

## Original Article

# Association between tumor necrosis factor alpha rs1800629 polymorphism and risk of cervical cancer

Lanling Wang<sup>1\*</sup>, Kunpeng Ma<sup>2\*</sup>, Zhiyong Wang<sup>3\*</sup>, Yingying Mou<sup>1</sup>, Li Ma<sup>1</sup>, Yong Guo<sup>4</sup>

<sup>1</sup>Department of VIP Obstetrics and Gynecology, Weifang Maternal and Child Health Hospital, Weifang 261011, Shandong, China; <sup>2</sup>Department of Otolaryngology, Affiliated Hospital of Weifang Medical University, Weifang 261011, Shandong, China; <sup>3</sup>Department of Pediatrics, Weifang Maternal and Child Health Hospital, Weifang 261011, Shandong, China; <sup>4</sup>Department of Pathological Obstetrics, Weifang Maternal and Child Health Hospital, Weifang 261011, Shandong, China. \*Equal contributors.

Received October 25, 2014; Accepted January 8, 2015; Epub February 15, 2015; Published February 28, 2015

**Abstract:** Purpose: The objective of this study was to evaluate the influence of the *TNF-α* rs1800629 polymorphism on the risk of cervical cancer. Methods: A comprehensive search of EMBASE, PubMed, Wanfang database and China National Knowledge Infrastructure (CNKI) was conducted and the papers published were retrieved. The fixed effects or random effects model was appropriately used to calculate the odds ratio (OR) along with 95% confidence interval (CI). Results: Data from sixteen articles containing nineteen studies were summarized in this meta-analysis. In general, we observed a marginal association between the *TNF-α* rs1800629 polymorphism and cervical cancer risk under the AA + AG vs. GG comparison model (OR=1.17, 95% CI=1.00-1.38, *P*=0.019 for heterogeneity). Interestingly, significantly increased risk was observed in the allele model (A vs. G: OR=1.19, 95% CI=1.02-1.38, *P*=0.006 for heterogeneity). In the stratified analysis by ethnicity, we found significant results in Caucasians. Conclusions: Our results reveal that the Caucasian population may be at increased risk of developing invasive cervical cancer associated with the *TNF-α* rs1800629 polymorphism.

**Keywords:** *TNF-α*, rs1800629, cervical cancer

## Introduction

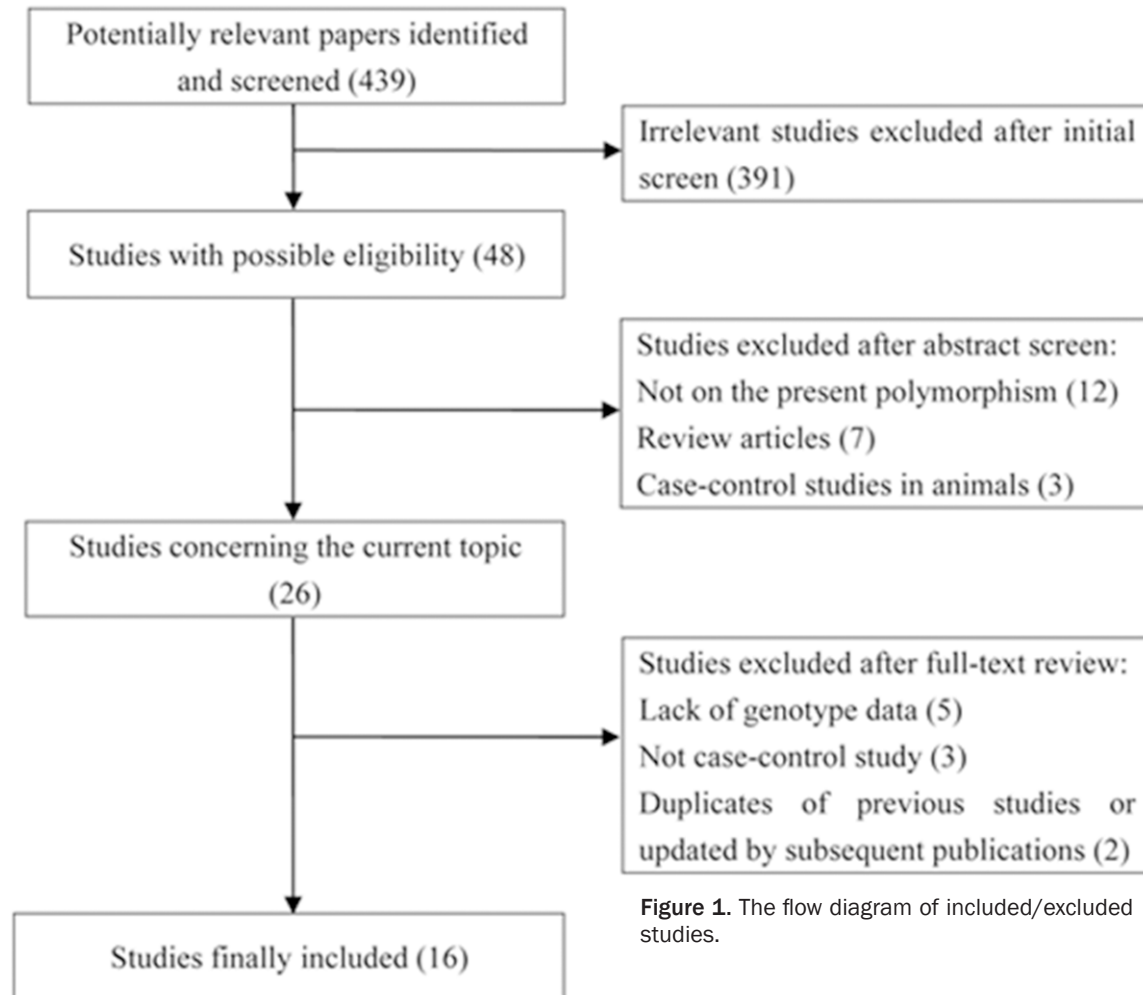
As the third most common cancer diagnosed in females, cervical cancer has been a severe health threat worldwide [1]. Decades of research has focused on the identification of etiological factors that determine the progression of this invasive cancer. It is found that human papillomavirus (HPV), a highly immunogenic virus that activates strong humoral and cellular immune responses, plays a predominant role in the initiation of cervical cancer [2, 3]. Multiple cytokines described as mediators of the immunologic response have been associated with the malignancy [4, 5].

Tumor necrosis factor alpha (*TNF-α*) is an important inflammation regulator cytokine associated with the B-cell-mediated immune response [6]. It is secreted by macrophages and functions as a proinflammatory molecular in response to immune injury and infection [7, 8]. Currently, little is known about the physiological role of this cytokine in various human

cancers. It is reported that lower expression of *TNF-α* induces invasion and transformation of cancer cells [9]. In addition, cancer type and serum level in tissues are key factors to determine how *TNF-α* acts in these cancers [10].

The *TNF-α* gene mapped on chromosome 6 is highly polymorphic. To date, there have been 16 single nucleotide polymorphisms identified in the *TNF-α* gene. A promotor polymorphism (rs1800629) which results in G to A transition at nucleotide position -308 of the transcriptional start site of the gene is directly and positively related to specific regulation of *TNF-α* synthesis at the transcriptional level [11]. Genotype-phenotype studies of the *TNF-α* rs1800629 polymorphism showed the G allele conferred two-fold lower effects on the transcription level when compared with the A allele [12]. But the transcription activity affected by the *TNF-α* rs1800629 polymorphism may vary due to different cell types [11-13].

The functional consequence of this polymorphism provided an impetus to researchers



interested in the role of *TNF-α* rs1800629 polymorphism in cervical cancer [14-16]. In general, the case-control studies have produced mixed results, with little consensus in most cases on whether the *TNF-α* polymorphism is actually associated with cervical cancer risk or not. Most importantly, several previous meta-analyses have examined the association of *TNF-α* rs1800629 polymorphism with cervical cancer risk and showed similar results [17-20], with incomplete data or duplicates included in these studies. We thereby performed a meta-analysis based on the data summarized from all available studies to evaluate the influence of the *TNF-α* rs1800629 polymorphism on the risk of cervical cancer.

## Materials and methods

### Literature search and selection

The identification of relevant studies was carried out in EMBASE, PubMed, Wanfang data-

base and China National Knowledge Infrastructure (CNKI). We retrieved the papers matching the following search terms: cervical cancer/polymorphism/polymorphisms/tumor necrosis factor-alpha/*TNF-α*/rs1800629. To assure the comprehensiveness of data, we also scanned the references of all retrieved articles. Selection of eligible studies for this meta-analysis was based on: (1) the paper must have a case-control design; (2) the influence of the *TNF-α* rs1800629 polymorphism on the risk of cervical cancer was evaluated; (3) the original article must present adequate genotype data to calculate the odds ratio (OR) along with 95% confidence interval (CI); (4) when several studies were conducted on the same case series, we considered the most recent or the largest one with complete data.

### Data extraction

Following the inclusion criteria mentioned above, two investigators selected all eligible

# Tumor necrosis factor alpha rs1800629 and cervical cancer

**Table 1.** Characteristics of all studies

First author	Year	Ethnicity	Country	HWE	Matching	Genotyping methods	Cases	Controls	Source of controls
Jang	2001	Asian	Korea	+	ND	PCR-RFLP	51	92	ND
Gostout	2003	Caucasian	USA	+	Residence area	PCR-RFLP	127	175	HB
Stanczuk	2003	African	Zimbabwe	+	ND	PCR-RFLP	103	139	HB
Deshpande*	2005	Caucasian	USA	+	ND	PCR-RFLP	115	213	HB
Deshpande*	2005	Caucasian	USA	+	ND	PCR-RFLP	143	194	HB
Govan&	2006	Mixed	South Africa	-	Age, ethnicity, residence area	PCR-RFLP	83	59	HB
Govan&	2006	African	South Africa	+		PCR-RFLP	161	169	HB
Kohaar	2007	Caucasian	India	+	Age, ethnicity	PCR-RFLP	120	165	HB
Singh	2009	Caucasian	India	-	Age, ethnicity	PCR-RFLP	150	162	HB
Wang	2009	Caucasian	Costa Rica	+	ND	PCR-RFLP	456	800	PB
Ivansson	2010	Caucasian	Sweden	+	Ethnicity	PCR-RFLP	1263	552	PB
Zu	2010	Asian	China	-	ND	PCR-RFLP	83	91	HB
Wang	2011	Asian	China	-	ND	PCR-RFLP	186	200	HB
Wang	2012	Asian	China	-	ND	PCR-RFLP	285	318	HB
Badano	2012	Caucasian	Argentina	+	age	PCR-RFLP	56	113	HB
Barbisan	2012	Caucasian	Argentina	+	ND	PCR-RFLP	122	176	HB
Huang	2012	Asian	China	+	ND	PCR-RFLP	42	87	HB
Sousa	2013	Caucasian	Portugal	+	ND	PCR-RFLP	223	205	PB

\*&, The same paper studied two distinct population; PB: population-based study; HB: hospital-based study; PCR-RFLP: polymerase chain reaction-restriction fragment length polymorphism; HWE: Hardy-Weinberg equilibrium.

**Table 2.** Meta-analysis results

Genetic contrast models	Groups	OR	95 % CI	$P_{OR}$	Heterogeneity analysis	
					$I^2$	$P$ values
AA vs. GG	Total	1.25	0.95-1.65	0.108	0.126	0.303
	Asian	1.01	0.58-1.75	0.976	0.000	0.527
	Caucasian	1.47	1.04-2.08 <sup>a</sup>	0.028	0.306	0.164
	HB	1.39	1.00-1.94 <sup>b</sup>	0.051	0.027	0.420
	PB	0.91	0.55-1.51	0.719	0.263	0.258
AG vs. GG	Total	1.13	0.95-1.34	0.160	0.433	0.026
	Asian	1.06	0.57-1.98	0.853	0.741	0.004
	Caucasian	1.09	0.91-1.30	0.336	0.305	0.165
	HB	1.15	0.91-1.46	0.234	0.505	0.016
	PB	1.11	0.87-1.41	0.406	0.432	0.172
A vs. G	Total	1.19	1.02-1.38 <sup>a</sup>	0.029	0.516	0.006
	Asian	1.10	0.75-1.60	0.636	0.582	0.048
	Caucasian	1.25	1.02-1.54 <sup>a</sup>	0.035	0.622	0.005
	HB	1.21	1.00-1.47 <sup>b</sup>	0.054	0.524	0.011
	PB	1.11	0.83-1.47	0.494	0.672	0.048
AA + AG vs. GG	Total	1.17	1.00-1.38 <sup>b</sup>	0.052	0.456	0.019
	Asian	1.09	0.68-1.75	0.711	0.639	0.026
	Caucasian	1.16	0.96-1.40	0.129	0.450	0.059
	HB	1.21	0.97-1.50	0.086	0.506	0.015
	PB	1.10	0.85-1.43	0.465	0.536	0.116
AA vs. AG + GG	Total	1.21	0.92-1.59	0.174	0.220	0.193
	Asian	0.90	0.53-1.55	0.716	0.211	0.280
	Caucasian	1.47	1.04-2.07 <sup>a</sup>	0.030	0.304	0.165
	HB	1.34	0.96-1.85	0.084	0.187	0.249
	PB	0.88	0.53-1.46	0.622	0.126	0.319

<sup>a</sup>significant results; <sup>b</sup>borderline results; PB: population-based study; HB: hospital-based study; OR: odds ratio; 95% CI: 95% confidence interval;  $P_{OR}$ :  $P$  value for the pooled ORs.

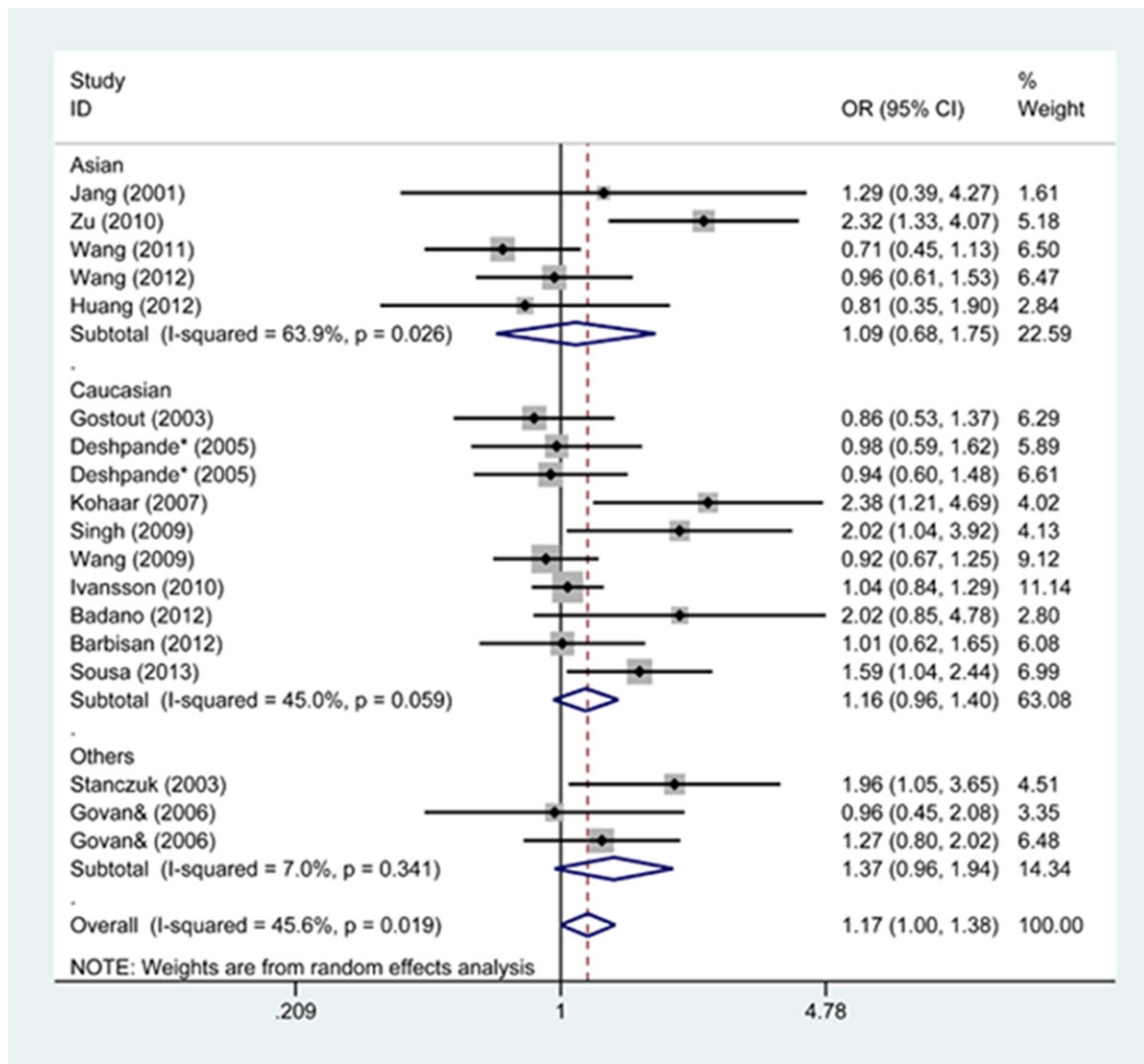
papers, from which they recorded the following information in duplicate: first author's surname, journal year of publication, ethnicity (Caucasian, Asian or African), study country, matching characteristics, source of controls (population-based or hospital-based) and genotype frequencies. An expert in this field was consulted whenever there was conflict.

#### Statistical analysis

Summary OR and 95% CI was calculated for all studies of the *TNF- $\alpha$*  rs1800629 polymorphism and the risk of cervical cancer under AA vs. GG model, AG vs. GG model, allele A vs. allele G model, AA + AG vs. GG model, and AA vs. AG +

GG model. Stratified analyses were performed in ethnicity (when the ethnicity was investigated in less than 3 papers, we categorized them into "other" group) and source of controls. Heterogeneity was measured by the  $X^2$  test and by the  $I^2$  statistic [21]. If the statistical significance level was reached ( $P < 0.1$  or  $I^2 > 50\%$ ), the values of each study were estimated using the random effects model [22]; otherwise the fixed effects model was more appropriate [23]. Publication bias was inspected by funnel plots. Egger's test was also performed to test the asymmetry or symmetry of the funnel plot [24].

Deviation from Hardy-Weinberg equilibrium (HWE) was checked using the Chi square good-



**Figure 2.** Forest plot of cervical cancer risk associated with the *TNF-α* rs1800629 polymorphism (AA + AG vs. GG) in the stratified analyses by cancer type. The squares and horizontal lines correspond to the study-specific OR and 95% CI. The area of the squares reflects the weight (inverse of the variance). The diamond represents the summary OR and 95% CI.

ness-of-fit test in each of the control populations. By sequentially omitting the single studies, we applied sensitivity analyses to detect the stability of our results. All analyses were performed using STATA version 12.0 (Stata Corporation, USA). The significant threshold was fixed at  $P < 0.1$ .

## Results

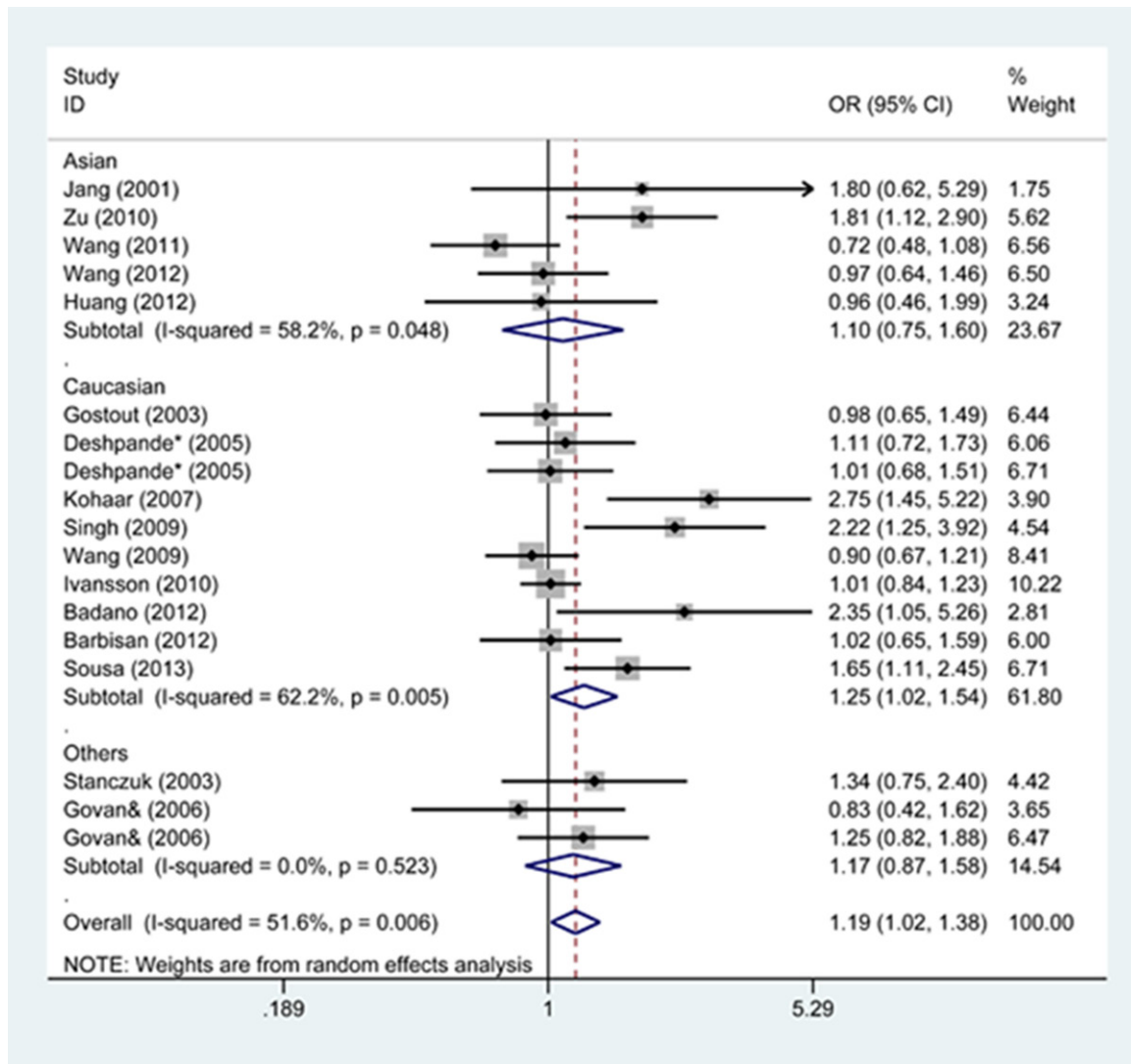
### Characteristics of the studies

As presented in **Figure 1**, we achieved a total of 16 papers [14-16, 25-37] after rigorous selection using the inclusion criteria. Of these, two

articles [25, 27] investigated the association of the *TNF-α* rs1800629 polymorphism with cervical cancer risk in two different populations, which were retrieved as separate dataset. Therefore, for the final meta-analysis, we included 19 studies, providing 3,769 cases and 3,910 controls. The details describing these included studies are summarized in **Table 1**.

### Meta-analysis results

The main meta-analysis results are listed in **Table 2**. When all of the eligible studies were pooled together, we observed a marginal association between the *TNF-α* rs1800629 poly-



**Figure 3.** Forest plot of cervical cancer risk associated with the *TNF-α* rs1800629 polymorphism (A vs. G) in the stratified analyses by cancer type. The squares and horizontal lines correspond to the study-specific OR and 95% CI. The area of the squares reflects the weight (inverse of the variance). The diamond represents the summary OR and 95% CI.

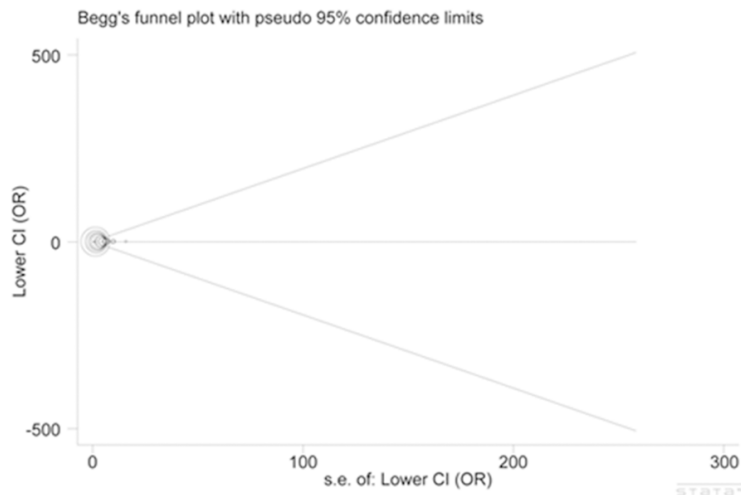
morphism and cervical cancer risk under the AA + AG vs. GG comparison model (OR=1.17, 95% CI=1.00-1.38,  $P=0.019$  for heterogeneity) (**Figure 2**). Interestingly, significantly increased risk was observed in the allele model (A vs. G: OR=1.19, 95% CI=1.02-1.38,  $P=0.006$  for heterogeneity) (**Figure 3**).

In the stratified analysis by ethnicity, we found significant results in Caucasians. The association strength was more pronounced in the AA vs. GG and the AA vs. AG + GG comparison models (OR=1.47, 95% CI=1.04-2.08,  $P=0.164$  for heterogeneity; OR=1.47, 95% CI=1.04-2.07,

$P=0.165$  for heterogeneity, respectively) (**Figure 2**), and less pronounced in the A vs. G contrast model (OR=1.25, 95% CI=1.02-1.54,  $P=0.005$  for heterogeneity) (**Figure 3**).

Finally, we estimated the association strength in stratification analysis according to source of controls. The AA vs. GG and A vs. G comparison models showed nonsignificant but borderline associations in the hospital-based studies (OR=1.39, 95% CI=1.00-1.94,  $P=0.420$  for heterogeneity; OR=1.21, 95% CI=1.00-1.47,  $P=0.011$  for heterogeneity, respectively) (**Table 2**).





**Figure 4.** Begg's funnel plot for publication bias test (model: AA vs. GG). Each point represents a separate study for the indicated association. Log [OR], natural logarithm of the odds ratio. Horizontal line, mean effect size.

#### Evaluation of heterogeneity

Significant heterogeneity was observed in this meta-analysis (**Table 2**). To identify the source of the between-study heterogeneity, we performed stratified analyses. As a result, we found both ethnicity and source of controls were contributing factors.

#### Sensitivity analysis and publication bias

We examined the stability of our results by performing the one-way sensitivity analysis. The sequential omission of the studies did not reflect any statistical alternation in the overall ORs, suggesting the evidence presented in this study was credible and compelling (data not shown). In terms of publication bias, no obvious asymmetry was revealed in the funnel plots for all comparison models. The Egger's test further confirmed the symmetry of the funnel plots with statistical evidence ( $P=0.494$  for AA vs. GG) (**Figure 4**).

#### Discussion

To date, a variety of polymorphisms in numerous genes have been investigated in cervical cancer. In the current study, the risk of cervical cancer associated with the *TNF-α* rs1800629 polymorphism was evaluated in 7,679 subjects summarized from sixteen papers with the aid of meta-analysis. Overall, we found significant results in the A vs. G allele model and nonsignificant but borderline association in the AA +

AG vs. GG model. In the stratified analysis by ethnicity, significantly increased risk of cervical cancer was observed in Caucasian population under the AA vs. GG and AA vs. AG + GG comparison models in addition to the allele model. Intriguingly, this significant association reduced to the borderline when performing the analysis according to source of controls.

As a result of the significant regulatory function in transcriptional activity of cytokine gene *TNF-α*, many clinical and epidemiological studies have focused on the rs1800629 polymorphism. The A allele and the G allele function differentially in

their effects on the transcription level of *TNF-α*. Compared with the G allele, the A allele is a much stronger transcriptional activator correlated with high *TNF-α* production [11, 12]. This is the most likely explanation for the increased risk of various diseases in relation to the rs1800629 polymorphism, especially the A allele. More specifically, the carriage of the AA genotype was discovered to have seven-fold elevated risk of death from cerebral malaria [11]. A recent statistical analysis carried out by Sousa et al. also revealed that the A allele carriers were associated with an almost 2-fold increased risk for invasive cervical cancer development [16]. Similar to the previous discoveries, in our meta-analysis, we found a fixed 19% increased risk to develop cervical cancer in the subjects carrying the A allele.

Although the rs1800629 polymorphism is a known risk factor for cervical cancer, not all association studies concerning their association showed significant results. Govan et al. associated the *TNF-α* rs1800629 polymorphism with two South African ethnic population groups, failing to find any significant association [27]. Similar results were also found in Korean population [29], Swedish population [28] and Chinese population [32]. The reported genetic effects varied across the published studies may attribute to the insufficient detection power of these relatively small sample-sized single studies.

Previously, several meta-analyses have been performed and estimated the association of the *TNF-α* rs1800629 polymorphism and cervical cancer risk. The first report gathered all case-control studies published from January 1989 to October 2010, and suggested that the *TNF-α* rs1800629 polymorphism may contribute to cervical cancer susceptibility [18]. This finding was replicated in the following studies with either a larger or relatively smaller study size [17, 19, 20]. However, considering that the overlapped data were included and more importantly, additional studies were published subsequently, we performed the quantitative analysis to further assess the association, though consistent results were revealed in our study.

We observed significant association in Caucasian population, but not in Asian population. One possibility is that *TNF-α* rs1800629 is a low-penetrant polymorphism, and the current sample size lacks statistical power to detect the slight effects. Another explanation may be the obvious heterogeneity indicated across studies. Therefore, the present findings may have been biased by the heterogeneity which may arise from differences in genetic background and source of controls. Because some studies did not report how the control subjects were recruited or selected and whether the controls were well matched with the patients [29, 31].

There are several weaknesses in this study. Firstly, publication bias may have occurred, as only published data were included in the meta-analysis. Secondly, in the subgroup of Asians, specially the Africans (the two studies were merged into "other" group for insufficient data), and the limited study size made it impossible to derive a precise estimate or even to perform a meta-analysis to assess the association. Thirdly, the impact of gene-to-gene and gene-to-environment interactions was not addressed in the analysis. Therefore, additional studies are warranted to evaluate the effect of this functional polymorphism on cervical cancer risk.

In conclusion, the evidence presented in this meta-analysis supported the consideration of a potential effect of the *TNF-α* rs1800629 polymorphism on the increased cervical cancer risk in Caucasian population. The genetic associa-

tion of the *TNF-α* rs1800629 polymorphism with cervical cancer risk requires to be further validated in larger studies in future.

## Disclosure of conflict of interest

None.

**Address correspondence to:** Dr. Yong Guo, Department of Pathological Obstetrics, Weifang Maternal and Child Health Hospital, Weifang 261011, Shandong, China. E-mail: guoyongwe@tom.com

## References

- [1] Jemal A, Bray F, Center MM, Ferlay J, Ward E and Forman D. Global cancer statistics. *CA Cancer J Clin* 2011; 61: 69-90.
- [2] Pinto LA, Castle PE, Roden RB, Harro CD, Lowy DR, Schiller JT, Wallace D, Williams M, Kopp W, Frazer IH, Berzofsky JA and Hildesheim A. HPV-16 L1 VLP vaccine elicits a broad-spectrum of cytokine responses in whole blood. *Vaccine* 2005; 23: 3555-3564.
- [3] zur Hausen H. Cervical carcinoma and human papillomavirus: on the road to preventing a major human cancer. *J Natl Cancer Inst* 2001; 93: 252-253.
- [4] Dranoff G. Cytokines in cancer pathogenesis and cancer therapy. *Nat Rev Cancer* 2004; 4: 11-22.
- [5] Stanczuk GA, Sibanda EN, Perrey C, Chirara M, Pravica V, Hutchinson IV and Tswana SA. Cancer of the uterine cervix may be significantly associated with a gene polymorphism coding for increased IL-10 production. *Int J Cancer* 2001; 94: 792-794.
- [6] Bidwell J, Keen L, Gallagher G, Kimberly R, Huizinga T, McDermott MF, Oksenberg J, McNicholl J, Pociot F, Hardt C and D'Alfonso S. Cytokine gene polymorphism in human disease: on-line databases. *Genes Immun* 1999; 1: 3-19.
- [7] Tracey KJ and Cerami A. Tumor necrosis factor, other cytokines and disease. *Annu Rev Cell Biol* 1993; 9: 317-343.
- [8] Mosaffa F, Kalalinia F, Lage H, Afshari JT and Behravan J. Pro-inflammatory cytokines interleukin-1 beta, interleukin 6, and tumor necrosis factor-alpha alter the expression and function of ABCG2 in cervix and gastric cancer cells. *Mol Cell Biochem* 2012; 363: 385-393.
- [9] Polunovsky VA, Wendt CH, Ingbar DH, Peterson MS and Bitterman PB. Induction of endothelial cell apoptosis by TNF alpha: modulation by inhibitors of protein synthesis. *Exp Cell Res* 1994; 214: 584-594.



- [10] Kroeger KM, Steer JH, Joyce DA and Abraham LJ. Effects of stimulus and cell type on the expression of the -308 tumour necrosis factor promoter polymorphism. *Cytokine* 2000; 12: 110-119.
- [11] Wilson AG, Symons JA, McDowell TL, McDevitt HO and Duff GW. Effects of a polymorphism in the human tumor necrosis factor alpha promoter on transcriptional activation. *Proc Natl Acad Sci U S A* 1997; 94: 3195-3199.
- [12] Kroeger KM, Carville KS and Abraham LJ. The -308 tumor necrosis factor-alpha promoter polymorphism effects transcription. *Mol Immunol* 1997; 34: 391-399.
- [13] Wu WS and McClain KL. DNA polymorphisms and mutations of the tumor necrosis factor-alpha (TNF-alpha) promoter in Langerhans cell histiocytosis (LCH). *J Interferon Cytokine Res* 1997; 17: 631-635.
- [14] Badano I, Stietz SM, Schurr TG, Picconi AM, Fekete D, Quintero IM, Cabrera MD, Campos RH and Liotta JD. Analysis of TNFalpha promoter SNPs and the risk of cervical cancer in urban populations of Posadas (Misiones, Argentina). *J Clin Virol* 2012; 53: 54-59.
- [15] Barbisan G, Perez LO, Contreras A and Golijow CD. TNF-alpha and IL-10 promoter polymorphisms, HPV infection, and cervical cancer risk. *Tumour Biol* 2012; 33: 1549-1556.
- [16] Sousa H, Oliveira S, Santos AM, Catarino R, Moutinho J and Medeiros R. Tumour necrosis factor alpha 308 G/A is a risk marker for the progression from high-grade lesions to invasive cervical cancer. *Tumour Biol* 2014; 35: 2561-2564.
- [17] Pan F, Tian J, Ji CS, He YF, Han XH, Wang Y, Du JP, Jiang FS, Zhang Y, Pan YY and Hu B. Association of TNF-alpha-308 and -238 polymorphisms with risk of cervical cancer: a meta-analysis. *Asian Pac J Cancer Prev* 2012; 13: 5777-5783.
- [18] Liu L, Yang X, Chen X, Kan T, Shen Y, Chen Z and Hu Z. Association between TNF-alpha polymorphisms and cervical cancer risk: a meta-analysis. *Mol Biol Rep* 2012; 39: 2683-2688.
- [19] Li M, Han Y, Wu TT, Feng Y and Wang HB. Tumor necrosis factor alpha rs1800629 polymorphism and risk of cervical lesions: a meta-analysis. *PLoS One* 2013; 8: e69201.
- [20] Zhang HL and Zhang YJ. A systemic assessment of the association between tumor necrosis factor alpha 308 G/A polymorphism and risk of cervical cancer. *Tumour Biol* 2013; 34: 1659-1665.
- [21] Higgins JP, Thompson SG, Deeks JJ and Altman DG. Measuring inconsistency in meta-analyses. *BMJ* 2003; 327: 557-560.
- [22] DerSimonian R and Laird N. Meta-analysis in clinical trials. *Control Clin Trials* 1986; 7: 177-188.
- [23] Mantel N and Haenszel W. Statistical aspects of the analysis of data from retrospective studies of disease. *J Natl Cancer Inst* 1959; 22: 719-748.
- [24] Egger M, Davey Smith G, Schneider M and Minder C. Bias in meta-analysis detected by a simple, graphical test. *BMJ* 1997; 315: 629-634.
- [25] Deshpande A, Nolan JP, White PS, Valdez YE, Hunt WC, Peyton CL and Wheeler CM. TNF-alpha promoter polymorphisms and susceptibility to human papillomavirus 16-associated cervical cancer. *J Infect Dis* 2005; 191: 969-976.
- [26] Gostout BS, Poland GA, Calhoun ES, Sohni YR, Giuntoli RL 2nd, McGovern RM, Sloan JA, Cha SS and Persing DH. TAP1, TAP2, and HLA-DR2 alleles are predictors of cervical cancer risk. *Gynecol Oncol* 2003; 88: 326-332.
- [27] Govan VA, Constant D, Hoffman M and Williamson AL. The allelic distribution of -308 Tumor Necrosis Factor-alpha gene polymorphism in South African women with cervical cancer and control women. *BMC Cancer* 2006; 6: 24.
- [28] Ivansson EL, Juko-Pecirep I and Gyllensten UB. Interaction of immunological genes on chromosome 2q33 and IFNG in susceptibility to cervical cancer. *Gynecol Oncol* 2010; 116: 544-548.
- [29] Jang WH, Yang YI, Yea SS, Lee YJ, Chun JH, Kim HI, Kim MS and Paik KH. The -238 tumor necrosis factor-alpha promoter polymorphism is associated with decreased susceptibility to cancers. *Cancer Lett* 2001; 166: 41-46.
- [30] Kohaar I, Thakur N, Salhan S, Batra S, Singh V, Sharma A, Sodhani P, Das BC, Sarkar DP and Bharadwaj M. TNFalpha-308G/A polymorphism as a risk factor for HPV associated cervical cancer in Indian population. *Cell Oncol* 2007; 29: 249-256.
- [31] Stanczuk GA, Sibanda EN, Tswana SA and Bergstrom S. Polymorphism at the -308-promoter position of the tumor necrosis factor-alpha (TNF-alpha) gene and cervical cancer. *Int J Gynecol Cancer* 2003; 13: 148-153.
- [32] Wang N, Yin D, Zhang S, Wei H, Wang S, Zhang Y, Lu Y, Dai S, Li W and Zhang Q. TNF-alpha rs1800629 polymorphism is not associated with HPV infection or cervical cancer in the Chinese population. *PLoS One* 2012; 7: e45246.
- [33] Wang Q, Zhang C, Walayat S, Chen HW and Wang Y. Association between cytokine gene polymorphisms and cervical cancer in a Chinese population. *Eur J Obstet Gynecol Reprod Biol* 2011; 158: 330-333.
- [34] Wang SS, Bratti MC, Rodriguez AC, Herrero R, Burk RD, Porras C, Gonzalez P, Sherman ME, Wacholder S, Lan ZE, Schiffman M, Chanock

- SJ and Hildesheim A. Common variants in immune and DNA repair genes and risk for human papillomavirus persistence and progression to cervical cancer. *J Infect Dis* 2009; 199: 20-30.
- [35] LiLi H and Lifa X. Research of gene polymorphism of cytokines on human papilloma virus induced cervical injury. *Journal of Bengbu Medical College* 2012; 37: 783-787.
- [36] Singh H, Jain M, Sachan R and Mittal B. Association of TNFA (-308G > A) and IL-10 (-819C > T) promoter polymorphisms with risk of cervical cancer. *Int J Gynecol Cancer* 2009; 19: 1190-1194.
- [37] Zu FY, A X, La L, Wu S and Gu ZL. A preliminary study of TNFa gene 308 single nucleotide polymorphism with cervical cancer risk and HPV subtype infection in southern Xinjiang Uygur patients. *Chinese Journal of Obstetrics and Gynecology* 2010; 45: 709-711.