# Review Article Mcl-1 intervention regulating macrophages apoptosis in vivo and in vitro TB model: a systematic review and meta-analysis

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Abstract: Tuberculosis has seriously affected human health, and current situation of prevention and control tuberculosis is becoming more and more urgent. Intervention the expression of Mcl-1 can control latent and persistent TB infection effectively, including virulent and attenuated MTB strains, and has divergent effects between different host macrophages environment and between different intervention times and methods. Herein, a meta-analysis was performed by independently searching databases including the Cochrane Library, PubMed, Springer, Embase, and China National Knowledge Infrastructure, to analyze effects of McI-1 intervention on MTB infection. Compared to controls, MTB infection induced macrophages apoptosis significantly increase in vivo and in vitro macrophages infected with different virulence of MTB strains (P < 0.0002), and short time's infection caused more host macrophages apoptosis in vivo TB model (P < 0.0002). After McI-1 intervention, compared with controls, McI-1 induction rate was significantly increase in H37Ra infection group (95% CI, 1.51, 3.86; Z = 2.69; P < 0.00002), Mcl-1 induction rate was also significantly increase in H37Rv infection group (95% Cl, 9.51, 24.28; Z = 16.91; P < 0.00002), whereas the induction relative weaker in other MTB strains infection group. Short Mcl-1 intervention time significantly increased host macrophages apoptosis infected BCG and H37Rv (P < 0.00001), but the effect was relatively decrease in other MTB strains infection group (P = 0.006), while it have no significantly differences in H37Ra infection group (P = 0.15). However, Long Mcl-1 intervention time increased macrophages apoptosis in all MTB strains (P < 0.00001). These findings may provide a theoretical basis for the interaction between host macrophages and MTB and Mcl-1 intervention introduce to control latent and persistent TB infection.

Keywords: Mcl-1, macrophages apoptosis, systematic review and meta-analysis

#### Introduction

Tuberculosis (TB) is a worldwide infectious diseases caused by Mycobacterium Tuberculosis (MTB). Based on the Bulletin of the World Health Organization (WHO), one-third of the world's population may be asymptomatically infected with tuberculosis (TB) [1]. TB has been a remarkable public health issue in mainland China, and 80% of new TB cases worldwide have been reported in China each year. In 2013, an estimated 9.0 million individuals developed TB and 1.5 million died. China alone accounted for 11% of the total cases worldwide [2]. However, MTB remains to survival within infected macrophages for prolonged periods by evading the elimination of host immune responses [3, 4]. As such, the long-term latent infection and persistent infection in host macrophages becomes the main difficulties of control and prevention TB. In addition, its incidence and the prevalence of Multidrug Resistant-TB (MDR-TB) have increased [5], while our tools to combat MDR-TB are unsatisfactory and not ideal. However, studies have found that inhibit the expression of McI-1 can effectively promote the MTB infection of host macrophage apoptosis, so as to achieve the purpose of persistent infection with the tuberculosis control in recent years. So the research on the basis of the literature retrieval using meta-analysis was carried out on the experimental data published at

First Author (Year)	Language	n	Type of TB model	Type of MTB	Time of Mcl-1 intervention	Type of McI-1 intervention	Outcome Indicators
Wang FY 2016 [7]	English	10	In vivo, in vitro	H37Rv	24 h; 3 d	RNAi	1, 2
Wang 2016 [8]	English	3	In vivo	BCG, H37Ra, H37Rv, XJ-MTB	1, 3, 5, 7 d	Signaling pathway intervention	1, 2
Marriott HM 2005 [9]	English	7	In vivo	Pneumoniae	<24 h	RNAi	1, 2
Sly LM 2003 [10]	English	5	In vivo, in vitro	H37Ra, H37Rv	4, 16, 24, 48 h	RNAi	1, 2
Kumar R 2016 [11]	English	3	In vivo, in vitro	H37Rv	<24 h	RNAi; Signaling pathway intervention	1, 2
Palaga T 2013 [12]	English	3	In vivo, in vitro	BCG	≤24 h	RNAi	1, 2
Wu XL 2014 [13]	English	15	In vitro	BCG	6, 12, 24, 36 h	Signaling pathway intervention	1
Wang FY 2015 [14]	Chinese	10	In vivo	BCG, H37Ra, H37Rv, XJ-MTB	1, 3 ,5, 7 d	RNAi	1, 2
Wu 2014 [15]	Chinese	15	In vitro	BCG	6, 12, 24, 36 h	Signaling pathway intervention	1
Zhang YQ 2015 [16]	Chinese	10	In vivo	H37Rv	5 d	Signaling pathway	1

 Table 1. Characteristics of the studies included in the meta-analysis

Note: n = the number in the experimental group; 1 = Apoptosis rate, 2 = Mcl-1 induction rate.



Figure 1. Flowchart.of search strategy.

home and abroad in recent years, comprehensive analysis, aimed to explore inhibit Mcl-1 expression for MTB infected macrophage apoptosis regulation function, for mycobacterium tuberculosis and the principle of the interaction between the host and provides the reference for the prevention and control of tuberculosis.

# Methods

# Search strategy

Using the PICO principle [6], searches were performed using the following electronic databases: the Cochrane Library, PubMed, Embase, Springer, Web of Science, China Science and Technology Journal Database (CSTJ), and China National Knowledge Infrastructure (CNKI) (last search conducted in July 2017). The key search string was (Mcl-1) AND (MTB) OR (Macrophages) OR (Infection) OR (Apoptosis) OR (TB) OR (Tuberculosis).

# Inclusion criteria

We systematically reviewed published studies according to the following inclusion criteria any animal and cells lines, category not limited, published in either Chinese or English. Mcl-1 intervened by any kind of method in TB model groups, and its compounds were used as the experimental groups, and the untreated served as control groups. If various intervention methods of Mcl-1 were used in the study, the best intervention was chosen for this analysis. If various intervention times of Mcl-1 were used in a study, the longest time was chosen for this analysis.

# Exclusion criteria

The exclusion criteria were as follows: (1) repeat publications; (2) incomplete information; (3) insufficient or insignificant statistical data; (4) unrelated to the study objectives; (5) lack of appropriate controls; and (6) review articles.

# Outcome indicators

Host macrophage apoptosis rate: including apoptosis rate detected by FCM (Flow of cytometry), and TUNEL technology (Terminal deoxynucleotidyl transferase-mediated dUTP nick end labelling) detected apoptosis rate. Mcl-1 induction rate: mainly including Western blot measured result of Mcl-1/ $\beta$ -actin.

# Data collection

Two reviewers (Xiao-fang Wang and Xin-min Wang) independently extracted data and crosschecked their data after aggregating the results. The following information was extracted from the complete manuscripts of each qualified study: publication characteristics (title of the study, first author, publication date, and journal/magazine title), baseline data (mean and standard deviation [SD]) for the experimental and control groups, subject characteristics (source of cells and tissue, arsenic doses and exposure times), outcome indicators, and the source of indicator estimates. This information is presented in **Table 1**. When the two reviewers' opinions differed, Disagreements were resolved by discussion with Professor Le Zhang, was asked to make the final decision regarding the results.

# Data analysis

Ten articles were analyzed in Review Manager Version 5.3 (The Nordic Cochrane Centre, The Cochrane Collaboration, 2012, Portland Oregon, OR, USA). Significant heterogeneity was detected (p < 0.05,  $l^2$ >75%) and a random effects model was therefore applied for the meta-analysis. A multivariate meta-regression analysis was performed to determine the source of heterogeneity and continuous variables were estimated as standardized mean differences (SMDs) with 95% confidence intervals (CIs) between the arsenic treated groups and control groups. All reported p-values are twosided and a significance level of 0.05 was used. For additional insight, subgroup analyses were performed based on TB models (MTB infection in vivo; MTB infection in vitro) and intervention time ( $\leq 24$  h or >24 h), based on the median of the indexes reported in the papers, to determine the factors associated with differences in the outcome indicators across different studies. Small-study effects were explored using funnel plots and Egger's tests and study sensitivities were assessed. Sensitivity analyses were performed using Stata 12.0 (StataCorp, College Station, Texas, TX, USA).

# Results

# Search result

**Figure 1** shows the study selection process, 10 relevant articles were identified. Initially, 162 articles were included in our search strategy. Finally, a total of 12 articles were included in the analysis, based on the inclusion and exclusion criteria. A total of 9 studies assessed Mcl-1 intervention in vitro TB model, 6 studies examined Mcl-1 intervention in vivo TB model (Table 1).

# Study characteristics

Using the search strategy described in Section 2, 162 relevant articles were identified (Figure 1), of which 10 were used for the meta-analysis based on the eligibility and exclusion criteria (Table 1). Various cell lines and animals were used as TB model groups in these studies, and in each study, the effect of Mcl-1 intervention on the MTB infection was assessed. The Mcl-1 intervention groups were primarily cell lines and animals treated with various MTB strains (e.g. BCG, H37Ra, H37Rv, XJ-MTB, Pneumococcus), and the control models were blank controls. Alone MTB infection was categorized as either in vivo TB model (n = 8) or in vitro TB model (n = 6). Mcl-1 intervention time varied among the studies and was categorized as  $\leq 24$ h (n = 9) or >24 h (n = 7), Mcl-1 intervention methods was categorized as RNAi (n = 5) or signaling pathway intervention (n = 5). Mcl-1 induction rate also varied among the studies, and was categorized as either in vivo TB model (n = 7) or in vitro TB model (n = 4).

# Results of the meta-analysis

# The influence of MTB infection alone on host macrophages apoptosis

A total of 8 studies were included in this study, and of these, 8 estimated the impact of alone MTB infection by host macrophages apoptosis. A pooled analysis showed that BCG, H37Ra, H37Rv, and other MTB strains infection were 28.64, 56.24, 42.07, 28.09-fold higher than the control group (BCG group: 95% Cl, 17.04-40.24; Z = 4.84; p < 0.00001; H37Ra group: 95% Cl, 26.25-86.23; Z = 3.68; p = 0.0002; H37Rv group: 95% Cl, 26.71-57.43; Z = 5.37; p

	MTB	infecti	on	C	ontrol			Std. Mean Difference	Std. Mean Difference
Study or Subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, Random, 95% CI	IV, Random, 95% CI
2.1.1 BCG infection in	n vivo m	odel							
Wang 2016	18.6	0.3	10	10.3	0.1	10	9.6%	35.55 [23.22, 47.88]	
Wang FY 2015	18.4	0.3	10	9.7	0.4	10	10.6%	23.57 [15.37, 31.76]	
Subtotal (95% CI)			20			20	20.2%	28.64 [17.04, 40.24]	•
Heterogeneity: Tau <sup>2</sup> =	43.27;	Chi <sup>2</sup> = 2	2.52, df	= 1 (P =	= 0.11)	; <b>I</b> <sup>2</sup> = 60	)%		
Test for overall effect:	Z=4.84	4 (P < 0	.00001	)		-			
2.1.2 H37Ra infection	ı in vivo	model							
Sly LM 2003	29	0.1	5	2	0.1	5	0.4%	243.87 [106.57, 381.17]	· · ·
Wang 2016	17	0.1	10	10.3	0.15	10	8.1%	50.34 [32.91, 67.77]	
Wang FY 2015	18	0.15	10	9.7	0.2	10	8.7%	44.97 [29.39, 60.54]	
Subtotal (95% CI)			25			25	17.2%	56.24 [26.25, 86.23]	
Heterogeneity: Tau <sup>2</sup> =	418.83	; Chi²=	8.02, 0	if = 2 (P	= 0.02	?); I² = 7	75%		
Test for overall effect:	Z = 3.68	8 (P = 0	.0002)						
2.1.3 H37Rv infection	in vivo	model							
Sly LM 2003	21	0.3	5	2	0.1	5	3.2%	76.75 [33.52, 119.97]	
Wang 2016	16.2	0.3	10	10.3	0.15	10	10.6%	23.83 [15.54, 32.11]	
Wang FY 2015	15.4	0.2	10	9.7	0.2	10	10.3%	27.30 [17.82, 36.78]	
Wang FY 2016	17.3	0.1	10	8.9	0.2	10	8.1%	50.88 [33.26, 68.50]	
Zhang YQ 2015	25.1	0.2	10	12	0.2	10	7.0%	62.73 [41.02, 84.44]	
Subtotal (95% CI)			45			45	39.1%	42.07 [26.71, 57.43]	-
Heterogeneity: Tau <sup>2</sup> = 214.56; Chi <sup>2</sup> = 20.85, df = 4 (P = 0.0003); l <sup>2</sup> = 81%									
Test for overall effect:	Z = 5.37	? (P < 0	.00001	)					
2.4.4 Other MTD info	otion in	uiuo m	adal						
2.1.4 Other MTB Inte	cuon in		Daei		~ 4	-	4.000	4.50.04.000.07.004.000	
Warriott HW 2005	28	0.2	10	40.2	0.1	10	1.0%	103.94 [80.07, 221.22]	
Wang 2016	15.8	0.3	10	10.3	0.15	10	10.7%	22.21 [14.48, 29.94]	
Wang FY 2015	14.1	0.3	10	9.7	0.2	10	11.1%	16.53 [10.75, 22.31]	
Subtotal (95% CI)			21			21	23.4%	28.09 [9.07, 40.51]	
Heterogeneity: Tau* =	176.91	Chir=	16.81,	df = 2 (	P = U.U	1002);1	r= 88%		
l est for overall effect:	Z = 2.99	9 (P = 0	.003)						
Total (95% CI)			117			117	100.0%	37.78 [28.77, 46.78]	•
Heterogeneity: Tau <sup>2</sup> =	180,90	: Chi <sup>z</sup> =	74.84	df = 12	(P < 0	00001	);   <sup>2</sup> = 849	6	
Test for overall effect	Z = 8.22	2 (P < 0	.00001	)			//· •//	-	-100 -50 0 50 100
Test for subgroup diff	erences	:Chi <sup>2</sup> =	4.39	, df = 3 (F	e = 0.2	2), <b> </b> ² =	31.6%		Favours [experimental] Favours [control]
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**Figure 2.** Effects of alone MTB infection in vivo TB model. Forest plot showing the impact of McI-1 intervention by different methods compared with controls. Abbreviations: SMD = standardized mean difference, IV = independent variable, 95% CI = 95% confidence interval.



**Figure 3.** Effects of alone MTB infection in vitro TB model. Forest plot showing the impact of McI-1 intervention by different methods compared with controls. Abbreviations: SMD = standardized mean difference, IV = independent variable, 95% CI = 95% confidence interval.

< 0.00001; Other MTB strains group: 95% Cl, 9.67-46.51; Z = 2.99; p = 0.003) in vivo TB

model, while only BCG group with no significant heterogeneity (p = 0.11;  $l^2 = 60\%$ ; Figure 2).

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Figure 4. Effects of alone MTB infection in vivo and in vitro TB model. Forest plot showing the impact of Mcl-1 intervention by different methods compared with controls. Abbreviations: SMD = standardized mean difference, IV = independent variable, 95% CI = 95% confidence interval.

Table 2. The apoptosis rate comparison of different models in MTB infection

Index			In Viv	/o TB mo	del	In Vitro TB model						
	N	n	р	SMD	95% CI	N	n	р	SMD	95% CI		
≤24 h	5	36	0.0002	37.1	13.84, 44.86	5	43	0.0002	56.79	26.59, 86.99		
>24 h	5	38	<0.0001	47.95	26.06, 69.85	1	5	0.0006	15.94	6.88, 25.00		
BCG group	2	20	0.005	28.64	17.04, 40.24	2	30	<0.0001	60.56	35.36, 85.75		
H37Ra group	3	25	0.0002	56.24	26.25, 86.23	1	5	0.0005	85.81	37.48, 134.13		
H37Rv group	5	45	<0.00001	42.07	26.71, 57.43	2	8	<0.0001	20.76	10.58, 30.94		
Other MTB group	3	27	0.003	28.09	9.67, 46.51	0	0	0	0	0, 0		

N = the number of documents; n = the number of samples. 95% CI = 95% confidence interval. SMD = standardized mean difference.

Pooled analysis showed that there were 60.56, 85.81, 20.76-fold higher than the control group (BCG group: 95% Cl, 35.86-85.75; Z = 4.71; p < 0.00001; H37Ra group: 95% Cl, 37.48-134.13; Z = 3.48; p = 0.0005; H37Rv group: 95% Cl, 10.58-30.94; Z = 4.0; p < 0.00001) in vitro TB model, but only BCG group with significant heterogeneity (p = 0.04;  $l^2 = 77\%$ ; Figure 3).

In subgroup analyses, the analysis based on the source of macrophages (in vivo TB model vs. in vitro TB model) and MTB infection time

	MTB	infecti	ion	С	ontrol			Std. Mean Difference	Std. Mean Difference
Study or Subgroup	Mean	SD	Total	Mean	SD	Total	Weight	IV, Random, 95% CI	IV, Random, 95% Cl
1.1.1 Mcl-1 induction	in BCG	infecti	on						
Wang 2016	1.2	0.15	3	1	0.01	3	10.5%	1.51 [-0.66, 3.67]	
Wang FY 2015	1.15	0.2	10	1.1	0.01	10	11.6%	0.34 [-0.55, 1.22]	•
Wu XL 2014	1.35	0.2	15	1.02	0.1	10	11.5%	1.90 [0.91, 2.88]	
Subtotal (95% CI)			28			23	33.6%	1.17 [0.01, 2.33]	
Heterogeneity: Tau <sup>2</sup> =	0.64; C	hi² = 5.	51, df=	= 2 (P =	0.06);	l <sup>2</sup> = 649	6		
Test for overall effect:	Z = 1.98	P = 0	.05)						
1.1.2 Mcl-1 induction	in H37R	ta infe	ction						
Wang 2016	1.35	0.15	3	1	0.01	3	9.5%	2.63 [-0.37, 5.64]	-
Wang FY 2015	1.2	0.1	10	1	0.01	10	11.3%	2.70 [1.42, 3.98]	
Subtotal (95% CI)			13			13	20.8%	2.69 [1.51, 3.86]	1
Heterogeneity: Tau <sup>2</sup> =	0.00; C	hi² = 0.	00, df=	= 1 (P =	0.97);	l² = 0%			
Test for overall effect:	Z= 4.47	' (P < 0	.00001	)					
1.1.3 Mcl-1 induction	in H37R	tv infec	ction						
Kumar R 2016	5.9	0.2	3	1	0.1	3	0.7%	24.79 [0.80, 48.79]	
Sly LM 2003	4.8	0.2	5	3	1	1	4.8%	7.20 [-0.08, 14.48]	-
Wang 2016	3.85	0.15	3	1.1	0.01	10	1.2%	39.60 [21.32, 57.88]	
Wang FY 2015	3.8	0.16	10	1.1	0.05	10	4.5%	21.82 [14.22, 29.41]	
Wang FY 2016	3.88	0.2	10	1.2	0.02	10	5.6%	18.06 [11.75, 24.37]	
Wang FY 2016	2.68	0.22	10	1.2	0.1	10	9.5%	8.30 [5.30, 11.29]	· · ·
Subtotal (95% CI)			41			44	26.3%	16.91 [9.54, 24.28]	◆
Heterogeneity: Tau <sup>2</sup> =	57.74; (	Chi <b>²</b> = 3	27.22.	df = 5 (P	< 0.0	001); P	= 82%		
Test for overall effect:	Z = 4.50	) (P < 0	.00001	)					
1.1.4 McI-1 induction	in other	r MTB i	nfectio	n					
Marriott HM 2005	1.3	0.2	7	1.15	0.01	7	11.4%	0.99 [-0.14, 2.13]	t
Wang 2016	4.6	0.15	3	1.1	0.05	3	0.7%	25.04 [0.81, 49.28]	
Wang FY 2015	4.2	0.3	10	1.1	0.06	10	7.2%	13.72 [8.90, 18.55]	+
Subtotal (95% CI)			20			20	19.3%	9.90 [-1.91, 21.72]	◆
Heterogeneity: Tau <sup>2</sup> =	82.79; (	Chi² = 3	28.91,	df = 2 (P	< 0.0	0001); P	<b>²</b> = 93%		
Test for overall effect:	Z=1.64	(P = 0	.10)						
Total (95% CI)			102			100	100.0%	6.01 [3.94, 8.07]	
Heterogeneity: Tau² =	9.35; C	hi² = 10	33.82, (	df = 13 (	P < 0.0	00001);	l² = 90%		
Test for overall effect:	Z= 5.71	(P < 0	.00001	)					Eavours [experimental] Eavours [control]
Test for subgroup diff	erences	: Chi² :	= 20.60	. df = 3	(P = 0.	0001).	I <sup>2</sup> = 85.49	6	r avours (experimental) - r avours (control)

**Figure 5.** Effects of Mcl-1 induction by different MTB. Forest plot showing the impact of Mcl-1 intervention by different methods compared with controls. Abbreviations: SMD = standardized mean difference, IV = independent variable, 95% CI = 95% confidence interval.

(>24 h vs.  $\leq$ 24 h) was conducted. The analysis demonstrated that short infection time ( $\leq$ 24 h) promote macrophages apoptosis in vitro TB model (95% Cl, 26.59-86.99; Z = 3.69; *p* = 0.0002), and there were 56.79-fold higher than the control group, with significant heterogeneity (*p* < 0.00001; l<sup>2</sup> = 92%; **Figure 4; Table 2**). Long infection time (>24 h) were 47.95-fold higher than the control group (95% Cl, 26.06-69.85; Z = 4.29; *p* < 0.00002) in vivo TB model, with significant heterogeneity (*p* = 0.0005; l<sup>2</sup> = 80%; **Figure 4; Table 2**).

#### The effects of Mcl-1 intervention on host macrophages apoptosis infected MTB

Considering special role of Mcl-1 in MTB infection, firstly, we pooled analyzed Mcl-1 induction rate in alone MTB infection. The meta-analysis results showed that H37Rv infection induced 16.91-fold Mcl-1 expression higher than the control group (BCG group: 95% Cl, 9.54-24.28; Z = 4.5; p < 0.00001), and with significant heterogeneity (p < 0.0001;  $l^2 = 82\%$ ; Figure 5).

However, BCG, H37Ra, and other MTB strains infection either induced less Mcl-1 expression compared with H37Rv infection group. The next analyses were the effects of Mcl-1 intervention.

The subgroup analysis based on Mcl-1 intervention time (>24 h vs. ≤24 h) and Mcl-1 intervention method (RNAi vs. Signaling pathway intervention) was conducted. The analysis showed that Mcl-1 intervention in short BCG, H37Rv, and other MTB strains infection time (≤24 h) caused 26.84, 38.11, 46.49-fold host macrophages apoptosis higher than the control group (BCG group: 95% CI, 17.37-36.11; Z = 5.59; *p* < 0.00001; H37Ra group: There was no significant difference; p = 0.15; H37Rv group: 95% CI, 24.61-51.62; Z = 5.53; p < 0.00001; Other MTB strains group: 95% CI, 13.27-79.72; Z = 2.74; p = 0.003), while only H37Rv group with no significant heterogeneity (p = 0.16;  $I^2 =$ 38%; Figure 6; Table 3). Mcl-1 intervention in long BCG, H37Ra, H37Rv, and other MTB

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11.1 Mcl-1 intervention in BCG infection(24h and 24h-)         Palaga T 2013       56       0.3       3       26       0.2       3       1.2%       94.14 [3.22, 185.05]         Wang 2016       29.9       0.4       3       17.5       0.3       3       5.7%       28.06 [0.2, 55.20]         Wang FY 2015       33.4       0.3       10       17.4       0.4       10       7.7%       43.34 [28.33, 58.36]         Wu 2014       52       0.8       15       35       0.6       15       8.8%       23.39 [17.00, 29.78]         Wu XL 2014       41.7       0.5       15       32.2       0.5       15       8.9%       18.49 [13.42, 23.56]         Subtotal (95% Cl)       46       46       32.3%       26.74 [17.37, 36.11]         Heterogeneity: Tau <sup>2</sup> = 56.75; Chi <sup>2</sup> = 12.39, df = 4 (P = 0.01); I <sup>2</sup> = 68%       20.22 [0.34, 3.70]         Yang FY 2015       29.3       0.2       10       16.8       0.1       3       5.2%       31.38 [1.04, 61.72]         Wang FY 2015       29.3       0.2       10       16.8       0.1       3       5.2%       35.21 [-12.39, 82.81]         Heterogeneity: Tau <sup>2</sup> = 1635.51; Chi <sup>2</sup> = 33.78, df = 2 (P < 0.00001); I <sup>2</sup> = 94%       35.21 [-12.39, 82.81]       46.63 [
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Subtotal (95% Cl)       46       46       32.3%       26.74 [17.37, 36.11]         Heterogeneity: Tau <sup>2</sup> = 56.75; Chi <sup>2</sup> = 12.39, df = 4 (P = 0.01); I <sup>2</sup> = 68%       Test for overall effect: Z = 5.59 (P < 0.00001)
Heterogeneily: Tau <sup>2</sup> = 56.75; Chi <sup>2</sup> = 12.39, df = 4 (P = 0.01); I <sup>2</sup> = 68% Test for overall effect: $Z = 5.59$ (P < 0.00001) 11.1.2 McI-1 intervention in H37Ra infection(24h and 24h-) Sly LM 2003 18.5 0.1 5 18 0.3 5 9.1% 2.02 [0.34, 3.70] Wang 2016 21.8 0.15 3 16.8 0.1 3 5.2% 31.38 [1.04, 61.72] Wang FY 2015 29.3 0.2 10 16.8 0.1 10 5.9% 75.72 [49.52, 101.92] Subtotal (95% Cl) 18 18 20.2% 35.21 [-12.39, 82.81] Heterogeneily: Tau <sup>2</sup> = 1635.51; Chi <sup>2</sup> = 33.78, df = 2 (P < 0.00001); I <sup>2</sup> = 94% Test for overall effect: $Z = 1.45$ (P = 0.15) 11.1.3 McI-1 intervention in H37Rv infection(24h and 24h-) Sly LM 2003 37 0.1 5 21 0.3 5 4.4% 64.63 [28.22, 101.04]
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Wang 2016       21.8       0.15       3       16.8       0.1       3       5.2%       31.38 [1.04, 61.72]         Wang FY 2015       29.3       0.2       10       16.8       0.1       10       5.9%       75.72 [49.52, 101.92]         Subtotal (95% Cl)       18       18       20.2%       35.21 [-12.39, 82.81]         Heterogeneity: Tau <sup>2</sup> = 1635.51; Chi <sup>2</sup> = 33.78, df = 2 (P < 0.00001); P <sup>2</sup> = 94%       75.72 [49.52, 101.92]         Test for overall effect: Z = 1.45 (P = 0.15)       11.1.3 Mcl-1 intervention in H37Rv infection(24h and 24h-)         Sly LM 2003       37       0.1       5       21       0.3       5       4.4%       64.63 [28.22, 101.04]
Wang FY 2015       29.3       0.2       10       16.8       0.1       10       5.9%       75.72 [49.52, 101.92]         Subtotal (95% Cl)       18       18       20.2%       35.21 [-12.39, 82.81]         Heterogeneity: Tau <sup>2</sup> = 1635.51; Chi <sup>2</sup> = 33.78, df = 2 (P < 0.00001); I <sup>2</sup> = 94%       Test for overall effect: Z = 1.45 (P = 0.15)         11.1.3 Mcl-1 intervention in H37Rv infection(24h and 24h-)       Sly LM 2003       37       0.1       5       21       0.3       5       4.4%       64.63 [28.22, 101.04]
Subtotal (95% Cl)       18       18       20.2%       35.21 [-12.39, 82.81]         Heterogeneity: Tau <sup>2</sup> = 1635.51; Chi <sup>2</sup> = 33.78, df = 2 (P < 0.00001); I <sup>2</sup> = 94%       Test for overall effect: Z = 1.45 (P = 0.15)         11.1.3 Mcl-1 intervention in H37Rv infection(24h and 24h-)       Sly LM 2003       37       0.1       5       21       0.3       5       4.4%       64.63 [28.22, 101.04]
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11.1.3 Mcl-1 intervention in H37Rv infection(24h and 24h-)           Sly LM 2003         37         0.1         5         21         0.3         5         4.4%         64.63 [28.22, 101.04]
11.1.3 Mcl-1 intervention in H37Rv infection(24h and 24h-)           Sly LM 2003         37         0.1         5         21         0.3         5         4.4%         64.63 [28.22, 101.04]
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Sly LM 2003 28 0.2 5 12 0.3 5 5.0% 56.68 [24.75, 88.62]
Wang 2016 23.7 0.4 3 15.4 0.2 3 6.8% 21.00 [0.66, 41.34]
Wang FY 2015 24.5 0.3 10 14.1 0.2 10 7.9% 39.07 [25.53, 52.61]
Wang FY 2016 25.1 0.3 3 14.4 0.3 3 5.6% 28.53 [0.93 56.13]
Subtotal (95% Cl) 26 26 29.8% 38.11 [24.61, 51.62]
Heterogeneity: Tau <sup>2</sup> = 88.40; Chi <sup>2</sup> = 6.50, df = 4 (P = 0.16); i <sup>2</sup> = 38%
Test for overall effect; Z = 5.53 (P < 0.00001)
11.1.4 McI-1 intervention in other MTB infection(24h and 24h-)
Siy LM 2003 28 0.2 7 2 0.2 7 2.8% 121.70 [68.51, 174.89]
Wang 2016 19.8 0.15 3 14.7 0.2 3 6.5% 23.08 [0.74, 45.42]
Wang FY 2015 24.4 0.4 10 13.3 0.3 10 8.4% 30.07 [19.63, 40.50]
Subtotal (95% CI) 20 20 17.6% 46.49 [13.27, 79.72]
Heterogeneity: Tau <sup>2</sup> = 649.56; Chi <sup>2</sup> = 11.66, df = 2 (P = 0.003); l <sup>2</sup> = 83%
Test for overall effect: Z = 2.74 (P = 0.006)
Total (95% Cl) 110 100.0% 35.28 [24.63, 45.93]
Heterogeneity: Tau <sup>2</sup> = 324.68; Chi <sup>2</sup> = 219.17, df = 15 (P < 0.00001); l <sup>2</sup> = 93%
Test for overall effect: Z = 6.49 (P < 0.00001) -100 -50 100
Test for subgroup differences: Chi <sup>2</sup> = 2.72. df = 3 (P = 0.44). I <sup>2</sup> = 0%

**Figure 6.** Effects of McI-1 was interfered within 24 h. Forest plot showing the impact of McI-1 intervention by different methods compared with controls. Abbreviations: SMD = standardized mean difference, IV = independent variable, 95% CI = 95% confidence interval.

Index			≤2	4 h			>24 h					
		n	р	SMD	95% CI	Ν	n	р	SMD	95% CI		
BCG treatment group	5	46	<0.00001	26.74	17.37, 36.11	4	43	<0.00001	30.46	18.06, 42.85		
H37Ra treatment group	3	18	0.15	35.21	-13.39, 82.81	3	16	<0.00001	61.96	42.5, 81.42		
H37Rv treatment group	6	26	<0.00001	38.11	24.61, 51.62	4	28	<0.00001	46.37	27.55, 65.19		
Other MTB treatment group	3	20	0.006	46.49	13.27, 79.72	2	13	<0.00001	40.11	27.01, 53.21		
Total effect	17	110	<0.00001	35.28	24.63, 45.93	13	100	<0.00001	40.13	30.63, 49.64		

	Table 3. T	he compai	rison of	different	Mcl-1	intervention	time
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N = the number of documents; n = the number of samples.95% CI = 95% confidence interval. SMD = standardized mean difference.

strains infection time (>24 h) caused 30.46, 61.96, 46.37, 40.11-fold higher than the control group (BCG group: 95% Cl, 18.06-42.85; Z = 4.82; p < 0.00001; H37Ra group: 95% Cl, 42.50-81.42; Z = 6.24; p < 0.00001; H37Rv group: 95% Cl, 27.55-65.19; Z = 4.83; p < 0.00001; Other MTB strains group: 95% Cl, 27.01-53.21; Z = 2.74; p < 0.00001) in vitro TB model, but BCG and H37Rv group with no significant heterogeneity (H37Ra group: p = 0.51;  $l^2 = 0\%$ ; Other MTB strains group: p = 0.79;  $l^2 =$  0%; **Figure 7**; **Table 3**). Obviously, the effect of signaling pathway intervention method was weak than RNAi, and RNAi included in every studies that we chose articles, while take into account the intersection of the two, so we didn't have any in-depth analysis.

#### Sensitivity analysis

As H37Rv infection for example, we conducted a sensitivity analysis for the TB model, Mcl-1



**Figure 7.** Effects of McI-1 was interfered after 24 h. Forest plot showing the impact of McI-1 intervention by different methods compared with controls. Abbreviations: SMD = standardized mean difference, IV = independent variable, 95% CI = 95% confidence interval.

intervention time, and Mcl-1 induction rate of H37Rv infection group, respectively (see Appendix 1). All of the included studies were distributed evenly from the central line, with no significant deviation. Therefore, no individual study affected the pooled effect results.

#### Discussion

Multidrug-resistant MTB in recent years as the popularity of mixed infection with HIV, TB control situation is more serious, tuberculosis (TB) has become a threat to people's life and health of all infectious disease leading killer [17, 18]. However, the detail interaction mechanism of Mycobacterium tuberculosis (MTB) with host macrophages is unclear. In our study found that MTB infection in a short time will cause more macrophages apoptosis in vitro TB model, and BCG induced host macrophages apoptosis was higher than other MTB strains in vitro TB model. While long time MTB infection cause more host macrophage apoptosis in vivo TB model, H37Ra and H37Rv infection were higher than other MTB strains in vivo TB model. It explained the consequences of mycobacterium tuberculosis infection and TB occurs or not is closely related to the host macrophages environment and infection time. These results will provide more theoretical basis for elucidating the mechanism of MTB interaction with host macrophages and prevention and control of tuberculosis.

Mcl-1 (Myeloid cell leukelmia-1) gene belongs to the members of the family of the Bcl-2, Mcl-1 protein by combined with promoting apoptosis proteins Bax delay cells apoptosis, increase cell survival time to play the role of resistant to apoptosis [19, 20], Mcl-1 is the key factor of the upstream regulation of apoptosis. Researchers suggest that Mcl-1 is a new target for control and prevention Tuberculosis [7-16]. The present study found that H37Rv infected host macrophages can induce higher expression of Mcl-1, and the role of other MTB strains are relatively weak. This may be the reported that the expression level of Mcl-1 is closely related to the MTB strains removal within the host macrophages [9]. While to Mcl-1 is the anti-apoptotic protein, so the expression level of Mcl-1 will fluctuate with the level of apoptosis. That indicates that by inhibiting Mcl-1 expression to promote host macrophage apoptosis infected MTB is a possible treatment for tuberculosis disease. These results provide a theoretical basis for the introduction of Mcl-1 to the treatment of latent infection of tuberculosis.

Mcl-1 are expressed in a variety of malignant tumor cells, high Mc1-1 expression not only inhibits tumor cell apoptosis, also increased the chemotherapy drug resistance [9, 21], reduce the Mcl-1 expression can lead to cell cycle arrest and apoptosis increase [21-23]. Mcl-1 lower expression can enhance the hypoxia induced by lung cancer cell apoptosis [23, 24]. Mcl-1 against Fas mediated apoptosis plays an important role in melanoma, the application of RNAi technology cut the Mcl-1 expression can increase cell apoptosis in the melanoma cells [24]. In recent years, studies have shown that inhibit Mcl-1 expression can effectively promote the host macrophage apoptosis, to protect against a latent MTB infection [7-16]. The study found that intervention Mcl-1 expression can significantly increase the MTB infected host macrophage apoptosis, but the effects of Mcl-1 intervention is closely related to MTB infection time. MTB infection for a short period of time, the effects of Mcl-1 intervention induced host macrophages apoptosis was weak in BCG, H37Ra, H37Rv infection group compared with other MTB strains group. However, as the time of infection increases, which host macrophage apoptosis rate were increased significantly, especially H37Ra and H37Rv infection group. These results suggest that the cell apoptosis pathways may be activated as the infection time increases. Considering that the function of the Mcl-1 is mainly involved in maintaining the stability of the mitochondrial membrane, inhibit the release of Cytochrome-c, so as to promote cell survival, prevent cell apoptosis [25]. So we speculate that intervention Mcl-1 expression for a long time, the stability of the mitochondrial membrane are destroyed, resulting in the extrinsic apoptosis pathways or the intrinsic apoptotic pathways was activated. so that the host macrophage apoptosis rate increased significantly, thus, latent infection and persistent infection of tuberculosis are controlled. These results indicate that the removal of mycobacterium tuberculosis within the host macrophages is closely related to the time of Mcl-1 intervention. Therefore, the effective use of the regulatory function of Mcl-1 intervention in mycobacterium tuberculosis infected host macrophage apoptosis might provide new ideas and targets for TB prevention and control, to speed up the pace of people against TB.

Of course, McI-1 may play double regulatory role or have no regulatory role on the same host macrophage in different intervention methods. Whereas the amount of studies we included this section is limited, hence the meta-analysis results did not show this inference. However, it should be noted that in the treatment of tuberculosis (TB) with McI-1 intervention, the dosage of intervention treatment, the intervention way and time is particularly important, an inappropriate choice may lead to an increase in the degree of infections.

To sum up, the study found that the consequences of mycobacterium tuberculosis infection and TB occurs or not is closely related to the host macrophages environment and infection time. By inhibiting Mcl-1 expression to promote host macrophage apoptosis infected MTB is a potential treatment for tuberculosis disease, but the removal of mycobacterium tuberculosis within the host macrophages is closely related to the time of McI-1 intervention. These results provide a theoretical basis in the Mcl-1 intervention treatment of latent tuberculosis infection and persistent infection and prevention and control Tuberculosis for our future. However, current study concluded articles exists heterogeneity, in addition to the associated with the factors of subgroup analysis, may be also related to the researchers choose the type of object of study, the type of MTB, and many other factors, but none of the existing studies on these factors to do a detailed description or part description only 1 paper, unable to conduct subgroup analysis, remains to be further research in the future.

# Conclusion

As described in the present study results demonstrate that inhibiting McI-1 expression to promote host macrophage apoptosis infected MTB is a potential treatment for tuberculosis disease, but the removal of mycobacterium tuberculosis within the host macrophages is closely related to the time of Mcl-1 intervention. These findings contribute to the latent infection and persistent infection of prevention and control Tuberculosis, and provide a theoretical basis for the implementation of new TB control strategy.

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#### Disclosure of conflict of interest

None.

# Abbreviations

HIV, Human immuno-deficiency virus; MDR, Mu-Iti drug resistant; MDR-MTB, Multidrug resistant Mycobacterium tuberculosis; MDR-TB, Mu-Itidrug resistant tuberculosis; MTB, Mycobacterium tuberculosis.

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#### References

- [1] Maher D, Dye C, Floyd K, Pantoja A, Lonnroth K, Reid A, Nathanson E, Pennas T, Fruth U, Cunningham J, Ignatius H, Raviglione MC, Koek I, Espinal M. Planning to improve global health: The next decade of tuberculosis control. Bull World Health Organ 2007; 85: 341-347.
- [2] Chatterjee A, Saranath D, Bhatter P, Mistry N. Global transcriptional profiling of longitudinal clinical isolates of mycobacterium tuberculosis exhibiting rapid accumulation of drug resistance. PLoS One 2013; 8: e54717.
- Holvast A, de Haan A, van Assen S, Stegeman CA, Huitema MG, Huckriede A, Benne CA, Westra J, Palache A, Wilschut J, Kallenberg CG, Bijl M. Cell-mediated immune responses to influenza vaccination in wegener's granulomatosis. Ann Rheum Dis 2010; 69: 924-927.

- [4] Dorhoi A, Kaufmann SH. Perspectives on host adaptation in response to mycobacterium tuberculosis: Modulation of inflammation. Semin Immunol 2014; 26: 533-542.
- [5] Gandhi NR, Nunn P, Dheda K, Schaaf HS, Zignol M, Soolingen DV, Jensen P, Bayona J. Multidrug-resistant and extensively drug-resistant tuberculosis: a threat to global control of tuberculosis. Lancet 2010; 375: 1830-1843.
- [6] Richardson WS, Wilson MC, Nishikawa J, Hayward RS. The well-built clinical question: a key to evidence-based decisions. ACP J Club 1995; 123: A12-3.
- [7] Wang FY, Zhang YQ, Wang XM, Wang C, Wang XF, Wu JD, Wu F, Zhang WJ, Zhang L. A small hairpin RNA targeting myeloid cell leukemia-1 enhances apoptosis in host macrophages infected with mycobacterium tuberculosis. J Microbiol 2016; 54: 330-337.
- [8] Wang FY, Wang XM, Wang C, Wang XF, Zhang YQ, Wu JD, Wu F, Zhang WJ, Zhang L. Suppression of mcl-1 induces apoptosis in mouse peritoneal macrophages infected with mycobacterium tuberculosis. Microbiol Immunol 2016; 60: 215.
- [9] Marriott HM, Bingle CD, Read RC, Braley KE, Kroemer G, Hellewell PG, Craig RW, Whyte MK, Dockrell DH. Dynamic changes in mcl-1 expression regulate macrophage viability or commitment to apoptosis during bacterial clearance. J Clin Invest 2005; 115: 359-368.
- [10] Sly LM, Hingley-Wilson SM, Reiner NE, Mcmaster WR. Survival of mycobacterium tuberculosis in host macrophages involves resistance to apoptosis dependent upon induction of antiapoptotic bcl-2 family member mcl-1. J Immunol 2003; 170: 430-437.
- [11] Kumar R, Sahu SK, Kumar M, Jana K, Gupta P, Gupta UD, Kundu M, Basu J. Microrna 17-5p regulates autophagy in mycobacterium tuberculosis-infected macrophages by targeting mcl-1 and stat3. Cell Microbiol 2016; 18: 679-91.
- [12] Palaga T, Ratanabunyong S, Pattarakankul T, Sangphech N, Wongchana W, Hadae Y, Kueanjinda P. Notch signaling regulates expression of mcl-1 and apoptosis in ppd-treated macrophages. Cell Mol Immunol 2013; 10: 444-452.
- [13] Wu X, Deng G, Hao X, Li Y, Zeng J, Ma C, He Y, Liu X, Wang Y. A caspase-dependent pathway is involved in wnt/β-catenin signaling promoted apoptosis in bacillus calmette-guerin infected raw264.7 macrophages. Int J Mol Sci 2014; 15: 5045-5062.
- [14] Wang FY, Wang XM, Wang C, Wang XF, Zhang YQ, Jiang-Dong WU, Fang WU, Zhang WJ, Zhang L, Pathophysiology DO. Effects of mcl-1 silencing on apoptosis of mouse peritoneal macrophages infected with different virulence of my-

cobacterium tuberculosis. Chinese Journal of Pathophysiology 2015; 31: 2195-2201.

- [15] Wu XL, Yang YJ. An immunoregulatory role of Wnt/β-catenin signaling in macrophage in response to Mycobacterium tuberculosis infection [D]. Ninxia University 2014.
- [16] Zhang YQ, Wang XM, Wang C, Wang FY, Zhao J, Wu F, Wu JD, Ji R, Zhang WJ, Zhang L. Ffect of Mcl-1 signaling pathway blockers on apoptosis of mouse macrophages infected with Mycobacterium tuberculosis H37Rv. Chinese Journal of Pathophysiology 2015; 31: 2059-2064.
- [17] Bruchfeld J, Correianeves M, Källenius G. Tuberculosis and hiv coinfection. Cold Spring Harb Perspect Med 2015; 5: a017871.
- [18] Dheda K, Rd BC, Maartens G. Tuberculosis. Lancet 2016; 387: 1211-1226.
- [19] Korkmaz D, Bastu E, Dural O, Yasa C, Yavuz E, Buyru F. Apoptosis through regulation of bcl-2, bax and mcl-1 expressions in endometriotic cyst lesions and the endometrium of women with moderate to severe endometriosis. J Obstet Gynaecol 2013; 33: 725-728.
- [20] Kodama T, Hikita H, Kawaguchi T, Shigekawa M, Shimizu S, Hayashi Y, Li W, Miyagi T, Hosui A, Tatsumi T. Mcl-1 and bcl-xl regulate bak/ bax-dependent apoptosis of the megakaryocytic lineage at multistages. Cell Death Differ 2012; 19: 1856.
- [21] Quinn BA, Dash R, Azab B, Sarkar S, Das SK, Kumar S, Oyesanya RA, Dasgupta S, Dent P, Grant S, Rahmani M, Curiel DT, Dmitriev I, Hedvat M, Wei J, Wu B, Stebbins JL, Reed JC, Pellecchia M, Sarkar D, Fisher PB. Targeting mcl-1 for the therapy of cancer. Expert Opin Investig Drugs 2011; 20: 1397-1411.

- [22] Yancey D, Nelson KC, Baiz D, Hassan S, Flores A, Pullikuth A, Karpova Y, Axanova L, Moore V, Sui G, Kulik G. Bad dephosphorylation and decreased expression of mcl-1 induce rapid apoptosis in prostate cancer cells. PLoS One 2013; 8: e74561.
- [23] Yi L, Ji XX, Tan H, Feng MY, Tang Y, Wen L, Su Q. Involvement of mcl1 in diallyl disulfide-induced g2/m cell cycle arrest in hl-60 cells. Oncol Rep 2012; 27: 1911-1917.
- [24] Harrison LR, Micha D, Brandenburg M, Simpson KL, Morrow CJ, Denneny O, Hodgkinson C, Yunus Z, Dempsey C, Roberts D, Blackhall F, Makin G, Dive C. Hypoxic human cancer cells are sensitized to bh-3 mimetic-induced apoptosis via downregulation of the bcl-2 protein mcl-1. J Clin Invest 2011; 121: 1075-87.
- [25] Chetoui N, Sylla K, Gagnonhoude JV, Alcaideloridan C, Charron D, Aldaccak R, Aoudjit F. Down-regulation of mcl-1 by small interfering rna sensitizes resistant melanoma cells to fasmediated apoptosis. Mol Cancer Res 2008; 6: 42.