Original Article GDF-15 promotes proliferation of vascular smooth muscle cells through MAPK-activated protein kinase pathways

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Abstract: Growth differentiation factor 15 (GDF-15) is a multifunctional member of the TGF-b superfamily. Accumulating evidence indicates that GDF15 is an independent biomarker for the cardiovascular disease, and is closely involved in the development of atherosclerotic lesions induced by ox-LDL, such as inhibition of macrophage infiltration and increase of endothelial cell proliferation, while whether GDF-15 also participates into the VSMCs proliferation is still lack of systemic analysis. In the present study, ox-LDL was found to induce GDF15 expression and secretion in cultured rat VSMCs in a p38- and ERK1/2-dependent manner. Inhibition of ox-LDL-induced GDF-15 expression by siRNA transfection by lentivirus demonstrated that adaptively induced GDF15 played a proliferative role in VSMCs, and overexpression of GDF-15 alone was adequate to induce VSMCs proliferation. Further application of inhibitors of p38- and ERK1/2 attenuated the GDF-15 expression induced by ox-LDL. All data suggest that GDF15 is a proliferative molecule that gives rise to VSMCs proliferation secondary to ox-LDL stimulation *in vitro*.

Keywords: Atherosclerosis, GDF-15, MAPK kinases, VSMCs proliferation

Introduction

Atherosclerosis is a progressive disease in which vascular smooth muscle cells (VSMCs) mainly derived from the medial layer of the blood vessel play a pivotal role in atherogenesis. VSMCs proliferation is one of the important components of atherosclerosis [1]. The oxidized low-density lipoprotein (ox-LDL) as a major contributing factor promotes the atherogenic actions, in part by stimulating proliferation in a variety of cell types of cells within the vessel wall, including endothelial cells, macrophages, and fibroblasts [2, 3]. However, the proliferative activity of ox-LDL in VSMCs appear contradictory. Experimental studies have shown that a mildly ox-LDL might evoke a proliferative response to contribute to the formation of intimal plague [4, 5]. Alternatively, high concentrations of ox-LDL cause LOX-1-mediated apoptosis of VSMCs to destabilize the atherosclerotic plaques [3, 6]. The mechanism whereby ox-LDL induces VSMCs proliferation and apoptosis might be dependent on the simultaneous induction of both cell cycle activators and suppressors [2]. In spite of the controversy, recent demonstration from our laboratory showed that ox-LDL evoked cell proliferation in a time- and concentration-dependent manner [7]. However, the mechanisms underlying the effect of ox-LDL on VSMCs proliferation are not fully understood.

Growth-differentiation factor-15 (GDF-15), a distant member of the transforming growth factor- β cytokine superfamily, is first found as macrophage inhibiting factor 1 (MIC-1) to inhibit tumor necrosis factor α (TNF- α) secreting from activated macrophages [8]. Accumulating evidence indicates that MIC-1/GDF-15 is induced in the myocardium following pathological stress associated with inflammation or tissue injury [9], and is closely related to the biomarkers of heart disease, such as B-type natriuretic peptide, N-terminal pro-BNP, cardiac troponin I and cardiac troponin T [10, 11]. Experimental studies have shown that, besides heart diseases, GDF-15 overexpression also occurs in other

cardiovascular cell types including macrophages, endothelial cells and vascular smooth muscle cells following pathological stresses [12, 13]. Consistent to correlation to common cardiac biomarkers, GDF-15 exhibits independent associations in those biomarkers of endothelial activation, both E- and P-selectin, VCAM-1, and ICAM-1. These adhesion molecules are involved in the recruitment of leukocytes into the vessel wall which is commonly regarded as a key step in the development and progression of atherosclerotic vascular disease [14]. These data suggest that GDF-15 may not only be a marker of cardiac abnormalities but also a marker of vascular pathologies, especially in atherogenesis.

In addition to overexpression of GDF-15 in activated endothelial cells, GDF-15 expression is also upregulated in macrophages activated by ox-LDL and is significantly correlated to MMP-9, probably reflecting the effects of macrophage activation on the progression of plaque development [15]. Given the prognostic value of GDF-15 in atherosclerotic plaques, a systematic analysis of the potential associations between GDF-15 and VSMCs proliferation is lacking so far. The aim of the current study was to illustrate the correlation of GDF-15 and VSMCs proliferation in the stimulation of ox-LDL and to explore the signaling pathways implicated in the expression of GDF-15.

Methods

Materials

Ox-LDL (YB-002) was purchased from the Yiyuan Biotechnologies Co. Ltd. Guangzhou, China. GDF-15 (1:1000, Abcam), ERK1/2 (1:1000, Abcam), p-ERK1/2 (1:1000, Abcam), GAPDH (1:2000, Santa Cruz) and detected using horseradish peroxidase-conjugated mouse IgG secondary antibodies (1:5000, Santa Cruz). ER-K1/2 (ab17942) and p38 (ab31828) inhibitors were purchased from Abcam. Other chemicals were dissolved in double-distilled water or experimental solutions.

Cell culture

VSMCs were utilized and isolated from Wistar rats. Aortic strips were cut into about 2 mm small rings and placed in 6-well culture plates. These explants were grown in DMEM (Hyclone, Utah, USA) following the instruction [5]. VSMCs were maintained in DMEM supplemented with 10% FCS (Invitrogen, Carlsbad, CA, USA) containing 100 mg/mL penicillin and 100 mg/mL streptomycin (Invitrogen), and cultured in a humidified atmosphere of 95% air/5% CO₂ at 37°C. The medium was changed after 24 hours and then every 3 days. These cells were identified as VSMCs by their characteristic hill-and-valley growth patterns and the presence of SMC-specific-actin.

ELISA assay

For quantitative measurement of GDF-15, the cell culture media was collected and diluted 20 times. Detailed procedures were conducted according to the protocol in the Mouse/Rat GDF-15 Quantikine ELISA Kit (MGD150, R&D system).

Plasmid construction and stable transfectants

The vector GV248 and helper plasmids pHelper were purchased from QIAGEN. The full-length CDS of rat GDF15 was cloned into the GV248 vector. GDF15-specific short hairpin RNAs were designed and cloned into GV248 (pFU-GW-007-hU6-Ubiguitin-EGFP-IRES-puromycin) vectors to inhibit GDF-15 expression. The viral particles were produced in 293T cells. A successful transfer of GDF-15 into VSMCs was achieved with a MOI (multiplicities of infection) of 100-200 viral particles/cell after 48 hour incubation in DMEM with 10% FBS. MOI 100 was the minimal concentration with the maximal transfer of the reporter gene into 100% of VSMCs. VSMCs were infected in DMEM with 10% FBS for 48 hour and cell numbers were determined by direct cell counting.

Western blot

The procedures were conducted according to our own publication [7]. Cultured VSMCs were lysed in lysis buffer (protease and phosphatase inhibitors), then lysates were centrifuged at 13, 500 g for 15 minutes at 4°C. Protein concentration was assessed using the BCA protein assay. The aliquots were then mixed with Laemmli sample buffer and boiled at 99°C for 5 min. The samples were resolved using 8-10% SDS/PAGE, followed by electrotransfer to polyvinylidene fluoride membranes. For visualization, blots were probed with antibodies against



Figure 1. ox-LDL promotes smooth muscle cell proliferation *in vitro*. The proliferation of rat aortic smooth muscle cells incubated with ox-LDL (50 μ g/ml and 100 μ g/ml) was evaluated by the increment of cell number (A). The viability was detected by CCK-8 assay (B). *P<0.05; n=3 different experiments. Values are means ± SEM.

GDF-15 (1:1000), ERK1/2 (1:1000), p-ERK1/2 (1:1000), GAPDH (1:2000) and detected using horseradish peroxidase-conjugated mouse IgG secondary antibodies (1:5000).

Statistics

All data are expressed as mean \pm standard error of the mean (SEM). One-way analysis of variance (ANOVA) was used to analyze the statistical differences among the groups. The *p*-values <0.05 were considered statistically significant. Statistical analyses and graphs were performed with SPSS 15.0 for Windows and GraphPad Prism 7.0 software.

Results

Ox-LDL induced VSMCs proliferation through MAPKs activation

Consistent with published data from our laboratory, ox-LDL promoted VSMCs proliferation in

a time- and concentration-dependent way. Compared with the control group, VSMCs proliferation was robust under the treatments of ox-LDL at 50 and 100 µg/ml for 24 hours. In addition, VSMCs maintained the proliferation with the time increasing to 48 hours (Figure 1A). However, VS-MCs proliferation was not a linear correlation with the concentrations of ox-LDL. The cell viability was tested using CCK-8 assay. After the exposure of ox-LDL for 24 hours, all cell suspensions were prepared in a 96-well plate to incubate with CCK-8 solution for 2 hours, and then the absorbance at 450 nm was measured using microplate reader (Figure 1B). ox-LDL enhanced VSMCs viability in a timeand concentration-dependent way.

MAPK activation, one of the major pathways involved in cell proliferation, was assessed through the protein expression level, including p38, ERK 1/2 and JNK (Figure 2). After 48 hour incubation with ox-LDL, VSMCs were collected

and isolated the total proteins. The protein productions of p38 and ERK 1/2 followed a similar trend, significantly increasing with the treatment of ox-LDL. In contrast, the expression level of JNK phosphorylation remained unchanged to ox-LDL treatment.

GDF-15 was upregulated in VSMCs after the treatment of ox-LDL through an ERK 1/2 and p38-dependent mechanism

The supernatants of VSMCs in each group were collected, and the GDF-15 concentration was measured by ELISA assay. After exposure of 50 μ g/ml ox-LDL for 24 hour, GDF-15 concentration in the supernatant of Control group was 0.375 \pm 0.023 ng/ml, and GDF-15 concentration in the supernatant of Control + ox-LDL group was 0.462 \pm 0.009 ng/ml; After 48 hours incubation without ox-LDL, GDF-15 concentration had no obvious change (0.349 \pm 0.048 ng/

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Figure 2. The effect of ox-LDL on the protein expression of MAPK families. Western blot analysis of protein expression (A) and relative quantity was calculated (B) in VSMCs exposed to 100 µg/ml ox-LDL for 48 h. *P<0.05; n=3 different experiments. Values are means ± SEM.





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ml), but ox-LDL dramatically increased the concentration level of GDF-15 to 0.637± 0.096 ng/ml. In this study, NC + ox-LDL group meant negative control of virus infection, a conclusion could be made that unpacked virus did not disturb the expression of GDF-15 stimulated by ox-LDL (Figure 3A). To further expose VSMCs to 100 µg/ml ox-LDL for 24 and 48 h, the concentration increased to 0.637±



Figure 4. GDF-15 protein expression in ox-LDL-treated VSMCs was dependent on p-38 and ERK1/2 MAPK activation. The inhibitors of p-38 and ERK1/2 inhibited the GDF-15 protein expression in ox-LDLtreated cells. P<0.05; n=3 different experiments. Values are means \pm SEM.

0.096 and 0.650±0.070 ng/ml in Control + ox-LDL and NC + ox-LDL groups respectively (**Figure 3B**).

The main objective of this study was to determine whether GDF-15 was involved in ox-LDLinduced proliferation. The GDF-15 expression in response to ox-LDL was stunted by ERK1/2 and p38 inhibition (**Figure 4**).

Gain- or loss-of-function of GDF-15 regulated the VSMCs proliferation in response to ox-LDL treatment.

Gain- or loss-of-function of GDF-15 was performed by the lentiviral infection of a full-length CDS of rat GDF-15 or GDF-15-specific short hairpin RNAs. Overexpression of GDF-15 significantly enhanced VSMCs proliferation. In contrast, knock-down GDF-15 followed a similar trend as utilization of ERK 1/2 and p38 inhibition significantly inhibited VSMCs proliferation with 100 μ g/ml ox-LDL treatment for 48 hours (**Figure 5**).

Discussion

TGF β has been intensively studied in atherosclerotic development [16], which is believed to be a critical factor in the formation of atherosclerotic plaque. GDF-15 is a member of the TGF-B cytokine superfamily and was first found as macrophage inhibiting factor 1, and it may inhibit tumor necrosis factor α (TNF- α) secreting from activated macrophages. TNF- α is important for recruiting macrophages into foam cells to form atherosclerotic plaque. Moreover, GDF-15 plays a pivotal role in ox-LDL-induced apoptosis of human macrophages in arteriosclerotic lesions [12, 17]. Thus, GDF-15 may orchestrate atherosclerotic lesion progression by regulating inflammatory responses to prevent atherosclerotic development [18, 19]. Overexpression of GDF-15 also protects the atherosclerotic process and reduces atherosclerotic lesion size [18]. Intriguing, it has also been found that deficiency of GDF-15 attenuates early atherosclerotic lesion formation and improves the stability of plagues by induction of collagen deposition [17, 20]. The diverse effects of GDF-15 may vary with the stage of the disease. GDF-15 mediates anti-atherosclerotic effects in mice by directly inhibiting myeloid cell recruitment, but indirectly promotes proinflammatory effects in atherosclerosis models [9, 17, 21-23].

In recent years, increasing investigation reveals that GDF-15 is an independent marker of cardiovascular dysfunction and disease [24-27]. Several lines of evidence reveal that GDF-15 is highly expressed in endothelial cells and VSMCs in normal and pathological conditions. In human endothelial cells, GDF-15 has independent associations with biomarkers of endothelial, such as E- and P-selectin, VCAM-1, and ICAM-1. These adhesion molecules are commonly regarded as an initial step in the formation of atherosclerosis by recruiting circulating leukocytes [14]. VSMCs proliferation is another crucial factor in developing atherosclerotic plaque [16, 28, 29]. ox-LDL has been thought to be mitogenic to VSMCs to induce VSMC proliferation through the activation of the ERK1/2 MAPK signaling pathway [5]. In addition, it has been reported that GDF-15 expression is upregulated by the treatment of ox-LDL, partially depending on the activation of ERK 1/2 [19]. However, a systematic analysis of the potential associations between GDF-15 levels and VS-MCs proliferation is lacking so far. Our data indicate that ox-LDL elicits expression of GDF-15 in rat VSMCs, and overexpression of GDF-15 by lentiviral infection has the same effects on VSMCs proliferation induced by ox-LDL. Con-



in vitro. NC, OE, and KD meant the negative control of virus infection, GDF-15 overexpression of virus infection and GDF-15 knockout of virus infection. *P<0.05; n=3 different experiments. Values are means ± SEM.

sistent with macrophages treated with ox-LDL, GDF-15 expression is upregulated in VSMCs incubated with ox-LDL *in vitro*, and is dependent on the activation of MAPK pathways. Unfortunately, correlation of increased expression of GDF-15 was not validated with proliferative actions *in vivo*. Future research is needed to study the potential actions of GDF-15 on the arterial wall thickness and plaque stability in an animal model of atherosclerosis.

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Conclusions

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The results of the present study demonstrate that GDF-15 is a proliferative molecule that gives rise to VSMCs proliferation secondary to ox-LDL stimulation *in vitro*. VSMCs treated with ox-LDL increase the induction of GDF-15, which is mediated through the activation of the p38 and ERK 1/2 MAPK pathways. A better understanding of the relationship between GDF-15 and VSMCs proliferation will provide us with further insight into the mechanism of plaque formation. It also offers new evidence that GDF-15 is a suitable biomarker for cardiovascular disease, and enables us to focus on this specific molecular as a potential therapeutic target for atherosclerosis.

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Disclosure of conflict of interest

None.

Abbreviations

GDF-15, Growth differentiation factor 15; TGF-β, Transforming Growth Factor-Beta; ox-LDL, Oxidized low-density lipoprotein; VSMC, Vascular smooth muscle cell; SMC, Smooth muscle cell; MAPK, Mitogen-activated protein kinase; ERK1/2, Extracellular signal-regulated kinase; LOX-1,

Lectin-type oxidized LDL receptor 1; FBS, Fetal bovine serum; MOI, Multiplicities of infection; TNF- α , Tumor necrosis factor alpha; VCAM-1, Vascular cell adhesion protein 1; ICAM-1, Intercellular adhesion molecule 1.

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References

- [1] Grootaert MOJ, Moulis M, Roth L, Martinet W, Vindis C, Bennett MR, De Meyer GRY. Vascular smooth muscle cell death, autophagy and senescence in atherosclerosis. Cardiovasc Res 2018; 114: 622-634.
- [2] Zettler ME, Prociuk MA, Austria JA, Massaeli H, Zhong G, Pierce GN. OxLDL stimulates cell proliferation through a general induction of cell cycle proteins. Am J Physiol Heart Circ Physiol 2003; 284: H644-53.
- [3] Kataoka H, Kume N, Miyamoto S, Minami M, Morimoto M, Hayashida K, Hashimoto N, Kita T. Oxidized LDL modulates Bax/Bcl-2 through the lectinlike Ox-LDL receptor-1 in vascular smooth muscle cells. Arterioscler Thromb Vasc Biol 2001; 21: 955-60.
- [4] Tsai S, Hollenbeck ST, Ryer EJ, Edlin R, Yamanouchi D, Kundi R, Wang C, Liu B, Kent KC. TGF-beta through Smad3 signaling stimulates vascular smooth muscle cell proliferation and neointimal formation. Am J Physiol Heart Circ Physiol 2009; 297: H540-9.

- [5] Yang CM, Chien CS, Hsiao LD, Pan SL, Wang CC, Chiu CT, Lin CC. Mitogenic effect of oxidized low-density lipoprotein on vascular smooth muscle cells mediated by activation of Ras/Raf/MEK/MAPK pathway. Br J Pharmacol 2001; 132: 1531-41.
- [6] Mehta JL, Sanada N, Hu CP, Chen J, Dandapat A, Sugawara F, Satoh H, Inoue K, Kawase Y, Jishage K, Suzuki H, Takeya M, Schnackenberg L, Beger R, Hermonat PL, Thomas M, Sawamura T. Deletion of LOX-1 reduces atherogenesis in LDLR knockout mice fed high cholesterol diet. Circ Res 2007; 100: 1634-42.
- [7] Lin J, Zhou S, Zhao T, Ju T, Zhang L. TRPM7 channel regulates ox-LDL-induced proliferation and migration of vascular smooth muscle cells via MEK-ERK pathways. FEBS Lett 2016; 590: 520-32.
- [8] Bootcov MR, Bauskin AR, Valenzuela SM, Moore AG, Bansal M, He XY, Zhang HP, Donnellan M, Mahler S, Pryor K, Walsh BJ, Nicholson RC, Fairlie WD, Por SB, Robbins JM, Breit SN. MIC-1, a novel macrophage inhibitory cytokine, is a divergent member of the TGF-beta superfamily. Proc Natl Acad Sci U S A 1997; 94: 11514-9.
- [9] Kempf T, Eden M, Strelau J, Naguib M, Willenbockel C, Tongers J, Heineke J, Kotlarz D, Xu J, Molkentin JD, Niessen HW, Drexler H, Wollert KC. The transforming growth factor-beta superfamily member growth-differentiation factor-15 protects the heart from ischemia/reperfusion injury. Circ Res 2006; 98: 351-60.
- [10] Wang TJ, Wollert KC, Larson MG, Coglianese E, McCabe EL, Cheng S, Ho JE, Fradley MG, Ghorbani A, Xanthakis V, Kempf T, Benjamin EJ, Levy D, Vasan RS, Januzzi JL. Prognostic utility of novel biomarkers of cardiovascular stress: the framingham heart study. Circulation 2012; 126: 1596-604.
- [11] Rohatgi A, Patel P, Das SR, Ayers CR, Khera A, Martinez-Rumayor A, Berry JD, McGuire DK, de Lemos JA. Association of growth differentiation factor-15 with coronary atherosclerosis and mortality in a young, multiethnic population: observations from the dallas heart study. Clin Chem 2012; 58: 172-82.
- [12] Schlittenhardt D, Schober A, Strelau J, Bonaterra GA, Schmiedt W, Unsicker K, Metz J, Kinscherf R. Involvement of growth differentiation factor-15/macrophage inhibitory cytokine-1 (GDF-15/MIC-1) in oxLDL-induced apoptosis of human macrophages in vitro and in arteriosclerotic lesions. Cell Tissue Res 2004; 318: 325-33.
- [13] Bermúdez B, López S, Pacheco YM, Villar J, Muriana FJ, Hoheisel JD, Bauer A, Abia R. Influence of postprandial triglyceride-rich lipoproteins on lipid-mediated gene expression in

smooth muscle cells of the human coronary artery. Cardiovasc Res 2008; 79: 294-303.

- [14] Galkina E1, Ley K. Vascular adhesion molecules in atherosclerosis. Arterioscler Thromb Vasc Biol 2007; 27: 2292-301.
- [15] Eggers KM, Kempf T, Lind L, Sundström J, Wallentin L, Wollert KC, Siegbahn A. Relations of growth-differentiation factor-15 to biomarkers reflecting vascular pathologies in a populationbased sample of elderly subjects. Scand J Clin Lab Invest 2012; 72: 45-51.
- [16] Bennett MR, Sinha S, Owens GK. Vascular smooth muscle cells in atherosclerosis. Circ Res 2016; 118: 692-702.
- [17] de Jager SC, Bermúdez B, Bot I, Koenen RR, Bot M, Kavelaars A, de Waard V, Heijnen CJ, Muriana FJ, Weber C, van Berkel TJ, Kuiper J, Lee SJ, Abia R, Biessen EA. Growth differentiation factor 15 deficiency protects against atherosclerosis by attenuating CCR2-mediated macrophage chemotaxis. J Exp Med 2011; 208: 217-25.
- [18] Bonaterra GA, Zügel S, Thogersen J, Walter SA, Haberkorn U, Strelau J, Kinscherf R. Growth differentiation factor-15 deficiency inhibits atherosclerosis progression by regulating interleukin-6-dependent inflammatory response to vascular injury. J Am Heart Assoc 2012; 1: e002550.
- [19] Xu X, Li Z, Gao W. Growth differentiation factor 15 in cardiovascular diseases: from bench to bedside. Biomarkers 2011; 16: 466-75.
- [20] Johnen H, Kuffner T, Brown DA, Wu BJ, Stocker R, Breit SN. Increased expression of the TGF-b superfamily cytokine MIC-1/GDF15 protects ApoE(-/-) mice from the development of atherosclerosis. Cardiovasc Pathol 2012; 21: 499-505.
- [21] Xu J, Kimball TR, Lorenz JN, Brown DA, Bauskin AR, Klevitsky R, Hewett TE, Breit SN, Molkentin JD. GDF15/MIC-1 functions as a protective and antihypertrophic factor released from the myocardium in association with SMAD protein activation. Circ Res 2006; 98: 342-50.
- [22] Rainer PP, Hao S, Vanhoutte D, Lee DI, Koitabashi N, Molkentin JD, Kass DA. Cardiomyocyte-specific transforming growth factor beta suppression blocks neutrophil infiltration, augments multiple cytoprotective cascades, and reduces early mortality after myocardial infarction. Circ Res 2014; 114: 1246-57.
- [23] Kempf T, Zarbock A, Widera C, Butz S, Stadtmann A, Rossaint J, Bolomini-Vittori M, Korf-Klingebiel M, Napp LC, Hansen B, Kanwischer A, Bavendiek U, Beutel G, Hapke M, Sauer MG, Laudanna C, Hogg N, Vestweber D, Wollert KC. GDF-15 is an inhibitor of leukocyte integrin activation required for survival after myocardial infarction in mice. Nat Med 2011; 17: 581-8.

- [24] Gohar A, Gonçalves I, Vrijenhoek J, Haitjema S, van Koeverden I, Nilsson J, de Borst GJ, de Vries JP, Pasterkamp G, den Ruijter HM, Björkbacka H, de Jager SCA. Circulating GDF-15 levels predict future secondary manifestations of cardiovascular disease explicitly in women but not men with atherosclerosis. Int J Cardiol 2017; 241: 430-436.
- [25] Adela R, Banerjee SK. GDF-15 as a target and biomarker for diabetes and cardiovascular diseases: a translational prospective. J Diabetes Res 2015; 2015: 490842.
- [26] Lind L, Wallentin L, Kempf T, Tapken H, Quint A, Lindahl B, Olofsson S, Venge P, Larsson A, Hulthe J, Elmgren A, Wollert KC. Growth-differentiation factor-15 is an independent marker of cardiovascular dysfunction and disease in the elderly: results from the prospective investigation of the vasculature in uppsala seniors (PI-VUS) study. Eur Heart J 2009; 30: 2346-53.

- [27] Zhu ZD, Sun T. Association between growth differentiation factor-15 and chronic heart failure in coronary atherosclerosis patients. Genet Mol Res 2015; 14: 2225-33.
- [28] Durham AL, Speer MY, Scatena M, Giachelli CM, Shanahan CM. Role of smooth muscle cells in vascular calcification: implications in atherosclerosis and arterial stiffness. Cardiovasc Res 2018; 114: 590-600.
- [29] Osonoi Y, Mita T, Azuma K, Nakajima K, Masuyama A, Goto H, Nishida Y, Miyatsuka T, Fujitani Y, Koike M, Mitsumata M, Watada H. Defective autophagy in vascular smooth muscle cells enhances cell death and atherosclerosis. Autophagy 2018; 14: 1991-2006.