

Original Article

Effect of diphenoxylate on CYP450 isoforms activity in rats

Peiwu Geng^{1*}, Jinzhang Cai^{2*}, Shuanghu Wang¹, Suping Yang³, Zezheng Liu³, Yingying Lin³, Congcong Wen³, Xianqin Wang^{3,4}, Yunfang Zhou¹, Meiling Zhang^{3,4}

¹Laboratory of Clinical Pharmacy, The People's Hospital of Lishui, Wenzhou Medical University, Lishui 323000, China; ²The Second Affiliated Hospital & Yuying Children's Hospital, Wenzhou Medical University, Wenzhou 325000, China; ³Analytical and Testing Center of Wenzhou Medical University, Wenzhou 325035, China; ⁴Shanghai Key Laboratory of Forensic Medicine, Institute of Forensic Science, Ministry of Justice, Shanghai 200063, China. *Equal contributors.

Received August 21, 2015; Accepted October 13, 2015; Epub October 15, 2015; Published October 30, 2015

Abstract: In order to investigate the effects of diphenoxylate on the metabolic capacity of cytochrome P450 (CYP) enzymes, a cocktail method was employed to evaluate the activities of CYP2B6, CYP2D6, CYP2C19, CYP1A2, CYP3A4, CYP2C9. The rats were randomly divided into diphenoxylate group (Low, Medium, High) and control group. The diphenoxylate group rats were given 12, 24, 48 mg/kg (Low, Medium, High) diphenoxylate by continuous intra-gastric administration for 7 days. Six probe drugs bupropion, metoprolol omeprazole, phenacetin, testosterone and tolbutamide were given to rats through intragastric administration, and the plasma concentrations were determined by UPLC-MS/MS. Statistical pharmacokinetics difference for omeprazole, phenacetin and tolbutamide in rats were observed by comparing diphenoxylate group with control group. Continuous 7 days-intragastric administration of diphenoxylate induces the activities of CYP2C19, CYP1A2 and CYP2C9 of rats. Induction of drug metabolizing enzyme by diphenoxylate would reduce the efficacy of other drug. Additionally, high dosage diphenoxylate may cause hepatotoxicity.

Keywords: CYP450, diphenoxylate, cocktail, UPLC-MS/MS, rat

Introduction

Diphenoxylate, a weak opioid agonist has been considered as a drug of low abuse potential. There have been however, only case reports of diphenoxylate being abused by patients with medical illnesses and substance users [1, 2]. It has also been tried during detoxification from opioids and has been shown to decrease withdrawal [3, 4]. Diphenoxylate is useful in the treatment of protracted diarrhea in children and disorders in which rapid gastrointestinal transit is the main cause of diarrhea. However, it is not recommended for acute infective diarrhea in childhood. Diphenoxylate has also been found to provide substantial relief from detoxification symptoms in opioid-dependent patients with no drug-associated adverse effects [1].

Cytochrome P450 (CYP) enzymes are responsible for most biotransformation steps of xeno-

biotics and endogenous molecules [5]. Variations of their activity by inhibition or induction can influence the pharmacokinetics and thereby the effect of drugs (of abuse). Enzyme inhibition by co-administered drugs (of abuse) and/or genetic variations of their expression can increase the risk of adverse reactions [6] or reduce the desired effect [7]. Such drug-drug interactions were described as a major reason for hospitalization or even death [8].

So far, no study on the effects of diphenoxylate on the metabolic capacity of CYP enzyme was reported. Therefore, in this study, six probe drugs were employed to evaluate effect of diphenoxylate on the metabolic capacity of CYP2B6, CYP2D6, CYP2C19, CYP1A2, CYP3A4, CYP2C9. The effects of diphenoxylate on rat CYP enzyme activity will be evaluated according to the pharmacokinetic parameters changes of

Effect of diphenoxylate on CYP450 isoforms activity of rats

Table 1. Pharmacokinetic parameters of bupropion and metoprolol from control group and diphenoxylate group rats (mean \pm SD, n=8)

Parameters		AUC _(0-t)	AUC _(0-∞)	t _{1/2}	CL	V	C _{max}
		ng/mL *h	ng/mL *h	h	L/h/kg	L/kg	ng/mL
Bupropion (CYP2B6)	Control	974.9 \pm 1167.4	1024.4 \pm 1147.6	1.8 \pm 1.1	15.8 \pm 7.8	46.6 \pm 42.6	527.0 \pm 623.5
	Low	1194.1 \pm 1025.4	1512.1 \pm 1817.8	1.8 \pm 1.6	12.6 \pm 7.6	24.5 \pm 15.1	348.3 \pm 238.1
	Medium	1106.6 \pm 449.0	1156.5 \pm 455.7	1.7 \pm 0.5	9.8 \pm 3.5	25.5 \pm 17.9	333.8 \pm 147.1
	High	1456.9 \pm 1014.0	2877.1 \pm 3298.5	5.6 \pm 5.6	10.2 \pm 11.2	48.8 \pm 43.0	369.7 \pm 285.1
Metoprolol (CYP2D6)	Control	890.7 \pm 848.9	906.1 \pm 849.2	1.2 \pm 0.4	16.7 \pm 9.9	28.8 \pm 13.8	463.5 \pm 468.4
	Low	1120.5 \pm 738.5	2045.7 \pm 3141.1	3.2 \pm 5.0	10.9 \pm 5.8	25.6 \pm 13.0	365.4 \pm 172.6
	Medium	1184.1 \pm 558.6	1295.7 \pm 552.3	2.2 \pm 0.9*	8.9 \pm 3.2	30.4 \pm 19.0	337.4 \pm 183.9
	High	1550.2 \pm 1010.5	2294.0 \pm 1697.1	3.9 \pm 2.2**	8.2 \pm 7.6	39.7 \pm 32.0	367.5 \pm 239.7

Diphenoxylate group was compared with the control group, *: P<0.05, **: P<0.01.

six specific probe drugs (bupropion, metoprolol omeprazole, phenacetin, testosterone and tolbutamide).

Material and methods

Chemicals

Bupropion, metoprolol omeprazole, phenacetin, testosterone and tolbutamide (all >98%) and the internal standard diazepam (IS) were obtained from Sigma-Aldrich Company (St. Louis, USA). Ultra-pure water was prepared by Millipore Milli-Q purification system (Bedford, USA). Methanol and acetonitrile (HPLC grade) were obtained from Merck Company (Darmstadt, Germany).

Animals

Sprague-Dawley rats (male, 220 \pm 20 g) purchased from Shanghai SLAC Laboratory Animal Co., Ltd. Animals were housed under a natural light-dark cycle conditions with controlled temperature (22°C). All forty rats were housed at Laboratory Animal Research Center of Wenzhou Medical University. All experimental procedures were approved ethically by the Wenzhou Medical University Administration Committee of Experimental Animals.

Pharmacokinetics

Forty rats (220 \pm 20 g) were randomly divided into four different dosages of diphenoxylate groups (Low-group, Medium-group, High-group and control group with 8 rats in each group). Diphenoxylate was dissolved in corn oil as suspension at three different concentrations (12,

24, and 48 mg/mL). Three different diphenoxylate group (Low-group, Medium-group, High-group) were respectively give diphenoxylate 12, 24, 48 mg/kg one time by intragastric administration at every morning, and last for 7 days. Control group were give saline by same administration method. At 8 days morning, six probe drugs bupropion, metoprolol omeprazole, phenacetin, testosterone and tolbutamide were mixed in corn oil and given to the rats of three diphenoxylate groups and control group by intragastric administration at a single dosage 10 mg/kg for bupropion, metoprolol omeprazole, phenacetin, testosterone, 1 mg/kg for tolbutamide.

Blood (0.3 mL) samples were collected into heparinized 1.5 mL polythene tubes from the tail vein at 0.0833, 0.5, 1, 2, 3, 4, 6, 8, 12, 24, 48 h after intragastric administration of six probe drugs. 100 μ L of plasma was obtained from blood sample after centrifugation at 4000 g for 10 min. In a 1.5 mL centrifuge tube, 200 μ L of acetonitrile (containing 50 ng/mL IS) was added into 100 μ L of collected plasma sample. After vortex-mixing for 1.0 min, the sample was centrifuged at 13000 g for 15 min. Then supernatant (2 μ L) was injected into the UPLC-MS/MS system for analysis.

Concentration of plasma probe drugs versus time was analyzed by Version 3.0 Data Analysis System (Wenzhou Medical University, China). The main pharmacokinetic parameters of the diphenoxylate group and control group were analyzed by SPSS 18.0 statistical software; statistical significance was assessed by t-test (P<0.05 was considered as statistically significant).

Effect of diphenoxylate on CYP450 isoforms activity of rats

Table 2. Pharmacokinetic parameters of omeprazole and phenacetin in control group and diphenoxylate group rats (mean \pm SD, n=8)

Parameters		AUC _(0-t)	AUC _(0-∞)	t _{1/2}	CL	V	C _{max}
		ng/mL *h	ng/mL *h	h	L/h/kg	L/kg	ng/mL
Omeprazole (CYP2C19)	Control	2347.9 \pm 2800.6	2555.9 \pm 3056.9	1.7 \pm 0.6	16.4 \pm 16.3	41.1 \pm 46.1	804.8 \pm 717.1
	Low	1070.2 \pm 889.1	1133.5 \pm 983.7	1.3 \pm 0.5	15.2 \pm 10.5	28.1 \pm 28.9	651.6 \pm 397.1
	Medium	772.0 \pm 470.5	797.0 \pm 462.3	1.2 \pm 0.3*	15.3 \pm 6.0	26.2 \pm 11.6	444.7 \pm 313.3
	High	285.4 \pm 274.1*	291.2 \pm 278.9*	0.9 \pm 0.2**	63.6 \pm 40.9**	87.1 \pm 65.1	169.7 \pm 161.5*
Phenacetin (CYP1A2)	Control	11054.1 \pm 5810.3	11062.4 \pm 5814.9	1.1 \pm 0.3	1.3 \pm 0.9	2.2 \pm 2.0	5773.8 \pm 1933.2
	Low	13016.2 \pm 11922.6	13789.8 \pm 12841.4	1.9 \pm 0.6	1.9 \pm 2.3	6.4 \pm 10.1	3962.2 \pm 2311.5
	Medium	5303.2 \pm 1837.5*	5382.5 \pm 1777.6*	3.3 \pm 2.6*	2.1 \pm 0.7	11.5 \pm 12.7	2575.2 \pm 984.1**
	High	1694.0 \pm 1909.1**	1698.4 \pm 1909.0**	1.3 \pm 0.5	12.2 \pm 9.6**	19.1 \pm 13.4**	1117.4 \pm 869.5**

Diphenoxylate group was compared with the control group, *: P<0.05, **: P<0.01.

Table 3. Pharmacokinetic parameters of testosterone and tolbutamide in control group and diphenoxylate group rats (mean \pm SD, n=8)

Parameters		AUC _(0-t)	AUC _(0-∞)	t _{1/2}	CL	V	C _{max}
		ng/mL *h	ng/mL *h	h	L/h/kg	L/kg	ng/mL
Testosterone (CYP3A4)	Control	317.1 \pm 553.3	363.2 \pm 650.9	2.4 \pm 1.4	107.9 \pm 75.2	318.9 \pm 302.0	148.0 \pm 222.2
	Low	121.1 \pm 67.4	129.6 \pm 69.5	2.0 \pm 1.1	103.1 \pm 59.3	258.6 \pm 141.2	37.8 \pm 22.0
	Medium	264.3 \pm 102.4	333.6 \pm 135.3	6.4 \pm 3.1*	35.3 \pm 15.8	325.7 \pm 269.2	116.3 \pm 90.7
	High	118.3 \pm 68.0	147.2 \pm 100.9	4.7 \pm 3.7	108.7 \pm 81.1	477.1 \pm 259.2	87.1 \pm 33.9
Tolbutamide (CYP2C9)	Control	49556.8 \pm 8970.1	64492.5 \pm 19288.1	10.3 \pm 4.4	0.02 \pm 0.01	0.23 \pm 0.05	3865.4 \pm 501.4
	Low	37873.9 \pm 10951.5*	43875.7 \pm 13099.8*	8.0 \pm 3.0	0.03 \pm 0.01*	0.28 \pm 0.12	3277.7 \pm 810.9
	Medium	30817.8 \pm 13569.0**	45061.2 \pm 24307.4*	8.3 \pm 5.2	0.03 \pm 0.01*	0.28 \pm 0.10	2970.9 \pm 1031.3*
	High	30583.3 \pm 10748.2**	35607.9 \pm 18796.1**	7.1 \pm 3.6	0.03 \pm 0.01**	0.29 \pm 0.03*	2633.3 \pm 888.8**

Diphenoxylate group was compared with the control group, *: P<0.05, **: P<0.01.

UPLC-MS/MS determination of probe drugs

The concentration of bupropion, metoprolol omeprazole, phenacetin, testosterone and tolbutamide in rat plasma were simultaneously determined by a sensitive and simple UPLC-MS/MS method [9]. The compounds were analyzed by a UPLC-MS/MS with ACQUITY I-Class UPLC and a XEVO TQD triple quadrupole mass spectrometer that equipped with an electrospray ionization (ESI) interface (Waters Corp., Milford, MA, USA). Data acquisition and instrument control were performed on the Masslynx 4.1 software (Waters Corp., Milford, MA, USA).

Bupropion, metoprolol omeprazole, phenacetin, testosterone, tolbutamide and diazepam (IS) were separated using a Waters BEH C18 column (2.1 mm \times 100 mm, 1.7 μ m) at constant temperature 40°C. The initial mobile phase consisted of 0.1% formic acid and acetonitrile with gradient elution at a flow rate of 0.4 mL/min and an injection volume of 2 μ L. Elution was in a linear gradient, with the acetonitrile changing from 30 to 60% in 0.3-1.8 min and

increasing up to 95% over 0.2 min. The acetonitrile content was maintained at 95% for 0.5 min and decreased to 30% within 0.1 min, and then maintained at 30% for 0.4 min.

The mass spectrometric detection was performed in a positive mode. Nitrogen was used as the cone gas (50 L/h) and desolvation gas (1000 L/h). The mass conditions were set as follows: source temperature 150°C; capillary voltage 2.5 kV; desolvation temperature 500°C. The multiple reaction monitoring (MRM) mode with m/z 240.1 \rightarrow 184.1 for bupropion, m/z 268.1 \rightarrow 115.8 for metoprolol, m/z 346.1 \rightarrow 197.8 for omeprazole, m/z 180.1 \rightarrow 109.9 for phenacetin, m/z 289.0 \rightarrow 97.0 for testosterone, m/z 271.2 \rightarrow 155.1 for tolbutamide, and m/z 285.1 \rightarrow 193.1 for IS was used for quantitative analysis.

The LLOQ for each probe drug in plasma was 2 ng/mL. The RSD of the six probe drugs were less than 15%. The calibration plot of the probe drugs is in the range of 2-2000 ng/mL (r>0.995). The intra-day and inter-day accuracy

Effect of diphenoxylate on CYP450 isoforms activity of rats

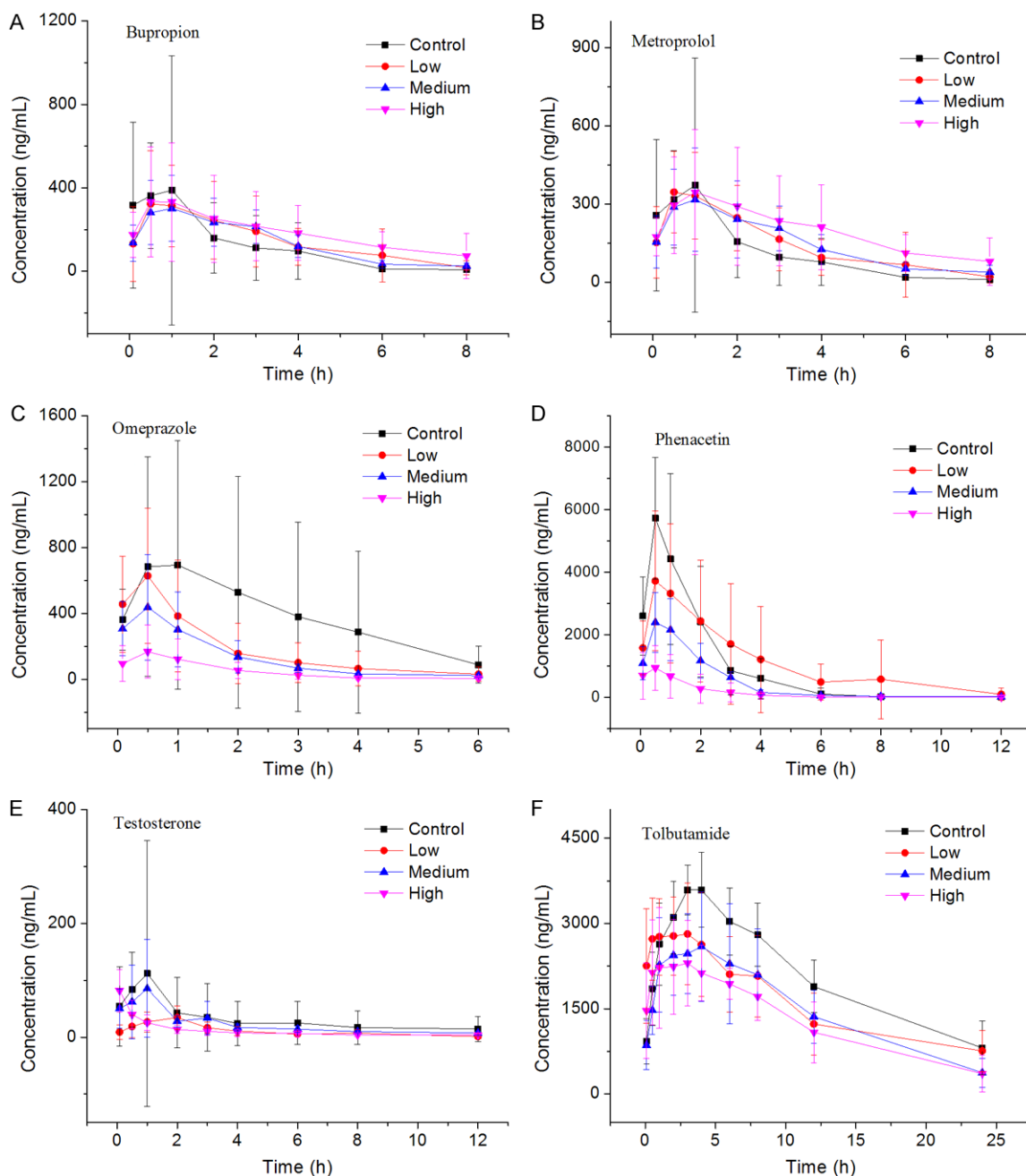


Figure 1. The pharmacokinetic profiles of bupropion (A), metoprolol (B), omeprazole (C), phenacetin (D), testosterone (E), tolbutamide (F) in control group and diphenoxylate group (low, medium, high) rats (n=8).

ranged from 90% to 115%. The matrix effects were more than 82% or less than 113%. The extraction recoveries were better than 85%.

Histopathology

After pharmacokinetic properties analysis, rats were deeply anesthetized with 10% chloral hydrate (i.p., 20 mg/kg). The liver of control

group and diphenoxylate treated groups were rapidly isolated and immersed in freshly prepared 4% w/v formaldehyde (0.1 M phosphate buffer, pH 7.2) for 48 h, and then embedded in paraffin. Then 5 μ m-thick histologic sections were prepared and stained with routine HE method (hematoxylin and eosin). The morphological changes of liver were observed under light microscope.

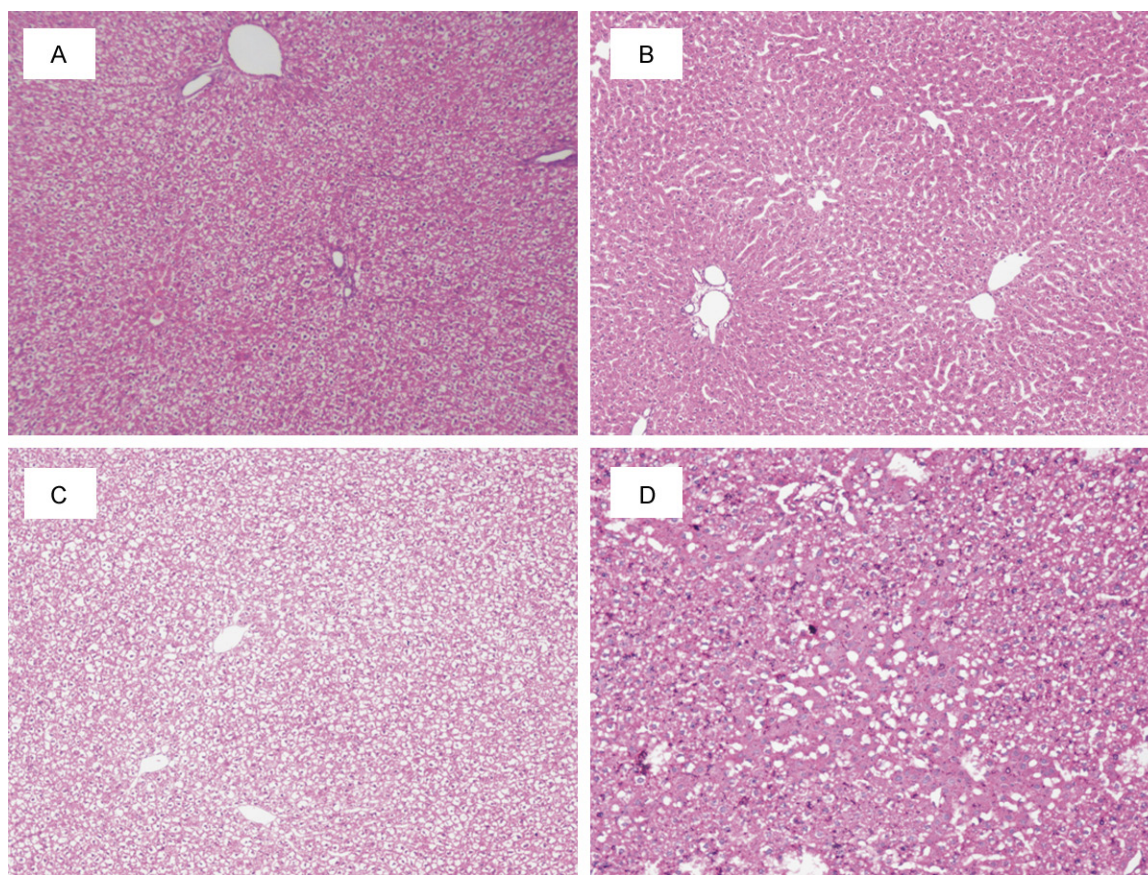


Figure 2. Morphological changes of liver in diphenoxylate treated group at low (A), medium (B), high (C) dosage and control group (D) (hematoxylin-eosin staining, $\times 100$).

Results

Pharmacokinetics

The main pharmacokinetic parameters of bupropion, metoprolol omeprazole, phenacetin, testosterone and tolbutamide calculated from non-compartment model analysis were summarized in **Tables 1-3**. The representative profiles of concentration of drugs (bupropion, metoprolol omeprazole, phenacetin, testosterone and tolbutamide) vs. time were presented in **Figure 1**.

From the **Table 1**, compared with the control group, no difference in pharmacokinetic behaviors can be observed between low, medium dosage group and control group for bupropion and metoprolol. While from the **Table 2**, the pharmacokinetic behaviors of omeprazole in high dosage group compared with the control group, $AUC_{(0-t)}$ decreased ($P<0.05$), CL increas-

ed ($P<0.01$), C_{max} decreased ($P<0.05$), and no difference in pharmacokinetic behaviors can be observed between low, medium dosage group and control group. The pharmacokinetic behaviors of phenacetin in medium and high dosage group compared with the control group, $AUC_{(0-t)}$ decreased (medium, $P<0.05$; high, $P<0.01$), CL increased (medium, $P>0.05$; high, $P<0.01$), C_{max} decreased (medium, $P<0.01$; high, $P<0.01$), and no difference in pharmacokinetic behaviors can be observed between low dosage group and control group. From the **Table 3**, compared with the control group, no difference in pharmacokinetic behaviors can be observed between low, medium dosage group and control group for testosterone. While for tolbutamide, compared with the control group, $AUC_{(0-t)}$ decreased (low, $P<0.05$; median and high, $P<0.01$), CL increased (Low and median, $P<0.05$; high, $P<0.01$), C_{max} decreased (low, $P>0.05$; median, $P<0.05$; high, $P<0.01$).

Morphological changes of liver

There is no significant difference between control group and low dose group in the liver lobules at low magnification, **Figure 2**. Both of them are intact and liver cells were arranged tightly along with central veins. In middle dose group, the structure of liver lobule is still intact and can be recognized easily. There liver cells arranged regularly, and there was no swelling in them. The nucleus of liver cells is round, clear and fine luster. However, the cell gap between liver cells seems becoming larger than control group and low dose group.

While, compared with control group the intact structures of liver lobule are damaged obviously in high dose group. The regular and tight arrangement of liver cells along with central veins was disappeared. Most of liver cells are arranged in disorder. Beside, the some gaps of hepatic sinusoid decreased and disappeared, and the liver cells became edema and appeared to be vacuolization.

Discussion

In general, changes in pharmacokinetics are thought to be caused by drug-drug or drug-food interactions [10]. In pharmacokinetic interactions, approximately 65% of drug-drug interactions occur in metabolic sites, and drug metabolic enzymes are considered to be the most important interactive sites. A large number of drugs are metabolized by CYP enzymes in the liver, and more than 90% of drug-drug interactions occur at the CYP-catalyzed step [11, 12]. Similarly, supplement-drug interactions involving CYP activity are occasionally found to cause considerable adverse events. For these reasons, we evaluated the effects of acute paraquat poisoning on the activity of CYP enzymes *in vivo*. We selected CYP isoforms CYP1A2, CYP2D6, CYP3A4, CYP2C19, CYP2C9 and CYP2B6 because more than 90% of drugs are known to be metabolized by these 6 CYP enzymes [13, 14].

There no significant difference for AUC, CL and C_{max} of bupropion, metoprolol and testosterone between the diphenoxylate group (low, median, high) and control group was observed. It suggested that the diphenoxylate was not able to induce or inhibit the activity of CYP2B6, CYP2D6 and CYP3A4 enzyme. The pharmacoki-

netic parameters of omeprazole experienced obvious change with decreased $AUC_{(0-t)}$ and increased CL after the dosage increase. This result indicates that the 7 days-intragastric administration of diphenoxylate with high dosage slightly induces the metabolism of omeprazole (CYP2C19) in rat. The pharmacokinetic parameters of phenacetin experienced obvious change with decreased $AUC_{(0-t)}$ and increased CL after the dosage increase. This result indicates that the 7 days-intragastric administration of diphenoxylate with medium dosage slightly induces the metabolism of phenacetin (CYP1A2) in rat, and the high dosage group induces the metabolism of phenacetin (CYP1A2), this results was consistent with **Figure 1D**. The pharmacokinetic parameters of tolbutamide experienced obvious change with decreased $AUC_{(0-t)}$ and increased CL after the dosage increase. This result indicates that the 7 days-intragastric administration of diphenoxylate with low dosage slightly induces the metabolism of tolbutamide (CYP2C9) in rat, the median and high dosage group induces the metabolism of tolbutamide (CYP2C9), this results was consistent with **Figure 1F**.

As diphenoxylate is always administrated in combination with other drugs, interactions between diphenoxylate and other drugs would increase the risk of either diminished efficacy or adverse effects. In our study, we found that 7 days-intragastric administration of diphenoxylate slightly induce the metabolism of omeprazole (CYP2C19), and induce phenacetin (CYP1A2) and tolbutamide (CYP2C9). Therefore, the metabolism and elimination of drugs would change if they are administrated in combination with diphenoxylate.

After the pharmacokinetic profiles evaluation by cocktail method, we also investigated the hepatotoxicity of diphenoxylate by observing the pathological changes of liver after diphenoxylate administration. The pathological changes of liver were observed at three difference dosages with small changes in high dosage and no change in low dosage, **Figure 2**. Therefore, high dosage diphenoxylate may cause hepatotoxicity. A more systematic and comprehensive study to investigate the hepatotoxicity of diphenoxylate will be carried out.

In conclusion, the results observed in this study would provide us valuable information regard-

ing the interactions of diphenoxylate with other drugs. Induction of drug metabolizing enzyme CYP2C19, CYP1A2 and CYP2C9 by diphenoxylate would reduce the efficacy of other drug. Additionally, high dosage diphenoxylate may cause hepatotoxicity.

Acknowledgements

This study was supported by grants from the Science and Technology Committee of Shanghai Municipality, China, No. KF1504.

Disclosure of conflict of interest

None.

Address correspondence to: Dr. Yunfang Zhou, The Laboratory of Clinical Pharmacy, The People's Hospital of Lishui, Wenzhou Medical University, Lishui 323000, China. Tel: (86) 578 2780158; E-mail: zyf2808@126.com; Dr. Meiling Zhang, Analytical and Testing Center of Wenzhou Medical University, Wenzhou 325035, China. Tel: (86) 57786699156; E-mail: meiling308@163.com

References

- [1] Firoozabadi A, Mowla A, Farashbandi H and Gorman JM. Diphenoxylate hydrochloride dependency. *J Psychiatr Pract* 2007; 13: 278-280.
- [2] Rao R, Agrawal A, Pal HR and Mohan I. Lomotil dependence: a note of caution. *Natl Med J India* 2005; 18: 330-331.
- [3] De S, Sircar BK and De SP. Diphenoxylate hydrochloride (Lomotil) in the management of cholera. *Indian J Med Res* 1975; 63: 469-474.
- [4] Ives TJ and Stults CC. Short-course diphenoxylate hydrochloride for treatment of methadone withdrawal symptoms. *Br J Psychiatry* 1983; 143: 513-514.
- [5] Nelson DR. Cytochrome P450: Structure, Mechanism, and Biochemistry. In: Paul R, editor. Ortiz de Montellano (University of California, San Francisco), editors. 3rd edition. New York: Kluwer Academic/Plenum Publishers; 2005. xx + 690 pp. \$149.00. ISBN 0-306-48324-6. *Journal of the American Chemical Society* 2005; 127: 12147-12148.
- [6] Gasche Y, Daali Y, Fathi M, Chiappe A, Cottini S, Dayer P and Desmeules J. Codeine intoxication associated with ultrarapid CYP2D6 metabolism. *N Engl J Med* 2004; 351: 2827-2831.
- [7] Stamer UM, Musshoff F, Kobilay M, Madea B, Hoeft A and Stuber F. Concentrations of tramadol and O-desmethyltramadol enantiomers in different CYP2D6 genotypes. *Clin Pharmacol Ther* 2007; 82: 41-47.
- [8] Dinger J, Meyer MR and Maurer HH. Development of an in vitro cytochrome P450 cocktail inhibition assay for assessing the inhibition risk of drugs of abuse. *Toxicol Lett* 2014; 230: 28-35.
- [9] Ma J, Wang S, Zhang M, Zhang Q, Zhou Y, Lin C, Lin G and Wang X. Simultaneous determination of bupropion, metoprolol, midazolam, phenacetin, omeprazole and tolbutamide in rat plasma by UPLC-MS/MS and its application to cytochrome P450 activity study in rats. *Biomed Chromatogr* 2015; 29: 1203-12.
- [10] Naramoto K, Kato M and Ichihara K. Effects of an Ethanol Extract of Brazilian Green Propolis on Human Cytochrome P450 Enzyme Activities in Vitro. *J Agric Food Chem* 2014; 62: 11296-302.
- [11] Lin GY, Ma JS, Xu RA, Hu LF, Wang Z and Wang XQ. Effects of Ougan juice on P450 activities using a cocktail method. *Pharmazie* 2012; 67: 242-246.
- [12] Qin CZ, Ren X, Tan ZR, Chen Y, Yin JY, Yu J, Qu J, Zhou HH and Liu ZQ. A high-throughput inhibition screening of major human cytochrome P450 enzymes using an in vitro cocktail and liquid chromatography-tandem mass spectrometry. *Biomed Chromatogr* 2014; 28: 197-203.
- [13] Borkar RM, Bhandi MM, Dubey AP, Nandekar PP, Sangamwar AT, Banerjee SK and Srinivas R. Plasma protein binding, pharmacokinetics, tissue distribution and CYP450 biotransformation studies of fidarestat by ultra high performance liquid chromatography-high resolution mass spectrometry. *J Pharm Biomed Anal* 2014; 102C: 386-399.
- [14] Tan ML and Lim LE. The effects of *Andrographis paniculata* (Burm.f.) Nees extract and diterpenoids on the CYP450 isoforms' activities, a review of possible herb-drug interaction risks. *Drug Chem Toxicol* 2014; 1-13.