# Original Article Serum miR-1826 expression serves as a diagnostic biomarker and inhibits cell proliferation, migration, and invasion of lung cancer

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**Abstract:** Many studies have indicated that abnormal expression levels of microRNAs play an important role in the development of several cancer types. The current study investigated the roles of miR-1826 in lung cancer. Expression levels of miR-1826 were detected using quantitative real-time polymerase chain reaction (qRT-PCR) in the serum of lung cancer patients and lung cancer cell lines. Association of miR-1826 with clinicopathological parameters of patients was analyzed using  $\chi^2$  tests. Receiver operating characteristic curve (ROC) analysis was adopted to estimate the potential diagnostic value of miR-1826. Cell proliferation was measured using MTT assays. Cell migration and invasion were identified by Transwell assays. Moreover, miR-1826 expression levels were significantly decreased in the serum of lung cancer patients and cell lines, compared to controls (all *P* < 0.05). Expression of miR-1826 was significantly associated with lymph node metastasis (*P* = 0.015) and TNM stage (*P* = 0.033). ROC curve analysis results showed that miR-1826 possesses diagnostic value in differentiating lung cancer patients from healthy individuals, with an area under the ROC curve (AUC) of 0.873, showing promising sensitivity and specificity. Using miR-1826 mimics and inhibitors, cell proliferation, migration, and invasion were inhibited by miR-1826 overexpression, but promoted by a reduction of miR-1826 (all *P* < 0.05). Downregulation of miR-1826 may serve as a novel marker for diagnosis of lung cancer. It may also be involved in tumor progression of lung cancer.

Keywords: miR-1826, lung cancer, diagnosis, progression

#### Introduction

Lung cancer, a common primary malignant tumor, is a leading cause of cancer-related deaths, worldwide, with high morbidity and mortality rates [1, 2]. Of all types of lung cancer, non-small cell cancer (NSCLC) accounts for approximately 85% [3]. Many studies have demonstrated that cigarette smoking is one of the leading high-risk factors in the development of lung cancer [4-6]. Lung cancer is a severe health burden in China, with incidence and mortality rates growing rapidly [7]. Although treatment strategies for lung cancer have greatly improved, prognosis of patients in the advanced stage remains unsatisfactory. Therefore, it is crucial to identify effective diagnostic biomarkers and novel therapies.

MicroRNAs (miRNAs) are a large class of small, non-coding, and endogenous RNA molecules that regulate gene expression through interaction with the 3' untranslated regions (3'UTRs) of target mRNAs [8]. Abnormal expression of miR-NAs has been strongly associated with tumor development and involved in the regulation of diverse physiological and pathological functions, including regulating cell proliferation, differentiation, migration, invasion, apoptosis, and tumor metastasis [9-11]. Many studies have demonstrated that miRNAs function as either oncogenes or tumor suppressors in the tumorigenesis process [12-16]. Other studies have indicated that miRNAs are associated with lung cancer [17, 18]. Accumulated evidence has suggested that miRNAs might be diagnostic or prognostic biomarkers for tumor detection and treatment therapies in cancers. Moreover, miR-1826 was found to be abnormally expressed and could serve as a non-invasive biomarker for diagnosis and prognosis in a number of cancers, including colorectal cancer and renal cell carcinoma [19, 20]. Abnormal

Parameters	Cases No. (n = 124)	miR-1826 expression		_
		Low (n = 74)	High (n = 50)	P
Age				0.338
< 60	58	32	26	
≥ 60	66	42	24	
Gender				0.702
Male	72	44	28	
Female	52	30	22	
Tumor size				0.709
< 5 cm	67	41	26	
≥ 5 cm	57	33	24	
Smoking status				0.202
No	73	47	26	
Yes	51	27	24	
Differentiation				0.10
Well + Moderate	63	42	21	
Poor	61	32	29	
Lymph node metastasis				0.015
Negative	68	34	34	
Positive	56	40	16	
TNM stage				0.033
-	70	36	34	
III-IV	54	38	16	

**Table 1.** Association between miR-1826 expression and
 clinicopathological variables of lung cancer patients

expression of miR-1826 was also found to be involved in the progression of various cancers, such as bladder cancer and renal cancer [21, 22]. However, the roles of miR-1826 in lung cancer remain elusive.

The present study detected expression levels of miR-1826 in the serum of lung cancer patients and healthy controls, as well as in cell lines. The effects of miR-1826 on biological behaviors of cancer cells were also assessed. The aim of this study was to identify the roles of miR-1826 in lung cancer.

#### Materials and methods

# Patients and specimens

A total of 124 patients with lung cancer and 115 healthy controls, hospitalized or physically examined, were collected at People's Hospital of Wenshang Country, from May 2010 to April 2017. The patients had not received chemotherapy, radiotherapy, or surgical resections before sampling. Whole blood specimens were collected from each participant during physical examinations. The serum was reserved by centrifugation. Serum samples were put into blood collection tubes of EDTA and stored at -80°C until further use. Detailed clinicopathological characteristics of the patients are collected and summarized in **Table 1**. This study was approved by the Ethics Committee of People's Hospital of Wenshang Country. Written informed consent was obtained from all participants in advance.

#### Cell lines and transfection

Human lung cancer cell lines (A549, H1299, H460, and H520) and normal bronchial epithelial 16HBE cell lines were obtained from American Type Culture Collection (ATCC, Manassas, VA, USA). These cells were grown in RPMI-1640 medium with 10% fetal bovine serum (FBS). They were incubated at 37°C in a humidified incubator with 5%  $CO_2$ . They were seeded into 6-well plates at a density of 1 × 10<sup>5</sup> cells per well for 24 hours, then transfected with

miR-1826 mimics, miR-1826 inhibitors, or respective negative controls (NC) using Lipofectamine 2000 (Invitrogen; Thermo Fisher Scientific, Inc.), according to manufacturer's instructions. Untreated cells were used as blank controls. After 48 hours of transfection, the transfection efficiency of cells was detected using quantitative real-time PCR (qRT-PCR). The cells were then used for further analysis.

# RNA isolation and qRT-PCR analysis

Total RNA was extracted from serum samples of lung cancer patients and healthy controls, as well as from cell lines, using Trizol Reagent (Invitrogen, Carlsbad, CA, USA), according to manufacturer's instructions. The RNA was purified to an OD A260/A280 ratio close to 2.0. Complementary DNA (cDNA) synthesis was performed using the Transcriptor First Strand cDNA Synthesis Kit (Roche, Vilvoord, Brussel, Belgium). Afterward, the qRT-PCR reaction was carried out with the SYBR Green PCR master mix (Applied Biosystems, USA) in an ABI PRISM 7900 HT system (Applied Biosystems), according to manufacturer's instructions. U6 was used as an endogenous control. Relative quantification of miR-1826 expression levels were calculated with the  $2^{-\Delta\Delta Ct}$  method.

### Cell proliferation assay

Cell viability was detected using the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assays. Cells were seeded in 96-well culture plates after transfection (5 ×  $10^4$  cells per well). After incubation for 0, 24, 48, 72, and 96 hours, 20 mµl MTT solution (Sigma-Aldrich, 5 mg/mL) was added to each well for 4 hours at 37°C. Subsequently, the medium with MTT was removed and 100 µl dimethyl sulfoxide (DMSO, Sigma-Aldrich) was added to each well, dissolving formazan crystals. Absorbance was measured at 490 nm using a spectrophotometer (Multiskan MK3, Thermo, Waltham, MA).

#### Cell migration and invasion assays

Cell migration and invasion assays were performed using Transwell chambers (Corning, Tewksbury, MA, USA). Eight nm pore size culture inserts were placed into the 24-well plates to separate the top and the lower chambers. For migration assays,  $1 \times 10^5$  cells were plated in the top chamber in 100 µl FBS-free medium. For invasion assays, 2 × 10<sup>5</sup> cells were plated in the top chamber precoated with Matrigel (Bedford, MA). The lower chambers were filled with 600 µl RMPI 1640 medium containing 20% FBS as a chemoattractant. Cells were incubated at 37°C for 24 hours. Next, nonmigrated/non-invaded cells were removed using a cotton swab. Migrated/invading cells on the lower side of the inserts were stained with formaldehyde and 0.1% crystal violet. Cells were then counted under a light microscope (Olympus, Tokyo, Japan).

# Statistical analysis

Statistical analyses were performed using SPSS 21.0 software (SPSS, Inc., Chicago, IL, USA) and GraphPad Prism 5.0 software (GraphPad Software, Inc., Chicago, USA). Data are presented as mean ± standard deviation (SD). Data for group comparisons were analyzed with paired Student's t-tests or one-way analysis of variance (ANOVA). The relationship between miR-1826 expression and clinicopathological

parameters of patients was analyzed using  $\chi^2$  tests. The diagnostic value of serum miR-1826 was evaluated through the establishment of a receiver operating characteristic curve (ROC), calculating the area under the ROC curve (AUC). P < 0.05 indicates statistical significance. All experiments were repeated at least three times.

### Results

# Expression of miR-1826 in the serum of lung cancer patients and cell lines

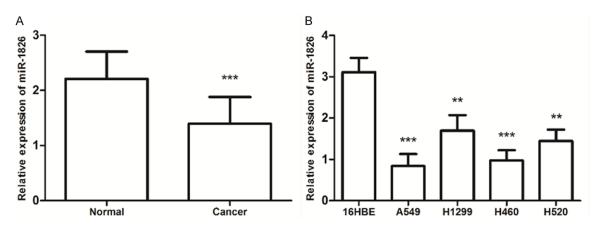
This study detected expression levels of miR-1826 in the serum samples of lung cancer patients and healthy controls, as well as in the cell lines, employing qRT-PCR. As shown in **Figure 1A**, serum miR-1826 expression was significantly downregulated in lung cancer patients, compared with healthy controls (P < 0.001). Similarly, compared to normal bronchial epithelial 16HBE cell lines, miR-1826 showed lower expression levels in lung cancer cell lines (all P < 0.01, **Figure 1B**).

Correlation of circulating miR-1826 with clinicopathological characteristics in lung cancer patients

Analyzing the association between miR-1826 expression and clinicopathological parameters of lung cancer patients, the lung cancer patients were divided into a high-expression group (n = 50) and low-expression group (n = 74), according to the mean value of miR-1826 expression levels (1.396). As shown in **Table 1**, expression of miR-1826 was associated with lymph node metastasis (P = 0.015) and TNM stage (P = 0.030). However, no positive correlation was observed between miR-1826 expression and other parameters, such as age, gender, tumor size, smoking status, and differentiation (all P > 0.05, **Table 1**).

# Diagnostic value of miR-1826 for lung cancer

The ROC curve was established to estimate the diagnostic value of miR-1826 using serum miR-1826 expression levels of lung cancer patients and healthy individuals. Corresponding healthy individuals were used as controls to produce the ROC curve (**Figure 2**). As shown in **Figure 2**, the AUC value was 0.873, yielding sensitivity of 70.2% and specificity of 90.4%, with a cutoff



**Figure 1.** Expression of miR-1826, measured by qRT-PCR, in the serum of lung cancer patients and cell lines. A. Serum expression of miR-1826 was decreased in lung cancer patients, compared to that in healthy controls (\*\*\*P < 0.001); B. Expression of miR-1826 was lower in lung cancer cell lines than in normal lung cancer cells (\*P < 0.05, \*\*P < 0.01, \*\*\*P < 0.001).

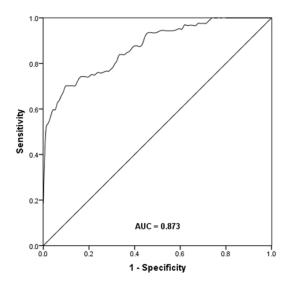


Figure 2. Diagnostic value of miR-1826 via ROC. It showed an AUC of 0.873, combined with a sensitivity of 70.2% and a specificity of 90.4%.

value of 1.570. Results showed that miR-1826 provided relative accuracy in differentiating lung cancer patients from healthy individuals.

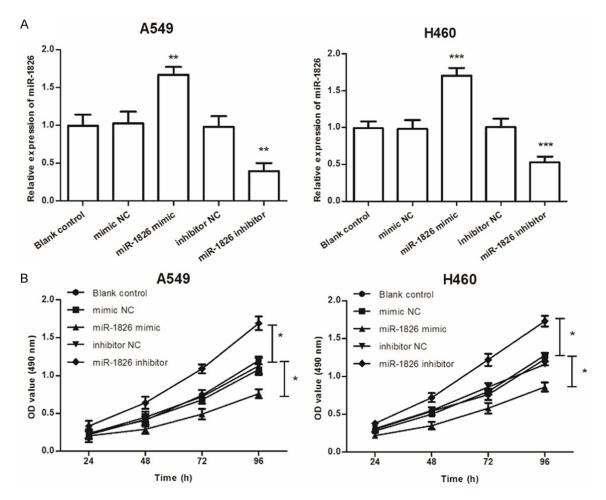
# Effects of miR-1826 on lung cancer cell proliferation, migration, and invasion

Investigating the roles of miR-1826 in lung cancer cells, the effects of miR-1826 on lung cancer cell proliferation were examined *in vitro*. Moreover, miR-1826 mimics or mimic NC and miR-1826 inhibitors or inhibitor NC were transfected into A549 and H460 cells, which showed extremely low miR-1826 expression. Subse-

quent to transfection, expression of miR-1826 in these cells was verified by gRT-PCR. Results showed that expression levels of miR-1826 were significantly increased in the cells transfected with the miR-1826 mimics, but expression levels of miR-1826 were decreased in cells transfected with miR-1826 inhibitors, compared with blank controls (all P < 0.01, Figure **3A**). MTT assays were performed to evaluate the effects of miR-1826 on lung cancer cell proliferation. Results showed that cell proliferation abilities were markedly inhibited by miR-1826 mimics, while promoted by miR-1826 inhibitors (all P < 0.05, Figure 3B). In addition to cell proliferation, the effects of miR-1826 on cell migration and invasion were examined using Transwell assays. As shown in Figure 4, overexpression of miR-1826 by miR-1826 mimics decreased cell migration and invasion abilities. Silencing of miR-1826 by miR-1826 inhibitors promoted the migration and invasion abilities in both A549 and H460 cells, compared with blank controls (all P < 0.05).

#### Discussion

Lung cancer continues to be the most common type of cancer and a major oncologic public health burden worldwide. Despite advances in standard treatments, such as surgery, radiotherapy, and chemotherapy, 5-year survival rates of patients at the advanced stage remain unsatisfactory [23-25]. Most patients are initially diagnosed at an advanced stage due to the lack of effective early diagnosis methods. Early detection and treatments are essential to



**Figure 3.** Effects of miR-1826 on cell proliferation in A549 and H460 cells. A. qRT-PCR analysis of miR-1826 in A549 and H460 cells. Expression of miR-1826 was decreased in cells transfected with miR-1826 mimics, while increased in cells transfected with miR-1826 inhibitors; B. Cell proliferation was suppressed by overexpression of miR-1826 but was enhanced by knockdown of miR-1826 in both A549 and H460 cells. (\*P < 0.05, \*\*P < 0.01, \*\*\*P < 0.001).

prolonging the overall survival of lung cancer patients. Thus, accurate noninvasive sensitive markers for lung cancer detection or treatments are urgently needed.

Substantial evidence has demonstrated that miRNAs have diagnostic or prognostic significance in a variety of human malignancies [26-30]. In lung cancer, some miRNAs have also been identified. Expression of serum and tissue miR-421 has been found to be upregulated in NSCLC patients, serving as a prognostic biomarker in NSCLC. Knockdown of miR-421 has been shown to suppress proliferation, migration, and invasion of NSCLC cells [31]. Serum miR-411 has been considered to have clinical potential as a non-invasive diagnostic biomarker for NSCLC patients [32]. In the present study, expression patterns of miR-1826 in the serum of lung cancer patients were investigated. This study showed that miR-1826 was significantly downregulated in the serum of lung cancer patients, compared with healthy controls, indicating that miR-1826 may act as a suppressor gene in lung cancer. Moreover, miR-1826 may be related to the development of lung cancer. Analysis results of the correlation between miR-1826 expression and patient clinicopathological characteristics showed that miR-1826 expression was significantly associated with lymph node metastasis and TNM stage. This suggests that miR-1826 expression was essential for the progression and metastasis of lung cancer. Based on altered miR-1826 expression in lung cancer patients and healthy controls, this study further analyzed whether miR-1826 has diagnostic value to distinguish lung cancer patients from healthy individuals. Based on

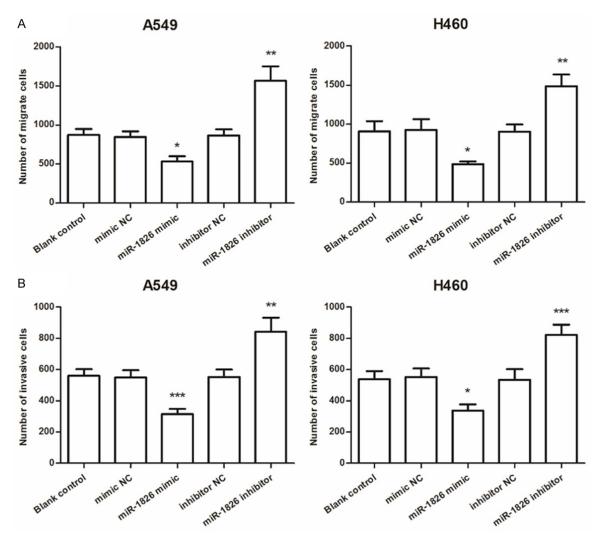


Figure 4. Results of migration and invasion assays for A549 and H460 cells. A, B. Overexpression of miR-1826 by miR-1826 mimics could inhibit cell migration and invasion, but downregulation of miR-1826 expression could promote cell migration and invasion (\*P < 0.05, \*\*P < 0.01, \*\*\*P < 0.001).

ROC curve analysis results with a high AUC value, as well as sensitivity and specificity, serum miR-1826 showed the ability to discriminate lung cancer patients from healthy individuals, with a relatively high diagnostic performance.

In addition, miRNAs have been shown to be involved in numerous aspects of cancer biology, including cell proliferation, apoptosis, migration, and invasion [33, 34]. For example, downregulation of miR-486-5p has been found in NSCLC. This could inhibit the proliferation and invasion of NSCLC cells [17]. Previous studies have indicated that identification of cancerrelated miRNAs and corresponding direct target genes is essential for lung carcinogenesis and progression, as well as improvement of lung cancer treatments. The current study explored the functional roles of miR-1826 in progression of lung cancer. The effects of miR-1826 on lung cancer cell proliferation, migration, and invasion were explored by regulating miR-1826 expression using miR-1826 mimics and inhibitors. Results showed that overexpression of miR-1826 by miR-1826 mimics could inhibit cell proliferation, migration, and invasion. Conversely, reduction of miR-1826 by miR-1826 inhibitors promoted cell proliferation, migration, and invasion, migration, and invasion in both A549 and H460 cell lines.

In recent years, the roles of miR-1826 have been reported in other malignancies. A systematic review study by Gu and co-workers [20] revealed that miR-1826 expression was downregulated in renal cell carcinoma and associated with poor prognosis. A study by Hu and his colleagues [35] indicated that miR-1826 was upregulated in colorectal cancer and miR-1826 inhibitors could suppress cell proliferation, migration, and invasion, as well as promote apoptosis. These studies have suggested that miR-1826 may act as either an oncogene or a tumor suppressor, depending on the cancer type. In the studies by Hiroshi Hirata and coworkers, they searched for miRNAs targeting beta-catenin, MEK-ERK pathway genes, and several growth factors (VEGFs). They observed that miR-1826 played an important role as a tumor suppressor by downregulating betacatenin and MEK1 in renal cancers and by CTNNB1/MEK1/VEGFC downregulation in BC [21, 22]. It was speculated that miR-1826 may also play a tumor suppressor role by targeting beta-catenin and MEK1 expression in lung cancer. However, potential molecular mechanisms of miR-1826 in lung cancer were not examined in the present study. Future studies should confirm the roles of miR-1826 in progression of lung cancer.

In conclusion, decreased expression of miR-1826 was shown to be tightly associated with cancer cell proliferation, migration, and invasion of lung cancer. Thus, it may be a potential predictive diagnostic biomarker for early detection of lung cancer.

#### Disclosure of conflict of interest

#### None.

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#### References

- [1] Miller KD, Siegel RL, Lin CC, Mariotto AB, Kramer JL, Rowland JH, Stein KD, Alteri R, Jemal A. Cancer treatment and survivorship statistics, 2016. CA Cancer J Clin 2016; 66: 271-89.
- [2] Nanavaty P, Alvarez MS, Alberts WM. Lung cancer screening: advantages, controversies, and applications. Cancer Control 2014; 21: 9-14.
- Pao W, Girard N. New driver mutations in nonsmall-cell lung cancer. Lancet Oncol 2011; 12: 175-80.
- [4] Shiono S, Katahira M, Abiko M, Sato T. Smoking is a perioperative risk factor and prognostic

factor for lung cancer surgery. Gen Thorac Cardiovasc Surg 2015; 63: 93-8.

- [5] de Groot P, Munden RF. Lung cancer epidemiology, risk factors, and prevention. Radiol Clin North Am 2012; 50: 863-76.
- [6] Molina JR, Yang P, Cassivi SD, Schild SE, Adjei AA. Non-small cell lung cancer: epidemiology, risk factors, treatment, and survivorship. Mayo Clin Proc 2008; 83: 584-94.
- [7] Hong QY, Wu GM, Qian GS, Hu CP, Zhou JY, Chen LA, Li WM, Li SY, Wang K, Wang Q, Zhang XJ, Li J, Gong X, Bai CX. Prevention and management of lung cancer in China. Cancer 2015; 121 Suppl 17: 3080-3088.
- [8] Shukla GC, Singh J, Barik S. MicroRNAs: processing, maturation, target recognition and regulatory functions. Mol Cell Pharmacol 2011; 3: 83-92.
- [9] Singh RP, Massachi I, Manickavel S, Singh S, Rao NP, Hasan S, Mc Curdy DK, Sharma S, Wong D, Hahn BH, Rehimi H. The role of miRNA in inflammation and autoimmunity. Autoimmun Rev 2013; 12: 1160-5.
- [10] Chen L, Song J, Cui J, Hou J, Zheng X, Li C, Liu L. microRNAs regulate adipocyte differentiation. Cell Biol Int 2013; 37: 533-46.
- [11] Takasaki S. Roles of microRNAs in cancers and development. Methods Mol Biol 2015; 1218: 375-413.
- [12] Lin HM, Castillo L, Mahon KL, Chiam K, Lee BY, Nguyen Q, Boyer MJ, Stockler MR, Pavlakis N, Marx G, Mallesara G, Gurney H, Clark SJ, Swarbrick A, Daly RJ, Horvath LG. Circulating microRNAs are associated with docetaxel chemotherapy outcome in castration-resistant prostate cancer. Br J Cancer 2014; 110: 2462-71.
- [13] Irmak-Yazicioglu MB. Mechanisms of MicroR-NA deregulation and MicroRNA targets in gastric cancer. Oncol Res Treat 2016; 39: 136-9.
- [14] Callegari E, Elamin BK, Sabbioni S, Gramantieri L, Negrini M. Role of microRNAs in hepatocellular carcinoma: a clinical perspective. Onco Targets Ther 2013; 6: 1167-78.
- [15] Clancy C, Joyce MR, Kerin MJ. The use of circulating microRNAs as diagnostic biomarkers in colorectal cancer. Cancer Biomark 2015; 15: 103-113.
- [16] Lodewijk L, Prins AM, Kist JW, Valk GD, Kranenburg O, Rinkes IH, Vriens MR. The value of miRNA in diagnosing thyroid cancer: a systematic review. Cancer Biomark 2012; 11: 229-38.
- [17] Yu S, Geng S, Hu Y. miR-486-5p inhibits cell proliferation and invasion through repressing GAB2 in non-small cell lung cancer. Oncol Lett 2018; 16: 3525-3530.
- [18] Zheng W, Zhao J, Tao Y, Guo M, Ya Z, Chen C, Qin N, Zheng J, Luo J, Xu L. MicroRNA-21: a promising biomarker for the prognosis and di-

agnosis of non-small cell lung cancer. Oncol Lett 2018; 16: 2777-2782.

- [19] Xu Z, Xi T, Han Y, Guo X, Liu F, Jiang M, Wan D, Xue X, He S, Ren R, Li W, Zhi Q. Circulating miR-1826 in plasma correlates with circulating tumor cells and is a prognostic marker in colorectal cancer. Tumour Biol 2017; 39: 1010428317705333.
- [20] Gu L, Li H, Chen L, Ma X, Gao Y, Li X, Zhang Y, Fan Y, Zhang X. MicroRNAs as prognostic molecular signatures in renal cell carcinoma: a systematic review and meta-analysis. Oncotarget 2015; 6: 32545-60.
- [21] Hirata H, Hinoda Y, Ueno K, Shahryari V, Tabatabai ZL, Dahiya R. MicroRNA-1826 targets VEGFC, beta-catenin (CTNNB1) and MEK1 (MAP2K1) in human bladder cancer. Carcinogenesis 2012; 33: 41-48.
- [22] Hirata H, Hinoda Y, Ueno K, Nakajima K, Ishii N, Dahiya R. MicroRNA-1826 directly targets beta-catenin (CTNNB1) and MEK1 (MAP2K1) in VHL-inactivated renal cancer. Carcinogenesis 2012; 33: 501-508.
- [23] Schabath MB, Nguyen A, Wilson P, Sommerer KR, Thompson ZJ, Chiappori AA. Temporal trends from 1986 to 2008 in overall survival of small cell lung cancer patients. Lung Cancer 2014; 86: 14-21.
- [24] Zhou C, Wu YL, Chen G, Feng J, Liu XQ, Wang C, Zhang S, Wang J, Zhou S, Ren S, Lu S, Zhang L, Hu C, Luo Y, Chen L, Ye M, Huang J, Zhi X, Zhang Y, Xiu Q, Ma J, You C. Final overall survival results from a randomised, phase III study of erlotinib versus chemotherapy as firstline treatment of EGFR mutation-positive advanced non-small-cell lung cancer (OPTIMAL, CTONG-0802). Ann Oncol 2015; 26: 1877-83.
- [25] Schad F, Thronicke A, Steele ML, Merkle A, Matthes B, Grah C, Matthes H. Overall survival of stage IV non-small cell lung cancer patients treated with Viscum album L. in addition to chemotherapy, a real-world observational multicenter analysis. PLoS One 2018; 13: e0203058.
- [26] Bertoli G, Cava C, Castiglioni I. MicroRNAs: new biomarkers for diagnosis, prognosis, therapy prediction and therapeutic tools for breast cancer. Theranostics 2015; 5: 1122-1143.

- [27] Deng Y, Chen Y. Increased expression of miR-29a and Its prognostic significance in patients with cholangiocarcinoma. Oncol Res Treat 2017; 40: 128-132.
- [28] Hussein S, Mosaad H, Rashed HE, El-Anwar MW. Up-regulated miR-221 expression as a molecular diagnostic marker in laryngeal squamous cell carcinoma and its correlation with Apaf-1 expression. Cancer Biomark 2017; 19: 279-287.
- [29] Yu Y, Zuo J, Tan Q, Zar Thin K, Li P, Zhu M, Yu M, Fu Z, Liang C, Tu J. Plasma miR-92a-2 as a biomarker for small cell lung cancer. Cancer Biomark 2017; 18: 319-327.
- [30] Huang LL, Huang LW, Wang L, Tong BD, Wei Q, Ding XS. Potential role of miR-139-5p in cancer diagnosis, prognosis and therapy. Oncol Lett 2017; 14: 1215-1222.
- [31] Li Y, Cui X, Zhang T, Li S. Upregulated expression of miR-421 is associated with poor prognosis in non-small-cell lung cancer. Cancer Manag Res 2018; 10: 2627-2633.
- [32] Wang SY, Li Y, Jiang YS, Li RZ. Investigation of serum miR-411 as a diagnosis and prognosis biomarker for non-small cell lung cancer. Eur Rev Med Pharmacol Sci 2017; 21: 4092-4097.
- [33] Xiang J, Wu Y, Li DS, Wang ZY, Shen Q, Sun TQ, Guan Q, Wang YJ. miR-584 suppresses invasion and cell migration of thyroid carcinoma by regulating the target oncogene ROCK1. Oncol Res Treat 2015; 38: 436-40.
- [34] Cheng JZ, Chen JJ, Wang ZG, Yu D. MicroR-NA-185 inhibits cell proliferation while promoting apoptosis and autophagy through negative regulation of TGF-beta1/mTOR axis and HOXC6 in nasopharyngeal carcinoma. Cancer Biomark 2018; 23: 107-123.
- [35] Hu Y, Yi B, He S, Gao L, Liu P, Wang L, Yu J, Wan D, Zhou J, Zhu X, Zhi Q. Clinical significance of miR-1826 as a novel prognostic biomarker in colorectal cancer. Anticancer Agents Med Chem 2016; 16: 1109-16.