Original Article Identification of B-cells participating in differentially-expressed pathways and hub genes in postmenopausal women with osteoporosis

Yong-Kun Wei¹, Bin-Hui Yang¹, Hui-Ling Ma², Li-Feng Yang¹, Wan-Jun Li³, Man Xiong¹, Zhen Ou-Yang¹, Yong-Cai Song¹

Departments of ¹Orthopedics and ³Pathology, 3201 Hospital, Hanzhong 723000, Shaanxi Province, China; ²Hanzhong Vocational and Technical College, Hanzhong 723000, Shaanxi Province, China

Received January 18, 2018; Accepted May 7, 2019; Epub July 15, 2019; Published July 30, 2019

Abstract: Objective: Osteoporosis (OP) can result in low bone mineral density (BMD) and reduced bone strength. This disease has been identified as a major public health problem around the world. Hence, it is necessary to find a proactive method, identifying high-risk OP patients, and to investigate the corresponding pathology. The objective of the current study was to reveal key pathways and hub genes associated with OP, utilizing Gibbs sampling. Methods: Informative pathways (IPs) with genes more than 5 were extracted, based on the KEGG database and microarray profiles. Obtained IPs were then converted into the Markov chain (MC). Afterward, Gibbs sampling was implemented to obtain a new MC. Subsequently, probabilities of IPs were counted through the MC Monte Carlo (MCMC) algorithm, followed by detection of differentially-expressed pathways (DEPs) based on adjusted probabilities of IPs higher than 0.65. Moreover, genes enriched in the DEPs were analyzed using the same sampling strategy. Hub genes were identified based on the threshold of adjusted probabilities greater than 0.80. Results: When the gene set was 5, a total of 278 IPs were extracted. After Gibbs sampling, only 1 DEP, mineral absorption, was identified according to the adjusted alfa.pi > 0.65. Moreover, after the probability of genes in this DEP was evaluated, a total of 8 hub genes were screened out, including VDR, ACP6, FTCD, ALDOB, ATIC, ALDH3A1, MAPK3, and OXCT2. Conclusion: Comprehensive approaches, including Gibbs sampling and Markov chain, might provide good reference for OP treatment in the future. Identified DEPs and hub genes might play pivotal roles in onset and progression of OP. They may be helpful in the development of available therapeutic drugs for treatment of OP in the future.

Keywords: Osteoporosis, Gibbs sampling, hub genes, differentially-expressed pathways

Introduction

Postmenopausal females have a high incidence of osteoporosis (OP). This is due to the co-existence of many independent predisposing factors, including estrogen deficiency, calcium loss, and aging [1-3]. OP, characterized by an imbalance between bone resorption and bone formation [4], results in low bone mineral density (BMD) and reduced bone strength. It also increases the risk of fragility fractures [5]. Significantly, OP is a major public health concern worldwide, particularly in China [6]. Currently, treatment of OP is mainly dependent on drugs, yet they come with a high cost. They are time consuming and have many side effects. Moreover, the curative effects are not ideal. Knowledge of molecular mechanisms concerning this disease remains poor. Thus, a proactive method, identifying high-risk OP patients and investigating the corresponding pathology, is urgent.

Apart from estrogen, calcium, and aging factors, genetic factors have been implicated in the progression of OP in postmenopausal women [7-9], including OPG [10] and ESR2 [11]. Additionally, B-cell precursors can differentiate into osteoclasts *in vitro* [12] and estrogen deficiencies trigger the production of B lymphopoiesis [13]. It has conclusively been revealed that estrogen suppresses B lymphocyte production during differentiation steps from pro-B cells to pre-B cells [13]. Thus, 17ß-Estradiol, as an

estrogen, excites antibody production by B-cells [14]. Compared to 17ß-Estradiol, bazedoxifene (an estrogen receptor modulator) restrains B lymphopoiesis generation at a later stage of B-cell differentiation [15]. Pineda et al. [16] attempted to reproduce one of the major risk factors (estrogen deficiencies after menopause or bilateral ovariectomy) for OP in women. However, the roles of B-cells in bone metabolism and OP remain largely unknown, especially at the systematic gene expression level in humans in vivo. At present, microarray technology is broadly used to detect key gene biomarkers for OP. Therefore, more investigators are using bioinformatics strategies to investigate the molecular mechanisms of OP, studying the microarray profiles of OP. GSE7429 is one of the microarray profiles of OP and was deposited by Xiao et al. [17]. They detected underexpression of ESR1 and MAPK3 in B-cells regulating the factor secretion, causing increased osteoclastogenesis or reduced osteoblastogenesis. In 2015, using the same data deposited by Xiao et al. [17], Yan et al. [18] extracted 238 differentially-expressed genes (DEGs) which were involved in OP. These included MAP-K3, MAP3K10, MAP3K9, COX10, COX15, ATIC, UMPS, and HPRT1. Ma et al. [19] utilized the same gene expression data to identify several crucial genes associated with OP, including CSTA, TUBA1B, and CCNE1. However, thus far, most studies involving OP have paid attention to several important genes. Of note, several of these gene signatures have poor reproducibility and overlapped among different studies, although using the same microarray profiles. Consequently, understanding the complex interaction among genes is very challenging. Generally, detecting pathways participated in a given phenotype is very important. As demonstrated, signaling pathways, instead of single genes, govern the process of diseases [20]. Thus, it is important to identify potential pathways related to OP, further exploring the pathology of OP. Gibbs sampling, a Markov chain Monte Carlo (MCMC) algorithm obtaining a sequence of observations that are approximated from a specified multivariate probability distribution [21], can be used to identify differentially-expressed pathways (DEPs).

In the current study, integrated approaches of Gibbs sampling and MC were utilized to predict hub genes and pivotal pathways of OP. This study converted 278 IPs based on gene set as 5. Gibbs sampling was then performed to obtain a new Markov chain (MC). Moreover, the MCMC algorithm was utilized to obtain hub genes with an expression probability > 0.8 and pivotal pathways with an expression probability > 0.65. Therefore, present outcomes provide novel pathway biomarkers as tools allowing for better diagnosis and prevention of OP in the future.

Materials and methods

In the current study, Gibbs sampling was utilized to explore the significance of pathways, examining their roles in OP. Gibbs sampling, a means of statistical inference, especially Bayesian inference, is an MCMC algorithm used to obtain a sequence of observations. These are approximated from a specified multivariate probability distribution [22-24].

Microarray data

Raw microarray data concerning PMOP (accession number: GSE7429) [17] were downloaded from Gene Expression Omnibus (GEO, http:// www.ncbi.nlm.nih.gov/geo/) based on the platform of GPL96 [HG-U133A] Affymetrix Human Genome U133A array. A total of 20 samples were available, including 20 B-cell samples isolated from whole blood, obtained from 10 postmenopausal females with low BMD, aged 56-60 years. Also, there were 10 samples having high BMD, aged 54-60 years. Inclusion criteria for PMOP were: Spine or hip Z-score < -0.84 for the low BMD group; Spine or hip Z-score > 0.84 for the high BMD group. This study was approved by the Institutional Review Board and informed consent was obtained, as mentioned in the article by Xiao et al. [17].

Probe IDs due to concentrated expression levels were transformed into gene symbols. Duplicated genes were then removed in the matrix. Overall, 12,437 genes were obtained for subsequent analysis.

Biological pathways

Kyoto Encyclopedia of Genes and Genomes (KEGG) database (www.genome.jp/kegg/) offers a reference knowledge base for understanding cellular processes. First, this study collected 300 original pathways (6919 human genes) from the KEGG database and named them as OPs. Next, this study mapped the

Table 1. Informative pathways (IPs) with ≥ 200 genes for osteoporosis

0	
IPs	Count
hsa05200: Pathways in cancer	366
hsa04151: PI3K-Akt signaling pathway	310
hsa04080: Neuroactive ligand-receptor interaction	281
hsa05166: HTLV-I infection	234
hsa04060: Cytokine-cytokine receptor interaction	233
hsa04010: MAPK signaling pathway	231

microarray genes (12,437 genes) to the OPs. As reported, pathways with too few genes may not have sufficient biological information [25]. Thus, a set of pathways was extracted by excluding pathways with less than 5 microarray genes. When removing the pathways having gene sizes < 5, a total of 278 informative pathways (IPs) were identified for further analysis.

In performing Gibbs sampling, the IPs needed to be converted into a data set with functional class expression measurements that were Markov chains (MCs). This was conducted using the Annotation Modified and Faster Global Optimization (MFGO) function of the Bayesian Approach for GeneSet Selection (BAGS) package [26].

Calculation of probabilities of IPs

After IPs were converted into MCs, posterior inferences for them were defined to measure probability distributions of IPs from OP [27].

In the current study, an empty set was first defined. Next, the MC dataset was deposited, including IPs with genes > 5 (N = 278) to this empty set. Afterward, Gibbs sampling was performed to establish the 10,000 dimensional-random vectors of N samples. Subsequently, these 10,000 dimensional were initiated into random vectors. Of these, one vector was extracted each time to produce the random number. This process was repeated 10,000 times. A new MC dataset, i.e. 10,000 probability of each IP, was received. Using the following formula, the probability of N IPs was computed:

Alfa.pi = the average probability of IPs (from 2000 to 10000) * 250/(10000 - 2000 + 1)

In this equation, "alfa.pi" denotes "posterior value of an MF". Afterward, researchers calculated the adjusted "alfa.pi" values for each IP based on the parameters of R values, *P* values, and rank order. Specifically, Student's t-test was used to calculate the *P*-values for each IP. The rank order of each IP was obtained based on the *P*-values. Next, R-values were computed based on the rank order and "alfa.pi" values.

Identification of DEPs

Empirically, the cut-off threshold of posterior probability was set as 0.05, which suggested the reliability of this sampler [28]. However, there was no obvious standard for high frequencies. As reported in a previous study [29], if the probability of a biological process was > 0.6, it was considered to be differentially-expressed. Thus, in the current study, relying on probabilities of IPs higher than 0.65, DEPs were identified. Moreover, genes in the differentially-expressed MF were believed to be DEGs. These DEGs were then merged. Statistical analysis was implemented on these merged genes that appeared in differentiallyexpressed MF.

Selection of hub genes in DEPs

The gene set enriched in DEPs was obtained, as with DEP analysis using Gibbs sampling. This study also implemented the same analysis for the pathway gene set. When the threshold of adjusted was set to 0.80, hub genes were identified.

Results

Identification of IPs

Under the criteria of IPs with at least 5 microarray genes, 278 IPs were identified. Of these IPs, 213 pathways displayed a gene number between 1 and 99. A total of 59 categories had more than or equal to 100 genes, but less than 200 genes. Four terms ranged from 200 to 300 genes and 2 terms had more than 300 genes (including hsa05200: Pathways in cancer with 366 genes and hsa04151: PI3K-Akt signaling pathway with 310 genes). Additionally, IPs with possessing gene count > 200 are shown in **Table 1.**

Detecting DEPs

Before assessing the probabilities of IPs based on Gibbs sampling using the MCMC algorithm, this study transformed the 278 IPs in expression data structure to the MC dataset. **Figure 1**



Figure 1. Probabilities for 278 informative pathways (IPs).



Figure 2. Expression levels of mineral absorption in high BMD and low BMD groups. From this figure, it can be seen that this pathway was downregulated in low BMD group.

displays the probability distribution of all IPs making use of the alfa.pi formula. According to adjusted alfa.pi > 0.65, only 1 DEP was identified, which was mineral absorption (adjusted alfa.pi = 0.7027). Expression levels of this DEP in the low and high BMD groups are shown in **Figure 2.** From this figure, it can be seen that this pathway was downregulated in the low BMD group. In this DEP, there were 41 genes.

Selection of hub genes

As with DEP identification, the same sampling method was used to detect hub genes from the



Figure 3. Probabilities for 41 genes in the DEP of mineral absorption.

41 genes within the DEP of mineral absorption. **Figure 3** illustrates the association of the probability distribution and each gene enriched in the DEP. Using the cut-off criteria of the adjusted alfa.pi > 0.80, 8 hub genes were extracted, including VDR, ACP6, FTCD, ALDOB, ATIC, ALDH3A1, MAPK3, and OXCT2.

Specific information is listed in Table 2.

Discussion

Gibbs sampling has been broadly utilized as way of statistical inference, including Bayesian inference [30]. Of note, Gibbs sampling, a Markov Chain Monte Carlo (MCMC) algorithm, can obtain a sequence of observations, approximated from a specified multivariate probability distribution [22-24]. Remarkably, on the basis of the probabilities, differentially-expressed biological processes and key genes are potentially identified. This might be important in revealing the pathology of disorders. Hence, in the current analysis employed Gibbs sampling to evaluate the significance of pathways, examining their function in OP. Consequently, only1DEP,namedasmineralabsorption,wasidentified according to the adjusted alfa.pi > 0.65. Moreover, after the probability of genes in this DEP was evaluated, a total of 8 hub genes were screened out, including VDR, ACP6, FTCD, ALDOB, ATIC, ALDH3A1, MAPK3, and OXCT2.

As demonstrated, calcium is the dominant mineral in bones. It has been regarded as a shortfall nutrient reported in the Dietary Guidelines

Hub gene	P value	Rank p	R value	alfa	Alfa-adj
VDR	0.016282	8	0.972318	0.905137	0.880081
ACP6	0.011281	4	0.986159	0.886014	0.873751
FTCD	0.01995	12	0.958478	0.911511	0.873663
ALDOB	0.012046	5	0.982699	0.873266	0.858157
ATIC	0.017116	10	0.965398	0.87964	0.849203
ALDH3A1	0.047386	29	0.899654	0.905137	0.814310
MAPK3	4.00E-05	1	0.99654	0.815898	0.813075
OXCT2	0.034067	20	0.930796	0.873266	0.812832

Table 2. List of hub genes based on the adjusted alfa.pi >0.80

for Americans [31]. Enhanced calcium intake has been suggested to be related to increased bone accrual [32]. Significantly, promoting calcium absorption, as well as other bone-related minerals, is an attractive strategy in reducing risks of OP [33]. Legette et al. [34] implicated that, during middle-age, mineral absorption decreases and bone resorption rates increase. Results suggested that this led to a higher risk for OP. Moreover, current results demonstrated that mineral absorption is very important for OP development.

VDR, a nuclear transcription factor, affects calcium absorption, mineralization, and bone remodeling [35]. Some studies have confirmed the correlation between VDR polymorphisms and occurrence of fractures. For example, VDR Focl and Tagl polymorphisms have been reported to be associated with low BMD at the lumbar spine and femoral neck, according to several studies [36-38]. Stathopoulou et al. [39] implicated that, under lower calcium intake (<680 mg/d), the presence of the B-allele of VDR Bsml polymorphisms enhanced the risk of OP by 118%. Thus, Horst-Sikorska et al. [40] concluded that adequate calcium intake "masked" the VDR genetic influence on bones. In the current study, VDR was the hub gene with the highest probability. Therefore, current results were in line with the above reports, suggesting an important role for VDR in OP development.

In general, the current study provides a comprehensive bioinformatics analysis (using the Gibbs sampling method), identifying DEPs and hub genes that might be involved in the development of OP. Current findings revealed that the pathway of mineral absorption might play key roles in the development of OP. In addition, present results might provide a better understanding for pathologies of OP, indicating potential targets for development of therapeutic drugs for OP in future. However, the current study had several disadvantages. The main drawback was the relatively small population. Another limitation is that additional experiments are necessary to confirm the results obtained above using bioinformatic approaches. Moreover, additional animal or tissue experiments should be conducted, validating current re-

sults using more samples based on Western blotting or PCR.

Disclosure of conflict of interest

None.

Address correspondence to: Yong-Kun Wei, Department of Orthopedics, 3201 Hospital, No. 783 Tianhan Road, Hanzhong 723000, Shaanxi Province, China. Tel: 86-0916-2383527; Fax: 86-0916-2383527; E-mail: weiyongkun2005@sohu.com

References

- [1] Kendler DL, Marin F, Zerbini CAF, Russo LA, Greenspan SL, Zikan V, Bagur A, Malouf-Sierra J, Lakatos P, Fahrleitner-Pammer A, Lespessailles E, Minisola S, Body JJ, Geusens P, Moricke R and Lopez-Romero P. Effects of teriparatide and risedronate on new fractures in post-menopausal women with severe osteoporosis (VERO): a multicentre, double-blind, double-dummy, randomised controlled trial. Lancet 2018; 391: 230-240.
- [2] Briot K and Roux C. [Post-menopausal osteoporosis: up-to-date]. Rev Med Interne 2016; 37: 195-200.
- [3] Ishtiaq S, Fogelman I and Hampson G. Treatment of post-menopausal osteoporosis: beyond bisphosphonates. J Endocrinol Invest 2015; 38: 13-29.
- [4] Khosla S and Hofbauer LC. Osteoporosis treatment: recent developments and ongoing challenges. Lancet Diabetes Endocrinol 2017; 5: 898-907.
- [5] Muschitz C, Kocijan R, Haschka J, Pahr D, Kaider A, Pietschmann P, Hans D, Muschitz GK, Fahrleitner-Pammer A and Resch H. TBS reflects trabecular microarchitecture in premenopausal women and men with idiopathic osteoporosis and low-traumatic fractures. Bone 2015; 79: 259-266.

- [6] Ni W and Jiang Y. Evaluation on the cost-effective threshold of osteoporosis treatment on elderly women in China using discrete event simulation model. Osteoporos Int 2017; 28: 529-538.
- [7] Yuan J, Tickner J, Mullin BH, Zhao J, Zeng Z, Morahan G and Xu J. Advanced genetic approaches in discovery and characterization of genes involved with osteoporosis in mouse and human. Front Genet 2019; 10: 288.
- [8] Guo Y, Dong SS, Chen XF, Jing YA, Yang M, Yan H, Shen H, Chen XD, Tan LJ, Tian Q, Deng HW and Yang TL. Integrating epigenomic elements and GWASs identifies BDNF gene affecting bone mineral density and osteoporotic fracture risk. Sci Rep 2016; 6: 30558.
- [9] Wang C, Zhang Z, Zhang H, He JW, Gu JM, Hu WW, Hu YQ, Li M, Liu YJ, Fu WZ, Yue H, Ke YH and Zhang ZL. Susceptibility genes for osteoporotic fracture in postmenopausal Chinese women. J Bone Miner Res 2012; 27: 2582-2591.
- [10] Song JF, Jing ZZ, Hu W and Su YX. Association between single nucleotide polymorphisms of the osteoprotegerin gene and postmenopausal osteoporosis in Chinese women. Genetics & Molecular Research Gmr 2013; 12: 3279-3285.
- [11] Kaminski A, Bogacz A and Czerny B. The rs1256044 polymorphism in the ESR2 gene and the risk for osteoporosis in Polish postmenopausal women. Gynecol Endocrinol 2018; 34: 579-583.
- [12] Attaianese C and Tomasso G. IL-7 induces bone loss in vivo by induction of receptor activator of nuclear factor kappa B ligand and tumor necrosis factor alpha from T cells. Proceedings of the National Academy of Sciences of the United States of America 2003; 100: 125-130.
- [13] Nordqvist J, Bernardi A, Islander U and Carlsten H. Effects of a tissue-selective estrogen complex on B lymphopoiesis and B cell function. Immunobiology 2017; 222: 918-923.
- [14] Erlandsson MC, Jonsson CA, Lindberg MK, Ohlsson C and Carlsten H. Raloxifene- and estradiol-mediated effects on uterus, bone and B lymphocytes in mice. J Endocrinol 2002; 175: 319-327.
- [15] Bernardi Al, Andersson A, Grahnemo L, Nurkkala-Karlsson M, Ohlsson C, Carlsten H and Islander U. Effects of lasofoxifene and bazedoxifene on B cell development and function. Immun Inflamm Dis 2014; 2: 214-225.
- [16] Pineda B, Serna E, Laguna-Fernández A, Noguera I, Panach L, Hermenegildo C, Tarín JJ, Cano A and García-Pérez MÁ. Gene expression profile induced by ovariectomy in bone marrow of mice: a functional approach to identify new

candidate genes associated to osteoporosis risk in women. Bone 2014; 65: 33-41.

- [17] Xiao P, Chen Y, Jiang H, Liu YZ, Pan F, Yang TL, Tang ZH, Larsen JA, Lappe JM and Recker RR. In vivo genome-wide expression study on human circulating B cells suggests a novel ESR1 and MAPK3 network for postmenopausal osteoporosis. J Bone Miner Res 2008; 23: 644-54.
- [18] Yan B, Li J and Zhang L. Identification of B cells participated in the mechanism of postmenopausal women osteoporosis using microarray analysis. Int J Clin Exp Med 2015; 8: 1027-34.
- [19] Ma M, Chen X, Lu L, Yuan F, Zeng W, Luo S, Yin F and Cai J. Identification of crucial genes related to postmenopausal osteoporosis using gene expression profiling. Aging Clin Exp Res 2016; 28: 1067-1074.
- [20] Sahakyan AB and Balasubramanian S. Long genes and genes with multiple splice variants are enriched in pathways linked to cancer and other multigenic diseases. BMC Genomics 2016; 17: 225.
- [21] Harms RL and Roebroeck A. Robust and fast markov chain monte carlo sampling of diffusion MRI microstructure models. Front Neuroinform 2018; 12: 97.
- [22] Kozumi H and Kobayashi G. Gibbs sampling methods for Bayesian quantile regression. J Stat Comput Simul 2011; 81: 1565-1578.
- [23] El-Hay T, Friedman N and Kupferman R. Gibbs sampling in factorized continuous-time Markov processes. arXiv preprint arXiv: 1206.3251 2012.
- [24] Chib S and Winkelmann R. Markov chain Monte Carlo analysis of correlated count data. Journal of Business & Economic Statistics 2012.
- [25] Ahn T, Lee E, Huh N and Park T. Personalized identification of altered pathways in cancer using accumulated normal tissue data. Bioinformatics 2014; 30: i422-i429.
- [26] Quiroz-Zarate A, Quiroz-Zarate MA, Gibbs CD-RGR and GrpMean RGRGR. Package 'BAGS'. 2013.
- [27] Moradkhani H, DeChant CM and Sorooshian S. Evolution of ensemble data assimilation for uncertainty quantification using the particle filter-Markov chain Monte Carlo method. Water Resources Research 2012; 48.
- [28] Yan Y and Zhang S. An improved estimation method and empirical properties of the probability of informed trading. Journal of Banking & Finance 2012; 36: 454-467.
- [29] Huang ZJ, Shen QH, Wu YS and Huang YL. A Gibbs sampling method to determine biomarkers for asthma. Comput Biol Chem 2017; 67: 255-259.
- [30] Walsh B. Markov chain monte carlo and gibbs sampling. 2004.

- [31] Report of the dietary guidelines advisory committee on the dietary guidelines for Americans, 2010. 2010.
- [32] Palacios C, Martin BR, Mccabe GP, Mccabe L, Peacock M and Weaver CM. Dietary calcium requirements do not differ between Mexican-American boys and girls. J Nutr 2014; 144: 1167-73.
- [33] Weaver CM. Diet, gut microbiome, and bone health. Curr Osteoporos Rep 2015; 13: 125-30.
- [34] Legette LL, Lee W, Martin BR, Story JA, Campbell JK, Weaver CM. Prebiotics enhance magnesium absorption and inulin-based fibers exert chronic effects on calcium utilization in a postmenopausal rodent model. J Food Sci 2012; 77: H88-94.
- [35] Nakamichi Y, Udagawa N, Horibe K, Mizoguchi T, Yamamoto Y, Nakamura T, Hosoya A, Kato S, Suda T and Takahashi N. VDR in osteoblastlineage cells primarily mediates vitamin D treatment-induced increase in bone mass by suppressing bone resorption. J Bone Miner Res 2017; 32: 1297-1308.
- [36] Gonzalez-Mercado A, Sanchez-Lopez JY, Regla-Nava JA, Gamez-Nava JI, Gonzalez-Lopez L, Duran-Gonzalez J, Celis A, Perea-Diaz FJ, Salazar-Paramo M and Ibarra B. Association analysis of vitamin D receptor gene polymorphisms and bone mineral density in postmenopausal Mexican-Mestizo women. Genet Mol Res 2013; 12: 2755-2763.

- [37] Tantawy M, Amer M, Raafat T and Hamdy N. Vitamin D receptor gene polymorphism in Egyptian pediatric acute lymphoblastic leukemia correlation with BMD. Meta Gene 2016; 9: 42-46.
- [38] Bao L, Chen M, Lei Y, Zhou Z, Shen H and Le F. Association between vitamin D receptor Bsml polymorphism and bone mineral density in pediatric patients: a meta-analysis and systematic review of observational studies. Medicine (Baltimore) 2017; 96: e6718.
- [39] Stathopoulou MG, Dedoussis GV, Trovas G, Theodoraki EV, Katsalira A, Dontas IA, Hammond N, Deloukas P and Lyritis GP. The role of vitamin D receptor gene polymorphisms in the bone mineral density of Greek postmenopausal women with low calcium intake. J Nutr Biochem 2011; 22: 752-757.
- [40] Horst-Sikorska W, Dytfeld J, Wawrzyniak A, Marcinkowska M, Michalak M, Franek E, Napiórkowska L, Drwęska N, Słomski R. Vitamin D receptor gene polymorphisms, bone mineral density and fractures in postmenopausal women with osteoporosis. Mol Biol Rep 2013; 40: 383-390.