Original Article Polymeric immunoglobulin receptor expression is predictive of poor prognosis in glioma patients

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Abstract: Although there have been recent advances in surgery, radiotherapy, and chemotherapy, the survival of patient with glioma remains poor. Increased expression of polymeric immunoglobulin receptor (plgR) in tumor tissue has been detected in various cancer forms. However, the clinical relevance of plgR in glioma remains unclear. The aim of this study was to assess the prognostic value of plgR in patients with glioma after surgical resection. plgR expression was evaluated by immunohistochemistry in paraffin-embedded glioma tissues from 146 patients. The relation between plgR expression and clinicopathologic factors and long-term prognosis in these 146 patients was retrospectively examined. The prognostic significance of negative or positive plgR expression in glioma was assessed using Kaplan-Meier survival analysis and log-rank tests. Positive expression of plgR was statistically significantly associated with poor prognosis of patients with glioma. Our results indicated that plgR could be a novel predictor for poor prognosis of patients with glioma after surgical resection.

Keywords: Glioma, polymeric immunoglobulin receptor (plgR), prognosis

Introduction

Gliomas are the most common primary brain tumors in the central nervous system and are also the most lethal and least successfully treated solid tumors [1]. The World Health Organization (WHO) classification grading system for human gliomas is usually used to evaluate the prognosis of glioma patients. According to the WHO guidelines [2], gliomas are histologically classified into four grades: pilocytic astrocytoma (grade I), diffuse astrocytoma (grade II), anaplastic astrocytoma (grade III) and glioblastoma multiforme (GBM, grade IV). Among these, the relatively slower-growing WHO I-II lesions are referred to as low grade gliomas and the more rapidly growing WHO III-IV lesions are referred to as high-grade gliomas. Gliomas arise from the constituent glial cells of the brain, or their precursors, and diffusely invade surrounding brain, making curative surgical resection almost impossible [3]. Despite progress in tumor diagnosis and treatment, including surgery, radiotherapy and chemotherapy, the median survival time is only one

year and few patients survive for two years [4]. However, some glioma patients with similar grades have obvious discrepancies in survival. It is therefore necessary to identify some new certain tumor biomarkers that are more suitable for the prognostic assessment of gliomas than the grading system.

The polymeric immunoglobulin receptor (plgR) is a transporter of dimeric IgA (dIgA) and pentameric IgM (pIgM), which are the first-line antibodies in response to initial infection. Widely expressed in epithelial cells, plgR expression is also commonly increased by proinflammatory cytokines in response to viral or bacterial infections, thus linking innate and adaptive immunity [5-8]. Up-regulation of plgR was detected in colon cancer [9], breast cancer [10, 11], endometrial carcinoma [12, 13], bladder carcinoma [14], and hepatocellular carcinoma (HCC) [15, 16]. High levels of the cleaved extracellular domain of plgR, designated as the secretory component, were also detected in the sera of patients with lung cancer [17, 18], pancreatic cancer [19], and colon cancer with liver metas-

Table 1. Childepathological features of 140 patients with glothas					
Factures		WHO grade			
reatures	WHO I	WHO II	WHO III	WHO IV	
Patients (n)	25	18	53	50	
Mean age (year)	39.7	45.9	44.1	44.2	
Gender					
Male (n)	15	8	26	34	
Female (n)	10	10	27	16	
KPS score					
≥ 80 (n)	21	15	12	12	
< 80 (n)	4	3	41	38	
Surgery					
Gross total resection (n)	25	18	30	38	
Partial resection (n)	0	0	23	12	
Adjuvant treatment					
Radiotherapy (n)	0	0	32	15	
Chemotherapy (n)	0	0	0	6	
Radiotherapy and chemotherapy combination (n)	0	0	7	23	

Table 1. Clinicopathological features of 146 patients with gliomas

tasis [20]. However, the clinical relevance of plgR in gliomas remains uncertain.

In the present study, formalin-fixed and paraffin-embedded tissues and the clinicopathological parameters from 146 patients with glioma were collected and the expression level of the plgR protein was evaluated by immunohistochemistry. Furthermore, the associations of patient prognosis and clinicopathological parameters with the expression of plgR protein were investigated. To the best of our knowledge, this is the first study to detect plgR expression in gliomas and to show a correlation between its expression level and patient prognosis.

Materials and methods

Patients and tumor tissue samples

Paraffin-embedded glioma tissue samples were obtained from 146 patients undergoing surgical resection at the Department of Neurosurgery, Sir Run Run Shaw Hospital, Zhejiang University School of Medicine and Department of Neurosurgery, Taizhou Hospital, Wenzhou Medical University from January 1998 to December 2010. None of 146 patients had received chemotherapy or radiotherapy before resection. This study was approved by the Research Ethics Committee of Sir Run Run Shaw Hospital, Zhejiang University School of Medicine and Taizhou Hospital, Wenzhou

Medical University. Written informed consent was obtained from all of the patients. All the slides were reevaluated according to WHO classifications [2, 21] by two pathologists. A total of 83 males and 63 females (1.32:1) were enrolled in this study, and the median age was 45 years (range, 15-78). Fortythree of the 146 gliomas were classified as low-grade [25] pilocytic astrocytomas (WHO I) and 18 diffuse as-

trocytomas (WHO II), and 103 were classified as high-grade gliomas [53] anaplasia astrocytomas (WHO III), and 50 primary glioblastomas (WHO IV). All patients were assessed by the Karnofsky Performance Status (KPS) scale: (1) minor disability (80 to 100 points); (2) moderate disability (60 to 70 points); and (3) severe disability (10 to 50 points) [22]. The clinicopathological features and the treatment strategies of all the patients were indicated in **Table 1**.

Immunohistochemistry

Selected tumor specimen were fixed in 10% neutral-buffered formalin and embedded in paraffin. Five micromolar sections were cut, dewaxed, rehydrated, and subjected to antigen retrieval. After blocking endogenous peroxidase activity, the sections were incubated with the primary antibody against plgR (Epitomics, Burlingame, CA) (1:100) (overnight at 4 at 4°C). Immunohistochemistry was performed using the streptavidin-biotin peroxidase complex method (Lab Vision, Fremont, CA). The slides were examined and pictures were taken using an Olympus BX60 (Olympus, Japan). Sections known to stain positively were incubated in each batch and negative controls were also prepared by replacing the primary antibody with preimmune sera.

Expression analysis of plgR in tumor tissue was performed by comparing staining intensity and the percentage of immunoreactive cells.



Figure 1. Representative immunohistochemical staining of plgR in glioma tissues. plgR protein was mainly expressed in the membrane with brown yellow (original magnification, A: × 100; B: × 400).

Staining intensity was arbitrarily scored on a scale of four grades: 0 (no staining of cancer cells), 1 (weak staining), 2 (moderate staining), and 3 (strong staining), and the percentage of positive cells was scored as follows: 0 (0%), 1 (1% to 25%), 2 (26% to 50%), and 3 (> 50%). plgR staining positivity was determined using the following formula: overall score = positive percentage score × intensity score. A score of 0 was defined as "0", > 0 to ≤ 2 as "1", > 2 to ≤ 6 as "2", and > 6 to ≤ 9 as "3". In the end, tumor samples rated as level 0 or 1 were defined as negative for expression, whereas samples rated as level 2 or 3 were defined as positive.

Follow-up

Patient follow-up consisted of assessment of CT and MRI every 3 months for the first 5 years. The patients were followed up until death or until the date of last follow-up. Follow-up was finished on December 31, 2013. The median follow-up was 28.2 months (range, 3-58 months).

Statistical analysis

All statistical analyses were performed by SPSS 16.0 software (SPSS, Chicago, IL). Data are expressed as a mean \pm SEM. Clinicopathologic parameters were analyzed using the two-tailed chi-square test, and the two-tailed *t* test was used to evaluate association between plgR expression and clinicopathologic parameters. Overall survival (OS) curves for positive- and negative-plgR patients were estimated with the Kaplan-Meier method, and the survival func-

tions were compared by the log rank test. Univariate and multivariate analyses were based on the Cox proportional hazards regression model. Factors that significantly influenced overall survival were used in the Cox proportional regression model for multivariate analysis. All *P*-values were considered statistically significant when the associated probability was less than 0.05.

Results

plgR expression in glioma

We evaluated the expression of plgR in 146 paraffin-embedded glioma tissue samples using the method of immunohistochemical staining. We found that among these 146 samples, plgR was positive in 101/146 (69.2%) glioma tissue samples (**Figure 1**). These results suggested that plgR might play a key role in glioma. **Table 2** showed the distribution of plgR expression level in 146 glioma tissue samples and the relationship between plgR expression level and clinicopathologic characteristics, including age, gender, and histological grade.

Positive-pIgR is associated with poor survival in patients with glioma

The OS curves for glioma patients subdivided on the basis of plgR expression are shown in **Figure 2**. Positive-plgR expression was associated with poor prognosis in glioma patients (log-rank test, P < 0.001). Univariate analysis showed that plgR-positive patients had a significantly poorer prognosis than plgR-negative

Clinicopathological	Detiente (n)	plgR expression		Dualua
features	Patients (n)	Positive (n, %)	Negative (n, %)	P value
WHO grade				
I	25	17	8	< 0.001
II	18	13	5	
III	53	37	16	
IV	50	34	16	
Gender				
Male	83	59	24	NS
Female	63	42	21	
Age (year)				
< 55	50	33	17	NS
≥ 55	96	68	28	
KPS score				
≥80	60	41	19	NS
< 80	86	60	26	

Table 2. Clinicopathological characteristics in relation to plgR expression in patients with glioma (n = 146)

Note: NS, the difference with no statistical significance.



Figure 2. Kaplan-Meier survival curves of patients with glioma undergoing surgical resection, grouped by plgR expression in tumor tissues. The survival rate for patients with glioma in the plgR-negative expression group (n = 45) was significantly higher than that for patients in the plgR-positive expression group (n = 101, log-rank, P < 0.001).

 Table 3. Univariate analysis of overall survival of glioma patients after surgical resection

Factor		Overall survival	
Factor		Patients (n)	P value
plgR expression in glioma tissue samples	Positive	101	< 0.001
	Negative	45	
	Total	146	

patients (*P* < 0.001; **Table 3**). Multivariate analysis showed that positive-plgR expression was an independent and significant predictor in OS (**Table 4**).

Discussion

Glioma accounts for nearly one-third of all intrinsic neoplasms in the central nervous system including well-differentiated low-grade astrocytomas, anaplastic astrocytomas, and glioblastoma [23]. This tumor is aggressive and has a tendency to invade the surrounding brain tissue. Although recent advances in surgery, radiotherapy, photodynamic therapy, and chemotherapy, survival of patients with gliomas remains poor. The median overall survival of patients with malignant gliomas is no more than 1 year and local recurrence occurs in more than 90% of patients [4]. It is extremely important to find biomarkers that can offer prognostic insight and eventually guide clinical treatment.

As the most common and deadly brain tumors, human gliomas have prompted many studies which focus on the genetic variation and molecular expression patterns in order to characterize different tumor subgroups, to understand the malignant tumor behavior and to identify valuable, reliable molecular targets for future targeted therapies. In the current study, our data showed that the abnormal expressions of plgR protein appeared to be correlated with the WHO grade of glioma, clinicopathological features, as well as patient survivals. To the best of our knowledge,

Table 4. Multivariate analysis of	f overall survival of	glioma patients after
surgicalresection		

Factor	HR (95% CI)	P value
plgR expression in glioma tissue samples (Positive)	2.371 (1.523-3.373)	< 0.001
HR, hazards ratio. Cl, confidence interval.		

this is the first study to demonstrate the prognostic value of the expression of plgR in gliomas.

The aim of this study was to evaluate the prognostic value of plgR in patients with glioma after surgical resection. plgR is a glycoprotein presents on glandular epithelial cells that functions as a receptor for polymeric immunoglobulin. plgR transports polymeric immunoglobulin A (IgA) into external secretions as secretory IgA (S-IgA), which is critical for the defence of mucosal tissues [24]. As mentioned above, plgR was overexpressed in tumor tissue of colon cancer [9], breast cancer [10, 11], endometrial carcinoma [12, 13], bladder carcinoma [14], and HCC [15, 16], but its clinical relevance remains uncertain. The prognostic value of plgR in patients with malignancy was also not ascertained. Ai and colleagues, for the first time, reported the clinical relevance of plgR in HCC [16]. In their study, plgR was identified as a prognostic biomarker for HCC and a molecular player in hepatitis B infection, chronic liver inflammation, the induction of the epithelialmesenchymal transition, HCC recurrence, and metastatic progression [16]. Since the role of plgR in glioma has not been studied so far, we investigated a set of 146 tumor samples immunohistochemically and correlated our findings with clinico-pathological parameters to also identify potential prognostic implications of plgR in this brain tumor.

We evaluated the plgR expression in paraffinembedded glioma tissue samples from 146 patients with glioma, which had clinical followup records. The result of positive expression of plgR was confirmed in 101 (69.2%) paraffinembedded glioma tissue samples. Univariate analysis indicated significantly worse OS for patients with a positive plgR expression in glioma tissues than for patients with a negative plgR expression. Multivariate analysis showed positive-plgR in glioma tissues to be an independent prognostic factor for OS after surgical resection (P < 0.001). These data, for the first time, imply that plgR has distinct roles in glioma and is worthy of further investigation.

In summary, this is the first study showing positive expression of plgR was sta-

tistically significantly associated with poor prognosis of patients with glioma. Our results indicated plgR can be a novel predictor for poor prognosis of patients with glioma after surgical resection and plgR might be a promising candidate for targeted therapy of glioma.

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Disclosure of conflict of interest

None.

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References

- Schneider T, Mawrin C, Scherlach C, Skalej M, Firsching R. Gliomas in adults. Dtsch Arztebl Int 2010; 107: 799-807.
- [2] Louis DN, Ohgaki H, Wiestler OD, Cavenee WK, Burger PC, Jouvet A, Scheithauer BW, Kleihues P. The 2007 WHO classification of tumours of the central nervous system. Acta Neuropathol 2007; 114: 97-109.
- [3] Ohgaki H. Epidemiology of brain tumors. Methods Mol Biol 2009; 472: 323-42.
- Parney IF, Hao C, Petruk KC. Glioma immunology and immunotherapy. Neurosurgery 2000; 46: 778-91.
- [5] Denning GM. IL-4 and IFN-gamma synergistically increase total polymeric IgA receptor levels in human intestinal epithelial cells. Role of protein tyrosine kinases. J Immunol 1996; 156: 4807-14.
- [6] Kvale D, Løvhaug D, Sollid LM, Brandtzaeg P. Tumor necrosis factor-alpha up-regulates expression of secretory component, the epithelial receptor for polymeric Ig. J Immunol 1988; 140: 3086-9.
- [7] Rojas R, Apodaca G. Immunoglobulin transport across polarized epithelial cells. Nat Rev Mol Cell Biol 2002; 3: 944-55.

- [8] Kaetzel CS. The polymeric immunoglobulin receptor: bridging innate and adaptive immune responses at mucosal surfaces. Immunol Rev 2005; 206: 83-99.
- [9] Poger ME, Hirsch BR, Lamm ME. Synthesis of secretory component by colonic neoplasms. Am J Pathol 1976; 82: 327-38.
- [10] Harris JP, Caleb MH, South MA. Secretory component in human mammary carcinoma. Cancer Res 1975; 35: 1861-4.
- [11] Harris JP, South MA. Secretory component: a glandular epithelial cell marker. Am J Pathol 1981; 105: 47-53.
- [12] DeSouza LV, Krakovska O, Darfler MM, Krizman DB, Romaschin AD, Colgan TJ, Siu KW. mTRAQ-based quantification of potential endometrial carcinoma biomarkers from archived formalin-fixed paraffin-embedded tissues. Proteomics 2010; 10: 3108-16.
- [13] DeSouza LV, Romaschin AD, Colgan TJ, Siu KW. Absolute quantification of potential cancer markers in clinical tissue homogenates using multiple reaction monitoring on a hybrid triple quadrupole/linear ion trap tandem mass spectrometer. Anal Chem 2009; 81: 3462-70.
- [14] Rossel M, Billerey C, Bittard H, Ksiazek P, Alber D, Revillard JP, Vuitton DA. Alterations in polymeric immunoglobulin receptor expression and secretory component levels in bladder carcinoma. Urol Res 1991; 19: 361-6.
- [15] Rossel M, Seilles E, Voigt JJ, Vuitton D, Legait N, Revillard JP. Polymeric Ig receptor expression in hepatocellular carcinoma. Eur J Cancer 1992; 28A: 1120-4.
- [16] Ai J, Tang Q, Wu Y, Xu Y, Feng T, Zhou R, Chen Y, Gao X, Zhu Q, Yue X, Pan Q, Xu S, Li J, Huang M, Daugherty-Holtrop J, He Y, Xu HE, Fan J, Ding J, Geng M. The role of polymeric immunoglobulin receptor in inflammation-induced tumor metastasis of human hepatocellular carcinoma. J Natl Cancer Inst 2011; 103: 1696-712.

- [17] Xiao T, Ying W, Li L, Hu Z, Ma Y, Jiao L, Ma J, Cai Y, Lin D, Guo S, Han N, Di X, Li M, Zhang D, Su K, Yuan J, Zheng H, Gao M, He J, Shi S, Li W, Xu N, Zhang H, Liu Y, Zhang K, Gao Y, Qian X, Cheng S. An approach to studying lung cancerrelated proteins in human blood. Mol Cell Proteomics 2005; 4: 1480-6.
- [18] Rossel M, Brambilla E, Billaud M, Vuitton DA, Blanc-Jouvan F, Biichle S, Revillard JP. Nonspecific increased serum levels of secretory component in lung tumors: relationship to the gene expression of the transmembrane receptor form. Am J Respir Cell Mol Biol 1993; 9: 341-6.
- [19] Makawita S, Smith C, Batruch I, Zheng Y, Rückert F, Grützmann R, Pilarsky C, Gallinger S, Diamandis EP. Integrated proteomic profiling of cell line conditioned media and pancreatic juice for the identification of pancreatic cancer biomarkers. Mol Cell Proteomics 2011; 10: M111.008599.
- [20] Kvale D, Norstein J, Meling GI, Børmer OP, Brandtzaeg P, Langmark F, Rognum TO. Circulating secretory component in relation to early diagnosis and treatment of liver metastasis from colorectal carcinomas. J Clin Pathol 1992; 45: 568-71.
- [21] Dunbar E, Yachnis AT. Glioma diagnosis: immunohistochemistry and beyond. Adv Anat Pathol 2010; 17: 187-201.
- [22] Wen PY, Brandes AA. Treatment of recurrent high-grade gliomas. Curr Opin Neurol 2009; 22: 657-64.
- [23] Rousseau A, Mokhtari K, Duyckaerts C. The 2007 WHO classification of tumors of the central nervous system - what has changed? Curr Opin Neurol 2008; 21: 720-7.
- [24] Mestecky J, McGhee JR. Immunoglobulin A (IgA): molecular and cellular interactions involved in IgA biosynthesis and immune response. Adv Immunol 1987; 40: 153-245.