

## Original Article

# Local transplantation of osteogenic pre-differentiated autologous adipose-derived mesenchymal stem cells may accelerate non-union fracture healing with limited pro-metastatic potency

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Received December 10, 2014; Accepted December 20, 2014; Epub January 15, 2015; Published January 30, 2015

**Abstract:** Fracture non-union is a serious complication in orthopedic clinical practice. Mesenchymal stem cells are believed to play a vital role in fracture healing process. Among various origins of mesenchymal stem cell, adipose derived stem cells hold great promise especially in clinical milieu. However, the wide spread application of mesenchymal stem cell based therapy is impeded by the pro-metastasis nature of the mesenchymal stem cell itself. Based on the findings from previous studies, we hypothesize that local transplanted osteogenic pre-differentiated adipose stem cell may promote the non-union fracture healing. Moreover, the pre-differentiation stem cells by down-regulating the expression of CCL5 and CCL2. This novel osteogenic pre-differentiation technique may help clinical orthopedists to resolve the refractory non-union cases and shed new light on other stem cell based therapies to counteract to avoid the pro-metastasis nature of the mesenchymal stem cells.

**Keywords:** ADSC, pre-differentiation, fracture non-union, pro-metastatic

## Introduction

Nonunion of bone fracture is a rare but refractory complication in orthopedic clinical practice, which can severely compromise prognostic function. Based on research datum, in the United states, 5% to 10% of all diaphyseal fractures will encounter delayed union or nonunion problem [1-5]. Major symptoms of fracture non-union include persistent pain, stiffness of surrounding joints and limb disability which ultimately causing unemployment. Therefore fracture nonunion imposes both economical and psychological burden on patients as well as their families [6, 7].

At least three cellular events are required to initiate fracture healing including chemo-attractive recruitment, inductive proliferation, and osteogenic differentiation [8-12]. Recently more and more evidence show that mesenchymal stem cell (MSC) population is the fundamental ancestor of these cellular assemblages and plays a vital role in fracture repair and nonunion

pathology. Following bone fracture, disrupted bone matrix and degranulated platelets release various cytokines to the fracture site forming a chemo-attractive environment which recruit MSC to the damaged area [13]. Moreover, communication with the local cell population occurs to stimulate MSC osteoblastic capabilities. Carter DR et al reported favorable biologic and mechanical environments result in proliferation and differentiation of MSC to osteoblasts and chondrocytes [14]. Furthermore, by studying 35 nonunion patient's bone marrow, Hernigou and Beaujean reported that reduction in bone-producing stem cell population in the fracture hematoma may contribute to bone consolidation malfunction [15]. Also, they found that there were smaller stem cell morphology in the marrow of synovial pseudarthrosis patient than that of control group patient which give a hint on the relationship between MSC disorder and nonunion development [16].

As a promising cell source, MSC are demonstrated capable to self-renew and possess

multi-potent differentiation properties [18]. Also, evidence show that MSC exhibit non-immunogenic or hypo-immunogenic properties [17]. Barrilleaux and colleagues isolated fibroblast-like MSC from the stromal cell population from various tissues [18]. Bone marrow has been used as the major source of MSC and under appropriate conditions bone marrow derived MSC can be selectively induced into osteogenic lineage [19]. However, the harvest of bone marrow MSC is a highly invasive and painful procedure which prompts the quest for alternative sources from which to isolate MSC. Moreover, Bone marrow cellularity declines with age, and there is also a decrease in the prevalence of connective-tissue progenitors with increasing age which hinder the bone marrow stem cells in a wide range of clinical applications [20].

Like bone marrow, adipose tissue is mesoderm-derived organ containing stromal population such as microvascular endothelial cells, smooth muscle cells and stem cells [21]. These cells can be enzymatically isolated from adipose tissue (commonly from lipoaspirate) and separated from the buoyant adipocytes by centrifugation. A more homogeneous population emerges in culture under conditions supportive of MSC growth. From the last decade, this population which is termed with generic nomenclature, adipose-derived stem cell (ADSC) has been identified as possessing many of the properties of its counterpart from bone marrow including extensive self-renewal potential and the capacity to undergo multilineage differentiation [22-25]. Furthermore, the phenotypic and gene expression profiles of ADSC are similar to MSC obtained from bone marrow [23, 25].

Zuk and colleagues reported that ADSC can be expanded in vitro for extended periods [22]. Since humans have abundant subcutaneous fat deposits, ADSC can easily be isolated by conventional liposuction procedures, thus overcoming the tissue morbidity associated with bone marrow aspiration. Furthermore, the MSC frequency in bone marrow is somewhere between 1 in 25,000 to 1 in 100,000 cells whereas ADSC constitute approximately 2% of lipoaspirate cells [25-28]. Due to its abundance, relatively easy harvest, and high MSCs frequency, adipose derived stem cell might be a solid starting basis for further development of stem cell therapies.

Despite all the advantages MSC therapies possess, the potential risks of inducing uncon-

trolled cell growth pose a serious threat to the recipient patient. In 2007, Weinberg and colleagues demonstrate that when mixed with otherwise weakly metastatic human breast carcinoma cells, human bone-marrow-derived mesenchymal stem cells greatly increase the metastatic potency of the cancer cells. Furthermore, they showed that mesenchymal stem cells when stimulated by the breast cancer cells produce chemokine CCL5 (also called RANTES) which in turn acts in a paracrine fashion on the cancer cells to enhance their motility, invasion and metastasis [29].

### Hypotheses

Local transplantation of autologous osteogenic pre-differentiated ADSCs to the fracture gap of non-union fracture might be able to promote non-union fracture healing by stimulating local angiogenesis, producing abundant calcium deposit in early phase of post-surgical fracture healing and matching the transplanted ADSCs osteogenic function to the fracture healing time frame. Moreover, by limiting cell self-renewal capacity, osteogenic pre-differentiation treatment might greatly prevent the transplanted mesenchymal stem cell from neoplasia and cancer metastasis promoting behavior which will potentially facilitate the wide spread use of mesenchymal stem cell therapy. Based on all the facts above, we hypothesize that local transplantation of autologous osteogenic pre-differentiated ADSCs holds the promise to enhance non-fracture healing and reduce risk of uncontrolled cell growth.

### Evaluation of the hypothesis

#### *Match ADSCs's osteogenic function to fracture repair time frame*

Previous studies have shown that it normally take more than two to three weeks for ADSCs to exhibit calcified extracellular matrix in osteogenic induction culture [30]. Moreover, in vivo study showed human ADSCs possesses the capacity to form osteoid on appropriate biomaterials [30]. Consistently, Jaiswal and colleagues demonstrated JNK activation occurred on day 13 to day 17 in the osteogenic differentiation process, which was associated with extracellular matrix synthesis and increased calcium deposition, the two hallmarks of bone formation [31]. Based on these evidence, in our hypothesis, we first isolate autologous ADSCs using the patient's own adipose tissue. Next,

## Osteogenic pre-differentiated of ADSCs promote fracture healing

the ADSCs are pre-differentiated with osteogenic induction medium in culture for 14 days. Then, the pre-differentiated ADSCs are locally transplanted into the the fracture gap of non-union patient in ten non-union repair surgery. This procedure may help to match the osteogenic function of ADSCs to the subsequent fracture healing time frame and aid to facilitate early production of osteoid and wound calcification, which may accelerate the stabilization of fracture site and enhance the prognosis of non-union repair surgery.

### *Avoid the risk of mesenchymal stem cell promoted cancer metastasis*

There is a long history of clinical and experimental observations showing that metastases frequently occur at sites of injury. As stem cells preferentially migrate to tumors and sites of tissue injury [32], they may prepare the injured sites for subsequent colonization. Once there, hematopoietic stem cells are recruited to a so-called 'premetastatic niche', where they reorganize the matrix and establish sites at which tumor cells proliferate more frequently than at other locales to develop tumor metastases [33]. Recent studies elucidated that the human bone-marrow-derived mesenchymal stem cell (MSC) is recruited in large numbers and integrate into the stroma of developing tumors [34]. Moreover, Weinberg's team revealed that that human mesenchymal stem cells, by secreting the CCL5, act in a paracrine fashion on the cancer cells to enhance their motility, invasion and metastasis [29]. Also, CCL2 was shown to partly mediate the interaction between breast cancer cells and MSCs [35]. These findings suggest a potential adverse effect of MSC mediated therapy, which may promote the otherwise weakly metastatic cancer cells in patients to increase metastatic potency and eventually cause the spread of cancer. On the other hand, Djouad's data demonstrated that, as the MSC differentiating into chondrocytes, CCL5 expression is downregulated on day 7 under chondrogenic induction [36]. In addition, Molloy's study showed that MSC's secretion of CCL2 is dramatically decreased on day 10 and 14 during differentiation into osteoblasts [35]. Accordingly, in our current hypothesis, a 14 days of osteogenic pre-differentiation would potentially abrogate the crosstalk effect between ADSCs and cancer cells via downregulating the CCL5 and CCL2 expression both of which play pivotal role in mediating metastasis of cancer cells. Furthermore, pre-differentia-

tion of ADSCs also serve to reduce the possibility of uncontrolled cell proliferation and increase the functional cell fraction in the total transplanted cells.

### **Conclusion**

Fracture non-union is a clinically refractory complication which seriously affecting the life quality of patients. Recently, MSC mediated therapy emerged as a promising solution to enhance the wound healing capacity. Among all source of MSC, ADSCs possess many advantages over other origins such as abundant sources, easy to harvest, and high MSCs frequency. However, despite MSC has been a hot spot of researches for over a decade, few have been interpreted into clinical therapy owing to its pro-metastatic nature. Based on reported studies, we present a hypothesis that local transplantation of autologous osteogenic pre-differentiated ADSCs into the fracture gap holds great promise to enhance non-union fracture healing while remitting the risk of systemic and local tumor metastasis. Our current hypothesis, if proven to be valid, will not only promote the outcome of fracture non-union cases with enhanced bone formation capacity, but also will shed new light on the other applications of stem cell based therapies.

### **Acknowledgements**

This work was funded by Chinese National Ministry of Science and Technology 973 Project Planning (No. 2014CB542200); The ministry of education innovation team (IRT1201); the National Natural Science Fund (No. 31271284, 31171150, 81171146, 31471144, 30971526, 31100860, 31040043, 31371210, 8137-2044), and the Educational Ministry New Century Excellent Talents Support Project (No. BMU20110270) and The Beijing Natural Science Foundation (7142164).

### **Disclosure of conflict of interest**

None.

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### **References**

- [1] Blick SS, Brumback RJ, Lakatos R, Poka A, Burgess AR. Early prophylactic bone grafting of

## Osteogenic pre-differentiated of ADSCs promote fracture healing

- high-energy tibial fractures. *Clin Orthop Relat Res* 1989; 240: 21-41.
- [2] DeLee JC, Heckman JD, and Lewis AG. Partial fibulectomy for ununited fractures of the tibia. *J Bone Joint Surg Am* 1981; 63: 1390-5.
- [3] Fernandez-Palazzi F. Fibular resection in delayed union of tibial fractures. *Acta Orthop Scand* 1969; 40: 105-18.
- [4] Hanson LW, Eppright RH. Posterior bone-grafting of the tibia for non-union: A review of twenty-four cases. *J Bone Joint Surg* 1966; 48A: 27-43.
- [5] Jones KG. Treatment of infected nonunion of the tibia through the posterolateral approach. *Clin Orthop* 1965; 43: 103-109.
- [6] Bondurant FJ, Cotler HB, Buckle R, Miller-Crotchett P, Browner BD. The medical and economic impact of severely injured lower extremities. *J Trauma* 1988; 28: 1270-3.
- [7] Johnson KD. Management of malunion and nonunion of the tibia. *Orthop Clin North Am* 1987; 18: 157-71.
- [8] In: Aaron AD. Bone Healing and Grafting. In: Kasser JR, editor. *Orthopaedic Knowledge Update 5*. Rosemont, IL: American Academy of Orthopaedic Surgeons; 1996. pp. 21-28.
- [9] Cook SD, Baffes GC, Wolfe MW, Sampath TK, Rueger DC, Whitecloud TS 3rd. The effect of recombinant human osteogenic protein-1 on healing of large segmental bone defects. *J Bone Joint Surg Am* 1994; 76: 827-38.
- [10] Konttinen Y, Imai S and Suda A. Neuropeptides and the puzzle of bone remodeling. *State of the art. Acta Orthop Scand* 1996; 67: 632-9.
- [11] Kristiansen TK. Accelerated healing of distal radial fractures with the use of specific, low-intensity ultrasound. A multicenter, prospective, randomized, double-blind, placebo-controlled study. *J Bone Joint Surg Am* 1997; 79: 961-73.
- [12] Lind M. Growth factors: possible new clinical tools. A review. *Acta Orthop Scand* 1996; 67: 407-17.
- [13] Yoo JU and Johnstone B. The role of osteochondral progenitor cells in fracture repair. *Clin Orthop Relat Res* 1998; Suppl 355: S73-81.
- [14] Carter DR, Beaupré GS, Giori NJ, Helms JA. Mechanobiology of skeletal regeneration. *Clin Orthop Relat Res* 1998; 355 Suppl: S41-55.
- [15] Hernigou P and Beaujean F. [Bone marrow in patients with pseudarthrosis. A study of progenitor cells by in vitro cloning]. *Rev Chir Orthop Reparatrice Appar Mot* 1997; 83: 33-40.
- [16] Hernigou P and Beaujean F. [Pseudarthrosis treated by percutaneous autologous bone marrow graft]. *Rev Chir Orthop Reparatrice Appar Mot* 1997; 83: 495-504.
- [17] Barry FP and Murphy JM. Mesenchymal stem cells: clinical applications and biological characterization. *Int J Biochem Cell Biol* 2004; 36: 568-84.
- [18] Barrilleaux B, Elçin YM. Review: ex vivo engineering of living tissues with adult stem cells. *Tissue Eng* 2006; 12: 3007-19.
- [19] Pittenger MF. Multilineage potential of adult human mesenchymal stem cells. *Science* 1999; 284: 143-7.
- [20] Muschler GF, Nitto H, Boehm CA, Easley KA. Age- and gender-related changes in the cellularity of human bone marrow and the prevalence of osteoblastic progenitors. *J Orthop Res* 2001; 19: 117-25.
- [21] Zuk PA, Zhu M, Mizuno H, Huang J, Futrell JW, Katz AJ, Benhaim P, Lorenz HP, Hedrick MH. Multilineage cells from human adipose tissue: implications for cell-based therapies. *Tissue Eng* 2001; 7: 211-28.
- [22] Zuk PA, Zhu M, Ashjian P, De Ugarte DA, Huang JI, Mizuno H, Alfonso ZC, Fraser JK, Benhaim P, Hedrick MH. Human adipose tissue is a source of multipotent stem cells. *Mol Biol Cell* 2002; 13: 4279-95.
- [23] De Ugarte DA, Morizono K, Elbarbary A, Alfonso Z, Zuk PA, Zhu M, Drago J, Ashjian P, Thomas B, Benhaim P, Chen I, Fraser J, Hedrick MH. Comparison of multi-lineage cells from human adipose tissue and bone marrow. *Cells Tissues Organs* 2003; 174: 101-9.
- [24] Gimble J and Guilak F. Adipose-derived adult stem cells: isolation, characterization, and differentiation potential. *Cytotherapy* 2003; 5: 362-9.
- [25] Strem BM, Hicok KC, Zhu M, Wulur I, Alfonso Z, Schreiber RE, Fraser JK, Hedrick MH. Multipotential differentiation of adipose tissue-derived stem cells. *Keio J Med* 2005; 54: 132-41.
- [26] Banfi A, Bianchi G, Galotto M, Cancedda R, Quarto R. Bone marrow stromal damage after chemo/radiotherapy: occurrence, consequences and possibilities of treatment. *Leuk Lymphoma* 2001; 42: 863-70.
- [27] D'Ippolito G, Schiller PC, Ricordi C, Roos BA, Howard GA. Age-related osteogenic potential of mesenchymal stromal stem cells from human vertebral bone marrow. *J Bone Miner Res* 1999; 14: 1115-22.
- [28] Muschler GF, Nitto H, Boehm CA, Easley KA. Age- and gender-related changes in the cellularity of human bone marrow and the prevalence of osteoblastic progenitors. *J Orthop Res* 2001; 19: 117-25.
- [29] Karnoub AE, Dash AB, Vo AP, Sullivan A, Brooks MW, Bell GW, Richardson AL, Polyak K, Tubo R, Weinberg RA. Mesenchymal stem cells within tumour stroma promote breast cancer metastasis. *Nature* 2007; 449: 557-63.
- [30] Hicok KC, Du Laney TV, Zhou YS, Halvorsen YD, Hitt DC, Cooper LF, Gimble JM. Human adi-

## Osteogenic pre-differentiated of ADSCs promote fracture healing

- pose-derived adult stem cells produce osteoid in vivo. *Tissue Eng* 2004; 10: 371-80.
- [31] Jaiswal RK. Adult human mesenchymal stem cell differentiation to the osteogenic or adipogenic lineage is regulated by mitogen-activated protein kinase. *J Biol Chem* 2000; 275: 9645-52.
- [32] Rafii S and Lyden D. Therapeutic stem and progenitor cell transplantation for organ vascularization and regeneration. *Nat Med* 2003; 9: 702-12.
- [33] Kaplan RN, Psaila B, Lyden D. VEGFR1-positive haematopoietic bone marrow progenitors initiate the pre-metastatic niche. *Nature* 2005; 438: 820-7.
- [34] Hall B, Andreeff M and Marini F. The participation of mesenchymal stem cells in tumor stroma formation and their application as targeted-gene delivery vehicles. *Handb Exp Pharmacol* 2007; 180: 263-83.
- [35] Molloy AP, Martin FT, Dwyer RM, Griffin TP, Murphy M, Barry FP, O'Brien T, Kerin MJ. Mesenchymal stem cell secretion of chemokines during differentiation into osteoblasts, and their potential role in mediating interactions with breast cancer cells. *Int J Cancer* 2009; 124: 326-32.
- [36] Djouad F, Delorme B, Maurice M, Bony C, Apparailly F, Louis-Plence P, Canovas F, Charbord P, Noël D, Jorgensen C. Microenvironmental changes during differentiation of mesenchymal stem cells towards chondrocytes. *Arthritis Res Ther* 2007; 9: R33.