	0.0197	
3+4	89 (61.81%)	55 (38.19%)		
N stage				
0	71 (71.72%)	28 (28.28%)	0.1699	
1+2+3	71 (62.83%)	42 (37.17%)		
TNM stage				
0+1+2	113 (69.75%)	49 (30.25%)	0.1224	
3+4	29 (58.00%)	21 (42.00%)		
Histological grade				
1	36 (83.72%)	7 (16.28%)	0.0089	
2+3	106 (62.72%)	63 (37.28%)		
ER status				
Positive	92 (65.71%)	48 (34.29%)	0.5844	
Negative	50 (69.44%)	22 (30.56%)		
PR status				
Positive	75 (68.81%)	34 (31.19%)	0.5608	
Negative	67 (65.05%)	36 (34.95%)		
HER2 status				
Positive	23 (69.70%)	10 (30.30%)	0.7181	
Negative	119 (66.48%)	60 (33.52%)		

Table 2. Relationship between clinical and pathological characteristics and LIMK2 expression

NC, USA). GraphPad Prism (version 6.0; GraphPad Software Inc., La Jolla, CA, USA) was used to analyze the expression of LIMK2 between breast cancer tissues and normal tissues. The expression of LIMK2 with clinicopathological parameters was analyzed by Pearson's Chi-squared tests and Fisher's exact test. Disease-free survival (DFS) was defined as the time from surgery to recurrence of breast cancer (in the breast or chest wall or at nodal or metastatic sites). Overall survival (OS) was defined as the time from surgery to any cause of death. Survival analyses were performed using the Kaplan-Meier method, and differences between groups were assessed using the log-rank test. Univariate analysis comparisons and multivariate survival comparisons were performed using Cox proportional hazard regression models. The estimated relative risks of deaths or relapse were expressed as adjusted hazard ratios (HRs) and corresponding 95% confidence intervals (CIs). A *P* value less than 0.05 was considered statistically significant.

Results

LIMK2 was highly expressed in breast cancer and mainly located in the cytoplasm

In normal breast tissues, the cells showed no or weak staining of LIMK2 (Figure 1A and 1B). In the breast cancer tissues, the positively reactive substance of LIMK2 was mainly located in the cytoplasm, and showed scarcely positive expression in the nucleus (Figure 1C-E). The positive rate of LIMK2 expression in the cytoplasm was notably higher in breast cancer tissues (33.02%, 70/212) than in normal breast tissues (5.882%, 1/17) (P<0.0001, Figure 1F).

The positive rate of LIMK2 was higher in breast cancers patients with tumor larger than 5 cm

The relationships between LIMK2 and clinicopathological parameters of breast cancer patients were analyzed (**Table 2**). The positive rate of LIMK2 was higher in breast cancers patients

with larger diameter (>5 cm) than in cases with smaller size (<5 cm) (P=0.0197). The positive rate of LIMK2 expression was also higher in breast cancers with high histological levels than in cases with low histological levels (P= 0.0089). There were no associations between LIMK2 expression and age, location, N stage, TNM stage, ER, PR and HER2 expression (P>0.05 for each).

High expression of LIMK2 was an independent prognostic factor for both DFS and OS

Breast cancer patients with high LIMK2 expression had significantly lower DFS than those with low LIMK2 expression (HR: 3.295, 95% CI: 2.251-7.156, *P*<0.0001; **Figure 2A**). Breast



Figure 2. Prognostic value of LIMK2 expression in breast cancer patients. Kaplan-Meier analysis of disease-free survival (A) and overall survival (B) for breast cancer patients with high and low LIMK2 expressing tumors.

cancer patients with high LIMK2 expression had significantly lower OS than those with low LIMK2 expression (HR: 6.251, 95% CI: 3.874-16.260, *P*<0.0001; **Figure 2B**). These data suggest that LIMK2 maybe a independent prognostic factor in breast cancer.

Multivariate analyses were performed using the Cox proportional hazards model. We found that tumor size, TNM stage and HER2 expression were proved to be independent prognostic factors for both DFS and OS. Most importantly, elevated LIMK2 expression emerged as an independent prognostic factor for both DFS and OS (for DFS, HR: 2.707, 95% CI: 1.546-4.740, *P*=0.0005; for OS, HR: 5.241, 95% CI: 2.436-11.277, *P*<0.0001) (**Table 3**).

High expression of LIMK2 mRNA indicated poor outcomes of breast cancer patients

To confirm the expression pattern of LIMK2 in breast cancer, we queried the METABRIC database, in which 160 normal breast tissues and

1161 breast cancer tissues were interrogated to evaluate the mRNA expression level of LIMK2. Expression of LIMK2 was significantly increased in breast cancer in comparison with normal tissue (**Figure 3A**, *P*<0.0001). To confirm the prognostic value of LIMK2 in breast cancer, the Kaplan Meier plotter, which could assess the effect of 54,675 genes on survival using 5,143 breast cancer patients, were explored. Elevated mRNA expression of LIMK2 predicted worse relapse-free survival (RFS) (HR: 1.13, 95% Cl: 1.01-1.27, *P*=0.038; **Figure 3B**) and OS (HR: 1.52, 95% Cl: 1.20-1.92, *P*< 0.001; **Figure 3C**).

Discussion

To our knowledge, this is the first study to explore the expression pattern of LIMK2 in breast cancers and correlate its expression level with clinicopathological characteristics and prognosis of breast cancer patients. Our results indicated elevated expression of LIMK2 was associated with large tumor size and high histological grade, and most importantly, it was an independent poor prognostic factor for breast cancer patients.

LIMK2 belongs to LIMK family, which includes LIMK1 and LIMK2. LIMK1 gene locates on human chromosome 7q11, which included 16 exons; whereas, LIMK2 gene locates on human chromosome 22g12.2, which included 19 exons. Both of them have a PDZ domain structure. Two nuclear signal output regions with leucine were found in this domain, which affected the LIMK nucleoplasm shuttle [1, 4, 20, 21]. LIMK1 was overexpressed in breast cancer and its important role in breast cancer tumor growth, angiogenesis and invasion had been demonstrated by numerous studies [16, 22-24]. In the meanwhile, LIMK2 could also contribute to chemotherapy resistance in neuroblastoma cell lines [10, 12] and p53-mediated survival of cancer cells following DNA damage [25]. Moreover, T56-LIMKi, an inhibitor of LIMK2, could effectively inhibit growth of pancreatic cancer [26]. These results indicated LIMK2 might play an important role in cancer cell progression, but little was known about the expression pattern and biological effects of LIMK2 in breast cancer.

Previous studied suggested that LIMK2 served as substrate and key oncogenic effecter of

	DFS			OS		
	HR	95% CI	Р	HR	95% CI	Р
Age	0.988	0.963-1.014	0.3803	1.002	0.972-1.033	0.8919
Location (right vs. left)	0.689	0.396-1.198	0.1866	0.904	0.452-1.809	0.7761
T (T3+T4 vs. T1+T2) ^a	3.049	1.249-7.441	0.0143	3.426	0.991-11.84	0.0516
N (N1-3 vs. N0) ^b	1.513	0.748-3.060	0.2497	1.332	0.543-3.271	0.5313
TNM (III+IV vs. I+II) ^c	2.975	1.594-5.552	0.0006	2.938	1.329-6.492	0.0077
Grade (II+III vs. I) ^d	0.617	0.245-1.552	0.3046	0.769	0.205-2.892	0.6981
ER (+ vs) ^e	0.973	0.509-1.861	0.9346	0.83	0.347-1.987	0.6763
PR (+ vs) ^f	0.659	0.346-1.258	0.2061	0.745	0.318-1.743	0.4971
HER2 (+ vs) ^g	2.219	1.139-4.324	0.0192	2.721	1.227-6.030	0.0137
LIMK2 (+ vs) ^h	2.707	1.546-4.740	0.0005	5.241	2.436-11.277	<.0001

Table 3. Multivariate analyses of disease-free survival and overall survival for positive (+) or netative(-) expression of LIMK2

DFS, disease-free survival; OS, overall survival. a: T1+T2 was reference group; b: N0 was reference group; c: I+II was reference group; d: I was reference group; e: ER- was reference group; f: PR- was reference group; g: HER2- was reference group; h: LIMK2- was reference group.



Figure 3. LIMK2 mRNA expression pattern and prognostic value in public databases. (A) LIMK2 mRNA expression levels were significantly increased in breast cancer tissues compared with normal in the METABRIC database. Kaplan-Meier survival curves indicated that increased LIMK2 mRNA expression predicted worse relapse-free survival (B) and overall survival (C) in breast cancer patients based on the Kaplan Meier plotter database. ***P*<0.0001.

OS

HR = 1.52 (1.2 - 1.92)

logrank P = 4e-04



80

547

169

100

370

107

120

292

82

60

741

239

Time (months)

Aurora A in breast cancer cells. Moreover, Aurora A and LIMK2 could be engaged in a positive-feedback loop, promoting Aurora-A-mediated oncogenic pathways [11]. In addition, recent report found LIMK2 was involved in the formation of invadopodia, matrix degradation and invasive migration in breast cancer cells. Nevertheless, to the best of our knowledge, the expression pattern and prognostic value of LIMK2 in breast cancer was still not clarified. To certify the biological role of LIMK2 in breast cancer, we collected specimens from 212 patients and correlated the LIMK2 expression with clinicopathological factors and survival of breast cancer patients using Kaplan-Meier analysis and Cox regression analysis. Our results suggested that LIMK2 was an independent poor prognostic factor for breast cancer patients, which was subsequently confirmed by bioinformatics analysis METABRIC and the Kaplan Meier plotter databases.

In conclusion, our data implied that elevated LIMK2 expression was an independent poor prognostic factor for breast cancer patients, which suggested LIMK2 might have a critical role in the progression of breast cancer. Detection of LIMK2 could provide some suggestions for diagnosis, prognostication and personalized treatment for breast cancer.

Acknowledgements

We would like to extend our sincere gratitude and appreciation to Ying Zhang from University of San Francisco, San Francisco, CA ,USA, for language editing and proof-reading.

Disclosure of conflict of interest

None.

Address correspondence to: Jun Jiang and Fan Zhang, Breast Disease Center, Southwest Hospital, Third Military Medical University, Chongqing 400038, China. E-mail: jcbd@medmail.com.cn (JJ); E-mail: zhangfan316@163.com (FZ)

References

- Scott RW and Olson MF. LIM kinases: function, regulation and association with human disease. J Mol Med (Berl) 2007; 85: 555-568.
- [2] Manetti F. Recent findings confirm LIM domain kinases as emerging target candidates for cancer therapy. Curr Cancer Drug Targets 2012; 12: 543-560.

- [3] Manetti F. LIM kinases are attractive targets with many macromolecular partners and only a few small molecule regulators. Med Res Rev 2012; 32: 968-998.
- [4] Okano I, Hiraoka J, Otera H, Nunoue K, Ohashi K, Iwashita S, Hirai M and Mizuno K. Identification and characterization of a novel family of serine/threonine kinases containing two Nterminal LIM motifs. J Biol Chem 1995; 270: 31321-31330.
- [5] Foletta VC, Moussi N, Sarmiere PD, Bamburg JR and Bernard O. LIM kinase 1, a key regulator of actin dynamics, is widely expressed in embryonic and adult tissues. Exp Cell Res 2004; 294: 392-405.
- [6] Honma M, Benitah SA and Watt FM. Role of LIM kinases in normal and psoriatic human epidermis. Mol Biol Cell 2006; 17: 1888-1896.
- [7] Salvarezza SB, Deborde S, Schreiner R, Campagne F, Kessels MM, Qualmann B, Caceres A, Kreitzer G and Rodriguez-Boulan E. LIM kinase 1 and cofilin regulate actin filament population required for dynamin-dependent apical carrier fission from the trans-Golgi network. Mol Biol Cell 2009; 20: 438-451.
- [8] Vlecken DH and Bagowski CP. LIMK1 and LIMK2 are important for metastatic behavior and tumor cell-induced angiogenesis of pancreatic cancer cells. Zebrafish 2009; 6: 433-439.
- [9] Hsu FF, Lin TY, Chen JY and Shieh SY. p53-Mediated transactivation of LIMK2b links actin dynamics to cell cycle checkpoint control. Oncogene 2010; 29: 2864-2876.
- [10] Po'uha ST, Shum MS, Goebel A, Bernard O and Kavallaris M. LIM-kinase 2, a regulator of actin dynamics, is involved in mitotic spindle integrity and sensitivity to microtubule-destabilizing drugs. Oncogene 2010; 29: 597-607.
- [11] Johnson EO, Chang KH, Ghosh S, Venkatesh C, Giger K, Low PS and Shah K. LIMK2 is a crucial regulator and effector of Aurora-A-kinase-mediated malignancy. J Cell Sci 2012; 125: 1204-1216.
- [12] Gamell C, Schofield AV, Suryadinata R, Sarcevic B and Bernard O. LIMK2 mediates resistance to chemotherapeutic drugs in neuroblastoma cells through regulation of drug-induced cell cycle arrest. PLoS One 2013; 8: e72850.
- [13] Lagoutte E, Villeneuve C, Lafanechere L, Wells CM, Jones GE, Chavrier P and Rosse C. LIMK regulates tumor-cell invasion and matrix degradation through tyrosine phosphorylation of MT1-MMP. Sci Rep 2016; 6: 24925.
- [14] Okamoto I, Pirker C, Bilban M, Berger W, Losert D, Marosi C, Haas OA, Wolff K and Pehamberger H. Seven novel and stable translocations associated with oncogenic gene expression in malignant melanoma. Neoplasia 2005; 7: 303-311.

- [15] Davila M, Frost AR, Grizzle WE and Chakrabarti R. LIM kinase 1 is essential for the invasive growth of prostate epithelial cells: implications in prostate cancer. J Biol Chem 2003; 278: 36868-36875.
- [16] Bagheri-Yarmand R, Mazumdar A, Sahin AA and Kumar R. LIM kinase 1 increases tumor metastasis of human breast cancer cells via regulation of the urokinase-type plasminogen activator system. Int J Cancer 2006; 118: 2703-2710.
- [17] Dan S, Tsunoda T, Kitahara O, Yanagawa R, Zembutsu H, Katagiri T, Yamazaki K, Nakamura Y and Yamori T. An integrated database of chemosensitivity to 55 anticancer drugs and gene expression profiles of 39 human cancer cell lines. Cancer Res 2002; 62: 1139-1147.
- [18] Fernandez-Banet J, Esposito A, Coffin S, Horvath IB, Estrella H, Schefzick S, Deng S, Wang K, AChing K, Ding Y, Roberts P, Rejto PA and Kan Z. OASIS: web-based platform for exploring cancer multi-omics data. Nat Methods 2016; 13: 9-10.
- [19] Gyorffy B, Lanczky A, Eklund AC, Denkert C, Budczies J, Li Q and Szallasi Z. An online survival analysis tool to rapidly assess the effect of 22,277 genes on breast cancer prognosis using microarray data of 1,809 patients. Breast Cancer Res Treat 2010; 123: 725-731.
- [20] Petrilli A, Copik A, Posadas M, Chang LS, Welling DB, Giovannini M and Fernandez-Valle C. LIM domain kinases as potential therapeutic targets for neurofibromatosis type 2. Oncogene 2014; 33: 3571-3582.

- [21] Mardilovich K, Baugh M, Crighton D, Kowalczyk D, Gabrielsen M, Munro J, Croft DR, Lourenco F, James D, Kalna G, McGarry L, Rath O, Shanks E, Garnett MJ, McDermott U, Brookfield J, Charles M, Hammonds T and Olson MF. LIM kinase inhibitors disrupt mitotic microtubule organization and impair tumor cell proliferation. Oncotarget 2015; 6: 38469-38486.
- [22] Nishimura Y, Yoshioka K, Bernard O, Himeno M and Itoh K. LIM kinase 1: evidence for a role in the regulation of intracellular vesicle trafficking of lysosomes and endosomes in human breast cancer cells. Eur J Cell Biol 2004; 83: 369-380.
- [23] Nishimura Y, Yoshioka K, Bernard O, Bereczky B and Itoh K. A role of LIM kinase 1/cofilin pathway in regulating endocytic trafficking of EGF receptor in human breast cancer cells. Histochem Cell Biol 2006; 126: 627-638.
- [24] McConnell BV, Koto K and Gutierrez-Hartmann A. Nuclear and cytoplasmic LIMK1 enhances human breast cancer progression. Mol Cancer 2011; 10: 75.
- [25] Croft DR, Crighton D, Samuel MS, Lourenco FC, Munro J, Wood J, Bensaad K, Vousden KH, Sansom OJ, Ryan KM and Olson MF. p53-mediated transcriptional regulation and activation of the actin cytoskeleton regulatory RhoC to LIMK2 signaling pathway promotes cell survival. Cell Res 2011; 21: 666-682.
- [26] Rak R, Haklai R, Elad-Tzfadia G, Wolfson HJ, Carmeli S and Kloog Y. Novel LIMK2 inhibitor blocks panc-1 tumor growth in a mouse xenograft model. Oncoscience 2014; 1: 39-48.