

## Original Article

# Exploring the molecular mechanism, biomarkers and prognostic values of HCC based on gene expression microarray

Ding Li<sup>1\*</sup>, Fu-Biao Ni<sup>1\*</sup>, Heng-Jie Tang<sup>1</sup>, Chong-Chu Huang<sup>1</sup>, Cheng-Yang Liu<sup>2</sup>, Ji-Cai Chen<sup>1</sup>

<sup>1</sup>Department of General Surgery, The First Affiliated Hospital of Wenzhou Medical University, Wenzhou, Zhejiang, China; <sup>2</sup>Department of Surgery, School of Medicine, University of Pennsylvania, Philadelphia, PA, USA. \*Equal contributors.

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**Abstract:** Liver cancer is a common malignant neoplasm worldwide, causing high morbidity and mortality globally. The molecular mechanisms of hepatocarcinogenesis remains unclear. The goal of our study is to elucidate the mechanism which could improve the prognosis of liver cancer. GSE121248 was downloaded from the GEO database, which is a gene expression profile data including 70 tumor samples and 37 adjacent normal samples from hepatocellular carcinoma (HCC). The differentially expressed genes (DEGs) between cancer tissues and normal tissues were screened. Subsequently, the enriched GO terms, KEGG pathways and Database for Annotation, Visualization and Integrated Discovery (DAVID) were analyzed by on-line tools. Finally, STRING database and Cytoscape software were used to construct protein-protein interactions (PPI) network and genes with high degree. Kaplan-Meier plotter (KM plotter) was used to explore the predictive prognostic value of gene expression to overall survival (OS). The results showed that there were 202 DEGs between the case samples and control samples, including 59 up-regulated genes and 143 down-regulated genes in HCC tissue samples. The study obtained a total of 132 enriched GO terms and 16 KEGG pathways, such as extracellular region, organelle membrane, extracellular space, epoxygenase P450 pathway, retinol metabolism, metabolic pathways, caffeine metabolism, tryptophan metabolism and drug metabolism-cytochrome P450. PPI suggests that BUB1B, CCNB1, EZH2, NUSAP1 and CDC20 were the top 5 core genes. The patients with high expression of 5 core genes had poor OS according to online database. Using comprehensive bioinformatics analyses, our study attempts to identify DEGs and find potential biomarkers to predict the occurrence and development of HCC.

**Keywords:** Molecular mechanism, biomarkers, prognostic values, HCC, gene expression microarray

## Background

Primary liver cancer (namely hepatocellular carcinoma, HCC) is one of the fifth most common cancer which takes up the third top cause of cancer mortality [1]. In 2016, it afflicted in excess of 1 million people and caused 800,000 deaths globally [2]. HCC has interesting epidemiologic features and a couple of well-substantiated environmental potentially preventable risk factors have been confirmed for the disease [3]. Despite active research [1, 4, 5], the molecular mechanisms that induce HCC remain unclear.

Gene expression profiles measure the expression of thousands of genes to build a global pic-

ture of cell function. Gene expression profiling may become an important diagnostic test [6, 7]. Such new technique may have positive influence in improving our knowledge of carcinogenesis and facilitate screening and early detection of diseases.

In our study, we analyzed gene expression profiles (GSE121248) downloaded from GEO database, to extract DEGs between liver cancer and normal samples. Moreover, KEGG pathways of those genes and most of the enriched GO terms that were correlated to HCC were also included in this research. In summary, our result may advance the knowledge of HCC and explore potential therapeutic targets for further studies.

## Materials and methods

### *Gene expression microarray data*

In this research, the gene expression microarray data set GSE121248 was downloaded from GEO database (GEO, <http://www.ncbi.nlm.nih.gov/geo/>), including 70 tumor samples from HCC patient and 37 adjacent normal samples [8]. We used k-nearest neighbors algorithm (k-NN) for classification and regression in pattern recognition [9]. In addition, a supplement work was conducted for other genes with similar expression profiles as those with deletion value.

### *Differentially expressed genes (DEGs)*

In the preprocessing section, the unwanted noise of the original microarray data was screened out. The original data and background correction were dealt with by the Affy [10] package in R. The *differentially* expressed mRNAs between tumor tissue and normal tissue samples were inspected using the Limma [11] package with the following criteria:  $|\text{Log}_2(\text{fold change})| > 1.5$  and false discovery rate  $< 0.05$ . Morpheus is an on-line tool used to deal with heatmap (<https://software.broadinstitute.org/morpheus>).

### *Functional and pathway enrichment analysis*

The gene product function is supplied from The Gene Ontology website [12] (GO, <http://www.geneontology.org>), a community-based bioinformatics resource using ontologies to represent ontologies to represent biological knowledge. GO provides a framework and a suit of concepts for describing the functions of gene products from all living organisms. KEGG (Kyoto Encyclopedia of Genes and Genomes, <http://www.kegg.jp/> or <http://www.genome.jp/kegg>) is used for interpretation of high-throughput data and other biological genome sequences [13]. DAVID [14] (<https://david.ncifcrf.gov>) is also a bioinformatic resources, consisted of analytic tools and biological knowledge base, aimed at organically extracting biological activity from huge gene protein statement. GO terms and KEGG pathway analyses were accomplished with DAVID to recognize DEGs.  $P < 0.05$  was executed as the cut-off value.

### *Protein-protein interactions (PPI) network construction and analysis of modules*

The STRING database (<http://string-db.org>) is a web resource aimed to predict protein-protein

interactions and provide critical assessment, which includes physical and functional associations [15]. Cytoscape [16] is an open-source software platform for visualizing biomedical networks exploration and offering researchers utility and coactive visualization interface. The tools is used for exploring interconnections of complex biological networks supported by varying annotation and experimental results, therefore accelerating research works such as forecasting gene function and reconstructing pathways. The STRING was used to evaluate PPI network mapped DEGs and Cytoscape for visualization. The combined score of greater than 0.15 was executed as the cut-off value. Finally the cut-off criterion for screening hub gene was defined as node degree 10.

### *Exploring overall survival*

'Kaplan-Meier plotter' (KM plotter), online database, was used to explore the predictive prognostic value of gene expression to overall survival (OS) in different clinical data, such as clinical grades, clinical stages, gender and race in liver cancer patients.

### *Statistical analysis*

Matrix data were analyzed by one-way ANOVA, and statistical significance was determined as  $p < 0.05$ . The results were compared between normal and tumor groups with Mann-Whitney Rank Sum Test.

## Results

### *Differentially expresses gene (DEGs)*

There were 202 DEGs between tumor samples and control samples. Comparing with adjacent tissue samples, 59 up-regulated and 143 down-regulated genes were selected in case tissue samples. The heatmap of the DEGs was shown in **Figure 1**.

### *GO enrichment terms and KEGG pathway analysis*

The study obtained totally of 132 GO enrichment terms and 16 KEGG pathways. The first 10 GO enrichment terms of the DEGs following  $P$  value were shown (**Table 1**). **Table 1** indicated clearly that the mainly GO enrichment term was the cellular components (CC), such as extracellular region, organelle membrane and extracellular space. In addition, epoxygenase P450



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**Table 1.** Top 10 enriched GO terms are sorted in ascending order by *P* value

Category	Term	Gene Number	<i>P</i> Value
CC	GO:0005576~extracellular region	47	7.04E-11
BP	GO:0019373~epoxygenase P450 pathway	8	4.33E-10
CC	GO:0031090~organelle membrane	12	1.25E-09
MF	GO:0008392~arachidonic acid epoxygenase activity	7	5.10E-09
MF	GO:0016705~oxidoreductase activity, acting on paired donors, with incorporation or reduction of molecular oxygen	10	6.28E-09
MF	GO:0005506~iron ion binding	13	6.16E-08
MF	GO:0004497~monooxygenase activity	9	1.34E-07
MF	GO:0020037~heme binding	12	1.75E-07
CC	GO:0005615~extracellular space	36	2.19E-07
BP	GO:0055114~oxidation-reduction process	23	4.31E-07

**Table 2.** The top 10 KEGG pathways in DEGs

Category	Pathway Name	Gene Number	<i>P</i> Value
KEGG_PATHWAY	hsa04610: Complement and coagulation cascades	8	3.84E-05
KEGG_PATHWAY	hsa00830: Retinol metabolism	7	2.20E-04
KEGG_PATHWAY	hsa05020: Prion diseases	5	1.11E-03
KEGG_PATHWAY	hsa01100: Metabolic pathways	30	1.26E-03
KEGG_PATHWAY	hsa00232: Caffeine metabolism	3	1.80E-03
KEGG_PATHWAY	hsa00380: Tryptophan metabolism	5	2.05E-03
KEGG_PATHWAY	hsa05204: Chemical carcinogenesis	6	4.59E-03
KEGG_PATHWAY	hsa04115: p53 signaling pathway	5	1.30E-02
KEGG_PATHWAY	hsa00982: Drug metabolism-cytochrome P450	5	1.37E-02
KEGG_PATHWAY	hsa04978: Mineral absorption	4	2.19E-02
KEGG_PATHWAY	hsa05323: Rheumatoid arthritis	5	3.19E-02
KEGG_PATHWAY	hsa00140: Steroid hormone biosynthesis	4	4.44E-02
KEGG_PATHWAY	hsa04060: Cytokine-cytokine receptor interaction	8	4.79E-02

ion binding, monooxygenase activity and heme binding composed the molecular function (MF) of the cell.

The KEGG pathways were demonstrated in **Table 2**. The top 10 KEGG enriched pathways were mostly related to metabolism, such as retinol metabolism, metabolic pathways, caffeine metabolism, tryptophan metabolism and drug metabolism-cytochrome P450. Furthermore, the other pathways may also influence on the progression of cancer via some biological process, such as the complement and coagulation cascades, Prion diseases, chemical carcinogenesis, p53 signaling pathway and mineral absorption.

### *PPI network and core genes in network*

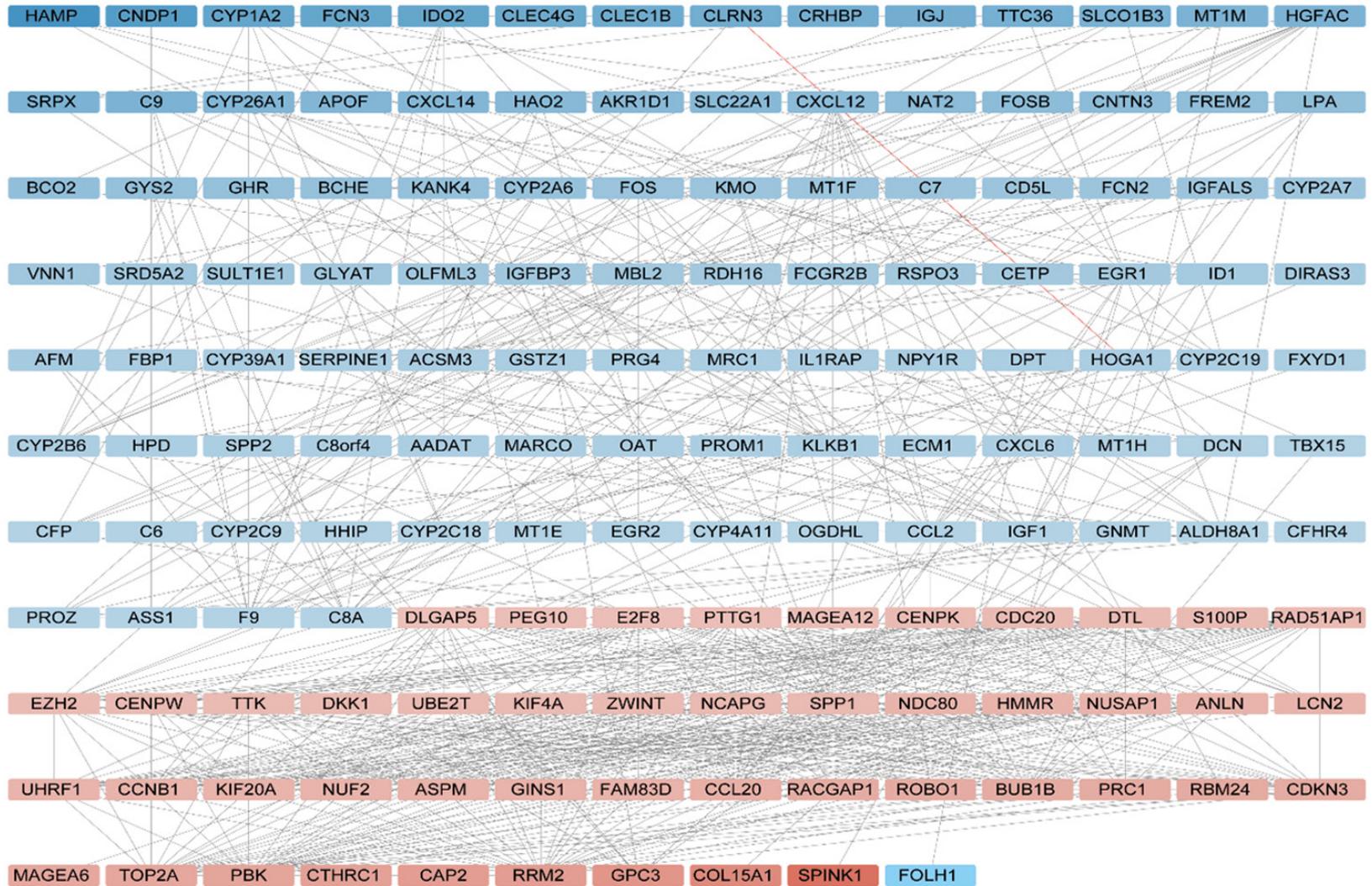
The PPI (**Figure 2**) network included 190 nodes (DEGs) and 756 edges (interactions) between

the DEGs. The core genes, had higher degree in the PPI network, might had a stronger correlation with liver cancer. **Table 3** has shown the cores genes' solid degree. Among those core genes, CDC20 had 33 degrees and 21 gene's node degree were beyond 30.

### *Overall survival based on core genes*

Firstly, the prognostic value of BUB1B was accessed in the database. The RNA-seq ID is 701 for BUB1B. OS curves were plotted for all liver cancer patients (n=364) (**Figure 3A**), BUB1B has related to poor OS in HCC patients, HR=2.01 (1.42-2.86), *P*=6.6e-05. Next, the prognostic significance of CCNB1 was evaluated in the database. The desired RNA-seq ID is 891. CCNB1 revealed a remarkable correlation with worse OS among liver cancer patients, HR=2.34 (1.55-3.54), *P*=3.4e-05 (**Figure 3B**). Similarly, EZH2 (RNA-seq ID 2146) exhibited a

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**Figure 2.** The PPI network of 190 nodes and 756 edges. The 47 red nodes had higher expression in the tumor samples; whereas the 103 blue nodes have lower expression values.

**Table 3.** The cores genes and solid degree according to the PPI network

Gene symbol	Node degree						
CYP26A1	10	C8A	15	CDKN3	30	ZWINT	31
CYP2B6	10	F9	16	HMMR	30	BUB1B	32
ALDH8A1	11	IGF1	16	KIF4A	30	CCNB1	32
C6	11	SPP1	17	PTTG1	30	EZH2	32
CYP1A2	11	GIN51	20	ASPM	31	NUSAP1	32
CYP2C9	11	CENPW	22	DLGAP5	31	CDC20	33
EGR1	11	E2F8	22	DTL	31		
HGFAC	11	CENPK	23	KIF20A	31		
FOS	12	UHRF1	23	NCAPG	31		
KLKB1	12	FAM83D	25	NDC80	31		
SERPINE1	12	ANLN	27	PBK	31		
MBL2	13	UBE2T	27	RAD51AP1	31		
CCL2	14	RACGAP1	28	RRM2	31		
CXCL12	14	NUF2	29	TOP2A	31		
SPP2	14	PRC1	29	TTK	31		

worse correlation with OS within liver cancer patients, HR=2.23 (1.56-3.19),  $P=6.8e-06$  (Figure 3C). The prognostic value of NUSAP1 was also determined in the database, RNA-seq ID 51203. A strong association was identified between NUSAP1 expression and poor OS for liver cancer, HR=1.67 (1.17-2.4),  $P=0.0046$  (Figure 3D). Figure 3E demonstrated the prognostic value of CDC20 in the database. The RNA-seq ID is 991. CDC20 was significantly correlated with poor OS for liver cancer patients, HR=2.49 (1.72-3.59),  $P=5.1e-07$ .

### Discussion

HCC's mortality has steadily increased in the last few years. It is now the fifth most common malignant tumor worldwide [17]. Hepatitis viruses, gene mutations, cell damage, alcoholic liver diseases and aflatoxin poisoning have all been identified as risk factors for HCC [18]. As a potential diagnostic tool, tumor markers have been widely used in the early diagnosis of HCC. Alejandro Forner claimed that  $\alpha$ -Fetoprotein was a brilliant star for HCC diagnosis [19]. Moreover, severe alpha-1-antitrypsin deficiency (AATD) played a key role in the development of liver disease [20]. However, further study on the mechanism of hepatocellular carcinoma is necessary for early diagnosis and optimal treatment.

In this work, a high-throughput method for genome-wide gene expression analysis was used

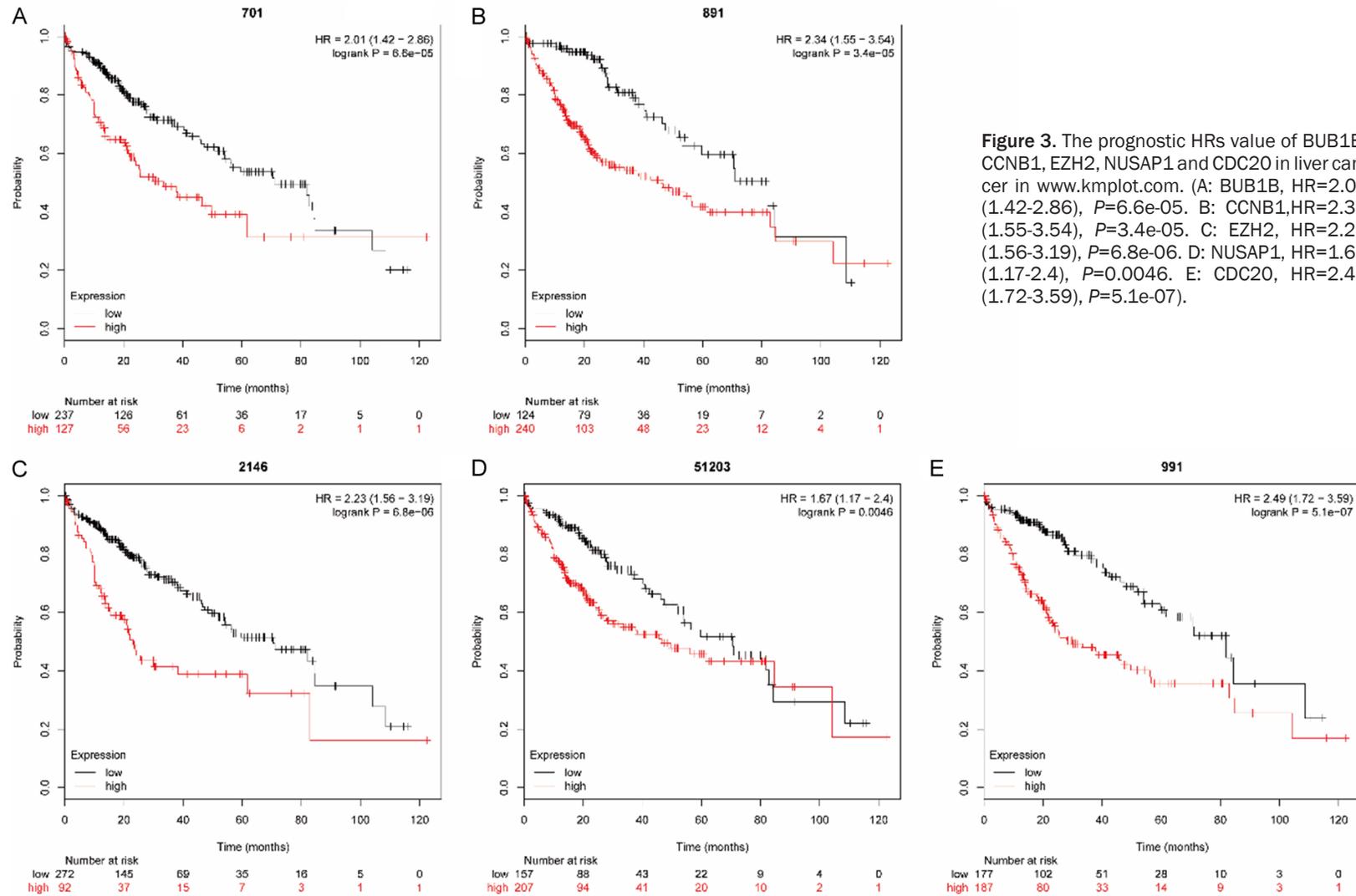
to screen 202 DEGs from HCC tissues and normal liver tissue, including 59 up-regulated genes and 143 down-regulated genes in tumor samples.

Enrichment GO analyses identified significant ontology categories including epoxygenase P450 pathway and oxidation-reduction process. The human liver microsome metabolizes arachidonic acid in NADPH and produces epoxyeicosapentaenoic acid and its hydrated metabolite dihydroxyeicosapentaenoic acid as the main reaction product [21].

These bioactive eicosanoids play a role in maintaining homeostasis in the liver. Under normal and pathophysiological conditions, human P450 cyclooxygenase (also known as CYP) and its derivatives arachidonic acid metabolites may be expressed differently, thus affecting regulation of vascular function [22]. In one study, it was found that CYP2C9 was highly expressed in patients with esophageal adenocarcinoma (including patients with early tumor stage and highly differentiated tumors). Additionally, selective inhibition of CYP2C9 was shown to reduce the proliferation of low tumor cells in in vitro experiments [23].

KEGG pathway analyzed DEGs and 13 pathways were screened out, such as retinol metabolism, tryptophan metabolism and drug metabolism-cytochrome P450, complement and coagulation cascades. Previous studies have covered that majority of these pathways were comprised in cancer progression. In human cancer cell lines of skin, oral cavity, kidney and breast, retinol acyltransferase (LRAT) was reduced compared with the normal counterparts, thus implicating aberrant retinoid metabolism in carcinogenesis [24]. Cytochrome P450 are hepatic enzymes that may activate some procarcinogens. Studies have shown that CYP1A1 increases the risk of peripheral adenocarcinoma type lung cancer [25]. And its polymorphism might be an important factor in the optimal use of selected anticancer drugs for cancer treatment [26].

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**Figure 3.** The prognostic HRs value of BUB1B, CCNB1, EZH2, NUSAP1 and CDC20 in liver cancer in [www.kmplot.com](http://www.kmplot.com). (A: BUB1B, HR=2.01 (1.42-2.86),  $P=6.6e-05$ . B: CCNB1, HR=2.34 (1.55-3.54),  $P=3.4e-05$ . C: EZH2, HR=2.23 (1.56-3.19),  $P=6.8e-06$ . D: NUSAP1, HR=1.67 (1.17-2.4),  $P=0.0046$ . E: CDC20, HR=2.49 (1.72-3.59),  $P=5.1e-07$ ).

The complement system is responsible for killing bacteria that infect the host. The secretion of proinflammatory mediators and ingestion of opsonized particles were led by complement activation [27]. On the other hand, coagulation activation occurs through two pathways, one exogenous and the other through an internal (contact activation) pathway that seems to be involved in inflammatory processes [28, 29]. In the occurrence and progression of tumor, tumor-promoting inflammation plays an important role [30, 31]. The connection between inflammation and cancer can be made both externally and internally. The external pathway increases the risk of inflammation driving cancer, while the internal pathway is driven by genes for inflammation and tumorigenesis [32].

In addition, PPI suggests that the top 5 core genes were BUB1B, CCNB1, EZH2, NUSAP1 and CDC20, which might serve as potential targets for therapy. In normal cells, BUB1B (coding BUBR1) has been shown to prevent duplication of chromosome segregation, but its role has been controversial in cancer pathogenesis. BUB1B has overexpression in prostate cancer [33], adenomatous polyposis coli [34], HCC [35] and other cancer [36, 37], which is consistent with our data; while low expression of BUB1B contributes to embryonal rhabdomyosarcoma [38], colorectal cancer [39] and other cancers [40, 41]. CCNB1 (Cyclin B1), tumor antigen, is overexpressed in many cancers [42]. The autoimmune response of CCNB1 in HCC may include the aberration of CCNB1 regulation leading to changes in product or its expression resulting in immune stimulation [43]. Wu suggest that CCNB1 may be a key target protein of human HCC cell line Lnc00312 [44]. Enhancer of zeste homolog 2 (EZH2) is a histone-lysine N-methyltransferase enzyme, which participates in histone methylation and transcriptional repression [45]. In some reports, inhibition of EZH2 function shrank malignant tumors because the tumor suppressor genes were not inhibited by EZH2 [46]. In HCC, EZH2 partially inhibits the immune response and plays a carcinogenic role [47]. Nucleolar and spindle-associated protein 1 (NUSAP1) maintains normal cellular division and participates in regulating spindle assembly through microtubule-binding and DNA-binding domains, which is an important regulator of mitosis and cell proliferation [48, 49]. Previous study indicated

that the expression of NUSAP1 at the margin of liver cancer surgery was closely related to early postoperative recurrence and can be used as an index to predict early recurrence of HCC [50]. As a cell cycle regulating kinase, CDC20, an essential cell cycle regulator, is necessary to complete mitosis. Some studies have reported that CDC20 plays a key role in gastric cancer [51] and HCC [52]. CDC20 may potentially be used as a biomarker and therapeutic target in HCC [53].

Finally, our results showed that high expression of the top 5 core genes had worsening effect on the prognosis of liver cancer patients. Zhuang [54] reported that BUB1B, CCNB1 and CDC20 could serve as predictive biomarkers for HCC, and they demonstrated that high expression of those genes was related to worse survival. As for NUSAP1, it was a valuable prognostic factor for hepatic carcinoma. Low NUSAP1 expression patients had better survival rate than high expression patients at both 6 months and 12 months (89.3% VS 33.3%, 53.6% VS 17.9%) [55]. It was reported that precise anti-tumor drugs can be developed because EZH2 promotes the occurrence and development of tumors [56]. This has the potential to improve survival in cancer patients.

### Conclusion

Our study screened DEGs and identified potential biomarkers to forecast the occurrence and development of HCC. A total of 202 DEGs were screened including BUB1B, CCNB1, EZH2, NUSAP1 and CDC20. The many functional partnerships and PPI are core of cellular processing and their classified characterization helps to deal with context in molecular system biology. Survival analysis identified 5 core genes as potential therapeutic targets in the management of HCC. However, our study lacks in vivo and in vitro validation. For future studies, the results of these bioinformatics analyses can be verified by experiments, such as Western Blot and qRT-PCR.

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**Disclosure of conflict of interest**

None.

**Address correspondence to:** Ji-Cai Chen, Department of General Surgery, The First Affiliated Hospital of Wenzhou Medical University, Wenzhou 325-000, Zhejiang, China. Tel: 13777760321; E-mail: cjc60321@163.com

**References**

[1] Parkin DM. Global cancer statistics in the year 2000. *Lancet Oncol* 2001; 2: 533-543.

[2] Global Burden of Disease Cancer Collaboration, Fitzmaurice C, Akinyemiju TF, Al Lami FH, Alam T, Alizadeh-Navaei R, Allen C, Alsharif U, Alvis-Guzman N, Amini E, Anderson BO, Aremu O, Artaman A, Asgedom SW, Assadi R, Atey TM, Avila-Burgos L, Awasthi A, Ba Saleem HO, Barac A, Bennett JR, Bensenor IM, Bhakta N, Brenner H, Cahuana-Hurtado L, Castañeda-Orjuela CA, Catalá-López F, Choi JJ, Christopher DJ, Chung SC, Curado MP, Dandona L, Dandona R, das Neves J, Dey S, Dharmaratne SD, Doku DT, Driscoll TR, Dubey M, Ebrahimi H, Edessa D, El-Khatib Z, Endries AY, Fischer F, Force LM, Foreman KJ, Gebrehiwot SW, Gopalani SV, Grosso G, Gupta R, Gyawali B, Hamadeh RR, Hamidi S, Harvey J, Hassen HY, Hay RJ, Hay SI, Heibati B, Hiluf MK, Horita N, Hosgood HD, Ilesanmi OS, Innos K, Islami F, Jakovljevic MB, Johnson SC, Jonas JB, Kasaieian A, Kassa TD, Khader YS, Khan EA, Khan G, Khang YH, Khosravi MH, Khubchandani J, Kopec JA, Kumar GA, Kutz M, Lad DP, Lafrancioni A, Lan Q, Legesse Y, Leigh J, Linn S, Lunevicius R, Majeed A, Malekzadeh R, Malta DC, Mantovani LG, McMahon BJ, Meier T, Melaku Y, Melku M, Memiah P, Mendoza W, Meretoja TJ, Mezgebe HB, Miller TR, Mohammed S, Mokdad AH, Moosazadeh M, Moraga P, Mousavi SM, Nangia V, Nguyen CT, Nong VM, Ogbo FA, Olagunju AT, Pa M, Park EK, Patel T, Pereira DM, Pishgar F, Postma MJ, Pourmalek F, Qorbani M, Rafay A, Rawaf S, Rawaf DL, Roshandel G, Safiri S, Salimzadeh H, Sanabria JR, Santric Milicevic MM, Sartorius B, Satpathy M, Sepanlou SG, Shackelford KA, Shaikh MA, Sharif-Alhoseini M, She J, Shin MJ, Shiue I, Shrimel MG, Sinke AH, Sisay M, Sligar A, Sufiyan MB, Sykes BL, Tabarés-Seisdedos R, Tessema GA, Topor-Madry R, Tran TT, Tran BX, Ukwaja KN, Vlassov VV, Vollset SE, Weiderpass E, Williams HC, Yimer NB, Yonemoto N, Younis MZ, Murray CJL, Naghavi M. Global, regional, and national

cancer incidence, mortality, years of life lost, years lived with disability, and disability-adjusted life-years for 29 cancer groups, 1990 to 2016 a systematic analysis for the global burden of disease study. *JAMA Oncol* 2018; 4: 1553-1568.

[3] Elserag HB. Hepatocellular carcinoma: epidemiology and molecular carcinogenesis. *Gastroenterology* 2007; 132: 2557-2576.

[4] Llovet JM, Brú C, Bruix J. Prognosis of hepatocellular carcinoma: the BCLC staging classification. *Semin Liver Dis* 1999; 19: 329-338.

[5] Chon YE, Jung KS, Kim MJ, Choi JY, An C, Park JY, Ahn SH, Kim BK, Kim SU, Park H, Hwang SK, Rim KS, Han KH, Kim DY. Predictors of failure to detect early hepatocellular carcinoma in patients with chronic hepatitis B who received regular surveillance. *Aliment Pharmacol Ther* 2018; 47: 1201-1212.

[6] Cheung NY. Molecular targets in gynaecological cancers. *Pathology* 2007; 39: 26-45.

[7] Magic Z, Radulovic S, Brankovic-Magic M. cDNA microarrays: identification of gene signatures and their application in clinical practice. *J BUON* 2007; 12: S39-44.

[8] Wang SM, Ooi LL, Hui KM. Identification and validation of a novel gene signature associated with the recurrence of human hepatocellular carcinoma. *Clin Cancer Res* 2007; 13: 6275-6283.

[9] Altman NS. An introduction to kernel and nearest-neighbor nonparametric regression. *Am Stat* 1992; 46: 175-185.

[10] Gautier L, Cope L, Bolstad BM, Irizarry RA. affy-analysis of affymetrix genechip data at the probe level. *Bioinformatics* 2004; 20: 307-315.

[11] Ritchie ME, Phipson B, Wu D, Hu Y, Law CW, Shi W, Smyth GK. limma powers differential expression analyses for RNA-sequencing and microarray studies. *Nucleic Acids Res* 2015; 43: e47.

[12] Gene Ontology Consortium. Gene ontology consortium: going forward. *Nucleic Acids Res* 2015; 43: D1049-1056.

[13] Kanehisa M, Sato Y, Kawashima M, Furumichi M, Tanabe M. KEGG as a reference resource for gene and protein annotation. *Nucleic Acids Res* 2016; 44: D457-462.

[14] Huang da W, Sherman BT, Lempicki RA. Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nat Protoc* 2009; 4: 44-57.

[15] Szklarczyk D, Franceschini A, Wyder S, Forslund K, Heller D, Huerta-Cepas J, Simonovic M, Roth A, Santos A, Tsafou KP, Kuhn M, Bork P, Jensen LJ, von Mering C. STRING v10: protein-protein interaction networks, integrated over the tree of life. *Nucleic Acids Res* 2015; 43: D447-452.

- [16] Su G, Morris JH, Demchak B, Bader GD. Biological network exploration with cytoscape 3. *Curr Protoc Bioinformatics* 2014; 47: 8.13.1-24.
- [17] El-Serag HB, Rudolph KL. Hepatocellular carcinoma: epidemiology and molecular carcinogenesis. *Gastroenterology* 2007; 132: 2557-2576.
- [18] Turner PC, Sylla A, Diallo MS, Castegnaro JJ, Hall AJ, Wild CP. The role of aflatoxins and hepatitis viruses in the etiopathogenesis of hepatocellular carcinoma: a basis for primary prevention in Guinea-Conakry, West Africa. *J Gastroenterol Hepatol* 2002; 17: S441-8.
- [19] Forner A, Reig M and Bruix J.  $\alpha$ -fetoprotein for hepatocellular carcinoma diagnosis: the demise of a brilliant star. *Gastroenterology* 2009; 137: 26-29.
- [20] Tanash HA, Piitulainen E. Liver disease in adults with severe alpha-1-antitrypsin deficiency. *J Gastroenterol* 2019; 54: 541-548.
- [21] Zeldin DC, Moomaw CR, Jesse N, Tomer KB, Beetham J, Hammock BD and Wu S. Biochemical characterization of the human liver cytochrome P450 arachidonic acid epoxygenase pathway. *Arch Biochem Biophys* 1996; 330: 87.
- [22] Martin S and Liao JK. Vascular protective effects of cytochrome p450 epoxygenase-derived eicosanoids. *Arch Biochem Biophys* 2005; 433: 413-420.
- [23] Schmelzle M, Dizdar L, Matthaehi H, Baldus SE, Wolters J, Lindenlauf N, Bruns I, Cadeddu RP, Kröppil F, Topp SA, Schulte am Esch J 2nd, Eisenberger CF, Knoefel WT, Stoecklein NH. Esophageal cancer proliferation is mediated by cytochrome P450 2C9 (CYP2C9). *Prostaglandins Other Lipid Mediat* 2011; 94: 25-33.
- [24] Guo X, Knudsen BS, Peehl DM, Ruiz A, Bok D, Rando RR, Rhim JS, Nanus DM and Gudas LJ. Retinol metabolism and lecithin:retinol acyltransferase levels are reduced in cultured human prostate cancer cells and tissue specimens. *Cancer Res* 2002; 62: 1654-1661.
- [25] Anttila S, Hietanen E, Vainio H, Camus AM, Gelboin HV, Park SS, Heikkilä L, Karjalainen A, Bartsch H. Smoking and peripheral type of cancer are related to high levels of pulmonary cytochrome P450IA in lung cancer patients. *Int J Cancer* 1991; 47: 681-685.
- [26] Rodriguez-Antona C, Gomez A, Karlgren M, Sim SC and Ingelman-Sundberg M. Molecular genetics and epigenetics of the cytochrome P450 gene family and its relevance for cancer risk and treatment. *Hum Genet* 2010; 127: 1-17.
- [27] Merle NS, Noe R, Halbwachs-Mecarelli L, Fremaux-Bacchi V, Roumenina LT. Complement system part II: role in immunity. *Front Immunol* 2015; 6: 257.
- [28] Davie EW, Ratnoff OD. Waterfall sequence for intrinsic blood clotting. *Science* 1964; 145: 1310-1312.
- [29] Adams RL, Bird RJ. Review article: coagulation cascade and therapeutics update: relevance to nephrology. Part 1: overview of coagulation, thrombophilias and history of anticoagulants. *Nephrology (Carlton)* 2009; 14: 462-470.
- [30] Coussens LM, Zitvogel L, Palucka AK. Neutralizing tumor-promoting chronic inflammation: a magic bullet? *Science* 2013; 339: 286-291.
- [31] Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell* 2011; 144: 646-674.
- [32] Mantovani A, Allavena P, Sica A, Balkwill F. Cancer-related inflammation. *Nature* 2008; 454: 436-444.
- [33] Fu X, Chen G, Cai ZD, Wang C, Liu ZZ, Lin ZY, Wu YD, Liang YX, Han ZD, Liu JC, Zhong WD. Overexpression of BUB1B contributes to progression of prostate cancer and predicts poor outcome in patients with prostate cancer. *Onco Targets Ther* 2016; 9: 2211-2220.
- [34] Abal M, Obrador-Hevia A, Janssen KP, Casadome L, Menendez M, Carpentier S, Barillot E, Wagner M, Ansoerge W, Moeslein G, Fsihi H, Bezrookove V, Reventos J, Louvard D, Capella G, Robine S. APC inactivation associates with abnormal mitosis completion and concomitant BUB1B/MAD2L1 up-regulation. *Gastroenterology* 2007; 132: 2448-2458.
- [35] Liu AW, Cai J, Zhao XL, Xu AM, Fu HQ, Nian H, Zhang SH. The clinicopathological significance of BUBR1 overexpression in hepatocellular carcinoma. *J Clin Pathol* 2009; 62: 1003-1008.
- [36] Slee RB, Grimes BR, Bansal R, Gore J, Blackburn C, Brown L, Gasaway R, Jeong J, Victorino J and March KL. Selective inhibition of pancreatic ductal adenocarcinoma cell growth by the mitotic MPS1 kinase inhibitor NMS-P715. *Mol Cancer Ther* 2014; 13: 307-315.
- [37] Crawford CV, Osborne MP and Katdare M. Identification of cell cycle and apoptosis related gene expression by curcumin in an in vitro model for ductal carcinoma in situ. *Cancer Res* 2005; 65.
- [38] Hanks S, Coleman K, Reid S, Plaja A, Firth H, Fitzpatrick D, Kidd A, Méhes K, Nash R, Robin N, Shannon N, Tolmie J, Swansbury J, Irrthum A, Douglas J, Rahman N. Constitutional aneuploidy and cancer predisposition caused by biallelic mutations in BUB1B. *Nat Genet* 2004; 36: 1159-1161.
- [39] Burum-Auensen E, DeAngelis PM, Schjøberg AR, Røislien J, Mjåland O, Clausen OP. Reduced level of the spindle checkpoint protein BUB1B is associated with aneuploidy in colorectal cancers. *Cell Prolif* 2008; 41: 645-659.

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- [40] Sánchez-Espiridión B, Sánchez-Aguilera A, Montalbán C, Martín C, Martínez R, González-Carrero J, Poderos C, Bellas C, Fresno MF, Morante C, Mestre MJ, Mendez M, Mazorra F, Conde E, Castaño A, Sánchez-Godoy P, Tomas JF, Morente MM, Piris MA, García JF; Spanish Hodgkin's Lymphoma Study Group. A TaqMan low-density array to predict outcome in advanced Hodgkin's lymphoma using paraffin-embedded samples. *Clin Cancer Res* 2009; 15: 1367-1375.
- [41] Park HY, Jeon YK, Shin HJ, Kim IJ, Kang HC, Jeong SJ, Chung DH, Lee CW. Differential promoter methylation may be a key molecular mechanism in regulating BubR1 expression in cancer cells. *Exp Mol Med* 2007; 39: 195-204.
- [42] None. Cyclin B1 (CCNB1). SciBX: Science-Business eXchange 2009; 2.
- [43] Covini G, Chan EK, Nishioka M, Morshed SA, Reed SI, Tan EM. Immune response to cyclin B1 in hepatocellular carcinoma. *Hepatology* 1997; 25: 75-80.
- [44] Wu J, Zhou X, Fan Y, Cheng X, Lu B, Chen Z. Long non-coding RNA 00312 downregulates cyclin B1 and inhibits hepatocellular carcinoma cell proliferation in vitro and in vivo. *Biochem Biophys Res Commun* 2018; 497: 173-180.
- [45] Viré E, Brenner C, Deplus R, Blanchon L, Fraga M, Didelot C, Morey L, Van Eynde A, Bernard D, Vanderwinden JM, Bollen M, Esteller M, Di Croce L, de Launoit Y, Fuks F. The polycomb group protein EZH2 directly controls DNA methylation. *Nature* 2006; 439: 871-874.
- [46] Ramakrishnan S, Granger V, Rak M, Hu Q, Attwood K, Aquila L, Krishnan N, Osiecki R, Azabdaftari G, Guru K, Chatta G, Gueron G, McNally L, Ohm J, Wang J, Woloszynska A. Inhibition of EZH2 induces NK cell-mediated differentiation and death in muscle-invasive bladder cancer. *Cell Death Differ* 2019; [Epub ahead of print].
- [47] Bugide S, Green MR, Wajapeyee N. Inhibition of Enhancer of zeste homolog 2 (EZH2) induces natural killer cell-mediated eradication of hepatocellular carcinoma cells. *Proc Natl Acad Sci U S A* 2018; 115: E3509-E3518.
- [48] Raemaekers T, Ribbeck K, Beaudouin J, Ann-aert W, Van Camp M, Stockmans I, Smets N, Bouillon R, Ellenberg J, Carmeliet G. NuSAP, a novel microtubule-associated protein involved in mitotic spindle organization. *J Cell Biol* 2003; 162: 1017-1029.
- [49] Vanden Bosch A, Raemaekers T, Denayer S, Torrekens S, Smets N, Moermans K, Dewerchin M, Carmeliet P, Carmeliet G. NuSAP is essential for chromatin-induced spindle formation during early embryogenesis. *J Cell Sci* 2010; 123: 3244-3255.
- [50] Zhang M, Yang D, Liu X, Liu Y, Liang J, He H, Zhong K, Lin L, Tao G, Zhang C, Zhou J. [Expression of Nusap1 in the surgical margins of hepatocellular carcinoma and its association with early recurrence]. *Nan Fang Yi Ke Da Xue Xue Bao* 2013; 33: 937-938, inside back cover.
- [51] Kim Y, Choi JW, Lee JH, Kim YS. Spindle assembly checkpoint MAD2 and CDC20 overexpressions and cell-in-cell formation in gastric cancer and its precursor lesions. *Hum Pathol* 2019; 85: 174-183.
- [52] Kato T, Daigo Y, Aragaki M, Ishikawa K, Sato M, Kaji M. Overexpression of CDC20 predicts poor prognosis in primary non-small cell lung cancer patients. *J Surg Oncol* 2012; 106: 423-430.
- [53] Wang Y, Ruan Z, Yu S, Tian T, Liang X, Jing L, Li W, Wang X, Xiang L, Claret FX, Nan K, Guo H. A four-methylated mRNA signature-based risk score system predicts survival in patients with hepatocellular carcinoma. *Aging (Albany NY)* 2019; 11: 160-173.
- [54] Zhuang L, Yang Z, Meng Z. Upregulation of BUB1B, CCNB1, CDC7, CDC20, and MCM3 in tumor tissues predicted worse overall survival and disease-free survival in hepatocellular carcinoma patients. *Biomed Res Int* 2018; 2018: 7897346.
- [55] Zhang M, Yang D, Liu X, Liu Y, Liang J. [Expression and clinical significance of Nusap1 in hepatic carcinoma]. *Zhong Nan Da Xue Xue Bao Yi Xue Ban* 2013; 38: 876-881.
- [56] Yamagishi M, Uchamaru K. Targeting EZH2 in cancer therapy. *Current Opinion in Oncology* 2017; 29: 375-381.