Review Article Stem cell and endometriosis: new knowledge may be producing novel therapies

Jing Yang, Fengying Huang

Department of Obstetrics and Gynecology, The Second Xiangya Hospital, Central South University, Changsha 410011, Hunan, China

Received August 31, 2014; Accepted October 23, 2014; Epub November 15, 2014; Published November 30, 2014

Abstract: The human endometrium is a dynamic tissue, which undergoes cycles of growth and regression with each menstrual cycle. Adult progenitor stem cells are likely responsible for this remarkable regenerative capacity; these same progenitor stem cells may also have an enhanced capacity to generate endometriosis if shed in a retrograde fashion. The progenitor stem cells reside in the uterus, and, however, may also travel from other tissues such as bone marrow to repopulate the progenitor population. Mesenchymal stem cells are also involved in the pathogenesis of endometriosis and may be the principle source of endometriosis outside of the peritoneal cavity when they differentiate into endometriosis in ectopic locations. The present short review mainly summarizes the latest observations contributing to the current knowledge regarding the presence and the potential contribution of stem cells in the etiology of endometriosis. All these data can have clinical implications and provide a basis for new potential therapeutic applications.

Keywords: Endometriosis, endometrial progenitor/stem cells, etiology, therapeutic applications, telomerase

Introduction

Endometriosis is a common, benign, chronic estrogen-dependent disease characterized by the presence of ectopic endometrial implant [1]. This disease affects approximately 10% of reproductive-aged women and 20% to 50% of infertile women [2, 3]. Despite its frequency and its impact on quality of life, our understanding of the pathogenesis of endometriosis remains incomplete. Endometrial lesions are primarily located on the pelvic peritoneum and ovaries but can also be found in the pericardium, pleura, lung parenchyma, and even the brain [4]. Implants can result in substantial morbidity, including pelvic adhesions, pain, fatigue, bowel disorders, and infertility requiring extensive and sometimes ineffective medical and surgical treatments [5]. This disease is not only costly but also physically and psychologically debilitating. To date, the leading theories are retrograde menstruation, coelomic metaplasia, embryonic cell rest, and lymphatic and vascular dissemination Endometriosis and infertility: a review of the pathogenesis and treatment of endometriosis-associated [6]. Current research involving stem cells should shed some light on the puzzling mechanisms of this disease [7-9]. However, data on stem or progenitor cell function in endometriosis are scarce due to the technical limitations to stem cell research.

Stem cells within the uterus

The human uterine endometrium is one of the most dynamic human tissues. It consists of a glandular epithelium and stroma that are completely renewed in each monthly menstrual cycle. Endometrial stem cells are thought to reside in the basalis layer and serve as a source of cells that differentiate to form the endometrium Cyclic endometrial renewal depends on a small pool of tissue-specific multipotential stem cells [10, 11]. Under systemic hormonal changes, stem cells migrate and give rise to a group of progenitor cells that become committed to specific types of differentiated cells, e.g. epithelial, stromal and vascular cells. These endogenous stem cells allow the rapid regeneration of the endometrium necessary to support pregnancy [12, 13]. To date, the mesenchymal origin for endometrial stem cells is the most accepted statement [14, 15]. Noteworthy, putative endometrial SSCs have been successfully differentiated into other mesodermal lineages including adipose, chondrogenic, and osteogenic tissues [16-18].

On the other hand, there is an increasing trend suggesting that endometrial regeneration arises from bone marrow derived cells. Different groups have shown endometrial chimerism in endometrial glands and stroma of women who were given male or HLA-mismatched bone marrow transplants [19-21]. Another recent study from Du and coworkers [22] has shown that, after uterine ischemia or reperfusion injury, the amount of bone marrow-derived cells recruited to the endometrium went through a 2-fold increase. In this sense, whether bone marrow cells are recruited for endometrial regeneration under physiological conditions or only after injury or special needs remains to be clarified. Moreover, if bone marrow derived cells could be transformed and give rise to endometriosis or in which stage of their differentiation should they be transformed is still an unresolved question.

Stem cells in endometriosis

The presence of stem progenitor cells in endometrium and in menstrual blood led to the hypothesis that these cells could be at least in part responsible for the development of endometriosis. Lots of studies in the last decade have contributed to the consolidation of this hypothesis, through different approaches. In one study, it was suggested that the basal layer of the endometrium was significantly shed in menstrual flow in women with endometriosis, in comparison with control women [23]. Interestingly, endometrial stem cells are particularly frequent in endometrial tissue during menstruation. It has been speculated that endometrial stem cells may play an important role in the development of endometrial implants [24].

Epithelial cells in some endometriosis lesions are monoclonal, suggesting a single cell origin, possibly by an endometrial stem/progenitor cell. Other endometrial lesions are polyclonal, suggesting contamination with polyclonal stromal cells, repeated seeding of the lesion with cells from other sources, such as bone marrow, Cindy M.P. Duke, et al [25] has provided novel evidence of endometrial regeneration in bone marrow transplant recipients who received marrow from a single-HLA (single-human leukocyte antigen) antigen mismatched bone marrow transplant for leukemia. Donor-derived endometrial epithelial cells and stromal cells were detected in endometrial samples of bone Marrow recipients by RT-PCR (real time-PCR) and immunohistochemistry. Cyclic mobilization of bone marrow derived stem cells may be a normal physiologic process [26]. These findings define a new theory for the etiology of endometriosis ectopic trans-differentiation of stem cells. We have shown that bone marrow derived stem cells can contribute to endometriosis, perhaps explaining the occurrence of disease outside of the peritoneal cavity. Levendecker et al. showed that significantly more basalis layer was shed in the menstrual flow suggesting an increased number of stem cells in this layer that can result in a propensity for endometriosis [27].

Laschke et al [26] devoted further to the knowledge of the role of stem/progenitor cells in endometriosis with an elegant study based on a model of intraperitoneal endometriosis surgically induced in irradiated FVB/N mice that were reconstituted with bone marrow from FVB/N-TgN (Tie2/GFP) 287 Sato mice These transgenic mice express the reporter gene GFP under the transcriptional control of the Tie2 promoter. Tie2 is a vascular-endothelial-cellspecific receptor tyrosine kinase controlling numerous signaling pathways that are involved in endothelial cell migration and proliferation. The analysis conducted by intravital fluorescence microscopy and immunohistochemistry over 4 weeks after surgery clearly revealed the presence of GFP-positive cells in the vasculature of the ectopic endometrium of mice, thus indicating the contribution of bone-marrowderived EPCs (endothelial progenitor cells), originating CD31-positive endothelial cells. These cells were accompanied by local tissueresident EPCs, co-expressing CD34 and VE-GFR2 (vascular endothelial growth factor receptor2) [26]. As expected, Laschke et al [28] also showed that the homing of bone marrowderived GFP-positive EPCs in ectopic endometrium in mice was mediated by the SDF-la (stromal-cell-derived factor la)/CXCR4 (CXC chemokine receptor 4) axis, with an increased expression of SDF-la in developing endometrial lesions. These observations are in accordance with others revealing an up-regulation of circulating EPCs in a mouse model of endometriosis, where they are incorporated in the growing vasculature of the endometrial lesion and whose concentration positively correlated with the amount of endometrial tissue [29].

Yong Song et al [30] focus on stemness-related genes, such as the transcription pluripotency factors SOX2 (sex-determining region Y-box 2), NANOG (Nanog homeobox), and OCT4 (octamer-binding protein 4) in the endometrium of reproductive-age women with and without ovarian endometriosis, which play a crucial role in the regulation of self-renewal and pluripotency in embryonic stem cells and primordial germ cells, in comparison with the control group, SOX2 mRNA and protein expression was significantly higher in the eutopic endometrium of participants in the endometriosis group. In the endometriosis group, SOX2 and NANOG mRNA and protein expression were markedly increased in ectopic endometrium compared with eutopic endometrium; there was a trend towards lower OCT4 mRNA expression and higher OCT4 protein expression in ectopic endometrium, thus supporting a role for stem cell origin of endometriosis.

It is well known that cigarette smoking leads to a reduction in female infertility, but also to a decreased incidence of endometriosis and of endometrial cancer, through the protective effect played by the increased expression of homeobox A 10 and of progesterone receptor [31, 32]. A recent study provided novel insights into this previous observation, demonstrating in a murine model that both leucocytes and bone-marrow-derived endometrial cells were reduced by 60% and 73% respectively, after 4 weeks exposure to cigarette smoke [28]. Smoking inhibited the recruitment of bone-marrow-derived stem cells to uterus, as well as stem cell differentiation, suggesting that the inhibition of stem cell recruitment might be a general mechanism by which smoking leads to long-term organ damage though the inability to repair or regenerate multiple tissues [33]. These data indirectly support a role for stem cells in endometriosis progression.

Stem cells attempt to compensate for telomere attrition through the action of telomerase. Since circulating stem cells may contribute to endometriosis, longer telomeres could be consistent with a stem cell origin of endometriosis.

Endometriosis has been reported to be characterized by the presence of endometrial cells with capacity to avoid apoptosis beyond the uterine cavity [34]. Apoptosis plays an important role in maintaining tissue homeostasis by striking a balance between proliferation and cell death [34]. Endometrial cells have been described to show the peculiar biological characteristics of resistance to apoptosis with the inability to transmit apoptotic signal and the ability to avoid cell death [34, 35]. Recently, endometriosis has been correlated with aberrant endometrial expression of telomerase and survivin implying modifications in cell fate [34, 35]. Telomerase is a specialized transcriptase that can prevent telomere shortening [35]. Telomeres are noncoding tandemly repeated DNA sequences that are vital for maintaining chromosomal integrity and cell stability [35]. The critical shortening of telomeres is linked to cell division and their senescence and death [35]. Consequently, telomerase activation allows cells to overcome apoptosis acquiring immortal capacity.

Dracxler RC et al [36] compared telomere content in lymphocytes of patients with (n = 86) and without endometriosis (n = 21). Patients with endometriosis had longer telomeres than that of matched, endometriosis-free controls (telomere to single copy gene ratio [T/S ratio] of 1.62 vs 1.34, respectively, P = 00002). Patients with endometriosis were 8.1-fold more likely to have long telomeres (Odds ratio = 8.1, 95% confidence interval: 1.28-51.57, P = 0264).

Novel therapies targeting stem cell

The study summarized so far demonstrating an increase in circulating progenitor cells after the induction of endometrial lesion; it will be of great interest to test whether the circulating progenitor cells and the elevated expression of telomerase in patients could serve as diagnostic makers or as indicators of therapeutic efficacy.

Owing to a lack of full understanding of the pathogenesis of endometriosis, the treatment options success rates of endometriosis are limited, a better comprehension of the pathogenesis of endometriosis could help in developing

new, more effective, non-hormonal medication, which can inhibit disease development and alleviate pain or infertility without inhibition of ovulation and menstruation [32]. The identification of the different stem cell populations in ectopic endometrium could provide a potential therapeutic target for this frequent and invaliding disease. For example, a recent study conducted on MSCs (mesenchymal cells) isolated from eutopic human endometrium and from ectopic endometrium revealed that ectopic endometrial MSCs from patients with endometriosis showed a higher proliferation, migration and angiogenic ability in comparison with eutopic MSCs from the same patient or with control MSCs from patients without endometriosis. Treatment with sorafenib, a tyrosine kinase inhibitor, was able to revert the increased proliferative, migratory and angiogenic phenotype of ectopic endometrial MSCs [37]. Similarly, Pittatore G et al. [38] also revealed that once in the abdominal cavity, these MSCs could proliferate, invade, and differentiate into endometrial cells, finally generating ectopic implants. In more detail, As only differentiated endometrial cells, and not endometrial MSCs, possess steroid hormone receptors, MSCs could be responsible for the high rate of persistence/recurrence of the disease after hypoestrogenism-inducing therapies. MSCs play an important role in the survival and proliferation of endometrial tissue through the formation of new blood vessels. In fact, inhibition of angiogenesis represents a new, promising therapeutic approach for the disease. Further, medications directly targeting endometriosis MSCs could be effective, alone or in association with hormonal treatments, in increasing the success of medical treatment.

Current research centers on elucidating the factors that enable these cells to engraft an ectopic site and drive them toward endometrial differentiation. Preliminary data are providing several pathways to target in order to prevent stem cell flux and the initiation and progression of endometriosis. For example, Laschke et al. [28] in a marine model of surgically induced intraperitoneal endometriosis, where the treatment with AMD3100, an antagonist of the SDF-la/CXCR4 axis, significantly decreased the number of recruited EPCs and the vascular ration of endometrial lesions.

On the basis of previous studies, tobacco use inhibits stem cell flux to endometrium and

endometriosis [33]. While we would not advocate tobacco use in the treatment of endometriosis, it may explain the lower incidence of this disease in smokers. Individual components of tobacco smoke may prove useful in the treatment of this disease. Of therapeutic interest, the molecular targeting of deregulated signaling pathways by these cells and their local microenvironment, blocking key self-renewal pathways and pro-survival-signaling pathways, preventing cell recruitment, flux and adhesion through interference with chemokines, and adhesion molecules that regulate these processes, as well as inhibiting abnormally activated pathways within the cells or surrounding niche cells, represent new potential strategies for the development of more effective clinical treatments.

Similarly, Sakr et al. [39] demonstrate that the selective estrogen receptor modulator bazedoxifene (BZA), administered with conjugated estrogens (CEs), leads to regression of endometriosis lesions as well as reduction in stem cell recruitment to the lesions. Female mice underwent transplantation of male bone marrow. Endometrium was transplanted in the peritoneal cavity of half to create experimental endometriosis. Mice with or without experimental endometriosis were randomized to BZA/CE or vehicle treatment. Endometriosis lesions, bone marrow-derived mesenchymal stem cell engraftment of the lesions, and eutopic endometrium as well as ovarian stimulation were assessed. BZA treatment significantly reduced lesion size, gland number, and expression of proliferation marker proliferating cell nuclear antigen. Ovarian weight was not affected. Stem cells were recruited to the endometriosis lesions, and this recruitment was dramatically reduced by BZA/CE treatment. Stem cell engraftment was reduced in the uterus of animals with endometriosis; however the number of stem cells engrafting the uterus was completely restored by treatment with BZA/CE. Competition between endometriosis and the eutopic endometrium for a limited supply of stem cells and depletion of normal stem cells flux to the uterus is a novel mechanism by which endometriosis interferes with endometrial function and fertility. BZA/CE not only treats lesions of endometriosis, it also dramatically reduces stem cell recruitment to the lesions and restores stem cell engraftment of the uterine endometrium.

Given the key role of telomerase in the regulation of stem cell in endometrial lesions, downregulating telomerase expression, may be candidate as a new therapeutic target for endometriosis [40].

In conclusion, all of these approaches represent good concrete examples of the novel potential therapeutic strategies possibly arising from the unequivocal identification of stem cell populations contributing to the development of endometrial lesions. Further, endometrial derived stem cells have been demonstrated to be useful in the treatment of several chronic and often debilitating diseases, including Parkinson's disease [41] and diabetes [42]. They demonstrate that the uterus is a dynamic organ permeable to fetal stem cells capable of trans-differentiation as well as a renewable source of multipotent stem cells. While we still have much to understand about stem cells, their potential applications in reproductive biology and medicine are countless.

Disclosure of conflict of interest

None.

Address correspondence to: Fengying Huang, Department of Obstetrics and Gynecology, The Second Xiangya Hospital, Central South University, 139 Renmin Road, Changsha 410011, Hunan, China. Tel: +8613874870394; E-mail: 805754296@ qq.com; hfy6697@163.com

References

- Gilmour JA, Huntington A, Wilson HV. The impact of endometriosis on work and social participation. Int J Nurs Pract 2008; 14: 443-448.
- [2] Taylor RN, Lebovic DI, Mueller MD. Angiogenic factors in endometriosis. Ann N Y Acad Sci 2002; 955: 89-100.
- [3] Krikun G, Lockwood CJ, Paidas MJ. Tissue factor and the endometrium: from physiology to pathology. Thromb Res 2009; 124: 393-396.
- [4] Sharpe-Timms KL, Young SL. Understanding endometriosis is the key to successful therapeutic management. Fertil Steril 2004; 81: 1201-1203.
- [5] Rogers PA, D'Hooghe TM, Fazleabas A, Gargett CE, Giudice LC, Montgomery GW, Rombauts L, Salamonsen LA, Zondervan KT. Priorities for endometriosis research: recommendations from an international consensus workshop. Reprod Sci 2009; 16: 335-346.
- [6] Macer ML, Taylor HS. Endometriosis and infertility: a review of the pathogenesis and treat-

ment of endometriosis-associated infertility. Obstet Gynecol Clin North Am 2012; 39: 535-549.

- [7] Oliveira FR, Dela CC, Del Puerto HL, Vilamil Q, Reis FM, Camargos AF. Stem cells: are they the answer to the puzzling etiology of endometriosis? Histol Histopathol 2012; 27: 23-29.
- [8] Schwab KE, Chan RWS, Gargett CE. Putative stem cell activity of human endometrial epithelial and stromal cells during the menstrual cycle. Fertil Steril 2005; 84: 1124-1130.
- [9] Sasson IE, Taylor HS. Stem cells and the pathogenesis of endometriosis. Ann N Y Acad Sci 2008; 1127: 106-115.
- [10] Maruyama T, Masuda H, Ono M, Kajitani T, Yoshimura Y. Human uterine stem/progenitor cells: their possible role in uterine physiology and pathology. Reproduction 2010; 140: 11-22.
- [11] Padykula HA. Regeneration in the Primate Uterus. Biology of the Uterus: Springer; 1989. pp. 279-288.
- [12] Chan RW, Schwab KE, Gargett CE. Clonogenicity of human endometrial epithelial and stromal cells. Biol Reprod 2004; 70: 1738-1750.
- [13] Dimitrov R, Timeva T, Kyurkchiev D, Stamenova M, Shterev A, Kostova P, Zlatkov V, Kehayov I, Kyurkchiev S. Characterization of clonogenic stromal cells isolated from human endometrium. Reproduction 2008; 135: 551-558.
- [14] Schwab K, Gargett C. Co-expression of two perivascular cell markers isolates mesenchymal stem-like cells from human endometrium. Hum Reprod 2007; 22: 2903-2911.
- [15] Schwab K, Hutchinson P, Gargett C. Identification of surface markers for prospective isolation of human endometrial stromal colonyforming cells. Hum Reprod 2008; 23: 934-943.
- [16] Cervelló I, Gil-Sanchis C, Mas A, Delgado-Rosas F, Martínez-Conejero JA, Galán A, Martínez-Romero A, Martínez S, Navarro I, Ferro J. Human endometrial side population cells exhibit genotypic, phenotypic and functional features of somatic stem cells. PLoS One 2010; 5: e10964.
- [17] Kato K. Stem cells in human normal endometrium and endometrial cancer cells: characterization of side population cells. Kaohsiung J Med Sci 2012; 28: 63-71.
- [18] Choi YH, Burdick MD, Strieter RM. Human circulating fibrocytes have the capacity to differentiate osteoblasts and chondrocytes. Int J Biochem Cell Biol 2010; 42: 662-671.
- [19] Cervello I, Gil-Sanchis C, Mas A, Faus A, Sanz J, Moscardo F, Higueras G, Sanz MA, Pellicer A, Simón C. Bone marrow-derived cells from male donors do not contribute to the endometrial side population of the recipient. PloS One 2012; 7: e30260.

- [20] Taylor HS. Endometrial cells derived from donor stem cells in bone marrow transplant recipients. JAMA 2004; 292: 81-85.
- [21] Ikoma T, Kyo S, Maida Y, Ozaki S, Takakura M, Nakao S, Inoue M. Bone marrow-derived cells from male donors can compose endometrial glands in female transplant recipients. Am J Obstet Gynecol. 2009; 201: 608.
- [22] Du H, Naqvi H, Taylor HS. Ischemia/reperfusion injury promotes and granulocyte-colony stimulating factor inhibits migration of bone marrow-derived stem cells to endometrium. Stem Cells Dev 2012; 21: 3324-3331.
- [23] Leyendecker G, Herbertz M, Kunz G, Mall G. Endometriosis results from the dislocation of basal endometrium. Hum Reprod 2002; 17: 2725-2736.
- [24] Sasson IE, Taylor HS. Stem cells and the pathogenesis of endometriosis. Ann N Y Acad Sci 2008; 1127: 106-115.
- [25] Duke CM, Taylor HS. Stem cells and the reproductive system: Historical perspective and future directions. Maturitas 2013; 76: 284-289.
- [26] Taylor H. Endometrial cell derived from donor stem cell in bone marrow transplant recipients. JAMA 2004; 292: 104-105.
- [27] Leyendecker G, Herbertz M, Kunz G, Mall G. Endometriosis results from the dislocation of basal endometrium. Hum Reprod 2002; 17: 2725-2736.
- [28] Laschke MW, Giebels C, Nickels RM, Scheuer C, Menger MD. Endothelial progenitor cells contribute to the vascularization of endometriotic lesions. Am J Pathol 2011; 178: 442-450.
- [29] Becker CM, Beaudry P, Funakoshi T, Benny O, Zaslavsky A, Zurakowski D, Folkman J, D'Amato RJ, Ryeom S. Circulating endothelial progenitor cells are up-regulated in a mouse model of endometriosis. Am J Pathol 2011; 178: 1782-1791.
- [30] Song Y, Xiao L, Fu J, Huang W, Wang Q, Zhang X, Yang S. Increased expression of the pluripotency markers sex-determining region Y-box 2 and Nanog homeobox in ovarian endometriosis. Reprod Biol Endocrinol 2014; 12: 42.
- [31] Zhou Y, Jorgensen EM, Gan Y, Taylor HS. Cigarette smoke increases progesterone receptor and homeobox A10 expression in human endometrium and endometrial cells: a potential role in the decreased prevalence of endometrial pathology in smokers. Biol Reprod 2011; 84: 1242-1247.
- [32] Forte A, Cipollaro M, Galderisi U. Genetic, epigenetic and stem cell alterations in endometriosis: new insights and potential therapeutic perspectives. Clin Sci 2014; 126: 123-138.

- [33] Zhou Y, Gan Y, Taylor HS. Cigarette smoke inhibits recruitment of bone-marrow-derived stem cells to the uterus. Reprod Toxicol 2011; 31: 123-127.
- [34] Watanabe A, Taniguchi F, Izawa M, Suou K, Uegaki T, Takai E, Terakawa N, Harada T. The role of survivin in the resistance of endometriotic stromal cells to drug-induced apoptosis. Hum Reprod 2009; 24: 3172-3179.
- [35] Hapangama D, Turner M, Drury J, Quenby S, Saretzki G, Martin-Ruiz C, Von Zglinicki T. Endometriosis is associated with aberrant endometrial expression of telomerase and increased telomere length. Hum Reprod 2008; 23: 1511-1519.
- [36] Dracxler RC, Oh C, Kalmbach K, Wang F, Liu L, Kallas E, Giret M, Seth-Smith M, Antunes D, Keefe D. Peripheral Blood Telomere Content Is Greater in Patients With Endometriosis Than in Controls. Reprod Sci 2014; [Epub ahead of print].
- [37] Moggio A, Pittatore G, Cassoni P, Marchino GL, Revelli A, Bussolati B. Sorafenib inhibits growth, migration, and angiogenic potential of ectopic endometrial mesenchymal stem cells derived from patients with endometriosis. Fertil Steril 2012; 98: 1521-1530. e1522.
- [38] Pittatore G, Moggio A, Benedetto C, Bussolati B, Revelli A. Endometrial adult/progenitor stem cells: pathogenetic theory and new antiangiogenic approach for endometriosis therapy. Reprod Sci 2014; 21: 296-304.
- [39] Sakr S, Naqvi H, Komm B, Taylor HS. Endometriosis Impairs Bone Marrow-Derived Stem Cell Recruitment to the Uterus Whereas Bazedoxifene Treatment Leads to Endometriosis Regression and Improved Uterine Stem Cell Engraftment. Endocrinology 2014; 155: 1489-1497.
- [40] Mormile R, Vittori G. MAPK signaling pathway and endometriosis: what is the link? Arch Gynecol Obstet 2013; 287: 837-838.
- [41] Wolff EF, Gao XB, Yao KV, Andrews ZB, Du H, Elsworth JD, Taylor HS. Endometrial stem cell transplantation restores dopamine production in a Parkinson's disease model. J Cell Mol Med 2011; 15: 747-755.
- [42] Santamaria X, Massasa EE, Feng Y, Wolff E, Taylor HS. Derivation of insulin producing cells from human endometrial stromal stem cells and use in the treatment of murine diabetes. Mol Ther 2011; 19: 2065-2071.