

## Original Article

# Expression of peroxisome proliferator-activated receptor (PPAR) family and survival analysis in patients with multiple myeloma using microarray gene expression profiles

Aikebaier Maimaiti<sup>1</sup>, Yong-Qin Zhao<sup>1</sup>, Zhang Ting<sup>1</sup>, Wei Huang<sup>2</sup>, Linsen Shi<sup>1</sup>

<sup>1</sup>Zhejiang Chinese Medical University, Hangzhou 311402, Zhejiang, China; <sup>2</sup>Division of Hematologic Malignancies and Cellular Therapy, Duke University, Durham, USA

Received June 12, 2017; Accepted December 7, 2017; Epub April 15, 2019; Published April 30, 2019

**Abstract:** Peroxisome proliferator-activated receptor (PPAR) family consists of 3 members: alpha, gamma, and delta (beta). PPARs play essential roles in cellular differentiation, development, and metabolism. However, little is known about the impact of PPARs on myeloma progression. The aim of this study is to systematically investigate the expression and survival rate of PPARs in myeloma. Using the public available datasets downloaded from Gene Expression Omnibus (GEO), gene expression of PPARs in clinical samples from human bone marrow and myeloma was obtained. Meanwhile, the prognostic value of PPARs in the primary myeloma was assessed using the Arkansas and Mulligan myeloma microarray datasets. mRNA levels of PPARs have a relatively lower expression in myeloma compared to bone marrow cells. High expression of PPARs predicts a good prognostic value for the overall survival rate. In conclusion, in this study, we elaborate the role of PPARs in myeloma progression, which may provide useful clinical significance for target/drug development for myeloma therapeutics.

**Keywords:** PPARs, myeloma, expression analysis, prognostic value, microarray expression analysis

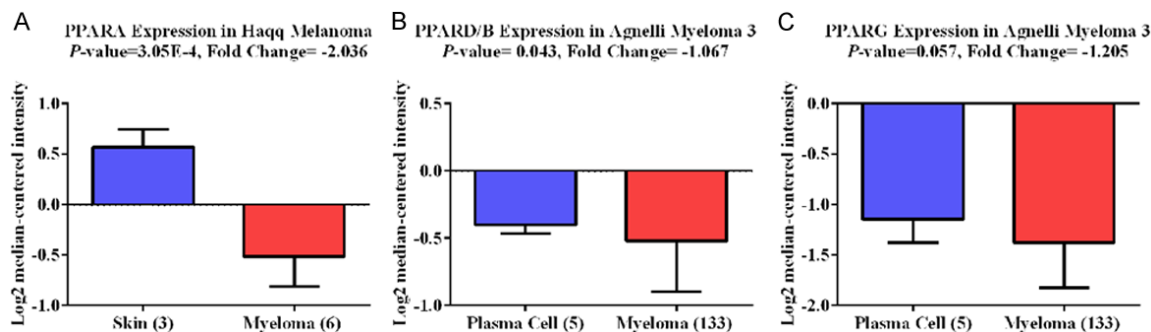
## Introduction

Multiple myeloma (MM), also known as plasma cell myeloma, is a cancer of plasma cells originally formed by malignant plasma cells [1]. From 2011 to 2013, there were an estimated 11,200 new MM cases per year in men and 8,500 new cases per year in women. By the end of 2016, it is estimated that 12,590 deaths (6,660 men and 5,930 women) occurred from this disease. The 5-year survival rate for people with multiple myeloma is about 49% [2]. Myeloma arises from an asymptomatic premalignant proliferation of monoclonal plasma cells that are derived from post-germinal-center B cells [1]. Several genetic abnormalities that occur in tumor plasma cells play major roles in the pathogenesis of myeloma, such as *MMSET*, *FGFR3* [3] and *MYC* [4] etc. Besides, alteration in microRNAs expression and gene methylation modifications also contributed to myeloma progression [5].

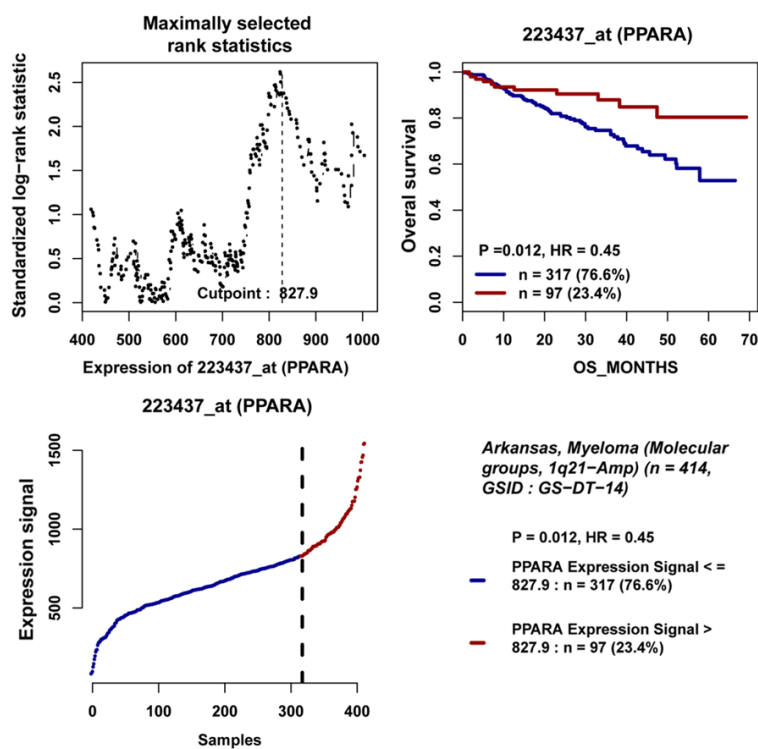
Gene expression microarrays provide a snapshot of all the transcriptional activity in a biological sample. Unlike most traditional molecular biology tools, which generally allow the study of a single gene or a small set of genes, microarrays facilitate the discovery of totally novel and unexpected functional roles of genes. The power of these tools has been applied to a range of applications, including discovering novel disease subtypes, developing new diagnostic tools, and identifying underlying mechanisms of disease or even drug response [6, 7].

The peroxisome proliferator-activated receptors (PPARs) are a group of nuclear receptor proteins that function as transcription factors regulating the expression of genes [8]. PPARs play essential roles in cell differentiation, development and metabolism [9], and have been shown to participate in tumorigenesis recently [10]. However, little is known about their impacts on myeloma. To systematically under-

## PPARs in myeloma progression



**Figure 1.** mRNA levels of PPAR gene family in patients with myeloma. A. mRNA level of *PPARA* in myeloma dataset obtained from Haqq myeloma. B. mRNA level of *PPARD* in myeloma dataset obtained from Agnelli myeloma. C. mRNA level of *PPARG* in myeloma dataset obtained from Agnelli myeloma. Expression levels are presented as box-plots and were compared using an unpaired Student's *t* test.



**Figure 2.** Overall survival analysis of *PPARA* in the Arkansas dataset. Survival analysis was performed using a log-rank test. \* $P < 0.05$ , \*\* $P < 0.01$  and \*\*\* $P < 0.001$  (one-way ANOVA with Tukey's post-test).

stand the role of PPARs, in this study, we investigate the expression level of PPARs in MM using the publicly available datasets downloaded from Gene Expression Omnibus (GEO) database. Besides, survival analysis in terms of PPAR family members also validated using Arkansas and Mulligan myeloma microarray datasets. According to our expression analysis, mRNA level of PPARs was decreased in MM patients compared to the vehicle control bone

marrow cells, illustrating their potential role of tumor suppressor in MM progression. In addition, high levels of PPARs in MM patients predict a good survival rate. More detailed information about our experimental setup is given below.

### Material and methods

#### Gene expression dataset

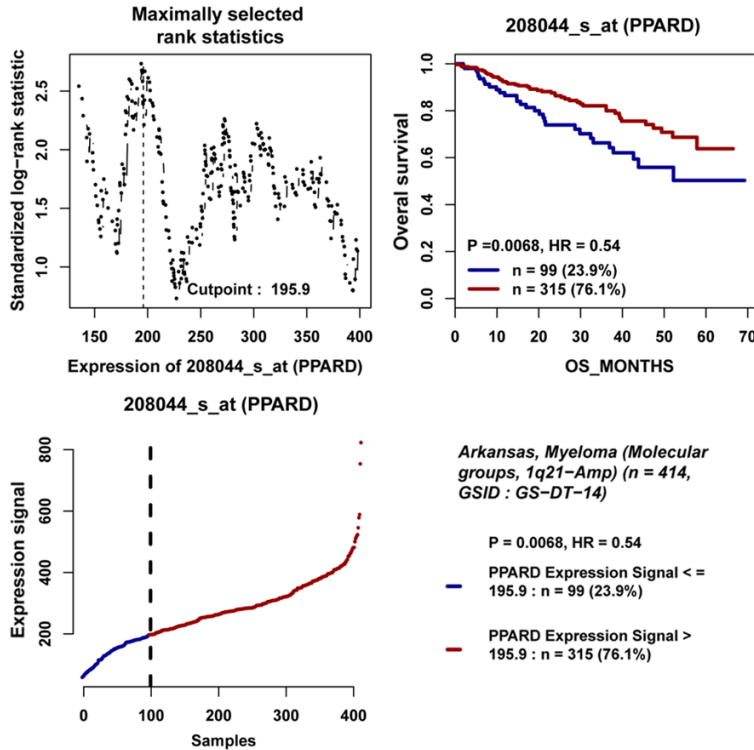
All the microarray datasets utilized for PPARs gene expression were downloaded from Gene Expression Omnibus database (GEO, <https://www.ncbi.nlm.nih.gov/geo/>) with a series matrix File (.txt). Datasets were processed basing on log2 transformation before conducting the expression analysis.

#### Clinical value of PPARs in myeloma patients

Overall survival (OS) of PPARs in myeloma patients was accessed using a data set of patients originally derived from Arkansas [11] and Mulligan myeloma [12] microarray as previous described [7].

#### Statistical analysis

For expression analysis, statistical analysis was performed using two class paired stu-



**Figure 3.** Overall survival analysis of *PPARD* in the Arkansas dataset. Survival analysis was performed using a log-rank test. \* $P < 0.05$ , \*\* $P < 0.01$  and \*\*\* $P < 0.001$  (one-way ANOVA with Tukey’s post-test).

dent’s t test. Gene expression was considered to be significant if the threshold of  $p$  value less than 5%. For survival analysis, Kaplan-Meier analysis of the over-all survival was performed in the Arkansas and Mulligan myeloma microarray datasets based on the *PPARs* gene expression. Survival analysis was performed using a log-rank test. \* $p < 0.05$ , \*\* $p < 0.01$  and \*\*\* $p < 0.001$  (one-way ANOVA with Tukey’s post-test).

**Results**

*PPARs* gene family functioned as tumor suppressors in myeloma progression

To better understand the role of *PPARs* in myeloma progression, firstly, Haqq myeloma microarray dataset was used for *PPARA* expression analysis [13]. As shown in **Figure 1A**, mRNA level of *PPARA* decreased from 0.567 to -0.517 (median), suggesting it functioned as a tumor suppressor in myeloma progression. Besides, we also analyzed mRNA level of *PPARD* (also called *PPARB*) in Agnelli myeloma, which derived from purified plasma cells obtained from 5 normal donors and 133 multiple

myeloma [14]. **Figure 1B** indicated that *PPARD* had a relatively lower expression in myeloma cells compared to the vehicle control. In addition, expression level of *PPARG* was also verified using Agnelli myeloma and it was found to be decreased dramatically as shown in **Figure 1C**. In conclusion, our expression analysis of *PPARs* gene family (*PPARA*, *PPARD* and *PPARG*) seemed to support the beneficial role of *PPARs* in myeloma initiation and progression.

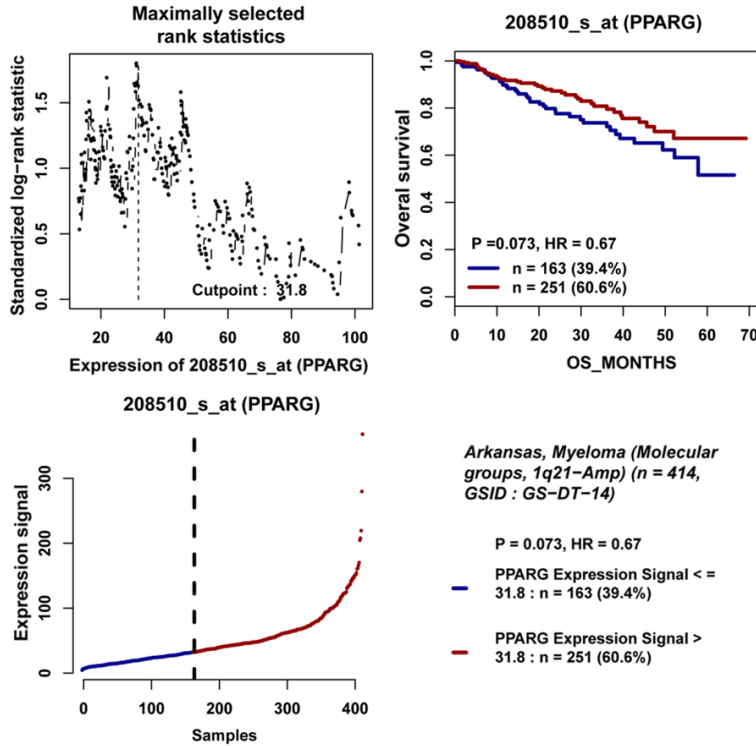
*High expression of PPARs gene family predicts a good prognostic value in myeloma*

To further determine the prognostic value of *PPARs* expression in primary myeloma, Arkansas and Mulligan myeloma microarray datasets were used for survival analysis. As shown in **Figures 2-4**, the Arkansas dataset suggests

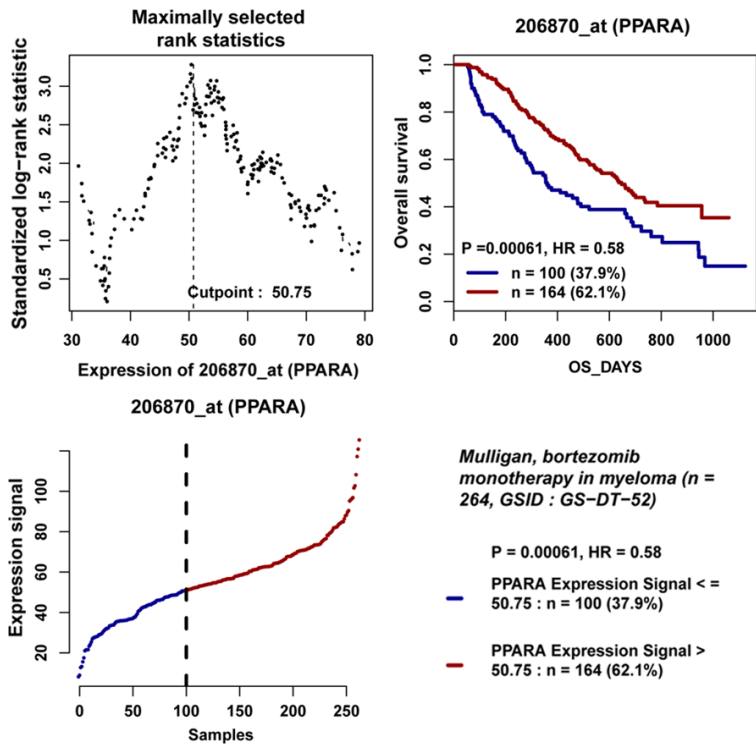
that the overall survival rate is higher in patients with high *PPARs* expression, compared to those patients with low *PPARA* (**Figure 2**), *PPARD* (**Figure 3**) and *PPARG* expression (**Figure 4**). Despite the separation or the difference between the two populations in terms of *PPARG* was not significant, we can still see the similar trends as shown in *PPARA* and *PPARD*. In addition, the Mulligan dataset revealed that myeloma patients with lower *PPARs* mRNA were significantly associated with decreased survival time (**Figures 5-7**). These findings showed the clinical significance for *PPAR* gene family in myeloma and validated the need to further understand the regulation of *PPARs* expression and function.

**Discussion**

With the help of high-through Screening (HTS) analysis, determination of gene expression for cells and tissue has become a major tool for scientific research in medicine [15-17]. Microarray experiments allow description of genome-wide expression changes in health and disease. The results of such experiments

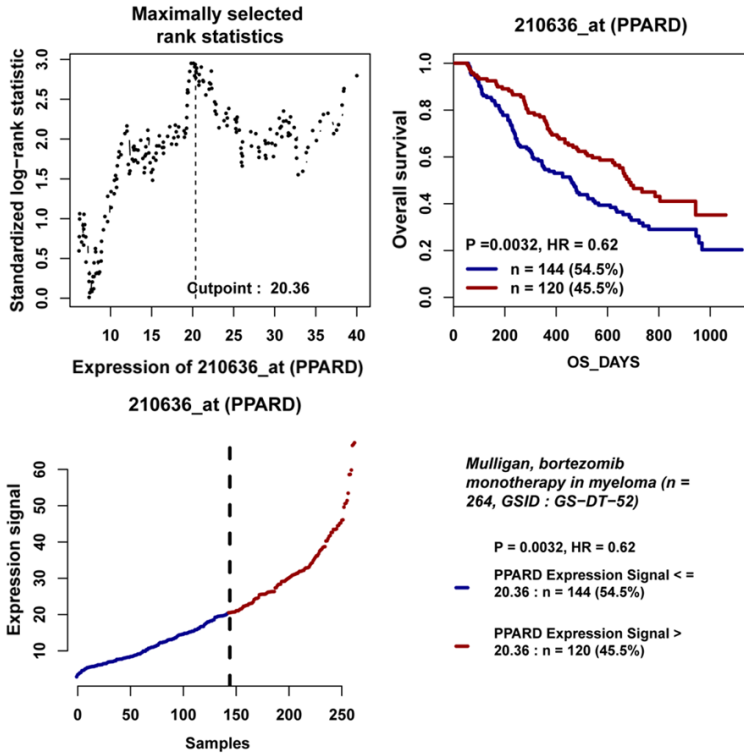


**Figure 4.** Overall survival analysis of *PPARG* in the Arkansas dataset. Survival analysis was performed using a log-rank test. \* $P < 0.05$ , \*\* $P < 0.01$  and \*\*\* $P < 0.001$  (one-way ANOVA with Tukey's post-test).

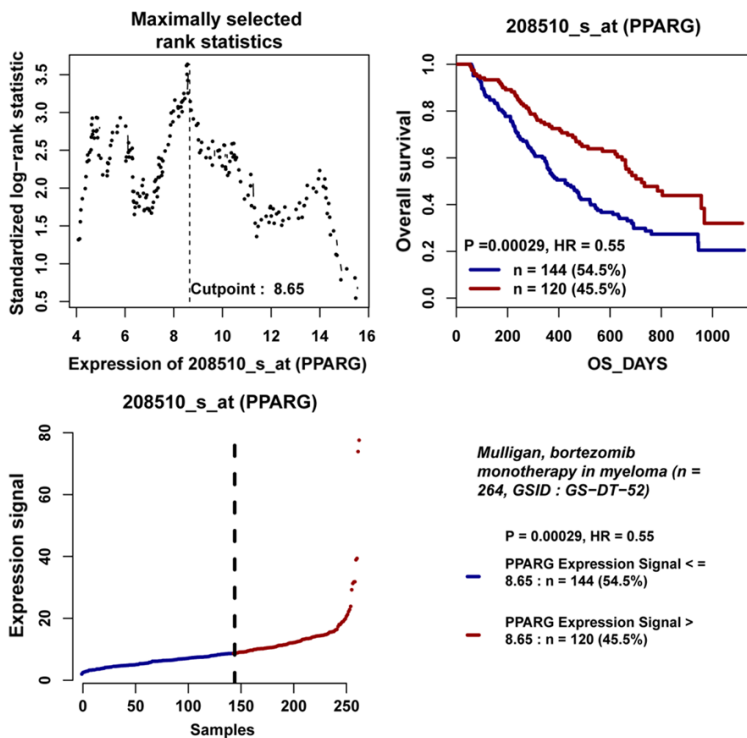


**Figure 5.** Overall survival analysis of *PPARA* in the Mulligan dataset. Survival analysis was performed using a log-rank test. \* $P < 0.05$ , \*\* $P < 0.01$  and \*\*\* $P < 0.001$  (one-way ANOVA with Tukey's post-test).

are expected to change the methods employed in the diagnosis and prognosis of disease, especially for cancers [18]. In the study, the microarray analysis confirmed that PPARs were decreased in myeloma. As ligand-activated transcription factors that belong to the nuclear hormone receptor superfamily, three subtypes of PPARs have been identified to be highly associated with cancer progression [19]. In some tissues, the expression level of PPARs and/or their activation correlates with a positive outcome against cancer, while, in other tissue types, their expression and activation have the opposite effect [20]. To determine if PPARs expression is of clinical relevance, we interrogated databases of gene expression in patient samples. Multiple independent datasets of patient samples indicated that PPARs expression was decreased in the primary tumors of patients with lung, liver and brain cancers. This is consistent with previous publications. In brain tumor, some studies have shown that PPAR $\alpha$  agonists interfere with glioblastoma growth and malignancy, as well as inhibit growth and expansion of brain tumor stem cells [21, 22]. For lung cancer, PPAR $\alpha$  activation generally inhibits tumorigenesis through its antiangiogenic and anti-inflammatory effects [23]. Pharmacological activation of PPARs could attenuate lung cancer [24, 25]. Other anti-cancer effects of PPARs ligands have been reported in several gastric cancer [26], liver cancer [27], pancreatic cancer [28] etc. As an anti-inflammatory nuclear receptor, Otsuyama and colleagues explored the expression level



**Figure 6.** Overall survival analysis of *PPARD* in the Mulligan dataset. Survival analysis was performed using a log-rank test. \* $P < 0.05$ , \*\* $P < 0.01$  and \*\*\* $P < 0.001$  (one-way ANOVA with Tukey's post-test).



**Figure 7.** Overall survival analysis of *PPARG* in the Mulligan dataset. Survival analysis was performed using a log-rank test. \* $P < 0.05$ , \*\* $P < 0.01$  and \*\*\* $P < 0.001$  (one-way ANOVA with Tukey's post-test).

of PPARs in myeloma and whether PPAR activation could inhibit the proliferation of myeloma cells *in vitro* [29]. Finally, authors concluded that primary myeloma cells from newly diagnosed MM patients as well as myeloma cell lines predominantly expressed the PPAR $\beta$  gene and also showed the weak expression of PPAR $\gamma$ . And activation of PPAR $\beta$  and PPAR $\gamma$  using carbacyclin and troglitazone would result in the suppressive effect on primary myeloma cells as well as myeloma cell lines. Thus, these studies confirmed the tumor suppressor role of PPARs in myeloma, which is consistent with our expression analysis.

To test the prognostic value of PPARs expression in Myeloma, we used two microarray datasets from Arkansas and Mulligan. The Arkansas dataset available indicated that high PPARs mRNA in the tumors of myeloma was associated with longer time to relapse. Despite the separation or the difference between the two populations in terms of *PPARG* was not significant, we can still see the similar trends as shown in *PPARA* and *PPARD*. Besides, the Mulligan dataset also confirmed that low expression of PPARs was significantly associated with decreased survival and shorter time to relapse, which was validated by previous groups [29].

In conclusion, in this study, we explored the expression of PPARs ( $\alpha$ ,  $\beta$  and  $\gamma$ ), and test the prognostic value of PPARs in patients with myeloma. Importantly, our data revealed that PPAR gene family served as tumor suppressors in myeloma initiation and pro-

gression and could be used as a predictor of cancer progression in patients suffering from myeloma.

### Acknowledgements

The authors want to thank GEO database, Arkansas and Mulligan myeloma microarray datasets for making their data readily available to the scientific community.

### Disclosure of conflict of interest

None.

**Address correspondence to:** Linsen Shi, Collge of Pharmaceutical Science, Zhejiang Chinese Medical University, Gaoke Road, Fuyang District, Hangzhou 311402, Zhejiang, China. Tel: +86-0571-61768135; Fax: +86-0571-61768135; E-mail: pjstone@163.com

### References

- [1] Palumbo A and Anderson K. Multiple myeloma. *N Engl J Med* 2011; 364: 1046-1060.
- [2] Siegel RL, Miller KD and Jemal A. Cancer statistics, 2017. *CA Cancer J Clin* 2017; 67: 7-30.
- [3] Bergsagel PL and Kuehl WM. Molecular pathogenesis and a consequent classification of multiple myeloma. *J Clin Oncol* 2005; 23: 6333-6338.
- [4] Manier S, Salem KZ, Park J, Landau DA, Getz G and Ghobrial IM. Genomic complexity of multiple myeloma and its clinical implications. *Nat Rev Clin Oncol* 2017; 14: 100-113.
- [5] Roccaro AM, Sacco A, Thompson B, Leleu X, Azab AK, Azab F, Runnels J, Jia X, Ngo HT, Melhem MR, Lin CP, Ribatti D, Rollins BJ, Witzig TE, Anderson KC and Ghobrial IM. MicroRNAs 15a and 16 regulate tumor proliferation in multiple myeloma. *Blood* 2009; 113: 6669-6680.
- [6] Slonim DK and Yanai I. Getting started in gene expression microarray analysis. *PLoS Comput Biol* 2009; 5: e1000543.
- [7] Fan S, Li X, Tie L, Pan Y and Li X. KIAA0101 is associated with human renal cell carcinoma proliferation and migration induced by erythropoietin. *Oncotarget* 2016; 7: 13520-13537.
- [8] Tyagi S, Gupta P, Saini AS, Kaushal C and Sharma S. The peroxisome proliferator-activated receptor: a family of nuclear receptors role in various diseases. *J Adv Pharm Technol Res* 2011; 2: 236-240.
- [9] Feige JN, Gelman L, Michalik L, Desvergne B and Wahli W. From molecular action to physiological outputs: peroxisome proliferator-activated receptors are nuclear receptors at the crossroads of key cellular functions. *Prog Lipid Res* 2006; 45: 120-159.
- [10] Belfiore A, Genua M and Malaguarnera R. PPAR-gamma agonists and their effects on IGF-I receptor signaling: implications for cancer. *PPAR Res* 2009; 2009: 830501.
- [11] Weinhold N, Heuck CJ, Rosenthal A, Thanendrarajan S, Stein CK, Van Rhee F, Zangari M, Horing A, Tian E, Davies FE, Barlogie B and Morgan GJ. Clinical value of molecular subtyping multiple myeloma using gene expression profiling. *Leukemia* 2016; 30: 423-430.
- [12] Mulligan G, Mitsiades C, Bryant B, Zhan F, Chng WJ, Roels S, Koenig E, Fergus A, Huang Y, Richardson P, Trepicchio WL, Broyl A, Sonneveld P, Shaughnessy JD Jr, Bergsagel PL, Schenkein D, Esseltine DL, Boral A and Anderson KC. Gene expression profiling and correlation with outcome in clinical trials of the proteasome inhibitor bortezomib. *Blood* 2007; 109: 3177-3188.
- [13] Haqq C, Nosrati M, Sudilovsky D, Crothers J, Khodabakhsh D, Pulliam BL, Federman S, Miller JR, 3rd, Allen RE, Singer MI, Leong SP, Ljung BM, Sagebiel RW and Kashani-Sabet M. The gene expression signatures of melanoma progression. *Proc Natl Acad Sci U S A* 2005; 102: 6092-6097.
- [14] Agnelli L, Mosca L, Fabris S, Lionetti M, Andronache A, Kwee I, Todoerti K, Verdelli D, Battaglia C, Bertoni F, Deliliers GL and Neri A. A SNP microarray and FISH-based procedure to detect allelic imbalances in multiple myeloma: an integrated genomics approach reveals a wide gene dosage effect. *Genes Chromosomes Cancer* 2009; 48: 603-614.
- [15] Li J, Fan S, Han D, Xie J, Kuang H and Ge P. Microarray gene expression profiling and bioinformatics analysis of premature ovarian failure in a rat model. *Exp Mol Pathol* 2014; 97: 535-541.
- [16] Fan S, Pan Z, Geng Q, Li X, Wang Y, An Y, Xu Y, Tie L, Pan Y and Li X. Layered signaling regulatory networks analysis of gene expression involved in malignant tumorigenesis of non-resolving ulcerative colitis via integration of cross-study microarray profiles. *PLoS One* 2013; 8: e67142.
- [17] Fan S, Geng Q, Pan Z, Li X, Tie L, Pan Y and Li X. Clarifying off-target effects for torcetrapib using network pharmacology and reverse docking approach. *BMC Syst Biol* 2012; 6: 152.
- [18] Tarca AL, Romero R and Draghici S. Analysis of microarray experiments of gene expression profiling. *Am J Obstet Gynecol* 2006; 195: 373-388.
- [19] Tachibana K, Yamasaki D, Ishimoto K and Doi T. The role of PPARs in cancer. *PPAR Res* 2008; 2008: 102737.

## PPARs in myeloma progression

- [20] Youssef J and Badr M. Peroxisome proliferator-activated receptors and cancer: challenges and opportunities. *Br J Pharmacol* 2011; 164: 68-82.
- [21] Chearwae W and Bright JJ. PPARgamma agonists inhibit growth and expansion of CD133+ brain tumour stem cells. *Br J Cancer* 2008; 99: 2044-2053.
- [22] Grommes C, Landreth GE, Sastre M, Beck M, Feinstein DL, Jacobs AH, Schlegel U and Heneka MT. Inhibition of in vivo glioma growth and invasion by peroxisome proliferator-activated receptor gamma agonist treatment. *Mol Pharmacol* 2006; 70: 1524-1533.
- [23] Lakshmi SP, Reddy AT, Banno A and Reddy RC. PPAR agonists for the prevention and treatment of lung cancer. *PPAR Res* 2017; 2017: 8252796.
- [24] Fukumoto K, Yano Y, Virgona N, Hagiwara H, Sato H, Senba H, Suzuki K, Asano R, Yamada K and Yano T. Peroxisome proliferator-activated receptor delta as a molecular target to regulate lung cancer cell growth. *FEBS Lett* 2005; 579: 3829-3836.
- [25] Tsubouchi Y, Sano H, Kawahito Y, Mukai S, Yamada R, Kohno M, Inoue K, Hla T and Kondo M. Inhibition of human lung cancer cell growth by the peroxisome proliferator-activated receptor-gamma agonists through induction of apoptosis. *Biochem Biophys Res Commun* 2000; 270: 400-405.
- [26] Lu J, Imamura K, Nomura S, Mafune K, Nakajima A, Kadowaki T, Kubota N, Terauchi Y, Ishii G, Ochiai A, Esumi H and Kaminishi M. Chemopreventive effect of peroxisome proliferator-activated receptor gamma on gastric carcinogenesis in mice. *Cancer Res* 2005; 65: 4769-4774.
- [27] Borbath I and Horsmans Y. The role of PPAR gamma in hepatocellular carcinoma. *PPAR Res* 2008; 2008: 209520.
- [28] Kumei S, Motomura W, Yoshizaki T, Takakusaki K and Okumura T. Troglitazone increases expression of E-cadherin and claudin 4 in human pancreatic cancer cells. *Biochem Biophys Res Commun* 2009; 380: 614-619.
- [29] Otsuyama KI, Ma Z, Abroun S, Amin J, Shamsa-senjan K, Asaoku H and Kawano MM. PPARbeta-mediated growth suppression of baicalein and dexamethasone in human myeloma cells. *Leukemia* 2007; 21: 187-190.