Original Article Upregulation of long non-coding RNA ATP6V0E2-AS1 predicts a poor prognosis in prostate cancer

Shengguo Hu*, Youkong Li*, Min Zhu

Department of Urology, Jingzhou Central Hospital, The Second Clinical Medical College, Yangtze University, Jingzhou 434020, Hubei, P. R. China. *Equal contributors.

Received December 5, 2019; Accepted January 3, 2020; Epub May 15, 2020; Published May 30, 2020

Abstract: Background: Prostate cancer is a common cancer in males. Previous studies had demonstrated more than 90% of the human genome transcribed non-coding RNA. Long non-coding RNAs could regulate many biological processes in human diseases, including epigenetic regulation, RNA splicing, and protein degradation. The present study explored the prognostic value of a novel IncRNA ATP6V0E2-AS1 in prostate cancer (PCa). Method: The TCGA database was used to explore the expression of ATP6V0E2-AS1 in PCa and normal prostate tissues. The expression of ATP6V0E2-AS1 in PCa clinical samples was detected by real-time PCR. Finally, co-expression analysis, together with bioinformatics analysis, was used to research the functions of ATP6V0E2-AS1 in PCa. Results: The present study showed that ATP6V0E2-AS1 was significantly upregulated in PCa tissues. The qRT-PCR assay showed that ATP6V0E2-AS1 was overexpressed in PCa cell lines compared to WPMY-1. Our results also showed that ATP6V0E2-AS1 was upregulated in high pathology stage PCa. Upregulated ATP6V0E2-AS1 was positively correlated with a shorter survival time in PCa patients by analyzing the Kaplan-Meier dataset. Bioinformatics analysis showed that ATP6V0E2-AS1 was involved in protein translation regulation in PCa. Conclusion: We believe this research will provide new ideas for the diagnosis and treatment of prostate cancer.

Keywords: ATP6V0E2-AS1, prostate cancer, TCGA, bioinformatics analysis

Introduction

Non-coding RNAs (ncRNAs), which were regarded as transcription noise, have been validated as key regulators in human disease progression [1-3]. ncRNAs could be divided into several types according to their lengths. Of note, IncRNAs, a type of ncRNAs longer than 200 bps, has been implicated in cancers by regulating many biological processes including epigenetic regulation, RNA splicing, and protein degradation [4, 5].

In the United States, prostate cancer (PCa) was the most commonly diagnosed cancer in males [6]. LncRNA HOTAIR, LOC440040, and PVT1 have been found to be related to PCa. HOTAIR was upregulated in PCa samples, whose downregulation could inhibit the proliferation of PCa cells [7]. LncRNA XIST was down-regulated and acted as a tumor suppressor in PCa [8]. Several independent groups demonstrated that PCA3 could be detected in blood, urine, and PCa tissues with better sensitivity and specificity than prostate-specific antigen (PSA) [9-13].

We analyzed publically available databases to explore the expression of ATP6V0E2-AS1. Then, the prognostic value of IncRNA ATP6V0E2-AS1 in PCa was also analyzed. Next, a series of bioinformatic analysis was performed to reveal the roles of IncRNA ATP6V0E2-AS1 in PCa. This study can further advance the understanding of IncRNA ATP6V0E2-AS1 in PCa.

Materials and methods

Patients and clinicopathological data

497 PCa patients with clinical data and detailed expression of ATP6V0E2-AS1 were downloaded from The Cancer Genome Atlas (TCGA) data portal. The inclusion criteria were: (1) histologic diagnosis was prostate cancer; (2) data of samples were available on the expression of genes and characteristics of patients. The exclusion criteria were: (1) first histological diagnosis was not prostate cancer; (2) patients suffered from other malignant neoplasms except prostate cancer. GSE6919 [14, 15] was contributed by Federico AM, which contained 233 normal and 267 PCa tissues. All the patients were staged according to the 7th edition of the TNM staging standard of 2009.

Patients and tissue samples

The Research Ethics Committee of Jingzhou Central Hospital approved this study. 18 tumor and 6 normal tissue samples were collected from Jingzhou Central Hospital. Written informed consent was obtained from the patients. The patients did not receive any other treatment. LNCaP, WPMY-1, PC-3, 22RV1, and DU145 cells were purchased from Cell Bank of Chinese Academy of Sciences and cultured in 1640-media (10% FBS ,Gibco, USA).

Real-time PCR (qRT-PCR) analysis

Total RNAs of samples and cell lines were extracted using TransZol Up Plus RNA Kit (TransGen BioTech, China). Reverse transcript PCR was carried out using BeyoRT First Strand cDNA Synthesis Kit (Beyotime, China). qRT-PCR was performed using SuperReal PreMix Plus (TIANGEN, China) on Roche LightCycler 480. β -actin was selected as a reference. The 2^{- $\Delta\Delta$ Ct} method was performed to analyze the data.

Co-expression network construction

The Pearson correlation coefficient of ATP6V0-E2-AS1-co-expressed genes pairs was calculated. The absolute value of the Pearson correlation coefficient LightCycler 480. β -actin was selected as Cytoscape software was used to perform the co-expression network.

Gene Ontology (GO)and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analysis

MAS3.0 system (http://mas.capitalbiotech. com/mas3/) was used to present the GO analysis. DAVID system (https://david.ncifcrf.gov/ home.jsp) [16, 17] was used to present the KEGG pathway analysis. P < 0.05 was considered as statistically significant.

Statistical analysis

The data were shown as the mean \pm SD. The chi-squared test, t-test, Fisher's exact test, or

Mann-Whitney U-test was used to analyze the data. The survival times were analyzed by Kaplan-Meier curves (log-rank test) and were used to analyze the association between IncRNA ATP6V0E2-AS1 and the prognosis of PCa. P < 0.05 was selected as a reference.

Results

ATP6V0E2-AS1 is upregulated in PCa

As shown in **Figure 1A**, our analysis showed that ATP6V0E2-AS1 was overexpressed in tumor tissues (P < 0.001). Then, the GSE6919 dataset analysis showed that ATP6V0E2-AS1 was overexpressed in primary PCa and metastatic PCa samples (**Figure 1B**, P < 0.05).

Furthermore, we observed that ATP6V0E2-AS1 was overexpressed in PCa cell lines LNCaP (P < 0.05), PC-3 (P < 0.001), DU145 (P < 0.05), and 22Rv1 (P < 0.001) compared to noncancerous prostatic cells WPMY-1 by using RT-qPCR assay (**Figure 1C**). We evaluated the expression of ATP6V0E2-AS1 in PCa samples to validate our results. Comparing to normal tissues, the expression of ATP6V0E2-AS1 was higher in PCa samples (**Figure 1D**, P < 0.001).

Upregulation of ATP6V0E2-AS1 was positively correlated with poor prognosis

We also evaluated the expression levels of ATP6V0E2-AS1 with clinical varies, including Gleason score, T staging, and N staging. Our analysis showed that ATP6V0E2-AS1 was upregulated in Gleason 8 (Gleason 8 VS Gleason 6, P < 0.05; Gleason 8 VS Gleason 7, P < 0.01) and Gleason 9 (Gleason 9 VS Gleason 6, P < 0.01; Gleason 9 VS Gleason 7, P < 0.01) PCa patients compared to Gleason 6 and Gleason 7 PCa samples (Figure 2A). The results also showed that ATP6V0E2-AS1 levels were overexpressed in T3/T4 PCa compared to T2 PCa (Figure 2B, P < 0.01). Furthermore, we found that the ATP6V0E2-AS1 was overexpressed in N1 stage PCa compared to N0 PCa (Figure 2C, P < 0.001). These results suggested ATP6V0E2-AS1 was involved in PCa prognosis.

As shown in **Figure 3**, the Kaplan-Meier curve method showed that the 5-year BCR-free survival and overall survival rates were higher in ATP6V0E2-AS1-low patients compared to ATP-6V0E2-AS1-high patients (**Figure 3A**, P < 0.001;



Figure 1. ATP6V0E2-AS1 is upregulated in prostate cancer. A. ATP6V0E2-AS1 was upregulated in tumor tissues compared with normal tissues by analyzing the TCGA dataset. B. ATP6V0E2-AS1 was overexpressed in tumor tissues compared with normal tissues by analyzing the GSE6919 dataset. C. ATP6V0E2-AS1 was significantly overexpressed in PCa cell lines compared to WPMY-1 by using qRT-qPCR assay. D. ATP6V0E2-AS1 was overexpressed in 18 prostate cancer tissues compared with 6 normal tissues. The Mann-Whitney U-test was used to analyze the expression levels of ATP6V0E2-AS1 between normal prostate and PCa samples. *, P < 0.05; **, P < 0.01; ***, P < 0.001.

3B, P < 0.01), indicating that ATP6V0E2-AS1 could serve as a biomarker for PCa.

GO and KEGG pathway analysis

The co-expression networks were constructed by analyzing the Pearson correlation coefficient of ATP6V0E2-AS1-targets pairs. The absolute value of the Pearson correlation coefficient tion coefficient of ATP6V0E2-AS1-targets pairs. The absolute value of the PCa (PCa. A total of 1638 mRNAs were selected to construct a co-expression network.

The DAVID system was used to perform GO analysis based on the co-expressed genes of



Figure 2. ATP6V0E2-AS1 was upregulated in high Gleason and high TNM stage PCa samples. A. ATP6V0E2-AS1 was up-regulated in Gleason 8 (Gleason 8 VS Gleason 6, P < 0.05; Gleason 8 VS Gleason 7, P < 0.01) and Gleason 9 (Gleason 9 VS Gleason 6, P < 0.01; Gleason 9 VS Gleason 7, P < 0.01) PCa patients compared to Gleason 6 and Gleason 7 PCa samples. B. ATP6V0E2-AS1 was significantly overexpressed in T3/T4 PCa samples compared to T2 samples. C. ATP6V0E2-AS1 was overexpressed in N1 stage PCa samples compared to analyze the expression levels of ATP6V0E2-AS1 between normal prostate and PCa samples. *, P < 0.05; **, P < 0.01, ***, P < 0.001.



ATP6V0E2-AS1. The analysis revealed that ATP6V0E2-AS1 was significantly associated with translation, translational initiation, rRNA processing, cytoplasmic translation, mRNA splicing, via spliceosome, protein targeting to mitochondrion, mitochondrial translation, DNA repair, ribosomal large subunit biogenesis, aerobic respiration, proteasome-mediated ubiquitin, ribosomal small subunit biogenesis, spliceosomal snRNP assembly, cell-cell adhesion, and tRNA processing (Figure 4A). KEGG analysis showed that ATP6V0E2-AS1 was related to the regulation Spliceosome, Pyrimidine metabolism, Notch signaling pathway, RNA polymerase, Base excision repair, Protein processing in endoplasmic reticulum, VEGF signaling

pathway, DNA replication, and RNA transport (Figure 4B).

Construction of ATP6V0E2-AS1 mediated PPI network in PCa

Furthermore, the protein-protein interacting network (PPI) was constructed to predict the interactions among ATP6V0E2-AS1 co-expressed proteins in PCa (combined score > 0.4) using the STRING database [18, 19]. The Mcode plugin (degree cut-off \geq 2 and the nodes with edges \geq 2-core) was used to module analysis the PPI network. Three key modules were identified. Module 1 included 79 nodes and 2898 edges (**Figure 5A**), Module 2 included 74



Figure 3. The upregulation of ATP6V0E2-AS1 predicts a poor prognosis in prostate cancer. The rates of 5-year BCRfree survival (A) and overall survival (B) were higher in ATP6V0E2-AS1-low patients compared to ATP6V0E2-AS1-high patients. The log-rank test was used to compare the differences in survival times.



Figure 4. Bioinformatic analysis of ATP6V0E2-AS1 in prostate cancer. A. The Gene Ontologies (GO) categories analysis of ATP6V0E2-AS1 in prostate cancer. B. The KEGG analysis of ATP6V0E2-AS1 in prostate cancer.

nodes and 1226 edges (Figure 5B), and Module 3 included 24 nodes and 231 edges (Figure 5C).

We performed ClueGO analysis to explore the potential roles of these key modules by using Cytoscape software. We found that Module 1 was associated with translation, negative regulation of protein ubiquitination, and cleavage involved in rRNA processing (Figure 6A). Module 2 was associated with protein ubiguitination, mRNA splicing, snRNA metabolic process, DNA-templated transcription, and regulation of mRNA processing (Figure 6B). Module 3 was associated with ATP metabolic process, oxidative phosphorylation, hydrogen ion transmembrane transport, and mitochondrial electron transport (Figure 6C).

Discussion

Prostate cancer was a common cancer in males worldwide [20]. LncRNAs have been





paid much attention in recent years. Emerging studies have revealed that IncRNAs could regulate a variety of important biological processes. For example, Inc-BM could promote breast cancer brain metastasis by increasing JAK2 kinase activity [21]. FILNC1 interacted with AUF1 and sequestered AUF1 from binding c-Myc mRNA, which could repress c-Myc-mediated energy metabolism and inhibit renal tumor development [22]. PSA was a common diagnostic target for PCa, but its specificity is not very good [23, 24]. Previous studies have demonstrated the prognostic value of IncRNAs in PCa. For example, several independent groups showed that low PCAT-14 expression is associated with poor outcomes in PCa [25]. LncRNA SChLAP1 was reported to be overexpressed in PCa, and high SChLAP1 expression independently predicted biochemical recurrence after radical prostatectomy [26-28].

The present study showed that ATP6V0E2-AS1 was upregulated in PCa by analyzing public datasets and detecting PCa samples. Moreover, ATP6V0E2-AS1 upregulation was associated with the advanced stage of PCa. Kaplan-Meier curve analysis showed that higher ATP6V0E2-AS1 expression levels were associated with



worse survival in patients with PCa. This study revealed that IncRNA ATP6V0E2-AS1 could be a biomarker for PCa. Furthermore, we performed a co-expression analysis for IncRNA ATP6V0E2-AS1. Bioinformatic analysis revealed that ATP6V0E2-AS1 was significantly associated with translation, rRNA processing, mRNA splicing, protein targeting to mitochondrion, mitochondrial translation, DNA repair, cell-cell adhesion, and tRNA processing. Moreover, we constructed an ATP6V0E2-AS1-related PPI network in PCa. Three key modules were identified. Bioinformatics analysis module 1 was associated with translation, and module 2 was associated with protein ubiquitination, mRNA splicing, while module 3 was associated with ATP metabolic process.

Several limitations should be noted in the present study. First, the present study showed that ATP6V0E2-AS1 played an important role in PCa, mainly based on bioinformatic analysis. Therefore, further validation of our analysis using large-scale clinical samples was still needed. Second, the molecular functions of ATP6V0E2-AS1 in PCa still need more exploring. The loss-of-function assays should be applied to explore the molecular functions of ATP6V0E2-AS1 in PCa. Finally, the mechanisms of ATP6V0E2-AS1 in PCa remained to be further investigated.

In conclusion, we for the first time revealed that the potential functional roles of ATP6V0E2-AS1 were involved in PCa. ATP6V0E2-AS1 was upregulated and involved in protein translation regulation in PCa. We believe this research will advance the diagnosis and treatment of PCa.

Acknowledgements

This study was supported by Hubei Province Key Laboratory Open Fund Project (No. SB20-1403), Hubei Provincial Health Department Youth Talent Project (No. QJX2012-47).

Disclosure of conflict of interest

None.

Address correspondence to: Youkong Li, Department of Urology, Jingzhou Central Hospital, The Second Clinical Medical College, Yangtze University, Jingzhou 434020, Hubei, P. R. China. Tel: +86-0716-8497031; Fax: +86-0716-8497031; E-mail: liyoukong@126.com

References

- Idda ML, Munk R, Abdelmohsen K and Gorospe M. Noncoding RNAs in Alzheimer's disease. Wiley Interdiscip Rev RNA 2018; 9.
- [2] Yang J, Li C, Mudd A and Gu X. LncRNA PVT1 predicts prognosis and regulates tumor growth in prostate cancer. Biosci Biotechnol Biochem 2017; 81: 2301-2306.
- [3] Jin Y, Lu J, Wen J, Shen Y and Wen X. Regulation of growth of human bladder cancer by miR-192. Tumour Biol 2015; 36: 3791-3797.
- [4] Prensner JR, Chen W, Iyer MK, Cao Q, Ma T, Han S, Sahu A, Malik R, Wilder-Romans K, Navone N, Logothetis CJ, Araujo JC, Pisters LL, Tewari AK, Canman CE, Knudsen KE, Kitabayashi N, Rubin MA, Demichelis F, Lawrence TS, Chinnaiyan AM and Feng FY. PCAT-1, a long noncoding RNA, regulates BRCA2 and controls homologous recombination in cancer. Cancer Res 2014; 74: 1651-1660.
- [5] Xiao H, Tang K, Liu P, Chen K, Hu J, Zeng J, Xiao W, Yu G, Yao W, Zhou H, Li H, Pan Y, Li A, Ye Z, Wang J, Xu H and Huang Q. LncRNA MALAT1 functions as a competing endogenous RNA to regulate ZEB2 expression by sponging miR-200s in clear cell kidney carcinoma. Oncotarget 2015; 6: 38005-38015.

- [6] Siegel RL, Miller KD and Jemal A. Cancer statistics, 2017. Ca Cancer J Clin 2017; 67: 7-30.
- [7] Zhang A, Zhao JC, Kim J, Fong KW, Yang YA, Chakravarti D, Mo YY and Yu J. LncRNA HOTAIR enhances the androgen-receptor-mediated transcriptional program and drives castrationresistant prostate cancer. Cell Rep 2015; 13: 209-221.
- [8] Du Y, Weng XD, Wang L, Liu XH, Zhu HC, Guo J, Ning JZ and Xiao CC. LncRNA XIST acts as a tumor suppressor in prostate cancer through sponging miR-23a to modulate RKIP expression. Oncotarget 2017; 8: 94358-94370.
- [9] Hessels D, Klein GJ, van Oort I, Karthaus HF, van Leenders GJ, van Balken B, Kiemeney LA, Witjes JA and Schalken JA. DD3(PCA3)-based molecular urine analysis for the diagnosis of prostate cancer. Eur Urol 2003; 44: 8-15.
- [10] Deras IL, Aubin SM, Blase A, Day JR, Koo S, Partin AW, Ellis WJ, Marks LS, Fradet Y, Rittenhouse H and Groskopf J. PCA3: a molecular urine assay for predicting prostate biopsy outcome. J Urol 2008; 179: 1587-1592.
- [11] de Kok JB, Verhaegh GW, Roelofs RW, Hessels D, Kiemeney LA, Aalders TW, Swinkels DW and Schalken JA. DD3(PCA3), a very sensitive and specific marker to detect prostate tumors. Cancer Res 2002; 62: 2695-2698.
- [12] de Kok JB, Verhaegh GW, Roelofs RW, Hessels D, Kiemeney LA, Aalders TW, Swinkels DW and Schalken JA. DD3(PCA3), a very sensitive and specific marker to detect prostate tumors. Cancer Res 2002; 62: 2695-2698.
- [13] Dijkstra S, Mulders PF and Schalken JA. Clinical use of novel urine and blood based prostate cancer biomarkers: a review. Clin Biochem 2014; 47: 889-896.
- [14] Huang DW, Sherman BT and Lempicki RA. Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. Nat Protoc 2009; 4: 44-57.
- [15] Huang DW, Sherman BT and Lempicki RA. Bioinformatics enrichment tools: paths toward the comprehensive functional analysis of large gene lists. Nucleic Acids Res 2009; 37: 1-13.
- [16] Chandran UR, Ma C, Dhir R, Bisceglia M, Lyons-Weiler M, Liang W, Michalopoulos G, Becich M and Monzon FA. Gene expression profiles of prostate cancer reveal involvement of multiple molecular pathways in the metastatic process. BMC Cancer 2007; 7: 64.
- [17] Chu K, Gao G, Yang X, Ren S, Li Y, Wu H, Huang Y and Zhou C. MiR-512-5p induces apoptosis and inhibits glycolysis by targeting p21 in nonsmall cell lung cancer cells. Int J Oncol 2016; 48: 577-586.
- [18] Franceschini A, Lin J, von Mering C and Jensen LJ. SVD-phy: improved prediction of protein functional associations through singular value

decomposition of phylogenetic profiles. Bioinformatics 2016; 32: 1085-1087.

- [19] Szklarczyk D, Morris JH, Cook H, Kuhn M, Wyder S, Simonovic M, Santos A, Doncheva NT, Roth A, Bork P, Jensen LJ and von Mering C. The STRING database in 2017: quality-controlled protein-protein association networks, made broadly accessible. Nucleic Acids Res 2017; 45: D362-D368.
- [20] Siegel RL, Miller KD and Jemal A. Cancer statistics, 2015. CA Cancer J Clin 2015; 65: 5-29.
- [21] Wang S, Liang K, Hu Q, Li P, Song J, Yang Y, Yao J, Mangala LS, Li C, Yang W, Park PK, Hawke DH, Zhou J, Zhou Y, Xia W, Hung MC, Marks JR, Gallick GE, Lopez-Berestein G, Flores ER, Sood AK, Huang S, Yu D, Yang L and Lin C. JAK2-binding long noncoding RNA promotes breast cancer brain metastasis. J Clin Invest 2017; 127: 4498-4515.
- [22] Xiao ZD, Han L, Lee H, Zhuang L, Zhang Y, Baddour J, Nagrath D, Wood CG, Gu J, Wu X, Liang H and Gan B. Energy stress-induced IncRNA FILNC1 represses c-Myc-mediated energy metabolism and inhibits renal tumor development. Nat Commun 2017; 8: 783.
- [23] Prensner JR, Rubin MA, Wei JT and Chinnaiyan AM. Beyond PSA: the next generation of prostate cancer biomarkers. Sci Transl Med 2012; 4: 123r-127r.
- [24] Jalloh M and Cooperberg MR. Implementation of PSA-based active surveillance in prostate cancer. Biomark Med 2014; 8: 747-753.

- [25] Prensner JR, Iyer MK, Balbin OA, Dhanasekaran SM, Cao Q, Brenner JC, Laxman B, Asangani IA, Grasso CS, Kominsky HD, Cao X, Jing X, Wang X, Siddiqui J, Wei JT, Robinson D, Iyer HK, Palanisamy N, Maher CA and Chinnaiyan AM. Transcriptome sequencing across a prostate cancer cohort identifies PCAT-1, an unannotated lincRNA implicated in disease progression. Nat Biotechnol 2011; 29: 742-749.
- [26] Mehra R, Shi Y, Udager AM, Prensner JR, Sahu A, Iyer MK, Siddiqui J, Cao X, Wei J, Jiang H, Feng FY and Chinnaiyan AM. A novel RNA in situ hybridization assay for the long noncoding RNA SChLAP1 predicts poor clinical outcome after radical prostatectomy in clinically localized prostate cancer. Neoplasia 2014; 16: 1121-1127.
- [27] Mehra R, Udager AM, Ahearn TU, Cao X, Feng FY, Loda M, Petimar JS, Kantoff P, Mucci LA and Chinnaiyan AM. Overexpression of the long non-coding RNA SChLAP1 independently predicts lethal prostate cancer. Eur Urol 2016; 70: 549-552.
- [28] Prensner JR, Zhao S, Erho N, Schipper M, Iyer MK, Dhanasekaran SM, Magi-Galluzzi C, Mehra R, Sahu A, Siddiqui J, Davicioni E, Den RB, Dicker AP, Karnes RJ, Wei JT, Klein EA, Jenkins RB, Chinnaiyan AM and Feng FY. RNA biomarkers associated with metastatic progression in prostate cancer: a multi-institutional highthroughput analysis of SChLAP1. Lancet Oncol 2014; 15: 1469-1480.